Supplementary data



Specimen ID: RS19058620FFP

BurningRock OncoScreen Plus™ Report

Report No: 32034501903190

Report Date: 2019/11/19 PM

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	k Dx	Specimen ID RS19058620FFP	Report Date 2019/11/19 PM	TEST OncoScreen	Plus™
PATIENT IN	IFORMATION				
Name	吴成云	Specimen ID	RS19058620FFP	Accession #	A00144591
Gender	Male	Specimen Type	FFPE slides	Accession Date	2019/11/11
Age	67	Biopsy Type	Puncture	Specimen Received	2019/11/13 AM
ID	3209211952***	*11 Specimen Site	lung	Report Date	2019/11/19 PM
Tumor Type*	Other	Hospital	江苏省人民医院		
Additional Information *	not provided				

*Note : Tumor Type and Additional Information are not from the test, but provided by the patient.

ABOUT THE TEST

BurningRock OncoScreen Plus[™] is a NGS based assay that identifies genomic alterations within 520 cancerrelated genes.

SUMMARY OF RESULTS

CATEGORIES	RESULTS		
Somatic Variants	12 variants in total, 3 of which with clinical significance		
Significant Results	MAP2K1: p.I103_K104del	PIK3CA: p.H1047L	
	TP53: p.G187S		
ТМВ	11.1 muts/Mbp		
MSI	Microsatellite Stable (MSS)		
Germline Variants	None		
Quality Control	Qualified		

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1. Somatic Variants	1. Somatic Variants and Interpretation				
Tier I: Variants with S	strong Clinical Signi	ficance			
AllelicRelevant TherapiesVariantFraction(Responsiveness , Evidence)					
No Tier I mutation detected					

Tier II: Variants of Potential Clinical Significance

Variant	Allelic Fraction	Interpretation	Relevant Therapies (Responsiveness , Evidence)
MAP2K1 Exon 3 p.I103_K104del disruptive_inframe_deletion c.306_311del p.Ile103_Lys104del	17.03%	MAP2K1 has the deletion between position 103 and position 104.The variant lies within protein kinase domain(UniProt.org: Q02750).	Trametinib(Resistance, Level D) RO5126766(Resistance, Level D)
PIK3CA Exon 21 p.H1047L missense_variant c.3140A>T p.His1047Leu	7.38%	PIK3CA has the substitution of Histidine with Leucine at position 1047.The variant lies within PI3K/PI4K domain(UniProt.org: P42336). The variant causes Gain- of-function[PMID: 20593314, 26627007, 22120714].	Alpelisib(Response, Level C)
TP53 Exon 5 splice_region_variant c.559G>A p.Gly187Ser	14.5%	TP53 has the substitution of the G nucleotide at c.559 by a A. The variant is located at the boundary of an exon, which may affect alternative splicing by prediction.	Adavosertib+Olaparib (Response, Level C)

Tier III: Variants of Unknown Clinical Significance

Gene	Variant Type	Exon	Nucleotide Change	Amino Acid Change	Allelic Fraction
ARID2	splice_acceptor_variant	3	c.187-2A>G	-	15.72%
ARID2	frameshift_variant	15	c.4709_4710del	p.Ile1570fs	6.87%
DICER1	missense_variant	22	c.4061G>T	p.Cys1354Phe	6.23%
HGF	missense_variant	5	c.610C>T	p.Pro204Ser	16.32%
MCL1	missense_variant	2	c.734C>T	p.Ser245Leu	6.73%
NOTCH3	missense_variant	3	c.304G>T	p.Ala102Ser	12.56%

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PIK3CG	stop_gained	2	c.1229G>A	p.Trp410*	16.28%
TET2	frameshift_variant	6	c.3733_3737del	p.Tyr1245fs	2.05%
TSC1	disruptive_inframe_deleti on	23	c.3459_3461del	p.Ile1153del	6.56%

Note : 1. The responsiveness to the relevant therapies corresponding to the genetic variation is derived from OncoDB, which is an internal database of Burning Rock Dx, with reference to public data such as NCCN guidelines. This data is for reference purpose by clinicians only. With the continuous improvement of the database and the update of clinical data, the grade of variants may change.

2. The evidence levels are grouped into four levels according to the AMP / ASCO / CAP guidelines [PMID: 27993330]: level A (FDA-approved therapies or have been included in professional guidelines), level B (based on well-powered studies with consensus from experts), level C (level A evidence in a different tumor type, or serve as inclusion criteria for clinical trials, or supported by multiple small studies), level D (preclinical studies , or supported by case reports). The variants are categorized into four categories based on their clinical impact: tier I, variants with strong clinical significance (level A and B evidence); tier II, variants with potential clinical significance (level C or D evidence); tier III, variants with unknown clinical significance; and tier IV, variants that are benign or likely benign.

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2. Germlir	ne Variants and	Interpretation				
Gene	Variant Type	e Exon	Nucleotide Change	Amino Aci Change	d Zygosity	Interpretati on
	No pathogenic	or likely patho	genic germline	variant deteo	cted in this sample	

Note : 1. Only pathogenic or likely pathogenic variants of hereditary cancer related genes will be reported.

2.Hereditary cancer related genes are defined by ACMG secondary finding guideline v2.0 [PMID: 27854360] and NCCN guidelines.This product tests 62 genes: APC、ATM、AXIN2、BARD1、BMPR1A、BRCA1、BRCA2、BRIP1、CDH1、CDK12、CDK4、CDKN2A、CHEK1、CHEK2、EGFR、EPCAM、FANCA、FANCI、FANCL、FH、FLCN、GREM1、HOXB13、KIT、MEN1、MET、MLH1、MSH2、MSH3、MSH6、MUTYH、NBN、NF1、NF2、NTHL1、PALB2、PDGFRA、PMS2、POLD1、POLE、PPP2R2A、PRKAR1A、PTCH1、PTEN、RAD51B、RAD51C、RAD51D、RAD54L、RB1、RET、SDHA、SDHAF2、SDHB、SDHC、SDHD、SMAD4、STK11、TP53、TSC1、TSC2、VHL、WT1.

3. According to ACMG guidelines[PMID:25741868], the pathogenicity of germline variant is classified into five categories : 1-benign; 2-likely benign; 3-variants with uncertain significance; 4-likely pathogenic; 5-pathogenic. The interpretation of variant is based on available literature and relevant databases and may change as research progresses.

	Specimen ID	Report Date	TEST
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3. Summary of Immun	otherapy Biomarkers		
ТМВ			
• RESULT	11.1 muts/Mbp		
• TMB Introduction	Tumor mutation burd that occur per million load of the genome l mutations) within reg	den (TMB) represen n bases in the geno based on non-silent jions of the PANEL c	ts the number of somatic mutations me. This test evaluates the mutation mutations (excluding tumor hotspot aptured.
• Clinical Significance	Previous retrospective studies have shown that a higher mutational burden in the tumor genome is correlated with a better efficacy of anti-PD-1/PD-L1 (± anti-CTLA-4) treatments. The TMB obtained from large NGS panels is highly correlated with that from whole exome sequencing (WES), confirming that selective sequencing of the specific genomic region is sufficient to obtain full understanding of a patient's TMB and can help predict the efficacy of immune checkpoint inhibitors. The CheckMate-227 study further confirmed that EGFR/ALK-negative advanced NSCLC patients with TMB≥10 have significant benefits in both objective response rate (ORR) and progression-free survival (PFS) when they were treated with first-line immune checkpoint inhibitor (Navurizumab plus Ipilimumab) regardless of their PD-L1 expression level compared to standard chemotherapy [PMID: 29658845].		

MSI	
• RESULT	Microsatellite Stable (MSS)
MSI Introduction	Microsatellite instability (MSI) refers to insertions and deletions of simple repetitive sequences in a microsatellite locus, resulting in genomic instability. Microsatellite instability (MSI) status is evaluated by the MSI detection algorithm independently developed by Burning Rock Dx.
Clinical Significance	No immunotherapy drugs are recommended for solid tumors with microsatellite stable (MSS).

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4. SNPs in Drug Metabolism Enzymes

SNP	Genotype	Zygosity	Clinical Significance
CYP2D6*10 rs1065852	G/A	Wild Type	Cytochrome P450 2D6 (CYP2D6) is involved in the metabolism of multiple drugs. Genetic polymorphisms cause substantial variation in CYP2D6 activity and serve as biomarkers guiding drug therapy. The *10 heterozygous variant alleles have partial activity and the exact effect on enzymatic activity is difficult to determine.
DPYD*13 rs55886062	A/A	Wild Type	DPD deficiency is associated with increased toxicity of fluorouracil (5-FU, capecitabine, or tegafur). Individuals with wild type DPD have normal DPD expression.
DPYD*2846A>T rs67376798	T/T	Wild Type	DPD deficiency is associated with increased toxicity of fluorouracil (5-FU, capecitabine, or tegafur). Individuals with wild type DPD have normal DPD expression.
DPYD*2A rs3918290	C/C	Wild Type	DPD deficiency is associated with increased toxicity of fluorouracil (5-FU, capecitabine, or tegafur). Individuals with wild type DPD have normal DPD expression.
UGT1A1*28 rs8175347	(TA) ₆ /(TA) ₇	Wild Type	UGT1A1-specific polymorphisms are associated with increased irinotecan toxicity. The precise dose reduction in heterozygous carriers of the variants UGT1A1*28 is not known and subsequent dose modifications should be considered based on the tolerance of individual patient.
UGT1A1*6 rs4148323	G/G	Wild Type	UGT1A1-specific polymorphisms are associated with increased irinotecan toxicity. It is less likely for individuals with wild-type UGT1A1 to have irinotecan-related toxicity.





Note : The figure above shows the copy number distribution for all detected genes. Each dot represents an interval from captured gene regions, and the highlighted dots represent those genes focused on copy number alterations. The horizontal axis represents the chromosome where a gene is located, and the vertical axis represents the copy number calculated based on the NGS data (the red horizontal line represents the normal copy number of a gene. Copy number

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obtained by this test cannot completely represent the gene copy number in tumor cells due to the dilution of normal cells.

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APPENDIX1 : QUALITY CONTROL

	Quality Parameters	Value	Criteria
Pathology	Tumor Purity (%) ¹	-	≥10%
	DNA Quantity(ng) ²	838	≥30
DNA Quality	DNA Degradation ³	А	A-B-C
	Pre-library Quantity (ng) ⁴	4490	≥300
	Mean Depth ⁵	1249	≥500
	Library Diversity ⁶	78%	≥20%
	Insert Length (bp) ⁷	242	≥150
Data Quality	Coverage Uniformity ⁸	97%	≥90%
	Mapping Ratio ⁹	100%	≥95%
	Q30 Percentage ¹⁰	92%	≥80%
	SNP Concordance ¹¹	99%	≥90%
Overall QC ¹²	Qu	ualified	

Note : 1. Tumor Purity: Tumor purity was assessed by Burning Rock using HE staining. This step will be omitted if the sample does not meet the requirement for tumor content assessment. This assessment is not performed on cfDNA samples.

2. DNA Amount: the amount of DNA extracted from the submitted specimen.

3. DNA Degradation: the level of DNA degradation assessed by gel electrophoresis; A-D indicate different levels of DNA degradation, A indicates the lowest degradation and D indicates the highest degradation. This step will be omitted if a sample does not need such assessment. This assessment is not performed on cfDNA samples.

4. Pre-library Amount: the amount of libraries for sequencing after adapter-ligated DNA fragments are amplified and purified.

5. Mean Sequencing Depth: the mean and median number of unique reads being mapped to a given nucleotide of a targeted gene

6. Library Diversity: the proportion of DNA libraries from the original DNA fragments.

7. Insert Length: the median length of the sequenced DNA fragments. For tissue samples, if the median insert length is less than 170bp, it indicates DNA degradation. This may introduce DNA degradation induced false positives. The insert length for cfDNA is 170bp.

8. Coverage Uniformity: the percentage of targeted base positions in which the read depth is greater than 0.2 times the mean coverage depth.

9. Mapping Ratio: the percentage of the number of sequences successfully aligned to the reference genome.

10. Q30 Percentage: the percentage of bases that have a Q-score above or equal to 30(below a probability of incorrect base calling of 1 in 1000).

11. SNP Concordance: the fraction of the genotype calls for SNPs from paired specimen.

12. Overall QC: the overall quality control based on multiple parameters; overall QC is divided into three levels: qualified, warning and fail. When the overall quality is warning or fail, the accuracy and sensitivity of this test may be affected.



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APPENDIX2 : METHODOLOGY AND LIMITATIONS

METHODOLOGY

This test performed hybrid capture-based next-generation sequencing (NGS) to detect genomic alterations. The detection method was independently developed, analyzed and validated by Burning Rock.Dx. Burning Rock has established CLIA-certified and CAP-accredited NGS laboratory, and passed the relevant quality assessment of National Center for Clinical Laboratories of China.

This test covers up to 20bp on either side of the intron/exon boundaries of the targeted genes. Variant types that this test can detect single nucleotide variation (SNV),small insertion or deletion, and some rearrangements.

LIMITATIONS

1. This report only provides reference for clinical diagnosis and treatment decisions-making. Clinical diagnosis and treatment decisions should be made by the clinician in combination with the clinical information of patient.

2. The analysis and interpretation of results are based on available literature and databases. With the advancements of relevant research and updates of databases, the interpretation of mutations may change.

3. This test is only applicable to the detection of DNA mutations. DNA methylation, RNA-level or protienlevel modifications are not inferred by this test.

4. If no muation detected (negative reult), we cannot rule out the possibility of the presence of mutations which are below the limit of detection.

5. Due to the complexity of the tumorigenic mechanism, the results of this test alone cannot confirm or exclude the presence of malignant tumors.

6. Acquired mutations may occur during treatment and tumor development, resulting in the change of mutation spectrum. Tumors also have spataial and temporal heterogeneitytest. Therefore, results from different specimens of the same patient may differ. This test is only responsible for the specimen submitted.

7. This test cannot determine chromosomal polyploidy which may result in copy number variations (CNV). The sensitivity of CNV detection is affected by the proportion of tumor cells in a specimen. If the proportion of tumor cells is <20%, the detection sensitivity is limited.

8. The sensitivity of TMB detection is affected by the proportion of tumor cells in a specimen which is correlated with the maximum allelic fraction (maxAF). If the proportion of tumor cells is <10% or maxAF <5%, the detection sensitivity is limited.

9. The sensitivity of MSI detection is affected by the proportion of tumor cells in a specimen which is correlated with the maximum allelic fraction (maxAF). If the proportion of tumor cells is <20% or maxAF <10%, the detection sensitivity is limited.



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APPENDIX3 : GENE LIST

ABL1 NM_005157.5 AKT1 NM_001014432.1 AMER1 NM_152424.3 **ARID1A** NM 006015.4 ASXL2 NM_018263.4 AURKB NM 001284526.1 BAP1 NM_004656.3 BCL2L1 NM 001317919.1 BCORL1 NM 021946.4 BRCA1 NM 007294.3 BRIP1 NM_032043.2 CARD11 NM_032415.5 CCND2 NM_001759.3 CD79A NM_001783.3 CDK12 NM 016507.3 CDKN1B NM_004064.4 **CEBPA** NM_004364.4 CHEK1 NM_001274.5 CRLF2 NM_022148.3 CTCF NM_006565.3 CUL4A NM_001008895.2 **DAXX** NM_001141970.1 DIS3 NM 0149534 DOT1L NM_032482.2 EIF4E NM 001130679.1 EPHA3 NM_005233.5 ERBB2 NM 004448.3 ERCC3 NM 000122.1 ESR1 NM 000125.3 EZH2 NM 004456.4 FANCE NM_021922.2 FANCM NM_020937.3 FGF12 NM_021032.4 FGF4 NM 002007.2 FGFR3 NM_000142.4 FLT3 NM_004119.2 FOXP1 NM_001244810.1 GATA1 NM_002049.3 **GEN1** NM_001130009.2 **GNAO** NM 002072.4 GRM3 NM_000840.2 HDAC1 NM 004964.2 HIST1H3A NM_003529.2

HIST1H3G NM_003534.2

HIST3H3 NM_003493.2

HOXB13 NM_006361.5

ID3 NM_002167.4 IGF1R NM_000875.4 IL7R NM_002185.3 INSR NM_000208.3 JAK1 NM_001320923.1 KDM5A NM_001042603.2 KEL NM_000420.2 KMT2C NM_170606.2 ABL2 NM_007314.3 AKT2 NM_001626.5 **APC** NM_000038.5 **ARID1B** NM 020732.3 **ATM** NM_000051.3 **AXIN1** NM 003502.3 BARD1 NM_000465.3 BCL2L11 NM_001204107.1 BIRC3 NM 001165.4 BRCA2 NM 000059.3 BTG1 NM_001731.2 CASP8 NM_001228.4 CCND3 NM_001760.4 CD79B NM_000626.3 CDK4 NM 000075.3 CDKN1C NM_000076.2 **CENPA** NM_001809.3 CHEK2 NM_007194.3 CSF1R NM_001288705.1 CTLA4 NM_005214.4 CXCR4 NM_003467.2 DCUN1D1 NM_020640.3 **DNAJB1** NM 006145.2 **DPYD** NM_000110.3 EMSY NM_001300942.1 EPHA5 NM_001281765.2 ERBB3 NM 001982.3 ERCC4 NM 005236.2 ETV4 NM 001079675.2 **EZR** NM_001111077.1 FANCF NM_022725.3 FAS NM_000043.5 FGF14 NM_175929.2 FGF6 NM 020996.2 FGFR4 NM_002011.4 FLT4 NM_182925.4 FRS2 NM_001042555.2 GATA2 NM_001145661.1 GID4 NM_024052.4 **GNAS** NM 080425.3 **GSK3B** NM_002093.3 HDAC2 NM 001527.3 HIST1H3B NM_003537.3 HIST1H3H NM_003536.2 HLA-A NM 001242758.1

IDH1 NM_005896.3 IGF2 NM_000612.5 INHA NM_002191.3 IRF2 NM_002199.3 JAK2 NM_004972.3 KDM5C NM_004187.3 KIT NM_000222.2 KMT2D NM_003482.3

HRAS NM_005343.3

ABRAXAS1 NM_139076.2 AKT3 NM_005465.4 **AR** NM_000044.3 **ARID2** NM 152641.2 **ATR** NM_001184.3 AXIN2 NM 004655.3 BBC3 NM_001127240.2 BCL2L2 NM 001199839.1 BLM NM 000057.3 BRD4 NM 058243.2 BTG2 NM_006763.2 **CBFB** NM_022845.2 CCNE1 NM_001238.3 CDC73 NM_024529.4 CDK6 NM 001145306.1 CDKN2A NM_000077.4 CHD1 NM_001270.2 **CIC** NM_015125.4 CSF3R NM_156039.3 **CTNNA1** NM_001323982.1 **CYLD** NM_015247.2 DDR1 NM_013994.2 **DNMT1** NM 001130823.2 **EED** NM_001308007.1 **EP300** NM 001429.3 EPHA7 NM_004440.3 ERBB4 NM_005235.2 ERCC5 NM 000123.3 ETV5 NM 004454.2 FANCA NM_000135.2 FANCG NM_004629.1 FAT1 NM_005245.3 FGF19 NM_005117.2 FGF7 NM 002009.3 FH NM_000143.3 FOXA1 NM_004496.3 FUBP1 NM_003902.4 GATA3 NM_001002295.1 GLI1 NM_005269.2 GPS2 NM 004489.4 H3F3A NM_002107.4 HGF NM 000601.5 HIST1H3C NM_003531.2 HIST1H3I NM_003533.2 HLA-B NM 005514.7 HSD3B1 NM_000862.2 IDH2 NM_002168.3 **IKBKE** NM 014002 3 **INHBA** NM_002192.3 IRF4 NM_002460.3 JAK3 NM 000215.3 **KDM6A** NM_001291415.1

ACVR1 NM_001105.4 ALK NM_004304.4 **ARAF** NM_001256196.1 **ARID5B** NM 032199.2 ATRX NM_000489.4 **AXL** NM 021913.4 BCL10 NM_003921.4 BCL6 NM 001130845.1 BMPR1A NM 004329.2 BRD7 NM 001173984.2 BTK NM_000061.2 **CBL** NM_005188.3 CD274 NM_014143.3 CDH1 NM_004360.4 CDK8 NM 001260.2 **CDKN2B** NM_004936.3 CHD2 NM_001271.3 **CREBBP** NM_004380.2 **CSMD1** NM_033225.5 CTNNB1 NM_001904.3 CYP17A1 NM_000102.3 DDR2 NM_001014796.1 **DNMT3A** NM 022552.4 EGFR NM_005228.3 **EPCAM** NM 002354.2 **EPHB1** NM_004441.4 ERCC1 NM_202001.2 ERG NM 001136154.1 ETV6 NM 001987.4 FANCC NM_000136.2 FANCI NM_001113378.1 FBXW7 NM_033632.3 FGF23 NM_020638.2 FGFR1 NM_023110.2 FLCN NM_144997.5 FOXL2 NM_023067.3 FYN NM_002037.5 GATA4 NM_001308093.1 GNA11 NM_002067.4 **GREM1** NM 013372.6 H3F3B NM_005324.4 HIST1H1C NM 005319.3 HIST1H3D NM_003530.4 HIST1H3J NM_003535.2 HLA-C NM 001243042.1 HSP90AA1 NM 001017963 2 IFNGR1 NM_000416.2 IKZF1 NM_006060.5 **INPP4A** NM_001134224.1 IRS1 NM_005544.2 JUN NM 002228.3

KDR NM_002253.2

KLHL6 NM_130446.2

LATS1 NM_004690.3

ACVR1B NM_020328.3 ALOX12B NM_001139.2 **ARFRP1** NM_001267547.2 **ASXL1** NM 015338.5 AURKA NM_001323303.1 B2M NM 004048.2 BCL2 NM_000633.2 BCOR NM 001123383.1 BRAF NM 004333.4 BRINP3 NM 199051.2 CALR NM_004343.3 CCND1 NM_053056.2 **CD74** NM_001025159.2 CDH18 NM_001291956.1 CDKN1A NM 001291549.1 CDKN2C NM_001262.2 CHD4 NM_001273.3 CRKL NM_005207.3 **CSMD3** NM_198123.1 CUL3 NM_001257198.1 CYP2D6 NM_000106.5 DICER1 NM_177438.2 DNMT3B NM 006892.3 EIF1AX NM_001412.3 EPHA2 NM 004431.4 EPHB4 NM_004444.4 ERCC2 NM 000400.3 ERRFI1 NM 018948.3 EWSR1 NM 013986.3 FANCD2 NM_001018115.2 FANCL NM_018062.3 FGF10 NM_004465.1 FGF3 NM_005247.2 FGFR2 NM 000141.4 FLT1 NM_002019.4 FOXO1 NM_002015.3 GABRA6 NM_000811.2 GATA6 NM_005257.5 GNA13 NM_006572.5 GRIN2A NM 000833.4 H3F3C NM_001013699.2 HIST1H2BD NM 021063.3 HIST1H3E NM_003532.2 HIST2H3D NM_001123375.2 HNF1A NM 000545.6 ICOSLG NM_001283050.1

IGF1 NM_001111285.2 IL10 NM_000572.2 INPP4B NM_001101669.1 IRS2 NM_003749.2 KAT6A NM_006766.4 KEAP1 NM_012289.3 KMT2A NM_001197104.1 LATS2 NM_014572.2

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KLF4 NM_001314052.1

KRAS NM_033360.3

MATE MATE MATERIA

LMO1 NM 002315.2 MAGI2 NM_012301.3 MAP3K1 NM_005921.1 MCL1 NM_021960.4 MEF2B NM_001145785.1 MIR21 NR 029493.1 **MPL** NM 005373.2 MST1 NM_020998.3 MYC NM_002467.4 NAV3 NM_001024383.1 NEGR1 NM_173808.2 NKX2-1 NM_001079668.2 NOTCH4 NM_004557.3 NSD2 NM_001042424.2 NTRK2 NM_006180.4 PAK1 NM_001128620.1 PARP2 NM_005484.3 PDCD1 NM 005018.2 PGR NM_000926.4 PIK3CA NM 006218.3 PIK3R2 NM_005027.3 PMS1 NM_000534.4 **PPARG** NM_015869.4 PRDM1 NM 001198.3 **PRKN** NM 004562.2 PTPRO NM 030667.2 RAC1 NM_018890.3 RAD51C NM_058216.2 RARA NM_000964.3 **REL** NM 002908.3 **RIT1** NM 001256821.1 **RPS6KB2** NM_003952.2 SDC4 NM_002999.3 **SDHD** NM_003002.3 SH2D1A NM_002351.4 SMAD2 NM_001003652.3 SMARCD1 NM 003076.4 SOX17 NM_022454.3 SPTA1 NM 003126.2 **STAT4** NM_001243835.1 SUFU NM_016169.3 TCF7L2 NM_001146274.1 TFT1 NM 030625.2 TMEM127 NM 017849.3 **TOP2A** NM_001067.3 TRIM58 NM_015431.3 TYRO3 NM_006293.3 VHL NM_000551.3 XPO1 NM 003400.3 **ZBTB16** NM_001018011.1

Specimen ID RS19058620FFP LRP1B NM 018557.2 MALT1 NM_006785.3 MAP3K13 NM 0012423141 MAPK1 NM 0027454 MDC1 NM_014641.2 MEN1 NM_000244.3 MITF NM_000248.3 MRE11 NM 005591.3 MST1R NM_002447.3 MYCL NM 001033082.2 NBN NM_002485.4 NF1 NM_000267.3 NKX3-1 NM_006167.3 NPM1 NM_002520.6 NSD3 NM_023034.1 NTRK3 NM_001012338.2 PAK3 NM_001128168.2 **PARP3** NM_001003931.3 PDCD1LG2 NM 025239.3 PHOX2B NM_003924.3 **PIK3CB** NM 006219.2 PIK3R3 NM_001303427.1 PMS2 NM_000535.6 **PPM1D** NM_003620.3 PREX2 NM 024870.3 PTCH1 NM 000264.3 PTPRS NM_002850.3 RAD21 NM_006265.2 RAD51D NM_002878.3 RASA1 NM_002890.2 **RET** NM 020975.4 RNF43 NM 017763.5 **RPTOR** NM_020761.2 **SDHA** NM_004168.3 SETD2 NM_014159.6 SHQ1 NM_018130.2 SMAD3 NM_005902.3 SMO NM 005631.4 SOX2 NM_003106.3 **SRC** NM 198291.2 **STAT5A** NM_001288718.1 **SYK** NM 001174167.2 TEK NM_000459.4 TET2 NM 001127208.2 TMPRSS2 NM_001135099.1 TNFAIP3 NM_001270507.1 TP53 NM_000546.5 TRPC5 NM_012471.2 **U2AF1** NM_001025203.1 WISP3 NM_198239.1 XRCC2 NM_005431.1 **ZBTB2** NM_020861.2

2019/11/19 PM LTK NM 002344.5 MAP2K1 NM_002755.3 MDM2 NM_002392.5 MERTK NM_006343.2 MKNK1 NM_003684.5 MSH2 NM 000251.2 MTAP NM_002451.3 MYCN NM_001293228.1 NCOA3 NM_181659.2 NF2 NM_000268.3 NOTCH1 NM_017617.4 NRAS NM_002524.4 NT5C2 NM_001134373.2 NUP93 NM_014669.4 PAK5 NM_020341.3 PAX5 NM_016734.2 **PDGFRA** NM 006206.4 PIK3C2B NM_002646.3 PIK3CD NM_005026.3 PIM1 NM_001243186.1 **PNRC1** NM 006813.2 **PPP2R1A** NM_014225.5 **PRKAR1A** NM 002734.4 **PTEN** NM 000314.6 **PTPRT** NM 133170.3 RAD50 NM_005732.3 RAD52 NM_001297419.1 **RB1** NM_000321.2 **RHEB** NM 005614.3 ROS1 NM 002944.2 RSPO2 NM_178565.4 **SDHAF2** NM_017841.2 SF3B1 NM_012433.3 SLC34A2 NM_006424.2 SMAD4 NM_005359.5 **SNCAIP** NM 001308100.1 **SOX9** NM_000346.3 SRSF2 NM 003016.4 **STAT5B** NM_012448.3 TAF1 NM_001286074.1 TENT5C NM_017709.3 **TGFBR1** NM 001306210.1 **TP63** NM 003722.4 TSC1 NM 000368.4 UGT1A1 NM_000463.2 WRN NM_000553.4 XRCC3 NM_001100118.1 ZNF217 NM_006526.2

Report Date

TEST

OncoScreen Plus™ LYN NM 002350.3 MAP2K2 NM_030662.3 MAPK3 NM 0027462 MDM4 NM_002393.4 MET NM_000245.3 MLH1 NM_000249.3 MSH3 NM 002439.4 MTOR NM_004958.3 MYD88 NM_002468.4 NCOR1 NM_006311.3 NFE2L2 NM_006164.4 NOTCH2 NM_024408.3 NRG1 NM_001322205.1 NTHL1 NM_002528.6 **NUTM1** NM_001284292.1 PALB2 NM_024675.3 **PBRM1** NM_018313.4 **PDGFRB** NM 002609.3 PIK3C2G NM_001288772.1 PIK3CG NM 001282426.1 PLCG2 NM_002661.4 POLD1 NM_001256849.1 PPP2R2A NM_002717.3 **PRKCI** NM 002740.5 PTPN11 NM 002834.3 **QKI** NM_006775.2 RAD51 NM_001164269.1 RAD54L NM_003579.3 RBM10 NM_001204468.1 **RHOA** NM_001664.3 **RPA1** NM 002945.3 RUNX1 NM_001754.4 **SDHB** NM_003000.2 SGK1 NM_001143676.1 SLIT2 NM_004787.3 SOCS1 NM 003745.1 **SPEN** NM_015001.2 STAG2 NM 001042749.2 **STK11** NM_000455.4 TBX3 NM 016569.3 TERC NR_001566.1 TGFBR2 NM 001024847.2 TNFRSF14 NM 003820.3 TRAF2 NM_021138.3 TSC2 NM_000548.4 VEGFA NM_001025366.2 WT1 NM_024426.4 **YAP1** NM_001282101.1 ZNF703 NM_025069.2

MAF NM 005360.4 MAP2K4 NM_001281435.1 MAX NM_002382.4 MED12 NM_005120.2 MGA NM_001164273.1 MLH3 NM 001040108.1 MSH6 NM 000179.2 MUTYH NM_001128425.1 MYOD1 NM_002478.4 NCOR2 NM_006312.5 NFKBIA NM_020529.2 NOTCH3 NM_000435.2 NSD1 NM_022455.4 NTRK1 NM_001007792.1 P2RY8 NM_178129.4 PARP1 NM_001618.3 PCDH11X NM_032968.4 PDK1 NM 001278549.1 PIK3C3 NM_002647.3 PIK3R1 NM 181523.2 PLK2 NM_006622.3 POLE NM_006231.3 **PPP6C** NM_001123355.1 **PRKDC** NM 006904.6 PTPRD NM 002839.3 RAB35 NM_006861.6 RAD51B NM_133509.3 RAF1 NM_002880.3 RECQL4 NM_004260.3 **RICTOR** NM_001285439.1 RPS6KA4 NM 003942.2 RUNX1T1 NM_001198679.1 **SDHC** NM_003001.3 SH2B3 NM_005475.2 SLX4 NM_032444.2 SMARCA4 NM_001128849.1 SMARCB1 NM_003073.4 SOX10 NM 006941.3 **SPOP** NM_001007226.1 **STAT3** NM 139276.2 STK40 NM_001282546.1 TCF3 NM_003200.3 TERT NM_198253.2 TIPARP NM_001184717.1 TOP1 NM 003286.2 TRAF7 NM 032271.2 **TSHR** NM_000369.2 VEGFB NM_003377.4 **XIAP** NM_001167.3 YES1 NM_005433.3 **ZNRF3** NM_001206998.1

Note : The NM number after the gene name is the transcript number (RefSeq) used when analyzing.

Gene List Focused on Rearrangements ALK BRAF CD274 **CD74** ETV4 ETV5 ETV6 EWSR1 EZR FGFR1 FGFR2 FGFR3 NRG1 NTRK1 NTRK2 NTRK3 RAF1 RET

	Specimen ID	Report Date	TEST	
	RS19058620FFP	2019/11/19 PM	OncoScreen Plus™	
ROS1 RSPO2	SDC4	SLC34A2	TMPRSS2	



Report Date 2019/11/19 PM

APPENDIX4 : REFERENCE

- 1. NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines $\ensuremath{\mathbb{R}}$)
- 2. Li MM et al. (2017) Standards and Guidelines for the Interpretation and Reporting of Sequence Variants in Cancer: A Joint Consensus Recommendation of the Association for Molecular Pathology, American Society of Clinical Oncology, and College of American Pathologists. J Mol Diagn [PMID: 27993330]
- 3. Kalia SS et al. (2017) Recommendations for reporting of secondary findings in clinical exome and genome sequencin g, 2016 update (ACMG SF v2.0): a policy statement of the American College of Medical Genetics and Genomics. Gen et. Med. [PMID: 27854360]
- 4. Richards S et al. (2016) Standards and guidelines for the interpretation of sequence variants: a joint consensus recom mendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet. Med. [PMID: 25741868]
- 5. Ng KP et al. (2012) A common BIM deletion polymorphism mediates intrinsic resistance and inferior responses to tyr osine kinase inhibitors in cancer. Nat. Med. [PMID: 22426421]



Specimen ID: RS19058622FFP

PD-L1 (Antibody Clone 22C3) Report

Burning Rock Dx | a CLIA certified lab

Report Serial Number 3131901668 Dat2019/11/15 AM

Anti-Forgery Port

	- 224	Specimen ID	Date	Test	版本号: 2.0 生	效日期:2020.09.30
	s 3 ck Dx	RS19058622FFP	2019/11/15 AM	PD-L1 (22C3	3)	
Patient In	formation					
Name	吴成云	Specimen ID	RS19058622FFP	Applicatio	on Form	A00144592
Gender	Male	Specimen Type	Section	Accession	Date	2019/11/11
Age	67	Biopsy Type	Puncture	Specimen	Received	2019/11/13 AM
Patient ID	3209211952** 1	Specimen Site	Lung	Report Da	ate	2019/11/15 AM
Tumor Type	Unknown	Hospital	江苏省人民医院			
Additional Information	Not provided					

*NOTE: Patient information was provided by the patient upon sample receiving. Above information is not conferred from this report therefore, this report is not responsible for the accuracy of the above information.

Introduction of Products

Protein	Clone number	Method	Reagent	Clinical interpretation
1 ו_חפ	2263	шс	Mouse anti-Human PD-L1	Keytruda (Pembrolizumab) Antibody for
FD-LI	PD-L1 22C3 IHC		Monoclonal Antibody	Companion Diagnostics

This report is only responsible for the detection of this specimen, and the results are only for medical reference.



Thechnician: 梁丽仪



Quality Control (QC)

Quality Parameter		Value	QC Standard
Pathological Assessment	Number of malignant cells	≥100	≥100
Overall Quality Assessment		Qualified	

Result

Evaluation Protein	TPS	CPS
PD-L1	90%	/

NOTE Please refer to the results of PD-L1 protein expression according to the corresponding instructions of immunotherapy treatments and the evaluation criteria of different types of tumors.

Result Analytical Reference:

Methods:

TPS: PD-L1 stained tumor cell /total tumor cell (PD-L1 stained and non-stained) x100%

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CPS: PD-L1 stained cell (tumor cell, lymphocyte, macrophage)/total tumor cell (PD-L1 stained and non-
```

stained) x100

Limitations:

The following samples are not suitable for CPS evaluation

- a. Cytological samples
- b. Shredded tissue samples
- c. Decalcification samples of bone metastases

Definition of boundary value:

Negative: < 1%

Low expression: 1-49%

High expression: $\geq 50\%$

NOTE This threshold is the definition of TPS test results for NSCLC samples.

NOTE 1. Due to tumor heterogeneity (temporal and spatial), and the way of sample preservation, the results only reflect the specimen sent for examination.

MACE学 Burning Rock Dx	Specimen ID	Date	Test	版本号: 2.0 生效日期: 2020.09.30
	RS19058622FFP	2019/11/15 AM	PD-L1 (2	2C3)

2. If the number of malignant tumor cells is less than 100, the accuracy of the detection may be affected.

3. The pharmacokinetics and the effect of its in vivo process are complex, and there are many factors affecting the efficacy and toxicity of the drug. Thus, doctors should consider the following factors prior to treatment decision-making, including but not limited to the pathophysiological characteristics of the patient, clinical manifestations, adjuvant therapy that is being used or to be used. This report serves as a reference for treatment-guidance based on the results of this test. Do not use this result as the sole basis for treatment-guidance.

Specimen ID	Date	Test	版本号: 2.	0 生效日期:	2020.09.30
RS19058622FFP	2019/11/15 AM	PD-L1 (22C	3)		

Result chart

Sample results:



TPS

NA

CPS

QC sample results:



Positive Control



Negative Control

	Specimen ID	Date	Test	版本号:	2.0 生效日期:	2020.09.30
	RS19058622FFP	2019/11/15 AM	PD-L1 (22)	C3)		

PD-L1 INTRODUCTION

PD-L1 (Programmed death-ligand 1, B7-H1 or CD274) protein, a type I transmembrane protein with a molecular weight of 40 kDa, is the ligand of PD-1 (programmed death-1). The encoding gene of PD-L1 is located on human chromosome 9p24.2 and its extracellular region of PD-L1 contains Ig-V and Ig-C-like domains. PD-1 is expressed in a wide range of cells, including T cells, B cells, dendritic cells, macrophages, mast cells, fibroblasts, and mesenchymal stem cells. PD-L1 is also widely expressed on the cell surface of tumor cells, and is proposed to be responsible for promoting tumor immune escape.

When binding to its ligand PD-L1, PD-1 can activate intracellular signaling pathways and inhibit the activity of killer T lymphocytes, thereby reducing the secretion of cytokines by immune cells. During infection and inflammation, interaction of PD-1 with PD-L1 helps to prevent autoimmunity to maintain immune homeostasis. In the tumor microenvironment, after binding to PD-L1 on the cell surface of tumor cells, PD-1 on the surface of tumor specific cytolytic T lymphocytes (CTL) can inhibit the activation of CTLs, and even promote the apoptosis of CTLs, ultimately leading to tumor immune escape.

Since the PD-L1/PD-1 signaling pathway plays a vital role in tumor immune escape, it becomes an important target of immunotherapy. Anti-PD-1 monoclonal antibodies (such as nivolumab and pembrolizumab) and anti-PD-L1 monoclonal antibodies (such as atezolizumab, durvalumab, and avelumab) could potentially restore the immune response of cytolytic T lymphocytes to tumor cells by blocking the PD-L1/PD-1 signaling pathway so as to prevent tumors from achieving immune evasion, and thus improve the prognosis of patients. Several antibodies targeting the PD-L1-PD-1 axis have been approved by the FDA for multiple cancers, including malignant melanoma(MM), non-small cell lung cancer (NSCLC), renal cell carcinoma(RCC), urothelial carcinoma, head and neck carcinoma, Hodgkin lymphoma, gastric cancer, hepatocellular carcinoma(HCC), small cell lung cancer and solid tumors with microsatellite instability-high(MSI-H). In addition, a large number of antibodies targeting the PD-L1-PD-1 axis are in clinical development.

Expression of PD-L1 is one of the more promising prognostic biomarker for predicting response to monotherapies blocking the PD-L1-PD-1 axis. In several clinical trials of antagonistic drugs designed to block PD-L1/PD-1, PD-L1 expression in tumor cells+/- immune cells has been used for stratifying patients. In addition, PD-L1 IHC is used as either the companion or the complementary diagnostic assay for several approved indications of PD-1/PD-L1 monoclonal antibodies.

Monoclonal antibody PD-L1 IHC 22C3 is the companion diagnostic antibody used in multiple registered clinical trials of pembrolizumab (KEYTRUDA) . It is also the first PD-L1 antibody approved for companion diagnosis of PD-1/PD-L1 immunotherapy in following indications: NSCLC, gastric or esophagogastric junction adenocarcinoma (GEJ), esophageal squamous cell carcinomas, cervical cancer, urothelial carcinoma, and head and neck cancers. In NSCLC, PD-L1 protein expression is accessed by tumor proportion score (TPS), which refers to percentage of complete or partial cell membrane staining of tumor cells with any intensity over total tumor cells in the denominator. PD-L1 positive is defined as TPS \geq 1% and high PD-L1 expression as TPS defin. While in

gastric or esophagogastric junction adenocarcinoma(GEJ), esophageal squamous cell carcinomas, cervical cancer, urothelial carcinoma, and head and neck cancers, PD-L1 expression is categorized by combined proportion score (CPS), which is defined as the sum of PD-L1 stained tumor cell and surrounding lymphocytes and macrophages divided by the total number of viable tumor cells multiplied by 100. In gastric or esophagogastric junction adenocarcinoma(GEJ), cervical cancer, and head and neck cancers, PD-L1 positive is defined as CPS \geq 1, and in esophageal squamous cell carcinomas and urothelial carcinoma, positive threshold is CPS \geq 10.

There is a lack of consensus on the correlation of the level of PD-L1 and efficacy of PD-1/PL-D1 inhibitors in solid tumors due to the presence of various anti-PD-L1 antibodies and multiple platforms for PD-L1 detection. Multiple clinical trials show that high expression of PD-L1 is a favorable prognostic biomarker, but regardless of how cutoff is defined, there is no definitive indication of whether the patient will benefit from the therapy. Thus, selecting threshold of positive PD-L1 expression for different cancer patients should be based on expected clinical benefit, patients 'ability to shoulder economically, and other factors, particularly for therapies that PD-L1 detection has not been approved as companion diagnostics.

Clinical trials suggested that expression of PD-L1 is not an optimal prognostic biomarker for predicting response to PD-L1/PD-1 immunotherapy. Patients with positive or high PD-L1 expression have a higher objective response rate compared with those with negative or low PD-L1 expression. Many PD-L1-positive tumors, however, do not respond, and a few PD-L1-negative tumors do respond to PD-L1/PD-1 immunotherapy. Thus, detection and assessment of PD-L1 expression in the ongoing clinical trials is just the first step in exploring the prognostic marker for predicting response of PD-1/PD-L1 inhibitors and it should not be the prerequisite and the only standard for selecting candidate patients receiving PD-1/PD-L1 immunotherapy.



Figure 1. PD-L1 positive patients had better objective response to anti-PD-1/PD-L1 therapy than PD-L1 negative patients. (Graph was cited from Joel Sunshine, Janis M Taube. PD-1/PD-L1 inhibitors. Current Opinion in Pharmacology 2015,23:32-38)