

The Effects of Luminescent CdSe Quantum Dot-Functionalized Antimicrobial Peptides Nanoparticles on Antibacterial Activity and Molecular Mechanism

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Background: With the development of bacterial resistance, the range of effective antibiotics is increasingly becoming more limited. The effective use of nanoscale antimicrobial peptides (AP) in therapeutic and diagnostic methods is a strategy for new antibiotics.

Methods: Combining both AP and cadmium selenide (CdSe) into a composite material may result in a reagent with novel properties, such as enhanced antibacterial activity, fluorescence and favorable stability in aqueous solution.

Results: AP-loaded CdSe NPs (AP-CdSe NPs) showed strong antibacterial activity against multidrug-resistant (MDR) *Escherichia coli* (*E. coli*) and *Staphylococcus aureus* (*S. aureus*) in vitro and in vivo. Colony-forming unit (CFU) and minimum inhibitory concentration (MIC) assays showed that AP-CdSe NPs have highly effective antibacterial activity. The quantitative analysis of apoptosis by flow cytometry analysis further confirmed that MDR *E. coli* and *S. aureus* treated with AP-CdSe NPs had death rates of 98.76% and 99.13%, respectively. Also, AP-CdSe NPs was found to inhibit bacterial activity in an in vivo bacteremia model in mice infected with *S. aureus*. In addition, the antibacterial mechanism of AP-CdSe NPs was determined by RNA sequencing analysis. Gene ontology (GO) analysis and Kyoto encyclopedia of genes and genomes (KEGG) pathway analysis revealed the molecular mechanism of the antibacterial effect of AP-CdSe NPs. Importantly, histopathology analysis, and hematological toxicity analysis indicated that AP-CdSe NPs had few side effects.

Conclusion: These results demonstrate that AP loaded on CdSe NPs had a higher water solubility, bioavailability and antibacterial effect compared with raw AP. This study reports findings that are helpful for the design and development of antibacterial treatment strategies based on AP.

Keywords: antibacterial peptides, CdSe QD, antibacterial mechanism, RNASeq, in vitro and in vivo

Introduction

Bacterial infections have become a global public health problem, and their mortality, morbidity and treatment costs during hospitalization increase every year.¹⁻³ With the rapid emergence of multidrug-resistant (MDR) “superbug” strains and the lack of new antibiotics, modern society is facing the post-antibiotic era.⁴⁻⁷ Therefore, there is an urgent need to develop alternatives to antibiotics.

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Antimicrobial peptides (AP) have become attractive potential alternative therapeutic compounds.⁸ AP have a wide range of activities, and a broad-spectrum antibacterial activity against Gram-negative and Gram-positive bacteria.^{9,10} AP kill or suppress benign and pathogenic organisms, thereby disrupting the homeostasis between a healthy microbiota and immune system.¹¹ The amino acid sequence and structure of different AP are obviously different, but they all have an amphipathic structure and can be combined with other compounds.^{12,13} Therefore, they can easily interact with the negatively charged components on the bacterial cell membrane and integrate into the lipid bilayer. However, their poor biocompatibility is one of the main obstacles to the clinical development of a large number of common AP.^{14,15} Target delivery of AP is necessary due to their short half-life and relative toxicity against healthy cells.¹⁶ The effective use of nanoscale proteins and peptides in therapeutic and diagnostic methods is a strategy to induce and maintain their functional conformation.

Research on nano-modification based on AP has shown that they can be combined with nanoparticles (NPs) to form nanomaterials with novel properties, due to their advantages in molecular recognition and antibacterial activity.^{15,17,18} Currently, quantum dots (QDs) not only have very important applications in solar cells and phototransistor components, but also in biological applications including biosensors,^{19,20} bioimaging²¹ and biomedical research.²² Cadmium selenide (CdSe) QDs with a size of 1–20 nm are currently used for in vivo imaging and diagnostic treatment.²³ Growing an outer shell of AP on the outside of the QDs can greatly improve its stability.²⁴ Therefore, the combined use of an AP and CdSe QDs (such as conventional antibiotics) definitely has the potential to improve the effectiveness of the two groups of compounds.^{25–27}

In this study, an AP, used as a stabilizer and active group, was modified on the surface of CdSe QDs to form fluorescent AP NPs (AP-CdSe NPs) with high antibacterial activity.²⁸ The AP-CdSe NPs showed strong antibacterial activity against MDR *Escherichia coli* (*E. coli*) and *Staphylococcus aureus* (*S. aureus*) in vitro and in vivo. The results of colony-forming unit (CFU) and minimum inhibitory concentration (MIC) assays showed that AP-CdSe NPs have a broad-spectrum antibacterial activity, and displayed highly effective antibacterial activity against MDR *E. coli* and *S. aureus*. Fluorescence microscopy analysis revealed that AP-CdSe NPs have a high cellular uptake rate and killing rate for *E. coli* and *S. aureus*. The quantitative analysis of live/dead cells by flow cytometry

analysis further confirmed the antibacterial activity of AP-CdSe NPs. Scanning electron microscopy (SEM) and transmission electron microscopy (TEM) were used to examine the morphological changes of bacteria after being destroyed by AP-CdSe NPs. In addition, through the use of a bacteremia model in mice infected with *S. aureus* to analyze the biodistribution of bacteria in mice and perform histological evaluation, it was determined that AP-CdSe NPs can inhibit bacterial activity and heal tissue damage through. Furthermore, gene expression profiling by RNA sequencing (RNASeq) analysis was used to determine the mechanism of antibacterial activity of AP-CdSe NPs.²⁹ Concurrently, we prepared peptide nanoparticles using traditional modification methods, and their effects were better than those of AP alone.

Methods and Materials

Materials

Selenium (Se) powder (99.99%), 11-Mercaptoundecanoic acid (99%), CdCl₂·2.5H₂O (99%), NaBH₄ (≥99%) were purchased from Aladdin Chemical Reagent Co., Ltd. (Shanghai, China). The beef extract peptone medium agar powder used to prepare the bacterial culture plates was obtained from the School of Biochemical Engineering, Anhui Polytechnic University (Wuhu, China).

Synthesis of AP, CdSe and AP-CdSe Nanoparticles

Peptides were synthesized by the fluorenylmethoxycarbonyl (Fmoc) solid-phase synthesis method. The 2-Cl-resin was selected as the carrier for amino acid binding. A mixture of N,N-diisopropylethylamine (DIEA), 1-hydroxybenzotriazole (HOBT), and N,N'-diisopropylcarbodiimide (DIC) were used as condensation reagents. The AP periplaneta-sin-4 (Leu-Arg-His-Lys-Val-Tyr-Gly-Tyr-Cys-Val-Leu-Gly-Pro-NH₂) was synthesized from the carboxyl end to the amino end in the indicated order.^{30,31} The structure of the AP is shown in [Supplementary Figure S1](#).

The CdSe NPs were prepared as previously reported by Sun et al.³² The AP-CdSe NPs were prepared with mini revise. Briefly, the AP (0.6 mM) and CdCl₂ (0.5 mM) were added into 60 mL of Milli-Q water. The pH of the mixture solution was adjusted to 11. Also, NaBH₄ (1 mM) was dissolved in 1 mL H₂O, and Se powder (0.5 mM) was added and mixed until the mixture was clear and transparent. The precursor is then quickly added to the above solution under N₂ flow. The nanomaterial (AP-CdSe NPs) was obtained after stirring for

2 h with a magnetic stirrer at 60°C. The AP-CdSe NPs were ultimately collected by centrifugation at 15,000 rpm, for 5 min, and washed twice with Milli-Q water. Amount of AP in the solutions was determined by measuring the absorbance at 234 nm after centrifugation. The [Table S1](#) showed the drug loading rate of AP-CdSe NPs was up to 85%.

Characterization

The purity of the AP was determined by mass spectrometry (MS) on a Q Exactive Plus Orbitrap mass spectrometer (Thermo Fisher Scientific Inc., Waltham, MA, USA), and nuclear magnetic resonance (NMR) spectroscopy on a picoSpin 80 NMR spectrometer (Thermo Fisher Scientific Inc.). The ultraviolet-visible (UV-vis) absorption spectra were recorded by UV-vis spectroscopy using a Shimadzu UV-2550 spectrometer (Shimadzu Corp., Kyoto, Japan). Fourier transform infrared (FT-IR) spectroscopy was performed on a IR Prestige-21 FT-IR spectrometer (Shimadzu Corp.). Fluorescence spectra of the AP-CdSe NPs were recorded by fluorescence spectroscopy using an RF-5301PC fluorescence spectrophotometer (Shimadzu Corp.). The morphology of the CdSe NPs and AP-CdSe NPs were examined by TEM on a JEOL 6300 F transmission electron microscope (JEOL Ltd., Tokyo Japan). The dynamic light scattering (DLS) of the NPs was measured using a Brookhaven Zeta PALS instrument (Brookhaven Instruments Corporation, Brookhaven, Holtsville, NY, USA).

Bacterial Culture

Escherichia coli ATCC 11303 (*E. coli*, Gram-negative), *Staphylococcus aureus* ATCC 25923 (*S. aureus*, Gram-positive), *Bacillus subtilis* ATCC 2233 (*B. subtilis*, Gram-positive), *Micrococcus tetragenus* ATCC 2835 (*M. tetragenus*, Gram-positive) and *Pseudomonas aeruginosa* ATCC 27853 (*P. aeruginosa*, Gram-negative) were obtained from Anhui Polytechnic University. MDR *E. coli* E76227, MDR *P. aeruginosa* BJ915, MDR *S. aureus* ATCC 25213, MDR *Acinetobacter baumannii* GIM (*A. baumannii*, Gram-negative) 1.650 were obtained from Jinan University (Guangzhou, China).

Minimum Inhibitory Concentration and Counting Colony-Forming Unit Test

The 96-well microtiter plate dilution method was used to determine the MIC of the AP-CdSe NPs. Logarithmic-phase bacteria were diluted with fresh medium and then two aliquots of 190 μ L of the bacterial culture suspension

were separately mixed with 10 μ L of two different concentrations of AP-CdSe NPs followed by incubation at 37°C for 12 h (the final density of bacteria was 1.0×10^6 CFU/mL). Then, 200 μ L of medium was added to each sample and in turn aliquots of 200 μ L were drawn from each sample and further diluted by double dilution, and ultimately the concentration of each sample set was determined in triplicate by measuring three parallel samples.³³

Log-phase MDR *E. coli* and MDR *S. aureus* were inoculated (1:40) in fresh medium containing AP-CdSe NPs solution at the corresponding concentrations (10, 20, 40 μ g/mL), or AP solution (40 μ g/mL), or CdSe NPs solution (40 μ g/mL) and subsequently incubated in a shaking incubator at 37°C, for 12 h. The bacterial culture treated with phosphate-buffered saline (PBS, pH 7.5, 0.1 M) were used as blank. The numbers of viable bacterial cells on solid medium were counted using the CFU counting method. The statistical analysis of the CFU assay data for each group was performed with the Origin software (OriginLab Corp., Northampton, MA, USA).

Cellular Uptake Assays

The cellular uptake assay was conducted using a staining test. Log-phase MDR *E. coli* and *S. aureus* were incubated with AP-CdSe NPs solution at the corresponding concentrations (10, 20, 40 μ g/mL) for 12h. The PBS-treated cells were used as the blank group. The CdSe NPs (40 μ g/mL) was set as the control group. Cells were collected and stained with 5 μ g/mL of 4'-6-diamidino-2-phenylindole (DAPI, Thermo Fisher Scientific Inc.). Bacterial culture suspension were also washed twice with PBS. Then, a fluorescence microscope was used to observe the cells in the red and blue channels.

LIVE-DEAD Assays

Log-phase MDR *E. coli* and MDR *S. aureus* were treated with AP-CdSe NPs (10, 20, 40 μ g/mL). AP solution (40 μ g/mL) and CdSe NPs solution (40 μ g/mL) were set as control group. The PBS was set as blank group. Cells were collected and stained with the reagent test kit (SYTO9 and propidium iodide; Thermo Fisher Scientific) for 30 min in the dark. Then, Used FL microscope observed the cell at red and green channels. The apoptotic cell death statistical test was analyzed by a CytoFLEX (Beckman Coulter).³⁴

Cell Membrane and Wall Integrity Studies

Log-phase MDR *E. coli* and MDR *S. aureus* were treated with AP-CdSe NPs solution at the corresponding

concentrations (10, 20, 40 µg/mL). The groups treated with AP solution (40 µg/mL) and CdSe NPs solution (40 µg/mL) were used as control groups. The PBS-treated group was used as blank group. All bacterial cells were collected by centrifugation at 3000 rpm, for 15 min and fixed by dehydrating in an ethanol gradient at 4°C, and then postfixed in a mixture of glutaraldehyde (2.5%) and paraformaldehyde (2%) for 12 h. Eventually, dried samples were examined by SEM using a Hitachi S-4800 scanning electron microscope (Hitachi Ltd., Tokyo, Japan).³⁵

Log-phase MDR *E. coli* and MDR *S. aureus* were treated with AP-CdSe NPs solution at the corresponding concentrations (10, 20, 40 µg/mL). The groups treated with AP solution (40 µg/mL) and CdSe NPs solution (40 µg/mL) were used as control groups. The PBS-treated group was used as blank group. All bacterial cells were collected by centrifugation at 3000 rpm, for 15 min, and fixed with a mixture of glutaraldehyde (2.5%) and paraformaldehyde (2%) for 24 h. Then, the MDR *E. coli* and MDR *S. aureus* cells were subjected to a series of biological dehydration tests, embedded, sectioned, mounted on 200-mesh copper grids and examined by TEM.³⁶

RNA Extraction and Quantification of Gene Expression Level

Log phase MDR *E. coli* cells were exposed to AP-CdSe NPs (40 µg/mL) for 12 h at 37°C in the shaking incubator. The PBS-treated cells were used as control. Total cellular RNA was extracted. After the library was constructed, Qubit3.0 was used for preliminary quantification, and then the Qsep 100 BioFragment Analyzer (BioOptic Inc., New Taipei City, Taiwan) was used to detect the insert size of the library. Once the insert size met expectations, the quantitative real-time polymerase chain reaction (qPCR) method was used to accurately quantify the effective concentration of the library in order to ensure the quality of the library.

Before differential gene expression analysis, for each sequenced library, the read counts were adjusted using a scaling normalization factor through the edge R package. Then, the DEGSeq (differentially expressed genes Seq) R package (1.20.0) was used to analyze the differential gene expression in the two conditions. The Benjamini & Hochberg method was used to adjust the P value. A corrected P value of 0.005 and a log₂ (fold

change) of 1 were set as the threshold for significant differential expression.

Gene Ontology and Kyoto Encyclopedia of Genes and Genomes Enrichment Analysis of Differentially Expressed Genes

The off-machine data was filtered to obtain clean reads. The clean reads were compared with the reference genome sequence, and the number of reads contained in each gene were counted, that is, the expression level, according to the genome annotation file and the position of the reads, and then statistical testing was performed. The differentially expressed genes (DEGs) were obtained and functionally annotated and ultimately Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genome (KEGG) enrichment analyses were performed on the DEGs to obtain the GO and KEGG sets enriched by the DEGs. Concurrently, according to reads and reference base due to group differences, together with statistical inference, single nucleotide variations, such as SNP and Indel, were found.

Animal Bacteremia Model

Six-week-old female SKH1 nude mice (~25 g) were obtained from the Model Animal Research Center of Nanjing University (Nanjing, China). All animals were maintained in accordance with the Animal Welfare Act and the Guide for the Care and Use of Laboratory Animals protocols. All animal studies were approved by the Suzhou University (Suzhou, China) Animal Care (Approval Number: 202010A415). The SKH1 nude mice were randomly and equally divided into 4 groups (n=21), on the 7th day of intravenous tail injection. The four groups of mice were separated into cages: blank, infection group (MDR *S. aureus*), and treatment groups (MDR *S. aureus*). At 24 h after the injection, the mice of the treatment groups were intravenous tail injections with a dose of 2 mg/kg AP or AP-CdSe NPs for treatment groups.

The mice of each group were sacrificed at 1 d, 3 d, 5 d and 7 d after treatment, and blood and organs (heart, liver, kidneys, spleen and lungs) were collected. The organs were harvested, weighed, and homogenized in sterile water. The number of surviving bacteria was counted by the CFU method. The weight and survival rate of the mice in each group were monitored daily.

Histological Analysis

The following organs (heart, liver, kidneys, spleen and lungs) were collected in each group after all treatment. The harvested organs were fixed in 4% formaldehyde for 24 h. Afterwards organ samples were embedded in paraffin, sectioned into 4- μ m thick slices, placed on slides and stained with hematoxylin and eosin (H&E). The stained slides were examined by light microscopy.

Safety Evaluation

To evaluate the biosafety of the AP-CdSe NPs, mice (Blank and AP-CdSe NPs treatment groups) were sacrificed after 30 days, and major organs (liver, spleen, kidneys, heart, and lungs) were collected and stained with H&E to evaluate histopathological changes *in vivo*. Blood samples were also collected and hematological parameters, including uric acid, blood glucose, cholesterol, and alanine

aminotransferase, were measured. Organs from healthy nude mice were used as blank.³⁷

Results and Discussion

Characterization of AP-CdSe NPs

AP-CdSe NPs were synthesized using a simple method as described above. The AP is mainly modified as a stabilizer active agent of CdSe NPs. The structure of the AP was also confirmed by MS analysis and NMR spectroscopy. The MS chromatogram showed the molecular weight of the AP was 1505.8, which is consistent with the design results (Supplementary Figure S2). The NMR spectroscopy results showed that the AP was highly pure (Supplementary Figure S3 and S4). The characterization of AP-CdSe NPs is shown in Figure 1. The extinction spectrum of CdSe NPs (red line) shows peaks at around 281 nm, and all shifted to the shorter wavelength after being modified by AP (Figure 1A). For the AP-CdSe NPs,

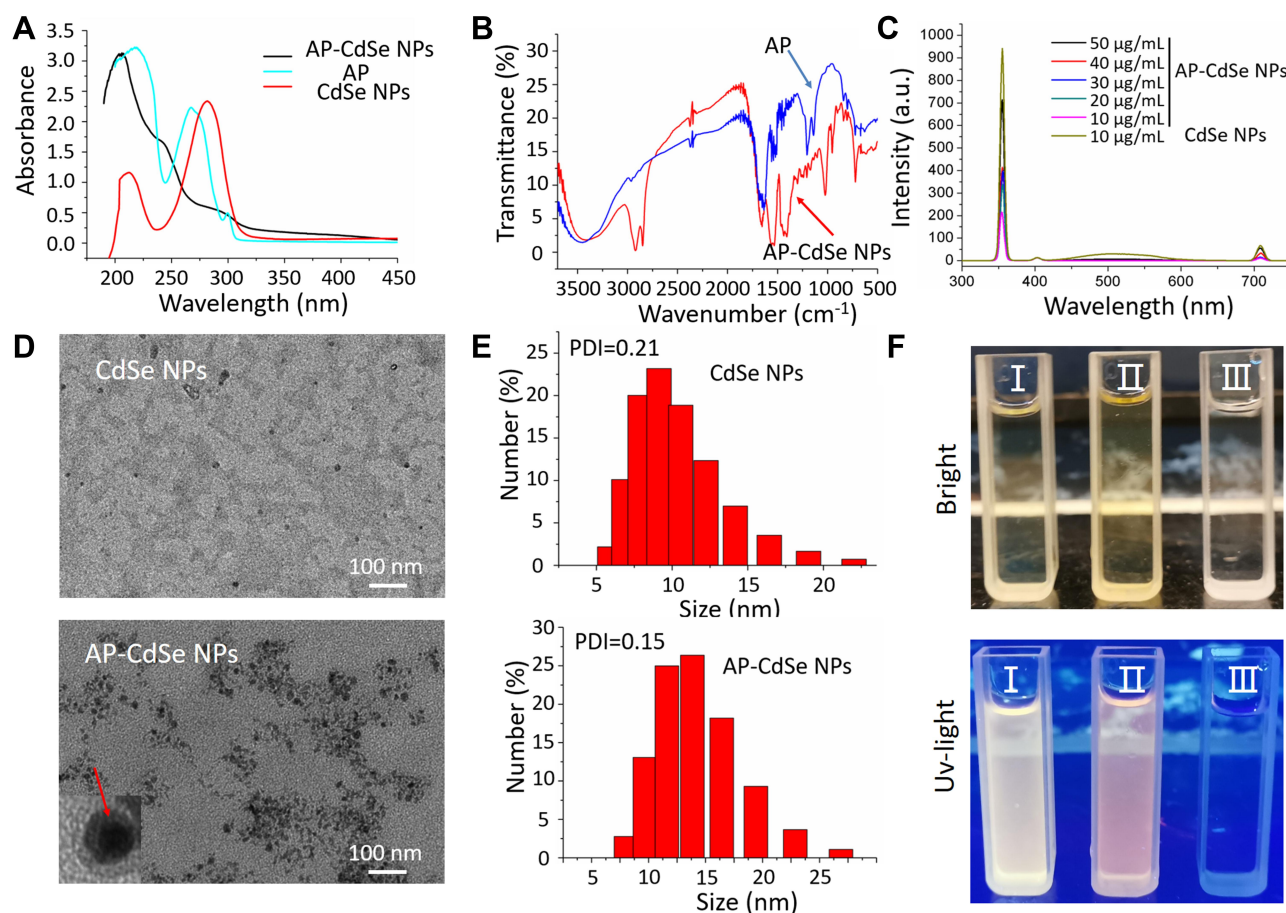


Figure 1 Characterization of AP-CdSe NPs. (A) UV-vis absorption spectra of AP, CdSe NPs and AP-CdSe NPs. (B) Infrared spectra of AP and AP-CdSe NPs. (C) Fluorescence emission spectra of AP-CdSe NPs at various concentrations (10, 20, 30, 40 and 50 μ g/mL) and CdSe NPs (10 μ g/mL). (D) TEM images of CdSe NPs and AP-CdSe NPs. (E) DLS of CdSe NPs and AP-CdSe NPs. In the inset, the red arrow points to a AP coated CdSe NPs. (F) Digital images of CdSe NPs (I) and AP-CdSe NPs (II) under bright and UV light.

the peak at 246 nm is blue-shifted, indicating the successful modification by AP on the CdSe NPs. Their structures were also confirmed by FT-IR spectroscopy, as shown by the difference changes observed in the peak of the fingerprint spectrum (Figure 1B). The fluorescence excitation spectra of AP-CdSe NPs at different concentrations, shown in Figure 1C, reveal that the fluorescence intensity of the solution became increasingly stronger with the increase of the concentration. The successful conjugation of the AP on the surface of the CdSe NPs also revealed the fluorescence property of QDs. AP is modified on the QDs without altering its fluorescence properties. The fluorescence property was confirmed in the image displayed in Figure 1F, which shows the fluorescence signal of CdSe NPs (I) and AP-CdSe NPs (II). The morphology and size distribution of the NPs are shown in Figure 1D and E, respectively. The average size of the obtained CdSe NPs and AP-CdSe NPs were 9 nm and 14 nm, respectively (Figure S7). Amount of AP released into the solutions was determined by measuring the absorbance at 234 nm using UV-vis spectroscopy at different time points. The Figure S5 showed the initial rate of release was much faster, the amount of release was much larger, and the time to reach equilibrium was at 12 h. This experiment demonstrated that the release rate of AP from nanoparticles was up to 82%. The stability of AP-CdSe NPs in bacterial culture suspension was evaluated by examining the size of NPs (Figure S6) in vitro. When the NPs were dispersed in PBS and bacterial culture medium for more than 196 h, no obvious size change was observed, indicating good stability in biological media. Taken together, these results suggest that APs were successfully decorated on the surface of CdSe NPs and exhibited fluorescence properties.

Antibacterial Activity of AP-CdSe NPs

AP have been reported as novel and effective antibacterial active short peptide fragments. However, since some APs have low water solubility, their bioavailability is extremely low.^{38,39} In this study, the polarity of an AP was successfully changed to improve its antibacterial activity, as indicated by the results shown in Figure 2 and Table 1. For instance, the MIC values (Table 1) for the AP against regular *E. coli*, *S. aureus*, *M. tetragenus*, *P. aeruginosa* and *B. subtilis* were relatively high at 48.0, 48.0, 64.0, 48.0 and 64.0 µg/mL, respectively. Also, there was only a little difference in the MIC values for the AP against MDR bacteria. On the other hand, AP-CdSe NPs not only showed relatively low MIC values against these regular

bacterial cells at 6.0, 4.0, 24.0, 16.0 and 24.0 µg/mL, but also against MDR bacteria at 16.0, 24.0, 8.0 and 8.0 µg/mL, respectively. By penetrating the bacterial cell membrane, the AP-conjugated CdSe NPs had enhanced antibacterial activity according to the MIC assay results. AP-CdSe NPs had a more prominent effect on the activity of *S. aureus* (Gram-positive bacteria) and *E. coli* (Gram-negative bacteria) than on the other bacterial cells.

These results were also consistent with those obtained by the CFU assays, shown in Supplementary Figure S8 and Figure 2. As shown in the Supplementary Figure S8, photographs of the inhibition zone in cultures of MDR *E. coli* and *S. aureus* after treatment with AP-CdSe NPs (40 µg/mL) were qualitative test of antibacterial drugs and measured. The diameter of the inhibition zone of AP-CdSe NPs can reach 27.5 mm for MDR *S. aureus* and 16.5 mm for MDR *E. coli*. The results showed that both the MDR *E. coli* and *S. aureus* are sensitive to AP-CdSe NPs, and the sensitivity of *S. aureus* is stronger than that of *E. coli*. The sensitivity of raw AP was bad. The CFU assays were used to evaluate the antibacterial activity of AP-CdSe NPs against MDR *E. coli* and *S. aureus*. The results showed that AP-CdSe NPs at 40 µg/mL against MDR *E. coli* and *S. aureus*, resulted in survival rates of 10.3 and 3.8%, respectively, whereas the survival rates of AP at 40 µg/mL were 43.7% and 22.3%, and those of CdSe NPs at 40 µg/mL were 96.7% and 95.3%, respectively (Figure 2). These results revealed that CdSe NPs alone had no antibacterial activity. In addition, the inhibitory effect of AP-CdSe NPs was higher than that of AP and CdSe NPs. Besides showing that AP-CdSe NPs have high antibacterial activity, these results also proved that AP-CdSe NPs exhibit a synergistically enhanced antibacterial performance against MDRs bacteria.

Cellular Uptake and Antibacterial Activity Assays

The drug delivery efficacy and antibacterial activity were investigated by fluorescence microscopy analysis. All MDR *E. coli* and MDR *S. aureus* cells were stained with DAPI, including the blank group, and with AP-CdSe NPs at various concentrations (10, 20, 40 µg/mL). The merged images indicated the drug delivery efficacy and antibacterial activity of AP-CdSe NPs. The results in Figure 3, where red cells are those stained with AP-CdSe NPs and blue cells are stained with DAPI, show that AP-CdSe NPs were taken up at the high rate of 100% in MDR *E. coli* and MDR *S. aureus* cells.

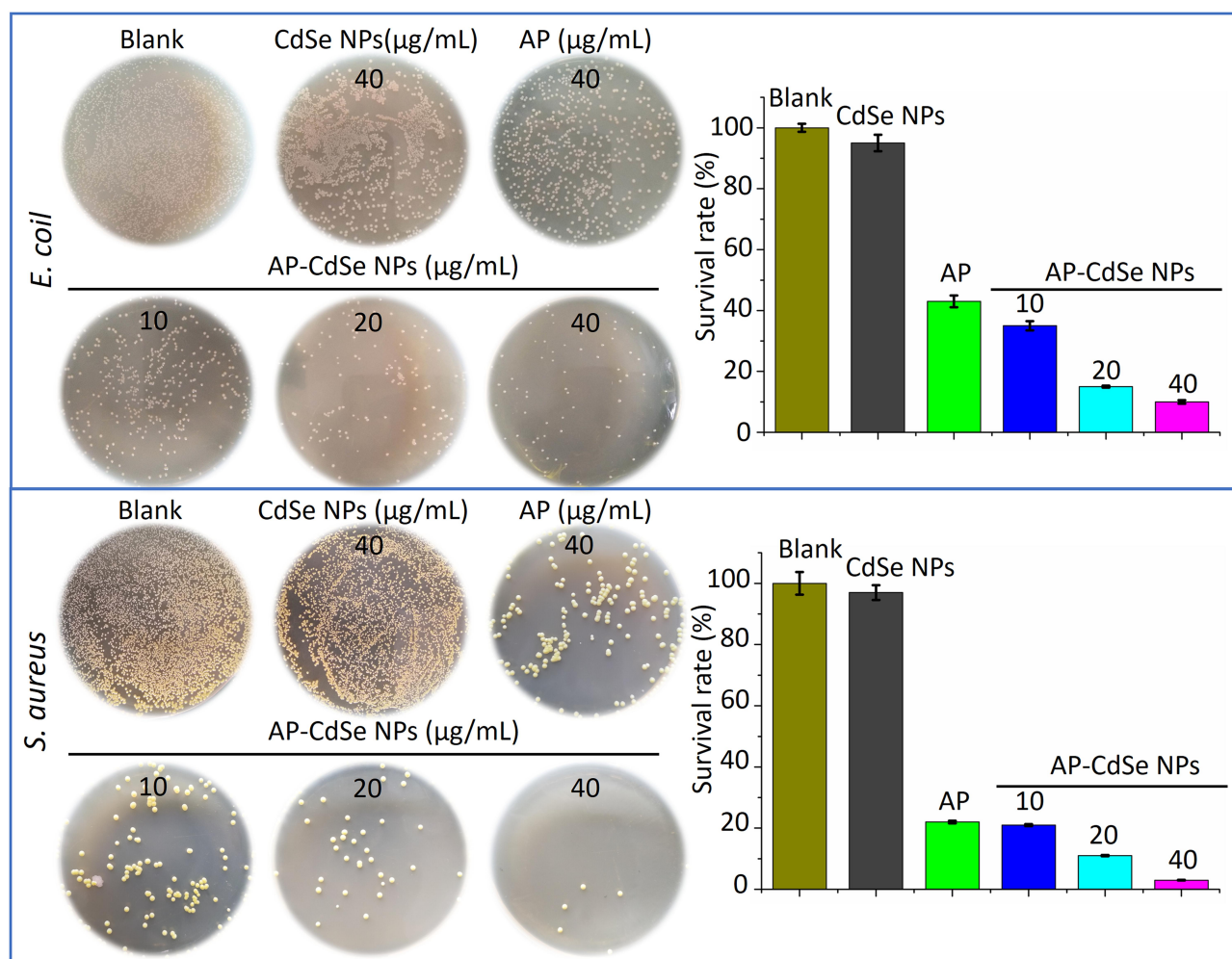


Figure 2 CFU assay of AP-CdSe NPs. Images of bacterial colonies (MDR *E. coli* and MDR *S. aureus*) formed on Lysogeny broth-agar plates. PBS was used for the blank group. Corresponding images were graphed using the Origin software. Data are expressed as the mean \pm SD (n=9).

Even at 10 $\mu\text{g/mL}$, AP-CdSe NPs could enter the cells, but the antibacterial effect is poor. At the concentration of 40 $\mu\text{g/mL}$ of AP-CdSe NPs, the number of cells in the field of view is smaller, but shows high antibacterial activity. In addition, the sampling time in cellular uptake assays had been set at the shorter time when most cells are still alive. It is clear that in just 1 hour, AP-CdSe NPs also completely enter the inside of the cell (Figure S9). Above all, the results show that AP-CdSe NPs are easily taken up by bacterial cells and have a good antibacterial activity against MDR *E. coli* and MDR *S. aureus*.

Fluorescence Microscopy Analysis of Antibacterial Activity

The CFU assay showed the remaining viable cells and evaluated antibacterial activity, but it did not reveal the dead cells. The uptake assay showed the decrease of cell

number under the microscope and evaluated antibacterial activity.^{37,38} Thus, the results of these assays only indirectly explain the antibacterial effect of the AP-CdSe NPs. Therefore, the live-dead assay was used to directly investigate the antibacterial activity of AP-CdSe NPs. In Figure 4, green fluorescent cells represented the living bacterial cells, while the red fluorescent cells represented dead bacterial cells. There were very few living MDR *E. coli* and MDR *S. aureus* cells (green fluorescent cell) in the blank group. Similar results were obtained for the group treated with CdSe NPs, which indicated that the NPs had no antibacterial activity against MDR *E. coli* and MDR *S. aureus*. There were some dead cells (red fluorescent cells) in the AP-treated groups, indicating that AP alone had antibacterial activity. However, the same concentration of AP-CdSe NPs displayed higher antibacterial activity compare with the AP-treated groups. All areas had

Table 1 MICs Values Test ($\mu\text{g/mL}$)

Bacteria	AP-CdSe NPs	CdSe NPs	AP
<i>E. coli</i>	6	128	64
<i>S. aureus</i>	4	>128	48
<i>M. tetragenus</i>	24	>128	64
<i>P. aeruginosa</i>	16	>128	48
<i>B. subtilis</i>	24	>128	64
MDR <i>E. coli</i>	8	>128	48
MDR <i>P. aeruginosa</i>	16	>128	48
MDR <i>S. aureus</i>	8	>128	64
MDR <i>A. baumannii</i>	8	>128	64

Note: Data are average values of at least three replicates.

dead cell (red fluorescent cells) and showed high killing rate. The reason might be that the AP loaded on CdSe NPs had a higher water solubility and bioavailability.

To further clarify that the AP-CdSe NPs reduce the survival rate of bacteria, a quantitative analysis of live/dead cells was performed by flow cytometry analysis

using the SYTO 9-PI apoptosis detection kit in MDR *E. coli* and MDR *S. aureus*. The results shown in [Figure 5A](#) and [B](#) revealed that the apoptosis rates of the blank group were 3.16% and 11.5%, and those of the group treated with CdSe (40 $\mu\text{g/mL}$) were 12.89% and 12.65% respectively. The results also revealed that CdSe NPs alone had no antibacterial activity. On the other hand, the AP (40 $\mu\text{g/mL}$) exhibited antibacterial activity with apoptosis rates of 61.36% and 59.47%, respectively. The result is consistent with the conclusion reported by others.^{40–42} Also, as shown in [Figure 5](#), after loading AP on CdSe NPs, the antibacterial activity of AP-CdSe NPs (40 $\mu\text{g/mL}$) was significantly higher than that of the AP-treated groups, with apoptosis rates of 98.76% and 99.13% for MDR *E. coli* and MDR *S. aureus*, respectively. The quantitative analysis of live/dead cells by flow cytometry analysis further confirmed the enhanced antibacterial activity of AP-CdSe NPs.

Cell Integrity Study

According to the results of the MIC assay, CFU assay, fluorescence microscopy analysis, and flow cytometry

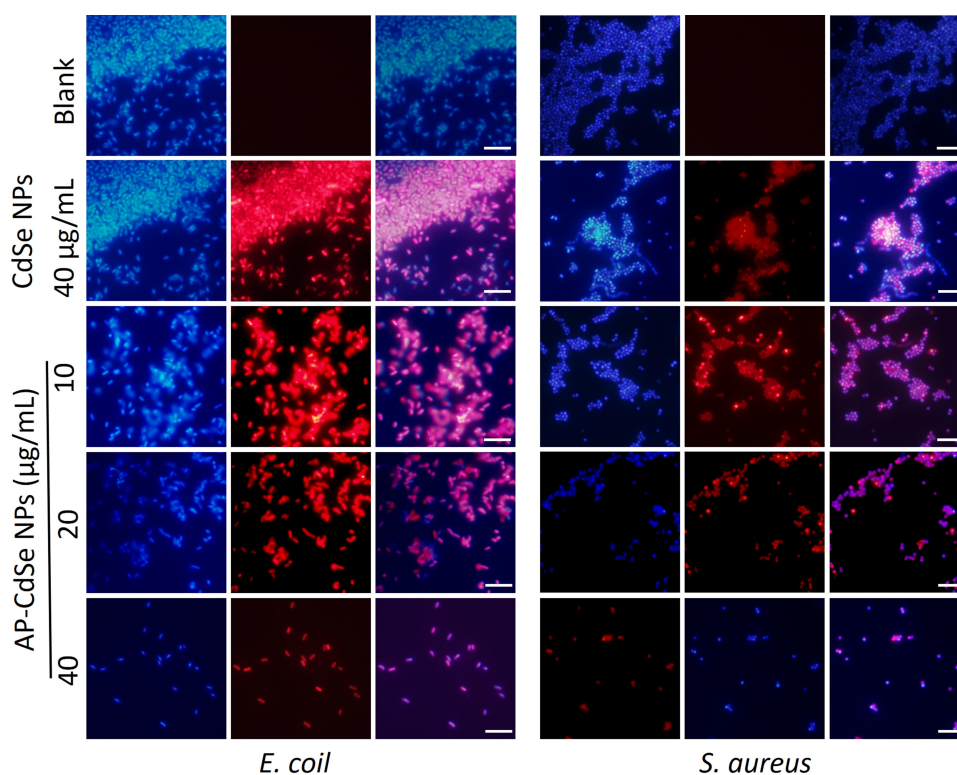


Figure 3 Fluorescence microscopy images of MDR *E. coli* and MDR *S. aureus* bacterial culture treated with AP-CdSe NPs at various concentrations (10, 20, 40 $\mu\text{g/mL}$) for 12 h. The PBS was used for the blank group. The CdSe NPs (40 $\mu\text{g/mL}$) was set as the control group. Cells were stained with DAPI for 30 min are fluorescent blue, and those stained with AP-CdSe NPs are fluorescent red, while those with fluorescent pink color appearance represent merged images of cells. Fluorescence images were captured using an UV illuminator with a 365 nm emission filter for AP-CdSe NPs and a 461 nm emission filters for DAPI. Scale bar = 10 μm .

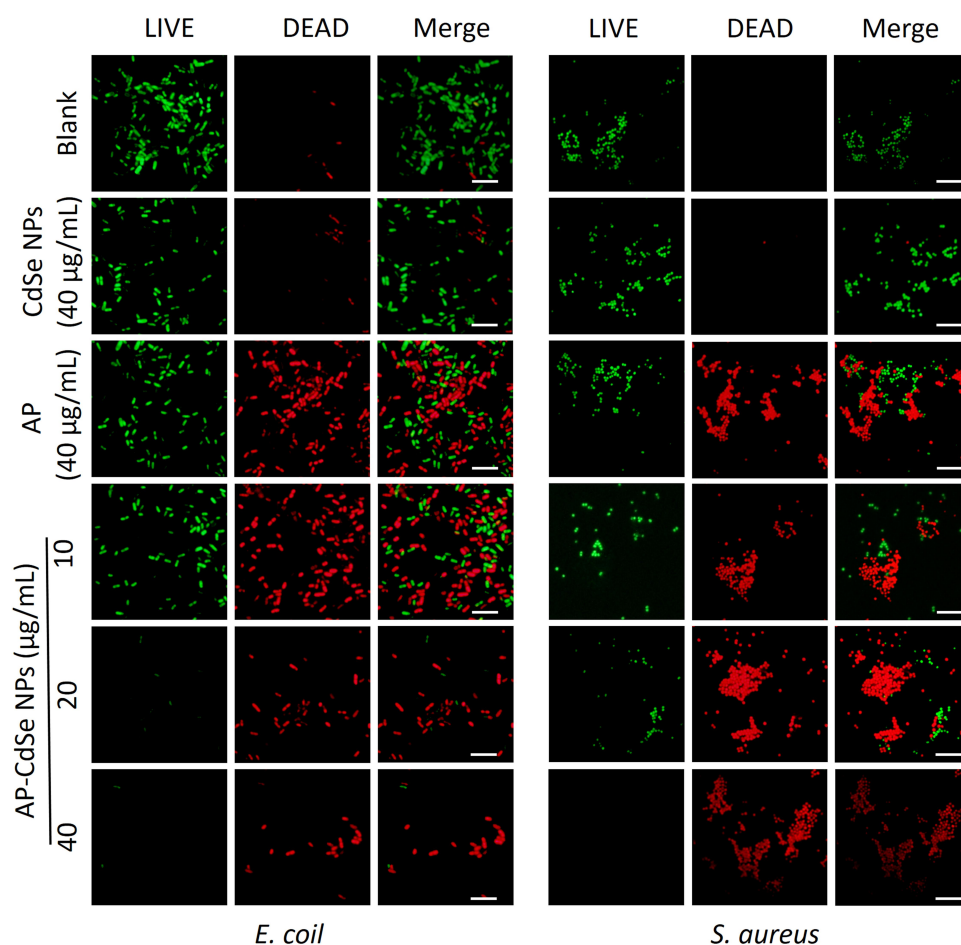


Figure 4 Confocal fluorescence microscopy assay. MDR *E. coli* and MDR *S. aureus*, after treatment with AP-CdSe NPs at various concentrations (10, 20, 40 µg/mL). AP solution (40 µg/mL) and CdSe NPs solution (40 µg/mL) were used as control groups. The PBS-treated group was used as blank group. All cells were stained for 30 min with SYTO 9 (green fluorescence) and PI (red fluorescence). Scale bar = 10 µm.

analysis, AP-CdSe NPs have highly effective antibacterial activity against MDR *E. coli* and MDR *S. aureus*. Changes in cell integrity may be the main mechanism of inducing bacterial cell apoptosis.⁴³ Furthermore, the effects of the antibacterial activity of AP-CdSe NPs on the cell morphology and structure were examined by SEM (Figure 6) and subcellular ultrastructure analysis by TEM (Figure 7).

The integrity of the bacterial cell, including the cell membrane, was examined by SEM, as shown in the SEM images of MDR *E. coli* and MDR *S. aureus* treated with AP-CdSe NPs, displayed in Figure 6. The *E. coli* and *S. aureus* cells in the blank group maintained complete cell morphology and the integrity of the membrane structure. The enlarged region confirmed that cells were smooth and had an intact cell membrane. Similar integrity of the membrane structure was observed in bacterial culture treated with CdSe NPs. Also, the results revealed that CdSe NPs alone had no antibacterial activity. The *E. coli* and

S. aureus cells in the AP-treated group showed damaged membrane structures, and the enlarged region revealed that cells had not intact cell membrane and leaked intracellular contents. The AP showed effective antibacterial activity against MDR *E. coli* and MDR *S. aureus* based on the damaged cell area. After the *E. coli* and *S. aureus* cells were treated with AP-CdSe NPs (40 µg/mL), the damaged cell area became larger in SEM images. The number of damaged cells increased significantly, and the results also indicated that the damage to the cells was more severe with increasing concentration of AP-CdSe NPs, indicating that the sensitivity of bacteria to AP-CdSe NPs is concentration-dependent. Taken together, these results indicated that AP-CdSe NPs had highly effective antibacterial activity compared with CdSe NPs and AP alone.

The evaluation of the antibacterial activity of AP-CdSe NPs through subcellular ultrastructure analysis by TEM revealed the changes in the internal structure of cells

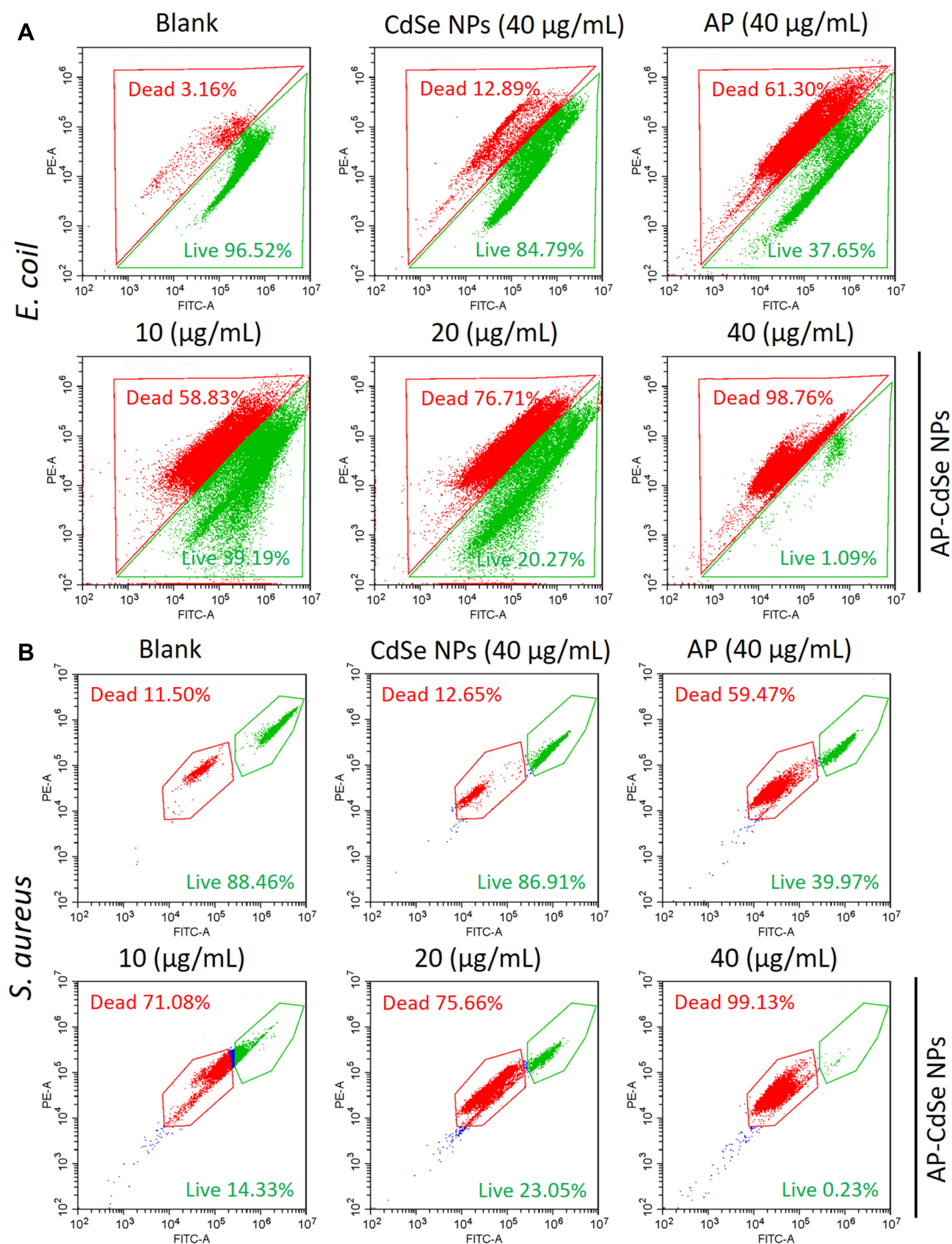


Figure 5 The results of the apoptotic cell death assay by flow cytometry analysis were statistically analyzed by the CytExpert software. MDR *E. coli* (A) and MDR *S. aureus* (B), after incubation with AP-CdSe NPs at various concentrations (10, 20, 40 $\mu\text{g/mL}$). AP solution (40 $\mu\text{g/mL}$) and CdSe NPs solution (40 $\mu\text{g/mL}$) were used as control groups. The PBS-treated group was used as blank group. All cells were subsequently briefly (30 min) stained with SYTO 9 and PI. Data are expressed as the mean \pm SD ($n=3$).

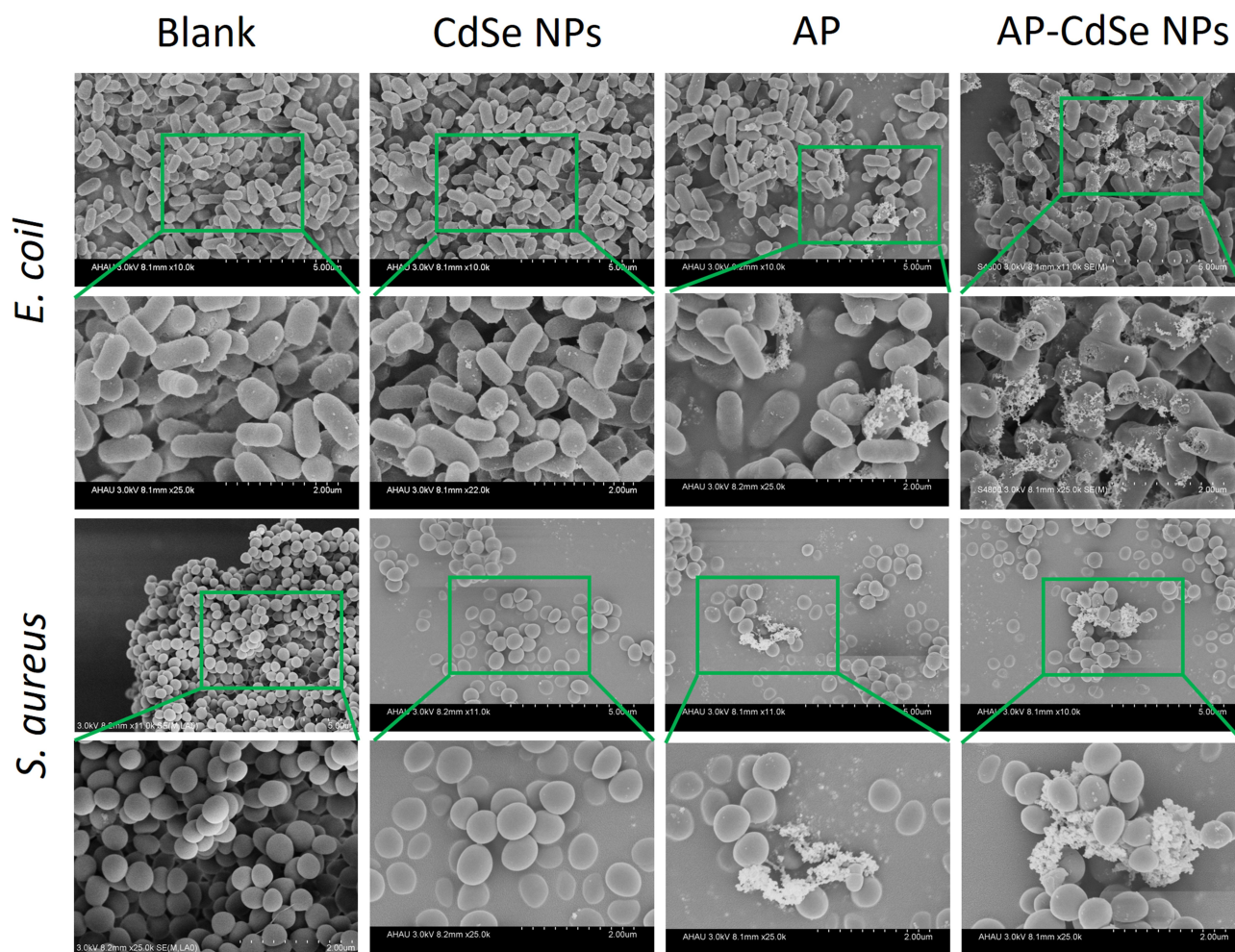


Figure 6 SEM images of MDR *E. coli* and MDR *S. aureus*. Bacterial culture treated with AP-CdSe NPs (40 $\mu\text{g/mL}$). Bacterial culture treated with AP solution (40 $\mu\text{g/mL}$) and CdSe NPs solution (40 $\mu\text{g/mL}$) were used as control groups. The PBS-treated group was used as blank group. Green squares indicated the enlarged regions.

compared with the SEM images.⁴⁴ As shown in the Figure 7, the internal structure of *E. coli* and *S. aureus* cells was intact in the blank group; the TEM image clearly show the bacterial nuclear and cytoplasmic areas inside the cell. Similarly, the internal structure of *E. coli* and *S. aureus* cells was also intact in bacterial culture treated with CdSe NPs. The subcellular ultrastructural analysis of bacterial culture treated with AP by TEM revealed that their internal morphology was significantly different from that of normal cells, as cells in the field of view appeared broken and the cytoplasm was missing. Moreover, in cells of the group treated with AP-CdSe NPs the cell damage was more pronounced than that in cells of the AP-treated group. Additionally, it was also found that the number of damaged cells in the AP-CdSe NPs group increased with the increase of the concentration of AP-CdSe NPs. The SEM and TEM images revealed that AP-CdSe NPs induce cell death combined with the destruction of the cell

integrity, suggesting a possible mechanism of cell death that needs further verification.

Gene Expression Analysis

Gene expression profiling, such as by RNASeq analysis, can be used to study the metabolism at the cellular molecular level. cDNA from *E. coli* (control and AP-CdSe NPs groups) was sequenced in triplicate. Statistical analysis revealed a high total number of reads of the sequencing samples and a high ratio of high-quality reads as shown in the Table 2. The results suggested that the sequencing data were of good quality. In order to identify DEGs, the expression of each gene can be analyzed by FPKM (the expected number of fragments per kilobase of the transcript sequence per kilobase pair). The HTSeq program was used to analyze differential gene expression. The density estimation of gene expression levels in the AP-CdSe NPs and control groups is compared in Figure 8A. The results showed that the

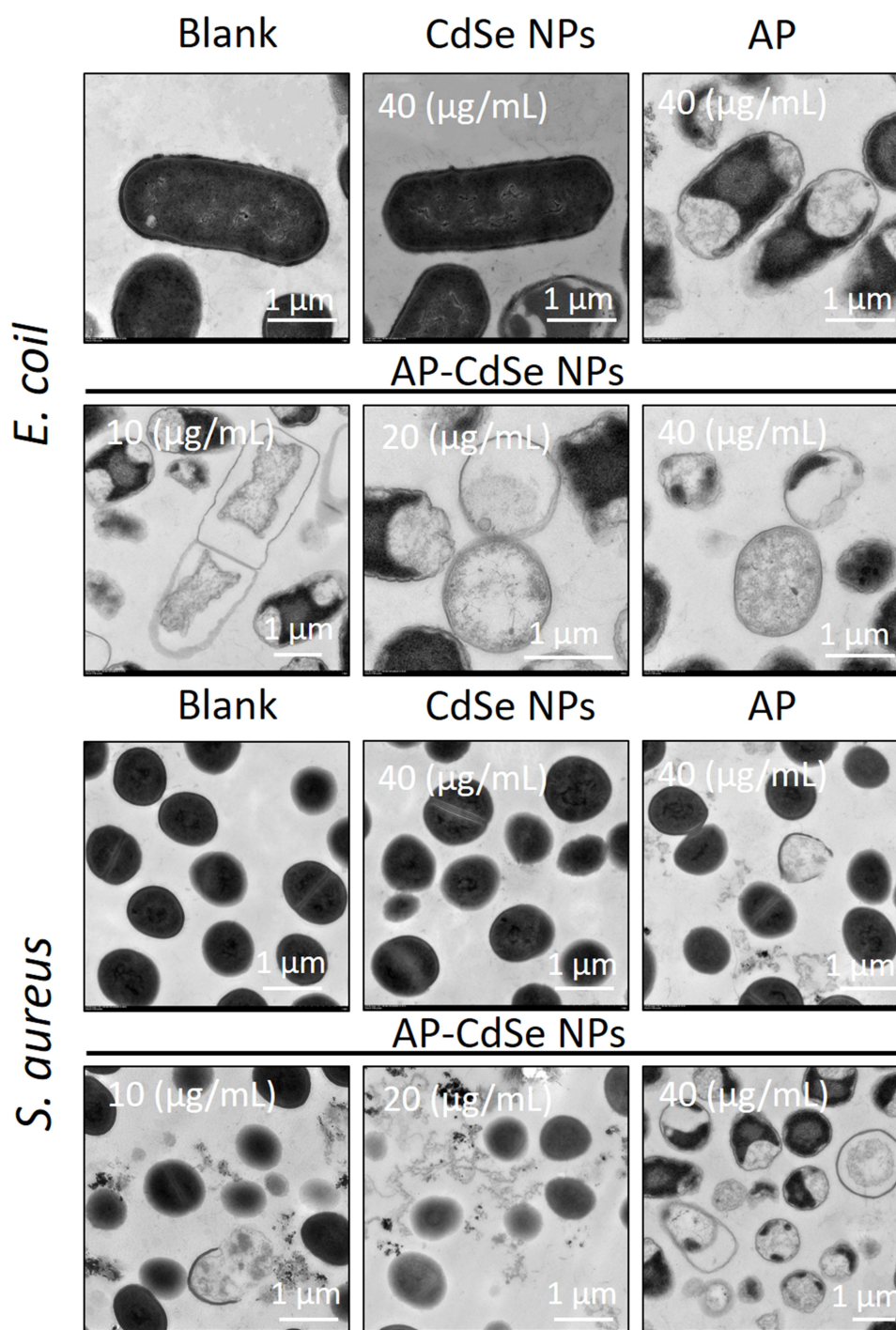


Figure 7 TEM images of MDR *E. coli* and MDR *S. aureus*. Cell treated with AP-CdSe NPs solutions at various concentrations (10, 20, 40 µg/mL). Bacterial culture treated with AP solution (40 µg/mL) and CdSe NPs solution (40 µg/mL) were used as control groups. The PBS-treated group was used as the blank group.

AP-CdSe NPs affected gene expression. To determine the differential gene expression in *E. coli* bacterial culture treated with AP-CdSe NPs, this study identified DEGs relative to control groups. As shown in Figure 8B, the number of DEGs was 373, including 216 upregulated genes and 157 down-regulated genes. Gene expression cluster analysis was used

to determine the clustering pattern of gene expression in the control group and AP-CdSe NPs group, and the results are shown in Figure 8C. We selected the top 50 DEGs with the largest expression variance for cluster analysis. These DEGs will be used for GO and KEGG enrichment analysis. The correlation analysis between samples is based on the

Table 2 The RNASeq Outcomes of Control and AP-CdSe NPs Samples

Sample Name	Raw Reads	Clean Reads	Error Rate (%)	Clean Q20(%)	Clean Q30(%)	GC Content (%)
Control	29120592	28992866	0.01	98.37	95.02	51.24
AP-CdSe NPs	28071310	27921738	0.01	98.16	94.54	51.36

expression levels of all genes or transcripts obtained by sequencing and is a way to analyze whether the expression profiles between two samples are correlated. As shown in Figure 8D, the results indicated a high degree of correlation between the control and AP-CdSe NPs. Also, the experiment with biological replicate samples is reliable and the sample selection is reasonable. The R^2 value (0.859) < 0.9 indicated a poor-level of correlation between the control group with the AP-CdSe NPs group. Such result may be due to the treatment of *E. coli* cells with AP-CdSe NPs. Taken together, these

results suggested that the *E. coli* bacterial culture treated with AP-CdSe NPs had shown differential expressions. This may be the antibacterial molecular mechanism of AP-CdSe NPs.

Antibacterial Pathway Analysis by GO and KEGG Enrichment Analysis

GO is often used to provide background knowledge of gene function classification labels and gene function research. The use of the GO database to search through species and gene information, thus provides the GO

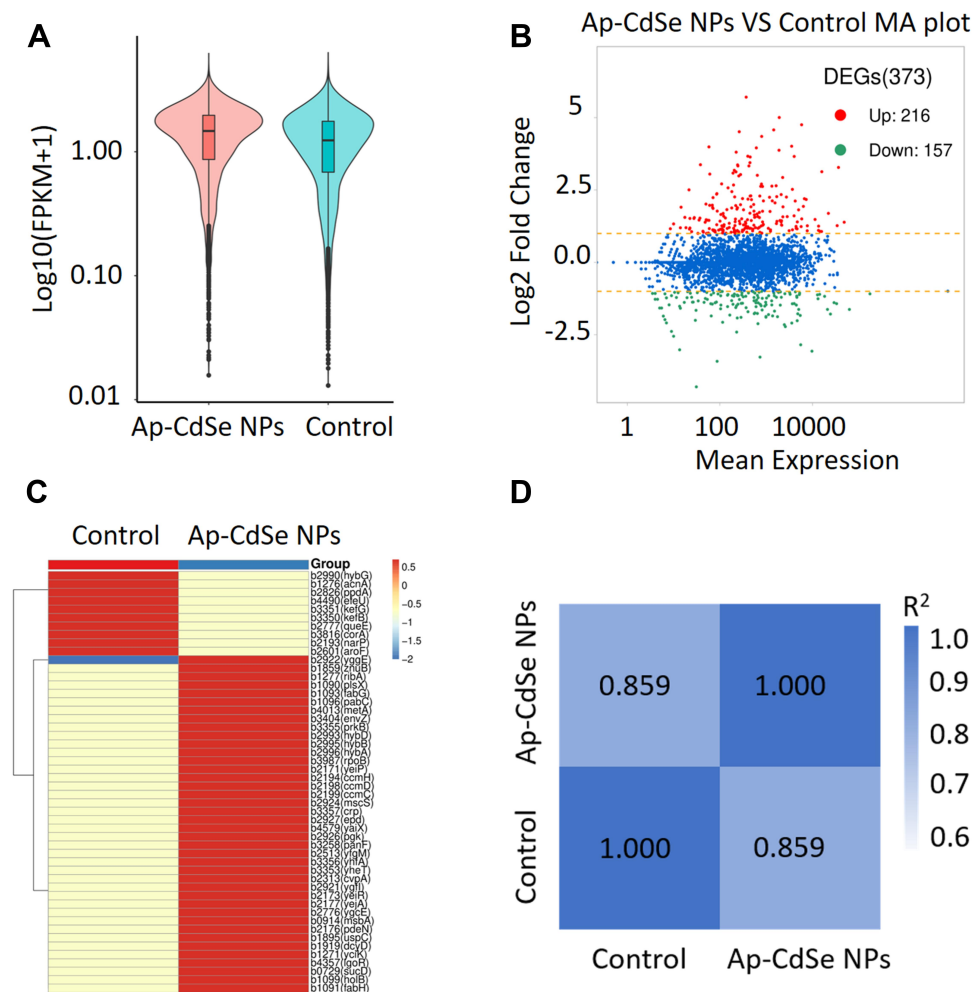


Figure 8 Differentially expressed genes (DEGs) analyzed in MDR *E. coli* (AP-CdSe NPs and control). **(A)** FPKM density distribution of *E. coli* genes in control and AP-CdSe NPs groups. **(B)** Number of DEGs comparing the AP-CdSe NPs group with the control group. **(C)** Gene expression cluster analysis is used to determine the clustering pattern of gene expression in different treatment groups. **(D)** Pearson correlation coefficient between samples.

annotation information (functional information) of the DEGs. The distribution information and significance of the gene set in the GO category are obtained based on the GO annotation of the gene. GO analysis identified a total of 6 terms related to cellular components (CC), 15 terms for biological processes (BP), and 15 terms for molecular functions (MF) after *E. coli* treatment with AP-CdSe NPs (Figure 9). Regarding CC ontology, the major represented categories were “small molecule catabolic process” (GO:0044282) and “organic acid catabolic process” (GO:0016054). Regarding BP ontology, the major represented category was “periplasmic space” (GO:0042597). Regarding MF ontology, the major represented categories were “small molecule binding” (GO:0036094) and “bacterial-type proximal promoter sequence” (GO:0000986).

The relative cDNA abundance of GO analysis terms was detected by RT-PCR analysis, and the results confirmed that AP-CdSe NPs significantly changed the cDNA abundance. All results showed that gene expression in *E. coli* bacterial culture treated with AP-CdSe NPs suggested a molecular mechanism for the antibacterial effect of AP-CdSe NPs.

KEGG is a database containing biochemical reactions, signaling pathways, metabolic pathways and biological processes.⁴⁵ The biological pathway enrichment analysis based on the KEGG database is performed on the DEG set, and pathways with significantly enriched DEGs are extracted, which is useful for the development of downstream experiments. Through the KEGG annotation of genes, the hypergeometric distribution is used to calculate

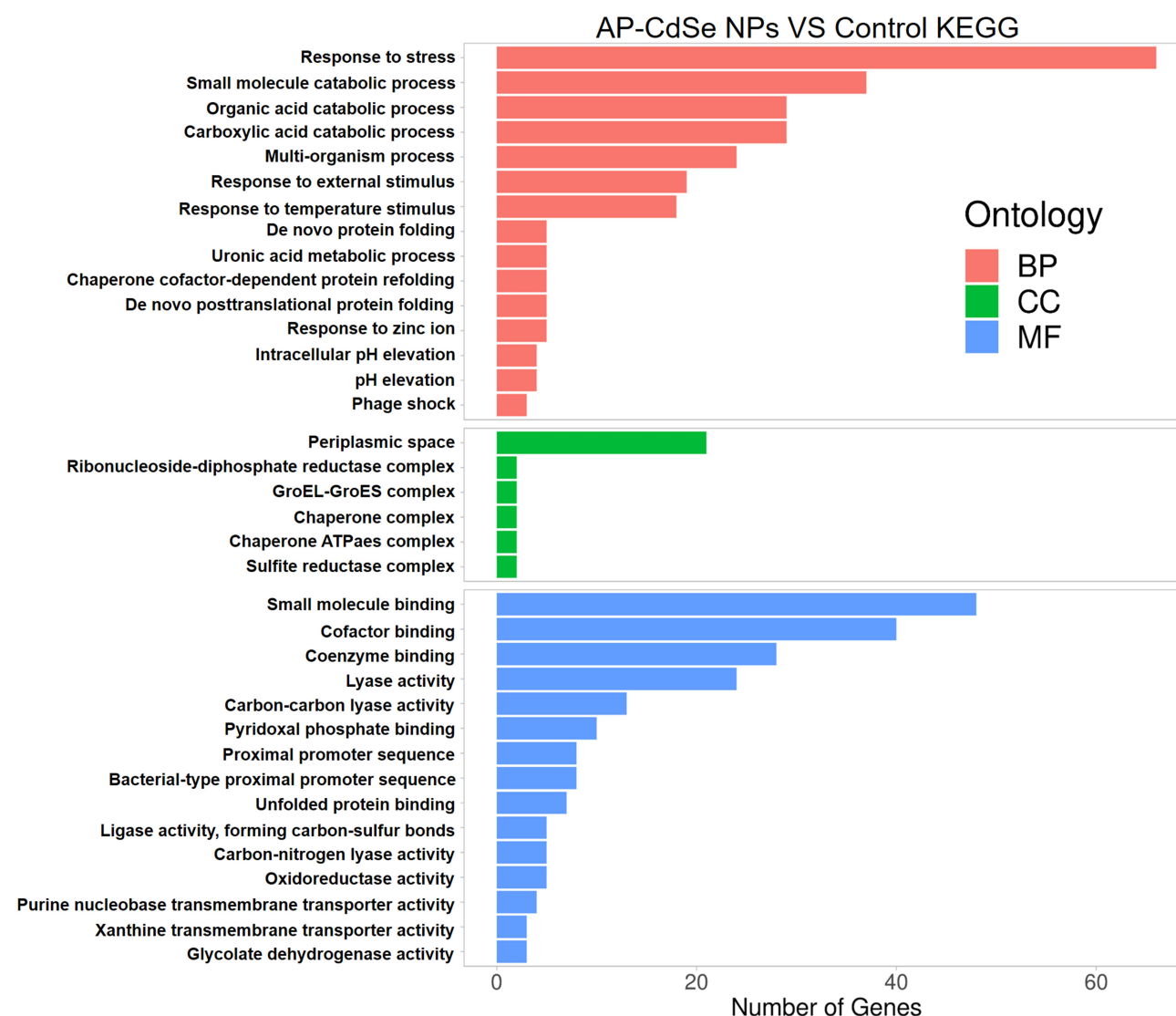


Figure 9 Differentially expressed genes (DEGs) GO enrichment analysis results.

the P value, and a $P < 0.05$ is used as the significance threshold to obtain statistically significant high-frequency annotations relative to the background, in order to obtain the distribution information and significant enrichment of genes in KEGG pathways (Figure 10). KEGG enrichment analyses of *E. coli* treated by AP-CdSe NPs transcriptome sequences revealed the presence of significant enrichment of DEGs in three pathways compared with the control group. The results showed that AP-CdSe NPs affected the bacterial metabolic pathways, thus inhibiting *E. coli* cell growth.

Antibacterial Activity of AP-CdSe NPs in vivo

Through in vitro antibacterial test, we confirmed that AP-CdSe NPs have high antibacterial activity. At the same time, transcriptome analysis was performed to determine the antibacterial molecular mechanism of AP-CdSe NPs. Further evaluation of the antibacterial activity of AP-CdSe NPs in vivo will greatly increase their potential clinical application. Due to its drug resistance and high mortality, *S. aureus* infections have become a major problem in public health worldwide. Therefore, we explored and established a bacteremia model in mice infected with *S. aureus*.

The biodistribution of bacteria in mice of different treatment groups (blank, infection and treatment groups) was analyzed at different times to determine the antibacterial

activity of AP-CdSe NPs in vivo. As shown in Figure 11A, the number of bacteria in blood and organs in the blank group was almost the same ($\sim 1.1 \times 10^4$ CFU/mL or g) and remained at such level for 7 days. After injecting bacteria into the mice through the tail vein, the number of bacteria in blood and organs was $\sim 3.3 \times 10^4$ CFU/mL or g in the infected group at 7 days. These results indicated that the bacteremia model was successfully developed. The AP alone treated mice with bacteremia exhibited antibacterial effect, as indicated by the significant decrease in the number of bacteria in these mice, although the clinical symptoms of bacterial infection were still present. This finding showed that AP alone was not enough to cure mice with bacteremia. However, when AP-CdSe NPs was injected into *S. aureus* infected mice, the number of bacteria in the blood and organs of the mice gradually decreased over time. The number of bacteria was almost the same as that in the blank group at 7 days, indicating that AP-CdSe NPs could also inhibit bacterial activity in vivo and cure mice with bacteremia. The survival rate and weight of mice were monitored after tail vein injection, as shown in Figure 11B. The survival rate of the AP-CdSe NPs group was 83%, which was significantly higher than that (31%) of the non-treated infected group. The results showed that AP-CdSe NPs improved the survival rate of mice with bacteremia compared with the AP-treated *S. aureus* infected group. This finding further confirmed the antibacterial effect of AP-CdSe NPs in vivo.

Clinical evaluation of the treatment of mice with bacteremia can be achieved not only through the biological distribution of bacteria, but also by immunostaining of major organs. The mice in the blank group did not have any histological damage or bacteremia. However, the organs and tissues of the *S. aureus* infected mice presented obvious symptoms of clinical bacteremia. Meanwhile, in the AP treatment group, the organ tissue damage of the mice was repaired, but there were obvious symptoms of clinical bacteremia. On the other hand, in the AP-CdSe NPs treatment group, the injured tissue completely recovered and appeared to be in a normal tissue state (Figure 11C). These results indicated that AP-CdSe NPs can heal tissue damage caused by bacteremia in mice. This result is consistent with the result of the biodistribution of bacteria shown in Figure 11A, confirming the highly effective antibacterial effect of AP-CdSe NPs in vivo.

Safety Evaluation

Side effects of the AP-CdSe NPs were examined to evaluate its safety. Blood biochemical assays were also

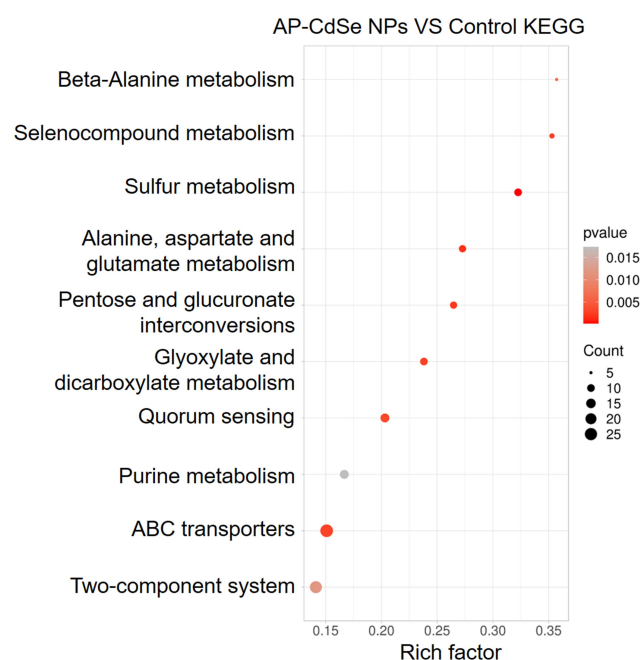


Figure 10 Differentially expressed genes (DEGs) KEGG enrichment analysis results.

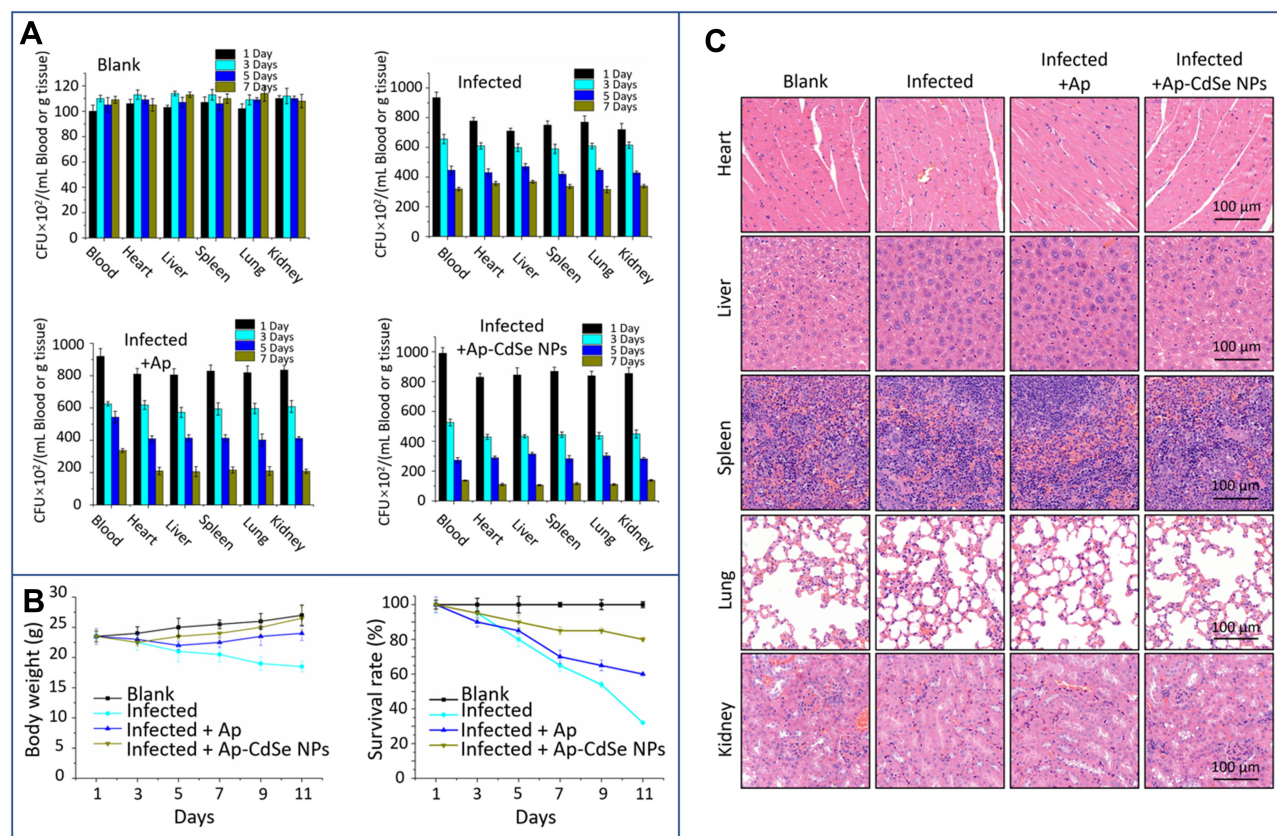


Figure 11 The antibacterial test in the animal bacteremia model. **(A)** Bacterial count in blood and major organs (heart, liver, spleen, lungs, and kidneys) in blank, infected, and treatment groups. **(B)** The survival rate and weight of mice were monitored after tail vein injection. **(C)** Images of H&E-stained tissues of major organs from mice in the blank, infected, and treatment groups.

performed to examine possible organ toxicity in mice after treatment with AP-CdSe NPs. As shown in Figure 12A, the levels of blood glucose, cholesterol, uric acid, and alanine aminotransferase (indexes for the function of kidney, liver, blood glucose, and blood lipids, respectively) all showed no significant changes, compared with the blank group, and were within normal range values. The assessment of histopathological changes by H&E staining of major mouse organs revealed that the AP-CdSe NPs treatment groups all showed similar histological features compared with the blank groups, and no obvious impairment or inflammation was observed in any of the groups (Figure 12B). These results confirmed that AP-CdSe NPs was non-toxic and had no side effects.

Conclusion

In this study, an AP was synthesized by the Fmoc solid-phase method and decorated on the CdSe NPs. The results of the characterization of the newly developed AP-CdSe NPs revealed that they had good monodispersity, size stability and consistent spectral characteristics. The AP-CdSe NPs

had a diameter of approximately 14 nm and showed excellent fluorescence properties. In addition, CFU assays showed that AP-CdSe NPs display a broad-spectrum antibacterial activity, and had highly effective antibacterial activities against MDR *E. coli* and *S. aureus*. The MIC assays showed that the AP-CdSe NPs had highly effective antibacterial activities at low dosages of 8.0 μg/mL against MDR *E. coli* and MDR *S. aureus*. Fluorescence microscopy analyses revealed that AP-CdSe NPs had high cellular uptake rate and killing rate for *E. coli* and *S. aureus*. The quantitative analysis of live/dead cells by flow cytometry analysis further confirmed the antibacterial activity of AP-CdSe NPs. Moreover, SEM and TEM observations were performed to investigate the mechanisms of cell integrity by examining changes in cell membrane and wall. Gene expression analysis of *E. coli* bacterial culture treated with AP-CdSe NPs indicated a molecular mechanism for the antibacterial effect of AP-CdSe NPs. GO and KEGG pathway enrichment analyses revealed key biological pathways involved in the metabolic pathways of *E. coli* treated with AP-CdSe NPs. In addition, AP-CdSe NPs inhibited bacterial

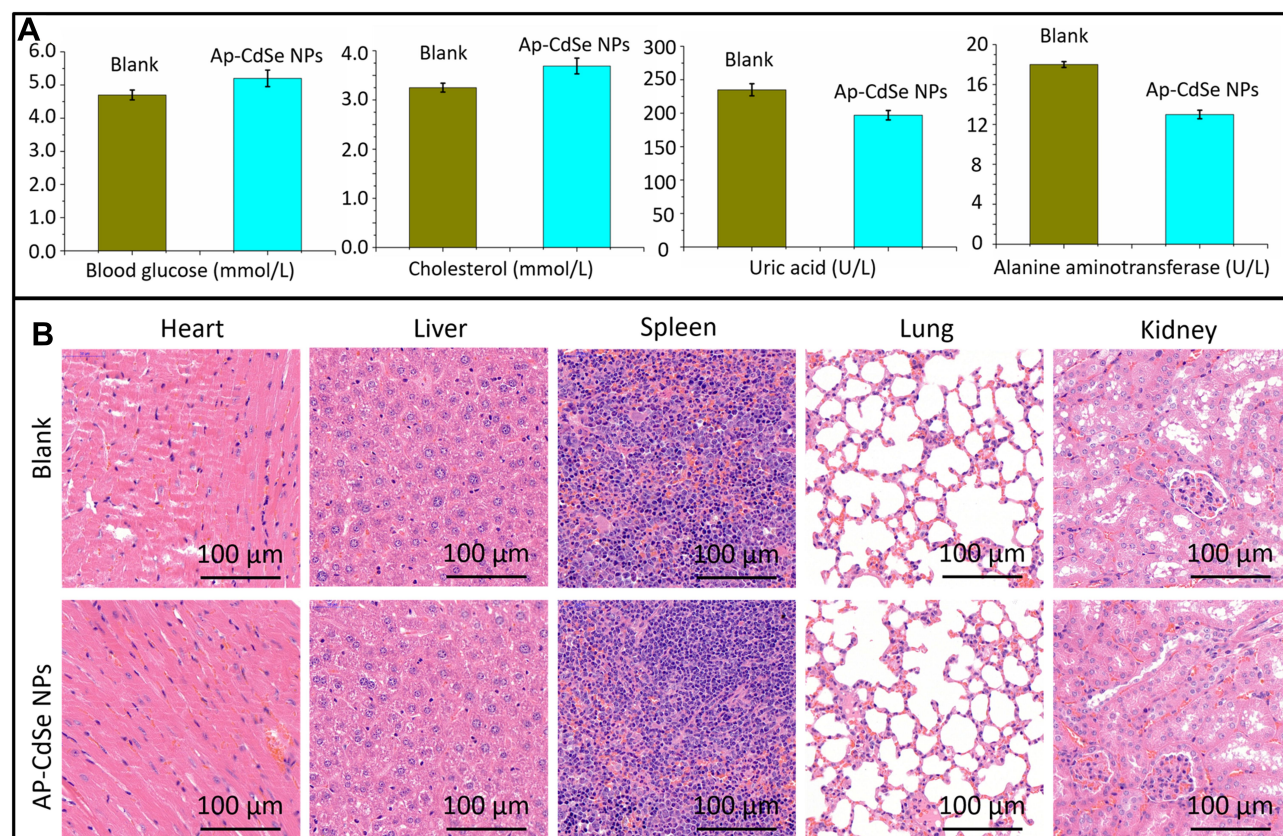


Figure 12 Biosafety evaluation of AP-CdSe NPs. **(A)** Blood biochemistry, liver function markers (alanine aminotransferase), kidney function markers (Uric acid), blood glucose, and cholesterol were all measured. **(B)** Histopathological changes in major mouse organs examined by H&E staining.

activity and healed tissue damage in a bacteremia model in mice infected with *S. aureus*. The evaluation of the biosafety of the AP-CdSe NPs showed that they do not have any side effects. This study also reports that the transcriptome analysis of the mechanism provides novel insights into the molecular mechanism of the antibacterial effects of AP-CdSe NPs. Both the in vitro and the in vivo studies suggested that AP-CdSe NPs can be a potential nanoscale antimicrobial agent against *E. coli* and *S. aureus*.

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Author Contributions

All authors contributed to data analysis, drafting or revising the article, have agreed on the journal to which the article will

be submitted, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Disclosure

The authors have declared that there are no competing interests in this work.

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