



The Influence of Nanostructured Hydroxyapatite Surface in the Early Stages of Osseointegration: A Multiparameter Animal Study in Low-Density Bone

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Background and Objective: The success rates of dental implants in low-density bone have been reported as a challenge, especially for early or immediate loading in the maxilla posterior area. Nanoscale architecture affects the roughness, surface area, surface energy of the implant and can enhance osseointegration. This study aimed to evaluate the implant-surface topography and biomechanical, histomorphometric, and histological bone responses to a new nanostructured hydroxyapatite surface placed in the iliac crest of sheep.

Methods: Ten female sheep (2–4 years) received 30 implants (n=10/group): HAnano[®] coated (Epikut Plus[®], S.I.N. Implant System, Sao Paulo, SP, Brazil), SLActive (BLX[®], Straumann, Basel, Switzerland), and TiUnite (NobelActive[®], Nobel Biocare, Göteborg, Sweden) surfaces. Scanning electron microscopy with energy-dispersive spectroscopy evaluated the implant surface topography, the insertion torque value, and resonance frequency analysis evaluated the primary stability, bone-implant contact, and bone-area fraction occupancy were evaluated after 14 and 28 days after implant placement.

Results: The surface morphology was considerably comparable between the implant groups^{*}; however, the TiUnite[®] group presented a remarkable different surface. The SLActive[®] and TiUnite[®] groups presented an insertion torque average of 74 (±8.9) N/cm that was similar to that of HAnano[®] 72 (±8.3) N/cm (p >0.05). The resonance frequency evaluated with Osstell[®]/SmartPeg[®] or Penguin[®]/MulTipeg[®] showed similar results when assessing implants from the same group. BIC and BAFO significantly increased (p<0.05) throughout the experimental periods to all groups, but BIC and BAFO values were similar among the implants at the same time point. After 4 weeks, bone-implant contact was higher than 80% of the total length analyzed. New bone occupies around 60% of analyzed area around the implants.

Conclusion: HAnano[®] coated surface promoted comparable osseointegration as SLActive and TiUnite in the sheep model. The three tested surfaces showed comparable osseointegration at the early stages of low-density bone repair in the sheep model.

Keywords: osseointegration, dental implant surface, nanotechnology, hydroxyapatite, sheep, bone response

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Introduction

Dental implantation in partial or totally edentulous patients is a predictable treatment with high rates of long-term success;¹ however, early osseointegration is still considered a challenge in areas with the most trabeculated bone (bone type IV)²

together with lower-density and thinner cortical bone, which is generally considered less suitable for supporting dental implants. In recent years, implant-surface technologies with micro- to nanotopography, chemical compositions, macro-scale implant designs, loading protocols, and new surgical procedures have been introduced to improve the osseointegration and reduce the treatment term, allowing immediate or early functional loading in patients with reduced bone density.^{3–5}

The implant's surface is critical for guaranteeing dental implants' osseointegration, preferably in the short term. Various approaches to improve these surfaces have been studied, including mechanical (eg, machining, grinding, polishing, blasting), chemical (eg, acid treatment, alkali treatment, hydrogen peroxide, and anodic oxidation), and physical (eg, plasma and flame spray, ion beam implantation)⁶ techniques.

Calcium phosphate has been widely used as a bone substitute,^{7,8} dental-implant coating,⁹ and carrier for proteins,¹⁰ growth factors,¹¹ and drugs^{12–14} due to its similarity to bone tissue's mineral content and suitable cell-attachment capacity. Among the calcium phosphates, hydroxyapatite in particular has been favored given its biocompatibility, safety, predictability, unlimited availability, lower morbidity for the patient, and cost-effectiveness, which offer significant advantages and make it a good choice for dental-implant coatings.¹⁵ The use of nanostructured hydroxyapatite when coating titanium-implant surfaces improves the characteristics of the implant surface, increases the strength of the bond between titanium and bone, enhances the rate of osseointegration, and reduces the length of the treatment period, especially in patients whose bone quality is poor.⁹

Early experiences with coatings applied to dental implants in the 1990s failed for various reasons, including a detachment of the hydroxyapatite coating and dissolution of the detached HA.^{16,17} However, more recently, coating methods have been improved and ion-sputtering or thermal-plasma treatment has been successfully applied.^{18,19} This has resulted in more durable coatings that adhere better to the titanium substrate and, therefore, show more significant promise for clinical use mainly in the posterior area of maxilla and immediate implant installation. The demands from specific locations suggest that coated implants might be necessary to ensure better clinical results, renewing the interest in coated dental implants.

Topographical modifications to dental-implant surfaces can vary from millimeter-wide grooves to nano-sized

structures.²⁰ The nanometric modifications on the surface of osseointegratable implants are made based on size-equivalent arrangements present in bone, which result in a well-organized, three-dimensional nanotopography. Besides, the biomolecules and cells involved in the early healing phase after implant installation will interact at the nanometer level.^{21–23}

There is a continuous increase in the effort to develop novel and bioactive surfaces to reduce the osseointegration period in low-density bone is ongoing. Since current coatings have not yet shown the necessary levels of improvement for bone with low rates of remodeling and in the early time, new implant surfaces have been researched and developed. This study aimed to compare the bone response of the HAnano[®] coated surface (S.I.N. Implant System, Sao Paulo, Brazil) to the well-known hydrophilic surfaces such as the SLActive[®] (Straumann, Basel, Switzerland) and TiUnite[®] (Nobel Biocare, Gothenburg, Sweden) through histomorphometric analysis of bone-implant contact (BIC) and bone area fraction occupancy (BAFO) in sheep model of low-density bone.

Materials and Methods

Implants and Surface Characteristics

Thirty titanium dental implants (3.5 mm in diameter and 10 mm in length) with the following three different surface modifications were used (n = 10/group): the HAnano group (Epikut Plus[®]; Sistema de Implante Nacional SA, São Paulo, Brazil) included implants with a nano-sized crystalline hydroxyapatite coating; the SLActive group (BLX[®]; Straumann, Basel, Switzerland) included implants with a hydrophilic sandblasted and acid-etched titanium surface; and the TiUnite group (NobelActive[®]; Nobel Biocare, Göteborg, Sweden) included implants with a highly crystalline and phosphate-enriched anodized titanium-oxide surface.

High-resolution scanning electron microscopy images obtained with FEI-QUANTA 450 (Thermo Fisher Scientific, Waltham, MA, USA) revealed the surface topography of the implants, one sample per group, at an accelerating voltage of 10 kV; focal width of 3.0; and magnifications of 50×, 3000×, and 15,000×. Energy-dispersive X-ray spectroscopy analysis determined each surface's chemical composition at an acceleration voltage of 20 kV and focal width of 40 using an EDAX detector equipped with a dual-beam electron microscope (AMETEK Materials Analysis Division, Mahwah, NJ,

USA) and the Genesis software program (EDAX, LLC, Mahwah, NJ, USA).

Animal Model

This study report followed the Animal Research: Reporting of in vivo Experiments (ARRIVE)²⁴ and Planning Research and Experimental Procedures on Animals: Recommendations for Excellence (PREPARE)²⁵ guidelines where appropriate. Ten adult female Santa Ines sheep were used for this study, with an average age of three years (2–4 years) and an average weight of 37.05 kg (31–42 kg). To minimize the effects of subjective bias when allocating animals to treatment, the animals were randomly allocated into two experimental periods of two and four weeks using the coin-toss method of randomization. Animal experiments and breeding occurred under conditions in compliance with the National Institutes for Health Guide for the Care and Use of Laboratory Animals²⁶ and relevant Brazilian legislation on animal use.²⁷ An experienced veterinarian conducted nutritional recommendations, animal care, and fasting in pre- and postoperative animals. The Ethics Committee for Animal Use of Fluminense Federal University in Cachoeira de Macacu, Rio de Janeiro, Brazil approved the protocol of this study (CEUA # 9,531,061,119). The animal experiments occurred during the first half of 2020. None of the animals were euthanized after the end of the study in compliance with the guidelines of the 3Rs Program (“Reduction, Refinement, and Replacement”), whose objective is to reduce the number of animals used during experimentation, minimize pain and discomfort, and avoid euthanasia at the end of experimentation.²⁸ All animals were immunized against common diseases for sheep and were submitted to tests to ensure good physical condition. The Veterinarian School Farm of Fluminense Federal University harbored the animals used in this study in a fenced semi-extensive system with native fodder and signal grass (*Brachiaria humidicola* and *Brachiaria decumbens*). In the preoperative period, the animals received food composed of the pastures above and, during the postoperative period, in addition to the mentioned pastures, nutritional supplementation proper for sheep. Salt mineral water ad libitum was available during the entire experimental period.

Aiming to reduce preoperative stress levels in the animals, the veterinarian moved the sheep two weeks before the surgery from the breeding site to the research center

for adequate acclimatization.²⁹ The animals were fasted for eight hours before the surgery.

Sample-Size Calculation

A significant effect of 15% or more 28 days after surgery in the BIC (primary outcome) would be of interest. Considering previous rates³⁰ for BIC at 28 days postsurgery in the control group and intervention group of 66.5% and 76.5%, respectively, with a two-sided significance of 0.05 and a power of 0.8, this study required a total of five sheep.^{31,32}

Surgical Model

The pelvic animal model was used for all experiments according to previous research.^{30,33–36} The cranial part of each animal’s right iliac wing (n = 5/group) received all implants (Figure 1). Five implants were installed in each animal, but only three were used for this study. The other two were used for another study.

Anesthetics and Analgesia

The animals were premedicated with 0.05 mg/kg of acepromazine intravenously (Acepran[®]; Vetnil, Louveira, São Paulo, Brazil), 0.2 mg/kg of diazepam intravenously (Teuto, Anapolis, Goiás, Brazil) and 0.4 mg/kg of morphine intramuscularly (Dimorf[®]; Cristalia, Itapira, São Paulo, Brazil). After 20 minutes, with the animals exhibiting no response to pain, cephalic-vein cannulation was initiated for the administration of 5 mL/kg/h of lactated Ringer’s solution intravenously (Baxter Hospitalar LTDA; São Paulo, São Paulo, Brazil). Anesthesia was induced with 4 mg/kg of propofol intravenously (Baxter Hospitalar LTDA; São Paulo, São Paulo, Brazil) and maintained using 1% isoflurane (Cristalia, Itapira, São Paulo, Brazil) following orotracheal intubation. Meanwhile, 4 mg/kg of lidocaine (Xylestesin[™]; Cristalia, Itapira, São Paulo, Brazil) and 0.1 mg/kg of morphine (Dimorf[®]; Cristalia, Itapira, São Paulo, Brazil) was used to block the epidural. A very experienced veterinarian managed the anesthesia and supervised all procedures throughout the surgery.

Surgical Procedures

The right side of the iliac crest was initially shaved with a razor blade, with subsequent application of an antiseptic chlorhexidine 0.5% solution. An incision of approximately 5 cm in length was made over the region of the animal’s iliac wing (right side) using scalpel handle no. 3 (Bard

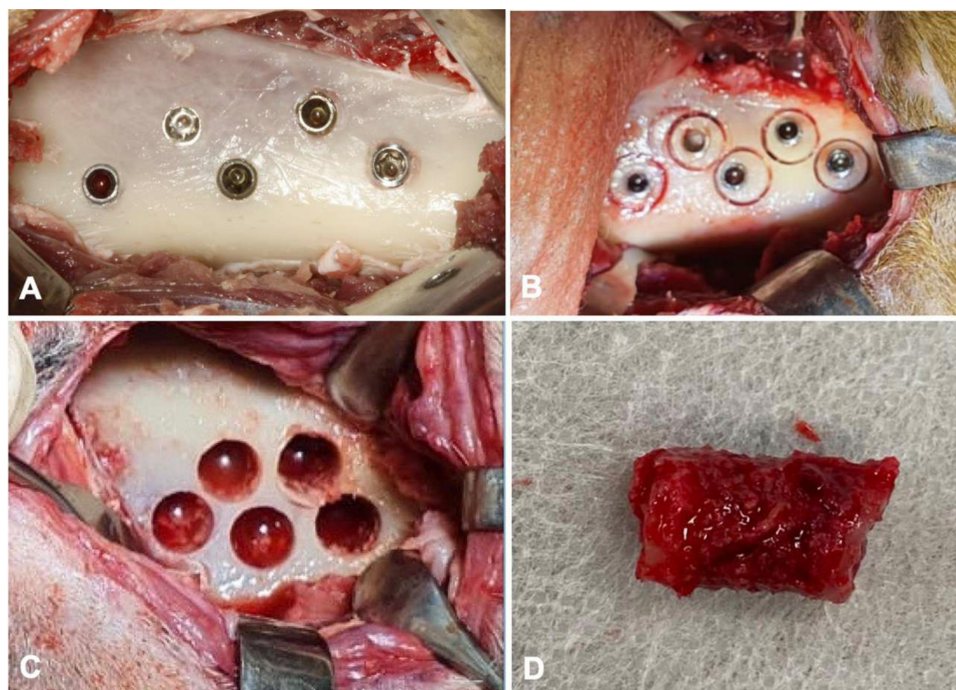


Figure 1 Surgical procedures for implant's installation of the 5 implants. **(A)** Implants installed in the iliac crest with a minimum distance of 5 mm between each implant; **(B)** after 2 and 4 weeks, the implants were trephined with a trephine drill (internal diameter = 5 mm) to remove the bone block containing the implant; **(C)** bone defects after the removal of the bone blocks and **(D)** bone block trephined with implant inside.

Parker[®], Aspen Surgical, Caledonia, MI, USA) and blade no. 15 (Solidor[®]; Lamedid, Osasco, São Paulo, SP, Brazil). After the incision was made, the skin, muscle, and periosteum were detached for bone perforations. The sequence of burs recommended by the manufacturer was used in low rotation (1200 rpm), with profuse irrigation with a 0.9% sodium chloride solution (Darrow Laboratórios SA, Rio de Janeiro, RJ, Brazil) to avoid tissue necrosis due to overheating. Five implants were installed (one for each group and two for another study) with the aid of a counter-angle coupled to a surgical electric micromotor (BLM 600 plus; K Driller, São Paulo, Brazil) with the capacity for 24 rpm. The implants were placed in the first sheep after randomly selecting the position for each using the sealed-envelope method. In the next sheep, the implant position was rotated clockwise, and all five implants were placed in different locations. At the end, no implant used the same implantation site as another. The inter-implant distance was at least 5 mm. The surgeons evaluated the resonance frequency analysis (RFA) for all implants using Osstell[®] and Penguin[®] with magnetic transducers SmartPeg[®] (Integration Diagnostics, Savedalen, Sweden) and MultiPeg[®] (Penguin Integration Diagnostics, Göteborg, Sweden), respectively.

Finally, absorbable sutures (Vicryl 4-0; Ethicon, Inc., Somerville, NJ, USA) were used to close the repositioned periosteal flap and skin (5.0 nylon sutures; Ethicon[®]; Somerville, NJ, USA). The operative wounds were left uncovered and the surgery region received no external immobilization.

Implant Stability Measurements

The surgeons carried out the RFA with the Osstell[®] ISQ and Penguin[®] RFA device. MultiPegs[®] magnetic transducers were mounted on each implant and tightened with hand pressure using the metallic screwdriver recommended by the manufacturer. The probe of the Penguin[®] RFA was held 1 mm from the MultiPeg[®] and the implant stability (ISQ) was registered on the digital display of the instrument for each implant. Three consecutive measurements from the lateral direction were recorded. The authors considered the mean of the three ISQ values to be the final ISQ value of each implant.

Also, the RFA measurements were collected with the Osstell[®] ISQ device using the SmartPeg[®] system and the plastic screwdriver supplied by the manufacturer. The authors considered the mean of the three ISQ values as the final ISQ of each implant.

Postoperative Care

All animals received antibiotic therapy by intramuscular injection of 0.1 mL/kg of oxytetracycline (Terramicina[®]; Pfizer, New York, NY, USA) every 48 hours for three days. Additionally, 4 mg/kg of Tramal[®] (Pfizer, New York, NY, USA) and 0.5 mg/kg of the anti-inflammatory meloxicam (Meloxivet[®]; Duprat, Rio de Janeiro, RJ, Brazil) were given over five days. Oxytetracycline spray with hydrocortisone was used daily at the wound site (Terra-Cortril[®] Spray; Zoetis, São Paulo, SP, Brazil). Zinc oxide ointment with cresylic acid was applied (Unguento Chemitec[®]; Chemitec, São Paulo, SP, Brazil) together with silver spray (Aerocid Total[®]; Agener União, Araçoiaba da Serra, SP, Brazil) to support healing and the prevention of insects.

Histological Preparation

After two and four weeks, the animals were re-operated on to remove the implants with a 5-mm internal diameter trephine drill (S.I.N. Implant System, São Paulo, SP, Brazil). The surgical procedures followed the protocol reported above and all sheep were subsequently returned to the farm, where they completely recovered. The samples containing bone and implants were fixed in 4% neutral-buffered formalin solution for 48 hours and then dehydrated in ascending alcohol solutions of 60%, 70%, 90%, and 100%. Thereafter, infiltration with a light-curing resin (Technovit 7200; Kulzer & Co., Wehrheim, Germany) was performed according to the manufacturer's instructions. The samples were then embedded in the same resin, cut in the mid-axial and apical-coronal planes using a macro-scale cutting and grinding technique (Exakt 310 CP series; Exakt Apparatebau, Norderstedt, Germany), and ground and polished to a final thickness of 30 to 40 μ m. Toluidine blue stained the samples to differentiate newly formed bone, and acid fuchsin were used to contrast the background. Light microscopy at 10 \times and 20 \times magnifications (Olympus BX43; Olympus Corporation, Tokyo, Japan) supported the analysis of the slices, with images acquired with the cellSens software (Olympus Corporation, Tokyo, Japan), and polarization light microscopy (Axioplan; Carl Zeiss AG, Oberkochen, Germany) enabling the visualization of the general orientation of collagen fibers.

Histomorphometry Analysis

From each slide obtained by histological processing, photomicrographs with 10 \times magnification were captured in sequenced fields to scan and reconstruct the total area of

the implant and adjacent bone (Figure 2A). Following the reconstruction of all images, the area of interest was determined and drawn, in the vertical direction, from the first thread of the implant to the beginning of the fourth thread (Figure 2B). This vertical delimitation was used to determine the BIC value, which was later transformed into a percentage. The implant-profile design was then duplicated and aligned at a distance of 270 μ m in the horizontal plane, thus completing the total area of interest (Figure 2C). The bone area fraction occupancy was determined manually with Image J (National Institutes of Health, Bethesda, MD, USA), which was later transformed into a percentage (Figure 2D).

Statistical Analysis

The data for RFA using Osstell[®] and Penguin[®] device and insertion torque values (IVT) did not pass the Shapiro–Wilk normality test; the log (ISQ) and log (IVT) for HANano[®], SLActive[®], and TiUnite[®] was evaluated using the one-way ANOVA test and Tukey's posttest ($p < 0.05$). The Student's *t*-test compared the data obtained by Osstell[®]/SmartPeg[®] and Penguin[®]/MultiPeg[®] for each implant surface, considering $\alpha = 0.05$.

For the evaluation of BIC and BAFO of the various surfaces in the same experimental period, the One-Way ANOVA parametric test and Tukey's posttest ($p < 0.05$) were applied. The Student's *t*-test ($p < 0.05$) evaluated the significance of BIC and BAFO of the same surface and at different experimental periods. The values are presented as mean \pm confidence interval ($n = 5$ animals per group/experimental period). The Prism Graph Pad 8.3[®] software (Inc. La Jolla, California, USA) enabled all the statistical analysis.

Results

Surface Characterization

Scanning electron microscopy micrographs of the implant's surface revealed the samples' textured microstructures and topography (Figure 3). The observed surface texture at intermediate (3000 \times) and high (15,000 \times) magnifications appeared considerably comparable in terms of the surface morphology between the implant groups; however, group TiUnite[®] exhibited a different remarkably surface morphology. Figure 3A and B present an example of a typical nano-HA surface with a well-defined micro-scale rough surface. The sandblasting and etched process's micro-scale rough surface texture can be seen in Figure 3C and D (SLActive[®]). The implants treated with the anodic plasma-chemical surface-modification method is shown in Figure 3E and F; the surface layer is composed of

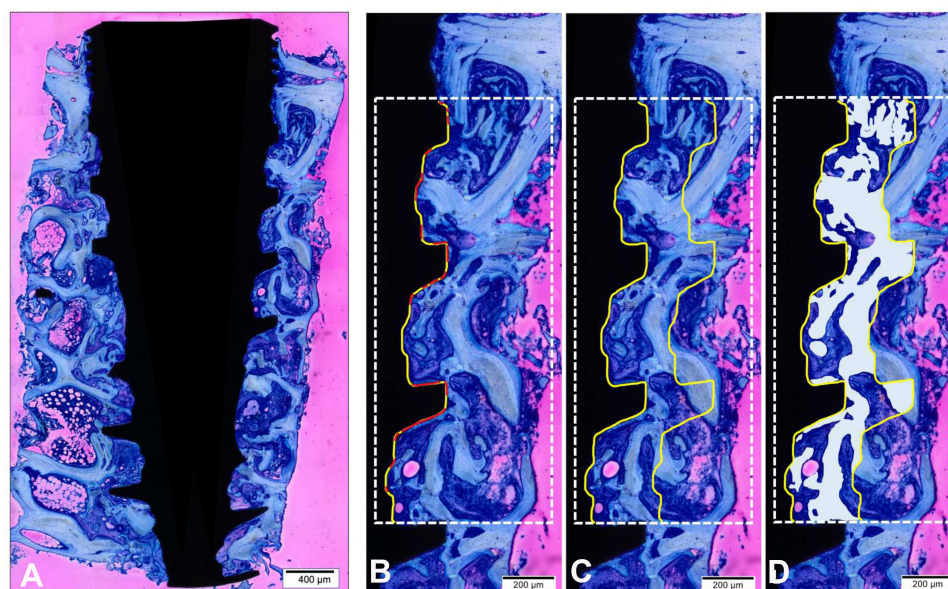


Figure 2 Histomorphometry for bone-implant contact and (BIC) and bone area formation (BAFO) analysis. (A) Histological reconstruction of the implant and adjacent bone area; (B) determination of the line of interest for BIC; in the long implant axis, the implant profile design was drawn from the first thread of the implant to the beginning of the fourth thread (yellow line) and direct bone-implant contact (red line); (C) determination of the area of interest for BAFO. An identical line of implant profile design was duplicate and aligned at a distance of 270 µm at the horizontal plane (total area); (D) the bone area formation (BAFO) manually determinates for posterior analysis (total area/BAFO) (%). Stain: Toluidine Blue and Acid Fuchsin stained. Scale bar: (A) 400µm; (B,C and D) 200µm.

small craters with holes in the center (TiUnite®). Energy-dispersive spectroscopy revealed the presence of calcium only in the nano-HA group and that of sodium only in the SLActive® group. Titanium, vanadium, and phosphorus peaks were observed in all groups ([Supplementary Figure 1](#)).

Resonance Frequency Measurements

The resonance frequency analysis occurred with the following devices and transducers Osstell®/SmartPeg®, and Penguin®/MultiPeg®. [Figure 4](#) shows the log values of ISQ obtained from the resonance frequency analysis of HAnano®, SLActive®, and TiUnite®. The Penguin®/MultiPeg® analysis showed significantly higher resonance frequency values than the Osstell/Smart Peg on all implants ($p < 0.05$). However, no significant ISQ difference occurred between the implants under the same device, Osstell®/SmartPeg® or Penguin®/MultiPeg® ($p < 0.05$) [Supplementary Table 1](#).

Implant Torque Value (ITV)

There was no statistical difference among the tested implants surfaces (HAnano®, SLActive®, and TiUnite®) regarding the insertion torque, and all groups presented a mean insertion torque between 72 and 74 N/cm ([Figure 5](#)) ([Supplementary Table 1](#)).

In vivo Study

Anesthesia, surgical intervention, and implant placement were completed uneventfully. All animals recovered quickly and could walk immediately after surgery. All animals gained weight after surgery except for one sheep that lost almost 10% of its initial weight 28 days after surgery. Still, no changes were observed in the histological results, so the weight loss probably did not impact the biological effects. No case of superficial nor deep infection happened. All animals were furnished daily with regular feed and water intake until the end of the study. All implants were able to be placed according to the intended technique and distribution. No implant exhibited clinical mobility, bone loss, or infection.

Histological Evaluation

Descriptive qualitative microscopic evaluation of the nondecalcified samples demonstrated time-dependent new bone formation in all groups, visible by way of blue staining, adjacent to the implant surface in all samples. After two weeks, all evaluated surfaces presented peri-implant bone healing with different bone and BIC volumes. The Epikut Plus® with HAnano® coating group showed large trabeculae of neoformed bone between the implant threads and islands of bone debris in contact with the trabecular bone. Several areas of

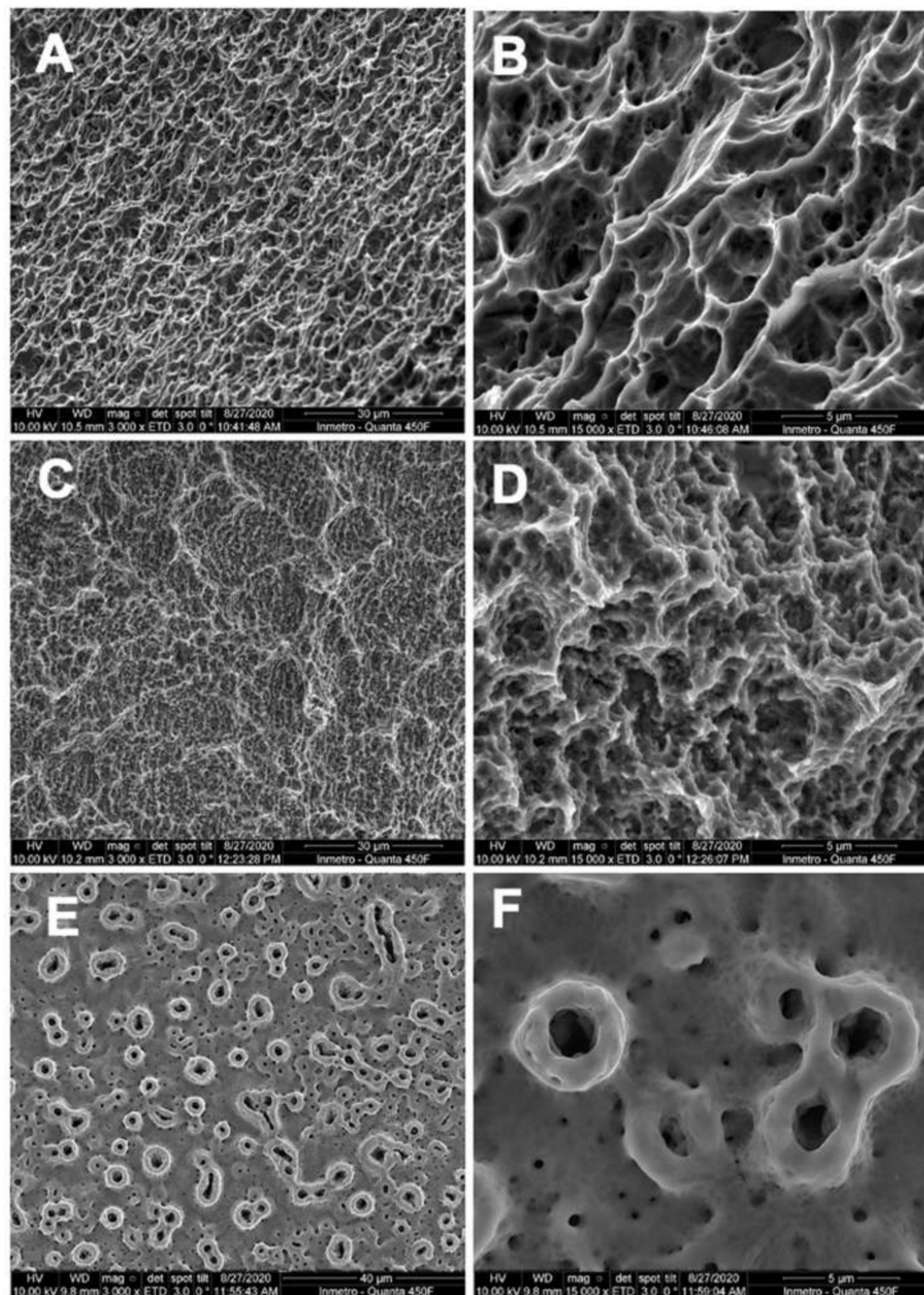


Figure 3 Scanning electron microscopy (SEM) micrographs of implants surface. (A and B) Hanano; (C and D) SLActive, and (E and F) TiUnite. (A, C and E) at 3000× magnification (scale bar = 30 μm) and (B, D and F) 15,000 × magnification (scale bar = 5 μm).

direct BIC were observed. The BLX[®] with SLActive[®] surface presented a similar degree of bone volume, less extensive trabeculae, bone debris between the threads, and evident BIC areas. The NobelActive[®] with the TiUnite surface showed bone trabeculae permeating the implant surface at a lower volume than either the nano-HA or SLActive surfaces but with a similar BIC area (Figure 6).

After four weeks, newly formed bone around the implants was clearly apparent. Moreover, a minimum amount of connective tissue was observed in all groups (Figure 7). Using polarization light microscopy, the general orientation of fibers could easily be seen when using a compensator crystal ($\lambda = 551$ nm). In Figure 8, the blue coloring indicates that fibers are parallel to the higher refraction index of the compensator.

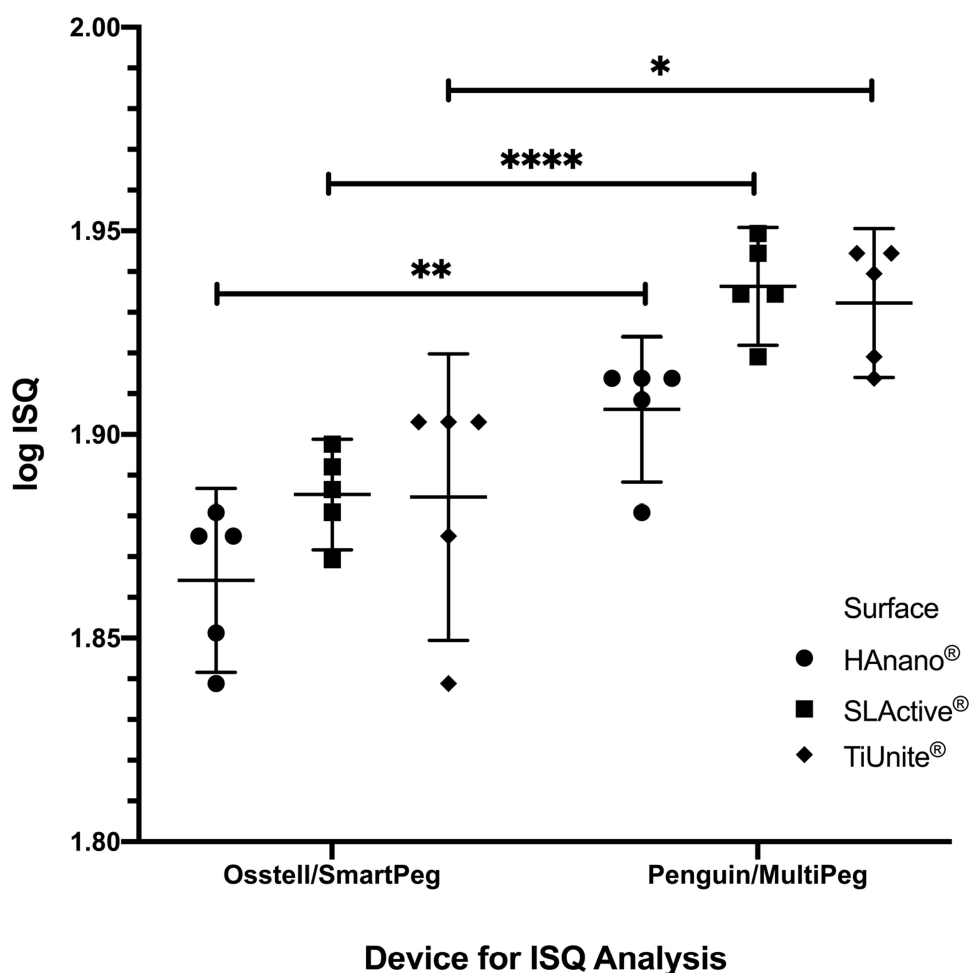


Figure 4 Implant stability quotient (ISQ) values of different implant surfaces. The devices and transducers grouping used were Osstell/Smart Peg (measured as implant stability quotient, ISQ) and Penguin/Multi Peg (measured as resonance frequency analysis, RFA). The graphic shows the distribution of all point values for different surfaces (HAnano, SLActive and TiUnite) with different devices and transducers grouping (n=5). After the normality test (Shapiro–Wilk), the values were transformed in LOG (Y). The groups were submitted to statistical analysis of Two-way ANOVA and Tukey post-test to evaluate the differences between different surfaces with the same device/transducer ($p < 0.05$). The Student's t-test was applied to assess the differences between the device/transducer at the same implant surface ($p < 0.05$); * ($p < 0.05$), ** ($p < 0.01$), and **** ($p < 0.0001$).

BIC

BIC significantly increased in a time-dependent fashion in all groups tested ($p < 0.05$) being higher than 80%. However, at each experimental time point, no statistical difference in BIC ($p > 0.05$) was observed between the experimental groups (Figure 9) (Supplementary Table 2)

Bone Area Fraction Occupancy

Histomorphometric analysis revealed clearly that all groups experienced a subsequent percental gain in new bone formation (Figure 10). There were no significant differences between the percentage of new bone formation at the implant interface between groups after two or four weeks ($p > 0.05$). Over time, the percentage of new bone formation increased at the interface and the surrounding tissue occupying around 60% of the analyzed area (Supplementary Table 2).

Discussion

With the development of nanotechnology, nanostructured HA surfaces are being recognized as promising to achieve rapid osseointegration in low-density bone, which is the main persisting challenge in implant dentistry. Thus, it is believed that the use of HA coatings on metallic implants would improve both the bone strength and speed of rehabilitation of patients by decreasing the time from implant insertion to receipt of the final prosthesis mainly in the posterior area of the maxilla where there the highest rate of failure exists due to progressive bone loss.⁹ Surface topography from the micro- to the nanoscale profoundly influences the bone cells attachment and osseointegration^{37–39} and the rough implant surfaces achieved by HA coatings offer osteoconductive properties⁴⁰ due to increased cell adhesion.

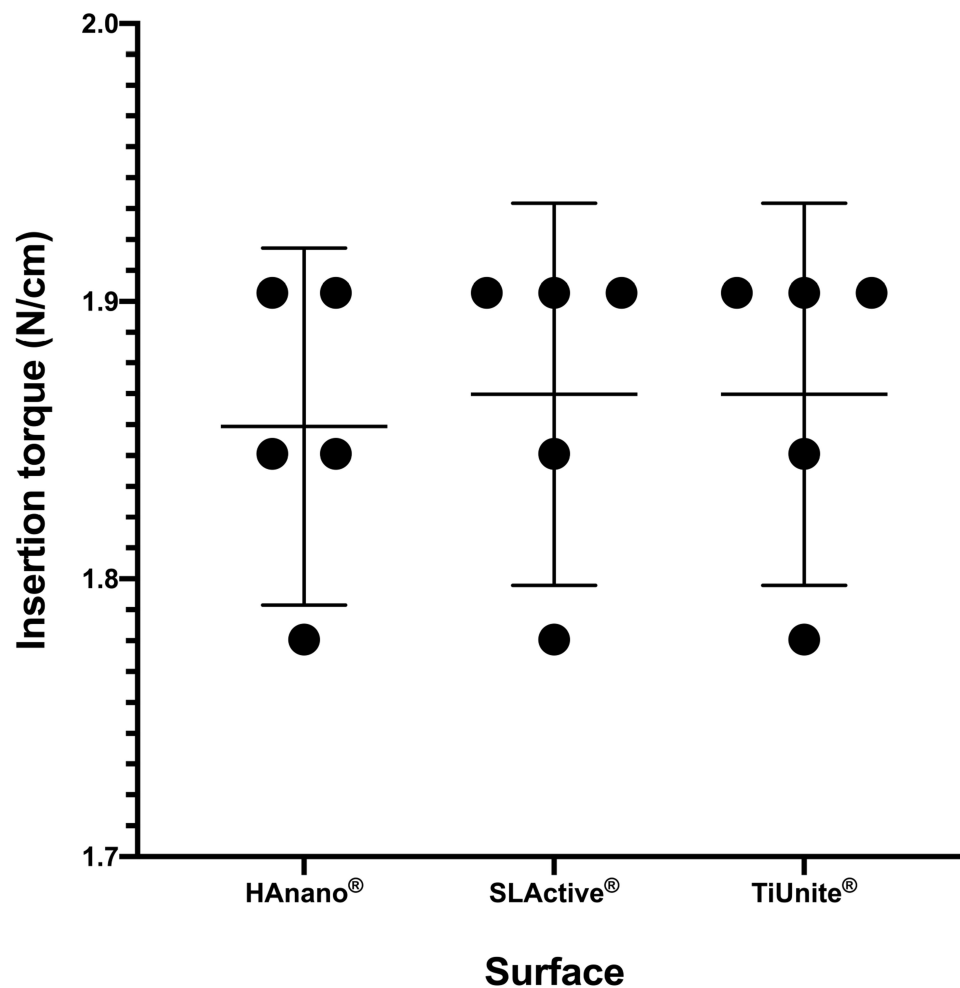


Figure 5 Insertion torque values (N/cm) of different implant surfaces (HAnano®, SLActive® and TiUnite®). The graphic shows the mean values and the confidence interval at 95% of different groups (n=5), the values were transformed in LOG (Y). The groups were submitted to statistical analysis of one-way ANOVA and Tukey post-test to evaluate the differences between surfaces ($p < 0.05$). There were no differences in the insertion torque between implant surfaces.

Surface interaction and changes at the implant–cell interface result in diverse cellular responses. Understanding intracellular signaling involved in the mechanisms of osteoblast adhesion, proliferation, and differentiation on dental implants is fundamental for the successful osseointegration. The first step in this response to implants involves the adsorption of specific proteins, lipids, sugar, and ions, establishing an organic coating responsible for guiding surrounding cell performance leading to activation of specific genes.⁴¹ A previous in vitro study has shown that HAnano-blasted titanium surface improved wettability and transformed the implant surface to super-hydrophilic ($<4^\circ$). This increased wettability facilitates the attachment process of bone cells to the implant surface, thereby improving the implant's integration.⁴² HAnano surface promoted increased cell proliferation, viability and cell spreading as well as type I collagen and osteopontin secretion favoring the early events of osseointegration.⁴³

In this preclinical study, we compared the osseointegration performance of a new nanostructured HA coating (Epikut Plus®, S.I.N. Implant System, Sao Paulo, Brazil) to a recently launched implant with hydrophilic SLActive® surface (BLX®, Straumann, Basel, Switzerland), and the well-known macrogeometry with highly crystalline and enriched TiUnite® surface (NobelActive®, Nobel Biocare, Göteborg, Sweden).

In general, all histological and biomechanical evaluations suggested no significant difference between the three surface modifications with respect to successful osseointegration. Given the plan to conduct this in vivo study on low-density bone, sheep ilium was chosen as the model for implantation.

The sheep model is considered to be suitable for biomedical research due to the similarities between sheep and humans regarding weight, joint structure, bone tissue, and bone

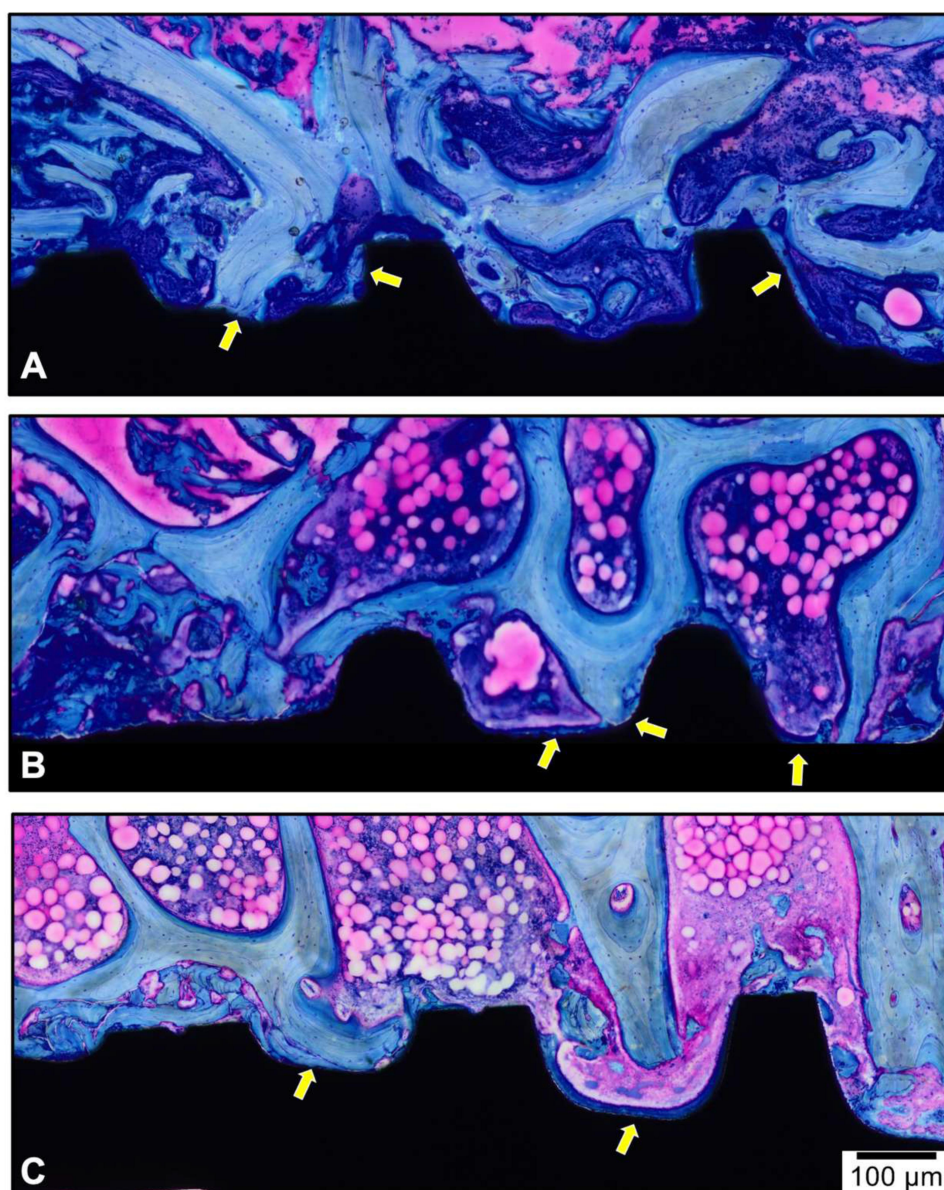


Figure 6 Representative photomicrographs of wound healing to different implant surfaces 14 days after implantation. (A) HANano®, (B) SLActive®, and (C) TiUnite®. Observe the three most coronally situated implant's threads that were located in the cancellous bone. The yellow arrows represent regions of bone-implant contact. Stain: Toluidine Blue and Acid Fuchsin stained. Magnification: 20×; Scale bar: 100µm.

regeneration.^{28,44} It also presents unique advantages such as bone composition, metabolism, remodeling, and regeneration time as compared with other experimental animal models.⁴³ Previous studies of dental-implant designs and surface have used sheep,^{30,33–36,45} rabbits,^{4,20,46} and dogs^{47–49} as animal models. The most used sites in these animals are the jaw^{47,48} or tibia⁴⁹ in dogs; the tibia^{4,20,46,50} in rabbits; and the tibia,^{34,35} iliac crest,^{30,33,36,45} and mandible⁵¹ in sheep. The iliac crest site offers the advantages of bone characteristics, the absence of postoperative morbidity, and the ability to conduct a high number of implants test simultaneously; however, although the iliac crest exhibits low bone density, the cortical is thicker

relative to the thin cortical of the posterior region of the maxilla, which also shows low bone density. Previous studies^{30,33,36,45} that used sheep models have reported that the animals were euthanized at the end of the experiments; in our study, all animals were kept alive and healthy.

The insertion torque value is a direct measure of the bone's cutting resistance during implant insertion surgery. It is a mechanical parameter that can be influenced by the surgical procedure and implant design and is reliant on the bone quality. Here, all tested implants displayed an insertion torque value of between 60 and 80 N/cm (min and max); notably, these high values are probably due to the resistance

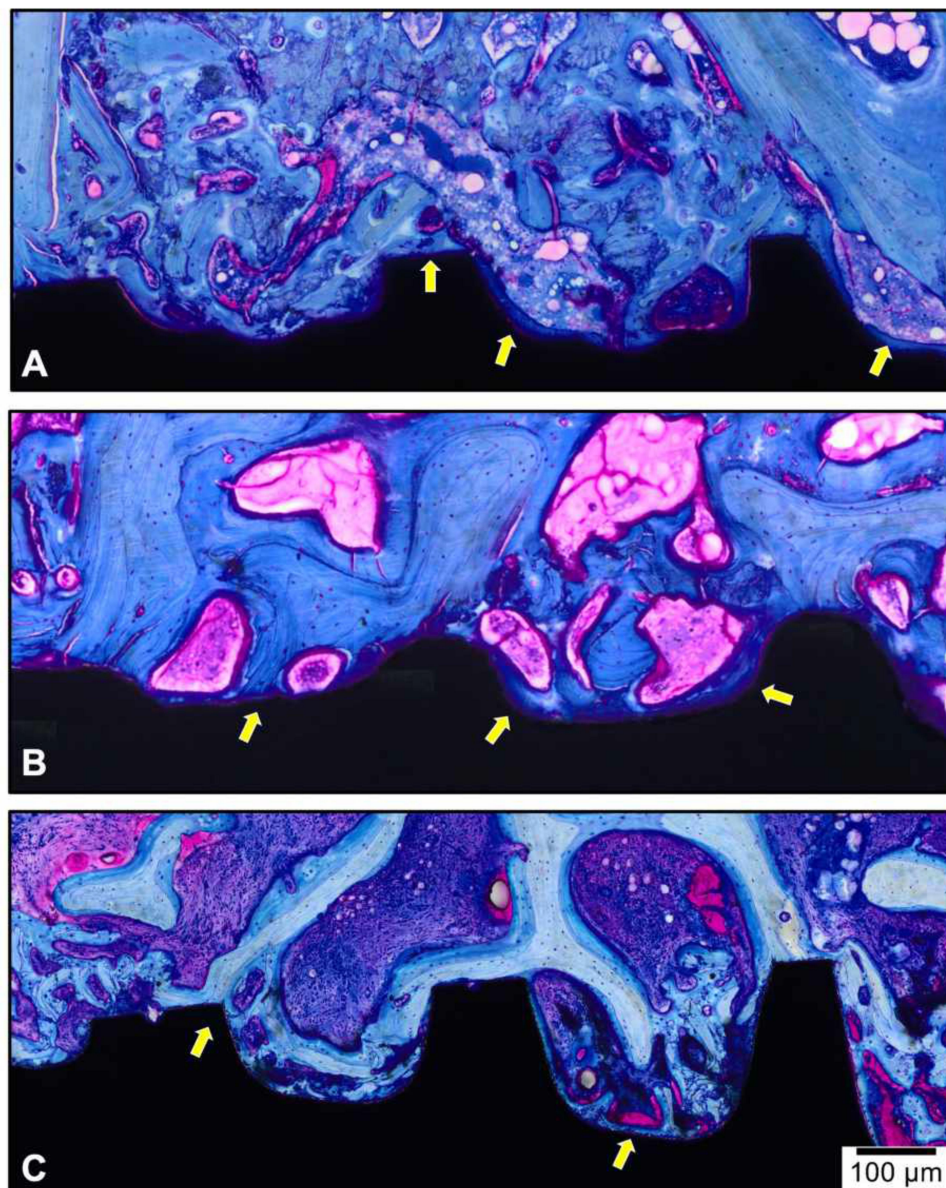


Figure 7 Representative photomicrographs of wound healing to different implant surfaces 28 days after implantation. (A) HANano®, (B) SLActive® and (C) TiUnite®. Observe the three most coronally situated implant's threads that were located in the cancellous bone. The yellow arrows represent regions of bone-implant contact. Stain: Toluidine Blue and Acid Fuchsin. Magnification: 20×; Scale bar: 100μm.

of the thicker cortical bone of the iliac crest region of the animal. In another study using sheep, a low value of insertion torque was observed in all groups (<21 N/cm), but the experimental implants used were characterized by a reduced thread profile that may have contributed to the low insertion torque values recorded.⁴⁴ Also, in this study, we confirmed the lack of effect of the implant surface on the insertion torque according to previous study.⁵²

Primary stability is defined as the absence of movement following intraosseous insertion of the implant. RFA is one of the most commonly used methods to

quantitatively evaluate the primary stability of implants. This analysis provides information regarding the stiffness of the bone-implant union and the results are recorded as the implant stability quotient. The present study is in agreement with findings of previous randomized clinical trials reporting acceptable primary stability with mean ISQ values of above 75 N/cm in humans.⁵³ The bone density, implant size and macro-scale geometry of the implant body greatly affect the ISQ values and, as we used the same implant size and receptor area, similar levels of ISQ between the evaluated groups were confirmed.⁵⁴ A clinical

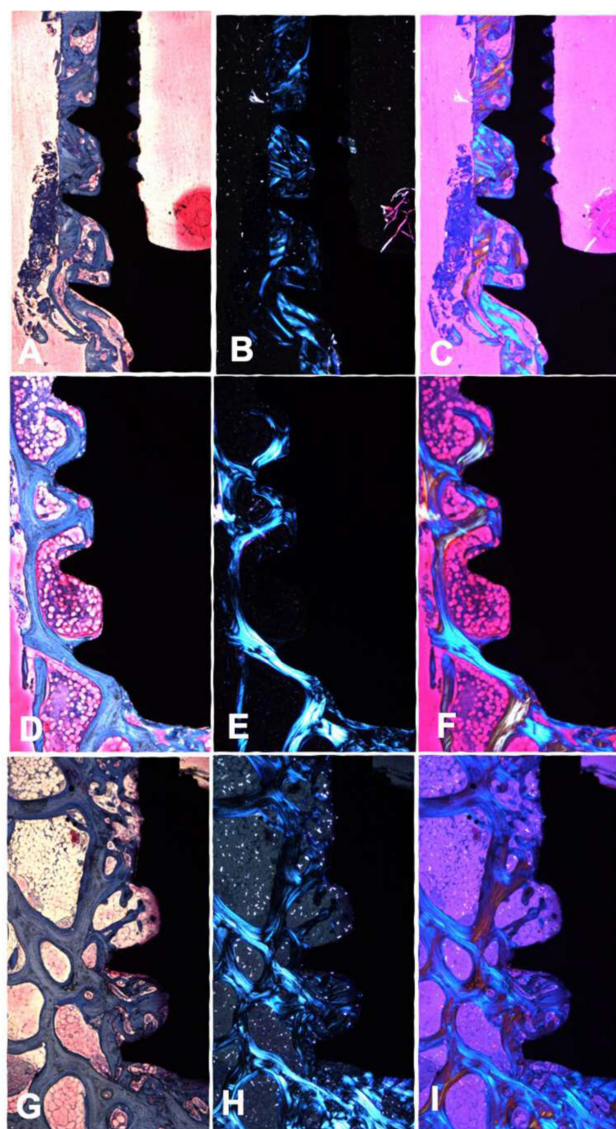


Figure 8 Polarization light microscopy. (A–C) HAnano® group; (D–F) SLActive® group, and (G–I) TiUnite® group. (A, D and G) bright filled microscopy; (B, E and H) Polarization light microscopy; (C, F and I) polarization light microscopy with compensator crystal ($\lambda = 551$ nm). The general orientation of bone collagen fibers is easily seen when using polarization light microscopy. Blue in the figure means that fibers are parallel to the higher refraction index of the compensator.

study using tapered implants with a wide thread in the posterior region of maxilla reported a mean ISQ value of $53.66 (\pm 12.04)$,⁵⁵ which is very close to our results.

The present study evaluated the resonance frequencies immediately after the implant's installation using two different devices (Osstell® and Penguin®) and two magnetic transducers (SmartPeg® and MulTipeg®), respectively. We observed statistical differences between the two devices, with the highest values recorded when using the MulTipeg® transducer in combination with the Penguin® device. These results were different from those in the

study by Bural et al⁵⁷ who evaluated the primary stability using the same devices and transducers but observed that the results were discretely higher in the Osstell® group; however, different from in our study, these authors performed the test in an ex vivo model. In our study, the resonance frequency was only evaluated on the day of implant installation; we did not evaluate after the experimental periods to limit any kind of damage caused by the insertion torque of the transducers into the implant and avoid impairing the histological and histomorphometric evaluations of the implant–bone interface.

Elsewhere, a recent systematic review by Lages et al⁵⁸ found no correlation existed between insertion torque and primary stability, suggesting that a high insertion torque does not necessarily correspond with a high ISQ value. In our study, we recorded high levels of insertion torque and primary stability.

Our results of BIC showed no statistical difference among the three groups in the same experimental period. All groups experienced a constant increase in trabecular bone fixation to the implant surface from two to four weeks, which corroborate data from a previous study which used the SLActive® and TiUnite® surfaces in the same animal/site model.³⁰

The bone area fraction occupancy increased from 2 to 4 weeks, but without significant difference between the HAnano®, SLActive® to TiUnite®. Our results corroborate with those of previous studies assessing HA-coated implants in rabbit tibia after two weeks,⁵⁹ in dogs' sockets after four weeks,⁶⁰ and in the iliac crests of goats after four weeks.⁶¹ In all these studies, the HA biomimetic coating was able to shorten the healing period of the implants by increasing the implant–bone interaction. Our BIC and BAFO values for HA coating results were similar to those of Van Oirschot et al⁶¹ who obtained values of $57.5\% (\pm 8.5\%)$ for BIC and $43.6\% (\pm 9\%)$ for BAFO in a study in iliac crests of goats after four weeks. Lee et al presented similar results for BAFO and BIC of $44.94\% (\pm 17.69\%)$ and $77.28\% (\pm 11.22\%)$ after four weeks in dogs as compared with our study, where the values were $65.53\% (\pm 6.22\%)$ and $82.27\% (\pm 3.38\%)$.

Conclusion

The three implant designs and improved surfaces proved comparable osseointegration at the early stages of low-density bone repair in the sheep model.

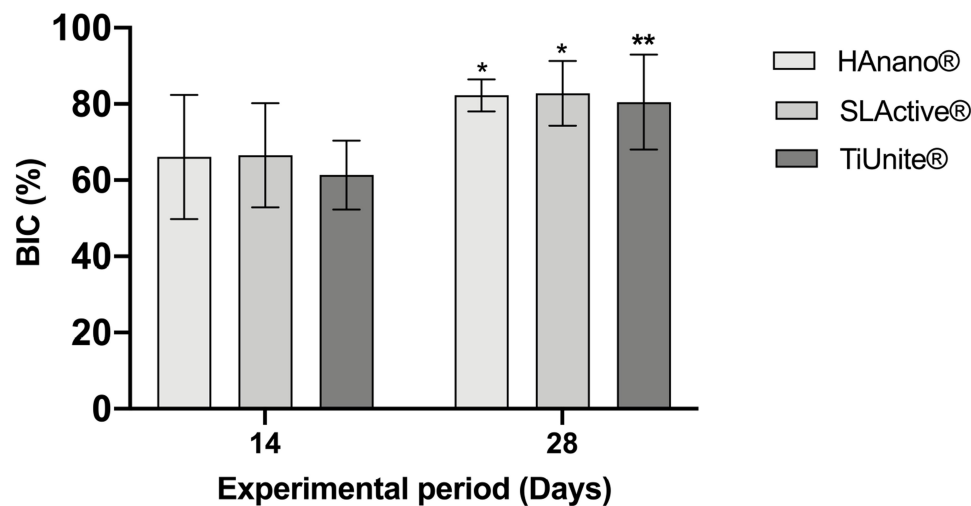


Figure 9 Bone-implant contact (BIC) percentage in the HAnano®, SLActive®, and TiUnite® surfaces groups, 14 and 28 days after implantation. The results are presented as a mean \pm confidence interval (n=5). After the normality test (Shapiro–Wilk), the groups were submitted to statistical analysis of One-way ANOVA and Tukey post-test ($p < 0.05$) to evaluate the differences between different surfaces at the same experimental time point. The Student's t-test was applied to assess the differences between the same surface groups at different time points ($p < 0.05$), represents by the (*, $p = 0.02$), (**, $p = 0.008$). There were no differences in the BIC between implant surfaces at the same time point.

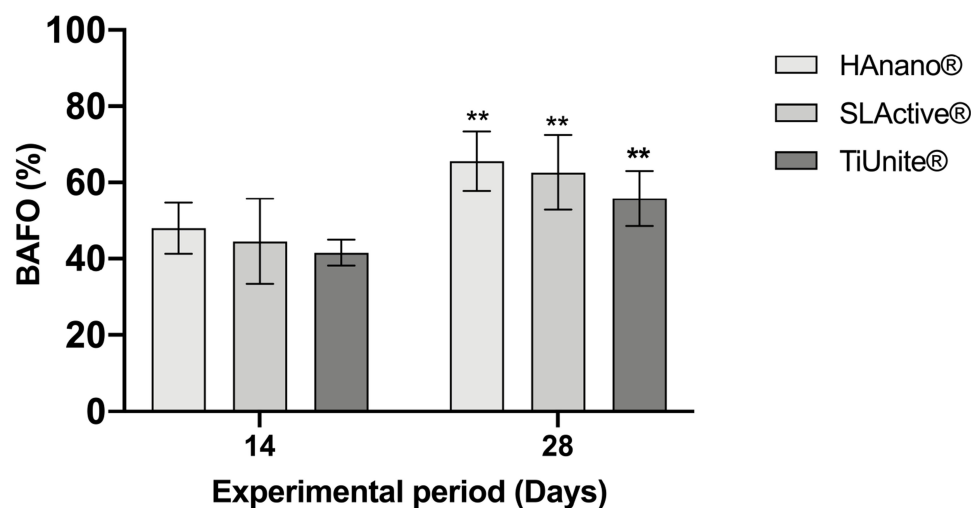


Figure 10 Bone area fraction occupancy (BAFO) percentage in the HAnano®, SLActive®, and TiUnite® surfaces groups, 14 and 28 days after implantation. The results are presented as a mean \pm confidence interval (n=5). After the normality test (Shapiro–Wilk), the groups were submitted to statistical analysis of One-way ANOVA and Tukey post-test ($p < 0.05$) to evaluate the differences between different surfaces at the same experimental time point; represents by horizontal bars. The Student's t-test was applied to assess the differences between the same surface groups at different experimental time points ($p < 0.05$), represents by the (**, p ranging from 0.001 to 0.009). There were no differences in the BAFO between implant surfaces at the same time point.

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Disclosure

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The authors report no other potential conflicts of interest for this work.

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