

Vaginal Microbiota and HPV Infection: Novel Mechanistic Insights and Therapeutic Strategies

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Yuanyue Li^{1,*}

Tao Yu^{1,*}

Huang Yan²

Duanduan Li²

Tang Yu²

Tao Yuan¹

Abdul Rahaman³

Shahid Ali³

Farhat Abbas²

Ziqin Dian⁴

Xiaomei Wu¹

Zulqarnain Baloch⁵

¹Department of Gynaecology, The First People's Hospital of Yunnan Province, Kunming 650032, Yunnan, People's Republic of China; ²South China Agricultural University, Guangzhou 510642, People's Republic of China;

³School of Food Science and Engineering, South China University of Technology, Guangzhou, People's Republic of China;

⁴Department of Clinical Laboratory, The First People's Hospital of Yunnan Province, Kunming, Yunnan 650032, People's Republic of China; ⁵Biomedical Research Center, Northwest Minzu University, Lanzhou 730030, People's Republic of China

*These authors contributed equally to this work

Abstract: Cervical cancer is a global public health concern. The complex interaction of genetic and environmental factors is critical for the progress of cervical cancer. Growing evidence suggests that microbes, human papillomavirus (HPV), and the immune system interact closely with each other to govern homeostasis of the vaginal environment and the health of the lower genital tract of females. Certain vaginal microbial strains may play either a protective or a pathogenic role in carcinogenesis of the cervix after HPV persistent infection. Probiotics can therefore present a putative therapeutic approach for cervical cancer. However, work in this field remains limited. Recent technological developments have allowed us to identify microbes and their products using culture-independent molecular detection techniques. In this review, we discuss the composition of the vaginal bacterial community, its commensal flora and the protective impact this has on the health of the female genital tract. This review will also describe critical immune factors in lower genital tract health and summarize the role of the vaginal microbiota in cervical carcinogenesis. Knowledge in this field has provided researchers with the clues and tools to propose the use of probiotics as a potential line of treatment for cervical cancer and has provided valuable insights into host–pathogen interaction dynamics within the female genital tract.

Keywords: vaginal microbiota, human papillomavirus, cervical cancer, immune system, probiotic therapy

Introduction

Cervical cancer is a global public health issue, having an estimated 530,000 annual cases and 270,000 deaths according to the World Health Organization.¹ Development of cervical cancer is a stepwise process by which localized cervical intra-epithelial neoplasia develops in the cervix and progresses into invasive and metastatic carcinoma forms. Human papillomavirus (HPV) is the major cause of cervical squamous cell carcinoma, its precursor lesions (cervical intraepithelial neoplasia), and several other benign and malignant clinical manifestations. HPVs are a family of circular double-stranded DNA viruses, 396 different subtypes of which have been reported.² Genital HPVs, subdivided into high- and low-risk types, are frequently associated with invasive cervical cancers.³ High-risk HPVs (HR-HPVs) are the major drivers of cervical carcinogenesis with integrated HPV DNA present in almost all cervical cancer biopsies. Major HPV oncogenic proteins, particularly E6 and E7, are involved in cervical cancer transformation, immortalization, and malignancy.^{4,5} The human vagina harbors affluent microorganisms that play essential roles in human health and disease.⁶ Work to date supports the notion that the vaginal microbiota could play a role in HPV acquisition and persistence in

Correspondence: Zulqarnain Baloch
Biomedical Research Center, Northwest
Minzu University, Lanzhou 730030,
People's Republic of China
Tel +86 18344564625
Email znbalooch@yahoo.com

cervical cancers.^{7–15} In this review, we discuss the composition, function and adaptive mechanisms of the vaginal microbiome in healthy females. We then discuss the impact of the vaginal microbiota on HPV infection acquisition and persistence, potential immune factors related to cervical carcinogenesis, and explore the potential use of probiotic therapy for the control of HPV infection in the cervix.

Vaginal Microbiota

Over time, humans have coexisted with complicated bacterial communities, which are unique to specific niches.¹⁶ The vaginal tract is colonized by various microorganisms forming the vaginal microbiota. In the past, the bacteria colonizing the vaginal tract were preliminarily isolated and confirmed by traditional bacterial culturing and biochemical identification methods. The composition of the vaginal microbiome remained obscure because the fraction of microbes failed to be cultured *ex vivo* until now. Present-day techniques, such as next-generation sequencing (NGS) and shotgun metagenomics sequencing, have helped us define the complex vaginal microbiota.¹⁷

The dominant microbial genus group found in healthy vaginal bacterial communities in women of childbearing age is *Lactobacillus*, commonly regarded as the first line of defense against pathogenic agents. This is different to the majority of body sites; to date, a variety of microbial communities is generally considered a signature of health. Ravel et al¹⁸ were the first to define the composition of the vaginal microbiota using bacterial 16S RNA sequencing technology. 16S RNA is the component of the 30S small subunit of a prokaryotic ribosome. 16S rRNA gene sequencing is highly useful in regard to bacterial classification; it has low phylogenetic power at the species level and poor discriminatory power for some genera. The genes coding for it are referred to as 16S rRNA genes and are used in reconstructing phylogenies due to the slow rates of evolution of this region of the gene. They defined the vaginal microbiota in to five community state types (CSTs) (I, II, III, IV, and V) based on the presence or absence of a particular *Lactobacillus* spp. CST I, II, III, and V microbiota belong to *L. crispatus*, *L. gasseri*, *L. iners*, and *L. jensenii*, respectively. CSTI V is heterogeneous on account of depletion of *Lactobacillus* spp. and the presence of strictly anaerobic species such as *Prevotella*, *Sneathia*, *Megasphaera*, and *Gardnerella*. Longitudinal studies found that the diversity and abundance of microbes within subjects vary widely both within and among subjects in the

vagina over time, which also presented superior longitudinal stability than other mucosal sites.⁶

The genus *Lactobacillus* is taxonomically complex and is composed of more than 170 species, which cannot be differentiated phenotypically by traditional molecular identification methods.¹⁹ *L. crispatus*, *L. gasseri*, *L. jensenii*, *L. iners*, and *L. vaginalis* are commonly populated in the vagina. *Lactobacillus* spp. are able to produce numerous protective peptides and metabolic products, such as lactic acid and other acidic compounds that are capable of inhibiting pathogenic bacteria adhesion and growth. In addition, bacteriocin-like compounds, such as bio-surfactants and hydrogen peroxide, may be key for *Lactobacillus* spp. persistence in the vagina.²⁰ These could explain why vaginal epithelial cells from healthy females are lightly colonized by *Lactobacillus* spp., as these metabolites counter the adhesion and growth of other bacteria.²¹ It also controls bacteria which ascend the cervix to the uterus and enhance sperm motility.²² Studies of chronic wounds showed that *Lactobacillus* spp. supernatants contain antimicrobials (mevalonolactone, benzoic acid, and 5-methyl-hydantoin), surfactants (distearin, dipalmitin, and 1,5-monolinolein), anesthetics (barbituric acid derivatives), and autoinducer type 2 precursors (4,5-dihydroxy-2,3-pentanedione and 2-methyl-2,3,3,4-tetrahydroxytetrahydrofuran). In addition to lactic acid, *Lactobacillus* spp. supernatant also contains phenolics, hydrogen peroxide, sodium, calcium, potassium, magnesium, and DNAases.²³ In all, *Lactobacillus* spp. play a protective function in maintaining vaginal health. As microbes struggle for nutrients and adherence to the vaginal epithelium, the local immune system may activate the production of host antibacterial substances to further control microbial growth.²⁴ Furthermore, a high lactobacilli population is crucial because it protects the host vaginal epithelia against sexually transmitted infections (STIs). Therefore, a complex interaction and synergy between host and microorganism to maintain the stability in the vaginal epithelia exists. A high population of *Lactobacillus* spp. has been reported in healthy vagina followed by other microorganisms²⁵ such as *Gardnerella*, *Candida*, *Prevotella*, *Ureaplasma*, *Peptostreptococcus*, *Mycoplasma*, *Streptococcus*, *Corynebacterium*, *Clostridium*, *Staphylococcus*, *Bacteroides*, *Enterococcus*, *Bifidobacterium*, *Veillonella*, and *Escherichia*. In short, the vaginal microbial population continually changes throughout life, but it may be consistent and stable in a certain period of time in healthy women. However,

when there is a disorder in the microbial flora, such as the loss of *Lactobacillus* spp. resulting in the overgrowth of anaerobic bacteria as in the case of bacterial vaginitis (BV), this is associated with the higher incidence, prevalence, and persistence of HPV infection, which can contribute to the development of cervical intraepithelial neoplasias.^{26–28}

Local Immunity in the Vaginal Microbiota

The vaginal mucosal barrier possesses specialized features to protect against infectious and noxious environmental damage, at the same time maintaining symbiotic mutualism with commensal microbes. Microbes, immune regulatory actions, and host genes interact closely to govern the homeostasis of the vaginal environment.²⁹ The cervicovaginal mucosa is a critical site for protection against urogenital pathogens. The female genital tract consists of two types of mucosal surface. The lower genital tract (vagina and ectocervix) represents the type II mucosal surface, lined by stratified squamous epithelia. Defining features include the presence of mucus-secreting cells and the expression of polymeric immune-globulin R (pIgR) on the basolateral surface of the epithelia. pIgR binds to polymeric IgA (pIgA) secreted by plasma cells and exports pIgA transepithelially.³⁰ Plasma cells also release secretory IgA (SIgA) into the lumen. In contrast, the upper genital tract (endocervix and endometrium) represents the type I mucosal surface, which is covered by a monolayer of columnar epithelial cells with tight junctions and SIgA present in the microenvironment. The boundary between type I and type II mucosa, also known as the cervical transformation zone, is the most vulnerable region invaded by pathogens and heavily populated with T cells and antigen-presenting cells (APCs).³¹

In the human vaginal mucosa, four unique myeloid-derived APC subsets have been characterized including, cervicovaginal langerhans cells (cvLCs), CD14[−] dendritic cells (DCs), CD14⁺, DCs, and CD14⁺ macrophages.³² Langerhans cells are also found in the epidermis (epLCs) and express the C-type lectin molecule. cvLCs are found in the human cervicovaginal epithelium, whereas CD14[−] DCs, CD14⁺, DCs, and CD14⁺ macrophages are found in the subepithelial lamina propria region.³² Transcriptional profiling showed that CD14[−] cells (cvLCs and CD14[−] DCs) displayed Th₂-inducing and regulatory properties, whereas the CD14⁺ APCs (CD14⁺ DCs and macrophages) exhibited

signatures of innate immune defense and pro-inflammatory responses.³³ Resident memory T cells in the vaginal mucosa mediate rapid protection upon infection. CD4⁺ T-cell entry into the cervicovaginal region facilitates the entry of circulating memory CD8⁺ T cells via the production of IFN- γ . Innate immune sensing and surveillance of commensal and pathogenic microbes in the female genital tract is achieved by microbial motif pattern recognition via pattern recognition receptors (PRRs), such as toll-like receptors,³⁴ dectin-1 receptor, and nucleotide-binding oligomerization domain receptors present in and on both squamous epithelial cells (Ref). PRR stimulation in the vaginal environment initiates various cytokine and chemokine signaling cascades, including secretion of interleukin (IL)-1 β , IL-6, IL-8, and tumor necrosis factor- α (TNF- α), to activate or recruit macrophages, natural killer cells, CD4⁺ and CD8⁺ T-cell lymphocytes, and B lymphocytes.²⁹ Other factors that strengthen vaginal defense system include IgA, IgG, vaginal antimicrobial peptides (AMPs) and mannose binding lectin (MBL). *N*-acetyl glucosamine and MBL bind mannose, which is present on the cell surface of microbes.³⁵ IgG and IgA may assist to avert vaginal epithelial cell adherence and uptake, playing a role in neutralizing and clearing infectious microbes from host sites.³⁶ Similarly, there are various classes of vaginal AMPs which normally engage immune cells via chemotaxis or possess anti-endotoxin activity. Mechanisms for AMP have been reported in detail previously.³⁷ However, the detailed relationship among AMPs and vaginal microbiota remains understudied. AMPs, such as defensins, are small cysteine-rich cationic proteins that have varied mechanisms of action against common vaginal microbes, including HPV (Ref). Other host-microbiota interactions occur at the epithelial cell interface; many of them are not well understood. For example, bacterial strains produce a range of metabolites, some of which could cross the epithelial barrier and affect cellular function in other sites, which are influenced by host factors such as hormones and mucins.³⁸

HPV and the Microbiota

It is well established that cervical cancer development is the final stage of a sequence of cellular and molecular changes instigated with HPV infection. Host immune response and evasion mechanisms involve a termination host response against tumor antigens. In order to know the local and systemic modifications during interactions between immune system and HPV-related cervical lesions progress to cancer it has already been reported that

elevated cervical leukocytes in cervical cancer patients infiltrate and imitate with increase of M2 macrophages, neutrophils and T lymphocytes. Additionally, a sturdy negative correlation among incidence of T cells, neutrophils, and cancer patients' samples has been reported. Similarly, neutrophils stop T-cell activity and show extended viability and CD16 expression in 3D tumor cell cultures compared to normal neutrophil culture. They also found a high rate of immature low-density neutrophils, elevated plasma concentration and tolerogenic monocyte-based dendritic cells in cancer cases compared to controls. These results showed that G-CSF and neutrophils might be a piece of the immune escape mechanism locally and systemically controlled by cervical cancer cells.³⁹

Women diagnosed with CST IV-like condition showed substantially increases in FMS-like tyrosine kinase 3 ligand, TNF- α , IL-1 α , IFN- γ , IL-1 β , IL-4, IL-12p70, IL-10, and IL-8, and then those of CST I. Additionally, compared with CST I, CST III showed significantly elevated IFN- γ . Recently, it has been shown that there is significant TNF- α , IL-1 β and IL-1 α longitudinally in cases which developed from a CST I, CST III and CST III to a CST IV.^{40,41} On the contrary, mock communities normally dominated by *L. jensenii* (CST V) and *L. crispatus* (CST I) on restructured 3D vaginal epithelial models did not sturdily evoke cytokine IL-8 or IL-1 β secretion compared with controls.⁴⁰ All observations persistently support the concept that the innate immune response is mainly operated with vaginal microbiota.

HPV infection associated with the vaginal microbiota. Cervical carcinogenesis related with the vaginal microbiota immune response in cervix. Hence, overall, microbes, environments, immune regulatory actions, and gene expression interact closely to govern the homeostasis of the vaginal environment.

Probiotic Therapy

According to the World Health Organization and the United Nations' Food and Agriculture Organization, probiotics are defined as "live microorganisms, which when administered in adequate amounts confer a health benefit on the host".⁴² Probiotics comprise living bacteria (eg, *Bifidobacteria* species, *Lactobacilli*, and *Streptococci*) that can modulate the composition of microbiota in an attempt to improve the health, immune system, inflammatory state, and bioavailability of micronutrients. They also reduce the risks of diabetes, and ameliorate dyslipidemia, allergic disorders, and certain cancers.⁴³ The mechanism by which probiotics confer their conducive

functions may be elaborated by alterations in pH, inhibition of pathogens (via the generation of antibacterial compounds, competitive elimination of pathogens in receptor binding sites, and contention for available nutrients), mutagenic, carcinogenic production, and maintenance of the mucosal barrier.⁴⁴

There is a potential to maximize a woman's first line of defense by understanding the vaginal microbiota in depth, which will ultimately help to develop practical and low-cost therapeutics to decrease the susceptibility of STIs, particularly HPV. According to the literature, fifty-four HPV-positive women diagnosed with low-grade squamous intraepithelial lesions were offered a probiotic containing *Lactobacillus casei* on a daily basis and interestingly *Lactobacillus casei* appeared to be helpful in HPV clearance.⁴⁵ In HIV-infected individuals, due to the impairment of the immune response and the loss of gut intraepithelial lymphocytes, lower rates of HPV clearance are observed and consequently spontaneous resolution of multiple large anal condylomas is rare among this population. For this reason, it has been suggested that direct efficacy of oral and rectal multi-strain probiotic administration in the treatment of anal condylomatosis in HIV-infected patients was effective.⁴⁶ In addition, it has been reported that appropriate probiotics would be most useful for women's health. It will be exciting and valuable to establish whether probiotics have the potential to reduce productive viral infection and lead to a clinically "latent" HPV stage.

In the last century, different research groups worked on urogenital lactobacilli based upon their global prevalence in the vagina. In 2002, it was found that fastidious *L. iners* is a highly populated microbe in the healthy vagina.^{47–49} In a clinical study, they successfully combined different microbial strains such as RC-14 with GR-1 and administered these orally to interfere with a range of urogenital pathogens to restore the host's own lactobacilli.⁵⁰ The idea of oral administration of probiotics came from the knowledge that many urogenital infections arise from the entrance of a pathogen from the rectum to the perineal skin and then the vagina. Therefore, if the other pathogens could do it, why could not lactobacilli? Different studies have already verified that orally administered lactobacilli can reach the vagina.^{51–54}

Probiotic lactobacilli have been frequently prescribed against urogynecologic infections. Although clinical data results show poor support, probiotic application effectively cures urogenital infections compared with other treatments. Additionally, probiotic application has no serious adverse

side effects. However, well-designed clinical trials of probiotics for curing urogenital infections are needed. Moreover, co-treatments suggested that probiotic and antibiotic application should be used at least 2 to 4 hours apart to prevent the live microorganisms' destruction in the gastrointestinal tract.⁵⁵ Thus, probiotics used for curing genitourinary infections contain *Lactobacillus* species, as the objective is the vaginal microflora. The mechanisms of action of probiotics include acidification of the mucosal surface, prevention of the adherence of pathogens, production of substances such as vitamins and immune modulators, and synergistic action with the host immune system.

It has also been suggested that the microbiome and radiation provoked enteritis. Radiation therapy, either alone or in combination with chemotherapy or surgery, is the standard treatment option for a clinician to treat advanced cervical cancer cases. Unluckily, radiation therapy can affect the bowel as a side effect. According to investigation, almost 80% of cases showed bowel symptoms such as urgency, abdominal pain, bloating and diarrhea during radiation treatment.⁵⁶ This dysbiosis and local inflammatory condition could contribute to cause sexual dysfunction and pelvic pain in cervical cancer cases. However, limited data have been reported on this topic, so additional research should be conducted in this area.⁵⁷

Together with the primary treatment, they are able to restore depleted microflora. Therefore, vaginal probiotics might be valuable for this purpose, and oral probiotics may be useful to conserve normal vaginal microflora and help preserve normal vaginal flora during antibiotic therapy.⁵⁰ Direct use of vaginal probiotics such as *Lactobacillus* species have been effectively used,⁵⁸ together with a reduction in BV.⁵⁹ Probiotics might have substantially higher effects on carcinogenesis. It has already been recommended that regular intake of oral probiotics can reduce the threat of gastrointestinal cancer.⁶⁰ We suggested that vaginal probiotics work simultaneously to decrease HPV infection rate and escalate HPV infection clearance. Moreover, during treatment, the microbiome can modulate the tumor microenvironment.⁶¹ Data suggested that CpG oligonucleotide or platinum chemotherapy regimens had better efficiency against tumors with regular gut micro-biomes as compared to antibiotic-treated mice.⁶² Further, the presence of microorganisms (*Bacteroides taiotaomicon* or *B. fragilis*) was required for CTLA-4 targeted immunotherapy.⁶³ Clinicians are looking to treat gynecological cancer using this immunotherapy, such that vaginal microbiomes could help influence the efficiency.^{64,65} Probiotics prevent sarcoma

by reducing the level of gut inflammatory (Th17) cells and promoting anti-inflammatory (Treg) cells in mice.^{63,64} The promotion of Treg cells leads to a reduction in gynecological cancers and chronic inflammation. The probiotic can change the tumor microenvironment by direct application to the gynecological tumor. Furthermore, in-vitro studies suggested that vaginal microbiota may increase the apoptosis of cancer cells or may provoke the anti-inflammatory cells like dendritic, Treg and cytokines.⁶⁶⁻⁶⁹

The lack of efficient treatment to counter antibiotic resistance to bacterial infections and the intravaginal application of lactobacilli and lactoferrin may be the narrative and capable therapeutic approach to reinstate the mucosal immune homeostasis.²⁵

Concluding Remarks

There are many imminent approaches that can be used to evaluate intestinal and related mucosal regions, but they require concrete capabilities that traverse mucosal immunology and reproductive biology. Modern technology like single-cell RNA sequencing, mass cytometry, deep sequencing of bacterial 16S rRNA, and internal transcribed spacer regions of fungal rRNA genes, metagenomics, and metabolomics play an important role in immunology and reproductive health. These may find unprecedented purposes. Human vaginal hosts possess millions of microorganisms, also collectively known as the bacterial flora. An increasing number of studies are progressively unraveling the fascinating interaction between hosts and microbiota. Probiotics can confer health by modulating the composition of gut microbiota and restoring the physiological bacterial flora. That being said, limited data are available in this area, and the relevant scientific work is still at the early stage. A lot of studies need to be carried out in the future, as below. Initially, a host provides a large and complex environment for gut bacterial flora. It is unclear yet whether the alteration of intestinal microbiota contributes to the development of eczema or its presence reflects the primary cause of changes in gut microbiota. There is need for a profound understanding of the interactive mechanism between vaginal microbiota and host. As such, it is critical to offer an entirely novel approach to cure diseases by monitoring and controlling vaginal microbiota. Compared with developing novel drugs for inflammation, it might cost less to seek novel approaches, such as monitoring and manipulating human vaginal microbiota if required using probiotics. The opportunity exists to develop updated probiotics in accordance with the interaction between specific microbiota and HPV infection. In addition, it is plausible to develop

probiotics from vaginal microbiota in healthy groups. Similarly, it is significant to determine how probiotics change the constitution of the vaginal microbiota (reconstructing bacterial flora) and how relevant it is to HPV infection. Given little relevant research in the area, more studies should be carried out to elucidate the interaction between specific microbiota and cervical carcinogenesis related with HPV infection and a wealth of well-controlled clinical studies on vaginal microbiota are required to ensure safety in patients. Synergistic efforts in vivo and in vitro are also required so as to advance our knowledge in the area, which is promising for its potential in biomarker reorganization and novel therapeutic targets. All in all, further understanding of the vaginal microbiota may uncover more mysteries of HPV persistent infection and related diseases and provide a promising therapeutic target.

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Author Contributions

All authors contributed to data analysis, drafting and revising the article, gave final approval of the version to be published, and agreed to be accountable for all aspects of the work.

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References

- Small W Jr., Bacon MA, Bajaj A, et al. Cervical cancer: a global health crisis. *Cancer*. 2017;123(13):2404–2412. doi:10.1002/cncr.v123.13
- Bzhalava D, Muhr LS, Lagheden C, et al. Deep sequencing extends the diversity of human papillomaviruses in human skin. *Sci Rep*. 2014;4:5807. doi:10.1038/srep05807
- N M, Bosch FX, de Sanjosé S, et al. International agency for research on cancer multicenter cervical cancer study group.: epidemiologic classification of human papillomavirus types associated with cervical cancer. *N Engl J Med*. 2003;348(6):10. doi:10.1056/NEJMicm020037
- Zur Hausen H. Papillomaviruses and cancer: from basic studies to clinical application. *Nat Rev Cancer*. 2002;2(5):342–350. doi:10.1038/nrc798
- Wilting SM, Steenbergen RDM. Molecular events leading to HPV-induced high grade neoplasia. *Papillomavirus Res*. 2016;2:85–88. doi:10.1016/j.pvr.2016.04.003
- Consortium TH. Project M. Structure, function and diversity of the healthy human microbiome. *Nature*. 2012;486(7402):207–214.
- Mitra A, MacIntyre DA, Marchesi JR, Lee YS, Bennett PR, Kyrgiou M. The vaginal microbiota, human papillomavirus infection and cervical intraepithelial neoplasia: what do we know and where are we going next? *Microbiome*. 2016;4(1):58. doi:10.1186/s40168-016-0203-0
- Seo SS, Oh HY, Lee JK, Kong JS, Lee DO, Kim MK. Combined effect of diet and cervical microbiome on the risk of cervical intraepithelial neoplasia. *Clin Nutr*. 2016;35(6):1434–1441. doi:10.1016/j.clnu.2016.03.019
- Audirac-Chalifour A, Torres-Poveda K, Bahena-Roman M, et al. Cervical microbiome and cytokine profile at various stages of cervical cancer: a pilot study. *PLoS One*. 2016;11(4):e0153274. doi:10.1371/journal.pone.0153274
- Oh HY, Kim BS, Seo SS, et al. The association of uterine cervical microbiota with an increased risk for cervical intraepithelial neoplasia in Korea. *Clin Microbiol Infect*. 2015;21(7):674e671–679. doi:10.1016/j.cmi.2015.02.026
- Mitra A, MacIntyre DA, Lee YS, et al. Cervical intraepithelial neoplasia disease progression is associated with increased vaginal microbiome diversity. *Sci Rep*. 2015;5:16865. doi:10.1038/srep16865
- Brotman RM, Shardell MD, Gajer P, et al. Interplay between the temporal dynamics of the vaginal microbiota and human papillomavirus detection. *J Infect Dis*. 2014;210(11):1723–1733. doi:10.1093/infdis/jiu330
- Nardis C, Mosca L, Mastromarino P. Vaginal microbiota and viral sexually transmitted diseases. *Ann Ig*. 2013;25(5):14.
- Lee JE, Lee S, Lee H, et al. Association of the vaginal microbiota with human papillomavirus infection in a Korean twin cohort. *PLoS One*. 2013;8(5):e63514. doi:10.1371/journal.pone.0063514
- Brotman RM. Vaginal microbiome and sexually transmitted infections: an epidemiologic perspective. *J Clin Invest*. 2011;121(12):4610–4617. doi:10.1172/JCI57172
- Martin DH. The microbiota of the vagina and its influence on women's health and disease. *Am J Med Sci*. 2012;343(1):2–9. doi:10.1097/MAJ.0b013e31823ea228
- Lamont RF, Sobel JD, Akins RA, et al. The vaginal microbiome: new information about genital tract flora using molecular based techniques. *BJOG*. 2011;118(5):533–549. doi:10.1111/j.1471-0528.2010.02840.x
- Ravel JGP, Abdo Z, Schneider GM, et al. Vaginal microbiome of reproductive-age women. *Proc Natl Acad Sci U S A*. 2011;108(Suppl 1):8. doi:10.1073/pnas.1002611107
- Goldstein EJ, Tyrrell KL, Citron DM. Lactobacillus species: taxonomic complexity and controversial susceptibilities. *Clin Infect Dis*. 2015;60(Suppl 2):S98–107. doi:10.1093/cid/civ072
- Reid G, Younes JA, Van der Mei HC, Gloor GB, Knight R, Busscher HJ. Microbiota restoration: natural and supplemented recovery of human microbial communities. *Nat Rev Microbiol*. 2011;9(1):27–38. doi:10.1038/nrmicro2473
- Swidsinski A, Mendling W, Loening-Baucke V, et al. Adherent biofilms in bacterial vaginosis. *Obstet Gynecol*. 2005;106:11. doi:10.1097/01.AOG.0000183594.45524.d2
- Barbonetti A, Cinque B, Vassallo MR, et al. Effect of vaginal probiotic lactobacilli on in vitro-induced sperm lipid peroxidation and its impact on sperm motility and viability. *Fertil Steril*. 2011;95(8):2485–2488. doi:10.1016/j.fertnstert.2011.03.066
- Ramos AN, Sesto Cabral ME, Arena ME, Arrighi CF, Arroyo Aguilar AA, Valdez JC. Compounds from lactobacillus plantarum culture supernatants with potential pro-healing and anti-pathogenic properties in skin chronic wounds. *Pharm Biol*. 2015;53(3):350–358. doi:10.3109/13880209.2014.920037

24. Aroutcheva A, Gariti D, Simon M, et al. Defense factors of vaginal lactobacilli. *Am J Obstet Gynecol*. 2001;185(2):375–379. doi:10.1067/mob.2001.115867
25. Valenti P, Rosa L, Capobianco D, et al. Role of lactobacilli and lactoferrin in the mucosal cervicovaginal defense. *Front Immunol*. 2018;9:376. doi:10.3389/fimmu.2018.00376
26. Gillet E, Meys JF, Verstraeten H, et al. Bacterial vaginosis is associated with uterine cervical human papillomavirus infection: a meta-analysis. *BMC Infect Dis*. 2011;11:10. doi:10.1186/1471-2334-11-10
27. Gillet E, Meys JF, Verstraeten H, et al. Association between bacterial vaginosis and cervical intraepithelial neoplasia: systematic review and meta-analysis. *PLoS One*. 2012;7(10):e45201. doi:10.1371/journal.pone.0045201
28. Guo YL, You K, Qiao J, Zhao YM, Geng L. Bacterial vaginosis is conducive to the persistence of HPV infection. *Int J STD AIDS*. 2012;23(8):581–584. doi:10.1258/ijsa.2012.011342
29. Smith SB, Ravel J. The vaginal microbiota, host defence and reproductive physiology. *J Physiol*. 2017;595(2):451–463. doi:10.1113/tjp.2017.595.issue-2
30. Iwasaki A. Mucosal dendritic cells. *Annu Rev Immunol*. 2007;25:381–418. doi:10.1146/annurev.immunol.25.022106.141634
31. Kumamoto Y, Iwasaki A. Unique features of antiviral immune system of the vaginal mucosa. *Curr Opin Immunol*. 2012;24(4):411–416. doi:10.1016/j.coi.2012.05.006
32. Duluc D, Gannevat J, Anguiano E, et al. Functional diversity of human vaginal APC subsets in directing T-cell responses. *Mucosal Immunol*. 2013;6(3):626–638. doi:10.1038/mi.2012.104
33. Duluc D, Banchereau R, Gannevat J, et al. Transcriptional fingerprints of antigen-presenting cell subsets in the human vaginal mucosa and skin reflect tissue-specific immune microenvironments. *Genome Med*. 2014;6(11):98. doi:10.1186/s13073-014-0098-y
34. Kang JY, Lee JO. Structural biology of the toll-like receptor family. *Annu Rev Biochem*. 2011;80:917–941. doi:10.1146/annurev-biochem-052909-141507
35. Neth O, Jack DL, Dodds AW, Holzel H, Klein NJ, Turner MW. Mannose-binding lectin binds to a range of clinically relevant microorganisms and promotes complement deposition. *Infect Immun*. 2000;68(2):688–693. doi:10.1128/IAI.68.2.688-693.2000
36. Wang YY, Kannan A, Nunn KL, et al. IgG in cervicovaginal mucus traps HSV and prevents vaginal herpes infections. *Mucosal Immunol*. 2014;7(5):1036–1044. doi:10.1038/mi.2013.120
37. Wilson SS, Wiens ME, Smith JG. Antiviral mechanisms of human defensins. *J Mol Biol*. 2013;425(24):4965–4980. doi:10.1016/j.jmb.2013.09.038
38. Reid G. Cervicovaginal microbiomes threats and possibilities. *Trends Endocrinol Metab*. 2016;27(7):9. doi:10.1016/j.tem.2016.04.004
39. Alvarez KLF, Beldi M, Sarmanho F, et al. Local and systemic immunomodulatory mechanisms triggered by human papillomavirus transformed cells: a potential role for G-CSF and neutrophils. *Sci Rep*. 2017;7(1). doi:10.1038/s41598-017-09079-3
40. Rose WA 2nd, McGowin CL, Spagnuolo RA, Eaves-Pyles TD, Popov VL, Pyles RB. Commensal bacteria modulate innate immune responses of vaginal epithelial cell multilayer cultures. *PLoS One*. 2012;7(3):e32728. doi:10.1371/journal.pone.0032728
41. Anahtar MN, Byrne EH, Doherty KE, et al. Cervicovaginal bacteria are a major modulator of host inflammatory responses in the female genital tract. *Immunity*. 2015;42(5):965–976. doi:10.1016/j.immuni.2015.04.019
42. Ebner S, Smug LN, Kneifel W, Salminen SJ, Sanders ME. Probiotics in dietary guidelines and clinical recommendations outside the European Union. *World J Gastroenterol*. 2014;20(43):16095–16100. doi:10.3748/wjg.v20.i43.16095
43. Pan W, Kang Y. Gut microbiota and chronic kidney disease: implications for novel mechanistic insights and therapeutic strategies. *Int Urol Nephrol*. 2018;50(2):289–299. doi:10.1007/s11255-017-1689-5
44. Behnsen J, Deriu E, Sassone-Corsi M, Raffatellu M. Probiotics: properties, examples, and specific applications. *Cold Spring Harb Perspect Med*. 2013;3(3):a010074. doi:10.1101/cshperspect.a010074
45. Verhoeven V, Renard N, Makar A, et al. Probiotics enhance the clearance of human papillomavirus-related cervical lesions: a prospective controlled pilot study. *Eur J Cancer*. 2013;22(1):46–51. doi:10.1097/CEJ.0b013e328355ed23
46. Ceccarelli G, Cavallari EN, Savinelli S, et al. Clearance of human papillomavirus related anal condylomas after oral and endorectal multistrain probiotic supplementation in an HIV positive male: a case report. *Medicine*. 2018;97(16):e0329. doi:10.1097/MD.00000000000010329
47. Burton JP, Reid G. Evaluation of the bacterial vaginal flora of 20 postmenopausal women by direct (Nugent score) and molecular (polymerase chain reaction and denaturing gradient gel electrophoresis) techniques. *J Infect Dis*. 2002;186(12):1770–1780. doi:10.1086/jid.2002.186.issue-12
48. Zhou X, Hansmann MA, Davis CC, et al. The vaginal bacterial communities of Japanese women resemble those of women in other racial groups. *FEMS Immunol Med Microbiol*. 2010;58(2):169–181. doi:10.1111/j.1574-695X.2009.00618.x
49. Gajer P, Brotman RM, Bai G, et al. Temporal dynamics of the human vaginal microbiota. *Sci Transl Med*. 2012;4(132):132ra152. doi:10.1126/scitranslmed.3003605
50. Macklaim JM, Clemente JC, Knight R, Gloor GB, Reid G. Changes in vaginal microbiota following antimicrobial and probiotic therapy. *Microb Ecol Health Dis*. 2015;26:27799. doi:10.3402/mehd.v26.27799
51. Reid G, Beuerman D, Heinemann C, Bruce AW. Probiotic lactobacillus dose required to restore and maintain a normal vaginal flora. *FEMS Immunol Med Microbiol*. 2001;32(1):37–41. doi:10.1111/fim.2001.32.issue-1
52. Reid G, Bruce AW, Fraser N, Heinemann C, Owen J, Henning B. Oral probiotics can resolve urogenital infections. *FEMS Immunol Med Microbiol*. 2001;30(1):49–52. doi:10.1111/fim.2001.30.issue-1
53. Reid G, Charbonneau D, Erb J, et al. Oral use of lactobacillus rhamnosus GR-1 and L. fermentum RC-14 significantly alters vaginal flora: randomized, placebo-controlled trial in 64 healthy women. *FEMS Immunol Med Microbiol*. 2003;35(2):131–134. doi:10.1016/S0928-8244(02)00465-0
54. Morelli L, Zonenenschain D, Del Piano M, Cognein P. Utilization of the intestinal tract as a delivery system for urogenital probiotics. *J Clin Gastroenterol*. 2004;38(6 Suppl):S107–110. doi:10.1097/01.mcj.0000128938.32835.98
55. Hanson L, VandeVusse L, Jerme M, Abad CL, Safdar N. Probiotics for treatment and prevention of urogenital infections in women: a systematic review. *J. Midwifery Women's Health*. 2016;61(3):339–355. doi:10.1111/jmwh.2016.61.issue-3
56. Andreyev J. Gastrointestinal complications of pelvic radiotherapy: are they of any importance? *Gut*. 2005;54(8):1051–1054. doi:10.1136/gut.2004.062596
57. Chase D, Goulder A, Zenhausern F, Monk B, Herbst-Kralovetz M. The vaginal and gastrointestinal microbiomes in gynecologic cancers: a review of applications in etiology, symptoms and treatment. *Gynecol Oncol*. 2015;138(1):190–200. doi:10.1016/j.ygyno.2015.04.036
58. Griffin C. Probiotics in obstetrics and gynaecology. *Aust N Z J Obstet Gynaecol*. 2015;55(3):201–209. doi:10.1111/ajo.2015.55.issue-3
59. Bayerdorffer E, Neubauer A, Rudolph B, et al. Regression of primary gastric lymphoma of mucosa-associated lymphoid tissue type after cure of helicobacter pylori infection. *Lancet*. 1995;345(8965):1591–1594.
60. Rauch M, Lynch SV. The potential for probiotic manipulation of the gastrointestinal microbiome. *Curr Opin Biotechnol*. 2012;23(2):192–201. doi:10.1016/j.copbio.2011.11.004

61. Viaud S, Saccheri F, Mignot G, et al. The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. *Science*. 2013;342(6161):971–976. doi:10.1126/science.1240537
62. Iida N, Dzutsev A, Stewart CA, et al. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. *Science*. 2013;342(6161):967–970. doi:10.1126/science.1240527
63. Vetizou M, Pitt JM, Daillere R, et al. Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. *Science*. 2015;350(6264):1079–1084. doi:10.1126/science.aad1329
64. Zamarin D, Jazaeri AA. Leveraging immunotherapy for the treatment of gynecologic cancers in the era of precision medicine. *Gynecol Oncol*. 2016;141(1):86–94. doi:10.1016/j.ygyno.2015.12.030
65. Li J, Sung CY, Lee N, et al. Probiotics modulated gut microbiota suppresses hepatocellular carcinoma growth in mice. *Proc Natl Acad Sci U S A*. 2016;113(9):E1306–E1315. doi:10.1073/pnas.1518189113
66. Nami Y, Abdullah N, Haghshenas B, Radiah D, Rosli R, Khosroushahi AY. Assessment of probiotic potential and anticancer activity of newly isolated vaginal bacterium *Lactobacillus plantarum* 5BL. *Microbiol Immunol*. 2014;58(9):492–502. doi:10.1111/mim.v58.9
67. Nami Y, Abdullah N, Haghshenas B, Radiah D, Rosli R, Yari Khosroushahi A. A newly isolated probiotic *Enterococcus faecalis* strain from vagina microbiota enhances apoptosis of human cancer cells. *J Appl Microbiol*. 2014;117(2):498–508. doi:10.1111/jam.2014.117.issue-2
68. Wang KD, Xu DJ, Wang BY, Yan DH, Lv Z, Su JR. Inhibitory effect of vaginal *Lactobacillus* supernatants on cervical cancer cells. *Probiotics Antimicrob Proteins*. 2017;10(2):236–242.
69. Eslami S, Hadjati J, Motevaseli E, et al. *Lactobacillus crispatus* strain SJ-3C-US induces human dendritic cells (DCs) maturation and confers an anti-inflammatory phenotype to DCs. *APMIS*. 2016;124(8):697–710. doi:10.1111/apm.2016.124.issue-8

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