Preservation of Small Extracellular Vesicle in Gelatin Methacryloyl Hydrogel through Reduced Particles Aggregation for Therapeutic Applications

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Figure S1: The pore size distribution of different concentrations GelMA hydrogels encapsulated with or without sEV.



Figure S2: General view of different concentrations of Gelma hydrogel encapsulating 1×10^9 particles μl^{-1} sEV in different crosslinking condition. Arrows: cross-linked gel.



Figure S3: The tracks of sEV particles in PBS and in 10% GelMA hydrogels in the different time period at 4°C, Scale bar=40 μ m.



Figure S4. The bioluminescence of different amount of DiO labeled sEV. (Correlation coefficient R = 0.9939), N=3.



Figure S5.The effect of preserved sEV on r-ASCs proliferation and migration. (A) r-ASCs proliferation curve, n=3 for each group. (B) Representative microscope images and number of

migrated r-ASCs in each group. Scale bar= 200μ m. n=3 for each group. (C) Representative microscope images of scratch assay in each group. Scale bar= 100μ m. Time transition of the percentage of cell-free zone against initial scratch area after 12 hr, N=3 for each group. The significance (Figure 4A, B, C, D, E) was tested with one-way ANOVA with Tukey posthoc test. (*P<0.05, **P<0.01, ***P<0.001)

Supplementary Video 1: Irregular Brownian motion of sEV in PBS. Supplementary Video 2: The movement of sEV encapsulated in hydrogels. Supplementary Video 3: The comparison of single particle moving range and track in 10% GelMA or in PBS. Left: sEV in 10% GelMA. Right: sEV in PBS.

Table S1. Oligonucleotide primer sequences.

Target cDNA	Primer sequence (5'-3')
Human-CD31	TCGTGGTCAACATAACAGAACT
	TTGAGTCTGTGACACAATCGTA
Human-VEGF	AGGGAAGAGGAGGAGATGAG
	GCTGGGTTTGTCGGTGTT
Human-FGF2	CATCAAGCTACAACTTCAAGCA
	CCGTAACACATTTAGAAGCCAG
Human-ANGIOGENIN	ACCCTCACAGAGAAAACCTAAG
	GACGACGGAAAATTGACTGATC
Human-GAPDH	CTTTGGTATCGTGGAAGGACTC
	GTAGAGGCAGGGATGATGTTCT