

Relationship Between Antibiotic Resistance, Biofilm Formation, and Biofilm-Specific Resistance in *Escherichia coli* Isolates from Ningbo, China

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Purpose: Several *Escherichia coli* pathotypes still constitute an important public health concern owing to its pathogenicity and antimicrobial resistance. Moreover, biofilm formation of *E. coli* can allow the strains to interfere with host and antimicrobial eradication, thus conferring additional resistance. The association between the formation of biofilm and antimicrobial resistance determinants has been extensively exploited; nevertheless, there is still no definite conclusion. The purpose of this study was to provide additional data to augment the present knowledge about the subject.

Methods: Antibiotic resistance/susceptibility profiles of 81 isolates from pediatric individuals in China between 2011 and 2014 against 20 antibiotics were assessed using the VITEK 2 system. Biofilm-forming capacities were evaluated using the crystal violet staining method, confocal laser scanning microscopy (CLSM), and field emission scanning electron microscopy. Biofilm compositions inside the biofilm formed by representative strains were assessed using CLSM. The effects of antibiotics on biofilms generated by *E. coli* strains of different biofilm-forming ability were examined using CLSM in combination with gatifloxacin. The relationships between antibiotic resistance, biofilm formation, and biofilm-specific resistance in *E. coli* isolates were investigated.

Results: The results showed that 23 isolates were classified as multidrug-resistant, and 57 isolates were classified as extensively drug-resistant (XDR). Among the 69 isolates with the ability to form biofilms, 46 isolates were stronger biofilm formers. Correlation analysis demonstrated that strain populations exhibiting more robust biofilm formation likely contained larger proportions of XDR isolates.

Conclusion: Together, our study implies that there was an association between biofilm-formation and resistance to several antibiotics for XDR-*E. coli* isolates, and would provide novel insights regarding the prevention and treatment against *E. coli*-related infections.

Keywords: *Escherichia coli*, biofilm, antibiotic resistance, biofilm-specific resistance

Introduction

Antimicrobial agents play an important part in the treatment of bacterial infections to significantly reduce mortality and morbidity globally.¹ However, the overuse of antibiotics in human and veterinary medicine has resulted in the selection of bacteria that are resistant to currently available antibiotics; in turn, this has allowed the propagation of drug-resistant strains in hospitals and communities, which poses a global challenge to public health.²⁻⁵ Among these infectious agents, one such bacterium is *Escherichia coli* (*E. coli*), a gram-negative, facultative anaerobic microbe that constitutes an intestinal commensal bacterium of humans and animals.⁶ However, infectious pathologies of *E. coli* also act as an opportunistic or true pathogen causing illnesses such as intestinal, extra-intestinal, urinary tract, and gastrointestinal infections, along with meningitis, peritonitis, and septicemia.⁷ For instance, uropathogenic *E. coli* (UPEC) is the most

frequent cause of urinary tract infections (UTIs) and a common cause of severe, life-threatening sepsis.^{8,9} Especially, in the past two decades, *E. coli* has surged to the primary cause of early-onset bloodstream infection (BSI) in preterm infants, and the common and recognized cause of late-onset BSI.^{10–13} Comparatively, *E. coli*-caused BSIs are markedly associated with enhanced mortality compared with those triggered by gram-positive bacteria in hospitalized infants.¹⁴ In addition, *E. coli* can lead to the potentially severe complication, including hemolytic uremic syndrome and acute diarrhea, which occurs most frequently in young children (age 5 and younger).^{15,16}

Antibiotic resistance primarily derives from changes, or mutations, in the DNA of the bacteria in addition to acquisition of antibiotic resistance genes from other bacterial species through plasmid transfer. Particularly, *E. coli* can exhibit a powerful ability to obtain antibiotic resistance determinants, resulting in few remaining effective treatment options against its pathogenic variants.¹⁷ For instance, previous studies displayed that the *E. coli* ST131 clone is associated with resistance to multiple classes of antibiotics, including fluoroquinolones, third-generation cephalosporins and more recently last-line carbapenems and polymyxins.^{9,18–20} Fighting *E. coli*-related infections has been compromised by the increased emergence of resistance to various first-line antibiotics. Decreased susceptibility of *E. coli* to cephalosporins, including imipenem, meropenem, and ertapenem, among members of *Enterobacteriaceae* has emerged mainly due to the diffusion of extended-spectrum β -Lactamases (ESBLs) such as CTX-M, TEM, and SHV, and metallo- β -Lactamases (MBLs).^{21,22} Recently, the emergence of ESBLs-producing *E. coli* has become a major concern and their emergence in infectious disease.²³ MBLs are founded increasingly in gram-negative organisms and are recognized mostly in *Pseudomonas* and *Klebsiella* species.^{24,25} The emergence of *Enterobacteriaceae* harboring ESBL has resulted in reliance on colistin treatment in humans. Further, since a plasmid-borne colistin resistance gene (*mcr-1*) was detected in an *E. coli* isolate from China, several families of genes (*mcr-1*, *mcr-2*, *mcr-3*, *mcr-4*, and *mcr-5*) in *E. coli* isolates have been reported worldwide. Meanwhile, *qnr* and *aac(6′)-Ib-cr* genes are known to mediate the resistance to quinolones in *Enterobacteriaceae*, and are commonly found on the same resistance plasmids as ESBL, and thus, could represent a serious challenge for the public health.

Various kinds of bacteria can form biofilms as a survival strategy, distinct from other multidrug resistance mechanisms; this is especially relevant with regard to *E. coli*, which represents one of the most common causes of UTIs,²⁶ and can cause biofilm-related contamination of medical devices, such as intravascular catheters, urinary catheters, and orthopedic implants.^{27,28} A biofilm is an assemblage of surface-associated microbial cells that are embedded in a self-produced matrix of extracellular polymeric substances. Importantly, bacterial cells within biofilms exhibit a set of emergent properties that differ substantially from their planktonic (freely suspended) counterparts.²⁹ The biofilm formation ability is a feature common to most microorganisms in natural and medical systems, constituting a protected mode of growth that allows survival in the associated relatively harsh environmental conditions.³⁰ Previous studies have demonstrated that extracellular matrix of biofilms aids in the adherence of *E. coli* to host cells, resists the shear forces associated with urine flow and contributes to the persistence of bacteria and chronicity of infection.^{31,32} Accordingly, biofilms were reported to be a major factor contributing to many chronic inflammatory diseases and involved in numerous serious consequences for public health.^{33,34} In China, some studies have reported the isolation of *E. coli* strains from humans; however, few studies have described *E. coli* strains isolated from infants and children younger than four years of age in conjunction with the associated biofilm formation ability.^{35,36}

In addition, the relationship between antibiotic resistance and biofilm-production has attracted considerable interests from researchers. Previous studies have pointed out that biofilm formation can be induced by certain antibiotics under conditions of low doses, suggesting that regulation of biofilms may be involved in the global response to external stress, including antibiotics.³⁷ Nevertheless, previous studies regarding quantitative correlation between biofilm formation and antibiotic resistance have yielded different and inconsistent results between different bacterial species, leaving researchers further explore the association.^{37,38} In this study, we investigated the relationship between antibiotic resistance, biofilm-formation, and biofilm-specific resistance in *E. coli* isolates derived from Chinese infants and children younger than four years of age in Ningbo. In addition, the possible relationship between the antibiotic resistance and the ability of biofilm-formation among these *E. coli* isolates was examined. The results could provide a foundation for further research regarding the potential mechanisms of biofilm-mediated enhanced resistance in *E. coli*.

Materials and Methods

Bacterial Strains and Growth Conditions

A total of 81 clinical strains were collected in the children's hospital in Ningbo, Zhejiang province, China during 2011–2014 (Table S1). The range of age of the pediatric patients was from 11 minutes after birth to 4-year-old child; 34.6% were female and 65.4% were male. All isolates were grown at 37 °C and identified as *E. coli* using Matrix-assisted laser desorption/ionization-time of flight (MALDI-TOF) mass spectrometry (MS) and Clinpro tools software (Bruker Biotyper; Bruker Daltonics, Bremen, Germany).³⁹ The *E. coli* ATCC 19606 was used as the reference strain. One single bacterial colony was inoculated into a test tube containing 3 mL of tryptic soy broth (TSB) and grown at 37°C.

Antibacterial Susceptibility Testing with *E. coli* Isolates

The antibacterial susceptibility testing was performed using the disk diffusion and the VITEK 2 system (bioMérieux, Marcy l'Etoile, France) and interpreted by the protocol specified in the guidelines of Clinical and Laboratory Standards Institute (CLSI, 2015). The antibacterial agents included cefoxitin, ceftriaxone, cefotaxime, cefepime, ceftazidime, cefazolin, moxifloxacin, ofloxacin, levofloxacin, gentamicin, amikacin, amoxicillin, minocycline, piperacillin, azithromycin, nitrofurantoin, polymyxin B, and meropenem (Thermo Fisher Scientific, USA). MDR isolates were defined as resistant to at least three different classes of antibiotics. Isolates were defined as extensively drug resistant (XDR) if they were not susceptible to at least one agent in all but two or fewer antibacterial categories (ie, bacterial isolates remained susceptible to only one or two antibacterial categories).

Biofilm Formation Assays

Biofilm production was assessed using a 96-well microtiter plate assay. After overnight growth in TSB, 150 µL of cell suspension were adjusted to the turbidity of a 0.5 McFarland standard and transferred into each microtiter plate well and incubated at 37 °C for 36 h. After three brief washes with 10 mM phosphate-buffered saline (PBS) solution and a 20-min fixation step with 180 µL methanol, the plate was stained with 180 µL 0.1% (v/v) crystal violet (CV) for 15 min and washed gently three times with 10 mM PBS. The formed biofilms were then dissolved with 180 µL 33% (v/v) acetic acid for 30 min. Sterile TSB was used as the negative control. The biofilm formation was measured at 550 nm optical density (OD₅₅₀) using a microtiter plate reader (Thermo Fisher Scientific, Vantaa, Finland). The biofilm assays were performed in triplicate. According to the cut-off ODc value (the cut-off ODc value = the mean OD of the negative control + three standard deviations (SD) of the negative control), the biofilms were divided into the following four categories: OD ≤ ODc (non-biofilm producers), ODc < OD ≤ 2 ODc (low-biofilm producers), 2 ODc < OD ≤ 4 ODc (medium-biofilm producers), and OD > 4 ODc (strong-biofilm producers).⁴⁰

Detection of Resistance Genes

Genomic DNA of *E. coli* isolates were extracted using the genomic DNA extraction kit (Magen Biotech, Guangzhou, China). All isolates were tested by PCR amplification of resistance genes that confer resistance to β-lactams (*bla*_{-TEM}, *bla*_{-SHV}, *bla*_{-OXA}, *bla*_{-PSE}, and *bla*_{-CTX-M}), quinolones (*qnrA*, *qnrB*, *qnrS*, *acc* (6')-Ib-cr, and *qepA*), aminoglycosides (*acc* (6')-Ib, *aac* (3)-I, *aac* (3)-II, *aac* (3)-III, *aac* (3)-IV, and *ant* (2'')), tetracyclines (*tetA* and *tetB*), and polymyxins (*mcr-1*, *mcr-2*, *mcr-3*, *mcr-4*, and *mcr-5*). PCR primers were synthesized by Sangon Co., Ltd. (Shanghai, China), and shown in Table S2.

Confocal Laser Scanning Microscopy (CLSM) Analysis of Biofilm Structure

CLSM (LSM800, Carl Zeiss AG, Oberkochen, Germany) was applied to observe biofilm structure.⁴¹ After incubation overnight at 37°C, 1 mL of cell suspensions (approximately 1×10⁶ CFU/mL) were added into each well of a 24-well microtiter plate containing a sterile coverslip and then cultured at 37 °C under 50 rpm for 36 h to form biofilms. Subsequently, the biofilms on the glass coverslips were washed three times with 10 mM PBS to remove unattached cells, and then stained with 5 µM SYTO 9 (Thermo Fisher Scientific, USA) for 15 min. Biofilms were visualized and imaged using CLSM, where the excitation/emission wavelength was 483/502 nm for SYTO 9.

Field Emission Scanning Electron Microscope (FESEM) Observations of Biofilms

For visualization of biofilms, biofilms of representative isolates were observed by FESEM (Nova Nano SEM-450, FEI, Hillsboro, OR, USA).⁴¹ One mL of cell suspension of *E. coli* isolates (approximately 1×10^6 CFU/mL) was transferred into each well of a 24-well microtiter plate containing a sterile glass coverslip, cultured at 37 °C under shaking conditions of 50 rpm for another 36 h, and fixed with 2.5% (v/v) glutaraldehyde solution for 4 h at 4 °C. The biofilms were then dehydrated in a series of washes with 30, 50, 70, and 90% ethanol for 10 min each, followed by 15 min rinses in 100% ethanol. Air-dried samples were immediately sputter-coated with platinum and subjected to FESEM.

CLSM Analysis of Biofilm Composition

CLSM analysis of the composition within biofilms was performed according to Qian et al with a few modifications.⁴¹ The biofilms of representative *E. coli* isolates were grown on a 24-well microtiter plate with glass coverslips for 36 h at 37 °C with the stirring speed of 50 rpm. Following incubation, the biofilms were rinsed with 10 mM PBS and stained with three types of fluorescent markers for 15 min in the dark: (i) 4', 6-diamidino-2-phenylindole (DAPI, Sigma-Aldrich, USA) to stain extracellular DNA (eDNA); (ii) Film Tracer SYPRO Ruby biofilm matrix stain (SYPRO Ruby, Invitrogen, Paisley, UK), which labels most classes of proteins; and (iii) wheat germ agglutinin (WGA, Invitrogen, Paisley, UK), which stains N-acetyl-D-glucosamine residues. Finally, samples were washed with 10 mM PBS to remove residual dyes and observed under CLSM. The fluorescences were detected using the following combination of laser excitation and emission band-pass wavelengths: 358/461 nm for DAPI, 450/610 nm for SYPRO Ruby, and 495/519 for WGA.

Diffusion Assay of Gatifloxacin Within Biofilms

To evaluate the diffusion of the gatifloxacin within biofilms produced by the strong, medium and weak biofilm producer of *E. coli*, gatifloxacin with the intrinsic fluorescence was used to evaluate antibiotics diffusion within the biofilm by CLSM.⁴¹ The biofilms were formed on the glass coverslips inside each well of a 24-well microtiter plate and cultured at 37 °C, 50 rpm for 36 h. The coverslips were washed gently three times with 10 mM PBS, and gatifloxacin was added at a final concentration of 0.4 mg/mL, and the coverslips were further incubated at 37 °C for 4 h. Next, 5 µM SYTO 9 was added and co-incubated for 15 min, and the biofilms were then washed three times with 10 mM PBS and observed using CLSM. The emission peak for gatifloxacin was recorded at 495 nm upon excitation at 291 nm. Finally, three random fields were visualized for each biofilm, and representative images were captured.

Statistical Analysis

Spearman's rank correlation tests were used for intergroup comparisons. The Wilcoxon rank sum test was used for comparison of biofilm formation between isolates susceptible/non-susceptible to each antimicrobial category. The chi-square test was used to analyze the correlation between biofilm formation ability, antibiotic resistance genes, and antibiotic resistance. Data analyses were performed using SPSS (version 20.0 SPSS Statistics, Inc., IBM, Armonk, NY, USA). $p < 0.05$ was considered statistically significant for all tests.

Results and Discussion

Antimicrobial Susceptibility of Planktonic *E. coli* Isolates

The resistance patterns of *E. coli* isolates to the tested antimicrobial agents are presented in [Figure 1B](#) and [Table 1](#). The phenotypic resistance profiles of the *E. coli* isolates were as follows: amoxicillin, 96.30% (78/81); cefotaxime, 69.14% (56/81); ceftriaxone, 66.67% (54/81); cefazolin, 64.20% (52/81); cefepime, 61.73% (50/81); azithromycin, 61.73% (50/81); moxifloxacin, 50.62% (41/81); ceftazidime, 33.33% (27/81); ofloxacin, 30.86% (25/81); gentamicin, 29.63% (24/81); levofloxacin, 24.69% (20/81); minocycline, 22.22% (18/81); nitrofurantoin, 14.81% (12/81); cefoxitin, 6.17% (5/81); amikacin, 6.17% (5/81), and piperacillin, 3.70% (3/81). The relatively high resistance rates for cephalosporins and ofloxacin among 81 *E. coli* isolates are similar to several recent reports, indicating that the resistance levels of uropathogenic *E. coli* against fluoroquinolones and cephalosporins in Iran is ranging between 15–66% and 30–60%, respectively.^{32,42} Moreover, the highest and the relatively lower resistance rate among 81 *E. coli* isolates was related to

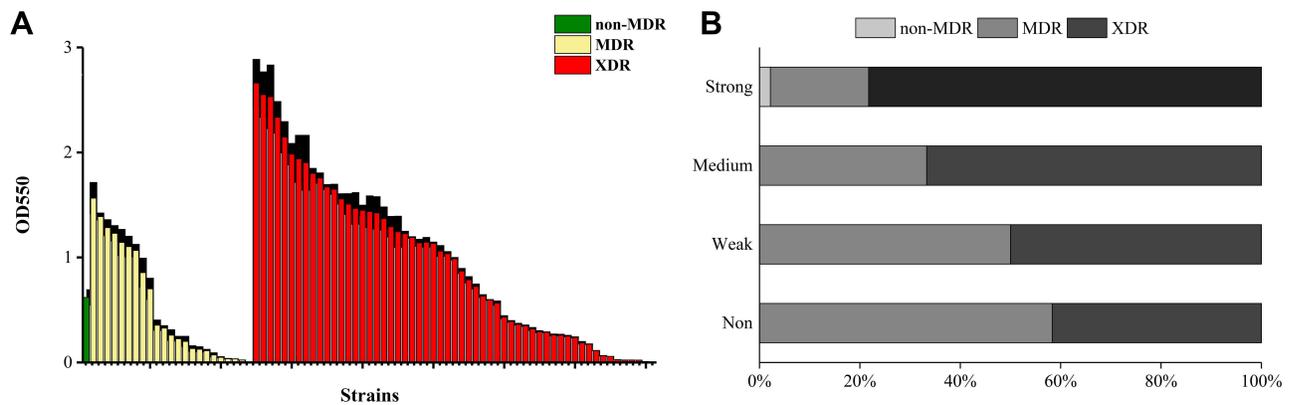


Figure 1 (A) The level of biofilm formation was assessed for *E. coli* isolate. (B) Distribution of resistance phenotypes among different biofilm production capacities displayed as a percentage stacked bar graph. Strains were divided into three groups according to their antibiotic resistance phenotypes: non-multidrug-resistant (non-MDR), multidrug-resistant (MDR), and extensively drug-resistant (XDR). Optical density at 550 nm (OD₅₅₀) represents biofilm forming capacity.

amoxicillin, as well as to piperacillin and amikacin, respectively, which is inconsistent with the studies performed by others.^{43,44} These studies demonstrated that resistance to piperacillin and amikacin antibiotics was lower than other antibiotics. For instance, Allami et al reported that the uropathogenic *E. coli* isolates from southern Iraq were most resistant to piperacillin, ticarcillin, amoxicillin/clavulanic acid (92%, 91%, and 88%) and most sensitive to amikacin and imipenem, respectively. Accordingly, both piperacillin and amikacin may be recommended as the last-line option for clinical treatment of infections caused by *E. coli* when there is no effective treatment available in Ningbo. In addition, a worrisome finding from this study is a relatively high resistance rate for nitrofurantoin (14.81%) in isolates of *E. coli*. This is similar to a previous study demonstrating that *E. coli* isolates were resistant to nitrofurantoin (21%),⁴⁵ but in contrary to a few previous studies with a low prevalence of nitrofurantoin resistance in *E. coli* isolates.^{46,47} The potential explanation of these resistance to antibiotics could be attributed to the inappropriate and excessive use of these antibiotics in the empirical treatment for *E. coli*-related infections in different parts of the world.⁴⁸ Additionally, according to the limited data, antibiotic resistance rates of cefotaxime, cefepime, ceftazidime, and ofloxacin were higher in Zhejiang (69.14, 61.73, 33.33, and 30.86%) than in Shanghai city (24.5, 23.5, 12.4, and 3.4%), respectively.³⁵ Thus, it is crucial to evaluate the local resistance rates of specific pathogens to antibiotics, and area-specific data on antimicrobial resistance could help physicians in prescribing choices for empirical antibiotic treatment, thereby reducing overuse of antibiotics for the treatment of infections. Meanwhile, among the 81 *E. coli* isolates tested, all isolates were resistant to at least one antibiotics; no isolates were resistant to all of the 18 antibiotics. Specifically, 23 isolates were classified as MDR, and 57 isolates were classified as XDR. As such, 98.77% (80/81) of the isolates were either MDR or XDR, indicating the prevalence rate of MDR or XDR isolates in Ningbo was higher in comparison to in North America and Europe. Such high resistance in *E. coli* isolates may be attributed to clinical pattern of overuse and inappropriate use of antibiotics in previous years.⁴⁹

The Relationship Between Antibiotic Resistance and Resistance Genes

The frequency of resistance genes is shown in Table 2. Of the *E. coli* isolates analyzed, 61.7% (50/81) carried one or more resistance genes. Among the 20 resistance genes investigated, *aac* (3)-II, *acc* (6')-Ib-cr, and *ant* (2'') were most frequently detected, demonstrating 32.10 (26/81), 20.99 (17/81), and 20.99% (17/81) frequency, respectively. The frequency of the other resistance genes was as follows: *tetA* (12.35%, 10/81), *bla*_{-TEM} (8.64%, 7/81), *bla*_{-OXA} (1.23%, 1/81), *qepA* (1.23%, 1/81), and *aac* (3)-IV (1.23%, 1/81). By contrast, *bla*_{-SHV}, *bla*_{-PSE}, *bla*_{-CTX-M}, *qnrA*, *qnrB*, *qnrS*, *aac* (3)-I, *aac* (3)-III, *acc* (6')-Ib, and *tetB* were not detected in *E. coli* isolates. Our results with low prevalence of ESBL-producing genes are inconsistent with several previous studies in different regions of the world, including Saudi Arabia, Vietnam, China, Iran, and Mexico, which highlighted the CTX-M, SHV, and TEM genes as the most critical mechanisms of ESBL production in *E. coli* strains.⁵⁰ The prevalence of *bla*_{TEM} and *bla*_{OXA} genes was more common in our settings compared with *bla*_{CTX-M} and *bla*_{SHV} genes. This is similar to the report by Sid Ahmed et al, revealing that the recent

Table 1 Correlation Between the Level of Biofilm Formation and Resistance to 16 Antibiotics in *E. coli* Clinical Isolates

Antimicrobial Category	Antimicrobial Agent	N (%) / OD ₅₅₀			r _s	p-value
		S	I	R		
Cephalosporins	Cefoxitin	74 (91.36%) / 0.617 (0.165, 1.289)	2 (2.47%) / 1.122 (0.292)	5 (6.17%) / 1.450 (0.914, 1.982)	0.233	0.036
	Ceftriaxone	21 (25.93%) / 0.281 (0.076, 1.035)	6 (7.41%) / 0.920 (0.153, 1.727)	54 (66.67%) / 1.024 (0.266, 1.507)	0.253	0.020
	Cefotaxime	19 (23.46%) / 0.609 (0.149, 1.232)	6 (7.41%) / 0.167 (0.018, 0.630)	56 (69.14%) / 0.933 (0.257, 1.488)	0.140	0.211
	Cefepime	28 (34.57%) / 0.472 (0.104, 1.231)	3 (3.70%) / 1.169 (0.426, 1.310)	50 (61.73%) / 0.384 (0.253, 1.507)	0.141	0.209
	Ceftazidime	2 (2.47%) / 1.773 (1.409)	52 (64.20%) / 0.593 (0.118, 1.213)	27 (33.33%) / 1.000 (0.267, 1.571)	0.116	0.304
	Cefazolin	29 (35.80%) / 0.367 (0.112, 1.107)	0 (0.00%) / -	52 (64.20%) / 1.066 (0.263, 1.519)	0.247	0.026
Quinolones and fluoroquinolones	Moxifloxacin	22 (27.16%) / 0.501 (0.052, 1.285)	18 (22.22%) / 0.492 (0.110, 1.180)	41 (50.62%) / 1.048 (0.279, 1.513)	0.189	0.091
	Ofloxacin	53 (65.43%) / 0.609 (0.122, 1.294)	3 (3.70%) / 0.042 (0.023, 0.245)	25 (30.86%) / 1.130 (0.376, 1.548)	0.187	0.094
	Levofloxacin	54 (66.67%) / 0.501 (0.125, 1.286)	7 (8.64%) / 0.326 (0.024, 1.647)	20 (24.69%) / 1.089 (0.464, 1.469)	0.187	0.094
Aminoglycosides	Gentamicin	56 (69.14%) / 0.501 (0.129, 1.234)	1 (1.23%) / 1.409	24 (29.63%) / 1.117 (0.300, 1.639)	0.236	0.034
	Amikacin	72 (88.89%) / 0.804 (0.248, 1.400)	4 (4.94%) / 0.398 (0.061, 1.171)	5 (6.17%) / 0.058 (0.023, 1.694)	-0.153	0.171
β-lactams	Piperacillin	65 (80.25%) / 0.626 (0.217, 1.331)	13 (16.05%) / 0.599 (0.048, 1.198)	3 (3.70%) / 1.952 (1.696)	0.051	0.652
	Amoxicillin	0 (0.00%) / -	3 (3.70%) / 0.626 (0.149)	78 (96.30%) / 0.834 (0.196, 1.589)	0.008	0.941
Tetracyclines	Minocycline	52 (64.20%) / 0.714 (0.154, 1.400)	11 (13.58%) / 0.333 (0.128, 1.048)	18 (22.22%) / 1.015 (0.369, 1.427)	0.049	0.661
Macrolides	Azithromycin	31 (38.27%) / 0.802 (0.204, 1.353)	0 (0.00%) / -	50 (61.73%) / 0.674 (0.171, 1.412)	-0.009	0.939
Nitrofurans	Nitrofurantoin	30 (37.04%) / 0.674 (0.196, 1.245)	39 (48.15%) / 0.609 (0.171, 1.525)	12 (14.81%) / 0.926 (0.158, 1.440)	0.050	0.660
Lipopeptides	Polymyxin B	78 (96.30%) / 0.762 (0.196, 1.358)	3 (3.70%) / 0.245 (0.023)	0 (0.00%) / -	-0.067	0.552
Carbapenems	Meropenem	80 (98.77%) / 0.762 (0.179, 1.560)	1 (1.23%) / 1.409	0 (0.00%) / -	-0.186	0.096

Abbreviations: OD₅₅₀, optical density at 550 nm; IQR, data shown in median interquartile range; R, resistance; I, intermediate; S, susceptible.

Table 2 The Frequency of Antibiotic Resistance Genes Among 81 Isolates

Antibiotic Resistance Gene	Percent
<i>bla</i> _{-TEM}	8.64%, 7/81
<i>bla</i> _{-OXA}	1.23%, 1/81
<i>acc</i> (6')- <i>IB-cr</i>	20.99%, 17/81
<i>qepA</i>	1.23%, 1/81
<i>tetA</i>	12.35%, 10/81
<i>aac</i> (3)- <i>II</i>	32.10%, 26/81
<i>aac</i> (3)- <i>IV</i>	1.23%, 1/81
<i>ant</i> (2'')	20.99%, 17/81

emergence of CTX-M group was predominantly mediated through the mutations of *bla*_{TEM} and *bla*_{SHV} genes.⁵¹ The resistance gene *tetA* (12.35%) detection rate of clinical *E. coli* isolates was lower, but the *aac* (3)-*IV* (1.23%) was higher than that reported by Momtaz et al.⁵² In addition, the percentage of isolates carrying the *bla*_{-TEM} gene was 8.64% in the present study, which is lower than that in a previous report by Nawaz et al.⁵³

The relationship between resistance genes and antibiotic resistance of *E. coli* isolates was further analyzed (Figure 2). Isolates containing the *acc* (6')-*IB-cr* gene were significantly more likely to belong to strains resistant to moxifloxacin and ofloxacin than susceptible strains ($p < 0.001$) or those exhibiting intermediate resistance (moxifloxacin: $p < 0.01$; ofloxacin: $p < 0.05$). Moreover, the detection rate of *acc* (3')-*II* gene in gentamicin-resistant strains was significantly higher than that of intermediate ($p < 0.001$) or susceptible strains ($p < 0.01$). These indicated that antibiotic resistance genes do confer the ability of resistant strains to survive compared with susceptible or intermediate strains.

The Association Between Biofilm Formation and Antibiotic Resistance

The biofilm production ability of MDR or XDR strains enhances the overall resistance, potentially resulting in treatment failure. In this context, awareness of the definite relationship between antibiotic resistance and biofilm formation ability of bacteria could help clinicians in prescribing regimes that specifically address biofilm to the treatment arsenal against MDR or XDR infections. The correlation between biofilm formation and the pattern of antibiotic resistance has been the focus of significant research in recent years; however, to date, definite conclusions cannot be drawn regarding the relationship since there have been some studies

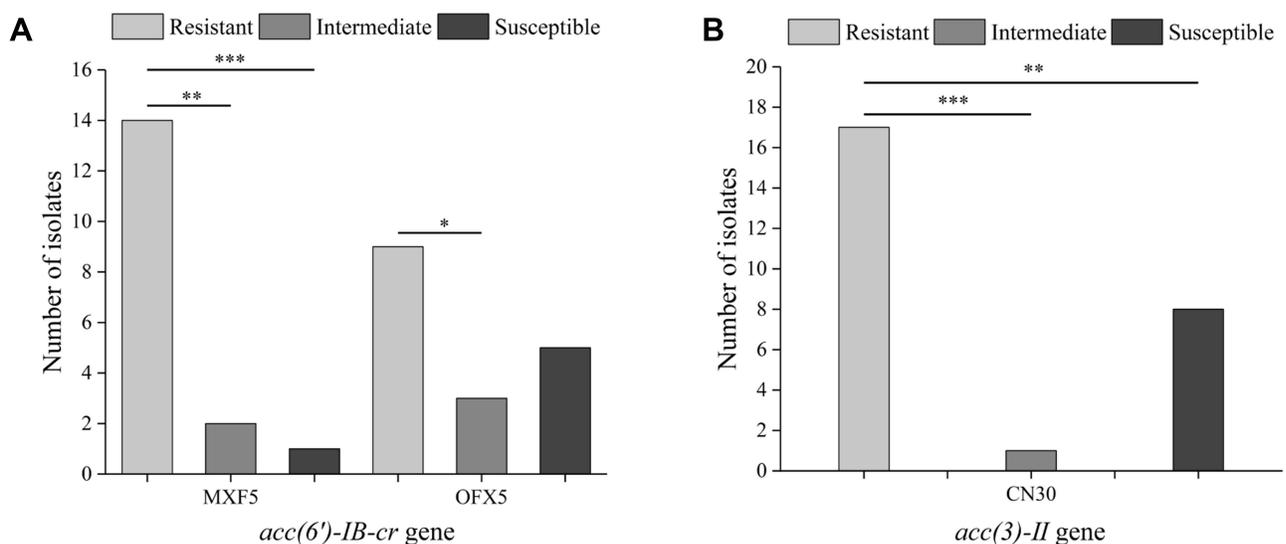


Figure 2 Relationship between resistance genes and antibiotic resistance. Correlation between the *acc* (6')-*IB-cr* (A) and *acc* (3)-*II* (B) gene and antibiotic resistance. MXF5, moxifloxacin; OFX5, ofloxacin; CN30, gentamicin. Statistics were achieved by the chi-square test (** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$).

that have reported conflicting results.³² In this study, the biofilm-forming capacity of *E. coli* isolates was summarized in Figure 1A. Sixty-nine isolates (85.19%, 69/81) exhibited the ability to produce biofilms, among which 46 isolates (56.79%, 46/81) were strong biofilm producers, 15 isolates (18.52%, 15/81) were medium biofilm producers, 8 isolates (9.88%, 8/81) represented weak biofilm producers, and 12 isolates (14.81%, 12/81) were non-biofilm producers. Among the 46 strong-biofilm producers, only 2.17% (1/46) were non-MDR isolates, 19.57% (9/46) constituted MDR isolates, and 78.26% (36/46) represented XDR isolates. The 15 medium-biofilm producers comprised 33.33% (5/15) MDR and 66.67% (10/15) XDR isolates. By contrast, the 8 weak-biofilm producers consisted of 50.00% MDR and 50.00% XDR isolates. The 12 strains that were negative for biofilm formation consisted of 58.33% (7/12) MDR and 41.67% (5/12) XDR isolates (Figure 1B). These compositional proportions indicated that XDR isolates tended to form stronger biofilms, revealing that the populations that exhibited more robust biofilm formation likely contained larger proportions of XDR isolates. The Spearman correlation coefficient (r_s) for this comparison was 0.280 ($p < 0.01$). Similarly, Dumaru et al, found that 62.7% of isolates were biofilm-producers in gut bacteria, and that there was strong association between the MDR-status and biofilm-production.⁵⁴ Avila-Novoa et al explored the correlation between biofilm-production and MDR in *Acinetobacter baumannii* (*A. baumannii*) using the phenotypic method, and found that 73.3% of isolates were biofilm-producers; however, in their study, there is no clear association between biofilm-formation and specific antibiotic resistance was revealed.⁵⁵ Next, XDR *E. coli* isolates tended to form stronger biofilms than MDR and non-MDR strains; this conclusion was also confirmed by statistical analyses ($r_s = 0.243$, $p < 0.05$; Table 2), indicating a positive correlation between biofilm formation capacity and antibiotic resistance phenotypes. Similarly, XDR *E. coli* isolates revealed a greater potential to be strong biofilm producers than MDR or non-MDR *E. coli* isolates.⁵⁶ These results were in agreement with a previous study displaying that biofilm-forming *E. coli* isolates from clinical isolates in Uganda were more resistant than the non-biofilm formers with 64% being MDR as compared to 36% among the non-biofilm forming *E. coli* isolates. Similarly, among 130 *E. coli* isolates from patients having UTI symptoms in Iran, 80 (61.53%, 80/130) were able to form biofilms, and maximum resistance to ampicillin (87.5%), followed by tetracycline (75%), nalidixic acid (72.5%) and co-trimoxazole (71.25%).⁵⁷ These observations indicate that the biofilm formation by *E. coli* isolates tested was potentially associated with MDR bacteria, as well as made their eradication difficult, suggesting that exploiting the factors associated with biofilm formation is key to the development of new therapies.⁵⁸

Meanwhile, we also investigated the relationship between biofilm formation and specific antibiotic resistance. The results revealed that isolates resistant to ceftazidime, ceftazidime-clavulanic acid, ceftriaxone, cefazolin, and gentamicin could form stronger biofilms than those susceptible or exhibiting intermediate resistance, being indicative of a positive correlation between biofilm quantity and resistance profile ($r_s = 0.233-0.253$, $p < 0.05$; Table 3). A similar report demonstrated that gentamicin-resistance and ceftazidime-resistance were related to biofilm-formation in *E. coli*.⁵⁹ Moreover, it was previously reported that, for *E. coli* strains with strong biofilm formation ability were more resistant to amoxicillin-clavulanic acid, norfloxacin, cotrimoxazole, gatifloxacin, and gentamicin, suggesting that the biofilms formed by these strains provide the ability to survive when exposed to these antibiotics.⁶⁰ The correlation between biofilm formation and resistance to the eight antimicrobial categories was also evaluated. For six of the categories, including cephalosporins and quinolones, fluoroquinolones, aminoglycosides, tetracyclines, macrolides, and nitrofurans, non-susceptible isolates could form stronger biofilms than susceptible isolates ($p < 0.05$; Figure 3A–F). Conversely, for β -lactams and lipopeptides, no significant difference in biofilm formation between susceptible and non-susceptible isolates was observed (Figure 3G, and H). Perez et al exploited the correlation of meropenem-resistance and the ability to form biofilms,⁶¹ and found an inverse relationship between carbapenem-resistance and biofilm-production. In addition, Fábrega et al demonstrated an inverse relationship between biofilm-formation and quinolone-resistance in the context of *Salmonella enterica*.⁶² Together, there

Table 3 Biofilm Forming Capacities of *E. coli* with Different Antibiotic Resistance Phenotypes

Resistance Phenotype	n	OD ₅₅₀	r_s	p-value
Non-MDR	1 (1.23%)	0.609	0.243	< 0.05
MDR	23 (28.4%)	0.281 (0.095, 1.111)		
XDR	57 (70.37%)	1.000 (0.264, 1.152)		

Abbreviations: OD₅₅₀, optical density at 550 nm; IQR, data shown in median interquartile range; MDR, multidrug-resistant; XDR, extensively drug-resistant.

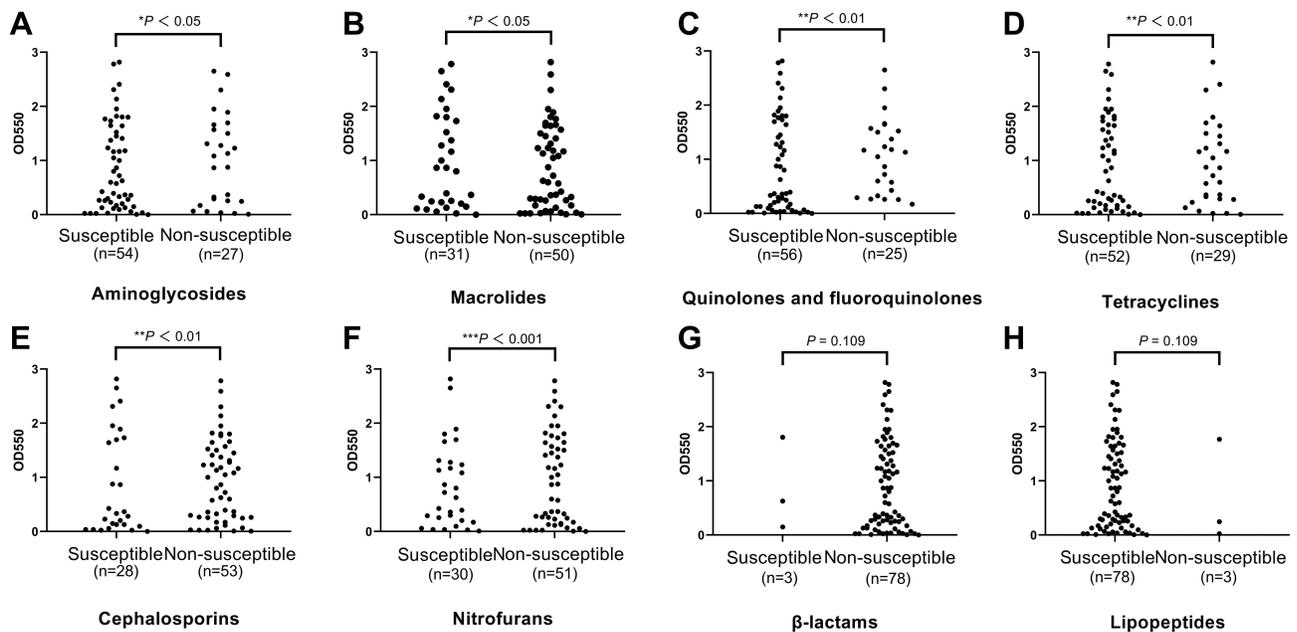


Figure 3 Relationship between biofilm formation and the resistance of *E. coli* isolates to each of the eight antimicrobial categories. Optical density at 550 nm (OD_{550}), represents biofilm forming capacity. (A–F): cephalosporins, quinolones and fluoroquinolones, aminoglycosides, tetracyclines, macrolides and nitrofurans; (G and H): β -lactams and lipopeptides.

are discrepancies in the relationship between antibiotic resistance and biofilm formation among different strains,³² but further efforts are still needed to optimize the treatment regime for clinician.

CLSM and FESEM Analysis of Biofilm Morphology and Structure

36-h biofilm structures of representative isolates observed using CLSM are presented in Figure 4A. Marked variability in three-dimensional biofilm architecture between the different isolates were displayed. Weak biofilm producers formed only a few, small scattered cell clusters, whereas medium biofilm producer formed rough biofilms containing several small aggregates and were of variable thickness. In contrast, strong biofilm producers displayed a high degree of variability in terms of biofilm structure. FESEM examination of biofilms cultivated for 36 h revealed that *E. coli* isolates exhibited disparate biofilm-forming abilities and adopted various structural conformations. Strong biofilm producers displayed thick-cell layers and medium biofilm producers shaped rough biofilms containing several small aggregates and

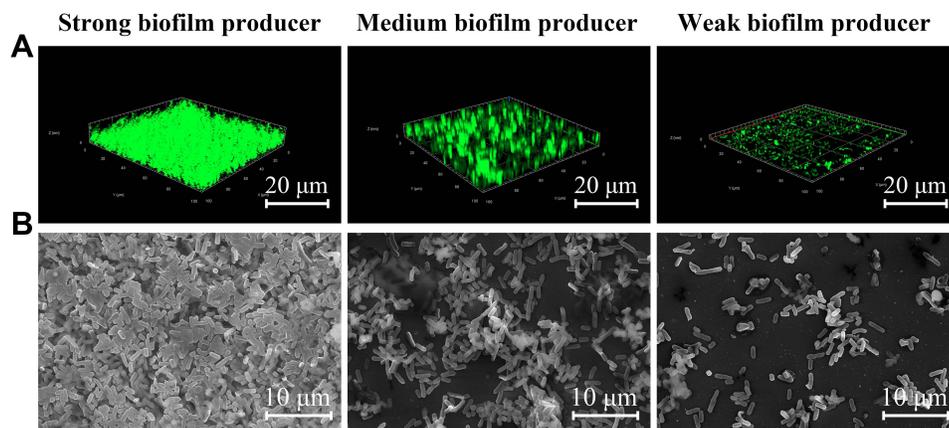


Figure 4 Observation of representative strong, medium, and weak biofilm producers via confocal laser scanning microscopy (CLSM, (A)) and field emission scanning electron microscopy (FESEM, (B)). scale bars represent 20 μ m for CLSM and 10 μ m for FESEM.

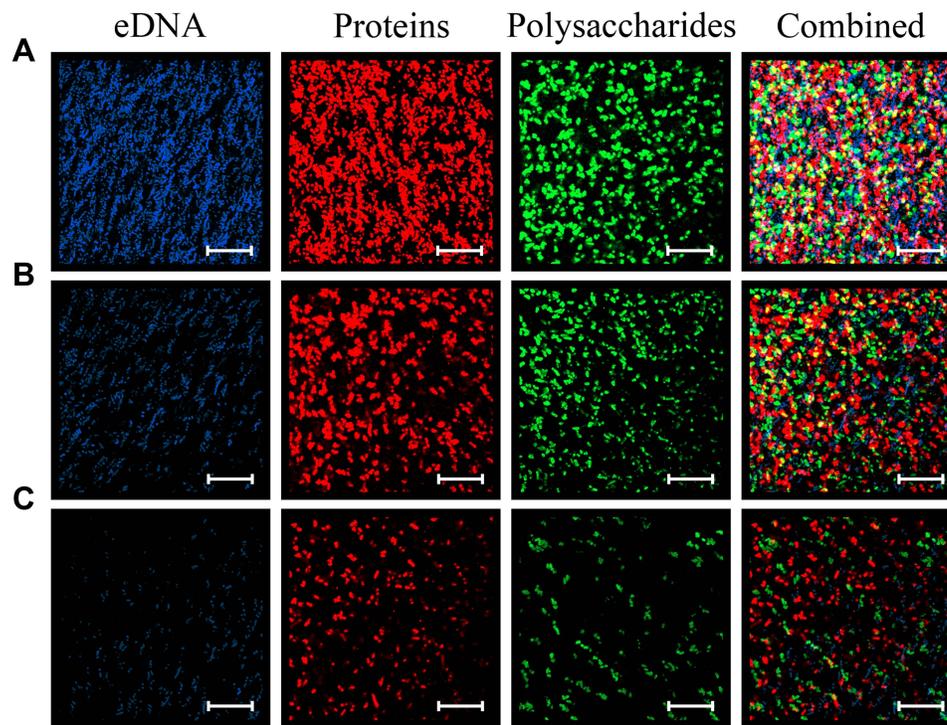


Figure 5 Visualization of the biofilm matrix levels of representative *E. coli* isolates with different biofilm-forming capabilities (strong **(A)**, medium **(B)** and weak **(C)**) by confocal laser scanning microscopy (CLSM) combined with three fluorescent dyes. Film Tracer SYPRO Ruby, WGA and DAPI were employed to stain proteins, polysaccharides and eDNA within the biofilm, respectively. Scale bars represent 10 μm .

were of variable thickness, while weak biofilm producers only a few, small scattered cell clusters (Figure 4B). Similarly, strong biofilm-producing bacteria such as *Vibrio parahaemolyticus* established thick 3-D structures, whereas poor-biofilm-forming strains produced thin and inconsistent biofilms.⁶³

Biofilm Composition by CLSM

CLSM in conjugation with three different fluorescent dyes was used to investigate the distribution of eDNA, proteins, and polysaccharides within the biofilm. As demonstrated in Figure 5A, eDNA, proteins, and polysaccharides were at high levels and uniformly distributed in strong biofilm producers, whereas weak biofilm producers revealed lower biomass level and the proteins and polysaccharides were the major components within biofilms (Figure 5B, and C).

Diffusion of Gatifloxacin Within Biofilms

To evaluate the contribution of the resistance of biofilms to antibiotics, gatifloxacin diffusion within biofilms was visualized using CLSM. As presented in Figure 6, CLSM revealed that for the biofilms produced by the strong biofilm

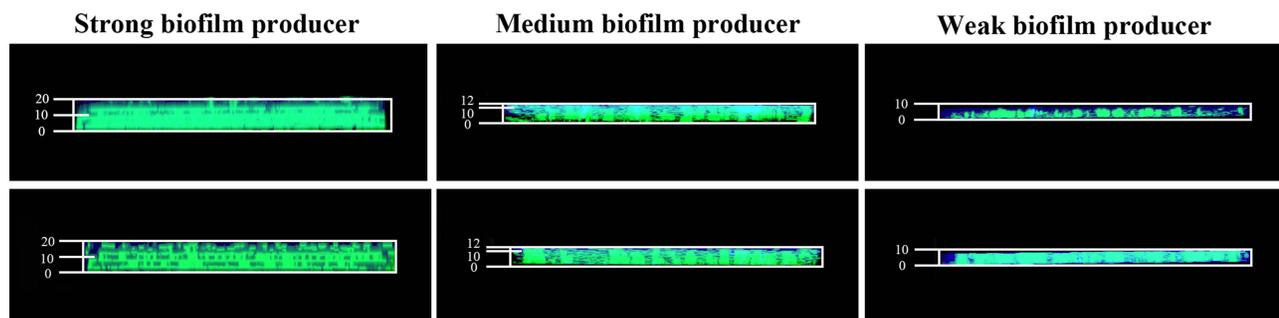


Figure 6 CLSM images assessing the diffusion of gatifloxacin through biofilms of three representative isolates. The biofilms were stained with SYTO 9 to visualize biofilm (green); the intrinsic fluorescence of gatifloxacin appeared blue.

producers, gatifloxacin (blue fluorescence) was mainly confined to the outer periphery of the biofilm, with little or no gatifloxacin penetration into the biofilm interior. In contrast, more penetration of gatifloxacin into the biofilm by the weak-biofilm producers was observed. Moreover, CLSM clearly revealed extensive gatifloxacin diffusion within biofilms by the weak-biofilm producers, almost reaching the basal layers. The results indicated that dense biofilms limited the penetration antimicrobial agents through the biofilm matrix due to their dense and thick structure, and thus could contribute to the antimicrobial tolerance of biofilms.⁶⁴

Conclusion

In this study, we evaluated the relationship between antibiotic resistance, biofilm formation, and biofilm-specific resistance of *E. coli* isolates from pediatric patients in Ningbo, China. The current study showed that the clinical isolates of *E. coli* from patients in Ningbo, China, demonstrated a positive association between the MDR or XDR phenotype and biofilm-production, and the correlation between biofilm-formation and cefoxitin, ceftriaxone, cefazolin, and gentamicin resistance was observed. Therefore, a local and regular surveillance of biofilm-formation in *E. coli* isolates and their antimicrobial resistance profiles should be performed. This might help to formulate therapeutic strategies against *E. coli*-related infections. Moreover, in this study, our sample size is a little small and only from a geographic location. Expanding the sample size and geographic area in order to cast light on how different geographical regions and patient populations contribute to the selection of pathogens with different profiles of biofilm and its resistance should await further study.

Ethics Approval and Consent to Participate

This study received approval from the Ethical Committee (IRB) of Ningbo Municipal Center for Disease Control and Prevention and was conducted in accordance with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards. Since this study was observational and strains from redundant, abandoned individual excreta were employed in clinical diagnosis, the confidentiality of patient data is preserved. Considering this study did not affect patients' health and privacy, the exemption criteria were met. After consulting with the IRB of the Ningbo Municipal Center for Disease Control and Prevention, written informed consent from the patient or parents was not required.

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Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval for the version to be published; have agreed on the journal to which the article has been submitted; and agreed to be accountable for all aspects of the work.

Disclosure

The authors claim no conflicts of interest in this study.

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