

Activation of Notch1 signaling alleviates dysfunction of bone marrow-derived mesenchymal stem cells induced by cigarette smoke extract

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Abstract: Bone marrow-derived mesenchymal stem cells (BM-MSCs) are considered attractive therapeutic agents for the treatment of COPD. However, little is known about the impact of Notch on the proliferation, migration, and survival of MSCs in a cigarette smoke (CS) microenvironment. Here, we used CS extract to mimic the CS microenvironment in vitro, with the intention to investigate the effect of Notch in regulating proliferation, migration, and survival of BM-MSCs. Rat bone marrow mesenchymal stem cells were infected with lentivirus vector containing the intracellular domain of Notch1 (N1ICD) and challenged with CS extract. Cell proliferation was detected by Ki67 staining and expression of cell cycle-related proteins. A transwell assay was used to measure cell migration and the expression of apoptotic proteins was examined. The proliferation of BM-MSCs overexpressing N1ICD significantly increased. Consistently, levels of cyclin D1, p-Rb, and E2F-1 increased in N1ICD overexpressing cells. N1ICD overexpression also increased cell migration compared with the control group. N1ICD overexpression equilibrated the expression of Bax and Bcl-2, and blocked caspase-3 cleavage, contributing to the inhibition of apoptosis. Moreover, blockade of the PI3K/Akt pathway suppressed the aforementioned cytoprotective effects of N1ICD. In conclusion, activation of Notch signaling improved proliferation, migration, and survival of BM-MSCs in a CS microenvironment partly through the PI3K/Akt pathway.

Keywords: mesenchymal stem cells, chronic obstructive pulmonary disease, cigarette smoke extract, Notch

Introduction

Mesenchymal stem cells (MSCs) are pluripotent stem cells with a fibroblast-like morphology and the capacity to differentiate into chondrocytes, osteoblasts, adipocytes, and muscle cells under different microenvironmental conditions, culture media, and supplements.^{1,2} MSCs are suitable for cell replacement. They do not possess the plasticity of embryonic stem cells, and they offer practical advantages in their ease of separation, propagation, and low immunogenicity. Their use also is not burdened by ethical issues often raised by the use of embryonic stem cells.³ In addition to their regenerative properties, MSCs have therapeutic potential for many diseases because of their anti-inflammatory, immunomodulatory, and anti-apoptotic properties.⁴

COPD is characterized by persistent inflammation and progressive alveolar destruction. It is becoming a major global health problem. Cigarette smoke (CS) is considered the chief causative agent in the development of COPD.⁵ In addition to the respiratory system, CS affects the function of numerous organs and systems.⁶ MSCs are known to possess potent therapeutic capacities in animal models of CS-induced

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or elastase-induced COPD/emphysema.^{7,8} However, little is known about the effect of CS on bone marrow (BM)-derived stem cells. Previous data indicate that BM is suppressed in patients with COPD, supported by the indirect finding that their circulating BM-derived progenitor cells were reduced.^{9,10} Furthermore, a direct effect of CS on the BM is supported by experimental studies showing that CS can adversely affect the performance of BM-derived stem cell recruitment to, and differentiation in, the uterus.¹¹ Despite the observed beneficial effects of cell therapy, improving the function of MSCs in a CS microenvironment is particularly important to improve the therapeutic potential of BM-MSC transplantation. Unfortunately, no potential remedies for CS-induced BM dysfunction have been identified.

Notch proteins belong to a family of transmembrane receptor proteins (Notch1–4) that are the pivotal components of the Notch signaling pathway. Tumor necrosis factor-converting enzyme and the γ -secretase/presenilin complex mediate activation of Notch signaling sequentially.¹² In the canonical Notch signaling pathway, activation of Notch receptors releases a signal in the form of the intracellular domain of Notch (NICD).¹³ In the absence of NICD, the DNA-binding protein CSL associates with ubiquitous co-repressor proteins to repress the transcription of target genes. When NICD enters the nucleus, its binding to CSL may trigger an allosteric change that facilitates displacement of transcriptional repressors with consequential activation of downstream effector proteins such as Hes and Hey protein,¹⁴ which are indispensable for the regulation of stem cell proliferation, differentiation, migration, and apoptosis.¹⁵ We hypothesized that the biological function of BM-MSCs would be improved by Notch signaling activation in the CS microenvironment. CS extract (CSE) contains all of the compounds inhaled by smokers and is frequently added to cells in culture to monitor specific cellular responses.¹⁶ To ascertain the particular biological effects of CS on MSCs, we used CSE to mimic the CS microenvironment *in vitro*. In the present study, we examined the beneficial effects of Notch1 on MSC survival, proliferation, and migration in the CS microenvironment, which partly depend on the PI3K/Akt signaling pathway.

Materials and methods

Ethics statement

The present study was conducted in accordance with the Declaration of Helsinki and the guidelines of the Ethics Committee of Xinhua Hospital affiliated to Shanghai Jiao Tong University School of Medicine, Shanghai, China.

The Ethics Committee of Xinhua Hospital Affiliated to Shanghai Jiao Tong University School of Medicine, Shanghai, China, approved all experimental protocols.

Cell culture

The rat alveolar epithelial cell line, RLE-6TN, which was characterized and found to be similar to alveolar type 2 cells, was purchased from the American Type Culture Collection (Manassas, VA, USA). The cells were cultured in DMEM: Nutrient mixture F-12 (DMEM/F-12; Gibco, Carlsbad, CA, USA) containing 10% fetal bovine serum (FBS, Gibco), 100 IU/mL penicillin, and 100 μ g/mL streptomycin (Gibco), in a humidified incubator with an atmosphere of 5% CO₂ and 95% air at 37°C.

Isolation, culture, and identification of BM-MSCs

Briefly, BM samples from male Sprague Dawley (SD) rats were diluted with PBS and centrifuged at 1,200 rpm for 10 min. The buffy coat containing mononuclear cells was washed twice with PBS and plated in MSC culture growth medium, which consisted of DMEM/F-12 supplemented with 10% (v/v) FBS and 1% (v/v) penicillin–streptomycin. Cultures were maintained at 37°C in a humidified atmosphere containing 5% CO₂. After 24 h, non-adherent cells were discarded. Fresh medium was added and replaced every 3 days.

MSCs were analyzed by flow cytometry (BD Biosciences, San Diego, CA, USA) using anti-CD29, anti-CD90, anti-CD80, anti-CD86, (eBioscience, San Diego, CA, USA), CD34, CD45, CD73, and CD105 antibodies (BD Biosciences) at passage 3. Antibody isotypes were used for negative controls. Cells were incubated in the dark at 4°C with the antibodies for 30 min, then washed 3 times with 10 volumes of PBS and centrifuged at 300 $\times$ g for 5 min. Next, the cells were resuspended in PBS supplemented with 1% FBS and measured by fluorescence-activated cell sorting (FACS). The acquired data from 4 independent experiments were analyzed using FCS Express V4 (BD Biosciences).

To determine tri-lineage differentiation capacity, MSCs were cultured in adipogenic, osteogenic, or chondrogenic medium (Cyagen Biosciences, Guangzhou, China) according to the manufacturer's protocol. To demonstrate adipogenic differentiation, the cells were stained with 0.3% Oil Red O (O0625; Sigma, St Louis, MO, USA) in isopropanol for 30 min and rinsed with PBS. For osteogenic differentiation, the cells were stained with 2% Alizarin Red (A5533; Sigma) for 5 min and rinsed with PBS. For chondrogenic

differentiation, cells were stained with 0.1% Alcian Blue (Cyagen Biosciences) for 30 min and rinsed with PBS. Stained cells were imaged under a Phase-contrast microscope (Leica, Wetzlar, Germany).

CSE preparation

CSE was prepared as previously described.^{17,18} Briefly, 400 mL of CS (containing 12 mg of tar and 2.5 mg of nicotine per cigarette; Da Qianmen, Shanghai, China) was drawn into a 50 mL plastic syringe through a 3-way stopcock and mixed with 20 mL of DMEM/F-12 by vigorous shaking. One cigarette was used per 20 mL CSE, and CSE was prepared no more than 30 min before being used in experiments. The CSE solution was filtered through an aseptic 0.22 µm filter and considered a 100% extract.

NIICD overexpressing lentivirus and transfection

Rat Notch-1 overexpressing lentivirus and lentivirus control were obtained from Genechem (Shanghai, China). Rat Notch-1 complementary DNA (cDNA) (NM_001105721) was generated by polymerase chain reaction (PCR) using the forward primer: 5'-GAGGATCCCCGGGTACCGGT CGCCACCATGCGCAAGCGCAGGCGGCAGCATG-3' and reverse primer: 5'-CACCCACATTCCAGAGGCAT TTAAGGGTATGGACTACAAGGA-3'. Rat BM-MSCs were transfected with NIICD overexpressing lentivirus (Lenti-NIICD-MSCs) and empty vector lentivirus carrying enhanced green fluorescence protein (Lenti-V-MSCs), according to the manufacturer's instructions. Cells were used for the experiments 72 h post-transfection.

Reagent treatment

To investigate the role of the PI3K/Akt pathway in NIICD-mediated MSC survival, proliferation, and migration, LY294002 (20 µM; Cell Signaling Technology, Danvers, MA, USA) was added to the MSC medium for 2 h before CSE treatment to block PI3K/Akt pathway activation.

Cell viability assay

MSC viability was assessed by using a cell counting kit (CCK-8; Dojindo, Kumamoto, Japan). Test cells (5×10^3) were plated in 96-well plates. After treatment with CSE for different intervals (6, 12, 24, and 48 h), the medium was removed, and 100 µL of fresh medium containing 10 µL water soluble tetrazolium (WST-8) reagent was added to each well. The cells were incubated with WST-8 reagent at 37°C for 2 h. The cell viability in each well was determined

by measuring the absorbance at 450 nm using a microplate reader. Data are expressed as the optical density value.

Cell proliferation

Cell proliferative capacities were determined by immunostaining. After treatment, the cells were fixed with 4% paraformaldehyde for 2 h at 37°C and permeabilized with 0.5% (v/v) triton. Then, the cells were incubated with a Ki67 antibody (BD Biosciences, 1:200) overnight at 4°C, followed by phycoerythrin (PE)-labeled goat anti-mouse secondary antibody (Jackson Immuno Research Laboratories, West Grove, PA, USA, 1:100) for 1 h at 37°C. The cells were washed 3 times with PBS, then stained with 4',6-diamidino-2-phenylindole (DAPI) (Sigma) for 5 min and immediately visualized under a fluorescence microscope (Olympus, Tokyo, Japan). The total number of double-positive cells in 5 randomly chosen fields was counted at 100× magnification.

Cell migration

Chemotaxis experiments were performed in polycarbonate transwell inserts (8-µm pores; Corning Costar Corp., Corning, NY, USA). RLE-6TN cells (1×10^6) were added in the lower chamber. MSCs (1×10^5) in 100 µL of culture medium were seeded into the upper chamber in serum-free medium and cultured at 37°C for 24 h. The non-migrating cells in the upper chamber were carefully removed with a cotton swab. Migrating cells were fixed in 4% paraformaldehyde, stained with 1% crystal violet (Beyotime Institute of Biotechnology, Shanghai, China), and photographed. The number of migrated cells was calculated as the ratio of the experimental samples to the control samples $\times 100$, by counting at least 5 randomly separated fields.

Western blot analysis

Following the appropriate treatments, cultured cells were resuspended in lysis buffer (10 mM Tris-HCl, pH 7.4, 150 mM NaCl, 0.5% Nonidet P (NP)-40, 1 mM EDTA, 1 mM Na_3VO_4 , and 1 mM phenylmethylsulfonyl fluoride) (Beyotime Institute of Biotechnology) containing protease and phosphatase inhibitors. The suspension was incubated for 30 min at 4°C and lysed by ultrasound pyrolysis for 30 s. The lysate was centrifuged at 14,000 rpm for 20 min. The protein concentration of each sample was determined with a bicinchoninic acid protein assay (Beyotime Institute of Biotechnology). An equal amount of each protein sample was electrophoresed on a 6%–12% gradient sodium dodecyl sulfate-polyacrylamide gel and transferred to a polyvinylidene difluoride membrane. The membranes were blocked

with 5% milk in Tris-buffered saline and 1% Tween at 25°C for 1 h and then incubated with primary antibodies overnight at 4°C. The antibodies used were: β -actin and Flag (Sigma); C-X-C chemokine receptor type 4 (CXCR4; Abgent, San Diego, CA, USA); cleaved caspase-3, Bcl-2, and Bax (Cell Signaling Technology); and p-21, p-Rb, cyclin D1, E2F-1, t-AKT, and p-AKT (Abcam, Cambridge, UK). The blots were washed 3 times with Tris-buffered saline containing 1% Tween and incubated with a horseradish peroxidase-conjugated secondary antibody (Beyotime Institute of Biotechnology) for 1 h at 25°C. Membranes were washed with TBST 3 times for 10 min and the expression signals were detected with an enhanced chemiluminescence reagent (Millipore, Billerica, MA, USA). The relative quantities of proteins were determined by scanning densitometry (ChemiDoc XRS + Systems; Bio-Rad Laboratories, Inc., Hercules, CA, USA) and analyzed using Image Lab 5.0 Software (Bio-Rad Laboratories, Inc.). β -actin was used as an internal loading control.

Real-time PCR (RT-PCR)

Total cellular RNA from MSCs was extracted with TRIzol reagent (Takara Biotechnology, Dalian, China), and used for reverse transcription to cDNA. cDNA was synthesized by using Prime Script RT Master Mix (Takara Biotechnology), according to the manufacturer's instructions. mRNA levels of *Hes-1*, *Hey1*, *HeyL*, *Sdf-1 α* , *Cxcr4*, *G-Csf*, and *Csf3r* were measured by quantitative PCR using SYBR Green Ex Taq™ (Takara) detected by an ABI 7500 real-time PCR system (Applied Biosystems, Foster City, CA, USA). The PCR protocol consisted of 95°C for 30 s, followed by 40 cycles of 95°C for 5 s, and 60°C for 34 s, with a final dissociation of 95°C for 30 s. The relative transcript levels were calculated by using the $2^{-\Delta\Delta CT}$ method, in which ΔCT is the difference between the threshold cycle for the gene of interest and that for glyceraldehyde-3-phosphate dehydrogenase. The results were normalized to untreated controls. Primer sequences are listed in Table 1.

Terminal deoxynucleotidyl transferase-mediated dUTP-biotin nick-end labeling (TUNEL) assay

MSCs were seeded onto cover slips and then treated with 10% CSE for 24 h to measure apoptosis. Subsequent to incubation, cover slips were processed for the TUNEL assay according to the manufacturer's protocol. Briefly, cells were fixed for 1 h in 4% (w/v) paraformaldehyde at 25°C. After specific labeling was performed, the cells were

Table 1 Primer sequences used for real-time polymerase chain reaction

Gene	Primer sequence	Product length
<i>Hes-1</i>	Forward: 5'TCGGTGGTTACTTTGTTGCTT3' Reverse: 5'AGGCGCAATCCAATATGAAC3'	122 bp
<i>Hey1</i>	Forward: 5'GCGCGGACGAGAATGGAAA3' Reverse: 5'TCAGGTGATCCACAGTCATCTG3'	231 bp
<i>HeyL</i>	Forward: 5'CAGCCCTT CGCAGATGCAA3' Reverse: 5'CCAATCGTCGCAATTGAGAAAG3'	101 bp
<i>SDF-1α</i>	Forward: 5'ATGCCCTGCGGATTCTTTG3' Reverse: 5'TGTTGTTGCTTTTCAGCCTTG3'	116 bp
<i>CXCR4</i>	Forward: 5'GAAGTGGGGTCTGGAGACTAT3' Reverse: 5'TTGCCGACTATGCCAGTCA AG3'	125 bp
<i>G-CSF</i>	Forward: 5'ACTTTCTGCTCAAGGACGCA3' Reverse: 5'AATGGCCTCTTGCTCTGACC3'	85 bp
<i>CSF3R</i>	Forward: 5'GGACAAGAAACCGACCCACT3' Reverse: 5'CCACTACTTCTGGGCTGGTG3'	191 bp

Abbreviations: CSF3R, colony stimulating factor 3 receptor; G-CSF, granulocyte-colony stimulating factor; SDF-1, stromal cell-derived factor-1.

exposed to DAPI in the dark for 5 min to stain nuclei. Then, the immunostained MSCs were observed by fluorescence microscopy. The degree of apoptosis was calculated as the number of double-positive cells per 500 MSC nuclei in 5 randomly chosen fields.

Immunocytochemistry (ICC)

Since caspase-3 activation represents a critical step in the apoptotic process, cleaved caspase-3 was detected in cells by ICC staining with an anti-cleaved-caspase-3 monoclonal antibody (Cell Signaling Technology) according to the manufacturer's protocol. Apoptotic cells were evaluated manually as the number of anti-cleaved-caspase-3 positive cells per 500 counted MSCs.

Statistical analysis

Data are expressed as the mean $\pm$ standard deviation. After comparison of the homogeneity of variance, 1-way analysis of variance was used to determine whether there were any significant differences among the means of the different groups. All statistical analyses were performed using the software SPSS Base 24.0 for Windows (SPSS Inc., Chicago, IL, USA). Probability values $P < 0.05$ were considered significant.

Results

MSC morphology and characterization

MSCs cultured in the medium demonstrated a spindle-like or fibroblast-like shape with directionality and regularity. After several days of induction toward the adipogenic

lineage, considerable morphological changes with lipid vacuole accumulation were observed. Following 14 days of osteogenic differentiation, positive staining for Alizarin Red S confirmed the differentiation of MSCs into osteocytes. After 4 weeks of chondrogenic differentiation, characteristic Alcian Blue staining confirmed the differentiation of MSCs into chondrocytes (Figure S1). MSCs were characterized by surface marker expression via FACS analysis. The results presented in Figure 1 indicate that MSCs expressed CD29, CD73, CD90, and CD105, but these cells were negative for CD34, CD45, CD80, and CD86, consistent with a previous report.¹⁹

Effects of CSE and N1ICD overexpression on MSC viability, Notch signaling pathway activity, and the PI3K/Akt pathway

First, we assessed whether CSE and Notch overexpression had toxic effects on MSCs. We used a single concentration of CSE over an extended period (6–48 h). As shown in Figure 2A, over the entire time course, 10% CSE had little influence on cell viability up to 24 h when compared with untreated cells. However, when MSCs were exposed to 10% CSE for 48 h, their viability decreased significantly. When exposed to 10% CSE for 48 h, the viability of N1ICD overexpressing cells did not significantly decrease compared with that of CSE-exposed MSCs. These results indicated that 10% CSE had little influence on cell viability when exposure was <24 h and N1ICD overexpression can improve the response of MSCs to CSE cytotoxic effects.

To determine the effect of CSE and N1ICD overexpression on Notch signaling pathway activity, we examined the mRNA expression level of Notch1 downstream target genes. N1ICD overexpression increased the *Hey1* mRNA gene expression level ($P < 0.05$), which was inhibited by incubation with CSE (Figure 2B). CSE treatment decreased the expression levels of *Hes-1* and *HeyL* compared with the control group ($P < 0.05$); however, there was no difference between the N1ICD and CSE groups ($P > 0.05$). These data suggest that the Notch1 signaling pathway was inhibited by exposure to CSE, while N1ICD overexpression can activate the endogenous Notch1 signaling pathway and *Hey1*, a target gene of Notch1, in MSCs.

The PI3K/Akt pathway exerts classical growth and survival effects on MSCs.²⁰ Thus, we examined the effects of Notch on the PI3K/Akt pathway in MSCs under CSE exposure. As shown in Figure 2C and D, CSE treatment down-regulated the level of phosphorylated Akt (p-Akt), while N1ICD overexpression increased p-Akt in MSCs (all $P < 0.05$). In the presence of PI3K/Akt inhibitors, p-Akt expression was strongly inhibited, indicating that Notch1 was the upstream activator of the PI3K/Akt pathway in MSCs.

PI3K/Akt is required for N1ICD-mediated proliferation

To establish the role of N1ICD in MSC growth, cells were cultured in the medium containing 10% CSE for 24 h. As shown in Figure 3A and B, the number of Ki67-positive cells was higher in the N1ICD group than that in the CSE group. However, pretreatment with LY294002 reversed this trend,

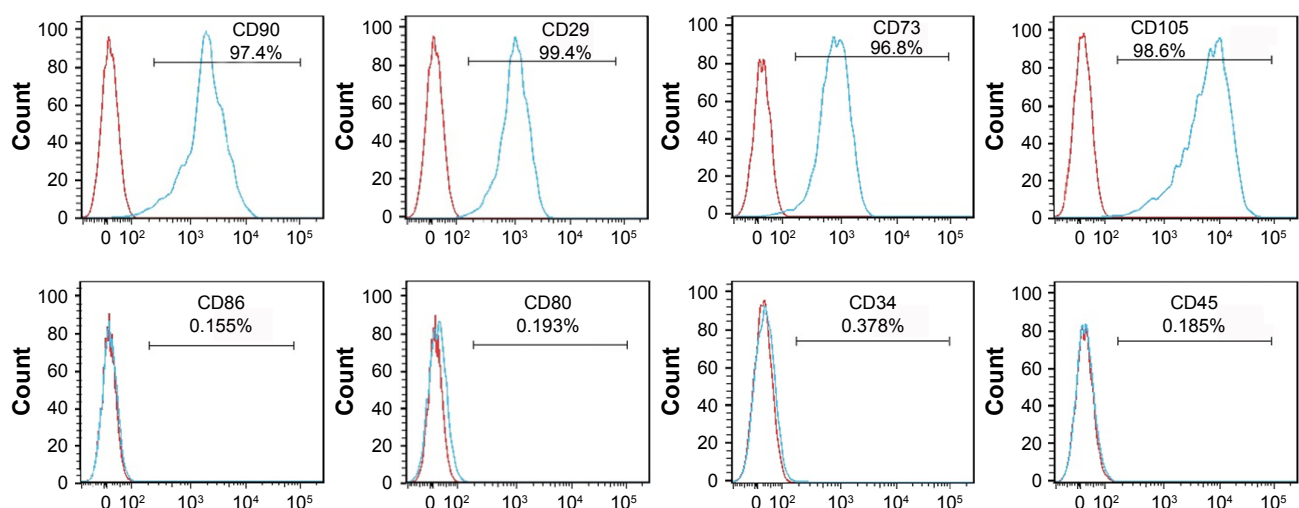


Figure 1 MSCs were phenotypically characterized by means of flow cytometry.

Note: The results demonstrated that MSCs were uniformly negative for CD34, CD45, CD80 and CD86, and positive for CD29, CD73, CD90, and CD105 expression.

Abbreviation: MSCs, mesenchymal stem cells.

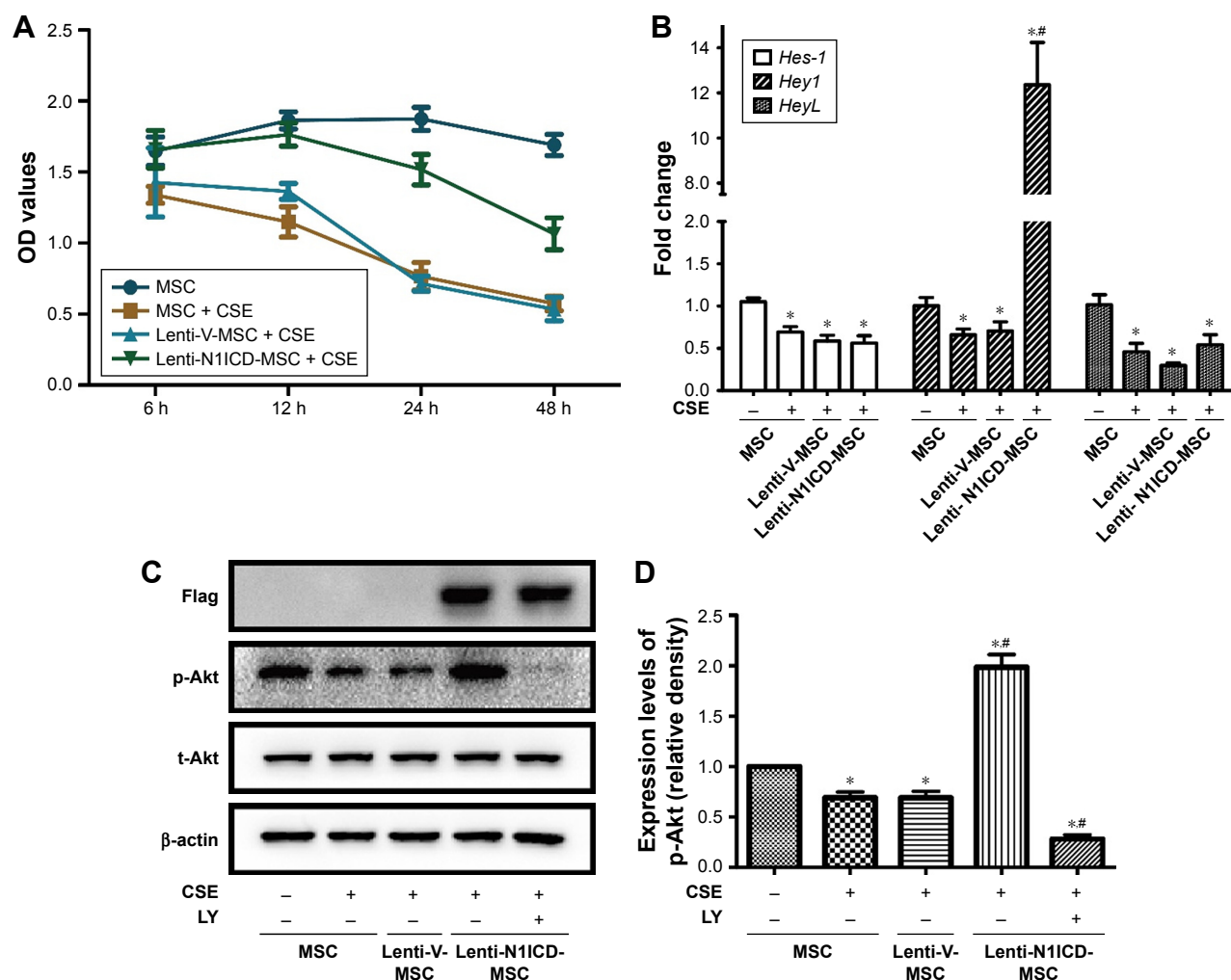


Figure 2 Effects of CSE and N1ICD overexpression on MSC viability, Notch signaling pathway activity, and the PI3K/Akt pathway.

Notes: MSCs were incubated with 10% CSE over an extended period. Their viability was assessed with CCK-8. Notch signaling pathway activity was detected by real-time polymerase chain reaction, and activation of the PI3K/Akt pathway was evaluated by Western blotting. **(A)** CSE exposure for <24 h had little influence on cell viability and N1ICD overexpression can improve MSCs' resistance to CSE cytotoxic effects. **(B)** The Notch signaling pathway was inhibited by incubation with CSE; N1ICD overexpression increased the *Hey1* gene expression level. **(C and D)** CSE treatment downregulated the p-Akt expression level, while N1ICD overexpression increased the level of p-Akt in MSCs. * $P < 0.05$ vs control group, ** $P < 0.05$ vs CSE group.

Abbreviations: CCK-8, cell counting kit-8; CSE, cigarette smoke extract; LY, LY294002; MSCs, mesenchymal stem cells; N1ICD, intracellular domain of Notch1; OD, optical density.

suggesting that N1ICD overexpression caused an increase in the number of dividing cells under CSE exposure. Furthermore, we examined the expression level of cell cycle-related proteins and determined that the expression levels of cyclin D1, p-Rb, and E2F-1 (Figure 3C and D) increased, while that of p21 decreased in N1ICD overexpressing MSCs when compared with that in the CSE group. PI3K/Akt blockade reversed these expression levels. The release of p-Rb activates the transcription factor E2F-1, which allows the expression of genes that are necessary for DNA replication and mitosis.²¹ These data suggested that N1ICD increased the proliferative capacity of MSCs through the PI3K/Akt pathway, which probably promoted cell proliferation via a cyclin D1-Rb-E2F-1-dependent pathway.

N1ICD improves CXCR4 expression and the subsequent migration capacity of MSCs

The stromal cell-derived factor 1 (SDF-1)/CXCR4 biological axis plays an important role in the mobilization and recruitment of MSCs to injured lung tissue after transplantation in a rat COPD model.²² In the present study, CSE treatment did not significantly alter CXCR4 protein expression in MSCs. There was no substantial difference between the control and CSE groups ($P > 0.05$). In contrast, N1ICD overexpression markedly increased the CXCR4 protein level compared with that in the CSE group ($P < 0.05$). However, pretreatment with a PI3K/Akt inhibitor partially decreased CXCR4 protein expression levels (Figure 4A and B), suggesting that the

up-regulation of CXCR4 in N1ICD overexpressing MSCs involved the PI3K/Akt pathway.

To determine whether altered CXCR4 expression in MSCs results in improved migration capacity, we performed a transwell assay to explore the chemoattractive response of MSCs. As shown in Figure 4C and D, no significant difference was observed between the control and CSE groups, and N1ICD overexpression in MSCs increased cell migration, indicating that MSCs with higher CXCR4 expression may have an increased migratory capacity. Interestingly, PI3K/Akt inhibition resulted in fewer cells located in the lower

chamber when compared with the CSE group, suggesting that, in addition to Notch1, other signaling pathways may also be involved in the PI3K/Akt-dependent chemoattractive response. Furthermore, the gene expression of colony stimulating factor 3 receptor (CSF3R) was also increased in N1ICD overexpressing MSCs when compared with that in the CSE group, which was not reversed by LY294002 pretreatment. In contrast, the expression of SDF-1 and granulocyte-colony stimulating factor (G-CSF) were similar between the CSE and N1ICD groups, but all significantly decreased when cells were pretreated with LY294002 (Figure 4E).

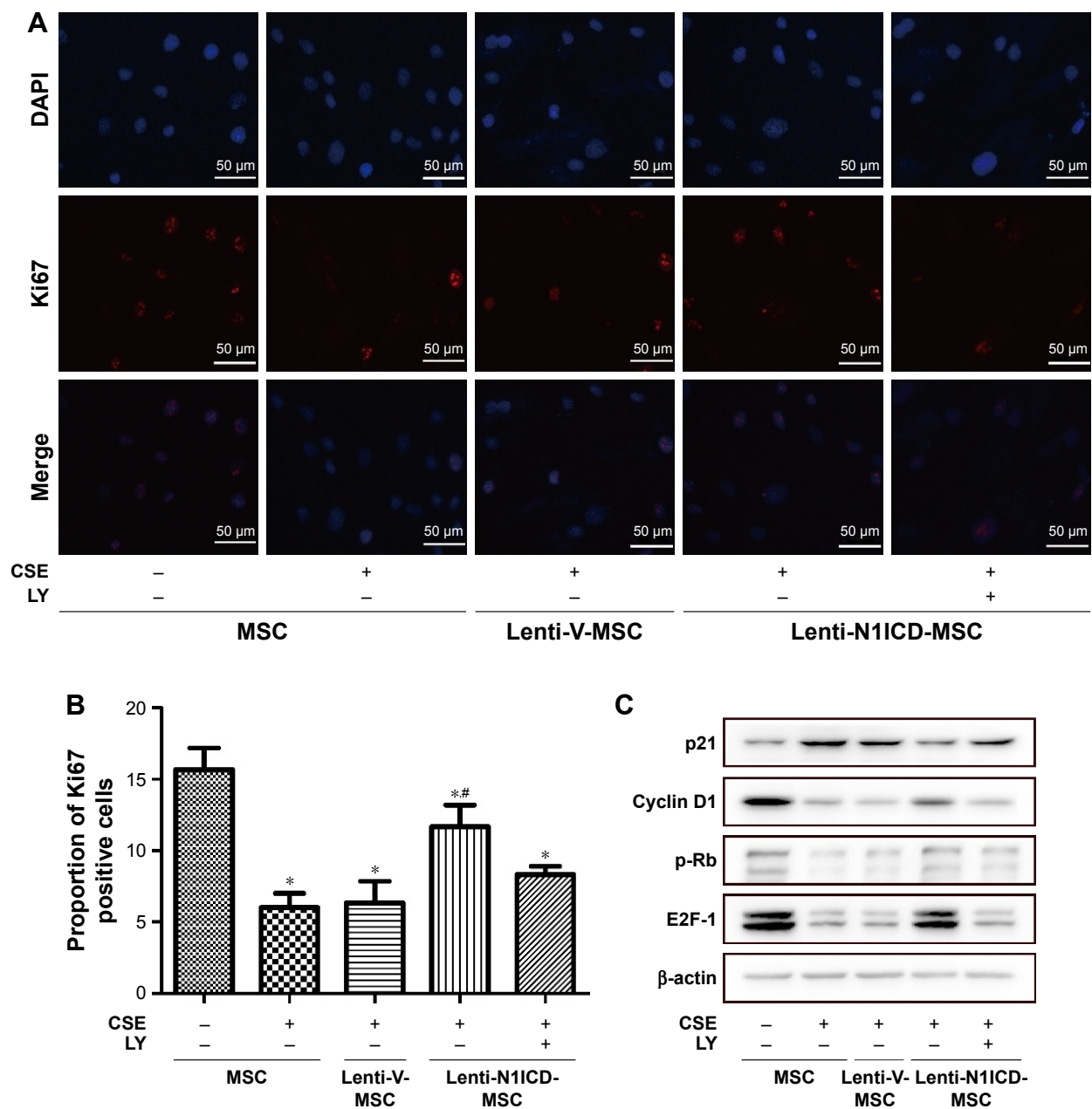


Figure 3 (Continued)

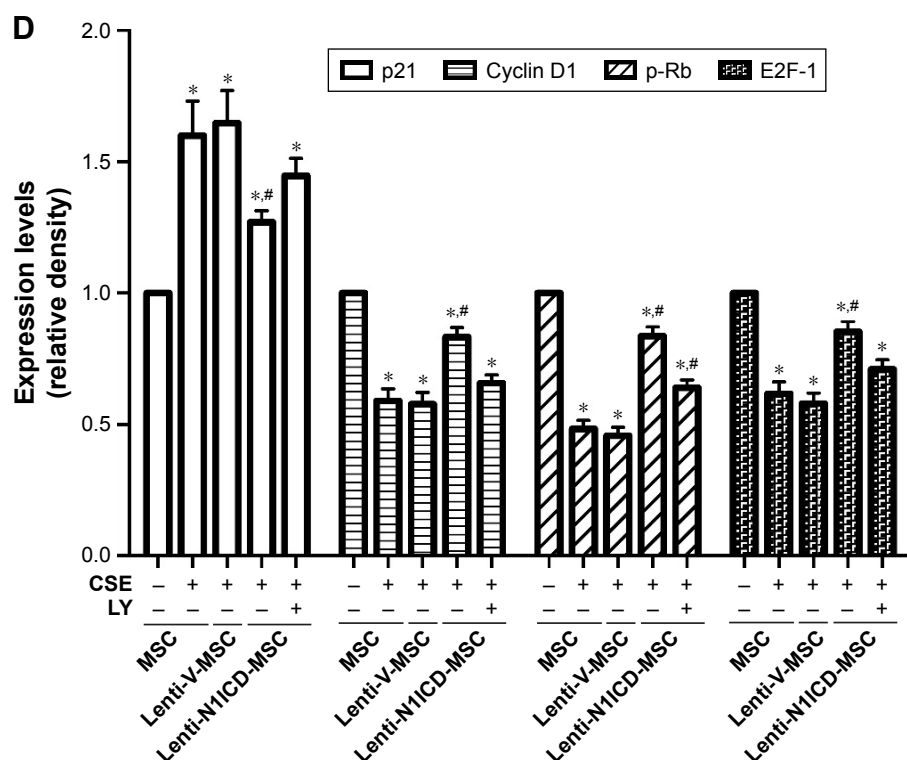


Figure 3 PI3K/Akt is required for N1ICD-mediated proliferation.

Notes: (A and B) Ki67 staining showed that N1ICD overexpression rescued the inhibition of proliferation induced by CSE, while blockade of the PI3K/Akt pathway decreased N1ICD-mediated MSC proliferation. (C and D) After treatment with CSE for 24 h, cyclin D1, p-Rb, and E2F-1 protein expression decreased, while that of p21 increased in the CSE group. N1ICD overexpression reversed this trend, but N1ICD effects were inhibited by blockade of PI3K/Akt. * $P < 0.05$ vs control group; # $P < 0.05$ vs CSE group.

Abbreviations: CSE, cigarette smoke extract; DAPI, 4',6-diamidino-2-phenylindole; LY, LY294002; MSC, mesenchymal stem cells; N1ICD, intracellular domain of Notch1.

Role of Notch and Akt in CSE-induced MSC apoptosis

CSE causes apoptotic cell death by inducing oxidative stress in vitro, which may contribute to the development of pulmonary emphysema.²³ Under our experimental conditions, CSE was applied to induce MSC apoptosis. We assessed whether N1ICD overexpression influenced apoptosis-related proteins in MSCs. First, the TUNEL assay was performed to demonstrate the presence of DNA fragmentation (Figure 5A and B). The analysis showed N1ICD overexpression decreased in TUNEL+ cells when cells were exposed to CSE. In addition, because caspase-3 activation represents a critical step in the apoptotic process, we evaluated cleaved caspase-3 staining by ICC (Figure 5C and D). The results showed that cleaved caspase-3 increased in the CSE group but decreased when MSCs were transfected with N1ICD overexpressing lentivirus. Furthermore, CSE led to an increase in the pro-apoptotic Bax/anti-apoptotic Bcl-2 ratio. In contrast, the Bax/Bcl-2 ratio decreased in N1ICD overexpressing cells (Figure 5E and F). These observations suggested that N1ICD overexpression could reverse CSE-mediated MSC apoptosis by modulating the balance between the expression of pro-apoptotic and

anti-apoptotic proteins belonging to the Bcl-2 family and by decreasing the level of cleaved caspase-3. LY294002 pretreatment to block Akt inhibited the protective effects of N1ICD on CSE-treated MSCs, as evidenced by an increase in the number of apoptotic cells, higher cleaved caspase-3 levels, and a higher ratio of Bax/Bcl-2. These results indicated that Akt was partly responsible for the anti-apoptotic role of N1ICD in CSE-mediated MSC death.

Discussion

Stem cells have recently been proposed as a promising strategy for the treatment of lung diseases. MSCs have been widely administered in experimental models of COPD. The CS microenvironment inhibits the therapeutic potential of MSCs against COPD, which is a critical issue, hence the need to improve the properties of MSCs for transplantation in COPD. Herein, we investigated the effects of Notch on MSC proliferation, migration, and apoptosis in a CS microenvironment in vitro.

The present study reveals several novel findings indicating that Notch1 signaling may represent a critical cell-signaling pathway for maintaining key quality-related properties of

MSCs. Our results are consistent with previous studies. Vujovic et al first investigated the role of Notch signaling in human MSCs by using γ -secretase inhibitors. They found that inhibiting γ -secretase reduced both the proliferation and differentiation capacity of hMSCs.²⁴ Na et al treated human umbilical cord-derived MSCs with γ -secretase inhibitor I (GSI-I) and found that GSI-I treatment led to apoptosis, inhibited osteogenic differentiation, and reduced immunomodulatory properties of MSCs.²⁵ However, the reagent inhibits both Notch signaling and the proteasome activities of MSCs.

To investigate how Notch signaling and its interaction with other potential signaling pathway(s) regulate these critical properties of MSCs, we treated rat BM-MSCs by overexpressing N1ICD and confirmed that N1ICD overexpression improves MSC proliferation. The underlying molecular mechanism may involve the regulation of the cell cycle by Akt through inhibition of p21 protein, which is associated with cell cycle arrest. The phosphorylation of p21 by Akt inhibits its interaction with proliferating cell nuclear antigen and promotes the assembly of the

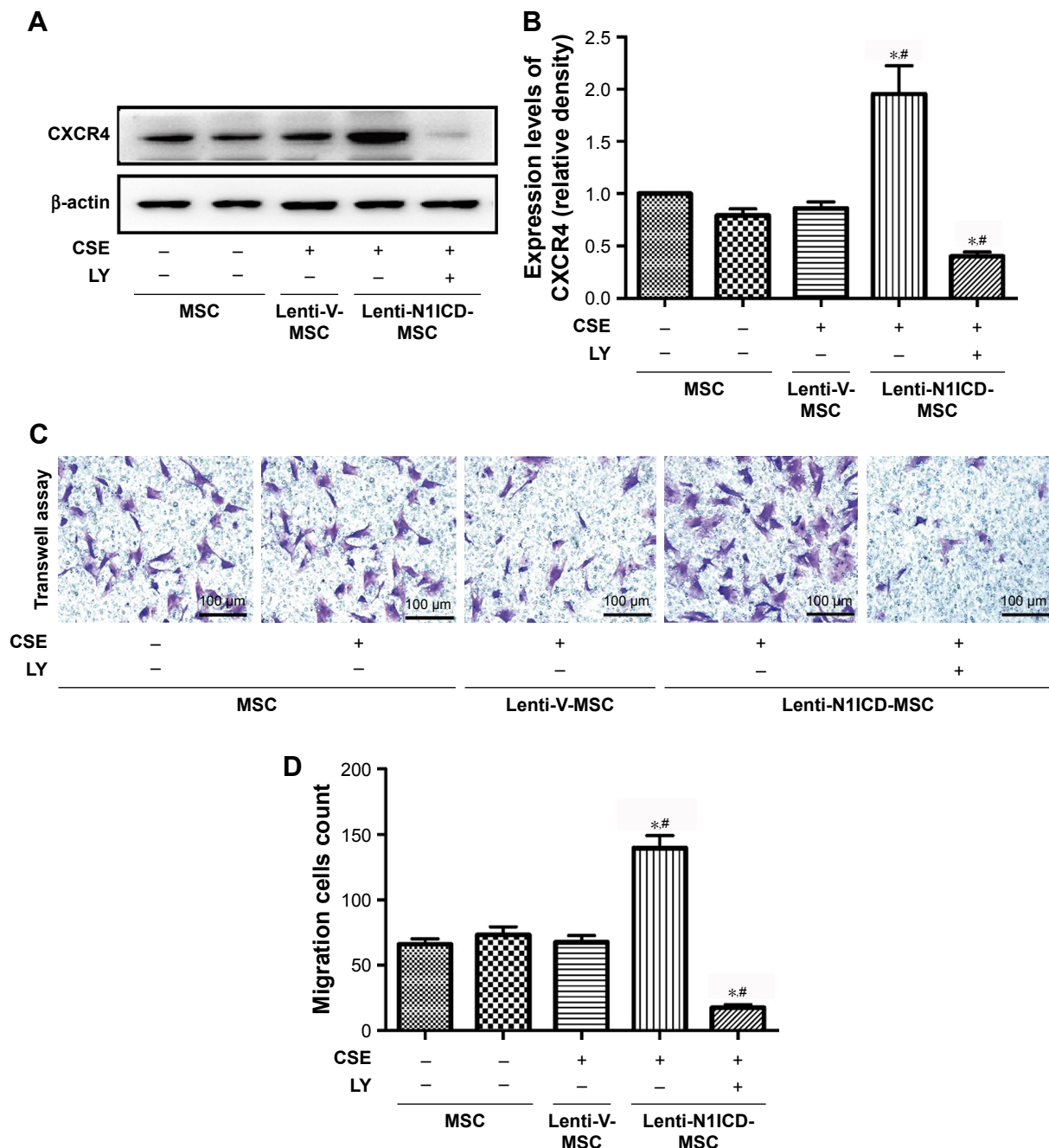


Figure 4 (Continued)

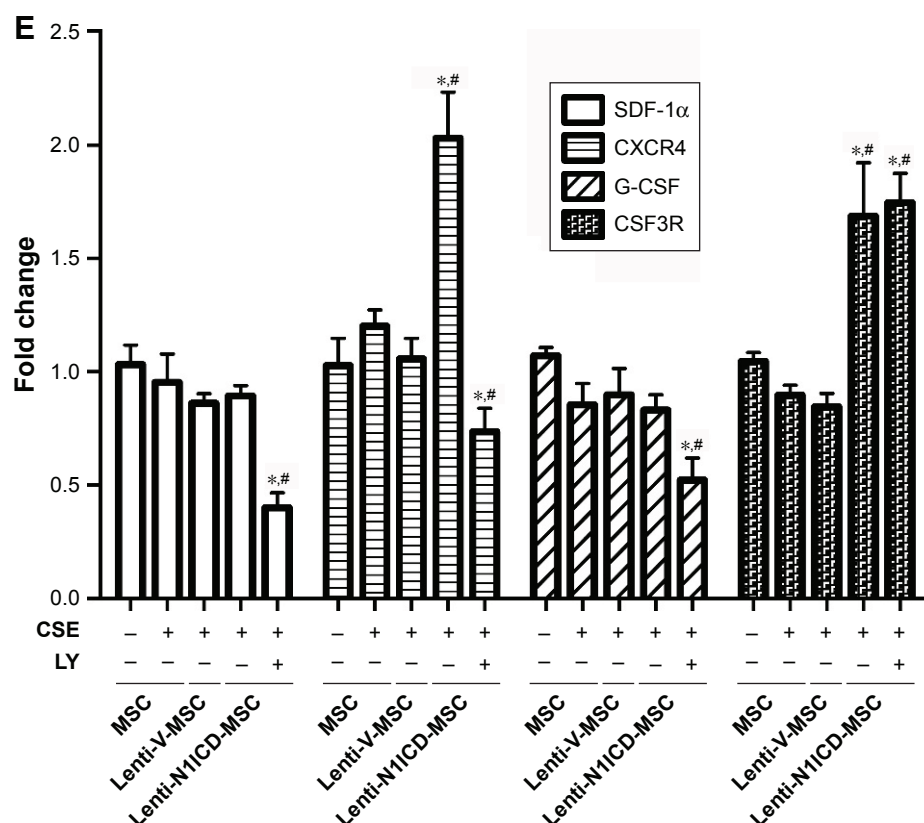


Figure 4 N1ICD improved CXCR4 expression and the subsequent migration capacity of MSCs via PI3K/Akt.

Notes: (A and B) CXCR protein expression was increased in the N1ICD group, but was inhibited by blockade of the PI3K/Akt pathway. (C and D) MSC migration capacity was analyzed with a transwell assay. (E) The expression of genes related to the SDF-1 α /CXCR4 and G-CSF/CSF3R pathways was detected by real-time polymerase chain reaction. * $P < 0.05$ vs control group; # $P < 0.05$ vs CSE group.

Abbreviations: CSF, colony stimulating factor; CSE, cigarette smoke extract; LY, LY294002; MSC, mesenchymal stem cells; N1ICD, intracellular domain of Notch1.

cyclin D1/cyclin-dependent kinase 4 complex.²⁶ Activation of the cyclin D1-P-Rb pathway facilitates the release of the transcription factor E2F-1 in MSCs, resulting in the expression of genes required for DNA synthesis. Moreover, PI3K inhibition reversed the effects of N1ICD on cell cycle protein expression and MSC growth.

Promoting BM stem cell mobilization is a common method to augment the cellular yield of peripheral blood apheresis for clinical stem cell transplantation.²⁷ Selected mobilizing agents have been tested for in vivo beneficial effects following experimental injury, with modulation of the SDF-1 α /CXCR4 axis being the most common strategy. Activation of Notch signaling up-regulated CXCR4 expression in BM-derived dendritic cells (DCs), and transduction with a CXCR4-expressing lentivirus relieved developmental arrest of recombination signal binding protein J κ (RBP-J κ)-deficient DCs.²⁸ Down-regulation of CXCR4 expression in BM-MSCs has been observed in global hemizygous deletion of Notch1 (Notch1^{+/-}) in mice.²⁹ In the present study, after CSE treatment, CXCR4 protein expression was not substantially altered in MSCs, while N1ICD overexpression

improved CXCR4 expression and the subsequent migratory response via the PI3k/Akt pathway as evidenced by transwell assay. Thus, our results provide a feasible means to improve the migration ability of MSCs during in vitro expansion. Furthermore, up-regulation of CSF3R expression in N1ICD overexpressing cells suggests that Notch1 may mediate the mobilization and migration of BM cells through CSF3R and CXCR4 signaling pathways. Indeed, G-CSF/CSF3R and SDF-1 α /CXCR4 signaling pathways contribute to the recruitment of BM-derived cells to damaged tissues.²⁹⁻³¹ Taken together, these findings suggest a critical role for Notch signaling in BM-derived MSC mobilization and migration.

Although the improved proliferative capacity and migration response of MSCs could contribute to higher transplantation efficiency in clinical applications, the hostile environment of injured lung tissue, including inflammation and lung tissue structural damage, may cause excessive cell death, leading to an urgent need to enhance the cells' resistance to apoptosis. CS-induced in vivo apoptosis can be simulated in vitro by CSE treatment of isolated MSCs. The Notch signaling pathway plays important roles in regulating

stem cell self-renewal and apoptosis. Notch1 signaling activation confers chemoresistance to rapamycin in lung cancer cells by inhibiting apoptosis via the PI3K-Akt pathway.³² Furthermore, activated Notch can protect against apoptosis

in physiological and pathological conditions by activating Akt signaling.^{33,34} In the current study, we explored the anti-apoptotic effect of Notch signaling in MSCs in conditions simulating the CS microenvironment in vitro. In addition,

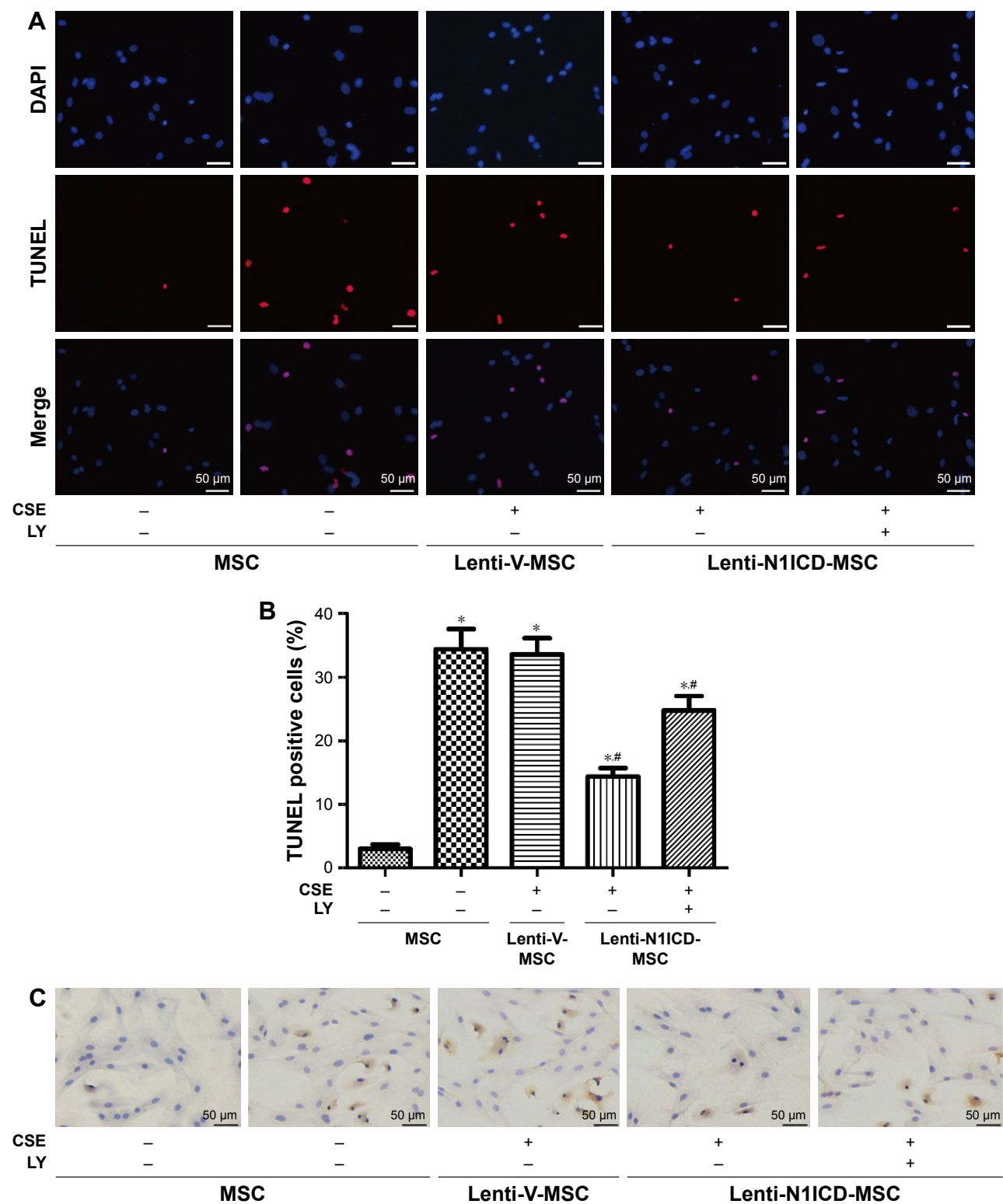


Figure 5 (Continued)

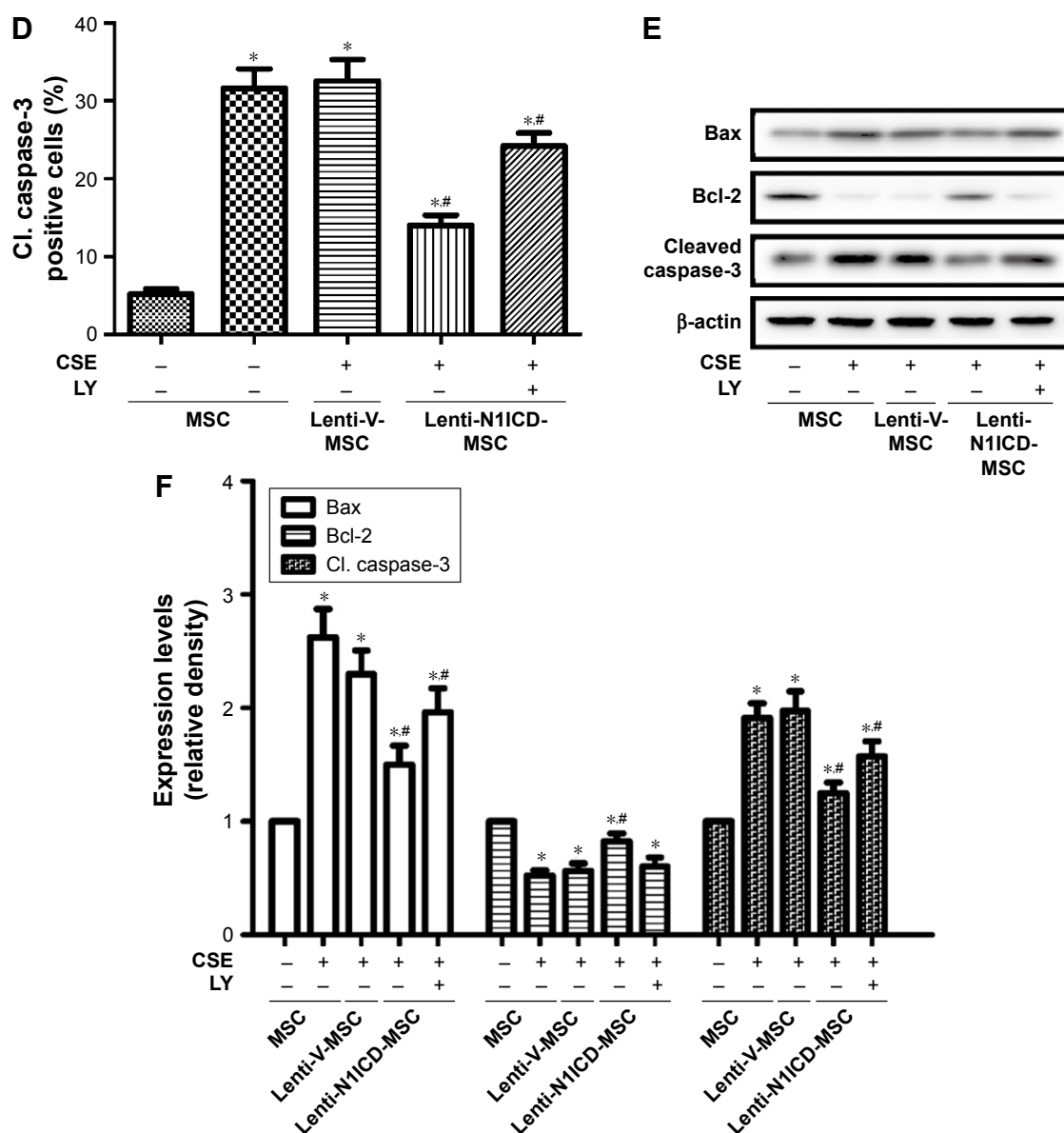


Figure 5 The PI3K/Akt pathway is involved in the anti-apoptotic effects of N1ICD in MSCs after CSE treatment.

Notes: Apoptosis was confirmed by TUNEL assay (A, B). Cleaved caspase-3 immunocytochemistry (C, D). The expression of proteins associated with the apoptotic pathway was also detected. CSE led to an increase in the pro-apoptotic Bax/anti-apoptotic Bcl-2 ratio and cleaved caspase-3 levels. N1ICD overexpression decreased the Bax/Bcl-2 ratio and reduced cleaved caspase-3 expression (E, F). * $P < 0.05$ vs control group; # $P < 0.05$ vs CSE group.

Abbreviations: CSE, cigarette smoke extract; Cl. caspase-3, cleaved caspase-3; DAPI, 4',6-diamidino-2-phenylindole; LY, LY294002; MSC, mesenchymal stem cells; N1ICD, intracellular domain of Notch1; TUNEL, terminal deoxynucleotidyl transferase-mediated dUTP-biotin nick-end labeling.

inhibition of the Akt pathway caused an increase in cellular apoptosis and a reduction in anti-apoptotic proteins.

The Bcl-2 protein family is highly involved in apoptosis. An increase in the Bax/Bcl-2 ratio and the level of cleaved caspase-3 induces cell apoptosis.³⁵ Previous studies reported that Notch1 down-regulation decreased Bcl-2 and Bcl-X (L) protein expression via inactivation of Akt, thereby contributing to apoptosis.^{36,37} In addition, Notch inhibition was associated with a decrease in Akt, which controls proliferation and/or cell death, and targets caspases.³⁸ Our data suggest that inhibition of the Bax/caspase-3 axis could be a

consequence of Notch activation, leading to the prevention of MSC apoptosis, which may help preserve MSC function. In hematopoietic stem cells, Notch signaling can induce growth arrest and apoptosis.^{39,40} This phenomenon could be explained by the fact that activation of the Notch pathway has various effects on proliferation and survival, which are highly cell-type specific, although these cell-specific mechanisms have not been elucidated in most systems.⁴¹ Nevertheless, accumulating evidence highlighted that MSCs therapeutic effects vary in response to local environmental cues.⁴ Moreover, activation of Notch signaling increased the p-Akt level,

and PI3K inhibitors partially blocked the anti-apoptotic effect of Notch signaling in MSCs, making PI3K/Akt signaling regulation an attractive alternative to targeting the Notch signaling pathway regulating MSC apoptosis.

In the present study, Notch was an upstream activator of the PI3K/Akt pathway, and Akt inhibition partially blocked the promoting effects of N1ICD on proliferation, migration, and apoptosis. Many studies have indicated that the accumulation of secreted frizzled-related protein 2⁴² in MSCs, induced by Akt activation, increases MSC self-renewal by inhibiting both Wnt and bone morphogenetic protein⁴³ and prevents the apoptosis of MSCs by triggering cytoplasmic β -catenin translocation into the nucleus.⁴⁴ Therefore, the activation of the PI3K/Akt pathway is probably involved in MSC proliferation and apoptosis. As reported, Akt activation also causes a dramatic increase in CXCR4 expression⁴⁵ and MSC migratory behavior.⁴⁶ Additionally, inhibiting Akt by LY294002 could not completely suppress the promoting effects of Notch signaling on MSCs, suggesting that other pathways may be involved in the action of Notch. Unfortunately, we have only revealed that N1ICD overexpression alleviates dysfunction of MSC induced by CSE in vitro. Whether N1ICD overexpression is involved in the enhanced proliferation, migration, and survival of MSCs in vivo remains to be determined.

In conclusion, these findings suggest that the Notch-PI3K/Akt signaling pathway may improve the biological function of MSCs by effectively reversing the limited proliferation, excessive apoptosis, and weak migration capacity induced by CSE. Therefore, approaches to activate the Notch pathway could potentially increase the efficiency of MSC-based transplantation therapies in the future.

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Disclosure

The authors report no conflicts of interest in this work.

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Supplementary material

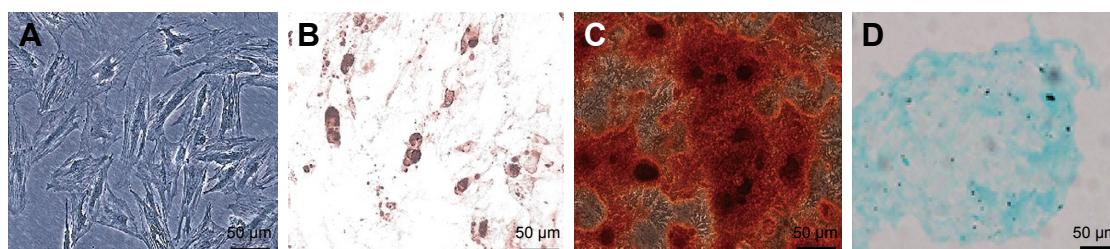


Figure S1 Isolated MSCs presented a fibroblast-like shape (A) and exhibited adipocyte (B), osteoblast (C), and chondrocyte (D) differentiation capacity.

Abbreviation: MSC, mesenchymal stem cells.

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