ORIGINAL RESEARCH

Green synthesis and characterization of gelatin-based and sugar-reduced silver nanoparticles

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Correspondence: Majid Darroudi Advanced Materials and Nanotechnology Laboratory, Institute of Advanced Technology (ITMA), Universiti Putra Malaysia, 43400 UPM Serdang, Selangor, Malaysia Tel +603 8946 6044 Fax +603 8946 6043 Email majiddarroudi@gmail.com **Abstract:** Silver nanoparticles (Ag-NPs) have been successfully prepared with simple and "green" synthesis method by reducing Ag⁺ ions in aqueous gelatin media with and in the absence of glucose as a reducing agent. In this study, gelatin was used for the first time as a reducing and stabilizing agent. The effect of temperature on particle size of Ag-NPs was also studied. It was found that with increasing temperature the size of nanoparticles is decreased. It was found that the particle size of Ag-NPs obtained in gelatin solutions is smaller than in gelatin–glucose solutions, which can be related to the rate of reduction reaction. X-ray diffraction, ultraviolet-visible spectra, transmission electron microscopy, and atomic force microscopy revealed the formation of monodispersed Ag-NPs with a narrow particle size distribution. **Keywords:** silver nanoparticles, green method, gelatin, glucose, UV-vis spectra

Introduction

Synthesis of nanomaterials with the desired quality and properties is one of the key issues in current nanotechnology.1 Today, the "green" synthesis of metallic nanoparticles has received increasing attention due to the development of eco-friendly technologies in materials science. Well-dispersed and ultrafine metal nanoparticles, especially transition metals, have great interest due to their distinctive physicochemical and thermodynamic properties, which have made them suitable for use in various fields, such as catalysis,² optics,³ and medicine.⁴ In green synthesis of nanoparticles, three important rules of green chemistry should be considered: (i) choice of the green solvents used in the synthesis, ii) choice of an eco-friendly benign reducing agent, and (iii) choice of a nontoxic material as a stabilizer. To date, most of the preparation methods published are based on using organic materials due to the hydrophobicity of the stabilizing agents used, such as natural polymers. Polymers are a category of macromolecules. Using a polymer as a matrix for preparing metallic nanoparticles, the repeat unit of the polymer should include metal species or have functional groups presented in order to bind the metal. When using polymers as soft support, the diameter of metallic nanoparticles can be logically controlled.

Gelatin is the protein from collagen and has a three-chain helical structure in which individual helical chains are stranded in a super-helix about the common molecular axis.^{5–7} Gelatin contains positively and negatively charged as well as hydrophobic domains folded into a stable, nonnative state that minimizes hydrophobic interactions with water (when dissolved in it). Gelatin can stabilize surfaces by the formation of a steric barrier.⁸ Therefore, the main function of gelatin is as a stabilizer. It is usually used

in applications such as food processing,⁹ the pharmaceutical industry,⁷ photography,¹⁰ and electrochemistry.¹¹

In many cases, the surface passivation reagents such as polymers and/or surfactant molecules are required to prevent the agglomeration of nanoparticles.¹²⁻¹⁶ For example, Polyvinylpyrrolidone (PVP) has been used as an effective passivation agent for preparing silver nanoparticles (Ag-NPs) and other metal nanoparticles.¹⁷ One of the green preparation methods of metallic nanoparticles is the polysaccharide method. In this process, Ag-NPs are synthesized in water as an eco-friendly benign solvent with polysaccharides as the stabilizer agent. In some cases, polysaccharides are used as both a reducing agent and a stabilizer. For example, preparation of Ag-NPs using soluble starch was carried out as a stabilizer with β -D-glucose as a reducing agent in low temperatures.¹⁸ Ag-NPs are prepared by the reduction of silver cations inside the starch templates. The extensive network of hydrogen bonds in the templates can be passivated by the surface nanoparticles and/or can be protected from aggregation.¹⁹ This method must normally be done with a gentle heat system because the binding interactions between polysaccharides and metallic nanoparticles are weak and can be reversible with high heat systems, allowing separation from the obtained particles. In this article, we have reported for the first time on the synthesis of Ag-NPs with gelatin as a reducing agent and capping agent. One of the advantages of this method is being able to use renewable materials like gelatin and glucose, which are eco-friendly agents.

Experimental procedure Materials

Chemicals were analytical grade and used as received without further purification. AgNO₃ (99.98%, Merck KGaA, Darmstadt, Germany), gelatin (type B, Sigma-Aldrich, St Louis, MO, USA), and glucose (BDH Chemical Ltd, Poole, England) were used as silver precursor, capping agent, and reducing agent, respectively. All glassware used in the laboratory experiments was cleaned with a fresh solution of HNO₃/HCl (3:1, v/v), washed thoroughly with doubly distilled water, and dried before use.

Synthesis of gelatin-reduced Ag-NPs

For synthesis of Ag-NPs, 2.0 g of gelatin was added to 190 mL of H_2O in a flask, and the solution was stirred to obtain a clear solution. Aqueous AgNO₃ (10 mL, 1 M) was added to gelatin solution with continuous stirring to obtain Ag⁺/gel-sol. The solution obtained was distributed into three cuvettes, and the prepared solutions were stirred and

maintained for 48 hours at different temperatures, ie, 28 (AG28), 40 (AG40), and 60°C (AG60), respectively.

Synthesis of gelatin–glucose-reduced Ag-NPs

For synthesis of Ag-NPs, 2.0 g of gelatin was added to 190 mL of H_2O in a flask, and the solution was stirred to obtain a clear solution. To obtain the Ag⁺/gelatin solution, the silver solution (10 mL, 1 M) was added to the gelatin solution with continuous stirring. Then, 20 mL of glucose solution (2 M) was added to the Ag⁺/gelatin solution. The solution obtained was distributed into three cuvettes, and the prepared solutions were stirred and maintained for 48 hours at different temperatures, ie, 28 (AGG28), 40 (AGG40), and 60°C (AGG60), respectively. All solutions were kept in dark to avoid any photochemical reactions during the experiment.

Characterization

The Ag-NPs prepared under the various temperatures were characterized by using ultraviolet-visible (UV-vis) spectroscopy, transmission electron microscopy (TEM), X-ray diffraction (XRD), and atomic force microscopy (AFM). The optical absorption properties of prepared samples were characterized using a Lambda 35[®] (PerkinElmer, Waltham, MS, USA) UV-vis spectrophotometer over the range of 300-700 nm. TEM images were obtained with a Hitachi H-7100[®] electron microscope (Hitachi High-Technologies Corporation, Tokyo, Japan), and the particle size distributions of nanoparticles were determined using the UTHSCSA Image Tool® Version 3.00 program (UTHSCSA Dental Diagnostic Science, San Antonio, TX, USA). The XRD patterns were carried out on a Philips X'pert (Cu K_{α}) and were recorded at a scan speed of 2°/min. The AFM image was obtained on an Ambios Q-scope[®] (Ambios Technology, Santa Cruz, CA, USA) (SPM) machine.

Results and discussion

The color of silver solutions in gelatin solutions at various temperature gradually changed from colorless to light brown and then to brown, and in gelatin–glucose solutions changed from colorless to light brown and then to brown and finally dark brown. This indicates the formation of Ag-NPs in both types of solutions. The prepared particles could dissociate due to further laser ablation to form smaller particles stabilized by the amine pendant groups on the gelatin backbone, which leads to the formation of gelatin-stabilized Ag-NPs. Glucose, as the aldehyde, can reduce silver cations to silver atoms and can be oxidized to gluconic acid. The preparation of Ag-NPs was studied by UV-vis spectroscopy, which has proven to be a useful spectroscopic method for the detection of prepared metallic nanoparticles. In UV-vis spectra, the Ag-NPs must display a surface plasmon resonance (SPR) band at around 400 nm. The small shift to the left (blue-shift) or to the right (red-shift) in the λ_{max} of the SPR peak could be related to obtaining Ag-NPs at various shapes, sizes, or solvent dependencies of formed Ag-NPs.

After reaction at 28°C, the Ag-NPs obtained showed a characteristic SPR band for Ag-NPs centered at about 445 nm and 438 nm for AG28 and AGG28, respectively (Figure 1A, D). As shown in Figure 1, the intensity of all the SPR peaks increased as the reaction temperature increased, which indicates the continued reduction of the silver ions, and the increase of the absorbance with the reaction temperature indicates that the concentration of Ag-NPs increased.^{20,21} When the reaction temperatures reached 40°C (Figure 1B,E), the absorbance intensities were increased, and the λ_{max} values were slightly blue-shifted to 443 nm and 431 nm for

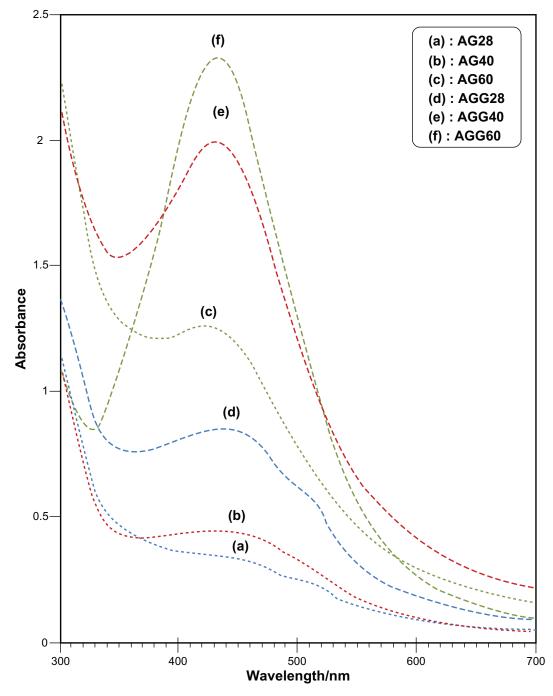


Figure 1 Ultraviolet-visible absorption spectra of Ag-NPs prepared in gelatin and gelatin-glucose solutions.

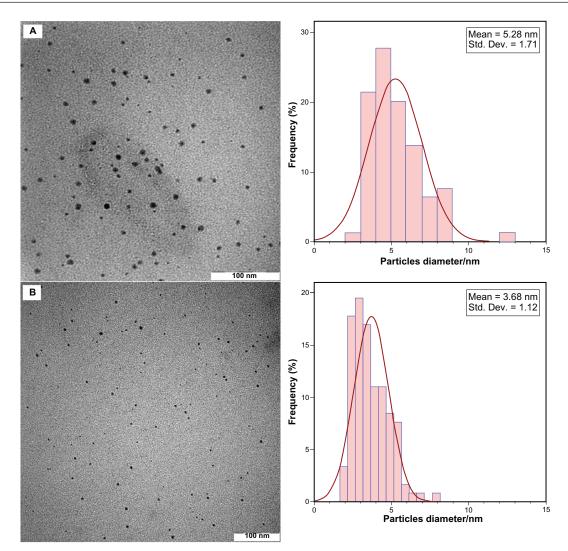


Figure 2 Transmission electron microscopy images and corresponding size distributions of AGG60 (A) and AG60 (B).

AG40 and AGG40, respectively. For AG60 and AGG60 (Figure 1C,F), the absorbance intensities was also increased and blue-shifted to 425 nm and 430 nm, respectively. This phenomenon indicated that the size of particles was decreased, because the absorbance peak due to the SPR of metallic

nanoparticles shows the blue-shift with decreasing particle size.²² The Ag-NPs could dissociate due to heating to form smaller particles stabilized by the amine pendant groups on the gelatin, which leads to the formation of gelatin-stabilized stable Ag-NPs.²³ Figure 2 shows typical TEM images and

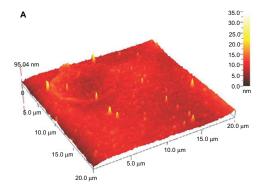
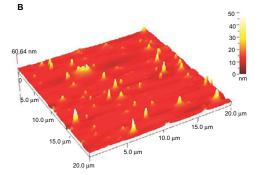


Figure 3 Atomic force microscopy images of AGG60 (A) and AG60 (B).



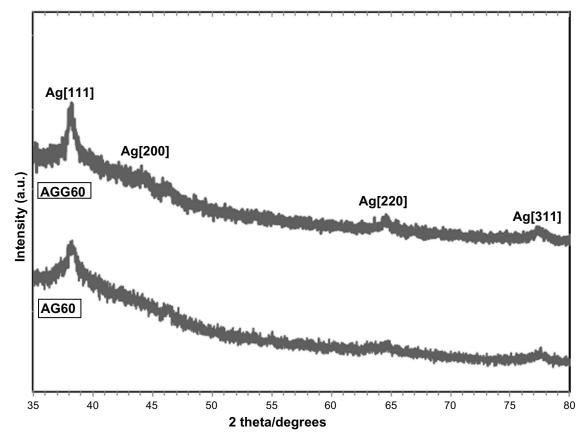


Figure 4 X-ray diffraction patterns of AGG60 and AG60.

the corresponding particle size distribution of the prepared Ag-NPs at different reaction media. The TEM results indicate that the samples obtained in gelatin and gelatin–glucose solutions retained a narrower particle size distribution. It was found that the particle size of Ag-NPs obtained in gelatin solutions is smaller than in the gelatin-glucose solutions, which can be related to rate of reduction reaction. The average size of all prepared Ag-NPs was less than 15 nm, and a smaller average size (about 3.7 nm) was obtained for AG60.

The AFM results display the surface morphology of the monodispersed Ag-NPs formed in gelatin and gelatin–glucose media. As observed in Figure 3, the value determined by the AFM was close to that determined by the TEM, and the films of gelatin and gelatin–glucose containing Ag-NPs showed a dense and uniform packed structure. Thus, the Ag-NPs and gelatin and Ag-NPs and gelatin–glucose films could provide a biocompatible and rough surface for biological uses, eg, cell immobilization. Figure 4 shows the XRD patterns of AG60 and AGG60, which show the synthesis of the silver crystalline structure. The XRD peaks at 20 degrees of 37.9, 44.1, 64.3, and 76.9 and can be attributed to the (111), (200), (220), and (311) crystalline planes of the face centered cubic (fcc) crystalline structure of metallic silver, respectively (JCPDS file No. 00-004-0783).

Conclusion

Briefly, we have investigated a green method to synthesize Ag-NPs in corresponding metal solution using green agents such as gelatin and glucose without special physical conditions. The Ag-NPs obtained in the presence of gelatin (in the absence of glucose) were smaller in particle size, which can be related to the rate of the reduction process. This preparation method is simple and eco-friendly and may be extended to other noble metals, such as gold (Au), palladium (Pd), and platinium (Pt), and may possibly find various additional medicinal, industrial, and technological applications.

Disclosure

The authors report no conflicts of interest in this work.

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