

Reversal of hepatic fibrosis: pathophysiological basis of antifibrotic therapies

Mona H Ismail¹
Massimo Pinzani²

¹Department of Internal Medicine, Division of Gastroenterology, King Fahad University Hospital, Al-Khobar, Saudi Arabia; ²Dipartimento di Medicina Interna Center for Research, High Education and Transfer, Università degli Studi di Firenze, Florence, Italy

Abstract: Chronic liver injuries of different etiologies eventually lead to fibrosis, a scarring process associated with increased and altered deposition of extracellular matrix in the liver. Progression of fibrosis has a major worldwide clinical impact due to the high number of patients affected by chronic liver disease which can lead to severe complications, expensive treatment, a possible need for liver transplantation, and death. Liver fibrogenesis is characterized by activation of hepatic stellate cells and other extracellular matrix producing cells. Liver fibrosis may regress following specific therapeutic interventions. Other than removing agents causing chronic liver damage, no antifibrotic drug is currently available in clinical practice. The extent of liver fibrosis is variable between individuals, even after controlling for exogenous factors. Thus, host genetic factors are considered to play an important role in the process of liver scarring. Until recently it was believed that this process was irreversible. However, emerging experimental and clinical evidence is starting to show that even cirrhosis in its early stages is potentially reversible.

Keywords: liver fibrosis, cirrhosis, fibrogenesis, antifibrotic agents

Introduction

Chronic liver disease is a major cause of mortality and morbidity worldwide. Most chronic liver diseases progress from mild inflammation to more severe inflammation, leading to fibrosis or cirrhosis. This can result in liver failure and portal hypertension, and is associated with an increased risk of liver cancer.¹ The development of advanced fibrosis, and particularly cirrhosis, is associated with significant and life-threatening complications, with liver transplantation being the only available treatment. However, transplantation is not always possible, due to limited organ availability and the presence of contraindicating comorbidities.

Fibrosis is a progressive pathological process in which the body's wound healing and tissue remodeling mechanisms respond to liver injury by promoting replacement of normal hepatic tissue with a scar-like matrix composed of cross-linked collagen.² Until recently, it was believed that this process was irreversible.³ However, emerging experimental and clinical evidence is starting to show that even early stages of cirrhosis are potentially reversible.

The cascade of events leading to hepatic fibrosis is complex, and is influenced by how different cell types in the liver interact in response to injury. The main cell type responsible for the development of fibrosis in chronic liver disease is the activated hepatic stellate cell. Chronically activated hepatic stellate cells proliferate and synthesize extracellular matrix proteins to produce the fibrous scar (Figure 1).⁴

Correspondence: Mona H Ismail
College of Medicine, University
of Dammam, Department of Internal
Medicine, Division of Gastroenterology,
King Fahad Hospital, PO Box 40149,
Al-Khobar 31952, Saudi Arabia
Tel +966 3896 6741
Fax +966 3896 6741
Email mismail.md@gmail.com

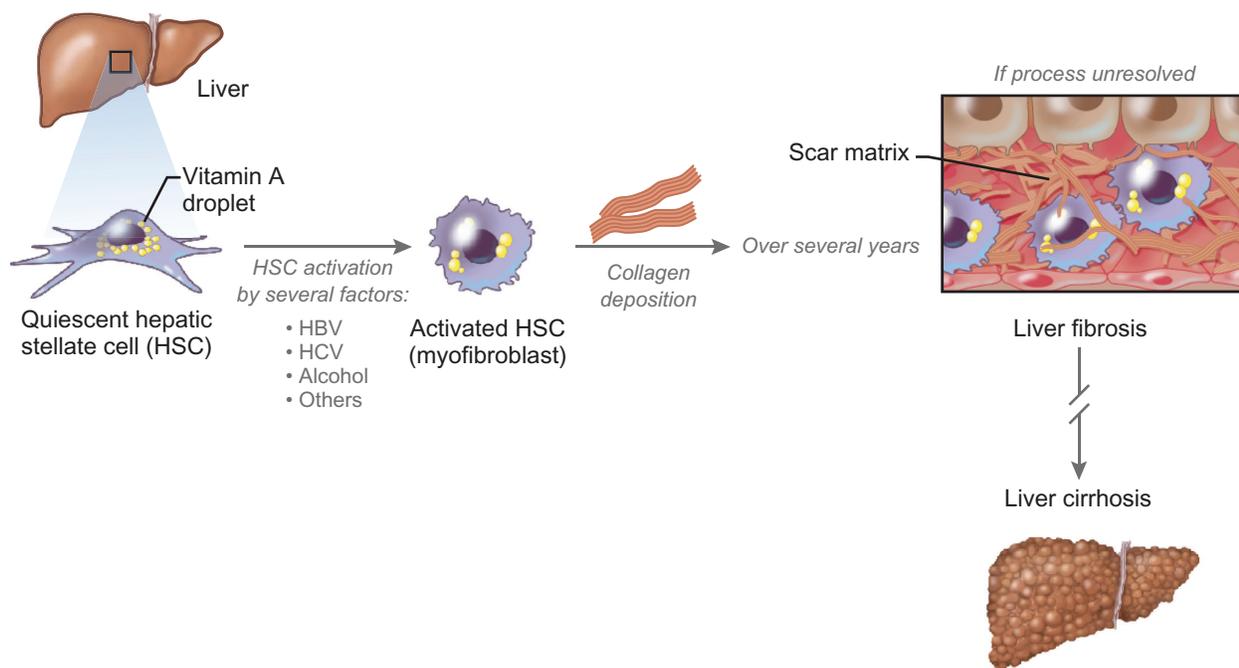


Figure 1 Schematic presentation of liver fibrogenesis.

Following any type of liver injury, hepatic stellate cells differentiate into myofibroblasts and acquire the typical “myofibroblast-like” phenotype characterized by a multifunctional profibrogenic, proinflammatory, and proangiogenic profile. Reported phenotypic changes garnered from hepatic stellate cell culture studies include increased proliferation, increased expression levels of platelet-derived growth factor- β and smooth muscle α -actin (α -SMA), and enhanced collagen secretion.^{5–8} More recent evidence implicates a role for leptin in hepatic stellate cell transdifferentiation by activating the hedgehog pathway, and galectin-3 for hepatic stellate cell activation *in vivo*.^{9,10}

In addition to hepatic stellate cells, other resident liver mesenchymal cells, such as portal fibroblasts and myofibroblasts, contribute to hepatic fibrogenesis, particularly in chronic liver diseases characterized by primary involvement of the portal and periportal areas (ie, chronic viral hepatitis and cholestatic diseases).¹¹

There is some evidence that bone marrow-derived fibrocytes or circulating mesenchymal cells can migrate through the injured liver and become myofibroblasts, which participate in the fibrotic process.^{12,13} In addition, there is controversial evidence that hepatocytes and cholangiocytes may undergo epithelial–mesenchymal transition to become activated myofibroblasts.¹⁴ Likewise, phenotypic agility associated with epithelial–mesenchymal transition allows for the reverse process whereby mesenchymal cells convert into epithelial derivatives.¹⁴

Among the three types of proposed epithelial–mesenchymal transitions, type 2 epithelial–mesenchymal transition refers to the process occurring in chronic fibrogenic disorders. In the context of fibrogenesis, synthesis and deposition of type I collagens most appropriately signal myofibroblast function *in vivo*.¹⁴ However, despite *in vitro* evidence supporting $1\alpha 1$ gene activation in cultured epithelial cells, including hepatocytes, these cells fail to generate collagen $1\alpha 1$ transcripts *in vivo*.^{14–16} In addition, some cells, particularly macrophages, internalize the collagen fibrillar extracellular matrix that leads to mistaken association of collagen accumulation with collagen synthesis. These contradictions beg the question of whether epithelial cells really play a role in fibrogenesis.^{14,15,17–19}

Many cellular markers have been wrongly defined as myofibroblast-specific based on gene expression patterns. For example, certain hepatocytes and cholangiocytes express “mesenchymal markers” typical of cell motility and survival. Hepatocytes when cultured *in vitro* and chronically stimulated with transforming growth factor- $\beta 1$ or serum factors exhibit genetic expression patterns analogous to myofibroblasts *in vivo* and connective tissue at the development stage.^{14,20–24} Some of the genes expressed include α -SMA, Slug, Twist, Snail, vimentin, and fibroblast-specific protein implicated in cell motility in general, although these genes do not mark myofibroblast or epithelial cell lineages specifically.^{14,15,18,25–28}

Oddly, α -SMA, that forms part of the cell's contractile machinery, constitutes a poor lineage marker for fibrogenesis, although it is the most widely used marker for recognizing myofibroblasts. Activation of this protein expression during disease or as part of an adaptive response to injury, compounded by the fact that it contributes minimally, if at all, to the synthesis and deposition of fibrillar extracellular matrix, explains the ambiguous role of α -SMA in marking myofibroblastic activity.¹⁴

A direct contribution of cholangiocytes to fibrosis via an epithelial–mesenchymal transition pathway has been proposed by Omenetti et al who demonstrated that immature cholangiocytes undergo a complete epithelial–mesenchymal transition when treated with activated hepatic stellate cell medium.²⁹ Others have shown coexpression of epithelial and mesenchymal markers in human liver cholangiocytes extracted from patients with cholestatic disease.^{30,31} However, recent reports have challenged the concept of cholangiocyte epithelial–mesenchymal transition, with conclusions pointing towards an absence of this process in liver fibrosis models.³² Specifically, fluorescently labeled cholangiocytes expressing the bile ductular cell-specific marker, K19, failed to coexpress myofibroblast markers in either bile duct-ligated or CCL4-induced fibrosis mouse models.³³ Clearly, further explorations beyond intuitive speculation are needed to unravel the role of epithelial cells and cholangiocytes in fibrogenesis, if any.

The development of significant tissue fibrosis usually requires several years of ongoing insult. However, not all patients exposed to a similar causal agent develop the same degree of liver fibrosis, ie, patients with similar risk factors have some variability in progression of liver fibrosis, which also may reflect host genotypic polymorphisms.³³ Evidence of fibrotic regression has now been documented in the entire spectrum of chronic liver diseases, including autoimmune hepatitis, biliary obstruction, iron overload, nonalcoholic steatohepatitis,³⁴ and viral hepatitis B (HBV) and C (HCV).^{35,36}

Patterns of fibrosis progression have been described on the basis of their disease origin. Among the contributors, chronic HCV poses a significant risk for developing fibrotic tissue, but only becomes lethal following cirrhosis onset. Factors affecting the rate of fibrosis progression in HCV-infected patients include duration of infection, age, male gender, consumption of alcohol, human immunodeficiency (HIV) coinfection, and low CD4 count.^{37–41} Metabolic conditions, including obesity, steatosis, and diabetes, are also surfacing as independent risk factors.⁴²

Both male gender and age strongly influence the speed of disease transition from infection to cirrhosis, with a 300-fold higher progression rate seen in men aged 61–70 years versus those 20–40 years of age.⁴¹ In contrast, HCV-infected women progress to cirrhosis at a much slower rate compared with their male counterparts, regardless of age. A multicenter study aimed at evaluating the natural history of liver fibrosis progression in HCV and associated risk factors reported a median 39% higher annual rate of fibrosis progression in men compared with women.⁴³ The significance of this result was partially rationalized by the greater alcohol consumption associated with men in this trial.

Antifibrotic effects from estrogen may also explain the fibrosis progression disparity seen between the genders. Long-term estrogen exposure reduced liver fibrosis progression in HCV-infected women who had either been pregnant, taken oral contraceptives, or received hormone replacement therapy in one retrospective study conducted across two centers in France.⁴⁴ Infected postmenopausal women not receiving hormone replacement therapy showed the highest rate of fibrosis progression.⁴⁴

Some speculative data have emerged demonstrating a correlation between insulin resistance and increased rate of fibrosis progression in nondiabetic individuals infected with chronic HCV, although a longitudinal study is needed to confirm this interesting observation.⁴⁵

Patients coinfecting with HIV and HCV have more rapid progression of cirrhosis that may be partially curbed with highly active antiretroviral therapies.^{46–50} In a recent study, Berenguer et al examined the effects of exposing coinfecting patients to nonnucleoside reverse transcriptase inhibitor and protease inhibitor therapy, and reported a successful outcome in reducing fibrosis progression with nonnucleoside reverse transcriptase inhibitors but not with protease inhibitors.⁴⁶

Other special populations on the radar for developing fibrosis-related complications include those with hemochromatosis or nonalcoholic steatohepatitis. Cirrhotic patients with nonalcoholic steatohepatitis have a reduced life expectancy and a concomitant 67% 5-year survival rate.^{51,52} Evaluation of pooled data from histological studies identified inflammation on the initial biopsy and age as independent predictors of progression of fibrosis to advanced nonalcoholic steatohepatitis.⁵²

Individuals suffering from hemochromatosis, characterized by iron deposition within tissues and related end-organ damage, develop cirrhosis, which may or may not present with symptoms or elevated liver enzymes.^{53–55} Some important risk factors identified for increased rate

of progression of fibrosis in this patient population include alcohol consumption, hepatic steatosis, HCV coinfection, male gender, oxidative injury, and genetic polymorphisms, particularly implicating transforming growth factor- β 1, tumor necrosis factor- β , and MCP1.⁵³

Although there is no doubt that liver tissue fibrosis can regress, the issue of regression/reversibility of cirrhosis is controversial and requires considerable clarification. This is particularly relevant after several claims of “cirrhosis reversal” in patients with chronic HBV and HCV infection after antiviral treatment.^{35,56} The concept of cirrhosis reversibility originates from evidence obtained in animal models upon the discontinuation of the cause of liver damage or following treatment with antifibrotic agents. Although cirrhosis tends to be defined as one stage, it is quite evident, both morphologically and clinically, that there are several stages of evolution following the initial morphological evidence of cirrhotic transformation of liver tissue. “Early” cirrhosis is often macronodular, with a limited thickness of fibrotic septa and a scarcity of neoangiogenesis. In addition, at this stage, the fibrotic tissue is still characterized by the presence of profibrogenic cells and an abundant inflammatory infiltrate. The evolution of this picture is characterized by increased thickness of the septa that become progressively acellular and by the development of micronodular cirrhosis. From the biochemical point of view, collagen fibers undergo extensive crosslinking and are wrapped in filaments of elastin. At this advanced stage, fibrosis becomes largely irreversible. Therefore, claims of “cirrhosis reversal” need to be framed in the stage of evolution of a cirrhotic liver and the term “reversal” used to denote a return to near-normal liver structure.

Establishing cirrhosis reversal in humans has been hampered by a lack of supportive evidence, compounded by the fact that few relevant experiments were carried out using animal models.^{57,58} It should be stressed that in the large majority of the available animal models (mostly in rodents), cirrhosis develops within weeks and is rather different both in morphological and biochemical (ie, composition and three-dimensional arrangement) terms from the cirrhosis observed in chronically ill patients who develop liver disease over several decades.

Mechanisms of fibrosis reversal

The first documented evidence for the reversal of liver fibrosis derived from studies showing a progressive decrease of fibrosis associated with apoptosis of hepatic stellate cells in rat models of chronic intoxication with CCl₄ following

suspension of toxic and bile duct ligation after recanalization of the bile duct.^{59,60} Altogether, these studies suggested that apoptosis of hepatic stellate cells plays a critical role in the recovery from biliary as well as toxic-induced liver fibrosis. In addition, hepatic stellate cell survival and apoptosis are regulated by growth factors expressed during fibrotic liver injury. Issa et al⁵⁸ went further to show that reversal was obtained in a chronic CCl₄ model, even when an advanced stage of micronodular cirrhosis was obtained after 12 weeks of administration of CCl₄. Indeed, 1 year after the cessation of administration of CCl₄, micronodular cirrhosis underwent remodeling to macronodular cirrhosis. In addition, expression of collagen type 1 and tissue inhibitor of metalloproteinases 1 (TIMP-1) mRNA decreased significantly, and active metalloproteinases were shown in liver tissue during fibrosis remodeling. This was paralleled by a significant loss of more recently formed fibrils, decreased perisinusoidal fibrosis, and decreased thickness of fibrotic septa. Once again, resolution was characterized by apoptosis of hepatic stellate cells, predominantly at the edges of fibrotic septa. Residual septa, not remodeled after 1 year, were characterized by transglutaminase-mediated crosslinking and relative hypocellularity. This experimental study highlighted a limited potential for the cirrhotic liver to engage in spontaneous regression of the mature matrix in the absence of a therapeutic intervention.^{57,58}

The relationship between fibrosis reversal and hepatic stellate cell apoptosis highlighted by these studies in animal models suggested that induction of hepatic stellate cell apoptosis could represent a potential antifibrogenic target. This process would necessarily remove the primary hepatic source of newly synthesized collagens as well as TIMP-1.⁶¹ Hepatic TIMP-1 inhibits metalloproteinases implicated in matrix degradation, and both collagen and TIMP-1 promote myofibroblast survival.^{62,63} Reverse differentiation of the myofibroblasts back to their original phenotype occurs in vitro and therefore constitutes an alternative mechanism for fibrosis regression, although as yet there is no in vivo evidence supporting this claim.^{57,64,65} In fact, Kim et al reported decreases in desmin-positive cells in rats recovering from dimethylnitrosamine-induced fibrosis.⁶⁶ This result argues for the apoptotic pathway because desmin marks both activated myofibroblasts and their hepatic stellate cell precursors.⁶⁶

However, the concept of hepatic stellate cell apoptosis in animal models is not likely to be completely reproducible in the human setting. Despite what we have learned about antifibrotic mechanisms from animal studies, the evidence is at best speculative and needs further support particularly with relevance to

clinical evidence. Although apoptosis seems to play a role in fibrosis regression, it may simply represent a symptom of extracellular matrix remodeling. This explanation garners validity from experimental evidence showing that expression levels of both collagen 1 and TIMP-1 ramp up during fibrogenesis, but then decrease in the recovering injured liver.^{57,63,67}

Different studies performed in human hepatic stellate cells and in liver tissue obtained from patients with chronic viral hepatitis have suggested that human profibrogenic cells, and particularly hepatic stellate cells, are characterized by a resistance to apoptosis once activated and fully involved in the fibrogenic process.^{68,69} It is therefore plausible that long-term fibrogenesis is characterized, in addition to biochemical evolution of scar tissue and lack of an appropriate degradation machinery, by the immovability of a critical mass of profibrogenic cells.

An important aspect of macrophage biology in the fibrogenic process is the observation that these cells are required for fibrosis resolution after withdrawal of the damaging agent. Along these lines, selective depletion of macrophages resulted in reduced scarring and a lower number of fibrogenic cells in the active phase of fibrogenesis, as expected based on previous work.⁷⁰ However, macrophage depletion during fibrosis resolution was associated with delayed matrix degradation. These data indicate the existence of distinct subpopulations of macrophages and the role of these cells in the recovery phase of the fibrogenic process. These data have been recently expanded with the observation that scar-associated macrophages that mediate fibrosis resolution express matrix metalloproteinase 13 allowing them to degrade complex extracellular matrices and remodeling of scar.⁷¹ The interaction between the chemokine system and macrophages in the process leading to fibrosis resolution has also been recently investigated. When chronically intoxicated with CCl₄, mice lacking chemokine (C-C motif) receptor 2 (CCR2) showed lower levels of fibrosis compared with wild-type animals, and reduced F4/80+, CD11b+, and CD11c+ populations at the sites of injury.⁷² However, upon discontinuation of toxin administration, fibrosis persisted in CCR2 knockout mice, and was correlated with sustained expression of TIMP-1 and with reduction in matrix metalloproteinase 13 expression. These data suggest that this chemokine system is one of the molecular effectors of both macrophage recruitment during active fibrogenesis and of macrophage-dependent fibrosis resolution. Accordingly, because therapies that interfere with chemokines or their receptors are being developed, it may be speculated that a therapy interfering with CCR2 could possibly retard the regression of fibrosis, especially when associated with a strategy that reduces the extent of damage.

Clinical evidence for reversal

Accordingly, when performing an accurate analysis of results of the clinical studies claiming reversal of cirrhosis after antiviral treatment, the only prudent conclusion is that, in most cases, there was a variable degree of fibrosis regression in cirrhosis but not a reversal of cirrhosis.^{73,74} For example, Poynard et al reported fibrosis reduction among cirrhotic patients with chronic HCV who received pegylated-interferon (IFN) and ribavirin regimens.⁷⁵ Of the 153 patients enrolled in their study, 49% showed downstaging of liver fibrosis from stage 4 to stage 3. A separate study looked retrospectively at liver improvements in 113 individuals with confirmed hepatic cirrhosis and receiving immunosuppressive therapies, including IFN- α , or a combination of IFN- α and ribavirin in the case of viral hepatitis infection, or steroids plus azathioprine in autoimmune patients. Among the 113 participants, 14 (12.4%) had biopsy-proven disappearance of cirrhosis.⁷⁶ One rare report describing spontaneous recovery in two young men who had developed HBV-related cirrhosis as children recently emerged. The investigators attributed their “cure” to cessation of viral replication.⁷⁷

Although fibrosis seems to be a reversible process accompanied by regression of scar tissue, establishing clear remodeling or an apoptotic mechanism underlying the regression process remains an elusive goal. However, the more challenging goal of reversing cirrhosis must take into account features other than simple regression alone. Additional factors, including what effects rapid clearance of the matrix would have on the highly altered vasculature structure of the cirrhotic liver, warrants special consideration.^{13,74,78} The availability of improved diagnostic techniques for monitoring disease progression should also enhance the evaluation of potential antifibrotic therapeutics in the clinic.⁷⁹

In other words, fibrotic deposition related to recent disease and characterized by the presence of thin reticulin fibers, often in the presence of a diffuse inflammatory infiltrate, is likely to be fully reversible, whereas longstanding fibrosis, characterized by extensive collagen crosslinking in a dense acellular/paucicellular extracellular matrix and decreased expression and/or activity of specific metalloproteinases, is not.^{58,80}

Assessment of fibrosis regression

Assessment of the fibrotic evolution of chronic liver disease has relied and still largely relies on histopathological scoring of liver tissue obtained by liver biopsy. However, liver biopsy is an invasive procedure limited by issues relating to sampling, cost, and morbidity, and only provides a static

measure of fibrosis. It was evident that liver biopsy was an imperfect gold standard when it was used to assess the extent of disease progression in terms of fibrotic transformation of liver tissue, and even more so when the fibrotic stage derived from histopathological evaluation was used as a gold standard to define the diagnostic accuracy of noninvasive methods.⁸¹ It is increasingly clear that the diagnostic accuracy of liver biopsy is limited by two major caveats, ie, sampling error and interobserver variation among pathologists, with an average 20% error rate in assessment of fibrosis stage.^{82,83}

Alternative methods to quantify liver tissue, such as computer-aided morphometric image analysis of hepatic collagen,⁸⁴ can provide an objective measurement of the proportion of liver with fibrous tissue. By comparison, routine histological assessment also takes into account other subjective factors, such as architectural distortion or nodule formation, but is clearly dependent upon the experience of the individual pathologist.⁸⁵ The coefficient of variation for image analysis compared with standard histological assessment remains unacceptably high, even for 25 mm biopsies.⁸² In addition, liver biopsy is not practical for monitoring disease progression or response to therapy in chronic HCV infection.⁸⁶ Thus, there is a need for noninvasive tests to assess liver fibrosis. Ideally these tests should be simple, readily available, inexpensive, and accurate. Over past years, several noninvasive tests have become available to assess liver fibrosis. In general terms, these are either “direct markers”, ie, proteins derived from the extracellular matrix assembly and remodeling, or “indirect markers” that are in general represented by algorithms, including biochemical tests that are commonly altered in chronic liver disease.⁸⁷

This decline in the diagnostic relevance of liver biopsy, particularly in chronic HCV, has led to the development of noninvasive methodologies for assessment of liver fibrosis or cirrhosis. There are two critical endpoints, ie, the presence of significant fibrosis, which is an indication for antiviral treatment in chronic HBV and HCV, and the presence of cirrhosis, which is an indication for specific monitoring of complications related to portal hypertension and to the increased risk of developing hepatocellular carcinoma. A summary of the more widely applied diagnostic techniques for evaluating stage of fibrosis are presented in Table 1.

The most commonly used noninvasive methods are measurement of liver stiffness using transient elastography (Fibroscan[®], Echosens, Paris, France)⁸⁸ and serum biomarkers of fibrosis, eg, the patented FibroTest algorithm.³⁶ Various noninvasive markers have been developed to differentiate mild from moderate-to-advanced

stage disease, mainly in HCV, but the most widely used and validated with transient elastography⁸⁸ by far are the aspartate to platelet ratio index (APRI, a free nonpatented index⁸⁹) and the FibroTest.⁹⁰

The APRI index has also been validated as a fibrosis predictor in HIV coinfection and chronic HCV, and also in alcoholic liver disease.^{91,92} A meta-analysis of 22 studies (most in chronic HCV) shows that, at an APRI threshold of 0.5, the sensitivity and specificity for significant fibrosis have been 81% and 50%, respectively, and that, at an APRI threshold of 1.0, the sensitivity and specificity for predicting cirrhosis have been 76% and 71%, respectively.⁹³

Other popular biochemical tests include the Forns' fibrosis index⁹⁴ and Enhanced Liver Fibrosis score.⁹⁵ The diagnostic performance of these indices is generally good, with a receiver-operating characteristic (ROC) curve ranging from 0.77 to 0.88. Comparative studies have noted that the performance characteristics of these biomarker panels are similar.^{96,97}

Imaging techniques are an attractive way of evaluating fibrosis. They have the advantages of being noninvasive and being able to detect structural changes, which serological-based tests of fibrosis and inflammation are unable to do. Using the modalities of ultrasound, computed tomography, or magnetic resonance imaging, it is possible to diagnose features of advanced chronic liver disease by recognizing surrogate markers of portal hypertension with a high degree of sensitivity and specificity. However, these techniques do not reliably detect lesser degrees of fibrosis.

Transient elastography has recently become available, which measures liver stiffness or elasticity to assess liver fibrosis.⁹⁸ The scan was developed on the principle that livers with increasing degrees of scarring or fibrosis have decreasing elasticity and that a shear wave propagating through stiffer material would progress faster than in one with more elastic material.⁹⁹ Although the reproducibility of transient elastography has been shown to be excellent in terms of interobserver and intraobserver agreement,⁴⁰ its applicability may not be as good as that of biomarkers.¹⁰⁰ The principal reasons are obesity, particularly increased waist circumference, and limited operator experience. The risk of overestimating liver stiffness values has been reported in the case of alanine aminotransferase flares in patients with acute viral hepatitis or chronic HBV,^{101–103} as well as in cases of extrahepatic cholestasis¹⁰⁴ and congestive heart failure.¹⁰⁵

Initial studies of noninvasive markers largely consisted of single components, but the field has evolved into combining these single components into panel markers.

Table 1 Common tests for evaluating liver fibrosis^{79,81,99,138–140}

Method	Measures	Advantage/Disadvantages
Biopsy		
METAVIR	Necroinflammation + fibrosis histological score: F0, no fibrosis F1, stellate enlargement of the portal tract without septa F2, stellate enlargement of the portal tract with septa F3, fibrosis without cirrhosis F4, cirrhosis	<ul style="list-style-type: none"> • Necessary for diagnosing autoimmune hepatitis, primary biliary cirrhosis, NASH, and Wilson's disease • Staging and grading for viral hepatitis and hemochromatosis • Invasive; patient may experience pain or bleeding and rarely death • Small sample size, may miss cirrhosis • Expensive
HAI	Summation of periportal/bridging necrosis, intralobular degeneration, portal inflammation and fibrosis (range 0–22)	<ul style="list-style-type: none"> • Invasive; patient may experience pain or bleeding and rarely death • Small sample size, may miss cirrhosis • Expensive
Noninvasive tests (panel)		
Fibrotest	Bilirubin, α 2-MC, gamma GT, haptoglobin, Apolipoprotein A1, age and gender AUC: $t = 0.84$, $v = 0.87$	<ul style="list-style-type: none"> • Separate mild from severe fibrosis with high diagnostic accuracy • Noninvasive
APRI	Platelets, AST AUC: $t = 0.80$, $v = 0.90$	<ul style="list-style-type: none"> • Simplistic and good alternative when biopsy is contraindicated
Forns' fibrosis index	Cholesterol, gamma GT, platelets, and age AUC: $t = 0.86$, $v = 0.81$	<ul style="list-style-type: none"> • Serial measurements possible
Fibrosis probability index	Plasma glucose, insulin, age at biopsy, AST, HOMA-IR, total cholesterol and past alcohol AUC: $t = 0.84$, $v = 0.77$	<ul style="list-style-type: none"> • Limitations in evaluating inflammation (biopsy better)
Transient elastography		
FibroScan®	Liver stiffness range 2.5–75.0 kPa (normal 5.5 kPa) AUC stage F3–F4 > 0.9	<ul style="list-style-type: none"> • Noninvasive tool for assessment of liver fibrosis by measuring liver stiffness • Immediate results • Painless procedure • Excellent performance in diagnosing severe disease

Abbreviations: α 2-MC, α 2-macroglobulin; AST, aspartate aminotransferase; AUC, area under the curve; GT, glutamyl transpeptidase; HAI, histological activity score; NASH, nonalcoholic steatohepatitis; HOMA-IR, homeostasis model assessment of insulin resistance.

The diagnostic performance of panel markers depends on the stage of fibrosis that is distinguished and disease origin. Although the diagnostic performance of the panel markers is broadly similar, there seems to be a trend of superior performance in alcohol-induced, followed by chronic HCV and nonalcoholic fatty liver disease, and then chronic HBV. In nonalcoholic fatty liver disease, a systematic review by Guha et al¹⁰⁶ suggested that the evolution of noninvasive markers lagged behind HCV with fewer panel markers. Variables found to be significant in the detection of fibrosis included the presence of diabetes, age, homeostasis model assessment of insulin resistance, ratio of aspartate to alanine transaminase, decreased platelet count, presence of hyaluronic acid, and increased body mass index. However, it remains difficult to draw any firm conclusions regarding the performance of these markers, given the heterogeneous fibrosis scoring systems and endpoints used in these studies. Transient elastography could be useful as a screening test to exclude advanced fibrosis in patients with nonalcoholic fatty liver disease.

Realistic endpoints for antifibrogenic therapy

The most effective antifibrotic strategy is to cure the underlying disease process before advanced fibrosis has developed. Existing treatments, particularly those that treat the primary cause of tissue injury, can allow extensive resolution.

Advances in our understanding of the pathogenesis of liver fibrosis have identified several potential feasible therapeutic targets, but unfortunately clinical development so far has been disappointing. One major limitation has been the often-prolonged natural history of fibrosis compared with that in experimental models, and difficulties in accurate noninvasive fibrosis assessment, thus making clinical trial design difficult. Furthermore, many patients are asymptomatic even when fibrosis is already advanced, or present with decompensated cirrhosis, which would require not only prevention of evolution in cirrhosis but also reversal of established fibrosis in a cirrhotic liver. Indeed, in broad terms, the two main endpoints of antifibrogenic therapy are reduction of fibrogenesis in the precirrhotic stage, in order to prevent

the establishment of cirrhosis, and reduction of fibrosis in a cirrhotic liver, leading to reduction of portal hypertension and expansion of the complication-free period (ie, compensated cirrhosis). Indeed, while the use of antifibrotic drugs in the precirrhotic stage is debatable and priority should be given to agents acting on the primary cause of disease (ie, antivirals), the single use of this class of drugs could be indicated in patients with cirrhosis, especially when not responding to treatments aimed at eliminating the cause of chronic damage. Several drugs with specific “antifibrotic activity” have been studied in human trials but were not shown to be clearly effective. The ideal antifibrotic agent which is safe when used over a long time, specific to the liver and nontoxic to hepatocytes, potent, orally bioavailable, and inexpensive, is not yet available. It is now well established that elimination of the fibrogenic stimulus can lead to regression of accumulated fibrosis, even in the setting of early cirrhosis. Examples include sustained virological clearance in HCV infection,^{107,108} durable viral suppression in patients with chronic HBV,^{109–111} venesection for hemochromatosis,¹¹² chelation for Wilson’s disease, immunosuppression for autoimmune hepatitis,¹¹³ and weight loss for nonalcoholic steatohepatitis.¹¹⁴ Unfortunately, curative therapy is not possible in many patients either because available disease-specific therapies have failed or because of late presentation

with established cirrhosis. These patients have limited therapeutic options to reduce their risk of developing complications from end-stage liver disease. Fortunately, a number of promising targeted approaches (Table 2) are in development.¹¹⁵ Such therapies have been targeted to any of several different biological targets (eg, inhibition of collagen synthesis, interruption of matrix deposition, stimulation of matrix degradation, modulation of stellate cell activation, induction of stellate cell death, or blocking receptor tyrosine kinases like vascular endothelial growth factor receptor and platelet-derived growth factor receptor).

In patients infected with HBV or HCV, regression of liver fibrosis usually happens naturally upon achieving a sustained viral response from antiviral therapy. Controlling HBV involves giving the patient nucleotide analogs, such as lamivudine, adefovir, and entecavir, while combined pegylated IFN (+ribavirin) therapy provides the best treatment protocol for eradicating HCV.¹¹⁶ Sustained response with this regimen has now reached 60%. IFN may also regulate promoters of collagen gene expression and reduce hepatic stellate cell proliferation with the help of intracellular signaling molecules, like microRNA-29b.^{116–118}

Therapeutics that target TIMP-1 provide a good strategy for antifibrotic development because TIMP-1 is implicated in extracellular collagen matrix turnover and antiapoptotic

Table 2 Potential antifibrotic agents and mechanism of action^{82,128,140}

Compound	Target	Mechanism of action
Endothelin antagonist A receptor antagonist (LU135252)	ET-I	Inhibits HSC activation and fibrogenesis
Angiotensin system inhibitors Captopril, enalapril, perindopril; losartan, irbesartan, telmisartan	RAS	Blocks profibrogenic effects of angiotensin II
PPAR- γ Glitazones (pioglitazone, rosiglitazone, troglitazone)		Unknown
Anti-TGF- β Anti-integrin $\alpha\omega\beta 6$ antagonist (EMD405270), TGF- β antisense oligonucleotides, soluble TGF- β decoy receptors	TGF- β I	Inhibits HSC fibrogenesis, apoptosis
Glucocorticoids MMP-inducers Halofuginone	HSC Extracellular matrix	Induces apoptosis Increase MMP fibrolytic activity
TIMP inhibitor Neutralizing anti-TIMP-I antibody	TIMP-I	Enhanced MMP activity
Sulfasalazine	NF- $\kappa\beta$	Induce HSC apoptosis
SiRNA	HSP47	Induces matrix degradation
Immunosuppressants Mycophenolate mofetil, rapamycin Angiogenesis inhibitors VEGF receptor 1 and 2 antagonists (PTK, sorafenib, vatalanib), anti-integrin $\alpha\upsilon\beta 3$ (EMD409915)		Inhibits HSC activation

Abbreviations: PPAR- γ , peroxisome proliferator-activated receptor-gamma; TGF- β , transforming growth factor beta; HSC, hepatic stellate cell/myofibroblast; MMP, matrix metalloproteinase; TIMP, tissue inhibitor of matrix metalloproteinase; HSP47, heat shock protein 47; VEGF, vascular endothelial growth factor; PTK, protein tyrosine kinase; NF, nuclear factor; ET-I, endothelin-I; RAS, renin-angiotensin system.

activity towards myofibroblasts.^{63,79,119} Of experimental relevance, a recent study showed reduced collagen accumulation in fibrotic liver tissue of rats treated with a TIMP-1-neutralizing antibody.¹²⁰ Because agents targeting TIMP-1 act extracellularly, drug design does not have to overcome the additional burden of crossing membrane barriers in this case.⁷⁹ Other agents that degrade the collagen-rich matrix may also offer alternative and viable antifibrotic treatment. As proof of concept, overexpression of matrix metalloproteinase 8 with adenovirus therapy led to fibrosis regression in two models of experimental liver fibrosis (CCl₄ and bile duct ligation).¹²¹ Adenoviral transfection of matrix metalloproteinase 1 similarly attenuated hepatic fibrosis in rats.¹²²

Perhaps some of the most interesting drug targets stem from inhibition of transforming growth factor- β 1, because this cytokine plays a critical role in the activation of myofibroblasts. Adenoviral constructs involved in transforming growth factor- β 1 signaling and other soluble molecules that modulate the expression or function of transforming growth factor- β 1 have shown antifibrotic efficacy in vitro and in vivo.^{123–130} However, the importance of this cytokine in homeostasis and repair, cautions against the long-term use of transforming growth factor- β 1 inhibitors, making this class of therapeutics less attractive. Indeed, complete chronic abrogation of the effects of transforming growth factor- β 1 on immunomodulation and regulation of epithelial cell apoptosis could result in autoimmunity and the emergence of neoplastic clones.

Angiotensin II accelerates fibrogenesis in response to liver injury. Vasoactive modulators like angiotensin, type 1 receptor antagonists, or angiotensin-converting enzyme inhibitors therefore represent attractive antifibrotic therapies by blocking angiotensin II receptor access, or decreasing its production.^{125,126,131–134} Such agents have effectively attenuated liver fibrosis in rat models and are suitable for long-term treatment because of their known safety profile.

Targeted antifibrotic therapies that block receptors on hepatic stellate cells have emerged with promising results. Target specificity is achieved by modifying drug protein carriers with molecules that recognize growth factors and collagen type IV receptors,¹³⁵ or by coupling apoptotic gliotoxin to mannose-6-phosphate-modified human serum albumin.¹³⁶ Alternatively, Sato et al observed complete resolution of hepatic fibrosis in rats treated with siRNA delivered with high specificity to stellate cells using vitamin A-coupled liposomes.¹³⁷

Conclusion

Our understanding of the mechanism of liver fibrosis has changed dramatically over the last decade. It is no longer viewed as passive or permanent, but as a dynamic process. Many mechanisms and potential therapies continue to be identified and more research is required. The number of potential therapeutic targets has exploded in recent years, and the realization that fibrosis can regress lends new urgency to their investigation. For the time being, greater emphasis must be placed on early identification of patients with potentially treatable chronic liver disease.

Disclosure

The authors report no conflicts of interest in this work.

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