# Dopamine-mediated vanillin multicomponent derivative synthesis via Grindstone method: Application of antioxidant, anti-tyrosinase, and cytotoxic activities

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<sup>1</sup>H NMR of the compound 1b



<sup>13</sup>C NMR of the compound 1b







<sup>13</sup>C NMR of the compound 1c



<sup>1</sup>H NMR of the compound 1d



<sup>13</sup>C NMR of the compound 1d



<sup>1</sup>H NMR of the compound 1e



<sup>13</sup>C NMR of the compound 1e







<sup>13</sup>C NMR of the compound 1f









<sup>13</sup>C NMR of the compound 1h



<sup>13</sup>C NMR of the compound 1i



<sup>1</sup>H NMR of the compound 1j



<sup>13</sup>C NMR of the compound 1j







<sup>13</sup>C NMR of the compound 1k





#### **Experimental for Biological activity**

## Antioxidant activity

### **DPPH method**

DPPH (2,2-diphenyl-1-picrylhydrazyl) antioxidant activity was screened for all compounds (**1a-l**). DPPH antioxidant activity was screened for various concentration (10, 25, 50, and  $100\mu$ g/mL) of tested compounds. Accordingly, extracts were examined for their ability to prevent the bleaching of the purple colored methanol solution of 1,1-diphenyl-1-picrylhydrazyl(DPPH). Aliquots (1.6 mL) teat sample were added to 1.6mL of 0.004% (w/v) methanol solution of DPPH at room temperature, absorbance was read against a blank at 517nm.

Percentage inhibition was calculated by.

DPPH Scavenging effect (%) =  $[(A_{control} - A_{sample}) / A_{control}] \times 100$ 

## H<sub>2</sub>O<sub>2</sub> scavenging activity

Hydrogen peroxide scavenging activity was screened for all compounds (**1a-1l**). A solution of  $H_2O_2(40 \text{ mM})$  was prepared in phosphate buffer (pH 7.4). Test compound (3.4 mL) (10, 25, 50, and  $100\mu$ g/mL) with phosphate buffer were added to  $H_2O_2$ solution (0.6 mL, 40 mM). Absorbance of reaction mixtures was recorded at 230 nm. Percentage inhibition of  $H_2O_2$  scavenging was calculated by.

% Inhibition =  $[(A_{control} - A_{sample} / A_{control}) \times 100]$ 

## NO scavenging activity

The compounds (**1a-1l**) was screened for Nitric oxide scavenging activity. Sodium nitroprusside (1 mL, 10 mM) and 1.5mL of phosphate buffer saline (0.2M, pH 7.4) were added to different concentrations (10, 25, 50, and  $100\mu$ g/mL) of test sample incubated for 150 min at 25°C. Aliquots (1 mL) of reaction mixtures were then treated with 1 mL of Griess reagent (1% sulfanilamide, 2% H<sub>3</sub>PO<sub>4</sub> and 0.1% naphthylethylene diamine dihydrochloride).

Percentage inhibition of H<sub>2</sub>O<sub>2</sub> scavenging was calculated by.

% NO Scavenging =  $[(A_{control} - A_{sample} / A_{control}) \times 100]$ 

Where A<sub>control</sub> is the absorbance of the control reaction, and A<sub>sample</sub> is the absorbance of the test compounds.

#### Antioxidant – (ABTS<sup>++</sup>) scavenging activity

The compounds (1a-11) were screened for ABTS<sup>++</sup> scavenging activity. The ABTS<sup>++</sup> radical cation was produced by the reaction of 7 mM aqueous ABTS with 2.45 mM potassium persulfate in the dark at room temperature for 12 h. Prior to the experiment, the ABTS<sup>++</sup> solution was diluted with phosphate buffer (0.1 M, pH 7.4) to an absorbance of  $0.700 \pm 0.025$  at 734 nm. Subsequently, 1 mL of the diluted solution was added to 1.5-mL aliquots of ethanolic solutions of (**1a-1l**) with different concentrations (10, 25, 50,100 µg/mL). After 30 min, the percentage of inhibition at 734 nm was calculated for each concentration relative to a blank sample (ethanol).

The capability of scavenging the ABTS<sup>++</sup> radical was calculated using the following equation:

ABTS<sup>++</sup> scavenging effect (%) =  $[(A_c - A_s)/A_c] \times 100\%$ 

where  $A_c$  is the initial absorbance of ABTS<sup>++</sup>, and  $A_s$  is the absorbance in presence of **2a–f** and **4a–f**.

## Inhibition of AAPH assays free radical analysis

All synthesised compounds (**1a-11**) were screened for linoleic acid peroxidation. The production of the conjugated diene hydroperoxide by oxidation of linoleic acid in an aqueous dispersion was monitored at 234 nm, with 2,2'-azobis(2-amidinopropane) dihydrochloride (AAPH) used as a free radical initiator. This assay can be used to follow oxidative changes and determine the inhibition of linoleic acid peroxidation induced by each compound tested. A 1-mL aliquot of a 16 mM linoleic acid dispersion was added to a UV cuvette containing 1 mL of 0.05 M phosphate buffer, pH 7.4, thermostatted at 37 °C. The oxidation reaction was initiated at 37 °C in air by adding 1 mL of the 40 mM AAPH solution. Oxidation was carried out in the presence of 1 mL of the tested compounds (10, 25, 50, and 100 $\mu$ g/mL) (**1a-11**). In the absence of antioxidants, the lipid oxidation was monitored in presence of the same amount of DMSO. The rate of oxidation at 37 °C was monitored by recording the absorption increase at 234 nm caused by the formation of a conjugated diene hydroperoxide, with the corresponding data provided in Table 3.

The percentage of linoleic acid oxidation inhibition was calculated as follows:

% Inhibition =  $[(1 - \text{rate of absorbance change with test compound})/\text{rate of absorbance change with solvent control}] \times 100\%$ 

#### Anti-tyrosinase activity

All compounds (1a-1j) were screened for anti-tyrosinase screening. The Mushroom tyrosinase (powder,  $\geq 1000$  unit/mg solid, EC 1.14.18.1) inhibitory activities were measured spectrophotmetrically.

Tyrosinase inhibitory activities were measured spectrophotometrically according to a modified method using Mushroom tyrosinase (EC 1.14.18.1, lyophilized powder, \_1000 unit/mg solid, Sigma Chemical Co.) and L-DOPA as substrate. Briefly, a reaction mixture (3.0 mL of final volume) containing 1.5 mM L-DOPA, 0.1 mM sodium phosphate buffer (pH 6.5), 12.428 U of mushroom tyrosinase, and the test sample was incubated at 30 °C for 2 min. The formation of dopachrome was determined spectrophotometrically by monitoring absorption at 475 nm using a lambda 850 spectrophotometer. Kojic acid was used as a positive control. Percentage tyrosinase inhibition was calculated using the

following equation:

Tyrosinase inhibitory activity (%) =  $[(A-B)-(C-D)]/(A-B) \times 100$ 

Where A is the absorbance of the blank solution after incubation, B is the absorbance of the blank solution before incubation, C is the absorbance of a sample solution after incubation, and D is the absorbance of the sample solution before incubation.

## Cell lines and cell culture

The cell lines, MCF-7 and normal cell lines were obtained from the American Type Cell Collection (ATCC; Manassas, VA, USA). The cells were cultured at 37°C and 5% CO<sub>2</sub> environment to get 70%–80% confluence in Dulbecco's Modified Eagle's Medium (DMEM; Gibco $\mathbb{R}$ , Thermo Fisher Scientific, Waltham, MA, USA) with 10% fetal bovine serum (FBS) (Gibco $\mathbb{R}$ ).

# Cytotoxic screening

The newly synthesized compounds (1a-11) were screened for cytotoxic activity according to a previously described procedure. MCF-7 and Vero cell lines were treated with these compounds at one primary cytotoxic assay dose of  $100\mu$ M for 48 h (MTT anticancer assay). Doxorubicin was used as a standard.

The screening results were expressed in terms of the growth inhibitor concentration (GI<sub>50</sub>), total growth of inhibition (TGI), and lethal concentration (LC<sub>50</sub>).

In the current protocol, all cell lines were pre-incubated on a microtiter plate. The results of each test were reported as the growth percentage of treated cells compared to untreated control cells.

Compounds reducing the growth of cell lines to approximately 32% or less were described as having cytotoxic activity. The cells lines were seeded into 96-well plates at a density of 5000 cells per well, A 0.1mL aliquot of the cell suspension and 0.1 mL of the test solution (6.25–100  $\mu$ g in 1% DMSO, with the final DMSO concentration in media less than 1%) were added to the wells, with the plates kept in an incubator (5% CO<sub>2</sub>) at 37 °C for 72 h. The blank sample contained only the cell suspension, and the control wells contained 1% DMSO and the cell suspension. After 72 h, 20  $\mu$ L of MTT was added, and the plates were kept in the CO<sub>2</sub> incubator for 2 h, followed by the addition of propanol (100  $\mu$ L). The plates were covered with aluminum foil to protect them from light and subsequently agitated in a rotary shaker for 10–20 min. Afterwards, the 96-well plates were processed on an ELISA reader to obtain absorption data at 562 nm.

The experiment was performed in triplicate. Percent proliferation inhibition was calculated using the following formula:

Viability cells inhibition (%) =100-[(At-Ab)/(Ac-Ab)]x 100%

At = absorption of test compound, Ab = absorption of blank, Ac = absorption of control.

The concentration of the synthesized compounds required to inhibit 50% of the growth of the cell

lines (IC50 values) was calculated by analyzing the relationship between concentrations and percent

(%) inhibitions using GraphPad Prism 7 version 7.00 forWindows, GraphPad Software, La Jolla, CA,

## USA, www.graphpad.com.

The NCI renamed the IC50 value, the concentration that causes 50% growth inhibition, the GI<sub>50</sub> value to emphasize the correction for the cell count at time zero; thus, GI50 is the concentration of test drug where  $100 \times (T - T0)/(C - T0) = 50 (1, 2)$ . The optical density of the test well after a 48-h period of exposure to test drug is T, the optical density at time zero is T0, and the control optical density is C. The "50" is called the GI50PRCNT, a T/C-like parameter that can have values from +100 to -100. The GI50 measures the growth inhibitory power of the test agent.

The TGI is the concentration of test drug where  $100 \times (T - T0)/(C - T0) = 0$ . Thus, the TGI signifies a cytostatic effect. The LC<sub>50</sub>, which signifies a cytotoxic effect, is the concentration of drug where 100  $\times (T - T0)/T0 = -50$