

# Clinical and Microbiological Characteristics of *Klebsiella pneumoniae* Bloodstream Infection in a Chinese Hospital: Hypervirulent and Multiclonal

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**Purpose:** Hypervirulent *Klebsiella pneumoniae* (hvKP) is emerging globally and can cause various infections. This study aimed to investigate the clinical and microbiological characteristics of bloodstream infection (BSI) caused by hvKP.

**Patients and Methods:** The clinical data of hospitalized patients with *K. pneumoniae* BSI were retrospectively analyzed. The *K. pneumoniae* strains were collected and re-identified, and antimicrobial susceptibility testing was performed using the broth microdilution method. Capsular serotypes and virulence genes were detected using polymerase chain reaction, and hvKP was defined as aerobactin positive. Molecular typing was done by multilocus sequence typing. The hvKP and classic *K. pneumoniae* (cKP) subgroups were compared.

**Results:** Of the 66 nonrepetitive BSI *K. pneumoniae* strains included, 29 (43.9%) were hvKP. In these BSI hvKP strains, salmochelin and yersiniabactin accounted for 86.2% and 72.4%, respectively. The prevalence of *rmpA*, *iroBCD* cluster, *ybtS*, *clbA*, and *allS* was 89.7%, 86.2%, 72.4%, 51.7%, and 41.4%, respectively, which were all significantly different between the hvKP and cKP subgroups. Serotypes K1 and K2 were strongly associated with hypervirulence ( $P < 0.05$ ). Nineteen sequence types were scattered in the 29 hvKP strains, and the most common was ST23 (24.1%). None of the hvKP strains were carbapenem resistant. Compared with cKP, hvKP was more capable of developing a liver abscess. However, the 30-day mortality rate was lower (13.8% vs 21.6%) in the hvKP subgroup than in the cKP subgroup.

**Conclusion:** This study demonstrated a high proportion of hvKP in BSI *K. pneumoniae*, most of which were RmpA and siderophore producing, and of multiclonal origin.

**Keywords:** capsular serotypes, hypervirulence, infection, mortality, sequence type

## Introduction

*Klebsiella pneumoniae* is one of the major nosocomial pathogens, which can cause various severe infections, including bloodstream infection (BSI). BSI is a systemic infection and usually results in relatively high mortality due to the virulence factors of *K. pneumoniae*.

Recently, hypervirulent *K. pneumoniae* (hvKP), as a variant of *K. pneumoniae*, has emerged as a major threat to individuals. hvKP has several unique characteristics compared with the classic *K. pneumoniae* (cKP). Hypermucoviscosity is regarded as an important feature of hvKP, which is mainly mediated by *rmpA* and/or *rmpA2* with overproduction of capsular polysaccharide. Previously, hypermucoviscosity has been used to define hvKP.<sup>1,2</sup>

Siderophore production is another distinguishing trait of hvKP. Siderophores are dominantly responsible for iron acquisition, contributing to bacterial virulence. Four kinds of siderophores have been reported in *K. pneumoniae*: aerobactin, salmochelin, yersiniabactin, and enterobactin. Of these, the siderophore aerobactin is dominantly produced (>90%) and has been established as an essential virulence factor in hvKP.<sup>3,4</sup> In addition, hvKP infections are more common in healthy individuals from the community, with invasive and metastatic lesions.

The definition of hvKP has not reached a consensus. The hypermucoviscous phenotype has been used to define hvKP in earlier publications.<sup>1,2</sup> However, this designation of hvKP by the mucoid phenotype alone is imperfect because not all hvKP strains are hypermucoviscous,<sup>5</sup> and some cKP can be hypermucoviscous.<sup>6</sup>

A few recent studies have defined hvKP based on the genetic background.<sup>7–11</sup> Still, many knowledge gaps exist regarding hvKP, and hence the awareness about hvKP needs to be increased urgently. Furthermore, the characteristics of hvKP strains and their differences from cKP are less well-known. Actually, data on hvKP in BSI are extremely limited.<sup>2,7</sup> This retrospective study mainly aimed to investigate the clinical and molecular characteristics of patients with *K. pneumoniae* BSI, especially to delineate the virulence-associated factors. The hvKP and cKP subgroups were compared.

## Patients and Methods

### Patients

Patients hospitalized at the First Hospital of Quanzhou, a teaching hospital located in Eastern China, from January 2019 to March 2020, with a positive blood culture of *K. pneumoniae* were enrolled. Clinical data were extracted from the electronic medical records system, including demographic characteristics, site of infections, underlying diseases, intensive care unit (ICU) stay, length of hospital stay, organ failure, prior hospitalization, and clinical outcomes. The 30-day mortality was used to evaluate the clinical outcomes of *K. pneumoniae* BSI. A community-acquired BSI was defined as a positive blood culture within 48 h after admission, and a hospital-acquired BSI was designated as a positive culture beyond 48 h after admission.<sup>2,10,12</sup> The clinical and microbiological data were compared between patients with cKP BSI and those with hvKP BSI. The present study complied with the Declaration of Helsinki and was approved by the Research Ethics Board at the First Hospital of Quanzhou (212–2018) with a waiver of informed consent for the following reasons: (1) the retrospective nature of the study, (2) the patient data was anonymous, and (3) the strains used in this study were part of the routine hospital laboratory procedure.

### Strains

During the study period, all *K. pneumoniae* strains isolated from blood cultures were collected, and only the first isolation of *K. pneumoniae* from the same patient was included for further assessment. The strains were re-identified using matrix-assisted laser desorption ionization–time-of-flight mass spectrometry (bioMérieux, Marcy-l'Étoile, France) and stored at  $-80^{\circ}\text{C}$  for further use. Strains with aerobactin positive were defined as hvKP.<sup>3,8,9</sup>

### Antimicrobial Susceptibility Testing

Antimicrobial susceptibility testing was carried out using the broth microdilution method, and 15 antimicrobial agents were tested, including cefotaxime, ceftazidime, cefepime, aztreonam, ampicillin-sulbactam, amoxicillin-clavulanic acid, piperacillin-tazobactam, imipenem, meropenem, amikacin, gentamicin, ciprofloxacin, levofloxacin, chloramphenicol, and colistin. The results were interpreted according to the Clinical and Laboratory Standards Institute.<sup>13</sup> *Escherichia coli* ATCC25922 was used as a quality control strain for determining antimicrobial susceptibility.

### Identification of the Hypermucoviscous Phenotype

The string test was used to detect the hypermucoviscous phenotype of *K. pneumoniae* strains as described in previous studies.<sup>1,14</sup> It was considered positive when the viscous string was more than 5 mm in length.

## Detection of Capsular Serotypes and Virulence Genes

Six capsular serotypes (K1, K2, K5, K20, K54, and K57) and 14 virulence genes (including *rmpA/rmpA2* for mucoviscosity, *iroBCD* cluster for siderophore salmochelin, *iucABCD* cluster for siderophore aerobactin, *ybtS* for siderophore yersiniabactin, *clbA/clbB/clbN/clbQ* for colibactin, *wabG* for the biosynthesis of the outer core lipopolysaccharide (LPS), *uge* for biosynthesis of the capsule and smooth LPS, *fimH* for type 1 fimbrial adhesin, *mrkD* for type 3 fimbrial adhesin, and *alls* for allantoin metabolism) were amplified by polymerase chain reaction (PCR) as described in previous studies.<sup>1,10,15–21</sup> The primers used in this study are summarized in [Table S1](#). The PCR products were visualized and analyzed by agarose gel electrophoresis and sequencing.

## Multilocus Sequence Typing

Multilocus sequence typing (MLST) was performed as described in a previous study.<sup>8</sup> New alleles and sequence types (STs), which did not completely match with those of STs in the MLST database, were temporarily designated as a new ST (nST) in this study.

## Statistical Analysis

Categorical variables were recorded as frequency rates and percentages, and continuous variables were expressed as median and interquartile range (IQR) values. SPSS (version 19.0, IBM SPSS Statistics for Windows, IBM Corp., Armonk, New York, United States) was used for analyzing the data. The chi-square test or Fisher's exact test was used for categorical variables. The Student *t*-test or Mann–Whitney *U*-test was used for continuous variables. A *P* value <0.05 indicated a statistically significant difference.

## Results

### Distribution and Prevalence of Virulence Genes in BSI *K. pneumoniae* Strains

During this study period, 66 patients with a positive blood culture of *K. pneumoniae* were included, and 66 nonrepetitive *K. pneumoniae* strains were collected. Among these strains, 29 (43.9%) were positive for siderophore aerobactin and consequently designated as hvKP strains. The remaining 37 (56.1%) *K. pneumoniae* were considered as cKP. Another two siderophores, salmochelin and yersiniabactin, accounted for 53.0% (n = 35) and 54.5% (n = 36), respectively, in all these BSI *K. pneumoniae* strains ([Table 1](#)). Forty-three (65.2%) strains were positive for *rmpA* and/or *rmpA2* (both *rmpA* and *rmpA2*, n = 32; *rmpA* only, n = 2; and *rmpA2* only, n = 9). Of note, 26 of the 32 (81.3%) strains co-carrying *rmpA* and *rmpA2* were hvKP strains, while 11 strains with *rmpA* or *rmpA2* only were all cKP strains. Two genes (*clbA* and *clbB*) of the *pks* colibactin gene cluster together with another two genes (*clbN* and *clbQ*) were simultaneously detected in 18 (27.3%) strains. A relatively low prevalence of *alls* (22.7%, n = 15) was noted. On the contrary, a high prevalence of *uge* (98.5%, n = 65) and *wabG* (84.8%, n = 56) was found. Interestingly, 65 (98.5%) strains had an *mrkD* gene, whereas 26 (39.4%) harbored a *fimH* gene.

As shown in [Table 1](#), the *iroBCD* gene cluster (86.2% vs 27.0%), *ybtS* (72.4% vs 40.5%), *rmpA* (89.7% vs 21.6%), *clbA* (51.7% vs 8.1%), and *alls* (41.4% vs 8.1%) were significantly more prevalent in hvKP than in cKP (*P* ≤ 0.001). Nonetheless, the prevalence of *uge* (96.6% vs 100%), *wabG* (86.2% vs 83.8%), *fimH* (41.4% vs 37.8%), and *mrkD* (96.6% vs 100%) genes was similar in both the hvKP and cKP groups.

### Antimicrobial Susceptibility of BSI *K. pneumoniae*

The tested antibiotics had good antibacterial activity against the BSI *K. pneumoniae* strains in vitro ([Table 2](#)). The susceptibility of hvKP to seven β-lactam agents was significantly higher than that of cKP (*P* < 0.05). Notably, no hvKP strain was found resistant to imipenem and meropenem.

### String Test

A total of 29 (43.9%) BSI *K. pneumoniae* strains were string test positive in the present study. The positive rate was 75.9% (n = 22) in hvKP strains and 18.9% (n = 7) in cKP strains (*P* < 0.001, [Table 1](#)), suggesting that defining hvKP by string test only was not suitable.

**Table 1** Microbiological Characteristics of *K. pneumoniae* Causing Bloodstream Infection

	No. (%) of Strains			
	Total (n=66)	hvKP (n=29)	cKP (n=37)	P value <sup>a</sup>
Age, median (IQR), y	63 (50–74)	58 (50–73)	64 (47–77)	0.936
Virulence genes				
<i>iucABCD</i> cluster	29 (43.9)	29 (100)	0 (0)	<0.001
<i>iroBCD</i> cluster	35 (53.0)	25 (86.2)	10 (27.0)	<0.001
<i>ybtS</i>	36 (54.5)	21 (72.4)	15 (40.5)	<0.001
<i>rmpA</i>	34 (51.5)	26 (89.7)	8 (21.6)	<0.001
<i>clbA</i>	18 (27.3)	15 (51.7)	3 (8.1)	<0.001
<i>allS</i>	15 (22.7)	12 (41.4)	3 (8.1)	0.001
<i>uge</i>	65 (98.5)	28 (96.6)	37 (100)	0.439
<i>wabG</i>	56 (84.8)	25 (86.2)	31 (83.8)	>0.999
<i>fimH</i>	26 (39.4)	12 (41.4)	14 (37.8)	0.770
<i>mrkD</i>	65 (98.5)	28 (96.6)	37 (100)	0.439
String test				
Positive	29 (43.9)	22 (75.9)	7 (18.9)	<0.001
Negative	37 (56.1)	7 (24.1)	30 (81.1)	<0.001
Serotypes				
K1	14 (21.2)	12 (41.4)	2 (5.4)	<0.001
K2	12 (18.2)	9 (31.0)	3 (8.1)	0.017
K5	3 (4.5)	0 (0)	3 (8.1)	0.250
K20	1 (1.5)	1 (3.4)	0 (0)	0.439
K54	1 (1.5)	0 (0)	1 (2.7)	>0.999
K57	2 (3.0)	2 (6.9)	0 (0)	0.189
K-nontypable	33 (50.0)	5 (17.2)	28 (75.7)	<0.001

**Notes:** <sup>a</sup>P values indicate differences between hvKP and cKP. P < 0.05 was considered statistically significant.

**Abbreviations:** hvKP, hypervirulent *K. pneumoniae*; cKP, classic *K. pneumoniae*; IQR, interquartile range.

## Serotype Identification

Of the 66 BSI *K. pneumoniae* strains, 14 (21.2%) K1, 12 (18.2%) K2, 3 (4.5%) K5, 1 (1.5%) K20, 1 (1.5%) K54, and 2 (3.0%) K57 capsular serotypes were identified. In hvKP strains, K1, K2, K5, K20, K54, and K57 capsular serotypes accounted for 41.4% (n = 12), 31.0% (n = 9), 0 (n = 0), 3.4% (n = 1), 0 (n = 0), and 6.9% (n = 2), respectively, and only 5 (17.2%) strains were K-nontypable (Table 1). Nevertheless, the prevalence of these six serotypes was quite low (5.4%, 8.1%, 8.1%, 0, 2.7%, and 0) in cKP BSI *K. pneumoniae* strains (Table 1). Taken together, these results suggested a good correlation between hypervirulence and K1/K2 capsular serotypes.

**Table 2** Antimicrobial Susceptibility of Hypervirulent and Classic *K. pneumoniae* Causing BSI

	No. (%) of Susceptible Strains			
	Total (n=66)	hvKP (n=29)	cKP (n=37)	P value <sup>a</sup>
Cefotaxime	54 (81.8)	29 (100)	25 (67.6)	0.001
Ceftazidime	55 (83.3)	29 (100)	26 (70.3)	0.002
Cefepime	57 (86.4)	29 (100)	28 (75.7)	0.004
Aztreonam	58 (87.9)	29 (100)	29 (78.4)	0.007
Ampicillin-sulbactam	17 (77.3)	13 (96.6)	4 (62.2)	0.002
Amoxicillin-clavulanic acid	54 (81.8)	29 (100)	25 (67.6)	0.001
Piperacillin-tazobactam	58 (87.9)	29 (100)	29 (78.4)	0.007
Imipenem	63 (95.5)	29 (100)	34 (91.9)	0.25
Meropenem	63 (95.5)	29 (100)	34 (91.9)	0.25
Amikacin	64 (97.0)	29 (100)	35 (94.6)	0.50
Gentamicin	62 (93.9)	29 (100)	33 (89.2)	0.125
Ciprofloxacin	54 (81.8)	27 (93.1)	27 (73.0)	0.035
Levofloxacin	59 (89.4)	28 (96.6)	31 (83.8)	0.124
Chloramphenicol	55 (83.3)	25 (86.2)	30 (81.1)	0.743
Colistin	66 (100)	29 (100)	37 (100)	NA

**Notes:** <sup>a</sup>P values indicate differences between hvKP and cKP.  $P < 0.05$  was considered statistically significant.

**Abbreviations:** BSI, bloodstream infection; hvKP, hypervirulent *K. pneumoniae*; cKP, classic *K. pneumoniae*; NA, not available.

## MLST

Among the 66 BSI *K. pneumoniae* strains, 43 had known STs (involving 30 STs), and the remaining 23 strains each had an nST (Table S2), indicating high diversity of these BSI strains. The most common ST was ST23 (n = 7, 10.6%) belonging to clonal complex (CC) 23, followed by ST86 (n = 3). Four strains (6.1%) were categorized as CC65, containing two ST65, one ST25, and one ST375. CC23 and CC65 were the most common CCs in this study.

Nineteen STs were scattered in the 29 hvKP strains, including ST23 (n = 7), ST65 (n = 2), ST86 (n = 2), ST380 (n = 2), ST412 (n = 2), and other 14 distinct STs (Table S2), while 39 different STs were distributed in the 39 cKP strains (Table S2).

Interestingly, of the 14 *K. pneumoniae* strains with K1 serotype (hvKP, n = 12), 7 (50%) belonged to ST23, which were all hvKP strains (Table S2), indicating a strong correlation between ST23 and K1 serotype in hvKP strains. Differently, among the 12 K2 serotype strains (9 hvKP and 3 cKP), ST86 (n = 3, 25%) was the most common ST, followed by ST380 (n = 2) and ST65 (n = 2). In a word, the present study indicated an association between certain STs and K serotypes (eg, ST23-K1 and ST86-K2).

## Clinical Characteristics of BSI Caused by *K. pneumoniae* Strains

The demographic and clinical data of the 66 enrolled patients with *K. pneumoniae* BSI are summarized in Table 3. These patients were divided into hvKP and cKP subgroups based on their corresponding isolated *K. pneumoniae* strains. Of the 66 patients, the median age was 63 years (ranging from 0 to 90 years; IQR, 50–74), and 39 patients (59.1%) were male. Forty-eight (72.7%) patients had a community-acquired BSI, of which 24 were in the hvKP subgroup. Although the percentage of the community-acquired BSI in the hvKP subgroup was higher than that in the cKP subgroup (82.8% vs 64.9%, respectively), no significant difference existed between the two subgroups ( $P > 0.05$ ; Table 3).

**Table 3** Clinical Characteristics of Patients with *K. pneumoniae* Bloodstream Infection

	No. (%) of Patients			P value <sup>a</sup>
	Total (n=66)	With hvKP (n =29)	With cKP (n =37)	
Age, median (IQR), y	63 (50–74)	58 (50–73)	64 (47–77)	0.936
Sex				
Male	39 (59.1)	15 (51.7)	24 (64.9)	0.281
Female	27 (40.1)	14 (48.3)	13 (35.1)	0.281
Community-acquired				
Hospital-acquired	48 (72.7)	24 (82.8)	24 (64.9)	0.105
Hospital-acquired				
Abscess	18 (27.3)	5 (17.2)	13 (35.1)	0.105
Abscess				
Liver abscess	17 (25.8)	13 (44.8)	4 (10.8)	0.002
Liver abscess				
Skin abscess	14 (21.2)	12 (41.4)	2 (5.4)	<0.001
Skin abscess				
Peripancreatic abscess	1 (1.5)	1 (3.4)	0 (0)	0.439
Peripancreatic abscess				
Abdominal wall abscess	1 (1.5)	0 (0)	1 (2.7)	>0.999
Abdominal wall abscess				
Other types of infections				
Pneumonia	1 (1.5)	0 (0)	1 (2.7)	>0.999
Pneumonia				
Urinary tract infection	26 (39.4)	10 (34.5)	16 (43.2)	0.470
Urinary tract infection				
Abdominal infection	15 (22.7)	5 (17.2)	10 (27.0)	0.346
Abdominal infection				
Skin and soft tissue infection	10 (15.2)	1 (3.4)	9 (24.3)	0.045
Skin and soft tissue infection				
≥2 sites of infection	3 (4.5)	2 (6.9)	1 (2.7)	0.578
≥2 sites of infection				
≥3 sites of infection	52 (78.8)	23 (79.3)	29 (78.4)	0.901
≥3 sites of infection				
Underlying diseases				
Diabetes	19 (28.8)	3 (10.3)	16 (43.2)	0.003
Diabetes				
Hypertension	11 (16.7)	3 (10.3)	8 (21.6)	0.375
Hypertension				
Coronary heart disease	7 (10.6)	4 (13.8)	3 (8.1)	0.690
Coronary heart disease				
Chronic kidney disease	6 (9.1)	3 (10.3)	3 (8.1)	>0.999
Chronic kidney disease				
Cancer	15 (22.7)	5 (17.2)	10 (27.0)	0.346
Cancer				
Hospitalization in recent 90 days	19 (28.8)	3 (10.3)	16 (43.2)	0.003
Hospitalization in recent 90 days				
ICU stay	20 (30.3)	9 (31.0)	11 (29.7)	0.909
ICU stay				
Septic shock	18 (27.3)	7 (24.1)	11 (29.7)	0.613
Septic shock				
Organ failure	14 (21.2)	5 (17.2)	9 (24.3)	0.485
Organ failure				
Median duration of hospital stay (IQR)	22.3 (9–24.5)	15.5 (9–22)	27.8 (9.5–29.8)	0.138
Median duration of hospital stay (IQR)				
Outcome				
Discharged	54 (81.8)	25 (86.2)	29 (78.4)	0.413
Discharged				
Died	12 (18.2)	4 (13.8)	8 (21.6)	0.413
Died				
Readmitted in the following 90 days	13 (24.1)	6 (24.0)	7 (24.1)	0.858
Readmitted in the following 90 days				

**Notes:** <sup>a</sup>P values indicate differences between hvKP and cKP. P < 0.05 was considered statistically significant.

**Abbreviations:** hvKP, hypervirulent *K. pneumoniae*; cKP, classic *K. pneumoniae*; IQR, interquartile range; NT, not tested.

Seventeen (25.8%) patients developed abscesses, including liver abscess (n = 14), skin abscess (n = 1), peripancreatic abscess (n = 1), and abdominal wall abscess (n = 1). The percentage of patients with a liver abscess was significantly higher in the hvKP subgroup than in the cKP subgroup ( $P < 0.001$ ), indicating the ability of hvKP to develop a liver abscess. Besides abscesses, pneumonia (n = 26), urinary tract infection (n = 15), abdominal infection (n = 10), and skin and soft tissue infection (n = 3) afflicted 39 patients (Table 3), which were all more common in the cKP subgroup, although no statistical difference except for abdominal infection was found between the two groups. Actually, most (n = 52, 78.8%) of the 66 patients had  $\geq 2$  sites of infections.

Underlying diseases were noted in all 66 patients (Table 3), including diabetes (n = 38, 57.6%), hypertension (n = 11, 16.7%), coronary heart disease (n = 7, 10.6%), chronic kidney disease (n = 6, 9.1%), and cancer (n = 15, 22.7%). Among these underlying diseases, diabetes, coronary heart disease, and chronic kidney disease were more prevalent in the hvKP subgroup, while hypertension and cancer were more prevalent in the cKP subgroup (Table 3). However, no significant statistical differences existed between the hvKP and cKP subgroups in these diseases ( $P > 0.05$ ). Interestingly, of the 38 patients with diabetes, 12 (31.6%) developed a liver abscess. Among the remaining 28 patients without diabetes, only 2 (7.1%) developed a liver abscess ( $P = 0.016$ ), demonstrating a strong association between diabetes and liver abscess.

Additionally, 19 (28.8%) patients had a history of hospitalization within 90 days prior to admission. A significant difference in hospitalization within recent 90 days was found between the hvKP subgroup and the cKP subgroup, with 10.3% and 43.2%, respectively ( $P = 0.003$ ; Table 3).

Although no statistical significance was noted, the cKP subgroup seemed to have a higher percentage of ICU stay, septic shock, and organ failure (30.3%, 27.3%, and 21.2%, respectively) and longer median hospital stay in this study (Table 3), suggesting that patients with cKP were prone to develop a severe disease and have a longer hospital stay.

The 30-day mortality rate of *K. pneumoniae* BSI was 18.2% in this study, and it was higher (21.6%) in the cKP subgroup (Table 3). After discharge from the hospital, 13 (24.1%) patients were readmitted to the hospital in the following 90 days, with 6 in the hvKP subgroup and 7 in the cKP subgroup (Table 3). Of them, four patients (4/13, 30.8%) were due to bacterial infection, including three (3/6, 50%) in the hvKP subgroup and one (1/7, 14.3%) in the cKP subgroup.

## Discussion

The present retrospective study provided insights into the clinical and microbiological characteristics of patients with *K. pneumoniae* BSI, especially revealing the distribution of virulence genes in hvKP. A relatively high percentage (43.9%) of hvKP in BSI *K. pneumoniae* strains was found to carry multiple virulence genes in this study, and these pathogens were of multiclonal origin.

The hypermucoviscosity and the iron acquisition systems (siderophores) are common in hvKP strains, which are considered as the main features of hvKP. In the present study, strains with *iucA* gene (for siderophore aerobactin) were defined as hvKP, and a relatively higher proportion of BSI hvKP was found compared with previous studies (21.6–31.4%).<sup>2,7,10</sup> The prevalence of hvKP in BSI was relatively high in China, but varied in different regions, which might partially result from the geographical difference and the inconsistent definition of hvKP. Recently, Harada et al revealed that 25.5% (n = 26) of BSI *K. pneumoniae* strains at hospitals across Japan were hvKP (defining hvKP as positive for any of *rmpA*, *rmpA2*, *iroBCDN*, *iucABCD*, and *iutA*).<sup>22</sup>

Previous studies showed that various virulence factors could concurrently be present in hvKP.<sup>3</sup> This study revealed that BSI hvKP strongly related to *rmpA*, *iroBCD* cluster, *ybtS*, *clbA*, and *allS*. The siderophore aerobactin is a dominant and critical siderophore for hvKP, which plays an important role in systemic infection.<sup>3</sup> In this study, 43.9% of BSI hvKP strains carried the *iucABCD* cluster, which was consistent with a recent study from South Korea (41.5% of BSI *K. pneumoniae* carried *iucA* gene).<sup>23</sup> Lan et al from China reported that a lower percentage of BSI *K. pneumoniae* was *iucA* positive (28.9%).<sup>10</sup> In this study, more than half of the BSI *K. pneumoniae* were also found to possess genes for other two siderophores (salmochelin and yersiniabactin). The exact role of these siderophores in BSI hvKP needs to be further investigated.

Colibactin, which is encoded by the *pks* gene cluster and exerts as genotoxins, was first found in *E. coli* and was associated with BSI.<sup>24</sup> The presence of the *pks* gene cluster has been related to early mortality in patients with BSI caused by *K. pneumoniae*.<sup>23</sup> The real role of colibactin in the pathogenesis of hvKP remains to be explored. Colibactin was detected in 27.3% of BSI *K. pneumoniae* strains in the present study, consistent with the studies by Lan et al and

Kim et al (26.8% and 27.3%, respectively).<sup>10,23</sup> Putze et al reported a much lower rate (3.5%) of clinical *K. pneumoniae* being colibactin positive in Europe.<sup>25</sup> Additionally, the colibactin-positive rate was 16.7% in *K. pneumoniae* isolated from various clinical samples in Taiwan.<sup>26</sup>

Interestingly, in this study, all colibactin-producing hvKP strains except one co-carried *ybtA*. Kim et al reported that 27.3% (n = 158) of BSI *K. pneumoniae* strains co-harbored the *pks* gene cluster and *ybtA*.<sup>23</sup> Additionally, although allantoin metabolism genes do not seem to be important for systemic infection,<sup>6</sup> this study demonstrated 22.7% of BSI *K. pneumoniae* strains with *allS*, suggesting a potential role of *allS* in BSI. Taken together, more studies are needed to clarify the real role of these virulence factors in BSI.

The present study suggested the association between hypervirulence and K1/K2/K20/K57 capsular serotypes, consistent with other studies from China.<sup>2,8,9</sup> Additionally, this study revealed that 72.4% of hvKP strains were K1 and K2 capsular serotypes, while only 13.5% of cKP belonged to these two serotypes. These findings indicated that K1 and K2 capsular serotypes played key roles in hvKP BSI. K5 and K54 capsular serotypes were not detected in BSI hvKP strains in the present study, consistent with previous studies.<sup>2,7</sup>

In this study, the BSI-causing *K. pneumoniae* strains were of multiclonal origin, despite the most common ST (ST23) accounting for 10.6%. Similarly, Liu et al reported that 42 STs were identified in the 70 BSI *K. pneumoniae* strains.<sup>2</sup> Wu et al demonstrated that more than 36 STs were established in 79 BSI *K. pneumoniae* strains.<sup>27</sup>

This study revealed that hvKP strains were more susceptible to tested antibiotics than cKP, consistent with previous studies.<sup>1,8,10</sup> The confluence of hypervirulence and multidrug resistance, especially carbapenem resistance, is extremely alarming. Although carbapenem-resistant hvKP has been described in previous studies,<sup>7,9,28–30</sup> it was not found in the present study. This might be partially attributed to the regional difference.

Although various infections had developed in patients with BSI in the present study, a liver abscess was more likely to be associated with hvKP, suggesting the capability of hvKP to develop a liver abscess, which was in agreement with previous studies.<sup>1,8,11</sup> The results of this study also indicated a strong association between diabetes and liver abscess (mainly by hvKP, 85.7%), in accordance with an earlier study.<sup>16</sup> Thus, it is reasonable to conclude that patients with diabetes may be prone to suffer from hvKP liver abscess and develop hvKP BSI afterward. This was in line with a theory that diabetes is a risk factor for an hvKP infection, and the confluence of diabetes and liver abscess is likely to develop secondary infections.<sup>31</sup> Although diabetes was identified as an independent risk factor for hvKP BSI in a previous study,<sup>7</sup> more studies are needed to clarify this.

Although no significant difference in the 30-day mortality rate was noted between cKP and hvKP subgroups, the percentage of mortality was higher in the cKP BSI subgroup than in the hvKP BSI subgroup in the present study, consistent with earlier studies in patients with BSI.<sup>2,7</sup> Recently, a study reported that the hypermucoviscosity (as a marker for defining hvKP in several studies) displayed an inverse association with 30-day mortality in BSI *K. pneumoniae* strains.<sup>23</sup> Similarly, Hwang et al recently reported that the 30-day mortality in patients with cKP pneumonia (26.9%) was higher than that in patients with hvKP pneumonia (17.9%).<sup>32</sup> This might be attributed to patients in the cKP subgroup usually having more and much severe underlying diseases, as well as greater age. Additionally, a higher percentage (43.2% in cKP vs 10.3% in hvKP) of a history of hospitalization in recent 90 days may be partially contributed to the higher mortality in the cKP subgroup. Finally, in the present study cKP BSI strains were more resistant to the common empiric antibiotics and then likely caused treatment failure, which might also be contributed to the higher mortality in those patients.<sup>33</sup> More prospective studies are needed to confirm the real role of hvKP in the mortality of patients with BSI.

This study had several limitations. First, the presence of the aerobactin gene was detected, and hvKP was defined on the basis of aerobactin positivity. However, the aerobactin production was not assessed. Further studies defining hvKP by assessing the quantitative production of aerobactin would be warranted, and studies in vitro and in vivo may be needed to confirm the real virulence of *K. pneumoniae*. Second, this was a single-center experience with a relatively small study sample. Multicenter studies with a larger population are required.

## Conclusions

This study demonstrated a high proportion of hvKP in BSI *K. pneumoniae*. These hvKP strains had a strong correlation with various virulence factors and were of multiclonal origin. It provided insights into the clinical and microbiological

features of BSI caused by *K. pneumoniae* strains, highlighting the significance of clinical awareness, and might be helpful in the clinical management of hvKP BSI.

## Ethics Statement

This study was approved by the Research Ethics Board at the First Hospital of Quanzhou (2012–2018) with a waiver of informed consent.

## Acknowledgments

This study was supported by the funding from the Science and Technology Project of Quanzhou (NO. 20181N043S to S. Chen), the National Natural Science Foundation of China (NO. 81703567 to J. Li, and NO. 81603166 to Z. Sheng), and the Shanghai Municipal Key Clinical Specialty (Infectious disease, YW20190002 to Q. Xie).

## Disclosure

The authors declare no conflicts of interest in this work.

## References

1. Li W, Sun G, Yu Y, et al. Increasing occurrence of antimicrobial-resistant hypervirulent (hypermucoviscous) *Klebsiella pneumoniae* isolates in China. *Clin Infect Dis*. 2014;58:225–232. doi:10.1093/cid/cit675
2. Liu YM, Li BB, Zhang YY, et al. Clinical and molecular characteristics of emerging hypervirulent *Klebsiella pneumoniae* bloodstream infections in mainland China. *Antimicrob Agents Chemother*. 2014;58:5379–5385. doi:10.1128/AAC.02523-14
3. Russo TA, Olson R, MacDonald U, et al. Aerobactin, but not yersiniabactin, salmochelin, or enterobactin, enables the growth/survival of hypervirulent (Hypermucoviscous) *Klebsiella pneumoniae* ex vivo and in vivo. *Infect Immun*. 2015;83:3325–3333. doi:10.1128/IAI.00430-15
4. Russo TA, Olson R, Macdonald U, et al. Aerobactin mediates virulence and accounts for increased siderophore production under iron-limiting conditions by hypervirulent (hypermucoviscous) *Klebsiella pneumoniae*. *Infect Immun*. 2014;82:2356–2367. doi:10.1128/IAI.01667-13
5. Shi Q, Lan P, Huang D, et al. Diversity of virulence level phenotype of hypervirulent *Klebsiella pneumoniae* from different sequence type lineage. *BMC Microbiol*. 2018;18:94. doi:10.1186/s12866-018-1236-2
6. Russo TA, Marr CM. Hypervirulent *Klebsiella pneumoniae*. *Clin Microbiol Rev*. 2019;32:e00001–e00019. doi:10.1128/CMR.00001-19
7. Li J, Ren J, Wang W, et al. Risk factors and clinical outcomes of hypervirulent *Klebsiella pneumoniae* induced bloodstream infections. *Eur J Clin Microbiol Infect Dis*. 2018;37:679–689. doi:10.1007/s10096-017-3160-z
8. Zhang Y, Zhao C, Wang Q, et al. High prevalence of hypervirulent *Klebsiella pneumoniae* infection in China: geographic distribution, clinical characteristics, and antimicrobial resistance. *Antimicrob Agents Chemother*. 2016;60(10):6115–6120. doi:10.1128/AAC.01127-16
9. Liu C, Shi J, Guo J. High prevalence of hypervirulent *Klebsiella pneumoniae* infection in the genetic background of elderly patients in two teaching hospitals in China. *Infect Drug Resist*. 2018;11:1031–1041. doi:10.2147/IDR.S161075
10. Lan Y, Zhou M, Jian Z, et al. Prevalence of pks gene cluster and characteristics of *Klebsiella pneumoniae* -induced bloodstream infections. *J Clin Lab Anal*. 2019;33:e22838. doi:10.1002/jcla.22838
11. Wu H, Li D, Zhou H, et al. Bacteremia and other body site infection caused by hypervirulent and classic *Klebsiella pneumoniae*. *Microb Pathog*. 2017;104:254–262. doi:10.1016/j.micpath.2017.01.049
12. Shi Q, Quan J, Lan P, et al. Prevalence and characteristics of pks gene cluster harbouring *Klebsiella pneumoniae* from bloodstream infection in China. *Epidemiol Infect*. 2020;148:e69. doi:10.1017/S0950268820000655
13. CLSI. *Performance Standards for Antimicrobial Susceptibility Testing*. 30th ed. CLSI supplement M100. Wayne, PA: CLSI; 2020.
14. Zhou C, Wu Q, He L, et al. Clinical and molecular characteristics of carbapenem-resistant hypervirulent *Klebsiella pneumoniae* Isolates in a tertiary hospital in Shanghai, China. *Infect Drug Resist*. 2021;14:2697–2706. doi:10.2147/IDR.S321704
15. Fang CT, Lai SY, Yi WC, et al. *Klebsiella pneumoniae* genotype K1: an emerging pathogen that causes septic ocular or central nervous system complications from pyogenic liver abscess. *Clin Infect Dis*. 2007;45:284–293. doi:10.1086/519262
16. Ye M, Tu J, Jiang J, et al. Clinical and genomic analysis of liver abscess-causing *Klebsiella pneumoniae* identifies new liver abscess-associated virulence genes. *Front Cell Infect Microbiol*. 2016;6:165. doi:10.3389/fcimb.2016.00165
17. Yeh KM, Kurup A, Siu LK, et al. Capsular serotype K1 or K2, rather than *magA* and *rmpA*, is a major virulence determinant for *Klebsiella pneumoniae* liver abscess in Singapore and Taiwan. *J Clin Microbiol*. 2007;45:466–471. doi:10.1128/JCM.01150-06
18. Turton JF, Baklan H, Siu LK, et al. Evaluation of a multiplex PCR for detection of serotypes K1, K2 and K5 in *Klebsiella* sp. and comparison of isolates within these serotypes. *FEMS Microbiol Lett*. 2008;284:247–252. doi:10.1111/j.1574-6968.2008.01208.x
19. Bachman MA, Oyler JE, Burns SH, et al. *Klebsiella pneumoniae* yersiniabactin promotes respiratory tract infection through evasion of lipocalin 2. *Infect Immun*. 2011;79:3309–3316. doi:10.1128/IAI.05114-11
20. Yu WL, Ko WC, Cheng KC, et al. Association between *rmpA* and *magA* genes and clinical syndromes caused by *Klebsiella pneumoniae* in Taiwan. *Clin Infect Dis*. 2006;42:1351–1358. doi:10.1086/503420
21. Luo Y, Wang Y, Ye L, et al. Molecular epidemiology and virulence factors of pyogenic liver abscess causing *Klebsiella pneumoniae* in China. *Clin Microbiol Infect*. 2014;20:O818–O824. doi:10.1111/1469-0691.12664
22. Harada S, Aoki K, Yamamoto S, et al. Clinical and molecular characteristics of *Klebsiella pneumoniae* isolates causing bloodstream infections in Japan: occurrence of hypervirulent infections in health care. *J Clin Microbiol*. 2019;57:e01206–e01219. doi:10.1128/JCM.01206-19
23. Kim D, Park BY, Choi MH, et al. Antimicrobial resistance and virulence factors of *Klebsiella pneumoniae* affecting 30 day mortality in patients with bloodstream infection. *J Antimicrob Chemother*. 2019;74:190–199. doi:10.1093/jac/dky397

24. Johnson JR, Johnston B, Kuskowski MA, et al. Molecular Epidemiology and Phylogenetic Distribution of the *Escherichia coli pks* Genomic Island. *J Clin Microbiol.* 2008;46:3906–3911. doi:10.1128/JCM.00949-08
25. Putze J, Hennequin C, Nougayre JP, et al. Genetic structure and distribution of the colibactin genomic island among members of the family Enterobacteriaceae. *Infect Immun.* 2009;77:4696–4703. doi:10.1128/IAI.00522-09
26. Chen YT, Lai YC, Tan MC, et al. Prevalence and characteristics of *pks* genotoxin gene cluster-positive clinical *Klebsiella pneumoniae* isolates in Taiwan. *Sci Rep.* 2017;7:43120. doi:10.1038/srep43120
27. Wu X, Shi Q, Shen S, et al. Clinical and bacterial characteristics of *Klebsiella pneumoniae* affecting 30-day mortality in patients with bloodstream infection. *Front Cell Infect Microbiol.* 2021;11:688989. doi:10.3389/fcimb.2021.688989
28. Zhang Y, Jin L, Ouyang P, et al. Evolution of hypervirulence in carbapenem-resistant *Klebsiella pneumoniae* in China: a multicentre, molecular epidemiological analysis. *J Antimicrob Chemother.* 2020;75:327–336. doi:10.1093/jac/dkz446
29. Yao H, Qin S, Chen S, et al. Emergence of carbapenem-resistant hypervirulent *Klebsiella pneumoniae*. *Lancet Infect Dis.* 2018;18:25. doi:10.1016/S1473-3099(17)30628-X
30. Gu D, Dong N, Zheng Z, et al. A fatal outbreak of ST11 carbapenem-resistant hypervirulent *Klebsiella pneumoniae* in a Chinese hospital: a molecular epidemiological study. *Lancet Infect Dis.* 2018;18:37–46. doi:10.1016/S1473-3099(17)30489-9
31. Choby JE, Howard-Anderson J, Weiss DS. Hypervirulent *Klebsiella pneumoniae*-clinical and molecular perspectives. *J Intern Med.* 2020;287:283–300. doi:10.1111/joim.13007
32. Hwang JH, Handigund M, Hwang JH, et al. Clinical features and risk factors associated with 30-day mortality in patients with pneumonia caused by hypervirulent *Klebsiella pneumoniae* (hvKP). *Ann Lab Med.* 2020;40:481–487. doi:10.3343/alm.2020.40.6.481
33. Chan KS, Shelat VG. *Klebsiella pneumoniae* bacteremia is associated with higher mortality in acute calculous cholangitis as compared to *Escherichia coli* bacteremia. *World J Surg.* 2022;46:1678–1685. doi:10.1007/s00268-022-06559-0

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