

Figure S1. Photograph of the solutions of GNCs and resulting FRNPs, respectively.



Figure S2. TEM image of FRNPs (scale bar = 200 nm).



Figure S3. Photographs of aqueous solution of FRNPs at 0 h, 24 h, and 48 h.



Figure S4. Photograph of FRNPs dispersed in water, saline, serum and cell culture medium.



Figure S5. XPS spectrum of Mn 2p (A) and Au 4f (B) for FRNPs.



Figure S6. Lattice of Au in the cage structure.

Time (h)	Hydrodynamic size (nm)	
0	$145.37 \pm 4.47$	
24	$146.23 \pm 3.62$	
48	$148.14 \pm 5.76$	

Figure S7. Dynamic light scattering (DLS) analysis of FRNPs at different time point.



**Figure S8.** Flow cytometric analysis of the mean fluorescence intensity for ECA 109 cells incubated with PBS or Cy7-FRNPs (100  $\mu$ g/ mL) for 24 h.



**Figure S9.** FRNPs in combined NIR photothermal and X-ray radiation exposure decreased ECA 109 cell survival rate detected by colony formation assay. Data represented were mean  $\pm$  SD of three identical experiments made in three replicates. Error bars were based on standard

deviation of three parallel samples. P-values were calculated by the student's test: \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001, \*\*\*\*p < 0.0001 compared to the control group.



Figure S10. Quantitative analysis of cell apoptosis assay. Data are shown as mean ± SD.



**Figure S11.** The fluorescence intensity of tumor area at different time intervals after injection of Cy7 -FRNPs.

T1 contrast agent	Commercial name	r <sub>1</sub> (mM <sup>-1</sup> s <sup>-1</sup> )	Ref
Gd-DTPA	Magnevist	4.32	[1]
Gd-DTPA-BMEA	OptiMARK	4.10	[2]
Gd-DTPA-BMA	Omniscan	3.80	[3]
Gd-DOTA	Dotarem	3.50	[4]
Gd-DO3A-Butriol	Gadavist	3.70	[5]
Gd-DO3A-HP	ProHance	3.60	[6]
Gd-BOPTA	Multihance	5.20	[7]
Gd-diphenyl-cyclohexy-	Vasovist	4.56	[8]
Fe3O4@SiO2(Gd-DTPA)-		4.20	[9]
ES-MIONs		2.40	[10]
FRNPs		4.76	

**Tabel S1.** Summary of longitudinal relaxation rate of T1 contrast agent used in clinic or biomedicine.



**Figure S12.** PA intensity at the tumor site before and after i.v. injection with FRNPs. P-values were calculated by the student's test: **\*\*\*\***p < 0.0001.

References

- Vecchione D, Aiello M, Cavaliere C, Nicolai E, Netti PA, Torino E. Hybrid core shell nanoparticles entrapping Gd-DTPA and 18F-FDG for simultaneous PET/MRI acquisitions. Nanomedicine. 2017;12(18):2223-2231.
- Periasamy M, White D, deLearie L, et al. The synthesis and screening of nonionic gadolinium (III) DTPA-bisamide complexes as magnetic resonance imaging contrast agents. Invest Radiol. 1991;26: S217-S220.
- Yim H, Yang S-G, Jeon YS, et al. The performance of gadolinium diethylene triamine pentaacetate-pullulan hepatocyte-specific T1 contrast agent for MRI. Biomaterials. 2011;32(22):5187-5194.
- 4. Silvio A, Lucio AP, Mauro B, et al. Synthesis, Characterization, and 1 /Tx NMRD Profiles of Gadolinium(III) Complexes of Monoamide Derivatives of DOTA-like Ligands.

X-ray Structure of the 10-[2-[[2-Hydroxy-I-(hydroxymethyl)ethyl]amino]-I-[(phenylmethoxy)methyl]-2-oxoethyl]-I,4,7,10-tetraazacyclododecane-I,4,7-triacetic Acid-Gadolinium(III) Complex. Inorganic Chemistry. 1992;31(12):2422-2428.

- Turowski PN, Rodgers SJ, Scarrow RC, Raymond KN. Ferric Ion Sequestering Agents.
  18.1 Two Dihydroxamic Acid Derivatives of EDTA and DTPA. Inorganic Chemistry.
  1988;27(3):474-481.
- Dischino DD, Delaney EJD, Emswiler JEE, et al. Synthesis of Nonionic Gadolinium Chelates Useful as Contrast Agents for Magnetic Resonance Imaging.I,4\*7-Tris(carboxymethyl)-10-substituted-I,4,7,10-tetraazacyclododecanes and Their Corresponding Gadolinium Chelates. Inorg Chem. 1991;30:1265-1269.
- Peter C, Ellison JJ, McMurry TJ, Lauffer RB. Gadolinium (III) Chelates as MRI Contrast Agents: Structure, Dynamics, and Applications. Chem Rev 1999;99:2293–2352.
- Blockley NP, Jiang L, Gardener AG, Ludman CN, Francis ST, Gowland PA. Field strength dependence of R1 and R relaxivities of human whole blood to prohance, vasovist, and deoxyhemoglobin. Magnetic Resonance in Medicine. 2008;60(6):1313-1320.
- Yang H, Zhuang Y, Sun Y, et al. Targeted dual-contrast T1- and T2-weighted magnetic resonance imaging of tumors using multifunctional gadolinium-labeled superparamagnetic iron oxide nanoparticles. Biomaterials. 2011;32(20):4584-4593.
- Shen Z, Chen T, Ma X, et al. Multifunctional Theranostic Nanoparticles Based on Exceedingly Small Magnetic Iron Oxide Nanoparticles for T1-Weighted Magnetic Resonance Imaging and Chemotherapy. ACS Nano. 2017;11(11):10992-11004.