

# Reduced dose maintenance eculizumab in atypical hemolytic uremic syndrome (aHUS): an update on a previous case report

Maro Ohanian  
Christian Cable  
Kathleen Halka

Department of Hematology/Oncology,  
Scott & White Healthcare and The  
Texas A&M Health Science Center,  
College of Medicine, Temple, TX, USA

**Objective:** To describe how maintenance eculizumab sustains improved renal function in severe atypical hemolytic uremic syndrome (aHUS).

**Case report:** A previously described 50-year-old woman with aHUS had a remarkable recovery with eculizumab, which safely reversed profound neurologic damage and eliminated the need for dialysis. Her recovery has been sustained on long-term eculizumab treatment. She initially received eculizumab 900 mg weekly for four doses. On week 5 she commenced maintenance therapy starting at 1200 mg every 2 weeks. Due to nausea and vomiting at that dose, the maintenance dosing was reduced to 600 mg weekly, beginning on dose seven. After receiving 600 mg weekly for nine doses, eculizumab was then reduced to 600 mg every 2 weeks, with continued improvement in renal function. This dosing is lower than the usual 1200 mg every 2 weeks described in the adult literature and used in current clinical trials of aHUS.

**Conclusion:** Six months after the initial diagnosis, our patient continues to have improved renal function on maintenance doses of eculizumab as low as 600 mg every 2 weeks.

**Keywords:** eculizumab, aHUS, thrombotic microangiopathy, atypical hemolytic uremic syndrome

## Introduction

Atypical Hemolytic Uremic Syndrome (aHUS) involves congenital or acquired chronic systemic alternative complement pathway dysregulation. This causes reduced host cell protection, dysfunctional complement activation of platelets and endothelial cells, inflammation, and thrombotic microangiopathy.<sup>1,2</sup> Terminal complement inhibition with eculizumab safely reversed our patient's neurologic changes and eliminated the need for dialysis.

While the optimal duration and frequency of therapy remains to be determined (and varies in the literature), complete and sustained terminal complement inhibition has been considered necessary for adequate control of aHUS.<sup>3</sup> Previous publications have indicated that long-term eculizumab therapy helps avoid aHUS relapse and maintenance therapy is used in current aHUS clinical trials.<sup>1-6</sup>

Eculizumab has been reported to be safe and effective in aHUS, a condition of chronic systemic uncontrolled complement activation.<sup>4-15</sup> Eculizumab is a high-affinity humanized monoclonal anti-C5 antibody that blocks terminal complement activity. Eculizumab binds to and blocks cleavage of the terminal complement protein C5 into its pro-inflammatory, prothrombotic, and lytic products: C5a and the cytotoxic

Correspondence: Maro Ohanian  
Department of Hematology/Oncology,  
Scott & White Healthcare, 2401 South  
31st Street Temple, TX 76508, USA  
Tel +1 281 217 1974  
Fax +1 254 771 4890  
Email ohanianus@yahoo.com

membrane-attack complex C5b-9. Upstream components of complement, most notably C3b, are left intact.<sup>3,7,15,16</sup>

While eculizumab is not yet FDA approved for aHUS, this update describes how long term eculizumab maintains and improves renal function, despite being used at doses lower than currently used in clinical trials. This update also reports the results of our patient's genetic and antibody tests, all of which were normal.

## Case summary and update

A previously described 50-year-old Caucasian woman was transferred to our intensive care unit with sepsis, pancolitis, acute renal failure, and thrombocytopenia. Low complement levels were noted: C3 was 35 (83–184) mg/dL and C4 was 6 (17–59) mg/dL. ADAMTS13 activity was adequate at 63%. Stool studies were all negative (including *Escherichia coli* 0157:H7 and *Clostridium difficile*). She underwent colectomy and was confirmed to have scattered fibrin microthrombi on her bowel specimen. Her renal failure worsened and continuous venovenous hemodialysis was initiated. A rapid neurologic decline occurred after surgery. Severe renal failure, profound neurologic changes, severe hemolysis, thrombotic microangiopathy, and low complement levels were all suggestive of atypical hemolytic uremic syndrome (aHUS). By hospital day 5, there were occasional schistocytes on the peripheral smear. Since aHUS is characterized by chronic systemic uncontrolled complement activation,<sup>2</sup> the anti-C5 monoclonal antibody eculizumab (900 mg IV) was administered on hospital day 6. It was continued weekly for four doses, followed by maintenance therapy that was gradually reduced.<sup>16</sup>

Because eculizumab increases the risk of *Neisseria meningitidis*, the meningococcal vaccine was given prior to the first dose, and 14 days of ciprofloxacin was prescribed for meningococcal prophylaxis.<sup>16</sup>

Her renal recovery was remarkable with eculizumab and she has remained off dialysis with long-term eculizumab treatment. Our patient's rapid and profound neurologic deterioration resulted from significant thrombotic microangiopathy. MRI showed a right parietal lobe infarction (images shown in original paper). Despite thrombotic microangiopathic brain injury, she had a swift and complete neurologic recovery with eculizumab.

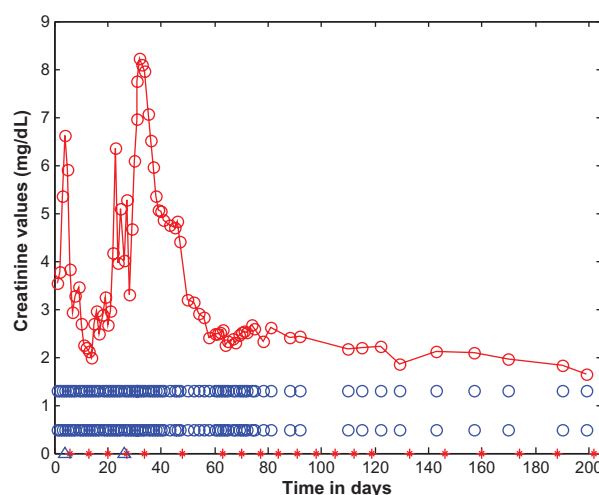
The initial dosing schema used for our patient is similar to that used in the clinical trials of eculizumab in aHUS, but maintenance therapy has gradually been reduced over time to a dose lower than what is used in the trials. Maintenance therapy, starting on week 5, was planned to be 1200 mg

every 2 weeks, as in the trials; however, this dose was only continued for doses five and six.<sup>17–19</sup> Due to nausea at the 1200 mg dose, the seventh dose was delayed by 1 day because the patient refused it, but then agreed to a reduced dose of 600 mg weekly. Her creatinine continued to improve despite the dose modification. She then remained on 600 mg weekly for nine doses with stable renal function. Her dose was subsequently reduced to 600 mg every 2 weeks and her improved renal function has been maintained despite the dose reduction. Additionally, her creatinine has continued to decrease on maintenance therapy with recent levels as low as 1.64 mg/dL (normal 0.5–1.3 mg/dL), despite maintenance dosing reduced to 600 mg every 2 weeks. Figures 1–6 show treatment responses over time with respect to creatinine, C3, C4, LDH, haptoglobin, and platelets.

## Discussion

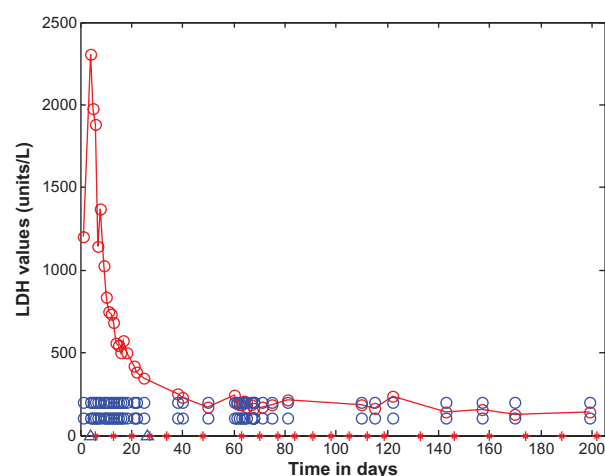
### Maintenance eculizumab

Several authors have described how long-term eculizumab therapy helps avoid aHUS relapse.<sup>1–6</sup> Mache et al describe how aHUS relapsed after discontinuation of eculizumab and resulted anuric end stage renal disease.<sup>1</sup> They describe another patient who experienced improvement in renal function, as well as normalization of platelets and haptoglobin, within 5 days of receiving eculizumab 600 mg. Unfortunately, atypical hemolytic uremic syndrome relapsed 2 weeks after recovery of complement-mediated hemolytic



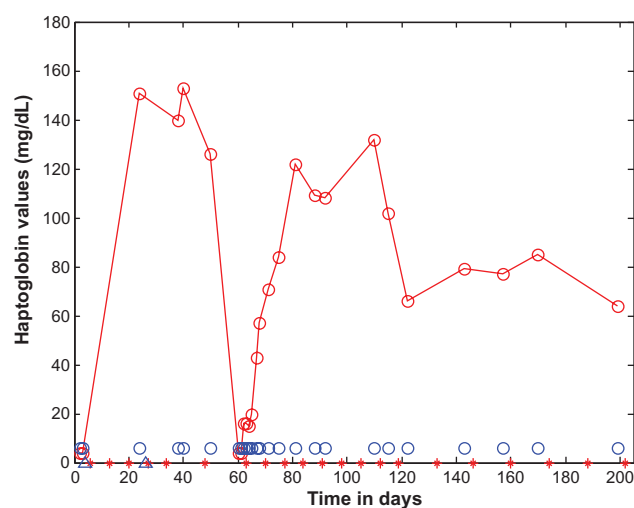
**Figure 1** Serum creatinine levels as a function of time (days).

**Notes:** Creatinine levels are marked with open red circles and the data points are connected with straight lines. For the purposes of reference, the normal creatinine range (0.5–1.3 mg/dL) is shown as open blue circles. The days corresponding to eculizumab administration are indicated by red asterisks along the horizontal time axis, appearing on days: 6, 13, 20, 27, 34, 48, 63, 70, 77, 84, 91, 98, 105, 112, 119, 133, 146, 160, 174, 188, and 202. The start and end dates of dialysis administration are marked on the time axis of the figure by two blue open triangles, on days 4 and 26.



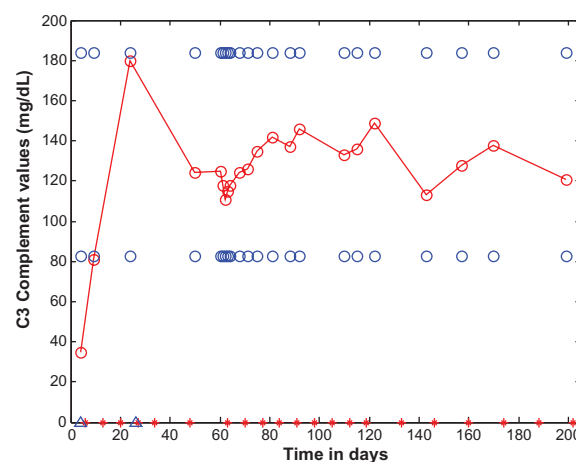
**Figure 2** Lactate dehydrogenase as a function of time (days).

**Notes:** Lactate dehydrogenase levels are marked with open red circles and the data points are connected with straight lines. For the purposes of reference, the normal range (108–202 U/L) is shown as open blue circles. Lactate dehydrogenase is shown as a function of time (days). Lactate dehydrogenase gradually dropped after initiating treatment with eculizumab. Eculizumab administration is denoted by vertical tick marks (days 6, 13, 20, 27, 34, 48, 63, 70, 77, 84, 91, 98, 105, 112, 119, 133, 146, 160, 174, 188, and 202). The start and end dates of dialyses administration are marked on the time axis of the figure by two blue open triangles, on days 4 and 26. Four days after the first dose, lactate dehydrogenase dropped to 837 U/L. After the second dose of eculizumab on day 13, lactate dehydrogenase dropped to 556 U/L. A week after the third dose, lactate dehydrogenase was down to 349 U/L. Three days after the sixth dose, lactate dehydrogenase was normal. It did rise slightly before the seventh dose was due.



**Figure 3** Haptoglobin as a function of time (days).

**Notes:** Haptoglobin levels are marked with open red circles and the data points are connected with straight lines. For the purposes of reference, the normal range (62–197 mg/dL) is shown as open blue circles. Eculizumab administration is denoted by vertical tick marks (days 6, 13, 20, 27, 34, 48, 63, 70, 77, 84, 91, 98, 105, 112, 119, 133, 146, 160, 174, 188, and 202). Initial haptoglobin was <6 mg/dL (normal range 62–197 mg/dL). Five days after the third dose, haptoglobin was normal and remained normal until the time the seventh dose was due. Haptoglobin then dropped precipitously, indicating an ongoing need for the drug. Haptoglobin did increase again after the seventh dose on day 63, and later normalized to 71 mg/dL 3 days after receiving the eighth dose on day 69. The start and end dates of dialyses administration are marked on the time axis of the figure by two blue open triangles, on the days 4 and 26.



**Figure 4** C3 as a function of time (days).

**Notes:** C3 levels are marked with open red circles and the data points are connected with straight lines. For the purposes of reference, the normal range (83–184 mg/dL) is shown as open blue circles. C3 is shown as a function of time (days). Eculizumab administration is denoted by vertical tick marks (days 6, 13, 20, 27, 34, 48, 63, 70, 77, 84, 91, 98, 105, 112, 119, 133, 146, 160, 174, 188, and 202). The start and end dates of dialyses administration are marked on the time axis of the figure by two blue open triangles, on the days 4 and 26. After the second dose of eculizumab on day 13, C3 rose to 81. Thirteen days after the sixth dose, C3 was normal and remained normal.

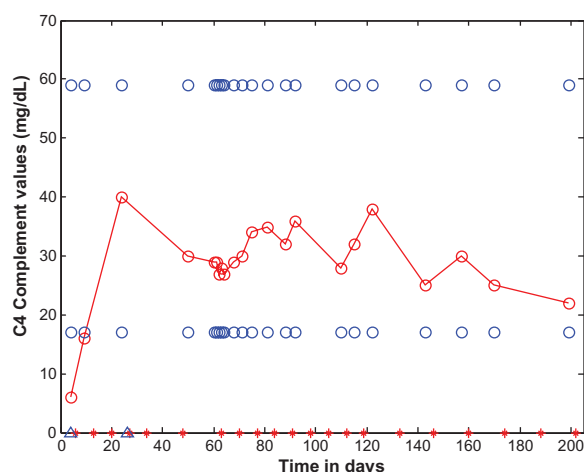
activity, suggesting the need for sustained complement inhibition to prevent relapse.

Châtelet et al described a transplant patient who was on a maintenance eculizumab 1200 mg every 14 days.<sup>2</sup> They reported that a 6-day delay in administering the ninth dose resulted in recurrent hemolysis and deterioration of transplant function. The authors consequently resumed eculizumab as permanent therapy for their patient, with recovery of renal function. Similarly, in our patient, when the seventh dose was due, laboratory investigations suggested relapsing hemolysis, with haptoglobin <6 mg/dL, indicating her continued need for the drug.

Zuber et al mention similar cases which emphasize the importance of long-term eculizumab therapy.<sup>3</sup> They describe two post-transplant aHUS cases in which a single dose of eculizumab resulted in long-term remission in both patients. However, in both cases relapse ultimately occurred. Relapse occurred after 11.5 months for one patient and eculizumab was resumed, resulting in a remission. The other patient relapsed after 21 months, but when eculizumab was resumed, graft loss could be not prevented. Zuber et al suggest that a single dose of eculizumab is less effective than prolonged therapy and argue that the interval between doses be no more than 14 days.<sup>3</sup>

## Genetic testing in aHUS

Approximately 60% of aHUS cases (familial and sporadic) have been associated with genetic mutations in complement



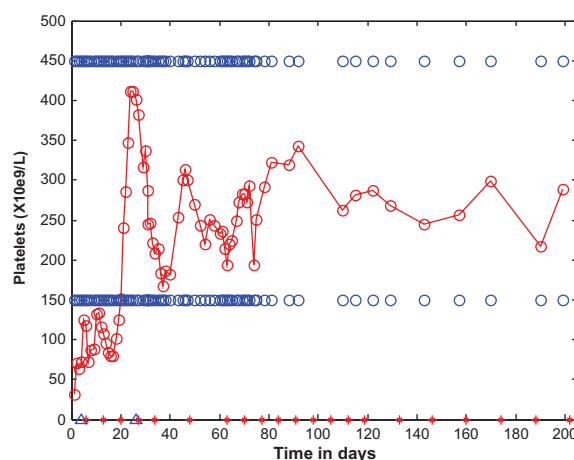
**Figure 5** C4 as a function of time (days).

**Notes:** C4 levels are marked with open red circles and the data points are connected with straight lines. For the purposes of reference, the normal range (17 to 59 mg/dL) is shown as open blue circles. C4 is shown as a function of time (days). Ecuzumab administration is denoted by vertical tick marks (days 6, 13, 20, 27, 34, 48, 63, 70, 77, 84, 91, 98, 105, 112, 119, 133, 146, 160, 174, 188, and 202). The start and end dates of dialyses administration are marked on the time axis of the figure by two blue open triangles, on days 4 and 26. The day after the second dose of ecuzumab on day 13, C4 rose to 16. Three days after the sixth dose, C4 was normal and remained normal.

regulatory proteins of the alternative complement pathway or auto-antibodies to the complement regulatory factor CFH.<sup>19,20</sup> Consequently, we ordered the following genetic and antibody tests for our patient: *CFH*, *CFB*, *CFI*, *CFHR3-CFHR1* (deletion testing), *MCP*, *THBD*, *C3*, and CFH auto-antibodies (serum test). The Complement Hemolytic assay (serum test) was also ordered. All these results were normal for our patient.

As gene sequencing is very time consuming, it can take several months for all of the genetic tests to return. In the case of our patient, all of the results were available 6 months after they were ordered. Furthermore, very few clinical laboratories do genetic testing for aHUS, but such labs may be found on the National Center for Biotechnology Information (NCBI) website: <http://www.genetests.org/>.<sup>21</sup> Our patient's genetic tests were performed at University of Iowa, which, to our knowledge, may be the only lab in the US that does these tests.<sup>22</sup>

Atypical HUS has been linked to the chromosome 1q32 locus containing genes for complement factor H (CFH), the plasma regulator of the alternative complement pathway, as well as for other complement regulators. Over 80 CFH mutations have been identified in aHUS patients. In familial aHUS the frequency of CFH mutations is 40%–50%. In the sporadic form, the frequency of CFH mutations is 20%.<sup>19,20</sup> The various genetic abnormalities and antibody abnormalities currently identified in aHUS are summarized below. More information



**Figure 6** Platelets as a function of time (days).

**Notes:** Platelet levels are marked with open red circles and the data points are connected with straight lines. For the purposes of reference, the normal range (150–450  $\times 10^9/L$ ) is shown as open blue circles. The days corresponding to ecuzumab administration are indicated by red asterisks along the horizontal time axis, appearing on days: 6, 13, 20, 27, 34, 48, 63, 70, 77, 84, 91, 98, 105, 112, 119, 133, 146, 160, 174, 188, and 202. On the day of transfer, hospital day 1, the patient underwent total abdominal colectomy. During surgery, she received 2U of pooled platelets. Initial platelets before surgery were  $28 \times 10^9/L$ . After surgery, her platelets rose, and continued to rise during the first 5 days of the admission, without additional transfusions. The rising platelets after colectomy suggested previous platelet consumption by a thrombotic microangiopathic process in the colon. After initiating ecuzumab, her platelets continued to rise.

on genetic mutations in aHUS can be obtained from two comprehensive papers by Noris and Remuzzi.<sup>19,20</sup>

- *CFH* regulates the alternative complement pathway (AP). In aHUS, most *CFH* mutations are heterozygous.<sup>19,20</sup> Mutant *CFH* heterozygotes usually carry normal levels of CFH but 50% of the molecules are dysfunctional. Only 15%–20% of *CFH* mutations in aHUS are homozygous, resulting in quantitative CFH deficiency and very low C3 levels.<sup>19,20</sup>
- 3%–5% of aHUS cases possess a heterozygous hybrid gene. The hybrid gene results from aberrant crossover between *CFH* and *CFHR1*, a gene located in tandem to *CFH*. *CFHR1* encodes complement factor H related protein (CFHR1). The gene product has decreased endothelial complement regulatory activity.<sup>19,20</sup>
- Deletion testing for *CFHR1* and *CFHR3* was ordered because these gene deletions are considered a risk factor for aHUS.<sup>23</sup>
- Approximately 6%–10% of aHUS patients acquire anti-CFH antibodies, which emulate *CFH* mutations by binding to the *CFH* C-terminus and decreasing CFH binding to C3b and surface cofactor activity.<sup>19,20</sup> Furthermore, some aHUS patients acquire autoantibodies to Factor H in the setting of deletions of *CFHR1* and *CFHR3*.<sup>24</sup>
- *CFI* is a plasma serine protease that regulates all three complement pathways by cleaving C3b and C4b.



4%–10% of aHUS cases involve a heterozygously mutated *CFI*.<sup>19,20</sup> As a proteolytic enzyme, CFI prevents hemolytic and immune-adherence activities of activated, cell-bound C3.<sup>22</sup>

- *MCP* encodes the membrane cofactor protein (MCP), which is a cofactor for CFI to cleave C3b and C4b on cell surfaces. Around 10% of aHUS cases involve *MCP* mutations, the majority of which are heterozygous.<sup>19,20</sup>
- Some patients have gain of function mutations that affect genes encoding CFB and C3, which are the AP C3 convertase components. *CFB* mutations, accounting for only 1%–2% of aHUS cases, result in chronic AP activation with low C3 level and normal C4 levels.<sup>19,20</sup> Approximately 4%–10% of aHUS cases have heterozygously mutated C3, associated with reduced C3b binding to CFH and MCP, significantly hindering degradation of mutant C3b and resulting in low C3 levels.<sup>19,20</sup>
- Around 5% of aHUS patients have heterozygously mutated *THBD*, resulting in inefficient inactivation of C3b. *THBD* encodes thrombomodulin that is involved in complement activation on cell surfaces.<sup>19,20</sup> Thrombomodulin is actually a negative regulator of the complement system, which suppresses leukocyte trafficking, inflammation, and complement activation.<sup>22,25</sup>
- Of note, 3%–5% of aHUS patients possess combined mutations, often in *CFH* along with either *MCP* or *CFI*. Few also possess anti-CFH antibodies.<sup>19,20</sup>

Although genetic testing did not change the management of our patient, genetic testing in aHUS can provide the clinician with information about disease process and long-term renal transplant outcome.<sup>19,20</sup> For example, mutations of *CFH*, *CFI*, *CFB*, and *C3* have been associated with post-transplant recurrence. This is due to the fact that the mutations give rise to abnormalities in circulating proteins that mostly come from the liver, and thus can exist even after kidney transplant.<sup>19,20</sup>

## Conclusion

Six months after initial diagnosis, our patient continues to have improved renal function on maintenance doses of eculizumab as low as 600 mg every 2 weeks. This dose is much lower than what is used in the current clinical trials, suggesting that complement inhibition needs vary among patients. Clinical trials are underway to determine the optimal dosing schema.

Although genetic abnormalities in complement regulatory factors or anti-CFH antibodies have been identified in approximately 60% of cases,<sup>19,20</sup> genetic testing did not change the management of this patient.

## Acknowledgment

The authors would like to acknowledge Camille Bedrosian, MD, for her important recommendations on the treatment of aHUS.

## Disclosure

The authors report no conflicts of interest in this work.

## References

1. Mache CJ, Acham-Roschitz B, Frémeaux-Bacchi V, et al. Complement inhibitor eculizumab in atypical hemolytic uremic syndrome. *Clin J Am Soc Nephrol*. 2009;4(8):1312–1316.
2. Châtelet V, Lobbedez T, Frémeaux-Bacchi V, Fichoux M, Ryckelynck JP, Hurault de LB. Eculizumab: safety and efficacy after 17 months of treatment in a renal transplant patient with recurrent atypical hemolytic-uremic syndrome: case report. *Transplant Proc*. 2010;42(10):4353–4355.
3. Zuber J, Le Quintrec M, Sberro-Soussan R, Loirat C, Frémeaux-Bacchi V, Legendre C. New insights into postrenal transplant hemolytic uremic syndrome. *Nat Rev Nephrol*. 2011;7(1):23–35.
4. Nurnberger J, Phillip T, Witzke O, et al. Eculizumab for atypical hemolytic-uremic syndrome. *N Engl J Med*. 2009;360(5):542–544.
5. Châtelet V, Frémeaux-Bacchi V, Lobbedez T, Fichoux M, Hurault de LB. Safety and long-term efficacy of eculizumab in a renal transplant patient with recurrent atypical hemolytic-uremic syndrome. *Am J Transplant*. 2009;9(11):2644–2645.
6. Gruppo RA, Rother RP. Eculizumab for congenital atypical hemolytic-uremic syndrome. *N Engl J Med*. 2009;360(5):544–546.
7. Davin JC, Gracchi V, Bouts A, Groothoff J, Strain L, Goodship T. Maintenance of kidney function following treatment with eculizumab and discontinuation of plasma exchange after a third kidney transplant for atypical hemolytic uremic syndrome associated with a CFH mutation. *Am J Kidney Dis*. 2010;55(4):708–711.
8. Zimmerhackl LB, Hofer J, Cortina G, et al. Prophylactic eculizumab after renal transplantation in atypical hemolytic-uremic syndrome. *N Engl J Med*. 2010;362(18):1746–1748.
9. Ardissino G, Testa S, Paglialonga F, et al. Remission of plasma-resistant atypical hemolytic uremic syndrome relapse on kidney graft with eculizumab. Presented at the 2nd International Conference on HUS-MPGN-PNH; June 13–15, 2010; Innsbruck, Austria. p. 17.
10. Haffner H, Hofer J, Zimmerhackl LB, Pohl M. Effective eculizumab therapy of familial atypical HUS in a 4 year old patient. Presented at the 2nd International Conference on HUS-MPGN-PNH; 2010 June 13–15; Innsbruck, Austria. p. 21.
11. Lapayraque AL, Phan V, Clermont MJ, et al. Effectiveness of eculizumab in a plasma infusion dependent patient with atypical haemolytic and uremic syndrome (a HUS) associated with heterozygous combined de Novo mutation in factor H gene. Presented at the 2nd International Conference on HUS-MPGN-PNH; June 13–15, 2010; Innsbruck, Austria. p. 24.
12. Tschumi S, Bucher BS, Sparta G, et al. Eculizumab in atypical hemolytic uremic syndrome: long-term clinical course and histological findings. Presented at the 2nd International Conference on HUS-MPGN-PNH; June 13–15, 2010; Innsbruck, Austria. p. 44.
13. Larrea CF, Cofan F, Oppenheimer F, Campistol JM, Escolar G, Lozano M. Efficacy of eculizumab in the treatment of recurrent atypical hemolytic-uremic syndrome after renal transplantation. *Transplantation*. 2010;89(7):903–904.
14. Legault DJ, Boelkins MR. Successful treatment of aHUS recurrence and arrest of plasma exchange resistant TMA post-renal transplantation with the terminal complement inhibitor eculizumab. Presented at the 51st ASH Annual meeting; December 6, 2009; Grand Rapids, MI. p. 2421.

15. Dmytrijuk A, Robie-Suh K, Cohen MH, Rieves D, Weiss K, Pazdur R. FDA report: eculizumab (Soliris) for the treatment of patients with paroxysmal nocturnal hemoglobinuria. *Oncologist*. 2008;13(9):993–1000.
16. Ohanian M, Cable C, Halka K. Eculizumab safely reverses neurologic impairment and eliminates need for dialysis in severe atypical hemolytic uremic syndrome. *Clinical Pharmacology: Advances and Applications*. doi: 10.2147/CPAA.S17904.
17. Alexion Pharmaceuticals. Open Label Controlled Trial of Eculizumab in Adult Patients With Plasma Therapy-sensitive Atypical Hemolytic Uremic Syndrome (aHUS). In: ClinicalTrials.gov [Internet]. Bethesda, MD: National Library of Medicine (US). 2009. Available from: <http://www.clinicaltrials.gov/ct2/show/NCT00838513?term=NCT00838513&rank=1>. Accessed on October 5, 2011. NLM identifier: NCT00838513.
18. Muus P, Legendre C, Douglas K, Hourmant M, Delmas Y, Herthelius B, et al. Safety and Efficacy of Eculizumab in aHUS Patients on Chronic Plasma Therapy: Interim Analysis from a Phase II Trial [abstract]. Presented at American Society of Nephrology Meeting, Renal Week 2010, Denver, CO. Available at: [http://www.abstracts2view.com/asn10/view.php?nu=ASN10L1\\_1338a](http://www.abstracts2view.com/asn10/view.php?nu=ASN10L1_1338a). Accessed on October 5, 2011.
19. Noris M, Remuzzi G. Genetics and genetic testing in hemolytic uremic syndrome/thrombotic thrombocytopenic purpura. *Semin Nephrol*. 2010;30(4):395–408.
20. Noris M, Remuzzi G. Atypical hemolytic-uremic syndrome. *N Engl J Med*. 2009;361(17):1676–1687.
21. GeneTests [homepage]. Available at: <http://www.ncbi.nlm.nih.gov/sites/GeneTests>. Accessed on October 5, 2011.
22. Molecular Otolaryngology and Renal Research Laboratories [homepage]. Available at: <http://www.healthcare.uiowa.edu/labs/morl/THBD.htm>. Accessed on October 5, 2011.
23. Zipfel PF, Edey M, Heinen S, et al. Deletion of complement factor H-related genes CFHR1 and CFHR3 is associated with atypical hemolytic uremic syndrome. *PLoS Genet*. 2007;3(3):e41.
24. Skerka C, Józsi M, Zipfel PF, Dragon-Durey MA, Frémeaux-Bacchi V. Autoantibodies in haemolytic uraemic syndrome (HUS). *Thromb Haemost*. 2009;101(2):227–232.
25. Delvaeye M, Noris M, De Vriese A, et al. Thrombomodulin mutations in atypical hemolytic-uremic syndrome. *N Engl J Med*. 2009;361(4):345–357.

## Clinical Pharmacology: Advances and Applications

### Publish your work in this journal

Clinical Pharmacology: Advances and Applications is an international, peer-reviewed, open access journal publishing original research, reports, reviews and commentaries on all areas of drug experience in humans. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use.

Submit your manuscript here: <http://www.dovepress.com/clinical-pharmacology-advances-and-applications-journal>

Dovepress

Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.