

Current Molecular and Clinical Landscape of ATRT – The Link to Future Therapies

Katharina Gastberger^{1,2}, Victoria E Fincke^{1,2}, Marlena Mucha^{1,2}, Reiner Siebert³,
Martin Hasselblatt⁴, Michael C Frühwald^{1,2}

¹Pediatrics and Adolescent Medicine, Swabian Children's Cancer Center, University Medical Center Augsburg, Augsburg, Germany; ²Bavarian Cancer Research Center (BZKF), Augsburg, Germany; ³Institute of Human Genetics, Ulm University & Ulm University Medical Center, Ulm, Germany; ⁴Institute of Neuropathology, University Hospital Münster, Münster, Germany

Correspondence: Michael C Frühwald, EU-RHAB Center, Swabian Children's Cancer Center, Pediatrics and Adolescent Medicine, University Medical Center Augsburg, and Bavarian Cancer Research Center (BZKF), Stenglinstr. 2, Augsburg, 86156, Germany, Tel +49 821 400 9340, Fax +49 821 400 179201, Email michael.fruehwald@uk-augsburg.de

Abstract: ATRT is a highly aggressive and rare pediatric CNS tumor of very young children. Its genetic hallmark is bi-allelic inactivation of *SMARCB1* encoding INI1. Rarely *SMARCA4* encoding BRG1 is affected. Up to 30% are associated with constitutional heterozygous pathogenic variants in one of the two genes, giving rise to the Rhabdoid-Tumor-Predisposition-Syndromes (RTPS) 1 and 2. Characteristic DNA methylation profiles distinguish ATRT from other *SMARCB1*-deficient entities. Three distinct subtypes ATRT-MYC, -TYR, and -SHH are on record. ATRT-SHH may be further divided into the subgroups ATRT-SHH1A, -SHH1B, and -SHH2. The cure of ATRT remains challenging, notwithstanding an increasing understanding of molecular pathomechanisms and genetic background. The implementation of multimodal institutional treatment protocols has improved prognosis. Regardless of treatment approaches, clinical risk factors such as age, metastases, and DNA methylation subtype affect survival probability. We provide a critical appraisal of current conventional multimodal regimens and emerging targeted treatment approaches investigated in clinical trials and entity-specific registries. Intense treatment approaches featuring radiotherapy (RT) and high-dose chemotherapy (HDCT) face the difficulty of balancing tumor control and treatment-related toxicity. Current approaches focus on minimizing radiation fields by proton beam therapy or to withhold RT in HDCT-only approaches. Still, a 40–75% relapse rate upon first-line treatment reveals the need for novel treatment strategies in primary and even more in recurrent/refractory (r/r) disease. Among targeted treatments, immune checkpoint inhibitors and epigenetically active agents appear most promising. Success remains limited in single agent approaches. We hypothesize that mechanism-informed combination therapy will enhance response, as the low mutational burden of ATRT may contribute to acquiring resistance to single targeted agents. As DNA methylation group-specific gene expression profiles appear to influence response to distinct agents, the future treatment of ATRT should respect clinical and biological heterogeneity in risk group adjusted treatment protocols.

Keywords: SWI/SNF related matrix associated, actin dependent regulator of chromatin, subfamily b, member 1, SMARCB1, atypical teratoid rhabdoid tumor, ATRT, central nervous system, CNS, treatment, pediatric, cell cycle

The Burden of Atypical Teratoid Rhabdoid Tumors ATRT – A Rare and Unique Entity

Atypical Teratoid Rhabdoid Tumors (ATRTs) are WHO grade 4 embryonal CNS tumors.¹ They predominantly affect children and young adults with incidences ranging from 0.3/100,000 to 0.6/100,000 in the first year of life and 0.03/100,000 to 0.26/100,000 for children between 5 and 9 years of age in the USA and Europe, respectively.^{2,3}

Typically, ATRTs are located infratentorial (60%) and in 22% are located in the cerebral hemispheres, with the mesencephalon and the pineal region being less common locations (each 4%). Primary intraspinal ATRTs are a rarity (1–2%)^{4,5} and occasionally cannot be distinguished clearly from spinal extracranial, extrarenal malignant Rhabdoid Tumors (eMRT).⁶ Rhabdoid tumors tend to appear synchronously: In 6% of all cases, two or more rhabdoid tumors are detected



at different anatomic sites, with the CNS being one in most instances.^{4,7–11} Dissemination and metastases along the cerebrospinal fluid (CSF) compartment are common and initially diagnosed in 20–30% of cases.^{4,5,12}

Diagnosis of ATRT can be confirmed immunohistochemically by the nuclear loss of SMARCB1 (INI1)-staining. In comparison to other *SMARCB1*-deficient CNS tumors, ATRT is presented with specific DNA methylation profiles. Histopathologic differential diagnoses in the scope of *SMARCB1*-deficient tumor entities are listed in Table 1 and comprise LGDIT,¹³ desmoplastic myxoid tumor, *SMARCB1*-mutant,^{14,15} CRINET,^{16,17} PXA,¹⁸ or *SMARCB1*-deficient malignant tumors in the context of Li-Fraumeni syndrome.¹⁹

ATRT Exhibit Uniform Inactivation of SMARCB1 or SMARCA4 but Comprise Distinct Molecular Subgroups

Presenting rather stable copy number variation profiles, rhabdoid tumors demonstrate pathogenic variants in only a few genes with *SMARCB1* or *SMARCA4* consistently inactivated in tumor cells. *SMARCB1* and *SMARCA4* encode for subunits of the SWI/SNF chromatin remodeling complex. ATRT is, thus, a disease intimately linked to epigenetic deregulation.^{27,28} ATRT can be grouped into three distinct major molecular subtypes, ATRT-MYC, -SHH and -TYR. Each of these is associated with distinct DNA-methylation and gene expression profiles as well as anatomic and clinical characteristics.^{24,27} The level of immune cell infiltration and immune checkpoint expression also differ between ATRT subgroups.²⁹ ATRT-MYC tumors exhibit similarities with the DNA methylation subgroups of extracranial rhabdoid tumors (group 3 = RTK like and group 4 = extra-renal MRT like).²⁹ ATRT-SHH was found to cluster into further subgroups. Federico et al described three distinct subgroups with respect to DNA-methylation profile: ATRT-SHH1A, -1B, and SHH2. ATRT-SHH1B are located primarily supratentorial, affect older children (median age for -SHH1B 107 months, -SHH1A 18 months, -SHH2 13 months), and are associated with a significantly improved survival probability compared to -SHH1A and -SHH2.³⁰ By converging imaging and multi-omics analyses, Lobón-Iglesias et al detected two distinct ATRT-SHH subgroups: CAL SHH ATRT (cerebral anterior lobe, overexpression in *EN2*) and BG/IV SHH ATRT (basal ganglia, intra ventricular, overexpression in *OLIG2*), which may correspond to ATRT-SHH2 and ATRT SHH-1A, respectively.³¹ Taken together, this strongly suggests the existence of distinct ATRT-SHH subgroups.

The relevance of DNA methylation subtypes as prognostic markers and therapeutic targets in an extremely rare disease remains currently unresolved. We could show that features of the ATRT-TYR group in combination with older age at diagnosis bear a favorable prognostic significance (5-year OS 0% and 70% in ATRT-non-TYR and <1 year and ATRT-TYR and ≥1 year, respectively; n=143). Upadhyaya et al confirmed this beneficial impact of ATRT-TYR but detected a diminished effect among ATRTs in M0 (n=22).³² For extracranial malignant rhabdoid tumors (eMRT), metastatic disease, GTR, and germ line mutation (GLM) distinguish high risk from standard risk groups.³³ Future research will need to focus on evaluating potential confounders. Nevertheless, the fifth edition of the WHO classification of Tumors of the Central Nervous System (CNS 5) sheds light on the descriptive importance of the three distinct methylation subtypes of ATRT.¹

Rhabdoid-Tumor-Predisposition Syndrome

SWI/SNF deficiency in rhabdoid tumors is a result of biallelic loss of *SMARCB1* or *SMARCA4*. In approximately 25–30%^{4,21–23} of the patients with ATRT the first hit is a constitutional event, ie, it is supposed to occur in the germline and leads to heterozygous inactivation of one allele of the respective gene in all cells of the body. Such constitutional inactivation (often also called germ line mutation, GLM) occurs de novo, as in most cases of constitutional inactivation of *SMARCB1*, or is inherited from a parent, as for most cases of *SMARCA4*. Constitutional inactivation of one copy of *SMARCB1* or *SMARCA4* gives rise to rhabdoid tumor predisposition syndrome (RTPS; RTPS1, GLM in *SMARCB1*; RTPS2, GLM in *SMARCA4*). Penetrance is considered high for RTPS1 but incomplete for RTPS2. Clinical features of RTPS include an earlier onset of disease and multiple synchronous tumor sites. Clustering of rhabdoid tumors and SWI/SNF deficient tumors in families has been reported, and frequently due to de novo mutation.^{34–36} Despite an inferior prognostic outlook and severe clinical courses, a significant number of long-time survivors affected by RTPS are on record.^{37–40}

Table 1 Differential Diagnoses of *SMARCB1* Deficient (Pediatric) CNS Tumors

Tumor Grade	Tumor Type	Histopathology	Localization	Genomic Alterations	DNA Methylation Profile	Age at Diagnosis	Prognosis
Low	LGDIT, <i>SMARCB1</i> mutant ¹³	Low to moderate cellularity, pleomorphic glial and neuronal cells, Ki67 8%	Supratentorial	Homozygous <i>SMARCB1</i> -deletion	Similarity to ATRT-MYC	Median 16 years	Good response to multimodal treatment, but potential to progress to ATRT
Low	Desmoplastic myxoid tumor, <i>SMARCB1</i> -mutant ¹⁵	Spindle and epithelioid cells embedded in a desmoplastic stroma, loose myxoid matrix; Ki67 3%	Pineal region	Heterozygous or homozygous <i>SMARCB1</i> deletion	Distinct DNA methylation profile in proximity to ATRT-MYC	Median 40 years (15–61 years)	Stable disease without CT, RT following complete surgical resection
High	ATRT	Poorly differentiated, multi-lineage phenotype, glial, mesenchymal, epithelial components, rhabdoid cells, small round blue cells ^{5,20}	Supratentorial, infratentorial and (less common) spinal ⁴	Biallelic alterations in <i>SMARCB1</i> / <i>SMARCA4</i> , GLM in 30% ^{4,21–23}	ATRT <ul style="list-style-type: none"> • TYR • SHH • MYC²⁴ 	Median 18 months ²⁵	5-year OS 0–71% dependent on risk factors ⁴
High	Cribriform neuroepithelial tumor (CRINET) ¹⁶	Cribriform growth pattern with neuroepithelial tumor cells; Ki67/MIB1 30%,	Often intraventricular	Large heterozygous deletions of 22q + <i>SMARCB1</i> mutations, often affecting the C-terminus ²⁶	Similarity with ATRT-TYR	10–26 months	Good response to multimodal treatment (I partial resection, CT, RT; I GTR, CT) ¹⁶
High	ATRT with molecular features of PXA ¹⁸	Rhabdoid cells, brisk mitotic activity, geographic necroses, Ki67 high 15–60%	Supratentorial, temporal lobe	Homozygous <i>SMARCB1</i> deletion, genetic features of PXA (homozygous <i>CDKN2A/B</i> deletion), gain of whole chromosome 7 <i>BRAF</i> V600E mutations)	Similarity with PXA	10–18 years, (I case report of a 62-year old female)	To be determined, <i>BRAF</i> mutation as novel therapeutic target?
High	<i>SMARCB1</i> -deficient malignant brain tumors in the context of Li-Fraumeni syndrome ¹⁹	Glial phenotype, pleomorphic nuclei, fibrillary process, no nuclear accumulation of p53 protein, scattered rhabdoid tumor cells ⁴	Supratentorial, temporal lobe	Complex copy number alterations and <i>TP53</i> mutation	Similarity with ATRT or malignant gliomas.	5–19 years	Depending on histopathological profile; to be determined

Germline predisposition in *SMARCB1/SMARCA4*-deficient ATRT is associated with younger age at diagnosis, which is also a common feature of other well-known genetic tumor syndromes like BRCA-related cancer predisposition or retinoblastoma. Nevertheless, in contrast to other genetic tumor syndromes, younger age itself may explain inferior prognosis as the spectrum of age adjusted treatment approaches is extremely limited (ie, aggressive surgery or radiotherapy). Moreover, it might have to be taken into account that *SMARCB1* and *SMARCA4* are more or less ubiquitously expressed and play a major role in chromatin remodeling and epigenetic control in various cell compartments. As an example, expression levels of *SMARCB1* have been linked to the immunoglobulin V-region mutational load in germinal center B-cells⁴¹ and somatic inactivation of *SMARCA4* is common in germinal center B-cell derived lymphomas like Burkitt lymphoma.⁴² It is, thus, intriguing to speculate, that haploinsufficiency, eg, in cell compartments other than tumor cells, like in cells of immunological compartments, might contribute to the aggressive biological behavior of ATRT in patients with constitutional *SMARCB1* alterations, eg, via contributing to immune escape.

Clinical Management of Atypical Teratoid Rhabdoid Tumors – Current Conventional Therapeutic Approaches

Reflecting the rarity of ATRT, international guidelines for clinical practice remain limited. The “Canadian Pediatric Neuro-Oncology Standards of Practice”, published in 2020, is one of the few detailed clinical practice guidelines publicly available.⁴³

Lessons from Phase III Clinical Trials and Entity Specific Registries

The diversity of data sets and treatment approaches in small cohorts explain the complexity of meta-analyses on which to base recommendations for therapy. Treatment elements such as maximal safe resection, conventional or high-dose chemotherapy (CT vs HDCT), and radiotherapy (RT) represent the most important components of first-line treatment in clinical trials and consensus therapy recommendations (eg, EU-RHAB, ACNS0333, Medical University of Vienna, DFCI, Head Start, HIT-SKK92).^{44–50} Controversy exists about the role of the extent of surgery, as some investigators describe a gross total resection (GTR) as a favorable prognostic factor,^{4,12,45,51–55} while others cannot confirm any statistically significant influence.^{44,56–58} The potential role of neoadjuvant chemotherapy in unresectable tumors awaits evaluation. A further, thus far unresolved, question is the role of radiotherapy in consolidation, ie, whether it may be postponed using intrathecal chemotherapy or at best replaced by HDCT.

Table 2 provides an overview of the results of inconsistently treated larger cohorts of ATRT such as phase III trials and demonstrates advantages and disadvantages. As results of these have been published several years ago, trial design and results are not the emphasis of this review. Instead, our aim is to focus on lessons learned from conventional treatment and to outline open tasks for future treatment approaches.

Multimodal Treatment Approaches in the Management of ATRT

Implementation of institutional multimodal approaches has remarkably improved the prognosis of selected cohorts of ATRT in 2-y and 5-year overall survival to almost 70%.^{4,56} It remains at 30–40% or less in unfavorable cohorts, including those with initial metastatic disease and age below 1 year.^{4,45,57}

The role of radiotherapy has been the subject of much debate, as it has been shown to be effective,^{12,51,58,60} while adverse effects on the physical and cognitive development, as well as severe complications such as radiation necrosis,^{61–65} have raised the question of viable alternatives.

Current investigations suggest that radiotherapy in younger children (<3 years) may be beneficial for tumor control⁶⁶ but bears the risk of disastrous late effects at this vulnerable age.⁶⁷ Sequencing RT to the end of treatment may be a solution to allow for further maturation of the CNS and thus reducing harm.⁶⁵ Studies by Yamasaki et al and Yang et al indicate that early vs late RT does not affect the treatment success in children below 3 years.^{58,66,68–71} Current evidence indicates that proton beam therapy (PBT) with doses equivalent to photons may achieve comparable results in terms of local tumor control and overall survival while potentially saving patients from toxicities, ie, symptomatic radiation necrosis. Still, a significant risk of distant treatment failures remains, ie, outside the former field of radiotherapy.^{69,70,72,73}

Table 2 Overview of the Results of Clinical Phase III Trials and Consensus Regimens Exclusively in ATRT

Phase	Title and Identifier	PI	Completion Date	Treatment	Results Published	Comments	Reference	Topic
III	Children's Oncology Group Trial ACNS0333 (NCT00653068)	A. Reddy	August, 2023	Maximal safe tumor resection 2 cycles induction 3 cycles consolidation plus RT (sequence dependent on age and tumor localization) 4-year Follow up period	65 patients 4-year EFS 37% 4-year OS 43% < 36 months 4-year EFS 35% (improved compared to historical cohort) 3 toxic deaths ≥ 36 months 4-year EFS 48% 4-year OS 57% 1 toxic death 4-year OS 49% for "RT before consolidation" 4-year OS 48% for "consolidation before RT" 50% treatment failure (16/33 local failure)	Two treatment arms, but non-randomized Challenging cohort (83% younger than 36 months, 30% younger than 12 months, 37% with M+, 38% with GTR) Improved survival for patients younger than 36 months High amount of safety relevant toxicity No information on cognitive and psychological late effects	[45]	Radiotherapy and HDCT

(Continued)

Table 2 (Continued).

Phase	Title and Identifier	PI	Completion Date	Treatment	Results Published	Comments	Reference	Topic
II	Multi-institutional Phase II clinical trial to test efficacy or aggressive multimodality approach for children with newly diagnosed ATRT (DFCI)	S N Chi	May, 2008	Maximal safe tumor resection Modified IRS-III including intrathecal chemotherapy: Pre-RT chemotherapy Chemoradiation post-RT induction Maintenance therapy Continuation therapy	20 patients 2-year PFS 53% (\pm 13%) 2-year OS 70% (\pm 10%) 5-year OS 49.6% 10-year OS 44.7% Median survival 5.2 years 58% objective response to induction therapy 38% response to RT 8 long-time survivors (median FU 14.7 years; median 1.9 years at diagnosis, localized disease, RT, 6/8 with upfront GTR/STR) Succumbed (10): 5/10 M+, 7/10 with upfront biopsy/partial resection 40% refractory or recurrent disease (max 4 years of data reporting) 1 toxic death (pneumococcal sepsis)	Prospective multicenter trial, single arm, Beneficial cohort (Median age at diagnosis 26 months (60% younger than 36 months, 20% younger than 12 months), 30% with M+, 50% with GTR) Short follow up period Small cohort size	[56,59]	Radiotherapy, intrathecal chemotherapy

registry	The Canadian Paediatric Brain Tumor Consortium experience (CBTC)	L. Lafay-Cousin		<p>Maximal safe tumor resection 10/50 in palliation 22/40 conventional chemotherapy (baby brain, IRS III like, ICE regimens, 9/22 anthracycline based) 18/40 HDCT (Head start, Triple Carboplatin/Thiotepa, MTX induction + Triple Carboplatin/Thiotepa) 9/40 intrathecal chemotherapy 21/40 radiation</p>	<p>50 patients Projected 2-year OS 36.4% ± 7.7 Median time of survival 13.5 months 75% with progress/relapse (median time to progression 5.5 months) Significant prognostic risk factors: GTR: 2-year OS (GTR) 60% ±12.6) 2-year OS (non GTR) 21.7% ± 8.5, p = 0.03 HDCT: 2-year OS (HDCT) 47.9% ±12.1 2-year OS (non HDCT) 27.3% ± 9.5, p= 0.036 Not-significant factors: Age: median survival time: 9.6 months in patients < 12 months 17.4 months in patients 12–36 months: 19.1 months in patients > 36 months: Upfront radiation no survival benefit! 6/12 survivors without radiation</p>	<p>Multicentric retrospective analysis Small cohort (n=50) Unfavorable cohort (median age at diagnosis 16.7 months, 76% younger than 36 months, 38% with M+, 30% with GTR, 2 synchronous MRT, 3 with diagnosed GLM) Best overall response not evaluated/not reported (radiographic CR, PR, SD etc) 75% refractory/ recurrent disease (max. 32 months) Multicenter Neuropsychological results in long-time survivors published three years later</p>		
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Table 2 (Continued).

Phase	Title and Identifier	PI	Completion Date	Treatment	Results Published	Comments	Reference	Topic
Registry	EU-RHAB	M C. Frühwald	May, 2023	Maximal safe tumor resection Anthracycline based chemotherapy (9 to 12 courses) Age-dependent radiotherapy (no RT in < 18 months) HDCT (Carboplatin/Thiotepa + ASCT) on physician's choice	143 patients 5-year OS 34.7 ± 4.5% 5-year EFS 30.5 ± 4.2% Median follow up 49.9 months 64% relapse or progression (mainly locally) 57% died due the time of analysis 3 cases of secondary malignancies (AML) No toxic death Independent significant prognostic factors: Age at diagnosis (< 1 year vs ≥ 1 year) DNA methylation subtype (TYR vs non-TYR) Dependent significant prognostic factors: Synchronous tumor (yes vs no) Metastases (M0 vs M+) Radiotherapy (yes vs no) Complete remission (yes vs. no) GLM (yes vs no)	Multicentric retrospective analysis Unfavorable cohort (median age at diagnosis 29.5 months, 86% younger than 36 months (35% younger than 12 months, 51% 12–35 months), 30% with M+, 34% with GTR) Large cohort Genetic and molecular information available	[4]	

Several international centers recommend the addition of intraventricular chemotherapy (IVCT) to conventional chemotherapy to avoid radiotherapy (EU-RHAB, DFCI-AT/RT, and MUV),^{46,50,56,74,75} as IVCT in metastatic ATRT has significantly improved 2-year OS (65% vs 17.3%) as suggested by a meta-analysis.⁵⁰ The risk of severe encephalopathy, mainly leukoencephalopathy, due to accidental intraparenchymal methotrexate instillation^{76,77} or diapedesis of MTX in ventricular obstruction⁷⁸ is a significant risk. There are no controlled or randomized phase III trials investigating the effect of IVCT in ATRT, thus evidence remains contentious. Currently recruiting clinical trials use the intrathecal route to administer experimental agents such as CAR-T cells, immunovirus, or immune-radiotherapy (see below).

Multimodal approaches including high-dose chemotherapy with autologous stem cell rescue (HDCT/ASCT) are associated with a significantly improved survival, especially in the first and second year from diagnosis.⁷⁹ HDCT may be considered an efficient treatment approach in children, who need bridging therapy until RT is feasible.⁴⁵ Reports from the Canadian Brain Tumor Consortium and the Head Start III trial describe encouraging success of radiotherapy-withholding HDCT in patients younger than 36 months (ie, prolonged survival even in patients with initial metastatic disease).^{47,54} As infants are an extremely challenging cohort,⁸⁰ appropriate treatment regimens minimizing toxicity are urgently needed. The significant therapeutic successes of ACNS0333 and CCG99703 were unfortunately accompanied by substantial treatment-related toxicities and even toxic deaths (n=2 CNS necrosis) predominantly in patients below 36 months.⁴⁵

The *Canadian Pediatric Neuro-Oncology Standards of Practice* features both CCG 99703 and ACNS0333 protocols as first-line therapy of ATRT, while radiotherapy instead serves as an additional option as a “physicians’-decision-only” approach. Recommendations reflect expert opinions and consensus statements rather than evidence-based data.⁴³ Data proving the superiority of HDCT approaches versus conventional chemotherapy and radiotherapy-based approaches, ie, derived from randomized controlled clinical trials are missing. In the largest cohort of consistently treated ATRT (n=143, EU-RHAB) HDCT did not significantly improve survival. Here, 5-year OS and 5-year EFS of 35% and 30% were achieved by multimodality treatment approaches based on physicians’ choice, according to a European consensus strategy (based on DFCI and IRS III protocols). This consisted of surgery, anthracycline-based chemotherapy, and age-dependent administration of RT (no RT in children < 18 months) or HDCT instead. In total, 34 patients were treated with consolidation therapy in the form of HDCT/ASCT (carboplatin, thiotepa), only. However, multivariate analysis could not demonstrate any significant influence of RT-withholding approaches.⁴

As baseline conditions and survival in this cohort were comparable to those of the cohort within ACNS0333 (see Table 2), we suggest that the question whether HDCT featuring protocols (eg, ASCNS0333,⁴⁵ Canadian Brain Tumor Consortium⁸¹) are preferable over those including radiotherapy (eg, DFCI⁵⁶ and EU-RHAB⁴) deserves validation in a randomized setting in larger cohorts. The ongoing SIOPE ATRT01 trial (Eudra-CT 2018-003335-29) investigates the impact of isolated HDCT with autologous stem cell rescue as consolidation therapy.

Remaining High Risk for Relapse and Refractory Disease

Even though institutional multimodal treatment approaches have improved survival probability, a significant risk of recurrent or refractory disease remains. Progressive disease still occurs in 40–75%.^{4,45,57} There is no international consensus on how to treat patients with r/r ATRT. This situation is the focus of Phase I and II clinical trials. Very limited data exist on real world experience such as individual experimental treatment approaches.

Therapeutic Approaches Supported by Genetic Analyses: MAPPYACT, eSMART, INFORM

To overcome limitations of conventional therapy, “one size does not fit all”, targeted anti-cancer medicine studies incl. MAPPYACT, eSMART, MATCH, and INFORM attempt to identify genetic and molecular targets for individualized treatment approaches.

The evidence available from these approaches reveals two major problems: On the one hand, apart from *SMARCB1* no other repetitive cancer-specific genetic alterations have been detected (INFORM⁸² and MAPPYACT⁸³). Identification of suitable genetic biomarkers to deduce specific treatment targets thus remains challenging.

On the other hand, albeit potential drug targets incl. CDKs, aurora kinase A, immune checkpoint receptors, and ligands are highly expressed in selected ATRT, targeting these induced significant treatment responses in very few ATRTs

only.⁸³ Expression profiles alone are thus not sufficient to predict response to single agent targeted therapy. Deciphering pathomechanisms of resistance to the EZH2 inhibitor tazemetostat, Kazansky et al identified specific acquired mutations in resistant rhabdoid tumor cell lines that appear to be responsible for treatment failure. At the same time, these analyses demonstrated potential targets for synthetic lethality (in cell lines and xenograft mouse model).⁸⁴ We discuss the use of synthetic lethality in the setting of clinical trials below.

Remaining Challenges in r/r ATRT

Randomized phase III trials may help consolidate evidence for risk factors and treatment elements. Future research will have to target the role of conventional compounds such as repurposed cytostatics (incl. IVCT) but also radiotherapy modalities, as well as smart combination elements.⁸⁵ The concept of future clinical trials depends on international (drug) availability and the capacity to recruit suitable cohorts. The fortunately increasing number of long-time survivors highlights the need to attend to and study the physical and mental development as well as aspects of quality of life in survivors. Treatment approaches for the special cohort of infants necessitate the highest level of diligence in designing safe and efficient therapeutic trials.⁸⁰

Potential Targeted Therapeutic Approaches to ATRT

Table 3 provides an overview of potential therapeutic targets investigated in ongoing clinical trials. These approaches target either cell cycle activating pathways and checkpoints, DNA-repair or histone modifications, the immune system–cancer interaction, or offer new options of radioimmunology. Figure 1 illustrates experimental agents and their targets within the cell cycle. Ongoing clinical trials using agents targeting the immune system-tumor cell interplay are displayed in Figure 2.

Targeting the Epigenome

Modifying DNA Methylation

Decitabine is a cytosine analogue and potent DNA-methyltransferase inhibitor that induces DNA hypomethylation, thus uncovering endogenous retroviral epitopes, which serve as targets for immune attack.^{87–89} This effect has been used in cancer with hypermethylated DNA profiles, including AML and MDS.⁹⁰ As rhabdoid tumors are an epigenetically driven disease with DNA-methylation-based subtypes (ATRT-TYR and -SHH present DNA-hypermethylation²⁴), the use of hypomethylating agents appears to be a promising approach. Retrospective analyses of decitabine in 22 patients with r/r rhabdoid tumors reported radiologic responses in 27% (total n=6, 4= ATRT).⁸⁵ Complete remission was achieved in one patient, who received further conventional chemotherapy, while in most patients, response was detected in isolated foci, only. As radiographic response was associated with prolonged time to progression and overall survival,⁸⁵ decitabine deserves further evaluation, eg, as bridging therapy in patients with r/r disease. Three current clinical trials employing decitabine in combination with a) immune checkpoint inhibitors (NCT05320640), b) gene-modified T Cells (NCT02650986), and c) a combination with pembrolizumab and hypofractionated radiotherapy (NCT03445858; primary CNS tumors not included) recruit children with solid tumors. Further approaches for children with neuroblastoma, rhabdomyosarcoma, Ewings sarcoma, and osteosarcoma combined decitabine with dendritic cell vaccination (NCT01241162).

Modifying Histone-Methylation

In *SMARCB1* or *SMARCA4* deficient tumors, overexpression of EZH2 leads to increased levels of H3K27me3 in promoter regions of tumor suppressor genes. It is thus a target for EZH2 inhibitors.^{91,92} In ATRT, targeted inhibition of EZH2 leads to repression of the Cyclin D1-E2F axis and thus reduced proliferation and apoptosis.^{92,93} Moreover, as recently reported, EZH2 deficiency correlates with increased origins of DNA replication in repressed motor regions via H3K27me3 loss.⁹⁴

The orally applicable specific EZH2 inhibitor tazemetostat has been FDA approved for the treatment of inoperable, metastatic epithelioid sarcoma in patients > 16 years.⁹⁵ Safety and efficacy have been shown in *SMARCB1*-negative tumors, including ATRT.^{96–98} Results from the MATCH study reported prolonged disease stabilization as the best overall

Table 3 Overview of Ongoing Clinical Trials Phases I and II Including Patients with ATRT

Inhibitor Group		Inhibitor	NCT Number	Phase	Study Completion
Immunotherapy	CAR T cells	B7-H3-specific	NCT04185038	I	2041
		C7R-GD2	NCT04099797	I	2039
		HER2-specific	NCT03500991	I	2039
		EGFR806-specific	NCT03638167	I	2040
	Immunovirus therapy	HSV G207	NCT03911388	I	2025
Immune-checkpoint inhibitors	PD-1/CTLA4-Inhibitors	Nivolumab + Ipilimumab + Tazemetostat	NCT05407441*	I/II	2029
	PD-1/CTLA4-Inhibitors	Nivolumab + Ipilimumab	NCT04416568*	II	2025
	PD-L1-Inhibitors	Atezolizumab + Tiragolumab	NCT05286801*	I/II	2025
	PD-1/HDI	Nivolumab + Entinostat	NCT03838042	I/II	2025
Cell cycle inhibitors	CDK4/6 inhibitor	Ribociclib + Gemcitabine Ribociclib + trametinib	NCT03434262	I	2025
		Ribociclib + TOTEM	NCT05429502	I/II	2028
		Abemaciclib (+RT)	NCT02644460	I	2024
		Abemaciclib (+ CTx)	NCT04238819	I/II	2028
		Palbociclib	NCT03709680**	I/II	2025
Targeted therapy	BRD9-degrader	FHD-609	NCT04965753**	I	2025
Targeted tumor cell radiotherapy	Phospholipid drug conjugate	CLR 131	NCT03478462	I	2024
	Radiolabeled monoclonal antibody	Iodine I131 MOAB 8H9	NCT00089245	I	2025
Protein kinase inhibitor	BCR-ABL tyrosine kinase inhibitor	Dasatinib	NCT00788125	I/II	June 20, 2023
	mTOR inhibitor	Sirolimus	NCT02574728	II	2024
	Aurora Kinase A inhibitor	Alisertib	NCT02114229*	II	2027
Replication	Folic acid analogue	Methotrexate	NCT02905110	Early Phase I	November 2023
Conventional therapy	Conventional chemotherapy, radiotherapy	Risk-adapted therapy	NCT00602667 (infants) NCT00085202 (children)	II	April 2026, December 2023
	HDCT/ASCT, radiotherapy	Controlled, randomized	EudraCT 2018-003335-29*	III	June 2029

Notes: *ATRT exclusive (including rhabdoid tumors, *SMARCB1*-deficient tumors); ***SMARCB1*-deficient tumors focusing on soft tissue tumors.

Abbreviations: AML, acute myeloid leukemia; ASCT, autologous stem cell transplantation; ATRT, atypical teratoid/rhabdoid tumor; CAR T cell, chimeric antigen receptor T cell; CD4T, CD4+ T cell; CDK, cyclin dependent kinase; cfDNA, cell free DNA; CNS, central nervous system; CNV, copy number variations; CRINET, cribriform neuroepithelial tumor; CSF, cerebrospinal fluid; CT, chemotherapy; ctDNA, circulating tumor DNA; CTLA4, cytotoxic T-lymphocyte associated protein 4; DFCI, Dana-Farber Cancer Institute; DIPG, diffuse intrinsic pontine glioma; DNA, deoxyribonucleic acid; DZNeP, 3-Deazoplanocin; EED, embryonic ectoderm development; eMRT, extracranial, extrarenal malignant rhabdoid tumor; EU-RHAB, European Rhabdoid Registry; EZH2, enhancer of zeste homolog 2; FDA, Food and Drug Administration; GEMM, genetically engineered mouse model; GLM, germ line mutation; GTR, gross total resection; HDCT, high-dose chemotherapy; HDI, histone deacetylase inhibitor; INI1, integrase interactor 1; IVCT, intraventricular chemotherapy; LGDIT, low grade diffusely infiltrative tumor; M, metastatic; MDM2, murine double minute 2 homolog; MDS, myelodysplastic syndrome; MEK, mitogen-activated protein kinase kinases; MOAB, monoclonal antibody; MRD, minimal residual disease; MTX, methotrexate; MV-NIS, measles virus (MV) expressing human thyroidal sodium iodide symporter (NIS); oHSV, oncolytic herpes simplex virus; OS, overall survival; PARR, Poly (ADO-ribose) polymerase; PBT, proton beam therapy; PD-1, programmed death protein 1; PD-L1, programmed death ligand 1; PDOX, patient derived orthotopic xenograft; PFS, progression free survival; PR, partial remission; PRC2, polycomb repressive complex; PXA, pleomorphic xanthoastrocytoma; RB, retinoblastoma protein; r/r, recurrent or refractory; RT, radiotherapy; RTK, rhabdoid tumor of the kidney; RTPS, rhabdoid tumor predisposition syndrome; SD, stable disease; SMARCA4, SWI/SNF related matrix associated, actin dependent regulator of chromatin, subfamily a, member 4; SMARCB1, SWI/SNF related matrix associated, actin dependent regulator of chromatin, subfamily b, member 1; SWI/SNF, switch/sucrose non-fermentable; TME, tumor microenvironment; TMB, tumor mutational burden.

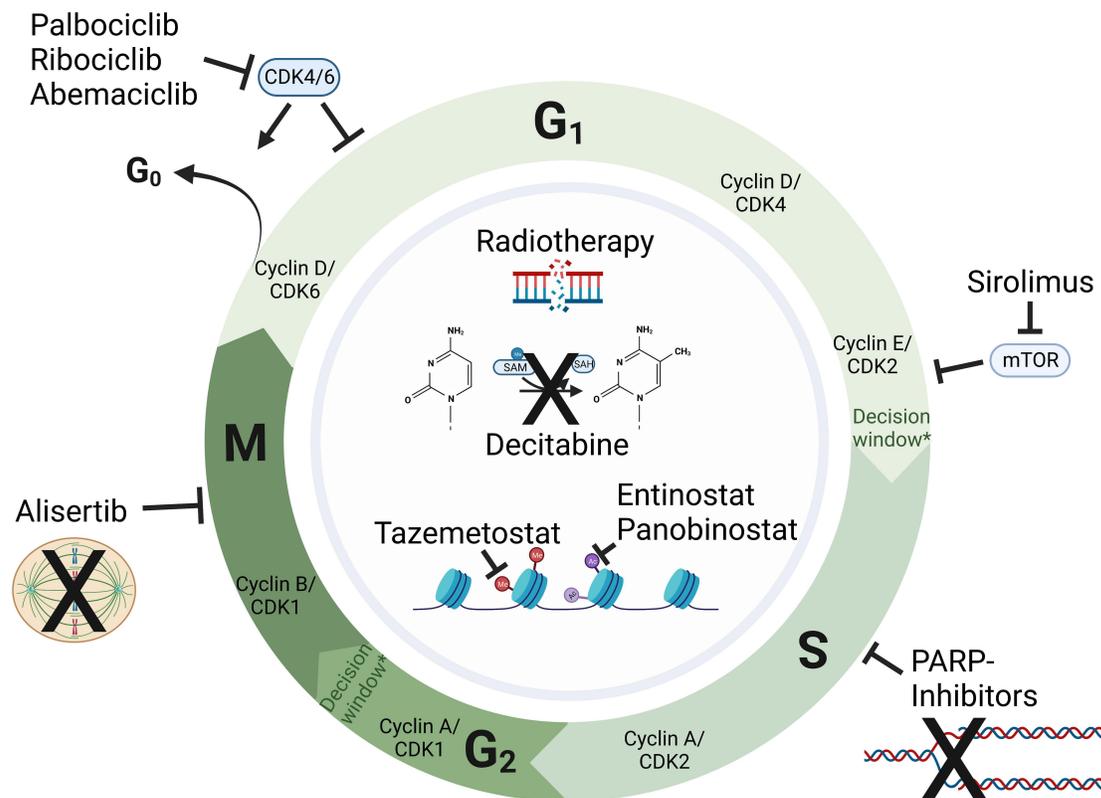


Figure 1 Novel agents for ATRT therapy in clinical phase I and II trials - correlation to the tumor cell cycle. Arrows with even tips indicate “inhibition”, arrows with sharp tips indicate “activation”, fading arrows with sharp tips indicate “cell cycle exit into G₀ phase”. Therapeutic approaches located in the center and surrounded by the grey circle affect more than one distinct phase of the cell cycle in the form of modified gene repression by epigenetically active drugs (decitabine (DNA-hypomethylation), tazemetostat (H3K27me3), entinostat and panobinostat (HDACI)), or DNA damage dependent cell cycle exit in G₁ and G₂ (radiotherapy induced double strand breaks). *Reaching and passing decision windows is driven by appropriate cyclin/CDK levels: G₁/S transition is regulated by Cyclin D/CDK2/4/6, while Cyclin A/B/CDK1/2 affect G₂/M transition.⁸⁶ A mitotic entry commitment point, a mitotic exit commitment point, or a S phase entry commitment point regulated by DNA damage, DNA replication stress, or spindle assessment are not shown in this figure. Created with Biorender.com.

response in 1 of 8 ATRT (13.7 months PFS) with confirmed loss-of-function alterations in the *SMARCB1* gene.⁹⁹ Ongoing clinical trials investigate the safety and efficacy of tazemetostat, as a single agent (NCT02601950) or in combinatorial approaches (NCT04537715, NCT05407441). NCT05407441 investigating the combination of tazemetostat with the immune-checkpoint inhibitors nivolumab (PD1 inhibitor) and ipilimumab (CTLA4 inhibitor) in INI1-negative/*SMARCA4*-deficient tumors has recently opened recruitment (estimated study completion 02/2029). Prescribing tazemetostat needs to consider the risk of development of secondary lymphoma described by Chi et al⁹⁶ as well as the development of drug resistance.¹⁰⁰ Trying to explain underlying mechanisms, Shinohara et al reported an increased expression of the EZH2 homologue EZH1 by depletion or chemical inhibition of EZH2 in cell lines and mouse xenograft models of rhabdoid tumors. These authors suggested decreased or repressed EZH2 levels as a cause for overexpression of EZH1, which in turn compensates the loss of function of EZH2. Targeting both EZH1 and EZH2, reduced H3K27me3 accumulation (in *CDKN2A*) as well as cell growth control.¹⁰¹ More recently, Kazansky et al proposed combined epigenetic therapy to overcome clinical resistance to EZH2 inhibition. The authors provide potential key requirements for the effective use of tazemetostat in *SMARCB1*-deficient tumors: (1) Intactness of drug binding sites of EZH2 and subsequent pathway components (including transcription factors and co-activators). This means that chromatin remodeling complexes must act on tumor suppressor loci that were aberrantly repressed by PRC2, and these must be upregulated for a response to tazemetostat. For example, mutations in *CDKN2A* and *CDKN2B* will lead to decreased *RB1* activation and subsequent reduced cell cycle inhibition at G₁/S transition. 2. Targeting the non-enzymatic core PRC2 subunit EED, as well as combining additional drugs affecting the cell cycle beyond the G₁/S checkpoint, could help overcome tazemetostat resistance.⁸⁴

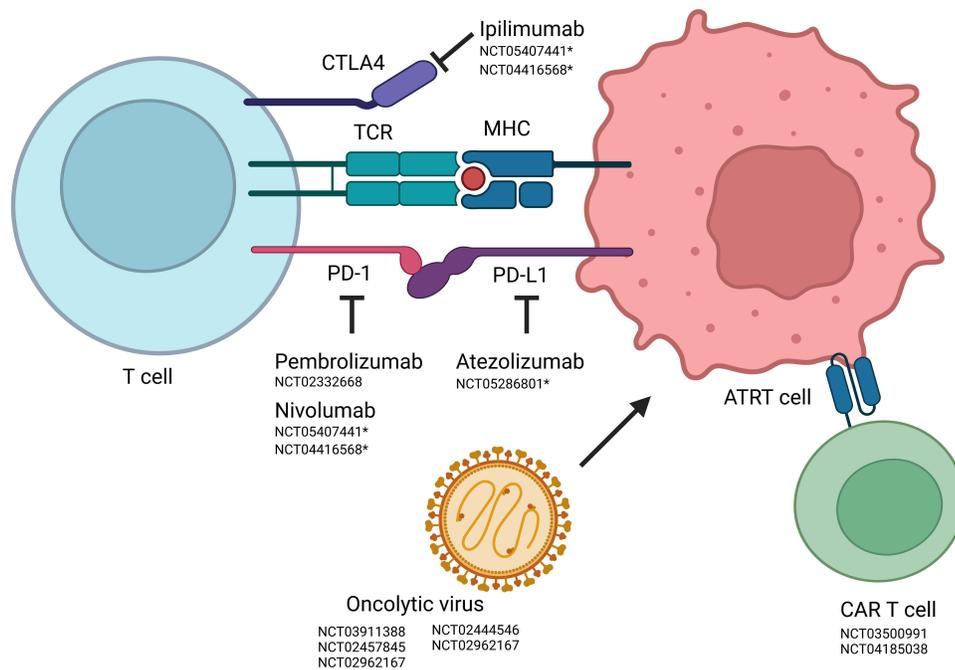


Figure 2 Novel targets affecting the interaction between ATRT tumor cells and the immune system. Structures investigated in clinical phase I and II trials targeting the interaction of T and ATRT tumor cells. Current clinical trials are provided for each agent in form of the NCT identifier. Trials including *SMARCB1/SMARCA4*-deficient entities only (focus on ATRT) are highlighted with *. Arrows with even tips indicate “inhibition”, arrows with sharp tips “uptake in”. Created with Biorender.com.

Abbreviations: CTLA4, cytotoxic T-lymphocyte-associated protein 4; TCR, T cell receptor; MHC, major histocompatibility complex; PD-1, programmed death protein 1; PD-L1, programmed death ligand 1.

Intratumor heterogeneity may provide a further explanation for drug resistance, as Qi et al supposed when describing regions of repressed *EZH2* in residual tumors. It cannot be determined whether this induces or is affected by therapy resistance.¹⁰²

Modifying Histone Acetylation

Recent data indicate the effect of panobinostat on histone-H4 acetylation leading to encouraging treatment stabilization when used as maintenance therapy.¹⁰³ Results of a current trial are pending, and the first-line pre-treatment is likely to substantially affect treatment response. INFORM 2 employs the histone-deacetylase inhibitor entinostat in combination with the immune checkpoint inhibitor nivolumab (NCT03838042, including ATRT).

Tyrosine Kinase Inhibition

Aurora Kinase A Inhibitors

The aurora kinase A inhibitor alisertib leads to impairment of mitotic spindle assembly (as reviewed in¹⁰⁴). A phase II study (SJATRT) investigated a cohort of rhabdoid tumors. Results for stratum A1 (recurrent or progressive ATRT) were published in 2023: Intermittent objective responses to treatment were achieved in 9/30 patients (n=8 stable disease (SD), n=1 partial remission (PR) in the 12-week assessment). A total of 25 patients eventually developed progressive disease. Five patients achieved a follow-up time of more than 50 weeks (4/5 ATRT-TYR, 1/5 ATRT-MYC, all > 1 year of age at enrolment). It should be stressed that two of them received alisertib for less than 2 months and never responded to treatment. All survivors received further salvage therapy. The authors suggest a potential role of alisertib in bridging or palliative care, only.¹⁰⁵ Anecdotally, long time remission for 3 years following alisertib was reported in an infant with ATRT-TYR in the treatment of its 2nd relapse.¹⁰⁶

BCR-ABL Tyrosine Kinase Inhibitor

The BCR-ABL tyrosine kinase inhibitor dasatinib was developed for the treatment of Philadelphia-chromosome-positive leukaemia¹⁰⁷ and investigated in a phase I/II clinical trial (NCT00788235) in children with metastatic or recurrent

malignant solid tumors including brain tumors as part of a combined anti-cancer treatment approach. According to the results published on *clinicaltrials.gov*, 6 out of 7 patients did not complete protocol treatment and succumbed due to disease (<https://clinicaltrials.gov/study/NCT00788125?term=NCT00788125&rank=1&tab=results>; no information on actually included tumor types, ATRT not an exclusion criterion).

Targeting the CDK4/6/Cyclin D/RB Pathway

During the early G₁ phase, increased levels of Cyclin D as well as Cyclin-dependent Kinases (CDK) 4 and 6 are needed to maintain the cell cycle and to prevent cell cycle exit (as reviewed in⁸⁶). Targeting CDK4/6 inhibitors aims at inducing cell cycle arrest in G₀/G₁ before reaching the G₁/S decision window.

Ribociclib penetrates the blood–brain barrier and is thus an interesting drug in CNS tumors.¹⁰⁸ Clinical phase I trials report weak therapeutic success. In 2017, a phase I trial demonstrated the safety of CDK4/6 inhibitors, but best overall response was stable disease in 2 of 13 patients with recurrent ATRT.¹⁰⁹ Recently published data report no objective responses in 18 children with recurrent or refractory malignant CNS tumors (no ATRT included) treated with a combination of ribociclib and everolimus. All patients, who completed follow-up, experienced disease progression.¹¹⁰ Investigating cell lines of adult CNS tumors, the combination of CDK4/6 inhibitors and other commonly used cytotoxic anti-neoplastic drugs could not provide any enhanced therapeutic success, as Jin et al reported in 2019.¹¹¹ Disease stabilization over a period of 32 to 48 weeks, as described by Georger et al,¹⁰⁹ still represents a remarkable achievement in the treatment of recurrent or relapsed ATRT.

The ongoing clinical trials NCT03434262 and NCT05429502 evaluate ribociclib as part of a combination therapy (combining with nucleoside analogues, inhibitors of the Hedgehog pathway, MEK inhibitors, or commonly used cytotoxic antineoplastic drugs such as topotecan and temozolomide (TOTEM)).

The phase I/II trial NCT02644460 evaluated abemaciclib as a single agent in children with CNS tumors and especially DIPG (ATRT not excluded) NCT04238819 (ATRT not excluded) looked into a combination with other anti-cancer treatment in pediatric patients with relapsed and refractory solid tumors.

Data from the Pediatric Brain Tumor Consortium study PBTC-042 investigating palbociclib as a single agent in patients with progressive CNS tumors showed limited treatment response, as none of the tumors responded and up to 90% progressed on treatment, although 80% of the tumors were tested positive for *RB* status before initiating treatment.¹¹² A phase I/II study currently investigated palbociclib combined with chemotherapy (irinotecan, topotecan, temozolomide, or cyclophosphamide) in children with recurrent or refractory solid tumors including eMRT (NCT03709680).

PARP-Inhibitors and DNA-Repair

Poly(ADP-ribose) polymerase (PARP) are multifunctional enzymes involved in DNA damage repair and maintaining genome stability. DNA damage leads to recruitment of PARP and DNA repair enzymes. PARP inhibitors prevent dissociation of the PARP-complex off the DNA strand thus inducing replication fork collapse.^{113,114}

Preclinical trials found decreased cell growth, reduced colony formation, cell cycle arrest, and apoptosis due to accumulation of DNA damage in the ATRT cell lines BT16 and MAF37 following treatment with the PARP inhibitor rucaparib. Prolonged survival in xenografted mice and potential synergistic effects increasing radiosensitivity suggest a potential therapeutic use.¹¹³ Until this day, there is no clinical trial confirming the pre-clinical effect of PARP inhibitors in ATRT, or rhabdoid tumors in general. However, the safety of the PARP inhibitors niraparib, talazoparib, and olaparib is under investigation in clinical trials NCT04544995 (no CNS-Tumors, eMRT not excluded), NCT023492793 (published for Ewing sarcoma, no objective response),¹¹⁵ and NCT01858168 (Ewing sarcoma and rhabdomyosarcoma), respectively. Despite dismal treatment responses upon single agent use, recent investigations in pediatric solid tumors provide information on beneficial effects of PARP inhibitors combined with conventional chemotherapy, ie, alkylating agents or topoisomerase inhibitors.¹¹⁶

Immunotherapy Approaches

Tumor immune microenvironment (TME) modulation is a successful approach in cancer treatment.¹¹⁷ Grabovska et al described immune cell infiltration in multiple pediatric brain tumors, including 229 ATRT and demonstrated distinct

immune cell infiltration profiles in the three ATRT subtypes (ATRT-MYC: CD8T, monocytes, Treg, B-cells, eosinophils; ATRT-SHH: CD8T, Treg, B-cells, monocytes, eosinophils; ATRT-TYR: Treg, B-cells, monocytes, eosinophils, NK cells, CD4T, neutrophils). The ATRT-TYR subgroup demonstrated infiltrations with NK- and CD4T-cells, which were neither found in ATRT-MYC nor -SHH.¹¹⁸ Several approaches integrating modulation of the TME in ATRT management have been developed (see Table 3).

Immune Checkpoint Inhibitors

Programmed death protein-1 (PD-1), programmed death ligand 1 (PD-L1) and cytotoxic T-lymphocyte associated protein 4 (CTLA4) are part of the tumor cell immune escape, as T cell mediated immune response is downregulated by PD-1-PD-L1 interaction¹¹⁷ or by activating CTLA4 (reviewed in¹¹⁹). Immune Checkpoint inhibitors block PD-1, PD-L1, or CTLA4 to sustain T cell mediated immune response against tumor cells.

The PD-1 inhibitor pembrolizumab has shown encouraging responses (partial response) in patients with confirmed PD-L1 expression in the tumor tissue, including 1 eMRT. Albeit PD-1 expression was confirmed in 95% of cases, 72% experienced progressive disease while on treatment. ATRT was not included in this investigation (NCT02332668).¹²⁰ Similar results were described for the PD-1 inhibitor nivolumab in a cohort of recurrent or refractory pediatric CNS tumors: transient radiographic responses were seen in 30%. Disease progression was found in all patients but occurred earlier in patients with negative PD-L1 expression compared to patients with positive PD-L1 expression (not statistically significant). The investigators suggest there might be a correlation between tumor mutation burden (TMB) and response to immune checkpoint inhibitors since nivolumab reached best efficacy in the patient with highest TMB.¹²¹

In a phase I/II study, the PD-L1 inhibitor atezolizumab elicited an objective response in 1 of 6 patients with relapsed extracranial malignant rhabdoid tumors. This patient exhibited high PD-L1 expression and experienced several months without tumor progression. eMRT with no or low PD-L1 expression did not respond to atezolizumab treatment.¹²² This effect has yet to be confirmed in ATRT and is under investigation in an ongoing clinical trial (NCT05286801, including ATRT) in a combined drug approach including tiragolumab.

Ongoing clinical trials in r/r high-risk pediatric cancer investigating immune checkpoint inhibitors in pediatric *SMARCB1*-deficient tumors mainly focus on the PD-1 inhibitor nivolumab in combination with the CTLA4 inhibitor ipilimumab with or without tazemetostat (NCT05407441, NCT04416568, both including ATRT), as well as in combination with the HDI entinostat (NCT03838042, including ATRT).

CAR T-Cells

Chimeric antigen receptor (CAR) T cells may target the epitopes of B7-H3, C7R-GD2, HER2, and EGFR806, which are specifically or highly overexpressed on CNS tumor cell surfaces.¹²³ Target selection has been predominantly driven by expression in adult glioblastoma.¹²⁴

Safety and feasibility of locoregional infusion of HER2-specific CAR T cells have been demonstrated in recurrent or refractory adult and pediatric CNS tumors (NCT03500991, including ATRT). Repeated administration of HER2-specific CAR T cells either into the tumor cavity or the ventricular system was safe in the first three patients enrolled. Immune activation was measured in the form of elevated cytokine (eg, CXCL10 and CCL2) levels in the cerebrospinal fluid (CSF).¹²⁵ However, assessment of treatment response and further safety profiles are not yet available.

B7-H3 is elevated in ATRT.¹²³ Following CAR T cell treatment, ATRT xenografted mice revealed a tumor cell purging effect.¹²⁶ Safety and efficacy of B7-H3-specific CAR T cells in children are currently being investigated in a phase I clinical trial (NCT04185038, including ATRT). Published results of three patients with DIPG showed the feasibility and safety of intraventricular B7-H3-specific CAR T cell administration in children with CNS tumors, with one patient providing a radiographic response for more than 12 months.¹²⁷

Glioblastomas mainly express EGFR806¹²⁸ and has been employed as a target in clinical phase I trial NCT03638167 for recurrent or relapsed pediatric brain tumors, including ATRT.

Radioimmunotherapy

In addition to being a target for CAR-T-cells B7H3 serves as a target for immunolabeled radioligands. Employing the radiolabeled monoclonal antibody ¹³¹I-omburtamab, a radiation source can be directed at B7H3 expressing cells. A phase

I trial in children with recurrent CNS malignancies including one ATRT infused ^{131}I -omburtamab into the ventricles. The radioactive compound barely passed the blood–brain barrier and exposure to organs at risk was low. Patients suffering from neuroblastomas experienced improved survival, while neither tumor remission nor prolonged disease stabilization was reached for the only patient with ATRT.¹²⁹ Ongoing clinical phase I trials such as NCT03478462 and NCT00089245 investigate the I-131 monoclonal antibody 8H9 and CLR 131 in children with recurrent CNS malignancies (ATRT not excluded).

Immunovirus Therapy

Modified oncolytic Herpes simplex virus-1 (oHSV-1) is capable of selectively infecting tumor cells with lasting responses. Antiviral treatment is employed as a safety insurance for healthy tissue. Anti-tumor effects are based on virus-induced oncolysis and stimulated anti-tumor immune responses. Cell surface structures including nectin (CD-111), expressed on CNS tumor cells, enable oHSV to attach to and enter tumor cells specifically.¹³⁰ It has been shown that the oHSV subtype G207 is able to switch the tumor microenvironment from immunological “cold” to “hot” by increasing lymphocyte infiltration. A phase I study of genetically engineered herpes simplex virus (HSV G207) has started in 2020 (NCT03911388). Due to a biallelic deletion of the neurovirulent gene $\gamma_134.5$ and insertion of the reporter *lacZ* into the U₃9 gene, which encodes ribonucleotide reductase inactivating ICP6,¹³¹ G207 is assumed to be non-neurovirulent, replication competent and selectively oncolytic. Safety of G207 was investigated in animal models and clinical trials recruiting adult patients. Importantly, a phase I trial, investigating G207 in pediatric malignant supratentorial tumors (glioblastoma, anaplastic astrocytoma; NCT02457845) showed convincing safety profiles and radiographic, neuropathological, or clinical response in 11 of 12 patients.¹³² An ongoing phase I trial investigates G207 in pediatric cerebellar tumors (not excluding ATRT).¹³³

Modified Oncolytic reovirus and measles virus have been investigated in recently completed clinical trials in ATRT (NCT02444546, NCT02962167). Wild-type reovirus was administered in combination with sargramostim in young patients with high-grade r/r CNS tumors (NCT0244546). In this trial, all patients (no ATRT thus far) experienced tumor progression and death of disease.¹³⁴ In the clinical phase I trial NCT02962167, modified measles virus (MV-NIS) was administered in patients with recurrent medulloblastoma or ATRT, either directly into the tumor bed during standard of care tumor resection or via lumbar puncture depending on tumor dissemination (results are pending). Oncolytic measles virus selectively infects cells with high CD46 surface expression, eg, tumor cells. CD46 is part of the complement inactivation system and therefore involved in tumor immune escape.^{130,135–137} The oncolytic effect of reoviruses is caused by tumor cell overexpression of Ras enabling active viral dsRNA translation, which is blocked in normal tissue cells.^{130,138}

Synthetic Lethality

Most novel therapeutic approaches in ATRT aim to inhibit overexpressed gene products. Due to the pronounced stability of the ATRT genome, the number of potential targets is limited. Identification of additional crucial gene products, eg, in the context of synthetic lethality, may present another approach to overcome therapeutic limitations. Synthetic lethality can be defined as lethality of cells or organisms caused by combined alterations of gene pairs, with single alterations in both maintaining viability.¹³⁹ CRISPR-Cas-9 screens identified *BRD9*, a subunit of the non-canonical *SWI/SNF* complex, as a potential target for synthetic lethality. In *SMARCB1*-deficient rhabdoid tumor cell lines *BRD9* knock-out decreased cell proliferation and viability.^{140,141} Clinical approaches evaluate the feasibility of the molecular *BRD9* degrader FHD-609 in adult and older pediatric (>16 years) patients with *SMARCB1*-deficient tumors (NCT04965753).

Preclinical Approaches to Increase Radiosensitivity

Histone Modifications Increase Radiosensitivity

Preclinical experiments attempt to induce radiation sensitivity by inhibition of histone modifying enzymes. EZH2 inhibition and treatment with the unspecific inhibitor of histone methylation DZNep increased the vulnerability of ATRT cell lines to radiotherapy. This finding is likely due to a general restriction of DNA synthesis and cell cycle progression by affecting EF2/c-Myc and Cyclin D1/CDK6/c-Myc/RB pathways caused by targeted molecular inhibition

(shRNA) of EZH2 and the S-adenosylhomocysteine hydrolase inhibitor DZNep, which induces apoptosis and cell cycle arrest in ATRT cell lines.⁹² We would like to point out that EZH2 inhibition by tazemetostat cannot match inhibition using CRISPR CAS9 or shRNA, as the latter is much more efficient. In patient-derived orthotopic xenograft mouse models, it was demonstrated that tazemetostat significantly prolonged survival compared to non-tazemetostat, but did not improve responses to RT.¹⁰² Therefore, *in vivo* effects were less obvious than anticipated from experimental investigations.

Histone deacetylase inhibitors (HDI) effects on apoptosis and cell cycle arrest may be explained by induction of p21 transcription by HDI SNDX-275.¹⁴² The HDI entinostat (SNDX-275), was safe in a recently completed phase I trial (NCT02780804), which included refractory primary CNS tumors (no ATRT), while response to treatment was weak, as 20 of 21 patients experienced progression within the first 3 courses. Only one patient with ependymoma reached a stable disease following 28 courses.¹⁴³ The currently recruiting INFORM2 (NivEnt) trial for refractory malignancies including primary CNS tumors combines entinostat and the PD-1 inhibitor nivolumab. An arm for patients with ATRT below 6 years of age is anticipated to open once an orally available formulation for entinostat can be provided.

Induction of Proapoptotic Pathways

Small-molecule MDM2 inhibitors such as nutlin 3 and idasanutlin activate the proapoptotic *TP53* pathway and increase the effect of radiation-induced DNA damage in ATRT *in vitro*.

Consistently combining idasanutlin with irradiation significantly reduced proliferation in ATRT cell lines.¹⁴⁴ A corresponding radiographic response was shown in orthotopic xenograft mouse models treated with idasanutlin including intratumor necrosis.¹⁴⁴ Further studies are needed to assess therapeutic success in clinical settings.

Future Directions – Discussion and Outlook

Within this review, we have given examples of the biological and clinical heterogeneity of ATRT and the associated challenges in therapy. Despite crucial improvements in therapeutic modalities and understanding of the molecular biology of ATRT results from clinical trials suggest that prognosis depends mainly on clinical characteristics such as age, metastatic disease, GLM, and extent of surgery. First attempts to classify risk groups, eg, ATRT-TYR as a positive predictor deserve further validation in a prospective fashion.

Treatment of relapsed and refractory ATRT remains an enormous challenge as single agents have not yet yielded any convincing responses in the limited number of phase I/II trials. We thus suggest that innovative, personalized treatment strategies need to include aspects of intra- and intertumoral heterogeneity. Such approaches, especially in r/r ATRT, need to respect clinical aspects including age at diagnosis and metastatic status. These will likely significantly influence the intensity and extent of conventional treatment elements (eg, fields of re-radiotherapy). Furthermore, molecular features such as genetic or epigenetic characteristics will guide individualized treatment approaches.

Rational drug combinations bear the potential to potentiate the therapeutic effects of single agents. Combinatorial approaches are currently investigated, eg, for immune checkpoint inhibitors and EZH2 inhibitors but also HDI (NCT05407441, NCT04416568, NCT05286801, and NCT03838042).

A potential model for therapeutic decision is given in [Figure 3](#). As knowledge increases rapidly, every model certainly warrants constant re-evaluation and adaptation.

The Challenge of Treating r/r ATRT

Despite first data that methylation subgroups may have prognostic significance, there remains a dearth of validated, reliable, and potent biomarkers predicting treatment response in general but also to specific targeting agents.

Several attempts have been made to correlate clinical and molecular characteristics with clinical outcome.^{4,30,32} First steps involving machine learning and artificial intelligence methods to generate prognostic algorithms have been initiated.¹⁴⁵ As this has identified prognostic treatment elements such as the extent of surgery, radiotherapy, and chemotherapy, it reproduces more or less the current knowledge obtained from retrospective analyses. We need to acknowledge these attempts as proof-of-principle approaches, which build the basis for improved and more complex AI models. These are anxiously awaited.

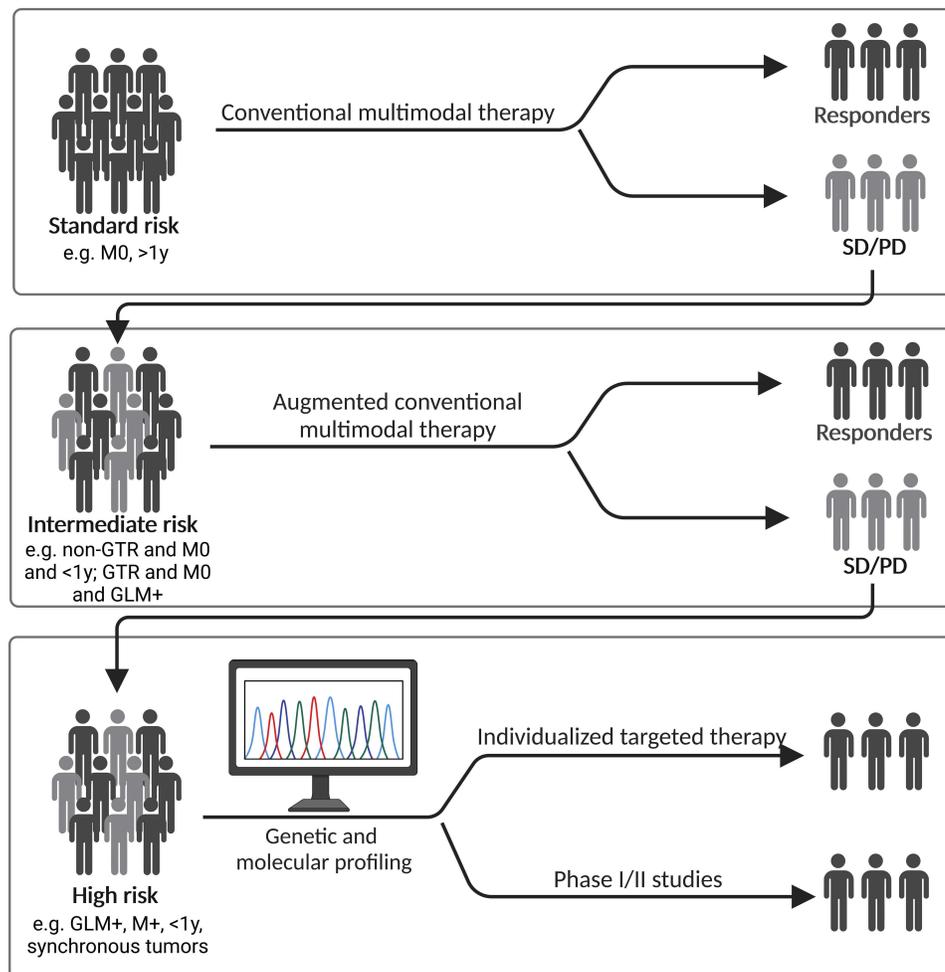


Figure 3 Risk factor-based model for treatment decision in ATRT. For standard risk patients, multimodal conventional therapy is the first line choice. Patients not achieving an objective response (SD, stable disease) and non-responders (PD, progressive disease) will be re-stratified into an intermediate risk group. Treatment of intermediate risk patients may add individual or combined novel targeting agents to conventional therapy. Delayed (SD) and non-responders (PD) will drop to a high-risk stratum. Treatment in the high-risk stratum will be individualized based on genetic and molecular profiling, and/or patients will be included in phase I/II clinical basket trials. Black characters represent responders, grey characters non-responders to current or previous treatment. Exemplary risk factors are provided for potential stratification and have to be defined by further research in form of international meta-analysis. Created with Biorender.com.

Abbreviations: GLM, germ line mutation; GTR, gross total resection; M, metastasis; y, year of age at diagnosis.

Treatment Adjustments – The Role of Molecular Subgroups

In 2020 we agreed on a consensus classification of ATRT into three separate molecular groups, each characterized by distinct clinical, molecular, and genetic characteristics. We suggest that divergent gene expression profiles²⁷ may be responsible for the heterogeneous response to conventional but also potential targeted therapy approaches. Recently, Johann and Altendorf elucidated distinct gene expression profiles in primary vs relapsed ATRT. For ATRT-MYC, differences were seen in the expression of cell cycle-dependent genes (eg, HDAC), while gene coding for components of the immune system was more likely to be downregulated in relapse compared to primaries. Tyrosine kinase receptor genes were overexpressed in relapse in ATRT-TYR tumors, while ATRT-SHH relapse displayed higher expression of genes associated with metastases.¹⁴⁶

Paassen et al developed a series of ATRT tumoroids and performed extensive drug screening. In contrast to suggestions by Altendorf et al, tyrosine kinase inhibitors were significantly more effective in ATRT-MYC tumoroids when compared to ATRT-SHH. For ATRT-SHH, gamma-secretase inhibitors affecting the NOTCH-pathway were most successful, even though differences between the different ATRT-SHH models were detected.¹⁴⁷ We would like to stress that different gene expression profiles of the distinct DNA methylation groups will likely play a crucial role for targeted

treatment approaches. Results from current studies will need to be confirmed in larger cohorts. Eventually clinical trials investigating novel therapeutic agents should correlate specifics of molecular subgroups with clinical data.

Improvements in the Diagnostic Workup

It remains to be determined whether our diagnostic portfolio is efficient enough to define treatment responses and, even more importantly, to detect relapse early. Standard of care surveillance methods mainly comprise MRI screens and occasionally CSF cytology, both demonstrating significant inter-examiner variability.

Ideally, diagnostic methods with a high specificity and sensitivity will enhance the screening for minimal residual disease (MRD) currently invisible to the human eye. Detecting cell free DNA (cfDNA) in body fluids reflects current attempts to gather diagnostic information by a non- or minimally invasive and technically standardizable system. These liquid biopsies will complement future diagnostic surveillance and initial workup. Such approaches have been investigated in different specimens. Sample volume and amount of DNA per volume have to be considered when designing experimental analyses. In the CSF of patients with CNS tumors, the amount of cfDNA is generally low. It appears to be significantly higher compared to serum/plasma, even though the absolute amount of DNA remains limited.¹⁴⁸ Nanopore sequencing has recently been shown to be an easy-to-use and reliable method to detect even small amounts of DNA in limited sample volumes.^{149–151} Nanopore provides a membrane-based whole sequencing approach without the need for polymerase-based amplification.^{149–153} Copy number variations (CNV) and DNA methylation profiles may be generated at the same time from the same sample.¹⁵³ Data by Afflerbach et al demonstrated encouraging success in detecting amounts of 5 ng of circulating cell free DNA in CSF volumes as little as 1 mL. These investigators were able to show that cfDNA was indeed circulating tumor DNA (ctDNA), as they presented identical CNV and DNA methylation patterns as in the tumor tissue. This was validated in eight patients suffering from ATRT. These methods were considered appropriate for the detection of ctDNA in pre-operatively collected CSF as well as in the longitudinal disease monitoring.¹⁵³

In summary, several lines of evidence demonstrate that liquid biopsy may be a suitable tool for MRD detection especially in high grade (embryonal) CNS tumors as 1) the amount of ctDNA is higher than in CNS tumors of lower grade and 2) a CSF access device (eg, Ommaya or Rickham) will be placed for intraventricular therapy in the first-line treatment.^{148,153} We anticipate a high potential for the establishment of liquid biopsies from ventricular reservoir CSF in patients with ATRT. Collecting samples within the scope of clinical trials may further be considered but requires a standardized procedure for both specimen collection and storage as well as laboratory analyses.

Preclinical Evaluation of Potential Novel Therapies

Traditionally, animal models have been an important tool for drug testing. ATRT animal models, most commonly immune-incompetent mice for xenografts or genetically engineered mouse models (GEMM), appear to be rather limited.^{154,155} Currently available models do not exactly recapitulate ATRT biology, eg, the models published by Roberts et al demonstrate rather long generation times and CNS tumors are rarely seen,¹⁵⁶ xenograft tumors exhibit an incomplete tumor immune infiltration due to the generation in nude mice and even the GEMM such as the one published by Han or Melcher do not give a perfect representation of the established methylation subtypes.^{155,157}

As an alternative to animal models, attempts have been made to create tumoroids based on patient-derived tumor samples or patient-derived orthotopic xenografts (PDOX). Paassen et al developed such tumoroid models and demonstrated the potential use in pre-clinical research, as these models are presented with stable genetic and epigenetic characteristics despite several cycles of tumor cell passage and expansion in cell culture.¹⁴⁷ Even if these authors used minced tumor tissue,¹⁴⁷ consisting of tumor cells and tumor microenvironment, these tumoroids are still not a perfect representation of the primary tissue. This may be explained by the lack of our knowledge on how to produce the 3-dimensional architecture within tumoroid models. Developing innovative tumor models will have to respect the spatial geometry of ATRT.

Outlook

Future research needs to focus on a correlation of molecular and genetic structures with response to treatment. This will be crucial to provide the best possible treatment for each patient, to design meaningful clinical trials, and to eventually improve survival.

Trial designs face the challenge of requirements for randomized controlled clinical trials to gather as much evidence as possible on the one hand, and individual genetic and molecular findings on the other small subcohorts of patients. International knowledge must be merged in common databases to overcome limited cohort sizes and to achieve maximum possible evidence.

Conclusion

Conventional, multimodal therapy in ATRT deserves further optimization. Regardless of the treatment approach, risk factors such as age, metastatic disease, and GLM affect survival probability. The clinical benefit and the risk of severe acute and late effects, especially in young children, stress the necessity to modify radiotherapy approaches. Proton beam therapy may cause less severe late effects and may therefore be an option in children younger than 18 months. It has been proposed that HDCT may replace radiotherapy; this issue remains unresolved.

While specific, mostly institutional approaches for the treatment of primary ATRT exist, the management of refractory and relapsed ATRT in general follows clinical phase I/II trials or individual treatment attempts. Agents under investigation include CDK4/6, PARP, or aurora kinase a inhibitors. Others belong to the group of epigenome modifying compounds (such as decitabine, or histone modifying agents). Inhibition of the PD-1/PD-L1 interaction or T cell activation via CTLA4 blockade targets the tumor cell directed immune responses. CAR-T-cells or oncolytic viruses are additional immune modulating agents. Recent basket trials to individualize targeted therapy based on expression profiles still face the difficulty of identifying reliable and targetable structures.

We suggest that a research-informed combinations of agents will improve treatment response in primary as well as r/r ATRT. Crucially, merging global knowledge in the form of internationally recruiting trials or meta-analyses is an important approach to reach significant numbers by building efficient collaborative networks.

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References

1. Louis DN, Perry A, Wesseling P, et al. The 2021 WHO classification of tumors of the central nervous system: a summary. *Neuro Oncol.* 2021;23(8):1231–1251. doi:10.1093/neuonc/noab106
2. Erdmann F, Kaatsch P, Grabow D, Spix C. *German Childhood Cancer Registry - Annual Report 2019 (1980–2018)*. Mainz: Institute of Medical Biostatistics, Epidemiology and informatics (IMBEI) at the University Medical Center of the Johannes Gutenberg University; 2020.
3. Ostrom QT, Price M, Neff C, et al. CBTRUS statistical report: primary brain and other central nervous system tumors diagnosed in the United States in 2015–2019. *Neuro Oncol.* 2022;24(Suppl 5):v1–v95. doi:10.1093/neuonc/noac202

4. Frühwald MC, Hasselblatt M, Nemes K, et al. Age and DNA methylation subgroup as potential independent risk factors for treatment stratification in children with atypical teratoid/rhabdoid tumors. *Neuro Oncol.* 2020;22(7):1006–1017. doi:10.1093/neuonc/noz244
5. Oka H, Scheithauer BW. Clinicopathological characteristics of atypical teratoid/rhabdoid tumor. *Neurol Med Chir.* 1999;39(7):510–517; discussion 517–518. doi:10.2176/nmc.39.510
6. Benesch M, Nemes K, Neumayer P, et al. Spinal cord atypical teratoid/rhabdoid tumors in children: clinical, genetic, and outcome characteristics in a representative European cohort. *Pediatr Blood Cancer.* 2020;67(1):e28022. doi:10.1002/pbc.28022
7. Bartelheim K, Sumerauer D, Behrends U, et al. Clinical and genetic features of rhabdoid tumors of the heart registered with the European Rhabdoid Registry (EU-RHAB). *Cancer Genet.* 2014;207(9):379–383. doi:10.1016/j.cancergen.2014.04.005
8. Pohl U, Dean AF, Ichimura K, et al. Genomic analysis of chromosome 22 in synchronous and histologically distinct intracranial tumours in a child. *Neuropathol Appl Neurobiol.* 2010;36(4):359–363. doi:10.1111/j.1365-2990.2010.01085.x
9. Litman DA, Bhuta S, Barsky SH. Synchronous occurrence of malignant rhabdoid tumor two decades after Wilms' tumor irradiation. *Am J Surg Pathol.* 1993;17(7):729–737. doi:10.1097/00000478-199307000-00011
10. Abu Arja MH, Patel P, Shah SH, et al. Synchronous central nervous system atypical teratoid/rhabdoid tumor and malignant rhabdoid tumor of the kidney: case report of a long-term survivor and review of the literature. *World Neurosurg.* 2018;111:6–15. doi:10.1016/j.wneu.2017.11.158
11. Frühwald MC, Biegel JA, Bourdeaut F, Roberts CW, Chi SN. Atypical teratoid/rhabdoid tumors-current concepts, advances in biology, and potential future therapies. *Neuro Oncol.* 2016;18(6):764–778. doi:10.1093/neuonc/nov264
12. Tekautz TM, Fuller CE, Blaney S, et al. Atypical teratoid/rhabdoid tumors (ATRT): improved survival in children 3 years of age and older with radiation therapy and high-dose alkylator-based chemotherapy. *J Clin Oncol.* 2005;23(7):1491–1499. doi:10.1200/JCO.2005.05.187
13. Hasselblatt M, Thomas C, Federico A, et al. Low-grade diffusely infiltrative tumour (LGDIT), SMARCB1-mutant: a clinical and histopathological distinct entity showing epigenetic similarity with ATRT-MYC. *Neuropathol Appl Neurobiol.* 2022;48(4):e12797. doi:10.1111/nan.12797
14. Matsumura N, Goda N, Yashige K, et al. Desmoplastic myxoid tumor, SMARCB1-mutant: a new variant of SMARCB1-deficient tumor of the central nervous system preferentially arising in the pineal region. *Virchows Arch.* 2021;479(4):835–839. doi:10.1007/s00428-020-02978-3
15. Thomas C, Wefers A, Bens S, et al. Desmoplastic myxoid tumor, SMARCB1-mutant: clinical, histopathological and molecular characterization of a pineal region tumor encountered in adolescents and adults. *Acta Neuropathol.* 2020;139(2):277–286. doi:10.1007/s00401-019-02094-w
16. Hasselblatt M, Oyen F, Gesk S, et al. Cribriform neuroepithelial tumour (CRINET): a nonrhabdoid ventricular tumor with INI1 loss and relatively favorable prognosis. *J Neuropathol Exp Neurol.* 2009;68(12):1249–1255. doi:10.1097/NEN.0b013e3181c06a51
17. Cai C. SWI/SNF deficient central nervous system neoplasms. *Semin Diagn Pathol.* 2021;38(3):167–174. doi:10.1053/j.semdp.2021.03.003
18. Thomas C, Federico A, Sill M, et al. Atypical Teratoid/Rhabdoid Tumor (AT/RT) with molecular features of pleomorphic xanthoastrocytoma. *Am J Surg Pathol.* 2021;45(9):1228–1234. doi:10.1097/PAS.0000000000001694
19. Hasselblatt M, Thomas C, Federico A, et al. SMARCB1-deficient and SMARCA4-deficient malignant brain tumors with complex copy number alterations and TP53 mutations may represent the first clinical manifestation of li-fraumeni syndrome. *Am J Surg Pathol.* 2022;46(9):1277–1283. doi:10.1097/PAS.0000000000001905
20. Nesvick CL, Lafay-Cousin L, Raghunathan A, Bouffet E, Huang AA, Daniels DJ. Atypical teratoid rhabdoid tumor: molecular insights and translation to novel therapeutics. *J Neurooncol.* 2020;150(1):47–56. doi:10.1007/s11060-020-03639-w
21. Bourdeaut F, Lequin D, Brugières L, et al. Frequent hSNF5/INI1 germline mutations in patients with rhabdoid tumor. *Clin Cancer Res.* 2011;17(1):31–38. doi:10.1158/1078-0432.CCR-10-1795
22. Eaton KW, Tooke LS, Wainwright LM, Judkins AR, Biegel JA. Spectrum of SMARCB1/INI1 mutations in familial and sporadic rhabdoid tumors. *Pediatr Blood Cancer.* 2011;56(1):7–15. doi:10.1002/pbc.22831
23. Hasselblatt M, Nagel I, Oyen F, et al. SMARCA4-mutated atypical teratoid/rhabdoid tumors are associated with inherited germline alterations and poor prognosis. *Acta Neuropathol.* 2014;128(3):453–456. doi:10.1007/s00401-014-1323-x
24. Johann PD, Erkek S, Zapatka M, et al. Atypical teratoid/rhabdoid tumors are comprised of three epigenetic subgroups with distinct enhancer landscapes. *Cancer Cell.* 2016;29(3):379–393. doi:10.1016/j.ccell.2016.02.001
25. Erdmann FKP, Grabow D, Spix C. *German Childhood Cancer Registry Annual Report 2019 (1980–2018)*. Mainz: Institute of Medical Biostatistics, Epidemiology and informatics (IMBEI) at the University Medical Center of the Johannes Gutenberg University; 2020.
26. Johann PD, Hovestadt V, Thomas C, et al. Cribriform neuroepithelial tumor: molecular characterization of a SMARCB1-deficient non-rhabdoid tumor with favorable long-term outcome. *Brain Pathol.* 2017;27(4):411–418. doi:10.1111/bpa.12413
27. Ho B, Johann PD, Grabovska Y, et al. Molecular subgrouping of atypical teratoid/rhabdoid tumors—a re-investigation and current consensus. *Neuro Oncol.* 2020;22(5):613–624. doi:10.1093/neuonc/noz235
28. Agaimy A, Foulkes WD. Hereditary SWI/SNF complex deficiency syndromes. *Semin Diagn Pathol.* 2018;35(3):193–198. doi:10.1053/j.semdp.2018.01.002
29. Chun HE, Johann PD, Milne K, et al. Identification and analyses of extra-cranial and cranial rhabdoid tumor molecular subgroups reveal tumors with cytotoxic T cell infiltration. *Cell Rep.* 2019;29(8):2338–2354 e2337. doi:10.1016/j.celrep.2019.10.013
30. Federico A, Thomas C, Miskiewicz K, et al. ATRT-SHH comprises three molecular subgroups with characteristic clinical and histopathological features and prognostic significance. *Acta Neuropathol.* 2022;143(6):697–711. doi:10.1007/s00401-022-02424-5
31. Lobón-Iglesias MJ, Andrianteranagna M, Han ZY, et al. Imaging and multi-omics datasets converge to define different neural progenitor origins for ATRT-SHH subgroups. *Nat Commun.* 2023;14(1):6669. doi:10.1038/s41467-023-42371-7
32. Upadhyaya SA, Robinson GW, Onar-Thomas A, et al. Relevance of molecular groups in children with newly diagnosed atypical teratoid rhabdoid tumor: results from prospective St. Jude multi-institutional trials. *Clin Cancer Res.* 2021;27(10):2879–2889. doi:10.1158/1078-0432.CCR-20-4731
33. Nemes K, Bens S, Kachanov D, et al. Clinical and genetic risk factors define two risk groups of extracranial malignant rhabdoid tumours (eMRT/RTK). *Eur J Cancer.* 2021;142:112–122. doi:10.1016/j.ejca.2020.10.004
34. Nemes K, Bens S, Bourdeaut F, et al. Rhabdoid tumor predisposition syndrome. In: Adam MP, Everman DB, Mirzaa GM, et al. editors. *GeneReviews*®. Seattle: University of Washington, Seattle. Copyright © 1993–2023, University of Washington, Seattle. GeneReviews is a registered trademark of the University of Washington, Seattle. All rights reserved; 2017.
35. Frühwald MC, Nemes K, Boztug H, et al. Current recommendations for clinical surveillance and genetic testing in rhabdoid tumor predisposition: a report from the SIOPE Host Genome Working Group. *Fam Cancer.* 2021;20(4):305–316. doi:10.1007/s10689-021-00229-1

36. Sredni ST, Tomita T. Rhabdoid tumor predisposition syndrome. *Pediatr Dev Pathol.* 2015;18(1):49–58. doi:10.2350/14-07-1531-MISC.1
37. Seeringer A, Reinhard H, Hasselblatt M, et al. Synchronous congenital malignant rhabdoid tumor of the orbit and atypical teratoid/rhabdoid tumor—feasibility and efficacy of multimodal therapy in a long-term survivor. *Cancer Genet.* 2014;207(9):429–433. doi:10.1016/j.cancergen.2014.06.028
38. Baker TG, Lyons MJ, Leddy L, Parham DM, Welsh CT. Epithelioid sarcoma arising in a long-term survivor of an atypical teratoid/rhabdoid tumor in a patient with rhabdoid tumor predisposition syndrome. *Pediatr Dev Pathol.* 2021;24(2):164–168. doi:10.1177/1093526620986492
39. Kordes U, Bartelheim K, Modena P, et al. Favorable outcome of patients affected by rhabdoid tumors due to rhabdoid tumor predisposition syndrome (RTPS). *Pediatr Blood Cancer.* 2014;61(5):919–921. doi:10.1002/pbc.24793
40. Fukushima H, Yamasaki K, Sakaida M, et al. Rhabdoid tumor predisposition syndrome with renal tumor 10 years after brain tumor. *Pathol Int.* 2021;71(2):155–160. doi:10.1111/pin.13056
41. Corinaldesi C, Holmes AB, Shen Q, et al. Tracking immunoglobulin repertoire and transcriptomic changes in germinal center B cells by single-cell analysis. *Front Immunol.* 2021;12:818758. doi:10.3389/fimmu.2021.818758
42. Kretzmer H, Bernhart SH, Wang W, et al. DNA methylome analysis in Burkitt and follicular lymphomas identifies differentially methylated regions linked to somatic mutation and transcriptional control. *Nat Genet.* 2015;47(11):1316–1325. doi:10.1038/ng.3413
43. Bennett J, Erker C, Lafay-Cousin L, et al. Canadian pediatric neuro-oncology standards of practice. *Front Oncol.* 2020;10:593192. doi:10.3389/fonc.2020.593192
44. Bartelheim K, Nemes K, Seeringer A, et al. Improved 6-year overall survival in AT/RT - results of the registry study Rhabdoid 2007. *Cancer Med.* 2016;5(8):1765–1775. doi:10.1002/cam4.741
45. Reddy AT, Strother DR, Judkins AR, et al. Efficacy of high-dose chemotherapy and three-dimensional conformal radiation for atypical teratoid/rhabdoid tumor: a report from the Children's Oncology Group Trial ACNS0333. *J Clin Oncol.* 2020;38(11):1175–1185. doi:10.1200/JCO.19.01776
46. Slave I, Chocholous M, Leiss U, et al. Atypical teratoid rhabdoid tumor: improved long-term survival with an intensive multimodal therapy and delayed radiotherapy. The Medical University of Vienna Experience 1992–2012. *Cancer Med.* 2014;3(1):91–100. doi:10.1002/cam4.161
47. Zaky W, Dhall G, Ji L, et al. Intensive induction chemotherapy followed by myeloablative chemotherapy with autologous hematopoietic progenitor cell rescue for young children newly-diagnosed with central nervous system atypical teratoid/rhabdoid tumors: the Head Start III experience. *Pediatr Blood Cancer.* 2014;61(1):95–101. doi:10.1002/pbc.24648
48. Hinkes BG, von Hoff K, Deinlein F, et al. Childhood pineoblastoma: experiences from the prospective multicenter trials HIT-SKK87, HIT-SKK92 and HIT91. *J Neurooncol.* 2007;81(2):217–223. doi:10.1007/s11060-006-9221-2
49. Müller K, Zwiener I, Welker H, et al. Curative treatment for central nervous system medulloepithelioma despite residual disease after resection. Report of two cases treated according to the GPHO Protocol HIT 2000 and review of the literature. *Strahlenther Onkol.* 2011;187(11):757–762. doi:10.1007/s00066-011-2256-0
50. Athale UH, Duckworth J, Odame I, Barr R. Childhood atypical teratoid rhabdoid tumor of the central nervous system: a meta-analysis of observational studies. *J Pediatr Hematol Oncol.* 2009;31(9):651–663. doi:10.1097/MPH.0b013e3181b258a9
51. Buscariollo DL, Park HS, Roberts KB, Yu JB. Survival outcomes in atypical teratoid rhabdoid tumor for patients undergoing radiotherapy in a surveillance, epidemiology, and end results analysis. *Cancer.* 2012;118(17):4212–4219. doi:10.1002/cncr.27373
52. von Hoff K, Hinkes B, Dannenmann-Stern E, et al. Frequency, risk-factors and survival of children with atypical teratoid rhabdoid tumors (AT/RT) of the CNS diagnosed between 1988 and 2004, and registered to the German HIT database. *Pediatr Blood Cancer.* 2011;57(6):978–985. doi:10.1002/pbc.23236
53. Schrey D, Carceller Lechón F, Malietzis G, et al. Multimodal therapy in children and adolescents with newly diagnosed atypical teratoid rhabdoid tumor: individual pooled data analysis and review of the literature. *J Neurooncol.* 2016;126(1):81–90. doi:10.1007/s11060-015-1904-0
54. Underiner RM, Eltobgy M, Stanek JR, Finlay JL, AbdelBaki MS. Meta-analysis of treatment modalities in metastatic atypical teratoid/rhabdoid tumors in children. *Pediatr Neurol.* 2020;108:106–112. doi:10.1016/j.pediatrneurol.2020.03.003
55. Silva AHD, Habermann S, Craven CL, et al. Atypical teratoid rhabdoid tumours (ATRTs)-A 21-year institutional experience. *Childs Nerv Syst.* 2023;39(6):1509–1518. doi:10.1007/s00381-023-05828-0
56. Chi SN, Zimmerman MA, Yao X, et al. Intensive multimodality treatment for children with newly diagnosed CNS atypical teratoid rhabdoid tumor. *J Clin Oncol.* 2009;27(3):385–389. doi:10.1200/JCO.2008.18.7724
57. Lafay-Cousin L, Hawkins C, Carret AS, et al. Central nervous system atypical teratoid rhabdoid tumours: the Canadian paediatric brain tumour consortium experience. *Eur J Cancer.* 2012;48(3):353–359. doi:10.1016/j.ejca.2011.09.005
58. Yamasaki K, Kiyotani C, Terashima K, et al. Clinical characteristics, treatment, and survival outcome in pediatric patients with atypical teratoid/rhabdoid tumors: a retrospective study by the Japan Children's Cancer Group. *J Neurosurg Pediatr.* 2019;1–10. doi:10.3171/2019.9.PEDS19367
59. Gonzalez G, Delgado M, Blanco A, Puentes M. SIOP ABSTRACTS. *Pediatr Blood Cancer.* 2021;68(Suppl 5):e29349. doi:10.1002/pbc.29349
60. Fischer-Valuck BW, Chen I, Srivastava AJ, et al. Assessment of the treatment approach and survival outcomes in a modern cohort of patients with atypical teratoid rhabdoid tumors using the national cancer database. *Cancer.* 2017;123(4):682–687. doi:10.1002/cncr.30405
61. Dahl NA, Liu AK, Foreman NK, Widener M, Fenton LZ, Macy ME. Bevacizumab in the treatment of radiation injury for children with central nervous system tumors. *Childs Nerv Syst.* 2019;35(11):2043–2046.
62. Plimpton SR, Stence N, Hemenway M, Hankinson TC, Foreman N, Liu AK. Cerebral radiation necrosis in pediatric patients. *Pediatr Hematol Oncol.* 2015;32(1):78–83. doi:10.3109/08880018.2013.791738
63. Nanda RH, Ganju RG, Schreiber E, et al. Correlation of acute and late brainstem toxicities with dose-volume data for pediatric patients with posterior fossa malignancies. *Int J Radiat Oncol Biol Phys.* 2017;98(2):360–366. doi:10.1016/j.ijrobp.2017.02.092
64. Dashti SR, Spalding A, Kadner RJ, et al. Targeted intraarterial anti-VEGF therapy for medically refractory radiation necrosis in the brain. *J Neurosurg Pediatr.* 2015;15(1):20–25. doi:10.3171/2014.9.PEDS14198
65. Baliga S, Gandola L, Timmermann B, et al. Brain tumors: medulloblastoma, ATRT, ependymoma. *Pediatr Blood Cancer.* 2021;68(Suppl 2):e28395.
66. Yang WC, Yen HJ, Liang ML, et al. Effect of early radiotherapy initiation and high-dose chemotherapy on the prognosis of pediatric atypical teratoid rhabdoid tumors in different age groups. *J Neurooncol.* 2020;147(3):619–631. doi:10.1007/s11060-020-03456-1
67. Squire SE, Chan MD, Marcus KJ. Atypical teratoid/rhabdoid tumor: the controversy behind radiation therapy. *J Neurooncol.* 2007;81(1):97–111. doi:10.1007/s11060-006-9196-z

68. Biewald E, Kiefer T, Geismar D, et al. Feasibility of proton beam therapy as a rescue therapy in heavily pre-treated retinoblastoma eyes. *Cancers*. 2021;13(8):1862. doi:10.3390/cancers13081862
69. McGovern SL, Okcu MF, Munsell MF, et al. Outcomes and acute toxicities of proton therapy for pediatric atypical teratoid/rhabdoid tumor of the central nervous system. *Int J Radiat Oncol Biol Phys*. 2014;90(5):1143–1152. doi:10.1016/j.ijrobp.2014.08.354
70. Jazmati D, Steinmeier T, Ahamd Khalil D, et al. Feasibility of proton beam therapy for infants with brain tumours: experiences from the Prospective KiProReg Registry Study. *Clin Oncol*. 2021;33(7):e295–e304. doi:10.1016/j.clon.2021.03.006
71. Upadhyay R, Liao K, Grosshans DR, et al. Quantifying the risk and dosimetric variables of symptomatic brainstem injury after proton beam radiation in pediatric brain tumors. *Neuro Oncol*. 2022;24(9):1571–1581. doi:10.1093/neuonc/noac044
72. De Amorim Bernstein K, Sethi R, Trofimov A, et al. Early clinical outcomes using proton radiation for children with central nervous system atypical teratoid rhabdoid tumors. *Int J Radiat Oncol Biol Phys*. 2013;86(1):114–120. doi:10.1016/j.ijrobp.2012.12.004
73. Kralik SF, Ho CY, Finke W, Buchsbaum JC, Haskins CP, Shih CS. Radiation necrosis in pediatric patients with brain tumors treated with proton radiotherapy. *Am J Neuroradiol*. 2015;36(8):1572–1578. doi:10.3174/ajnr.A4333
74. Seeringer A, Bartelheim K, Kerl K, et al. Feasibility of intensive multimodal therapy in infants affected by rhabdoid tumors - experience of the EU-RHAB registry. *Klin Padiatr*. 2014;226(3):143–148. doi:10.1055/s-0034-1368719
75. Mestnik S, Wilson S, Huang A, Sato M. Prolonged remission achieved with maintenance intraventricular chemotherapy in young patient with recurrent atypical teratoid rhabdoid tumor. *Pediatr Blood Cancer*. 2023;70(6):e30225. doi:10.1002/pbc.30225
76. Gottschling S, Reinhard H, Meyer S, Krenn T, Graf N, Strowitzki M. Severe encephalopathy caused by intraparenchymal methotrexate instillation due to the design of the catheter. *Med Pediatr Oncol*. 2003;41(5):491–492. doi:10.1002/mpo.10117
77. Packer RJ, Zimmerman RA, Rosenstock J, Rorke LB, Norris DG, Berman PH. Focal encephalopathy following methotrexate therapy. Administration via a misplaced intraventricular catheter. *Arch Neurol*. 1981;38(7):450–452. doi:10.1001/archneur.1981.00510070084016
78. Shapiro WR, Chernik NL, Posner JB. Necrotizing encephalopathy following intraventricular instillation of methotrexate. *Arch Neurol*. 1973;28(2):96–102. doi:10.1001/archneur.1973.00490200044005
79. Bachu VS, Shah P, Jimenez AE, et al. Clinical predictors of survival for patients with atypical teratoid/rhabdoid tumors. *Childs Nerv Syst*. 2022;38(7):1297–1306. doi:10.1007/s00381-022-05511-w
80. Nemes K, Johann PD, Steinbügl M, et al. Infants and Newborns with Atypical Teratoid Rhabdoid Tumors (ATRT) and Extracranial Malignant Rhabdoid Tumors (eMRT) in the EU-RHAB registry: a unique and challenging population. *Cancers*. 2022;14(9):2185. doi:10.3390/cancers14092185
81. Lafay-Cousin L, Fay-McClymont T, Johnston D, et al. Neurocognitive evaluation of long term survivors of atypical teratoid rhabdoid tumors (ATRT): the Canadian registry experience. *Pediatr Blood Cancer*. 2015;62(7):1265–1269. doi:10.1002/pbc.25441
82. van Tilburg CM, Pfäff E, Pajtler KW, et al. The pediatric precision oncology INFORM registry: clinical outcome and benefit for patients with very high-evidence targets. *Cancer Discov*. 2021;11(11):2764–2779. doi:10.1158/2159-8290.CD-21-0094
83. Berlanga P, Pierron G, Lacroix L, et al. The European MAPPYACTS trial: precision medicine program in pediatric and adolescent patients with recurrent malignancies. *Cancer Discov*. 2022;12(5):1266–1281. doi:10.1158/2159-8290.CD-21-1136
84. Kazansky Y, Cameron D, Demarest P, et al. Overcoming clinical resistance to EZH2 inhibition using rational epigenetic combination therapy. *bioRxiv*. 2023. doi:10.1101/2023.02.06.527192
85. Steinbügl M, Nemes K, Johann P, et al. Clinical evidence for a biological effect of epigenetically active decitabine in relapsed or progressive rhabdoid tumors. *Pediatr Blood Cancer*. 2021;68(12):e29267. doi:10.1002/pbc.29267
86. Matthews HK, Bertoli C, de Bruin RAM. Cell cycle control in cancer. *Nat Rev Mol Cell Biol*. 2022;23(1):74–88. doi:10.1038/s41580-021-00404-3
87. Roulois D, Loo Yau H, Singhania R, et al. DNA-demethylating agents target colorectal cancer cells by inducing viral mimicry by endogenous transcripts. *Cell*. 2015;162(5):961–973. doi:10.1016/j.cell.2015.07.056
88. Janin M, Esteller M. Epigenetic awakening of viral mimicry in cancer. *Cancer Discov*. 2020;10(9):1258–1260. doi:10.1158/2159-8290.CD-20-0947
89. Deblais G, Tonekaboni SAM, Grillo G, et al. Epigenetic switch-induced viral mimicry evasion in chemotherapy-resistant breast cancer. *Cancer Discov*. 2020;10(9):1312–1329. doi:10.1158/2159-8290.CD-19-1493
90. Patel AA, Cahill K, Saygin C, Odenike O. Cedazuridine/decitabine: from preclinical to clinical development in myeloid malignancies. *Blood Adv*. 2021;5(8):2264–2271. doi:10.1182/bloodadvances.2020002929
91. Kakkar A, Biswas A, Goyal N, et al. The expression of Cyclin D1, VEGF, EZH2, and H3K27me3 in atypical teratoid/rhabdoid tumors of the CNS: a possible role in targeted therapy. *Appl Immunohistochem Mol Morphol*. 2016;24(10):729–737. doi:10.1097/PAL.0000000000000247
92. Alimova I, Birks DK, Harris PS, et al. Inhibition of EZH2 suppresses self-renewal and induces radiation sensitivity in atypical rhabdoid teratoid tumor cells. *Neuro Oncol*. 2013;15(2):149–160. doi:10.1093/neuonc/nos285
93. Knutson SK, Warholc NM, Wigle TJ, et al. Durable tumor regression in genetically altered malignant rhabdoid tumors by inhibition of methyltransferase EZH2. *Proc Natl Acad Sci U S A*. 2013;110(19):7922–7927. doi:10.1073/pnas.1303800110
94. Prorok P, Forouzanfar F, Murugarren N, et al. Loss of Ezh2 function remodels the DNA replication initiation landscape. *Cell Rep*. 2023;42(4):112280. doi:10.1016/j.celrep.2023.112280
95. Straining R, Eighmy WT. Tazemetostat: EZH2 inhibitor. *J Adv Pract Oncol*. 2022;13(2):158–163. doi:10.6004/jadpro.2022.13.2.7
96. Chi SN, Bourdeaut F, Laetsch TW, et al. Phase I study of tazemetostat, an enhancer of zeste homolog-2 inhibitor, in pediatric pts with relapsed/refractory integrase interactor 1-negative tumors. *J Clin Oncol*. 2020;38(15_suppl):10525. doi:10.1200/JCO.2020.38.15_suppl.10525
97. Chi SN, Bourdeaut F, Casanova M, et al. Update on Phase I study of tazemetostat, an enhancer of zeste homolog 2 inhibitor, in pediatric patients with relapsed or refractory integrase interactor 1-negative tumors. *J Clin Oncol*. 2022;40(16_suppl):10040. doi:10.1200/JCO.2022.40.16_suppl.10040
98. Gounder M, Schoffski P, Jones RL, et al. Tazemetostat in advanced epithelioid sarcoma with loss of INI1/SMARCB1: an international, open-label, Phase 2 basket study. *Lancet Oncol*. 2020;21(11):1423–1432. doi:10.1016/S1470-2045(20)30451-4
99. Chi SN, Yi JS, Williams PM, et al. Tazemetostat for tumors harboring SMARCB1/SMARCA4 or EZH2 alterations: results from NCI-COG pediatric MATCH APEC1621C. *J Natl Cancer Inst*. 2023;115(11):1355–1363. doi:10.1093/jnci/djad085
100. Gibaja V, Shen F, Harari J, et al. Development of secondary mutations in wild-type and mutant EZH2 alleles cooperates to confer resistance to EZH2 inhibitors. *Oncogene*. 2016;35(5):558–566. doi:10.1038/ncr.2015.114
101. Shinohara H, Sawado R, Nakagawa M, et al. Dual targeting of EZH1 and EZH2 for the treatment of malignant rhabdoid tumors. *Mol Ther Oncolytics*. 2022;27:14–25. doi:10.1016/j.omto.2022.09.006

102. Qi L, Lindsay H, Kogiso M, et al. Evaluation of an EZH2 inhibitor in patient-derived orthotopic xenograft models of pediatric brain tumors alone and in combination with chemo- and radiation therapies. *Lab Invest.* 2022;102(2):185–193. doi:10.1038/s41374-021-00700-8
103. Wood P, Desai J, Waldeck K, et al. ATRT-17. A phase II study of continuous low dose panobinostat in paediatric patients with malignant rhabdoid tumours and atypical teratoid rhabdoid tumours. *Neuro-Oncology.* 2022;24(Supplement_1):i6–i7. doi:10.1093/neuonc/noac079.016
104. Liewer S, Huddleston A. Alisertib: a review of pharmacokinetics, efficacy and toxicity in patients with hematologic malignancies and solid tumors. *Expert Opin Investig Drugs.* 2018;27(1):105–112. doi:10.1080/13543784.2018.1417382
105. Upadhyaya SA, Campagne O, Billups CA, et al. Phase II study of alisertib as a single agent for treating recurrent or progressive atypical teratoid/rhabdoid tumor. *Neuro Oncol.* 2023;25(2):386–397. doi:10.1093/neuonc/noac151
106. Howden K, McDonald PJ, Kazina C, et al. Sustained and durable response with Alisertib monotherapy in the treatment of relapsed Atypical Teratoid Rhabdoid Tumor (ATRT). *Neurooncol Adv.* 2022;4(1):vdac090. doi:10.1093/nojnl/vdac090
107. Aplenc R, Blaney SM, Strauss LC, et al. Pediatric phase I trial and pharmacokinetic study of dasatinib: a report from the children's oncology group phase I consortium. *J Clin Oncol.* 2011;29(7):839–844. doi:10.1200/JCO.2010.30.7231
108. Patel YT, Davis A, Baker SJ, Campagne O, Stewart CF. CNS penetration of the CDK4/6 inhibitor ribociclib in non-tumor bearing mice and mice bearing pediatric brain tumors. *Cancer Chemother Pharmacol.* 2019;84(2):447–452. doi:10.1007/s00280-019-03864-9
109. Georger B, Bourdeaut F, DuBois SG, et al. A Phase I Study of the CDK4/6 Inhibitor Ribociclib (LEE011) in pediatric patients with malignant rhabdoid tumors, neuroblastoma, and other solid tumors. *Clin Cancer Res.* 2017;23(10):2433–2441. doi:10.1158/1078-0432.CCR-16-2898
110. DeWire MD, Fuller C, Campagne O, et al. A Phase I and surgical study of ribociclib and everolimus in children with recurrent or refractory malignant brain tumors: a Pediatric Brain Tumor Consortium Study. *Clin Cancer Res.* 2021;27(9):2442–2451. doi:10.1158/1078-0432.CCR-20-4078
111. Jin D, Tran N, Thomas N, Tran DD. Combining CDK4/6 inhibitors ribociclib and palbociclib with cytotoxic agents does not enhance cytotoxicity. *PLoS One.* 2019;14(10):e0223555. doi:10.1371/journal.pone.0223555
112. Van Mater D, Gururangan S, Becher O, et al. A phase I trial of the CDK 4/6 inhibitor palbociclib in pediatric patients with progressive brain tumors: a Pediatric Brain Tumor Consortium study (PBTC-042). *Pediatr Blood Cancer.* 2021;68(4):e28879. doi:10.1002/pbc.28879
113. Alimova I, Murdock G, Pierce A, et al. The PARP inhibitor Rucaparib synergizes with radiation to attenuate atypical teratoid rhabdoid tumor growth. *Neurooncol Adv.* 2023;5(1):vdad010. doi:10.1093/nojnl/vdad010
114. Keung MYT, Wu Y, Vadgama JV. PARP inhibitors as a therapeutic agent for homologous recombination deficiency in breast cancers. *J Clin Med.* 2019;8(4):435. doi:10.3390/jcm8040435
115. Schafer ES, Rau RE, Berg SL, et al. Phase 1/2 trial of talazoparib in combination with temozolomide in children and adolescents with refractory/recurrent solid tumors including Ewing sarcoma: a Children's Oncology Group Phase 1 Consortium study (ADVL1411). *Pediatr Blood Cancer.* 2020;67(2):e28073. doi:10.1002/pbc.28073
116. Keller KM, Koetsier J, Schild L, et al. The potential of PARP as a therapeutic target across pediatric solid malignancies. *BMC Cancer.* 2023;23(1):310. doi:10.1186/s12885-022-10319-7
117. Lv B, Wang Y, Ma D, et al. Immunotherapy: reshape the tumor immune microenvironment. *Front Immunol.* 2022;13:844142. doi:10.3389/fimmu.2022.844142
118. Grabovska Y, Mackay A, O'Hare P, et al. Pediatric pan-central nervous system tumor analysis of immune-cell infiltration identifies correlates of antitumor immunity. *Nat Commun.* 2020;11(1):4324. doi:10.1038/s41467-020-18070-y
119. Korman AJ, Garrett-Thomson SC, Lonberg N. The foundations of immune checkpoint blockade and the ipilimumab approval decennial. *Nat Rev Drug Discov.* 2022;21(7):509–528. doi:10.1038/s41573-021-00345-8
120. Georger B, Kang HJ, Yalon-Oren M, et al. Pembrolizumab in paediatric patients with advanced melanoma or a PD-L1-positive, advanced, relapsed, or refractory solid tumour or lymphoma (KEYNOTE-051): interim analysis of an open-label, single-arm, phase 1–2 trial. *Lancet Oncol.* 2020;21(1):121–133. doi:10.1016/S1470-2045(19)30671-0
121. Gorski HS, Malicki DM, Barsan V, et al. Nivolumab in the treatment of recurrent or refractory pediatric brain tumors: a single institutional experience. *J Pediatr Hematol Oncol.* 2019;41(4):e235–e241. doi:10.1097/MPH.0000000000001339
122. Georger B, Zwaan CM, Marshall LV, et al. Atezolizumab for children and young adults with previously treated solid tumours, non-Hodgkin lymphoma, and Hodgkin lymphoma (iMATRIX): a multicentre phase 1–2 study. *Lancet Oncol.* 2020;21(1):134–144. doi:10.1016/S1470-2045(19)30693-X
123. Haydar D, Houke H, Chiang J, et al. Cell-surface antigen profiling of pediatric brain tumors: B7-H3 is consistently expressed and can be targeted via local or systemic CAR T-cell delivery. *Neuro Oncol.* 2021;23(6):999–1011. doi:10.1093/neuonc/noaa278
124. Rao P, Furst L, Meyran D, et al. Advances in CAR T cell immunotherapy for paediatric brain tumours. *Front Oncol.* 2022;12:873722. doi:10.3389/fonc.2022.873722
125. Vitanza NA, Johnson AJ, Wilson AL, et al. Locoregional infusion of HER2-specific CAR T cells in children and young adults with recurrent or refractory CNS tumors: an interim analysis. *Nat Med.* 2021;27(9):1544–1552. doi:10.1038/s41591-021-01404-8
126. Theruvath J, Sotillo E, Mount CW, et al. Locoregionally administered B7-H3-targeted CAR T cells for treatment of atypical teratoid/rhabdoid tumors. *Nat Med.* 2020;26(5):712–719. doi:10.1038/s41591-020-0821-8
127. Vitanza NA, Wilson AL, Huang W, et al. Intraventricular B7-H3 CAR T cells for diffuse intrinsic pontine glioma: preliminary first-in-human bioactivity and safety. *Cancer Discov.* 2023;13(1):114–131. doi:10.1158/2159-8290.CD-22-0750
128. Ravanpay AC, Gust J, Johnson AJ, et al. EGFR806-CAR T cells selectively target a tumor-restricted EGFR epitope in glioblastoma. *Oncotarget.* 2019;10(66):7080–7095. doi:10.18632/oncotarget.27389
129. Kramer K, Pandit-Taskar N, Kushner BH, et al. Phase 1 study of intraventricular (131)I-omburtamab targeting B7H3 (CD276)-expressing CNS malignancies. *J Hematol Oncol.* 2022;15(1):165. doi:10.1186/s13045-022-01383-4
130. Ghajar-Rahimi G, Kang KD, Totsch SK, et al. Clinical advances in oncolytic virotherapy for pediatric brain tumors. *Pharmacol Ther.* 2022;239:108193. doi:10.1016/j.pharmthera.2022.108193
131. Mineta T, Rabkin SD, Yazaki T, Hunter WD, Martuza RL. Attenuated multi-mutated herpes simplex virus-1 for the treatment of malignant gliomas. *Nat Med.* 1995;1(9):938–943. doi:10.1038/nm0995-938
132. Friedman GK, Johnston JM, Bag AK, et al. Oncolytic HSV-1 G207 immunovirotherapy for pediatric high-grade gliomas. *N Engl J Med.* 2021;384(17):1613–1622. doi:10.1056/NEJMoa2024947
133. Bernstock JD, Bag AK, Fiveash J, et al. Design and rationale for first-in-human Phase 1 immunovirotherapy clinical trial of oncolytic HSV G207 to treat malignant pediatric cerebellar brain tumors. *Hum Gene Ther.* 2020;31(19–20):1132–1139. doi:10.1089/hum.2020.101

134. Schuelke MR, Gundelach JH, Coffey M, et al. Phase I trial of sargramostim/pelareorep therapy in pediatric patients with recurrent or refractory high-grade brain tumors. *Neurooncol Adv.* 2022;4(1):vdac085. doi:10.1093/noajnl/vdac085
135. Dörig RE, Marciel A, Chopra A, Richardson CD. The human CD46 molecule is a receptor for measles virus (Edmonston strain). *Cell.* 1993;75(2):295–305. doi:10.1016/0092-8674(93)80071-L
136. Anderson BD, Nakamura T, Russell SJ, Peng KW. High CD46 receptor density determines preferential killing of tumor cells by oncolytic measles virus. *Cancer Res.* 2004;64(14):4919–4926. doi:10.1158/0008-5472.CAN-04-0884
137. Fishelson Z, Donin N, Zell S, Schultz S, Kirschfink M. Obstacles to cancer immunotherapy: expression of membrane complement regulatory proteins (mCRPs) in tumors. *Mol Immunol.* 2003;40(2–4):109–123. doi:10.1016/S0161-5890(03)00112-3
138. Müller L, Berkeley R, Barr T, Ilett E, Errington-Mais E-MF. Past, present and future of oncolytic reovirus. *Cancers.* 2020;12(11):3219. doi:10.3390/cancers12113219
139. Setton J, Zinda M, Riaz N, et al. Synthetic lethality in cancer therapeutics: the next generation. *Cancer Discov.* 2021;11(7):1626–1635. doi:10.1158/2159-8290.CD-20-1503
140. Wang X, Wang S, Troisi EC, et al. BRD9 defines a SWI/SNF sub-complex and constitutes a specific vulnerability in malignant rhabdoid tumors. *Nat Commun.* 2019;10(1):1881. doi:10.1038/s41467-019-09891-7
141. Michel BC, D'Avino AR, Cassel SH, et al. A non-canonical SWI/SNF complex is a synthetic lethal target in cancers driven by BAF complex perturbation. *Nat Cell Biol.* 2018;20(12):1410–1420. doi:10.1038/s41556-018-0221-1
142. Knipstein JA, Birks DK, Donson AM, Alimova I, Foreman NK, Vibhakar R. Histone deacetylase inhibition decreases proliferation and potentiates the effect of ionizing radiation in atypical teratoid/rhabdoid tumor cells. *Neuro Oncol.* 2012;14(2):175–183. doi:10.1093/neuonc/nor208
143. Bukowinski A, Chang B, Reid JM, et al. A phase I study of entinostat in children and adolescents with recurrent or refractory solid tumors, including CNS tumors: trial ADVL1513, Pediatric Early Phase-Clinical Trial Network (PEP-CTN). *Pediatr Blood Cancer.* 2021;68(4):e28892. doi:10.1002/pbc.28892
144. Alimova I, Wang D, Danis E, et al. Targeting the TP53/MDM2 axis enhances radiation sensitivity in atypical teratoid rhabdoid tumors. *Int J Oncol.* 2022;60(3). doi:10.3892/ijo.2022.5322
145. Liu Y, Peng X, Zeng T. Development and validation of a nomogram for predicting overall survival in pediatric patients with atypical teratoid/rhabdoid tumors. *Turk Neurosurg.* 2021. doi:10.5137/1019-5149.JTN.33034-20.2
146. Johann PD, Altendorf L, Efreanova EM, et al. Recurrent atypical teratoid/rhabdoid tumors (AT/RT) reveal discrete features of progression on histology, epigenetics, copy number profiling, and transcriptomics. *Acta Neuropathol.* 2023;146(3):527–541. doi:10.1007/s00401-023-02608-7
147. Paassen I, Williams J, Rios Arceo C, et al. Atypical teratoid/rhabdoid tumoroids reveal subgroup-specific drug vulnerabilities. *Oncogene.* 2023;42(20):1661–1671. doi:10.1038/s41388-023-02681-y
148. Pagès M, Rotem D, Gydush G, et al. Liquid biopsy detection of genomic alterations in pediatric brain tumors from cell-free DNA in peripheral blood, CSF, and urine. *Neuro Oncol.* 2022;24(8):1352–1363. doi:10.1093/neuonc/noab299
149. Katsman E, Orlanski S, Martignano F, et al. Detecting cell-of-origin and cancer-specific methylation features of cell-free DNA from Nanopore sequencing. *Genome Biol.* 2022;23(1):158. doi:10.1186/s13059-022-02710-1
150. Patel A, Dogan H, Payne A, et al. Rapid-CNS(2): rapid comprehensive adaptive nanopore-sequencing of CNS tumors, a proof-of-concept study. *Acta Neuropathol.* 2022;143(5):609–612. doi:10.1007/s00401-022-02415-6
151. Euskirchen P, Bielle F, Labreche K, et al. Same-day genomic and epigenomic diagnosis of brain tumors using real-time nanopore sequencing. *Acta Neuropathol.* 2017;134(5):691–703. doi:10.1007/s00401-017-1743-5
152. Bowden R, Davies RW, Heger A, et al. Sequencing of human genomes with nanopore technology. *Nat Commun.* 2019;10(1):1869. doi:10.1038/s41467-019-09637-5
153. Afflerbach AK, Rohrandt C, Brändl B, et al. Classification of brain tumors by nanopore sequencing of cell-free DNA from cerebrospinal fluid. *Clin Chem.* 2023. doi:10.1093/clinchem/hvad115
154. Elghetany MT, J-M H, Shi-Qi LH, et al. Maximizing the potential of aggressive mouse tumor models in preclinical drug testing. *Sci Rep.* 2021;11(1):11580. doi:10.1038/s41598-021-91167-6
155. Han ZY, Richer W, Fréneaux P, et al. The occurrence of intracranial rhabdoid tumours in mice depends on temporal control of Smarcb1 inactivation. *Nat Commun.* 2016;7:10421. doi:10.1038/ncomms10421
156. Roberts CW, Galusha SA, McMenamin ME, Fletcher CD, Orkin SH. Haploinsufficiency of Snf5 (integrase interactor 1) predisposes to malignant rhabdoid tumors in mice. *Proc Natl Acad Sci U S A.* 2000;97(25):13796–13800. doi:10.1073/pnas.250492697
157. Melcher V, Graf M, Interlandi M, et al. Macrophage-tumor cell interaction promotes ATRT progression and chemoresistance. *Acta Neuropathol.* 2020;139(5):913–936. doi:10.1007/s00401-019-02116-7

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