REVIEW

Nano Encapsulated Curcumin: And Its Potential for Biomedical Applications

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Abstract: Curcumin, a yellow-colored polyphenol extracted from the rhizome of turmeric root, is commonly used as a spice and nutritional supplement. It exhibits many pharmacological activities such as anti-inflammatory, anti-bacterial, anti-cancer, anti-Alzheimer, and anti-fungal. However, the therapeutic application of curcumin is limited by its extremely low solubility in aqueous buffer, instability in body fluids, and rapid metabolism. Nano delivery system has shown excellent potential to improve the solubility, biocompatibility and therapeutic effect of curcumin. In this review, we focus on the recent development of nano encapsulated curcumin and its potential for biomedical applications.

Keywords: curcumin, nanoformulations, bioavailability, nanoparticles, therapy

Introduction

Curcumin (Cur), 1,7-bis(4-hydroxy-3-methoxy-phenyl)-1,6-heptadiene-3,5-dione,¹ is a yellow-colored polyphenol extracted from the rhizome of turmeric root (Curcuma longa).^{2,3} Commercial curcuminoid powder is a mixture of curcumin, demethoxycurcumin and bisdemethoxycurcumin in a ratio of 77:17:3 (Figure 1). It has been used as a spice and dietary supplement as well as a component of many traditional medicines in the oriental world for many centuries.^{4,5} In recent decades, curcumin is shown to exhibit anti-inflammatory, anti-bacterial, anti-cancer, anti-Alzheimer, anti-fungus and so on.^{6–12}

However, its poor bioavailability, extremely low solubility in aqueous buffer, instability in body fluids, and rapid metabolism have jeopardized the therapeutic applications of curcumin in clinic.^{13,14} Its water solubility is only at 0.0004 mg/mL at pH 7.3.¹⁵ Besides this, soluble curcumin molecules are extremely sensitive at physiological conditions owing to rapid hydrolytic degradation.¹⁶ Numerous preclinical and clinical studies in mice and humans showed a low bioavailability of curcumin.^{17,18} Although 12 g of curcumin administered orally in humans was still in the safe range, the final curcumin level existed in serum was approximately only 50 ng/mL, this reflected a minimum availability of curcumin in the systemic circulation to achieve its therapeutic effects.^{2,19}

To circumvent these obstacles, researchers encapsulate curcumin into nanocarriers, such as liposomes, polymeric micelles, polymeric nanoparticles, mesoporous silica nanoparticles, protein-based nanocarriers, solid lipid nanoparticles, cyclodextrins, nanogels, nanocrystals etc.^{20,21} What are nanocarriers in pharmaceutical materials? Nanocarriers generally refer to pharmaceutical materials and products of approximately 1 nm to 100 nm;²² however, the US Food and Drug Administration

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Figure I Chemical structures of (A) curcumin, (B) demethoxycurcumin and (C) bisdemethoxycurcumin.

(FDA) often considers 1000 nm as an upper limit.²³ Why researchers encapsulate drugs into nanocarriers? Because of the high surface area to volume ratio offered by nanocarriers, both the solubility and dissolution rate of the drugs can be increased. Furthermore, the small particle size can prolong the drug's maintenance in the systemic circulation, modify drug distribution, and permit drug targeting and transport across barriers. Due to the aforementioned reasons, curcumin encapsulated in nanoformulations can effectively improve its solubility and bioavailability.^{24,25}

In this review, we will retrospect the different nano encapsulated curcumin formulations and their potential for biomedical applications.

Nano Formulations

Liposomes

Liposomes are the first nanocarriers approved by FDA for clinical application.²⁶ As a spherical bilayer vesicle with an aqueous core, a liposome is composed of phospholipid bilayer, and the particle size is generally between 25–1000 nm.²⁷ Hydrophilic drugs can be loaded in the aqueous cores and lipophilic drugs can be incorporated into liposome bilayers. According to the structure, liposomes can be divided into single unilamellar vesicles (SUVs), large unilamellar vesicles (LUVs) and multilamellar vesicles (MLVs).²⁸ The preparation methods of liposome include

3100 submit your manuscript | www.dovepress.com DovePress thin-film dispersion method, ultrasonic dispersion method, injection method, reverse phase evaporation method etc.²⁹ Since their compositions are the same as human cell membranes, liposomes have incomparable advantages as targeted drug carriers. Owing to their properties of non-toxic, non-immunogenicity, assorted carrying capacity and easy preparation, numerous liposomal formulations have been developed and some liposomal drugs have been launched.^{26,30,31} However, the shortcomings of low encapsulation efficiency, easy fusion and leakage are potential obstacles for further application.³²

In recent years, plenty of Cur-loaded liposomal formulations have been developed. Shi et al prepared curcumin-loaded liposomes (Lipo-cur) composed of lecithin and cholesterol to inhibit radiation pneumonitis and sensitize lung carcinoma to radiation by thin-film dispersion method.33 Lipo-cur had an average particle size of 114.9 nm, a zeta potential of -2.62 mV, a drug loading of 5% and an encapsulation efficiency of 90.1%, respectively. Liposomal formulation significantly increased the water solubility and reduced the potential longterm toxicity of curcumin. Radiation pneumonitis (RP) is a major dose-limiting side effect during thoracic radiotherapy.³⁴ It has been reported that curcumin was a radioprotective agent and could reduce the mortality rate in the irradiated mice.³⁵ In this study, it was observed that Lipocur inhibited nuclear factor-kappaB (NF-kB) pathway and downregulated relevant inflammatory factors induced by

thoracic irradiation, including transforming growth factor-B (TGF- β), tumor necrosis factor- α (TNF- α), interleukin-6 (IL-6) and IL-8. In addition, the combined treatment of Lipo-cur and radiotherapy exhibited enhanced anti-cancer efficiency in LL/2 Lewis lung carcinoma cell model. Increased intratumoral apoptosis and microvessel responses to irradiation were also reported. In another study, curcumin-loaded cationic liposomes were produced with the addition of didodecyldimethylammonium bromide (DDAB).³⁶ Compared to DDAB-free liposomes, DDAB-containing liposomes improved cell uptake and cytotoxicity of curcumin, likely due to the increased cell internalization capability of cationic liposomes. Dhule et al prepared curcumin-loaded 2-hydroxypropyl-y-cyclodextrin liposomal nanoparticles (HPyCD-curcumin liposomes) for the treatment of osteosarcoma.³⁷ HPyCD-curcumin system was first made to improve the solubility of Cur and then was encapsulated in the aqueous core of liposomes. The results of the cytotoxicity study showed that HPyCD-curcumin liposomes exhibited increased anticancer effect against KHOS osteosarcoma cells and MCF-7 breast cancer cells. In addition, more apoptotic were observed in the tumors treated with HPyCD-curcumin liposomes in vivo.

In order to enhance the antitumor efficiency of liposomal formulations, some new functions, such as long-circulating, ligand-mediated targeting, and responsive drug release were introduced into the design of liposomes.^{38,39} Mahmud et al prepared different types of long-circulating Cur-loaded liposomes by thinfilm hydration followed by extrusion technique.⁴⁰ 1,2-distearoylsn-glycero-phosphoethanolamine-N-(poly [ethylene glycol]₂₀₀₀) (DSPE-PEG₂₀₀₀) was used as the long-circulating lipid material. By comparing the parameters of particle size, zeta potential, shape and stability, it was found that PEGylated, cholesterol-free liposomes based on hydrogenated soy phosphatidylcholine (HSPC) at a drug-to-lipid molar ratio of 1:19 was the best formulation. This formulation had a low drug release in plasma and high stability in vitro. Furthermore, Cur-loaded long-circulating liposomes significantly induced the reactive oxygen species (ROS) level and upregulated the expression of caspase 3/7, leading to higher cytotoxic activity on AsPC-1 and BxPC-3 pancreatic cancer cells. Folate receptor (FR)-targeted liposomal Cur (F-CUR-L) was reported in our previous research (Figure 2).⁴¹ F-CUR-L composed of cholesterol, Egg phosphatidylcholine (Egg PC), DSPE-PEG₂₀₀₀, folate-poly [ethylene glycol]₃₃₅₀-cholesteryl hemisuccinate (folate-PEG3350-CHEMS) was prepared by thin-film dispersion. In a cellular uptake experiment, the mean fluorescence intensity of F-CUR-L was higher than that of non-targeted CUR-L both in KB and Hela cells, which overexpress FRs. However, in A549 cells that lack FRs, the uptake amounts of F-CUR-L, CUR and F-CUR-L+1 mM folic acid were all extremely low and nearly the same. Moreover, F-CUR-L exhibited enhanced antitumor activity than non-targeted CUR-L, demonstrating the increased tumor targeting by a FR-mediated endocytosis pathway. Ju et al fabricated a novel pH-sensitive liposomes using N-(3- Aminopropyl) imidazole-cholesterol (IM-Chol) and phosphatidylcholine (PC).⁴² Imidazolyl group has an electron lone pair on the unsaturated nitrogen and a pKa of about 6.0. It was



Figure 2 The uptake of F-CUR-L through folate receptor-mediated endocytosis pathway.

Liposomes are superior nanocarriers for combined drug delivery due to the ability of co-encapsulating both hydrophilic and lipophilic drugs. Ruttala et al developed PEGylated liposomes co-loading Cur and Paclitaxel (PTX) for enhanced synergistic antitumor efficacy.43 In this research, PTX-loaded albumin nanoparticles (APN) were encapsulated in the aqueous core and Cur was loaded in the liposome bilayers. Also, the ratio of PTX and Cur at 1:1 was chosen to prepare co-loaded liposomes (CL-APN) because of the significant inhibition of cell proliferation compared to other ratios. CL-APN had an average particle size of 250 nm and a zeta potential of 7 mV. In an MTT assay, CL-APN showed more cytotoxicity than individual drug-loaded liposomes in MCF-7 and B16F10 cells. It was believed that Cur downregulated the expression of NF-KB and protein kinase B (Akt), resulting in an increased therapeutic efficacy of PTX. Moreover, more early and late apoptosis, stronger G2/M arrest, and enhanced subG1-cell population were observed upon the treatment of co-loaded liposomes. In our previous work, Cur and cisplatin co-loaded liposomes were prepared and showed a synergistic effect against hepatocellular carcinoma (HCC).⁴⁴ Furthermore, enhanced intracellular ROS was also detected during the treatment of liposomes.

Polymeric Micelles

Polymeric micelles were used as nanoscale drug delivery system (nano-DDS) in the late 1980s.45 Micelles are thermodynamically stable colloidal solutions, formed by selfassembly of amphiphiles in water above the critical micelle concentration (CMC).⁴⁶ Structurally, the micelles are composed of hydrophilic shell and hydrophobic core which can load hydrophobic drugs. Various amphiphilic polymers, such as diblock, triblock and graft copolymers have been synthesized to prepare micelles.^{47–49} Among these, common hydrophilic blocks included poly (ethylene glycol) (PEG), chitosan and polyvinyl pyrrolidone (PVP). In addition, hydrophobic materials such as distearoyl phosphoethanolamine (DSPE), poly (lactic acid) (PLA), dioleoyl phosphatidylethanolamine (DOPE), poly (ε-caprolactone) (PCL), poly (lactide-co-glycolide) (PLGA), cholesteryl hemisuccinate (CHEMS) and vitamin E were used to form the cores of micelles. The size of micelles is generally ranging between 20 and 100 nm, which allows micelles overflow from tumor vessel walls and enter cancer cells.⁵⁰ Meanwhile, with the incorporation of an outer hydrophilic shell, micelles are able to escape the clearance of reticuloendothelial system (RES) and increase systemic circulation time of drugs.

As shown in Table 1, numerous Cur-loaded polymeric micelles have been developed. Liu et al synthesized monomethyl poly (ethylene glycol)-poly (ɛ-caprolactone) copolymer (MPEG-PCL) and prepared Cur-loaded micelles (Cur-M)



Figure 3 Schematic representation of pH-triggered release of IM-Chol liposomes. Reprinted from *Int J Pharm.* 518. Ju L, Cailin F, Wenlan W, et al. Preparation and properties evaluation of a novel pH-sensitive liposomes based on imidazole-modified cholesterol derivatives, 213–219, Copyright 2017, with permission from Elsevier.⁴²

Types of Micelles	Composition	Preparation Methods	Major Outcome	References
Normal micelles	MPEG-PCL	Solid dispersion method	Improved water solubility Inhibition of breast tumor growth and spontaneous pulmonary metastasis	[51]
EGFR-targeted micelles	PLGA-PEG GEII peptides	Emulsification solvent evaporation method	Increased intracellular uptake Prolonged circulation A reduction in primary tumor burden	[58]
FR-targeted micelles	Folate-PEG ₃₀₀₀ - PLA ₂₀₀₀ mPEG ₂₀₀₀ - PLA ₂₀₀₀	Thin film-hydration method	Increased intracellular uptake Sustained drug release Improved solubility Cancer targeting Enhanced anti-cancer activity	[13]
pH-sensitive micelles	TPGS-PAE	Solvent evaporation method	Low polydispersity High encapsulation efficiency Enhanced release in the acidic environment Improved pro-apoptotic and antiangiogenic activities	[60]
Redox-sensitive micelles	C ₁₆ -SS-CS- mPEG	Dialysis method	Controlled drug release Enhanced antitumor and anti-inflammation efficacy	[61]

Table I Different Types of Cur-Loaded Polymeric Micelles

by one-step solid dispersion method.⁵¹ Cur-M had a small particle size (28.2 nm) with good dispersity, whereas the drug loading capacity was 14.84% and the encapsulation efficiency was 98.91%, respectively. Compared to free curcumin, Cur-M showed a sustained release behavior at pH 7.4 in vitro and $\sim 60\%$ of drugs were released in one week. Moreover, IC₅₀ value for Cur-M was a little lower than that for free drugs on 4T1 cells since the enhanced cellular uptake of micelles. The similar result was also reported by Manjili et al using the MPEG-PCL micelles.⁵² Furthermore, enhanced chemotherapeutic efficacy, improved anti-metastasis activity and prolonged survival were observed in 4T1 xenograft tumorbearing nude mice. Another Cur-loaded polymeric micelle consisted of PLGA-PEG-PLGA copolymer was prepared by a solvent-dialysis method and the CMC of PLGA-PEG-PLGA was at a lower concentration $(2.82 \times 10^{-5} \text{ g/mL})$ at room temperature which indicated the strong hydrophobicity of copolymer.⁵³ The average particle size, zeta potential, drug loading capacity and encapsulated efficiency were 26.29 nm, -0.71 mV, 70% and 6.4%, respectively. In pharmacokinetics analysis, the plasma parameters including distribution half-life $(t_{1/2\alpha})$, elimination half-life $(t_{1/2\beta})$, area under the curve $(AUC_{0-\infty})$ and mean residence time $(MRT_{0-\infty})$ of Cur micelles were 2.48, 4.54, 1.31, and 2.67-times than that of free Cur solution. The biodistribution study showed that Cur-loaded

micelles reduced drug distribution in liver and spleen, and increased distribution in lung and kidney. Moreover, enhanced accumulation of Cur in the brain was detected and it may be related to the breakthrough of blood-brain barrier (BBB) because of the smaller diameter and the nearly neutrally charged property of Cur-loaded polymeric micelles. Manjili et al prepared poly (caprolactone)-poly (ethylene glycol)-poly (caprolactone) (PCL-PEG-PCL) nanoparticles with a nanoprecipitation method.⁵⁴ The results showed that CUR-loaded nanoparticles significantly inhibited cell proliferation, induced apoptosis and enhanced anti-tumor immunity stimulation. Phan et al synthesized PLA-PEG copolymers at the weight ratio of PLA/PEG in 3:1, 2:1, 1:1, 1:2 and 1:3, and then loaded curcumin in micelles by emulsification/solvent evaporation method.55 It was reported that decreasing the amount of PLA (from 3: 1 to 1: 3) resulted in the reduction of drug loading capacity. However, a higher loading capacity of PLA-PEG micelles did not lead to greater cytotoxicity of the samples. Cur/PLA-PEG micelles achieved the most effective inhibition on HepG2 cells at the PLA/PEG ratio of 1:3. It may be attributed to the smaller size, higher release rate and better biocompatibility.

In order to increase the intracellular uptake and improve therapeutic efficacy, the surface of micelles was modified with various targeted ligands to recognize tumor cell surface-overexpressed receptors.^{56,57} Jin et al have developed an epidermal growth factor receptor (EGFR)-targeted polymeric micelle to deliver curcumin for breast cancer therapy.⁵⁸ Cur-loaded PLGA-PEG nanoparticles (Cur-NPs) were synthesized by the emulsification solvent evaporation method. Then EGFR-targeted GE11 peptides were chemically conjugated to Cur-NPs to prepare GE11-modified Cur-loaded PLGA-PEG nanoparticles (GE11-Cur-NPs). The release profiles of Cur-NPs and GE11-Cur-NPs at pH 7.4 PBS were similar for approximately 24 hours. Intracellular uptake test showed that GE11-Cur-NPs delivered more drugs to EGFR-overexpressing MCF-7 cells compared with EGFR-low-expression MCF-10A cells. Furthermore, GE11-Cur-NPs exhibited higher intracellular fluorescence intensity than Cur-NPs after MCF-7 cells were incubated with both drug formulations for 2, 6 and 12 h. In the studies of pharmacodynamics, loading Cur into targeted PLGA-PEG NPs significantly enhanced the anti-cancer effect both in vitro and in vivo. Yang et al investigated the antitumor activity of FR-targeted micelles loaded with curcumin.¹³ In this study, Folate-PEG₃₀₀₀-PLA₂₀₀₀, a type of FR-targeted material was first synthesized. Then two kinds of micelles were prepared with mPEG₂₀₀₀-PLA2000 (Cur-PPs) or mPEG2000-PLA2000/Folate-PEG3000-PLA₂₀₀₀ (Cur-FPPs) through the thin film-hydration method. In an in vitro cellular uptake study, coumarin 6 was used as a model drug. Results showed that modification with folate on the surface of micelles could increase cellular uptake both on MCF-7 or HepG2 cells through folate mediated targeting. Meanwhile, enhanced uptake of Cur resulted in increased anti-cancer activity. The IC₅₀ value of Cur-FPPs was 42.9 µg/mL while that of Cur-PPs was 52.9 µg/mL on MCF-7 cells, and the IC₅₀ values were 43.3 and 54.3 µg/mL for Cur-FPPs and Cur-PPs on HepG2 cells, respectively.

In recent years, tumor microenvironment-responsive polymeric micelles have been widely investigated to deliver Cur and other drugs.⁵⁹ Zhang et al developed pH-sensitive micelles prepared from D-a-tocopheryl polyethylene glycol 1000-block-poly (B-amino ester) (TPGS-PAE) polymers for combined delivery of Dox and Cur.60 TPGS-PAE was synthesized by the Michael addition polymerization method, and Dox/Cur co-loaded NPs (D + C/NPs) were prepared by a one-step solvent evaporation method. D + C/NPs could rapidly release entrapped drugs under acidic environment (Figure 4). After incubation of 192 h, 59.37% of Dox and 66.63% of Cur in NPs were released at pH 7.4, whereas over 90% of drugs were released at pH 5.8.60 In another study, redox-responsive polymeric micelles, composed by monomethoxy-poly (ethylene glycol)-chitosan-S-S-hexadecyl (C16-SS-CS-mPEG) were designed for enhanced anticancer efficacy.⁶¹ Moreover, monomethoxy-poly (ethylene glycol)-chitosan-C-C-hexadecyl (C16-CC-CS-mPEG) without a redox-responsive disulfide bond was synthesized as a control using the same method (Figure 5). In the presence of reducing agents in vitro, it was observed that more drugs were released from C16-SS-CS-mPEG@Cur micelles compared with C16-CC-CS-mPEG@Cur micelles. The similar results were found in the intracellular drug release study. Furthermore, C16-SS-CS-mPEG@Cur efficiently enhanced cellular uptake of Cur, leading to improved bioactivity both in vitro and in vivo.

Polymeric Nanoparticles

Polymeric nanoparticles are solid colloidal particles formed from polymer materials. Owing to their good biodegradability, excellent biocompatibility, controllable structure,



Figure 4 In vitro release profiles of (A) Dox and (B) Cur from D + C/NPs at pH 7.4 or 5.8. Reprinted from Acta Biomater. 58. Zhang J, Li J, Shi Z, et al. pH-sensitive polymeric nanoparticles for co-delivery of doxorubicin and curcumin to treat cancer via enhanced pro-apoptotic and anti-angiogenic activities, 349–364, Copyright 2017, with permission from Elsevier.⁶⁰





Figure 5 Chemical structures of (A) C₁₆-SS-CS-mPEG and (B) C₁₆-CC-CS-mPEG.

and easy industrial production, polymeric nanoparticles have attracted widespread attention and research.^{62–64} In recent years, numerous natural and synthetic polymers have been used to prepare polymeric nanoparticles, for example, PLGA, poly(butyl cyanoacrylate) (PBCA), N-isopropylacrylamide (NIPAAM), polyvinyl alcohol (PVA), polyamidoamine (PAMAM)⁶⁵ and chitosan (CS).

PLGA

PLGA is polymerized from lactic acid and glycolic acid. It is an organic polymer with good biocompatibility and biodegradability. In the United States, PLGA passed FDA certification and was officially included in the United States Pharmacopoeia as a pharmaceutical excipient.⁶⁶

The improved oral bioavailability of Cur encapsulated by PLGA nanoparticles was reported. Xie et al produced Cur-loaded PLGA nanoparticles (CUR-PLGA-NPs) with a particle size of about 200 nm by a solvent evaporation method.⁶⁷ The drug loading capacity and encapsulation efficiency were 5.75% and 91.96%, respectively. In addition, the solubility of CUR-PLGA-NPs was 4.35 mg/mL which was much higher than that of free Cur (6.79 μ g/mL). In a pharmacokinetic study in vivo, rats were orally administrated of CUR-PLGA-NPs and free Cur at a dose of 100 mg/kg. The results indicated that PLGA NPs improved the oral bioavailability of Cur at 5.6-fold and had a longer half-life compared to free Cur. The authors thought that the improved oral bioavailability was associated with increased water solubility, higher release rate, improved absorption, inhibition of P-glycoprotein-mediated efflux and increased residence time in the intestinal cavity. Meanwhile, a similar result of improved oral bioavailability of Cur-loaded PLGA NPs was reported by Shaikh et al.⁶⁸ The relative bioavailability of Cur entrapped nanoparticles was 9-fold compared

with that of curcumin administered with piperine as an absorption enhancer.

PLGA nanoparticles are widely used in cancer therapy due to their good biocompatibility and stability.⁶⁹⁻⁷¹ Yallapu et al prepared Cur-loaded PLGA nanoparticles using PVA and poly (L-lysine) (PLL) as stabilizers by a nano-precipitation technique.⁷² The particle size of Curloaded PLGA nanoparticles was decreased from 560.4 nm to 95.6 nm with the increased concentration of PVA from 0 to 1% (w/v). Moreover, the addition of PLL further reduced the size of nanoparticles (76.2 nm). The zeta potential values of all nanoformulations were close to 0 mV, indicating a better stability.⁷³ Furthermore, higher drug encapsulation, increased Cur solubility, sustained drug release and enhanced intracellular uptake in A2780CP and MDA-MB-231 cells were observed when Cur-loaded nanoparticles were prepared with 1% PVA in the presence of PLL (Nano-CUR6). Thus, Nano-CUR6 formulation was chosen for further pharmacodynamics studies. In an in vitro cytotoxicity study, IC₅₀ values for nano-CUR6 and free Cur were found to be 13.9 and 15.2 µM in A2780CP cells, and 9.1 and 16.4 µM in MDA-MB -231 cell, respectively. In addition, nano-CUR6 significantly inhibited colony formation, suppressed prosurvival genes and enhanced apoptosis of cancer cells. These data clearly indicated the superior efficacy of Curloaded PLGA nanoparticles for cancer therapy.

NIPAAM

NIPAAM is one of the most commonly used temperaturesensitive polymers with a low critical solution temperature (LCST) at 32 $^{\circ}C^{74}$ It is hydrophobic when the temperature is higher than LCST while it is hydrophilic when the temperature is lower than LCST. Owing to this reversible sensitivity to environmental temperature stimuli, NIPAAM copolymers are widely used in drug delivery, chromatographic analysis, and other fields.^{75–77}

Nanoparticles made from NIPAAM were widely investigated. Lim et al synthesized nanocurcumin using NIPAAM, vinylpyrrolidone (VP) and acrylic acid (AA).⁷⁸ MTT assays showed that curcumin nanoparticles significantly inhibited the growth of DAOY, D283Med and HSR-GBM1 brain tumor cells due to apoptotic induction and G2/M cell cycle arrest. In addition, nanocurcumin reduced the clonogenicity of DAOY, D283Med, U87, and HSR-GBM1 brain tumor cells, and the CD133-positive stem-like fraction in JHH-GBM14 and HSR-GBM1 glioblastoma cells. Downregulation of IGF, STAT3, Akt and Hh were also observed, indicating that Cur-loaded NPs could modulate the signaling pathways affecting both survival and neoplastic stem cell phenotype. Zeighamian et al prepared Cur-loaded nanoparticles from N-isopropylacrylamide-methacrylic acid (NIPAAm-MAA) using a post-polymerization method.⁷⁹ NPs showed higher cytotoxicity against MCF-7 breast cancer cells. After 72h incubation, the IC50 values of free Cur and Cur-loaded NIPAAm-MAA NPs were 13.7 and 7.3µM, respectively.

PBCA

PBCA nanoparticles were widely used in targeted drug delivery systems due to their good biocompatibility, biodegradability, and non-immunogenicity.^{80,81} The commonly used preparation methods of PBCA nanoparticles are emulsion polymerization method and interfacial polycondensation method. Drugs are loaded in the porous structure of nanoparticles.

To enhance the transport of curcumin to the brain, Sun et al prepared Cur-loaded PBCA nanoparticles (CUR-PBCN) using an anionic polymerization method.⁸² CUR-PBCN had a core-shell structure with an average particle size of 152.0 nm and a drug loading of 21.1%. Moreover, CUR-PBCN slowly released 75% of drugs without a burst within 24 h, indicating a sustained-release property of nanoparticles. In an in vivo pharmacokinetic and biodistribution study, Kunming mice were given curcumin solution at a dose of 10 mg/kg or CUR-PBCN at a dose of 5 mg/kg by tail intravenous injection. The results showed that the mean residence time of CUR-PBCN was 14-fold that of free Cur in plasma. Furthermore, nanoparticles improved $AUC_{0-\infty}$ of free Cur solution at 2.53-fold in the brain. In conclusion, the research demonstrated that PBCA nanoparticles could enhance the transport of Cur to the brain and have the potential to cross BBB. A novel PBCA nanoparticles coated with chitosan was synthesized through the emulsion polymerization method for the treatment of hepatocellular carcinoma.⁸³ In vitro, the results showed that curcumin nanoparticles could suppress the expression of COX-2 and VEGF, leading to increased anti-angiogenic activity. In vivo, Cur-loaded PBCA nanoparticles inhibited the growth of hepatocellular carcinoma in mice xenograft models and suppressed tumor angiogenesis.

Chitosan

Chitosan, a semi-synthetic polysaccharide is a linear polymer derived from chitin. Due to its good biodegradability, biocompatibility, non-immunogenicity, non-irritating, nonpyrogenic and non-toxicity, chitosan can be used as an ideal carrier material for drug delivery.⁸⁴ Recently, chitosan-based nanoparticles have been widely reported for pharmaceutical and biomedical applications.^{85,86}

Chuah et al produced curcumin-containing chitosan nanoparticles with a mean particle size of 340 nm, polydispersity index of 0.352 and zeta potential of 43.7 mV.⁸⁷ In a cytotoxicity assay, the IC₅₀ was about 10 μ M for free curcumin and less than 10 μ M for CUR-CS-NP after 72 h incubation on HT-29 cells. In addition, increased cellular uptake and induced cell apoptosis were also observed upon the treatment of nanoparticles, indicating the potential of CUR-CS-NP for colorectal cancer therapy. In a recent study, Cur was encapsulated into nanoparticles based on folate modified chitosan for FR-targeting.⁸⁸ The results of the cytotoxicity test showed that curcumin loaded folate-modified-chitosan-nanoparticles are potential carriers for targeted therapy of breast cancer.

Mesoporous Silica Nanoparticles

In recent years, porous materials, such as mesoporous silica nanoparticles (MSNs), metal-organic frameworks⁸⁹ and iron oxide-gold nanoparticles⁹⁰ have been widely used in drug delivery. Kresge et al first used surfactants as structure directing agents to prepare highly ordered MSNs with uniform pore sizes in 1992.91 In the past decade, MSNs have been immensely explored due to their unique properties such as good biocompatibility, large specific surface area, tunable pore sizes and volumes and easily surface modification. MSNs were firstly introduced as promising carriers for drug delivery by Vallet-Regi et al in the early 2000s.⁹² In addition, they have been projected to be a type of nontoxic candidate for biocatalysis, bioimaging and biosensor.^{93,94} Microwave synthesis, template synthesis, hydrothermal synthesis and sol-gel method are the most common method to prepare MSNs.⁹⁵ Among these, template synthesis method using surfactants is the most commonly used method due to the mild conditions and convenience.

The improved solubility and enhanced cell cytotoxicity of Cur encapsulated by mesoporous silica nanoparticles (MSNs) were reported. Jambhrunkar et al prepared CURloaded mesoporous silica nanoparticles (MCM-41-CUR) with a mean size of 190 nm through a rotary evaporator technique.⁹⁶ The adsorption of curcumin in the nanopores of MSNs was achieved via hydrogen-bond interaction (Figure 6). MCM-41-CUR improved the curcumin solubility and increased the in vitro release compared with free Cur and MCM-41-CUR PM, the physical mixture of MCM-41. Consequently, Cur loaded MCM-41 nanoparticles significantly enhanced the cell cytotoxicity in SCC-25 cells due to the activation of caspase-3 and the inhibition of PcG proteins. Moreover, the amino ligands modificated mesoporous silica nanoparticles were also investigated.^{97,98} The drug loading capacity of MSNs functionalized with polyamino ligands (MSU-2-CP-TR-Curc and MCM-41-CP-TR-Curc) was 4.3% and 3.6% respectively, which was higher than that of MSNs functionalized with monoamino ligands (MSU-2-AP-CUR: 3.5%; MCM-41-AP-CUR:3.5%). The results indicated that modification of polyamino ligands on the surface of MSNs increased the drug loading capacity. Thus, they proposed that curcumin was encapsulated in MSNs by the

interactions between the amino groups of silica materials and the polar groups of curcumin. Chen et al constructed a novel type of folate-receptor targeted mesoporous silica nanoparticles loaded curcumin via pH-sensitive schiff base reactions (FA-MSN-N=C-Cur).99 It was found that the average size of the nanoparticles increased from 136 nm to 165 nm with the modification of 3-aminopropyl trimethoxysilane (APTES), Cur and folate. In a study of pH-responsive release in vitro, the released amount of curcumin from nanoparticles at pH 7.4, 6.5 and 5.0 was 11.7%, 32.5% and 58.6%, indicating the pH-responsive release property of FA-MSN-N=C-Cur. In addition, FA-MSN-N=C-Cur could effectively target to FA receptor over-expressed MCF-7 cells. The MTT assay showed that FA-MSN-N=C-Cur increased the cell cytotoxicity in MCF-7 cells through FA-mediated endocytosis and tumor acidic environment-responsive release.

In a recent study, curcumin was loaded into MSN grafted of polyethyleneimine (PEI) and folic acid (FA) for targeted drug delivery.¹⁰⁰ The enhanced cellular uptake and increased cytotoxicity of MSN-PEI-FA confirmed the significance of nanocarriers conjugated with folic acid. Similarly, in another study, Li et al developed MSN modified with PEI-FA (MSN-PEI-FA) and hyaluronic acid (MSN-HA) via disulphide bonds to achieve FA receptor-or CD44-targeting.¹⁰¹



Figure 6 Schematic representation comparing cytotoxic effect of (A) pure curcumin and (B) curcumin loaded MCM-41on SCC-25 cells. MCM-41-CUR was achieved via hydrogen-bond interaction and showed higher cytotoxicity. Reprinted from *RSC Adv.* 4. Jambhrunkar S, Karmakar S, Popat A, et al. Mesoporous silica nanoparticles enhance the cytotoxicity of curcumin, 709–712, Copyright 2014, with permission from The Royal Society of Chemistry.⁹⁶

Surface modification of mesoporous silica nanoparticles with polymers plays an effective role in improving biocompatibility, providing controlled drug release, enhancing therapeutic effect and decreasing systemic toxicity. Elbialy et al prepared Cur-loaded PEGylated MSNs (PEG-MSNPs-Cur) with an average particle size of 184.6 nm, zeta potential of 20.8 mV and loading efficiency of 92.4%.¹⁰² The nanoparticles released about 52% of loaded drugs within 192 h at pH 5.5 while only 1% of Cur was released at pH 7.4. In addition, IC₅₀ values for PEG-MSNPs-Cur after 24 h incubation were 20 and 28 µg/mL for HepG2 and HeLa cells, which were much lower than free Cur. In another study, Mamani et al produced guanidine functionalized PEGylated MSNs for the treatment of breast cancer.¹⁰³ The characteristics of the nanoparticles, including high loading efficiency, biocompatibility, pH-sensitive, and penetrability made it a promising system for delivery of curcumin. Lipid bilayer and chitosan were also used to modify MSNs for enhanced transport of curcumin.^{104,105} Nasab et al developed the surface-modified MSNs with chitosan (CS-MCM-41) for glioblastoma treatment.¹⁰⁶ In this study, CS-MCM-41 slowly released about 42.72% of their content at pH 5.5 in 96 h due to covering the porous surface of MCM-41. Moreover, this nanocarrier increased the anti-cancer efficiency of curcumin against U87MG cells. The IC50 of CS-MCM-41 and free curcumin were 5.21 and 15.20 µg/mL, respectively.

Protein-Based Nanocarriers

As a natural biomaterial with unique advantages, protein has excellent non-immunogenicity, biocompatibility, biodegradability and low cost. Protein-based nanocarriers have been widely used in the field of biomedicine.^{107,108} In recent years, many forms of protein nanoparticles for Cur delivery were developed, such as bovine serum albumin (BSA), human serum albumin (HSA), ovalbumin (OVA), zein, casein and silk fibroin (SF) nanoparticles.

BSA

Albumin nanoparticle has a lot of superior properties that make it an attractive nanocarrier system. Albumin is biocompatible, nontoxic, non-immunogenic, biodegradable, easy to purify and soluble in water. The preparation of albumin nanoparticles can combine the advantages of both nanoparticles and albumin. Also, a noteworthy amount of drugs can be embedded in the hydrophobic pocket of albumin through hydrophobic interaction. BSA has a molecular weight of 69,323 Da and an isoelectric point of 4.7 in water at 25 °C.¹⁰⁹ BSA has been widely concerned and applied for drug delivery due to its low cost, easy access and ease of purification.

It was reported that polyphenols-BSA bindings occurs by hydrophilic and hydrophobic interactions.¹¹⁰ The binding constants of genistein, resveratrol and curcumin were 1.26 $\times 10^{4} \text{ M}^{-1}$, 2.52 $\times 10^{4} \text{ M}^{-1}$ and 3.33 $\times 10^{4} \text{ M}^{-1}$, respectively. These results indicated that the affinity of curcumin to BSA was stronger. The desolvation method was widely used to prepare BSA nanoparticles. Sadeghi et al investigated the effect of different desolvating agents on Cur-loaded BSA nanoparticles.¹¹¹ They found that using ethanol could produce more uniform spherical nanoparticles and acetone resulted in higher encapsulation efficiency. Salehiabar et al synthesized Cur-loaded BSA nanoparticles (BSA@CUR NPs) with a mean size of about 99.78 nm, a zeta potential of -9.19 mV, an entrapment efficiency of 78.12% and a loading efficiency of 2.61%.¹¹² The in vitro release behavior of BSA@CUR NPs was time and pH dependent due to the degradation of BSA in acidic medium. Moreover, an in vitro hemolysis assay on the human red blood cells (HRBCs) showed the biocompatibility of the nanoparticles. The same research group also prepared bovine serum albumin-coated magnetic nanoparticles to delivery curcumin (F@BSA@CUR NPs).¹¹³ The results of MTT assay revealed the significant cytotoxicity of nanoparticles on MCF-7 cells. In a recent study of Zhang et al, curcumin/BSA nanoparticles were used to suppress the toxicity of CuO.¹¹⁴ Nanoparticles were prepared by co-precipitation of BSA and curcumin and followed by glutaraldehyde cross-linking. Results showed that particles could reduce cytotoxicity and genotoxicity of CuO nanoparticles by decreasing the ROS level and Cu²⁺ concentration. In addition, in vivo instillation and inflammation assessment further demonstrated the inhibitory effect of CUR/BSA NPs on the toxicity of metal-based nanomaterials.

BSA-saccharide conjugates were produced to achieve excellent stability or targeted ability.¹¹⁵ Huang et al prepared Cur-loaded galactosylated BSA nanoparticles (Gal-BSA-Cur NPs) to target asialoglycoprotein receptor (ASGPR) over-expressed on HepG2 cells.¹¹⁶ BSA-galactose conjugate was first synthesized using the reductive amination method and then Gal-BSA-Cur NPs were prepared by the desolvation method. The intracellular uptake of Gal-BSA-Cur NPs, BSA-Cur NPs and free Cur in HepG2 cells were 240.0 \pm 18.2, 133.6 \pm 7.9 and 46.8 \pm 14.8 ng/10⁵ cells, respectively, indicating the importance of galactose-mediated endocytosis. Fan et al produced Cur-loaded BSA-dextran nanoparticles with a particle size of 115 nm.¹¹⁷ BSA-dextran nanoparticles significantly induced the cellular antioxidant activity of Cur because of the increased uptake.

HSA

In order to avoid a possible immunologic response, BSA can be replaced with HSA, the most abundant protein in human serum. HSA has a molecular weight of 66,500 Da and is consisted of 585 amino acid residues.¹⁰⁹ HSA nanoparticles have attracted great intention in the field of drug delivery.

Kim et al prepared curcumin-loaded HSA nanoparticles (CCM-HSA-NPs) with a mean size of 135.5nm using albumin bound technology.¹¹⁸ The solubility of CCM-HSA-NPs in water was 851.416 µg/mL while that of free curcumin was only 2.792 µg/mL. At all-time points of biodistribution study, tumor accumulations of CCM-HSA-NPs were remarkably higher than that of free Cur, probably due to the EPR effects and enhanced penetration of albumin. Moreover, the in vivo antitumor activity of nanoparticles was investigated in HCT116 and MiaPaCa2 xenograft tumor-bearing nude mice. CCM-HSA-NPs had a greater therapeutic effect than free drug without observable toxicity. In a recent study, folate-conjugated HSA nanoparticles loaded with curcumin were also reported.¹¹⁹ HSA NPs significantly improved the antitumor activity of free drug both in vitro and in vivo. Saleh et al fabricated Cur-loaded, redox-responsive HSA nanoparticles (CCM/ HSA NPs) by using a modified desolvation method.¹²⁰ GSH, a reducing agent was used to break down the disulfide bonds of HSA and curcumin was entrapped in the hydrophobic cavity to form HSA-CCM nanoparticles. The release rate of curcumin in CCM/HSA NPs was promptly increased when it was incubated with 10 mM GSH, indicating the property of reductive disintegration.

OVA

Ovalbumin, a highly functional food protein is usually obtained from egg white.¹²¹ It is consisted of 385 amino acid residues with a molecular weight of 47,000 Da and isoelectric point of 4.8.¹⁰⁹ Recently, the application of ovalbumin for drug delivery has gained interest owing to its inexpensive and biocompatible properties. Moreover, OVA is a promising carrier for controlled drug release because of its temperature- and pH- sensitive properties.

Liu et al prepared OVA/curcumin complex powder by freeze-drying.¹²² The mean particle size of OVA/curcumin complex in aqueous solution was 103.57 nm by dynamic light scattering (DLS) measurement. The authors reported that the complex of OVA and Cur resulted in the secondary structure change of OVA and the crystal structure disappear of Cur, leading to more prone to pyrolyze. Compared with free curcumin, OVA/curcumin complex showed a higher antioxidant activity, representing the potential applications of OVA/curcumin complex in functional food. Recent study reported that OVA nanoparticles could improve the oral bioavailability of curcumin.¹²³ In an in vitro gastrointestinal tract (GIT) model, the transformation of curcumin in ovalbumin-dextran nanogels was significantly higher than that in ovalbumin nanoparticles.

Zein

Zein, an alcohol-soluble protein is obtained from corn and has a molecular weight of 22 to 27 kDa.¹²⁴ The major fractions of zein include α -, β -, γ - and δ -zein, and hydrophobic amino acid residues account for more than 50%. Zein is one of the few proteins that can be solubilized in 60–90% ethanol solutions but not in pure water.¹²⁵ Due to its hydrophobicity, biodegradability and biocompatibility, zein has been widely investigated to encapsulate the hydrophobic bioactive compounds. Moreover, zein can easily form self-assembled nanoparticles by the method of antisolvent co-precipitation.

In the present work, Cur was successfully incorporated into zein fibers by electrospinning technique.¹²⁶ The encapsulation efficiency of CUR-zein fibers was close to one hundred percent. In an antibacterial test, Escherichia coli and Staphylococcus aureus were used as Gram-positive and Gramnegative model bacteria, respectively. The results showed that CUR-zein fibers have good antibacterial activity towards both two kinds of bacterias, indicating the potential to inhibit bacterial growth and propagation in food packaging. In addition, PEG, shellac, chitosan and pectin were used to coat and stabilize zein nanoparticles.^{125,127-129} Sun et al produced curcumin-loaded zein-shellac nanoparticles with a higher encapsulation efficiency than pure zein particles.¹³⁰ Also, the nanoparticles showed a sustaining drug release in PBS and simulated gastrointestinal fluids.

Casein

Casein, an amphiphilic protein with an isoelectric point of 4.8 is the major protein of milk.¹³¹ It is composed of α_{s1} -, α_{s2} -, β and κ -casein. Casein can spontaneously forms micelles and the average particle size is 150 nm.¹³² Esmaili et al used beta casein-micelle as a nanocarrier for hydrophobic drugs.¹³³ When Cur was loaded into the micellar nanostructures, its solubility enhanced at least 2500 fold. The IC₅₀ values for B-CN encapsulated and free Cur were 17.7 and 26.5 µmol/L for K562 cells, respectively. Antioxidant activity of Cur was also improved via the process of drug loading into beta caseinmicelle. Similarly, in another study, curcumin-loaded casein nanoparticles not only increase the solubility and bioactivity, but also improve the dispersibility of pristine Cur.¹³⁴

SF

Silk, an insoluble natural polymeric biomaterial, is obtained from Bombyx mori silkworm.¹³⁵ Silk consists of an inner core of silk fibroin and an outer layer of silk sericin. Because of the biocompatibility, controllable degradability, processibility and excellent mechanical properties, SF has been engineered for sustained and controlled drug delivery systems. Several curcumin-loaded SF nanoparticles have been reported.^{136,137} For example, Xie et al prepared Curcumin-silk fibroin nanoparticles (CM-SF NPs) using a solution-enhanced dispersion by supercritical CO₂ process.¹³⁸ The size distribution, particle size, drug loading and encapsulation efficiency were influenced by various parameters, including the ratio of Cur and SK, pressure, and final concentration of NPs. Gupta et al simultaneously fabricated pure SF and silk fibroin-chitosan (SFCS) nanoparticles to encapsulate curcumin using a capillary-microdot technique.¹³⁹ The encapsulation efficiency of SF-coated curcumin nanoparticles was higher than that of SFCS-coated nanoparticles. This may be because curcumin is a hydrophobic drug, so the presence of CS, a hydrophilic polymer reduces the encapsulation efficiency of curcumin by SK. In addition, Cur-loaded SF nanoparticles showed higher intracellular uptake and cytotoxicity against breast cancer cells.

Solid Lipid Nanoparticles

Solid lipid nanoparticles (SLNs) are novel colloidal drug delivery system composed of biodegradable solid lipids.¹⁴⁰ The drugs are wrapped or embedded in the lipid core consisted of the phospholipid hydrophobic chains and high melting fat matrix. The particle sizes are usually between 50 and 1000 nm.¹⁴¹ Previous studies have reported that SLNs exhibited unique advantages, such as high biocompatibility, physical stability, drug targeting and controlled drug release.

The encapsulation of Cur in SLNs have been widely investigated. Wang et al synthesized SLN-curcumin by sol-gel method for non-small cell lung cancer treatment.¹⁴² It was found that SLN-curcumin increased the antitumor effect both in vitro and in vivo. Similarly, in another study, Sun et al prepared Cur-loaded SLNs by high-pressure homogenization to improve the dispersibility, stability and bioavailability of curcumin, and enhance its cellular uptake and anti-cancer efficacy.¹⁴

The burst release of drugs from solid lipid nanoparticles in acidic environment limits the usage as an oral delivery system.

In order to inhibit the rapid release of curcumin from SLNs in acidic environment, N-carboxymethyl chitosan (NCC) coated curcumin-loaded SLN (NCC-SLN) was prepared.¹⁴³ NCC-SLN (182.0 ± 6.7 nm) has a larger particle size than curcumin-loaded SLN (C-SLN) (166.5 ± 5.4 nm) due to the successful surface modification. In an in vitro release study, 40.4% of encapsulated Cur in C-SLN was released while only 1.5% Cur was released from NCC-SLN in simulated gastric fluid (SGF) within 2 h. In addition, NCC-SLN improved the lymphatic uptake of free Cur at 6.3-fold and increased the oral bioavailability at 9.5-fold. These results indicated that NCC-SLN could be a promising oral delivery carrier for Cur.

Cyclodextrins

Cyclodextrins (CDs) are cyclic oligosaccharides with a structure of hollow truncated cone.¹⁴⁴ Due to its hydrophilic outer surface and lipophilic cavity, CDs are able to solubilize hydrophobic drugs. β -CD, γ -CD, hydroxypropyl- β -CD and methyl- β -CD are the most frequently used compounds (Figure 7). Yallapu et al developed β -cyclodextrin- curcumin (CD-CUR) inclusion complexes by a solvent evaporation method.¹⁴⁵ The IC₅₀ values of CD30 and free Cur were 16.8 and 19.6 µM for C4-2 prostate cancer cells, and 17.6 and 18.4 µM for DU145 prostate cancer cells, respectively. In another study, CUR was loaded into hydroxypropyl-β-CD (HP-β-CD) in the presence of different stabilizers.¹⁴⁶ It was found that PVA was the most suitable stabilizer, resulting in the highest encapsulation efficiency $(\sim 60\%)$ with lower nanoparticle size (40 nm). In addition, the results indicated that prepared HP-\beta-CD-curcumin nanoparticles could effectively overcome the multidrug resistance of DOX in COLO205 colorectal adenocarcinoma cells.

Nanogels

As a type of promising drug delivery carriers, nanogels have attracted a lot of attention.^{147–149} They are usually formed from amphiphilic or hydrophilic polymer networks by physical self-assembly or chemical crosslinking. The preparation of nanogels combines the advantages of both hydrogels and nanoparticles. Similar to other nanoparticles, nanogels can easily pass through tumor vessel walls and be taken up by cancer cells. Owing to their high loading capability, biocompatibility and responsive drug release property, nanogels are ideal candidates for drug delivery. In addition, the presence of large surface area and functional groups for further modification with targeted agents or other biomolecules is another advantage.

Mangalathillam et al developed Cur-loaded chitin nanogels (CCNGs) for skin cancer therapy.¹⁵⁰ The values of particle size, zeta potential and entrapment efficiency were 70–80 nm, 49.34



n=1 R ₁ =H R ₂ =H	β-CD
n=2 R ₁ =H R ₂ =H	γ-CD
n=1 R_1 =CH ₂ CHOHCH ₃ R_2 =H	Hydroxypropyl-β-CD
n=1 R_1 =H R_2 =C H_3	Methyl-β-CD

Figure 7 Chemical structures of most frequently used cyclodextrins

mV and 95%, respectively. The swelling studies showed that Cur conjugation decreased the swelling ratio of chitin nanogels both at alkaline, acidic and neutral conditions due to the reduction of free reactive functional groups. When exposed to fresh human blood, the hemolytic ratio of CCNGs was 2.45% which is far below 5%, the critical safe hemolytic ratio for biomaterials according to ISO/TR 7406.151 indicating the biocompatibility and biological safety of nanogels. In skin permeation studies in vivo, CCNGs increased the penetration of Cur. By histopathological evaluations, it was observed that the keratin was comparatively thin and stratum corneum was fragmented upon the treatment of nanogels. All these data indicated that CCNGs offered an effective transdermal delivery for the treatment of skin cancer. Wu et al designed and produced core-shell nanogels, which were consisted of Ag/Au nanoparticles as core and polystyrene-poly (ethylene glycol) gels as shell.¹⁶ The Ag/Au core exhibited strong absorption in the near-infrared region (NIR) for photothermal conversion and fluorescence for cellular imaging. Cur was loaded in the hydrophobic polystyrene chain networks through hydrophobic interactions. Temperature rise or NIR irradiation can accelerate the release of curcumin from nanogels. Furthermore, Cur-loaded hybrid nanogels showed higher therapeutic efficacy upon combined chemophotothermal therapy, indicating the potential for cancer treatment. In a recent study, Cur was conjugated to cholesterylhyaluronic acid (CHA) nanogel for targeted delivery to CD44-expressing cancer cells.¹⁵² Curcumin and CHA-CUR significantly down-regulated the expression of COX-2, TNF- α , and NF- κ B. In a tumor growth inhibition study using MiaPaCa-2 xenograft model, CHA-CUR resulted in 5-fold

tumor suppression than free drug, making this Cur-loaded nanogel a potential candidate for cancer therapy.

Environment-responsive nanogels were also prepared for anticancer drug delivery. A novel interpenetrating polymeric network nanogels (IPN-NGs) composed of gelatin (GL) acrylamidoglycolic acid (AGA) were produced by emulsion polymerization.¹⁵³ The IPN-NGs showed pH-sensitive properties due to dual crosslinking and the hydrogen bonds between polymer chains. Luckanagul et al developed thermo-responsive nanogels from chitosan-grafted poly-(N-isopropylacrylamide) (CS-g-pN) by sonication method.¹⁵⁴ The authors mentioned that pNIPAM has a low critical solution temperature of 32 °C. When the surrounding temperature is higher than LCST, drugs will release from the pNIPAM nanogels. However, no data regarding responsive drug release property were reported in this study.

Nanocrystals

Nanocrystals are carrier-free nanoparticles composed of pure drug crystals and minimum stabilizer.¹⁵⁵ Nanocrystals have small particle size and high drug loading capacity. Owing to the properties of increased surface area and saturation solubility, nanocrystals can solve the solubility and dissolution problems of most hydrophobic drugs, resulting in increased bioavailability and improved biodistribution. Nanocrystals were initially investigated to increase the bioavailability of poorly soluble oral medication. Nowadays, nanocrystals were widely developed for intravenous injection, transdermal delivery, ocular delivery, and pulmonary administration.^{156–159} The preparation methods of nanocrystals include bottom-up and top-down approaches¹⁶⁰ (Table 2). The former is a method in which nanoparticles are precipitated or crystallized through

Types of Preparation Methods	Preparation Methods	Particle Size (nm)	Stabilizers	References
Top-down	High-pressure homogenization method	500–800	PVA, PVP, TPGS, sodium carboxyl methyl cellulose (Na- CMC) or sodium dodecyl sulfate (SDS)	[163]
	Grinding method	250	Hydroxypropyl cellulose SL (HPC-SL) and SDS	[164]
	Grinding and high-pressure homogenization method	181	Stabilizer solution	[165]
Bottom-up	Precipitation method	104 or 135	Sodium lauryl sulfate or poloxamer 188	[166]
	Thin-film dispersion method	161.9	Pluronic F127	[167]

Table 2 Preparation Methods of Cur-Loaded Nanocrystals

the combination of molecules in solution, mainly including precipitation method and emulsification method. The latter is a method of dispersing large drug particles into small particles, mainly including grinding method and high-pressure homogenization method. In addition, nanocrystals are highly dispersed heterogeneous systems with a large specific surface area and require stabilizers to maintain stability.^{161,162} Surfactants and polymers, such as Tween 80, cetyltrimethy-lammonium bromide (CTAB), TPGS, PVA and pluronic were usually used to stabilize curcumin molecules.

Rachmawati et al prepared curcumin nanocrystals with different stabilizers by high-pressure homogenization (HPH).¹⁶³ It was found that PVP is the most efficient polymer to stabilize curcumin nanocrystal. The grinding method was also used to develop curcumin nanocrystals.¹⁶⁴ There appeared to be a significant improvement in oral bioavailability. In a recent study, novel curcumin nanocrystals were produced for dermal penetration enhancement by a smart crystal technology, which is a combination of grinding and HPH.¹⁶⁵ The average particle size and zeta potential were about 180 nm and -30 mV, respectively. The skin penetration profile was performed for curcumin nanocrystal with decreasing concentrations. The results showed that curcumin nanocrystals at 2%, 0.2% and 0.02% (w/w) revealed strong fluorescence intensity in the epidermis while at 0.002% revealed a very faint fluorescence.

In a study of Moorthi et al, the nanoprecipitation method with the assistance of sonication was used to prepared curcumin nanocrystals.¹⁶⁶ Sodium lauryl sulfate and poloxamer 188 were used as the stabilizers. Sonication could inhibit the growth of crystal at the initial stage and stabilizers inhibited further growth. In addition, curcumin nanocrystal stabilized by sodium lauryl sulfate produced a smaller particle size, higher surface area and higher negative potential than that stabilized by poloxamer 188. The authors considered that the anionic property of sodium lauryl sulfate overcame van der Waals force and gravitational force, resulting in the prevention of nanocrystal aggregation. Recently, curcumin nanocrystals coated by hyaluronic acid (HA@Cur-NCs) has been developed for the targeted drug delivery.¹⁶⁷ The curcumin nanocrystals (Cur-NCs) were first synthesized by thin-film dispersion and then coated with HA using a nanoprecipitation method. It was observed from transmission electron microscopy (TEM) images that HA@Cur-NCs showed a more regular structure after surface modification with HA (Figure 8). DLS analysis showed that the average particle size and zeta potential of HA@Cur-NCs were 161.9 \pm 1.7 nm and -25.0 \pm 0.8 mV, respectively. Compared to Cur-NCs, HA@Cur-NCs exhibited increased cellular uptake and cytotoxicity in breast cancer cells. It was also observed that HA@Cur-NCs showed better anticancer effects compared with Cur-NCs and free Cur in an in vivo animal experiments, demonstrating the necessity and importance of HA targeting.

Biomedical Applications

As described above, plenty of studies reported that different Cur-loaded nanoformulations showed anti-tumor effects for many types of cancer. Furthermore, curcumin nanoformulations exhibited the potential applications for the prevention and treatment of other diseases, such as brain injury,^{168,169} wound healing,^{170,171} malaria,^{172,173} asthma,¹⁷⁴ psoriasis,¹⁷⁵ arteriosclerosis,¹⁷⁶ leishmaniasis,¹⁷⁷ bacterial infection⁸ (Table 3), etc.

Due to its promising therapeutic effect, several curcumin formulations based on free form and nanoparticles have been under investigation in clinical trials.^{178–181} In recent years, some clinical trials were performed to evaluate the improved bioavailability of curcumin nanoformulations. In a human pharmacokinetic study, healthy subjects were divided into three groups and then were given single oral medication of



Figure 8 Characterization of formulations. (A) Appearances of free Cur, Cur-NCs, and HA@Cur-NCs. TEM images of (B) Cur-NCs and (C) HA@Cur-NCs. Reprinted from *Biomater Sci.* Ji P, Wang L, Chen Y, et al. Hyaluronic acid hydrophilic surface rehabilitating curcumin nanocrystals for targeted breast cancer treatment with prolonged biodistribution, 462-472. Copyright 2020, with permission from The Royal Society of Chemistry.¹⁶⁷

curcuminoids, micronized powder and Cur-loaded micelles at a dose of 500 mg, respectively.¹⁸² The results showed that the AUC of micronized curcumin was 14-, 5-, and 9-fold than curcumin in women, men and all subjects. Furthermore, curcumin micelles improved the oral bioavailability of Cur at 277-, 114- and 185-fold, respectively. In another study, Kanai et al evaluated the safety and pharmacokinetics of THERACURMIN, a novel nanoparticle curcumin.¹⁸³ THERACURMIN increased the curcumin concentration in plasma in a dose-dependent manner. In addition, no toxicities of curcumin nanoformulations were observed in this study.

Although some preclinical researches showed potent anticancer effects of curcumin nanoformulations, clinical findings have not been conclusive. But several clinical trials were conducted to investigate the reduced adverse effects caused by curcumin nanoformulations in cancer treatment. Meriva®, a nano delivery system composed of curcuminoids and phosphatidylcholine, has exhibited a boosted pharmacokinetic profile in both rats and human.¹⁸⁴ In a clinical trial, patients with histologically documented solid tumors were randomized to receive either Meriva® or placebo for 8 weeks. It was found that Meriva[®] significantly improved the health-related quality of life (QoL) compared with the placebo. Moreover, some markers associated with systemic inflammation including TNF-α, TGF-β, IL-6, substance P, calcitonin gene-related peptide (CGRP), high-sensitivity C-reactive protein (hs-CRP) and monocyte chemotactic protein-1 (MCP-1) were significantly decreased upon the treatment of curcumin nanoformulations. The results indicated that adjuvant therapy with curcumin nanoformulations can improve quality of life and suppress systemic inflammation in patients with solid tumors who are under treatment with standard chemotherapy protocols. Furthermore, a similar study showed that Meriva® could decrease the side effects after chemo- and radiotherapy.¹⁸⁵ In another clinical trial, nano-curcumin was combined with ω -3 fatty acids to treat migraines.¹⁸⁶ Eighty patients were divided into four groups and given the treatment of different drug formulations. It has been reported that TNF- α plays an important role in neuroimmunity pathogenesis of migraine. The combination of nano-curcumin and ω -3 fatty acids significantly decreased the expression of TNF- α in serum while nanocurcumin or ω -3 fatty acids alone did not exhibit a significant reduction in the levels of TNF- α . This combinational therapy represented a promising approach for migraine management.

Conclusions and Future Perspectives

Curcumin is a natural polyphenolic compound that has been widely used in traditional Chinese medicine. The main pharmacological activities of CUR are anti-inflammatory, antioxidation, anti-tumor, anti-bacterial, liver protection, etc. However, the poor solubility and low bioavailability of Curcumin result in poor intestinal absorption, rapid metabolism in plasma and liver, and fast systemic clearance. Thus, the in vivo activities of Cur are limited. Nanoparticles have shown many advantages as drug carriers, including increased drug stability, slowed drug degradation, enhanced drug solubility, and improved pharmacokinetics. Several types of Curloaded nanoparticles have been developed for the treatment of different diseases. As we discussed in this review, nanoformulations not only increase the solubility and bioavailability but also improve the therapeutic effect of curcumin. As a research focus, the anti-tumor efficacy of CUR has been widely studied. The preclinical and clinical trials of curcumin-loaded nanoformulations that have been carried out indicated that curcumin nanoparticles might act as enhanced

Disease	Cur-Loaded Nanoformulation	Animal	Major Outcome	References
Cancer	MPEG-PCL micelles	BALB/c mice	Sustained drug release Enhanced anti-cancer efficacy Improved anti-metastasis activity	[51]
Brain injury	PLGA nanoparticles	Sprague Dawley rats	Down-regulated expression of NF-ĸB Decreased early brain injury induced by subarachnoid hemorrhage	[168]
	PLGA nanoparticles	Sprague Dawley rats	Improved neurological deficit Attenuated brain edema Enhanced therapeutic potential against subarachnoid hemorrhage- induced blood-brain barrier disruption	[169]
Wound healing	Dextran hydrogel incorporated PEG–PLA micelles	BALB/c mice	Acceptable biocompatibility Accelerated angiogenesis, fibroblast accumulation and wound healing	[170]
	PVA/Chitosan/ Curcumin patches	Albino wistar rats	Increased bioavailability Improved anti-bacterial activity Enhanced wound healing	[171]
Malaria	PLGA nanoparticles	C57BL/6 mice	Enhanced and delayed uptake in the brain. Preventing degenerative changes in cerebral malaria	[172]
	PLGA nanoparticles	Swiss male albino mice	Higher potency against malaria parasite Higher safety	[173]
Asthma	Solid lipid nanoparticles	BALB/c mice Sprague Dawley rats	Enhanced bioavailability Increased tissue concentrations Suppressed airway hyperresponsiveness and inflammatory cell infiltration	[174]
Psoriasis	Hydrogel coated PLGA nanoparticles	C57/BL6 mice	Enhanced topical penetration and system exposure Improved anti-psoriasis activity	[175]
Arteriosclerosis	MPEG-PCL micelles	ApoE ^{-/-} mice	Inhibition of intraplaque neovascularization Reduced matrix metalloproteinases 2/9 activity and inflammatory response Regulation of lipoprotein cholesterol metabolism	[176]
Leishmaniasis	Chitosan nanoparticles	Male albino rats	Satisfactory loading efficiency, stability and drug release characteristics Higher anti-leishmaniasis efficiency	[177]
Bacterial infection	PVP nanoparticles	C57BL/6 mice	Improved anti-bacterial and anti-inflammatory effects Enhanced therapeutic potency of vancomycin	[178]

chemotherapeutic and chemopreventive agents. Moreover, by down-regulating the expression of P-glycoprotein (P-gp), multidrug resistance protein (MRP), breast cancer resistance protein (BCRP) and other drug efflux transporter proteins, CUR can reverse the resistance of common chemotherapeutic drugs. Therefore, combination of curcumin and other anticancer drugs using nano delivery systems is a promising therapeutic strategy.

However, there are still some problems that need to be solved. The foremost is the toxicity of nanocarriers. Although in most cases materials used to prepare the carriers are biodegradable, biocompatible and non-toxic, some reagents added during the preparation process, such as solvents and surfactants, may cause some damage to the human body. Moreover, the lack of tissue specificity is also a big problem. In tumor treatment, nanoparticles not only deliver drugs to cancer cells, but also to normal tissues. Therefore, targeted nanoparticles should be investigated. Additionally, small and large cohorts, as well as patients in Phase I/II clinical trials are still needed to evaluate the pharmacological and toxicological effects of Curloaded nanoparticles. Various curcumin nanoformulations are still at the stage of laboratory research, and there are some problems in large-scale preparation. Since the preparation of most nanoparticles is relatively complicated and difficult to scale up, we also need to make breakthrough progress in processes and technologies of large-scale production, stability, and quality control of drug loading.

In summary, nanoformulations improve the solubility and biocompatibility, increase the therapeutic effect, and lay the foundation for further clinical research and applications of curcumin.

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Disclosure

The authors have no conflicts of interest to declare.

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