

#### ORIGINAL RESEARCH

# Lactosylated IR820/DOX Co-Assembled Nanodrug for Synergetic Antitumour Therapy

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Introduction: Synergistic treatment integrating photothermal therapy (PTT) and chemotherapy is a promising strategy for hepatocellular carcinoma (HCC). However, the most commonly used photothermal agent, IR820, and chemotherapeutic drug, doxorubicin hydrochloride (DOX), are both hydrophilic molecules that suffer from the drawbacks of a short circulation time, rapid elimination and off-target effects.

Methods and Results: Herein, a novel nanodrug that combined HCC-targeted IR820 and DOX was developed based on excipient-free co-assembly. First, lactosylated IR820 (LA-IR820) was designed to target HCC. Then, the LA-IR820/DOX nanodrug (LA-IR820/DOX ND) was purely self-assembled without excipient assistance. The physicochemical properties and the chemo-photothermal antitumour activity of the excipient-free LA-IR820/DOX ND were evaluated. More importantly, the obtained LA-IR820/DOX ND exhibited 100% drug loading, remarkable HCC targeting and excellent antitumour efficacy.

Conclusion: This excipient-free LA-IR820/DOX ND may be a promising candidate for the synchronous delivery and synergistic targeting of IR820 and DOX as a combined chemophotothermal therapy.

**Keywords:** excipient-free, hepatoma cell targeting, self-assembly nanodrug, photothermal therapy, chemotherapy

#### Introduction

Hepatocellular carcinoma (HCC) is one of the leading causes of cancer-related death throughout the world. 1,2 Its insidious onset, rapid progress and severe liver dysfunction lead to high mortality.<sup>3</sup> To date, commonly used anthracyclines have been unsatisfactory in the treatment of advanced HCC patients, whose survival time is only three to four months in most Asian countries.<sup>4,5</sup> Therefore, there is an urgent need to explore more efficient and safer treatments for HCC. Photothermal therapy (PTT) employs optical absorbing agents to induce local hyperthermia to ablate tumours by near-infrared (NIR) light irradiation.<sup>6</sup> It has attracted significant attention due to its non-invasiveness, high spatiotemporal selectivity and limited therapeutic resistance. Remarkably, in addition to causing direct tumour cell death, PTT can also decrease interstitial fluid pressure and increase blood perfusion.<sup>8,9</sup> Moreover, the destruction of cancer cells by hyperthermia causes immunogenic cell death (ICD), which can release tumour-specific antigens to initiate the antitumour immune response. 10,11 However, PTT alone is insufficient to eradicate tumours due to the limited penetration depth of NIR light. The surviving tumour cells from PTT will continue to proliferate, leading to cancer recurrence and metastasis. 12,13

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From this viewpoint, it is essential to combine other therapeutic strategies with PTT in a coordinated way. The combination of PTT and chemotherapy has been shown to be an encouraging approach resulting in superior synergistic effects and lower side effects. 14-17 The introduction of chemotherapy provides an opportunity to kill deep tumour cells, and some chemotherapeutic agents, such as DOX, can also induce ICD, consolidating the antitumour immunity invoked by PTT. Some challenges, however, still need to be overcome to achieve the optimal therapeutic effect. In many reported systems, IR820 and DOX are commonly used small hydrophilic molecules. 18,19 However, their theranostic effect is limited due to shortcomings such as instability, short lifetime, rapid elimination and non-specific tumour-homing ability.<sup>20</sup> <sup>22</sup> It is therefore, highly desirable to exploit an IR820 and DOX co-assembled nanosystem that provides increased stability, tumour targeting and prolonged circulation time.

In recent years, tremendous efforts have been made to develop nanodrug delivery systems, which are anticipated to optimize cancer therapy.<sup>23–25</sup> Among them, selfassembly has attracted considerable interest due to its facile fabrication and flexibility in the organization of ordered nanostructures. Nevertheless, excipient-assistant nanodrugs suffer from low drug-loading capacity (< 20%), native toxicity of excipients and difficulty in scaling up production, all of which hinder their commercialization.<sup>26</sup> To address these concerns, the innovative drug delivery system of excipient-free nanodrugs self-assembled by active small molecules without inert excipients have been developed and attracted extensive attention. 27-29 Excipient-free nanodrugs can simultaneously achieve high drug-loading efficiency, display good pharmacokinetics and avert the safety problems caused by excipients. 30,31 Shim et al developed a excipient-free DOX prodrug, in which the cathepsin B-cleavable peptide was conjugated to DOX for tumour targeting, tumour-specific DOX release and reduction of systemic toxicity.<sup>26</sup> However, the types of tumourresponsive linkers are limited and the conjugation often needs tedious synthesis. Fan et al reported a excipientfree nanodrug formed by pure ursolic acid to treat the nonsmall cell lung cancer.<sup>32</sup> Feng et al reported an indocyanine green (ICG)-templated self-assembly of paclitaxel (PTX), achieving 90.7% PTX and 9.2% ICG loading.<sup>28</sup> This co-assembly nanosystems may offer new inspiration for synergistic therapy. However, the pure and co-assembly nanodrug both suffer from certain systemic toxicity due to the lack of active targeting. Therefore, it is essential to develop an active targeting excipient-free self-assembly for synergistic therapy. Because the asialoglycoprotein receptor (ASGPR) is exclusively expressed in hepatoma cells and can recognize and internalize glycoproteins bearing terminal lactose moieties, we previously developed a lactosylated IR820 (LA-IR820) that enabled HCC targeting.<sup>33</sup> Here, we developed an excipient-free nanodrug composed of only the HCC-targeted IR820 and chemotherapeutic DOX via pure self-assembly. The physicochemical properties and the chemophotothermal antitumour activity of the excipient-free LA-IR820/DOX nanodrug (LA-IR820/DOX ND) were evaluated. The results revealed that LA-IR820/DOX ND displayed 100% drug loading, remarkable HCC targeting and excellent antitumour efficacy. Thus, this excipientfree LA-IR820/DOX ND may be a promising candidate for targeted chemo-photothermal therapy against hepatocellular carcinoma.

## **Experimental Section**

#### **Materials**

IR820 was purchased from Beijing J & K Technology Co., Ltd. (China). Lactobionic acid was supplied by Sigma-Aldrich (USA). Doxorubicin hydrochloride (DOX) was obtained from Dalian Meilun Biological Technology Co., Ltd. (China). Trypan blue was received from Beijing Solarbio Technology Co., Ltd. (China). Methylene dichloride (>99.5, CH<sub>2</sub>Cl<sub>2</sub>) and triethylamine (Et<sub>3</sub>N, >99.0%, (C<sub>2</sub>H<sub>5</sub>)<sub>3</sub>N) were provided by Tianjin Fuyu Fine Chemical Co., Ltd. (China).

# Preparation of LA-IR820/DOX ND

LA-IR820 was synthesized as previously described.<sup>33</sup> The self-assembly process of LA-IR820 and DOX is illustrated in Figure 1A. Briefly, LA-IR820 (1 mM, 1.0 mL) and DOX (2 mM, 1.0 mL) were individually dissolved in distilled water, and the DOX solution was added dropwise to LA-IR820 solution with 200 rpm stirring, and yielding spherical LA-IR820/DOX ND.

# Morphology of LA-IR820/DOX ND

Transmission electron microscopy (TEM, JEM-200CX II, Electronics Corporation, Japan) and dynamic light scattering (DLS, Zetasizer Nano ZS, Malvern Panalytical, England) were used to characterize the morphology and particle size of LA-IR820/DOX ND. The UV-visible absorption spectra of IR820 and LA-IR820 were recorded

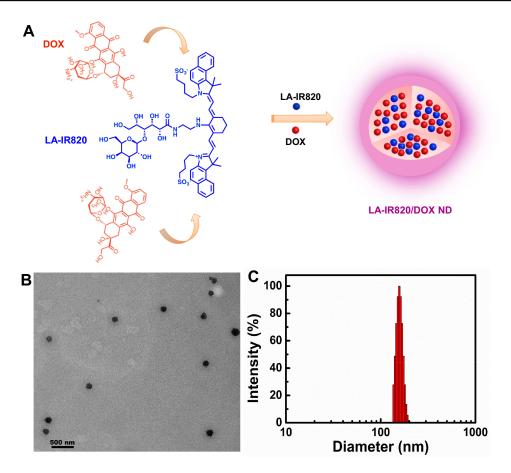


Figure 1 (A) Schematic representation of the self-assembly process of LA-IR820/DOX ND with the molar ratio of 1:2 (LA-IR820:DOX); (B) TEM image of LA-IR820/DOX ND, and the scale bar is 500 nm; (C) size distribution of LA-IR820/DOX ND.

by UV-visible spectrophotometer (TU-1810, General Analysis Instrument Co., Ltd., China). All samples were tested at room temperature.

# In vitro Drug Release Studies

The DOX release from LA-IR820/DOX ND was studied using the dialysis method. One millilitre of LA-IR820/DOX ND ( $c=150~\mu g/mL$ ) was put into the dialysis bag (MWCO  $\sim 3500~Da$ ) and incubated at  $37.0\pm0.5~^{\circ}C$  in 20 mL PBS (pH 7.4) with 100 rpm stirring. At specific time points, 1.0 mL of the release medium was taken out and replaced with the same volume of fresh medium. The content of DOX was analysed using a fluorescence spectrophotometer at an excitation wavelength of 496 nm and an emission wavelength of 558 nm. The accumulated DOX release was calculated by the following equation:

Cumulative release amount(%) =  $M_t/M_{\text{total}} \times 100\%$ 

where  $M_t$  is the amount of DOX released into the medium at time point t, and  $M_{total}$  is the total amount of DOX in the dialysis bag.

#### In vitro Photothermal Performance

The photothermal effect of IR820 and LA-IR820/DOX ND was conducted using distilled water as a control, and the concentrations of the photothermal agents ranged from 10  $\mu$ M to 100  $\mu$ M. One millilitre of IR820 and LA-IR820/DOX ND were added to a 12-well plate and irradiated at 808 nm (1.0 W·cm<sup>-2</sup>) and 660 nm (1.0 W·cm<sup>-2</sup>), respectively, for 5 min. During laser irradiation, the temperature was monitored every 30 seconds.

## Cell Culture

Human HepG2 hepatoma cells and murine Hepa1-6 hepatoma cells were purchased from American Type Culture Collection (ATCC). HepG2 cells were cultured in RPMI-1640 medium (Gibco, Thermo Scientific, USA) and Hepa1-6 cells were cultured in high glucose DMEM (Gibco, Thermo Scientific, USA) at 37 °C with 5% CO<sub>2</sub>. Both mediums contained 10% foetal bovine serum (Qingdao Haosail Technology Co., Ltd, China) and 1% penicillin-streptomycin antibiotic mixture (Beijing solarbio science and technology co., Ltd, China).

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# Intracellular Uptake of LA-IR820/DOX ND

HepG2 cells ( $2 \times 10^5$  cells/well) were incubated in a 6-well plate for 24 h. Then, the medium was removed and replaced with the medium containing free DOX, IR820 and DOX co-assembly nanodrug (IR820/DOX ND) and LA-IR820/DOX ND, respectively (DOX:  $10~\mu g/mL$ ). After 4 h incubation, the medium was discarded, and the cells were rinsed three times with pH 7.4 PBS. Then, the cells were analysed by fluorescence inverted microscopy (ECLIPSE-Ti, Nikon, Japan) and flow cytometry (BD FACSAria III, USA).

## Cytotoxicity Assay

Hepa1-6 and HepG2 cells were chosen to estimate the dark cytotoxicity of IR820 and LA-IR820 by the methyl tetrazolium (MTT) method. Hepa1-6 and HepG2 cells were seeded in 96-well plates at a density of  $5\times10^3$  cells for overnight incubation. Afterwards, the medium was replaced with fresh medium containing IR820 and LA-IR820 at concentrations ranging from 0.076  $\mu$ M to 38  $\mu$ M. After 24 h, 20  $\mu$ L MTT (5%) was added into each well for another 4 h incubation. The supernatants were then discarded and 150  $\mu$ L DMSO was added to each well to dissolve formazan crystals. The absorbance was measured by PerkinElmer ELISA at a test wavelength of 570 nm.

To evaluate the chemo-photothermal antitumour efficacy, cells were incubated with different concentrations of DOX, IR820, LA-IR820 and LA-IR820/DOX ND with equivalent DOX ( $c=0.1,\ 1,\ 10,\ 20,\$ and 50  $\ \mu g\cdot mL^{-1}$ ) for 4 h. Subsequently, the drug-containing medium was substituted by the fresh drug-free medium. The cells treated with IR820 were irradiated by an 808 nm laser while the LA-IR820 and LA-IR820/DOX ND-treated cells were irradiated by a 660 nm laser at a power of 1.0 W·cm<sup>-2</sup> for 2 min. After 24 h, the cytotoxicity was tested by MTT. The cell inhibition ratio was calculated by the following formula:

Cell inhibition ratio(%) = 
$$\left(1 - \frac{A_{\text{sample}}}{A_{\text{negative}}}\right) \times 100\%$$

where  $A_{\text{sample}}$  and  $A_{\text{negative}}$  are the absorbance values from each sample and negative control, respectively.

# Trypan Blue Staining

Trypan blue staining can directly determine whether cells are alive. HepG2 cells ( $5 \times 10^4$  cells) were seeded in 24-well plates and incubated overnight. Then, the medium

was substituted for fresh medium with free DOX, IR820 and LA-IR820/DOX ND with the same DOX concentration (50  $\mu$ g·mL<sup>-1</sup>). After incubation for 4 h, the medium was replaced with drug-free medium and the photothermal agent-treated cells received irradiation at 808 nm or 660 nm at a power of 1.0 W·cm<sup>-2</sup> for 2 min. After another 24 h, the cells were rinsed with fresh PBS, digested and suspended. Then, the cells were observed under a microscope (NIKON Ti-U, Japan) after trypan blue staining.

## Haemolysis Test

The biocompatibility of LA-IR820/DOX ND was evaluated by determining the haemolysis ratio (HR). Firstly, 2% (v/v) red cell suspension was prepared. Normal saline (1.1 mL) and 1.25 mL red cell suspension were mixed and then 0.15 mL LA-IR820/DOX ND was added with DOX concentrations ranging from 1 µg·mL<sup>-1</sup> to 100 µg·mL<sup>-1</sup>. Each sample was bathed at 37 °C for 3 h and then centrifuged at 1500 rpm for 15 min. The supernatant was collected and absorbance was measured at 576 nm with a UV-visible spectrophotometer. Red cell suspension (1.25 mL) mixed with 1.25 mL distilled water was used as the positive control, and 1.25 mL red cell suspension mixed with 1.25 mL normal saline was the negative control. Counteracting the interference absorption of LA-IR820 at 576 nm, equivalent LA-IR820 samples with different concentrations were prepared. The 2% red cell suspension was replaced by an equal volume of normal saline, and the other steps were conducted under the same conditions. The absorbance difference between LA-IR820/DOX ND and LA-IR820 was considered the true absorbance of each sample. HR was counted as follows:

$$HR(\%) = \frac{A_{\text{sample}} - A_{\text{negative}}}{A_{\text{positive}} - A_{\text{negative}}} \times 100\%$$

where  $A_{\text{sample}}$   $A_{\text{negative}}$  and  $A_{\text{positive}}$  are the absorbances from each sample, negative control and positive control, respectively.

# Statistical Analysis

Data were expressed as the mean  $\pm$  SD and analysed by nonlinear regression using GraphPad Prism 7.0 software.

## **Results and Discussion**

The HCC-targeted LA-IR820 was synthesized according to our previous report.<sup>32</sup> The assembly process of LA-IR820/DOX ND is presented in Figure 1A. We obtained LA-IR820/DOX ND through a simple procedure that only required

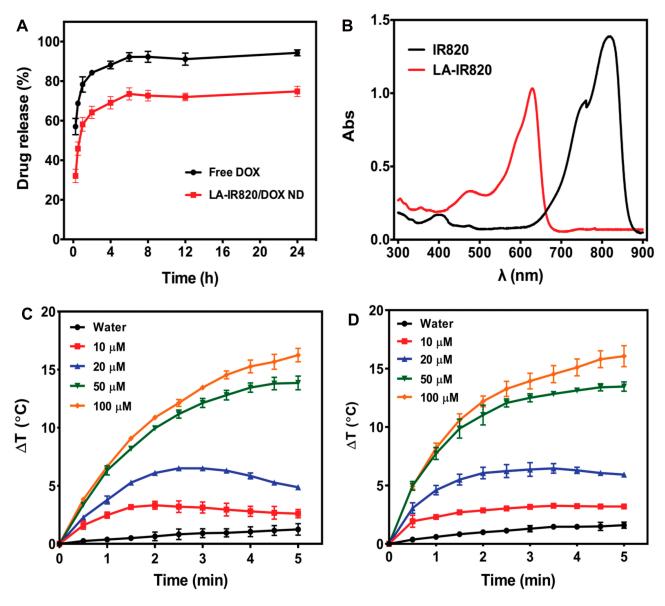


Figure 2 (A) The release curves of free DOX and LA-IR820/DOX ND in pH 7.4 PBS; (B) the UV-visible spectra of IR820 and LA-IR820; temperature increase profiles of different concentrations of (C) IR820 and (D) LA-IR820/DOX ND after irradiation with a laser wavelength of 808 nm and 660 nm, respectively (P = 1.0 W cm<sup>-2</sup>, n = 3).

mixing the two components in an aqueous solution. To determine the morphology of aggregates, TEM was used. The appearance of spherical nanoparticles in the TEM image proved the successful self-assembly of LA-IR820/DOX ND without any excipients (Figure 1B). The organization of the ordered nanostructures was most likely driven by  $\pi$ - $\pi$  stacking interactions, hydrogen bonds, electrostatic interactions, and hydrophobic interactions. The hydrophobic aromatic ring structures of LA-IR820 and DOX can induce the formation of hydrophobic cores through hydrophobic interactions and  $\pi$ - $\pi$  stacking interactions. At the same time, the hydrophilic fragment sticks out of the core to further stabilize the formed nanostructure. This 100% drug-loaded system

contained 48.9 wt % LA-IR820 and 51.1 wt % DOX, which are much higher values than those of excipient-assisted nanosystems. DLS was further used to survey the size and size distribution. As shown in Figure 1C, the hydrodynamic diameter of LA-IR820/DOX ND was  $174.0 \pm 10.2$  nm with quite a narrow PDI of 0.169. Zeta potential of LA-IR820/DOX ND was -15.33 mV. Previous studies have shown that 10-200 nm nanoparticles can passively target tumour sites through the enhanced permeability and retention (EPR) effect. Therefore, our LA-IR820/DOX ND is appropriate for both passive and active tumour targeting.

The release behaviour of DOX from LA-IR820/DOX ND was investigated in PBS, pH 7.4. Release profiles are

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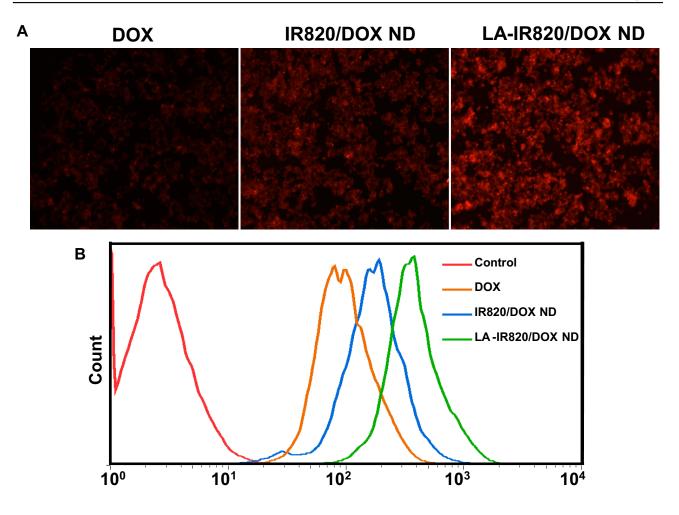


Figure 3 Intracellular uptake of DOX, IR820/DOX ND and LA-IR820/DOX ND in HepG2 hepatoma cells: (A) fluorescence microscopy images, (B) flow cytometry results.

shown in Figure 2A. The release rate of DOX from the LA-IR820/DOX ND was slower than that of free DOX. Compared to the 78.3% cumulative release of free DOX in 1 h, the LA-IR820/DOX ND did not reach achieve the same amount of DOX release in 24 h. Therefore, sustained LA-IR820/DOX ND exhibited its potential in prolonging circulation time and reducing clearance rate to a certain extent. 35-37 The UV-vis spectra (Figure 2B) show that the maximum absorption peak of IR820 was at 821 nm while the peak absorption of the HCC-targeted LA-IR820 had a blueshift to 630 nm. Therefore, using a 660 nm laser is much more reasonable for LA-IR820/DOX ND irradiation, and an 808 nm laser is suitable for IR820 irradiation. To evaluate the photothermal efficacy of LA-IR820/DOX ND in PTT, the temperature increase after laser irradiation was tested in vitro. For comparison, different concentrations of free IR820 were investigated as the control group.

IR820 and LA-IR820/DOX ND were irradiated for 5 min at 808 nm and 660 nm, respectively. As illustrated in Figure 2C and D, it was obvious that the temperature-increasing capability correlated with concentration, indicating their concentration-dependent photothermal effect during the tested time interval. Notably, IR820 and the LA-IR820/DOX ND had similar photothermal efficacy under laser irradiation, which demonstrated that the lactobionic acid modification had negligible effects on the photothermal properties of IR820.

Effective intracellular uptake is a precondition for satisfactory chemotherapeutic effect of DOX. Therefore, the intracellular internalization of LA-IR820/DOX ND was investigated qualitatively and quantitatively in HepG2 hepatoma cells. In the fluorescence microscope experiment, the fluorescence of DOX was used to indicate the intracellular uptake. To verify the targeting effect of the LA-modified nanodrug, the IR820

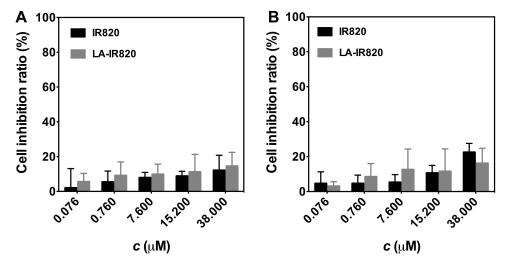


Figure 4 Dark toxicity of IR820 and LA-IR820 at various concentrations to (A) HepaI-6 cells and (B) HepG2 cells.

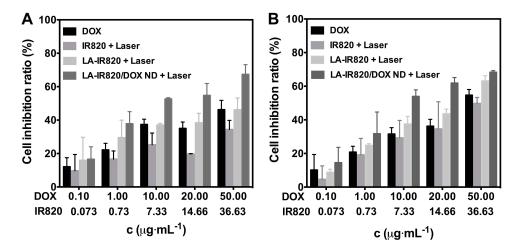


Figure 5 Cell inhibition ratio of DOX, IR820 + laser (808 nm, 1.0 W cm<sup>-2</sup>, 2 min), LA-IR820 + laser (660 nm, 1.0 W cm<sup>-2</sup>, 2 min) and LA-IR820/DOX ND + laser (660 nm, 1.0 W cm<sup>-2</sup>, 2 min) at various concentrations incubated with (**A**) Hepa1-6 cells and (**B**) HepG2 cells.

and DOX co-assembly nanodrug (IR820/DOX ND) was prepared as an untargeted control. As shown in Figure 3A, the LA-IR820/DOX ND group exhibited much stronger fluorescence intensity than not only the free DOX group but also the IR820/ DOX ND group. The increased IR820/DOX ND uptake in HepG2 cells was mainly because IR820/DOX ND entered cells through endocytosis instead of passive diffusion as free DOX. More importantly, the significantly improved LA-IR820/DOX ND internalization could be attributed to the LA ligand on the surface of this nanodrug, which can bind to the overexpressed ASGP receptor on the surface of hepatoma cancer cells. Quantitative analysis by flow cytometry agreed with the qualitative (Figure 3B) analysis. Consequently, these results obviously suggested that LA- IR820/DOX ND is competent to act as both the cargo and carrier, actively targeting drug delivery into hepatic carcinoma cells and guaranteeing the efficacy of chemotherapy and PTT.

To evaluate the safety of LA-IR820, murine Hepa1-6 cells and human HepG2 cells were selected for the cytotoxicity assay. The cytotoxicity of IR820 and LA-IR820 at different concentrations was assessed in the absence of laser irradiation, and the LA-IR820 showed rather low cytotoxicity on both cell lines within the studied concentration range (Figure 4). LA-IR820 displayed a small increase in toxicity compared with IR820, which was most likely caused by improved intracellular uptake through receptor-mediated endocytosis and could also be due to the HCC-targeting property of LA. LA-IR820 would reduce the damage to

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**Table I**  $IC_{50}^{a}$  Values of DOX, IR820 + Laser, LA-IR820 + Laser and LA-IR820/DOX ND + Laser Against Hepa1-6 and HepG2 Cells

Cell Line	Groups	IC <sub>50</sub> (μM)
Hepal-6	DOX IR820 + Laser LA-IR820 + Laser LA-IR820/DOX ND + Laser	175.8 588.5 82.43 6.349
HepG2	DOX IR820 + Laser LA-IR820 + Laser LA-IR820/DOX ND + Laser	93.19 59.21 20.81 5.884

Note: <sup>a</sup>The half-maximal inhibitory concentrations.

normal cells, which suggests that LA-IR820 could be a high-safety therapeutic agent for biomedical applications. To further assess the safety of the LA-IR820/DOX ND, a hae-molysis test was performed. As shown in <u>Figure S1</u>, the haemolysis ratios of the LA-IR820/DOX ND were all less than 5% at different concentrations, indicating its good biocompatibility and safety.

The antitumour activity of DOX, IR820 with laser, LA-IR820 with laser, and LA-IR820/DOX ND with laser was investigated. As shown in Figure 5, the antitumour ability of each group was in a dose-dependent manner. Notably, different concentrations of LA-IR820/DOX ND in the presence of laser exhibited the highest inhibition ratios of both cells among all groups. This result can be explained by the fact that the LA-IR820/DOX ND are efficiently internalized by hepatoma cells through ASGP receptormediated endocytosis due to the specific binding ability originating from the LA moiety. After endocytosis, DOX was released intracellularly, achieving its therapeutic effect. To quantitatively analyse the cytotoxicity, the halfmaximal inhibitory concentrations (IC<sub>50</sub> values) were calculated. As shown in Table 1, the IC<sub>50</sub> value of IR820 was approximately 7.1 times and 2.8 times higher than that of LA-IR820 for the two cell lines under the same laser irradiation, which proved the excellent tumour targeting ability endowed by the LA motif. Specifically, the IC<sub>50</sub> value of LA-IR820/DOX ND in the presence of the laser was 6.349 µM and 5.884 µM, which confirmed that LA-IR820/DOX ND could produce a marked antitumour effect under laser irradiation. The cytotoxicity of LA-IR820/ DOX ND in the presence of laser was also verified by trypan blue staining (Figure S2). The results were consistent with those of the MTT assay. The maximum quantity of stained cells was observed in the LA-IR820/DOX ND with laser treatment. Therefore, synergistic antitumour activity was successfully achieved by the combination of tumour-targeted photothermal therapy and chemotherapy.

#### **Conclusion**

In summary, we developed an excipient-free nanodrug composed only of HCC-targeted IR820 and chemotherapeutic DOX via pure self-assembly. In the present study, LA-IR820 and DOX co-assembled into well-defined nanospheres with optimal size and distribution, ensuring passive tumour-targeting ability. The 100% drug loading (48.9 wt % LA-IR820 and 51.1 wt % DOX) resolved the extremely low drug loading and excipient-induced toxicity problems of excipient-assistant systems. Specifically, LA-IR820 endowed the LA-IR820/DOX ND with active targeting capability. The sustained LA-IR820/DOX ND release made it potential to prolong circulation time and reduce clearance rate of the hydrophilic IR820 and DOX. The assembled LA-IR820/DOX ND exhibited remarkably enhanced accumulation in hepatoma cells, providing an opportunity to overcome the undesirable side effects. This excipient-free synergistic nanodrug demonstrated higher antitumour activity than individual treatment with the same dose. Thus, this excipient-free LA-IR820/DOX ND may be a promising candidate for synchronous drug delivery and synergistic chemo-photothermal therapy targeting hepatoma cells.

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#### **Disclosure**

The authors report no conflicts of interest in this work.

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