

# Development of polymer-bound fast-dissolving metformin buccal film with disintegrants

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**Abstract:** Fast-dissolving drug-delivery systems are considered advantageous over the existing conventional oral dosage forms like tablets, capsules, and syrups for being patient friendly. Buccal films are one such system responsible for systemic drug delivery at the desired site of action by avoiding hepatic first-pass metabolism. Metformin hydrochloride (Met), an antidiabetic drug, has poor bioavailability due to its high solubility and low permeability. The purpose of the study reported here was to develop a polymer-bound fast-dissolving buccal film of metformin to exploit these unique properties. In the study, metformin fast-dissolving films were prepared by the solvent-casting method using chitosan, a bioadhesive polymer. Further, starch, sodium starch glycolate, and microcrystalline cellulose were the disintegrants added to different ratios, forming various formulations (F1 to F7). The buccal films were evaluated for various parameters like weight variation, thickness, folding endurance, surface pH, content uniformity, tensile strength, and percentage of elongation. The films were also subjected to in vitro dissolution study, and the disintegration time was found to be less than 30 minutes for all formulations, which was attributed to the effect of disintegrants. Formulation F6 showed 92.2% drug release within 6 minutes due to the combined effect of sodium starch glycolate and microcrystalline cellulose.

**Keywords:** chitosan, sodium starch glycolate, microcrystalline cellulose, drug-delivery system, immediate release

## Introduction

Presently, pharmaceutical research is focusing on the development of new drug-delivery systems as the conventional mode of drug administration suffers from several disadvantages. The rate of drug absorption is also dependant on various factors, like the nature of the drug, its formulation, and its interaction in in vivo conditions, which necessitates the development of fast-dissolving systems using polymers for their effective role as drug carriers. This would increase the bioavailability of the drug at the required site of action and bring about immediate pharmacological action. In addition, these systems ensure greater patient compliance, especially in geriatrics and pediatrics. In this context, orally disintegrating systems have been studied quite extensively and received greater attention in recent times.

Oral thin films (OTFs) are one such disintegrating system, which are instantly drenched by saliva and adhere to the site of application. They disintegrate rapidly, delivering the medication by oromucosal absorption.<sup>1</sup> Several drugs that would otherwise undergo degradation in the gastrointestinal tract have been successfully administered by this route.<sup>2-8</sup> This suits patients suffering from repeated emesis, dysphagia, etc, who cannot swallow a large quantity of water. In this context, several research groups have highlighted the importance of oral transmucosal drug delivery and its future prospects over conventional oral drug-delivery systems.<sup>9-11</sup>

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Therefore, the purpose of the study reported here was to develop a polymer (chitosan)-based oral disintegrating systems of metformin in the form of thin films. Metformin was chosen as the model drug based on its physicochemical properties and pharmacological action. The main limitations of metformin toward therapeutic effectiveness – its poor bioavailability (50%–60%), short biological half-life (5 hours), having the proximal small intestine as its main site of absorption, and the maintenance of adequate plasma levels of the drug<sup>12,13</sup> – are overcome by this method of systemic drug delivery quite satisfactorily. Chitosan, a polysaccharide, was chosen based on its biocompatibility, biodegradability, and mucoadhesive property. The bioavailability and disintegration properties of OTFs in saliva are increased by adding different ratios of super-disintegrants such as starch, sodium starch glycolate (SSG), and microcrystalline cellulose (MCC). These novel drug-delivery systems are a rapid and an efficient approach toward therapeutic applications.<sup>14,15</sup>

## Materials and methods

### Materials

Analytical grade metformin and MCC were obtained from Cipla Limited, Mumbai, India. AR-grade low-molecular-weight chitosan with a viscosity of 20–300 cps, starch, and SSG were purchased from Sigma-Aldrich Co (St Louis, MO, USA) and used as received. All other chemicals and reagents used were of analytical grade. The assay of the chosen drug (metformin hydrochloride) was validated and found to be 98.96% as per Indian pharmacopoeia (IP) 1996 and pre-formulation study was also carried out for the drug. Dissolution medium (pH 6.8) was also prepared as per IP 1996.

### Methods

#### Preparation of fast-dissolving buccal films

Different formulations of metformin fast-dissolving films (F1–F7) were prepared from metformin, chitosan, various disintegrants, sucrose, citric acid, and glycerol by the solvent-casting technique (Table 1). Chitosan (204 mg) was dissolved in 1% acetic acid and stirred until a clear solution formed. To this, a calculated amount of metformin (500 mg), disintegrating agents – sucrose, citric acid, and glycerol – were added and diluted to 20 mL using double-distilled water. The solution was stirred continuously to obtain a clear bubble-free viscous solution, transferred into a clean petri dish (area of 13.995 cm<sup>2</sup>), and kept at room temperature for approximately 24 hours. The prepared films were then cut into a size of 2×2 cm<sup>2</sup>, packed in aluminum foil, and stored in a desiccator.

**Table 1** Formulations of fast-dissolving buccal films of metformin and their compositions

Formulation	Metformin (mg)	CH (mg)	Starch (mg)	SSG (mg)	MCC (mg)	Sucrose (mg)	Citric acid (mg)	Glycerol 5% (mL)	Water (mL)
F1	500	204	44	–	–	64	34	4.2	20
F2	500	204	–	44	–	64	34	4.2	20
F3	500	204	–	–	44	64	34	4.2	20
F4	500	204	22	22	–	64	34	4.2	20
F5	500	204	22	–	22	64	34	4.2	20
F6	500	204	–	22	22	64	34	4.2	20
F7	500	204	–	–	–	64	34	4.2	20

**Abbreviations:** CH, chitosan; MCC, microcrystalline cellulose; SSG, sodium starch glycolate.

## Characterization of buccal films

### Fourier transform infrared spectroscopy

The infrared absorption spectra of OTFs (F1–F7) were analyzed, using a Fourier transform infrared spectrophotometer (IR-Affinity-1, Shimadzu Corporation, Kyoto, Japan).

### Mass uniformity and thickness

Three randomly selected different films from each batch (2×2 cm<sup>2</sup>) were weighed individually on an electronic balance (AUY220 Analytical Balance, Shimadzu Corporation). The average weight of the films was calculated. The thickness of the films was measured by vernier caliper at three different positions on the film and the average was calculated.

### Uniform drug content of the buccal films

The amount of drug present in the films was determined by dissolving the film in 100 mL of phosphate buffer (pH 6.8) for 8 hours under occasional shaking. Of this solution, 5 mL was taken out and diluted to 25 mL, and filtered through a 0.45 μm Whatman filter paper. The drug content was then determined by UV spectrophotometer at 232 nm. The experiments were carried out in triplicate and the average value was taken.

### Determination of surface pH

Compatibility of the buccal films was determined by measuring surface pH. The films were allowed to dip in 40 mL of phosphate buffer (pH 6.8) at 37°C±5°C for 2 hours. The surface pH of the buccal films was confirmed with a pH meter.

## Evaluation of physical stability

### Folding endurance

Folding endurance of the buccal films was determined by repeatedly folding one film<sup>16</sup> at the same place till it broke or was folded up to 350 times manually, which is considered

satisfactory to reveal good film properties. The value of the folding endurance was calculated by counting the number of folds made at the same place without the film breaking.

## Tensile strength and percentage elongation studies of buccal films

The strength of the buccal films was determined by measuring tensile strength (TS) and percentage elongation. A universal testing machine (H5K-S UTM, Tinius Olsen Testing Machine Company, Horsham, PA, USA) was used for the determination of TS by maximum stress applied to the buccal film until it reached the point at which it would break. TS was computed by using the cross-sectional area of the buccal film as per Equation 1.<sup>17</sup> The average of triplicate readings was taken (Table 2).

$$\text{Tensile strength} = \frac{\text{Force at break}}{\text{Initial cross-sectional area of film (cm}^2\text{)}} \quad (1)$$

Percentage elongation was determined by measuring the distance obtained by the maximum length just before the breaking point of the film on the scale, as per Equation 2. Percentage elongation can also be useful to find out the elasticity as well as the strength of a film.

$$\% \text{ elongation} = \frac{\text{Increase in length}}{\text{Original length}} \times 100 \quad (2)$$

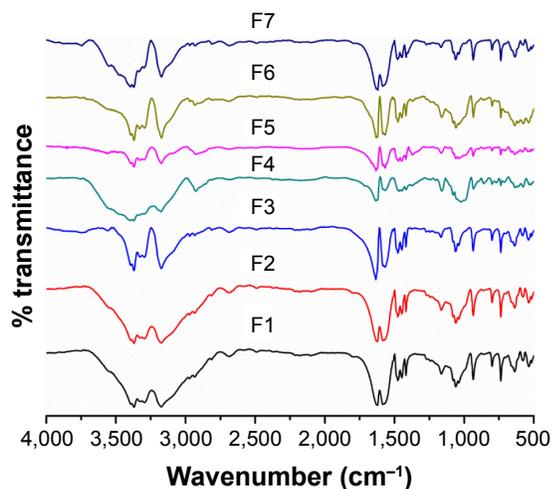
## In vitro dissolution studies

Drug-release studies of the fast-dissolving buccal films of metformin were carried out by using a single-bucket United States Pharmacopeia type I basket apparatus containing 500 mL of pH 6.8 phosphate buffer as the dissolution medium maintained at 37°C±0.5°C.<sup>18,19</sup> It was stirred at 100 rpm and 5 mL samples were withdrawn at 2-minute intervals and replaced with fresh phosphate buffer. Absorbance was measured at 232 nm.

**Table 2** Evaluation of fast-dissolving buccal films of metformin

Formulation	Weight (mg)	Thickness (mm) (±0.05)	Folding endurance (±5)	Surface pH range	Drug content (±1)	Disintegration time (min) (±2)
F1	85.92±0.25	0.188	297	6–7	97	18
F2	77.82±0.68	0.187	335	6–7	98	8
F3	85.05±0.29	0.187	290	6–7	97	11
F4	81.25±1.33	0.188	286	6–7	98	17
F5	76.13±0.32	0.188	210	6–7	95	19
F6	82.58±0.33	0.182	291	6–7	95	20
F7	71.90±0.28	0.176	298	6–7	97	18

**Note:** Results are expressed as of mean ± standard deviation (n=3).



**Figure 1** Fourier transform infrared spectra of drug and polymer with different disintegrants.

The physical parameters, like folding endurance, thickness, TS and % elongation, provide insight into the stability of the films with regard to their utility in pharmaceutical applications.

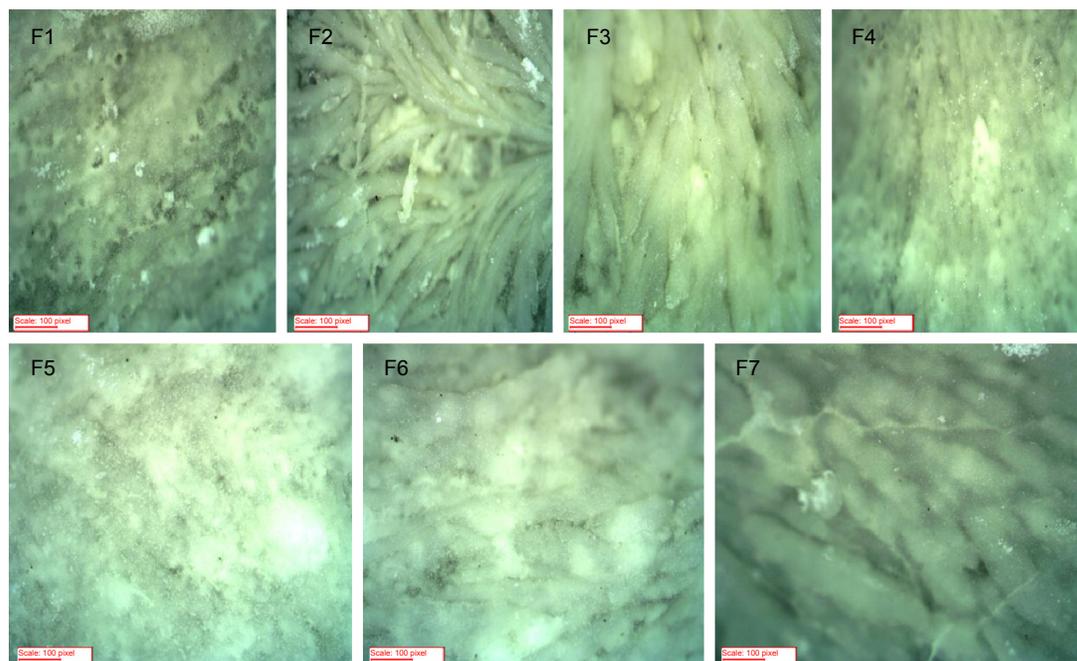
## Results and discussion

We obtained desirable compatibility between a drug-polymer and disintegrants with favorable surface properties. Compatibility between a drug-polymer and different disintegrants

was compared using Fourier transform infrared spectra. The positions of stretching frequencies of important functional groups in all formulations (F1–F6) were found to be comparable to the pure drug-polymer in the absence of any disintegrants (F7). This indicates that there was no chemical interaction of the drug-polymer with the disintegrants (Figure 1).

In addition, mass uniformity, thickness, folding endurance, surface pH, % elongation, and TS values of films were evaluated and were found to be comparable to those of standard limiting values. The surface morphology of the films of different formulations was analyzed using a 100× magnification microscope (RS Microscope System for spectro meter model -PRO-532; PeakSeeker Pro™-532, Raman Laboratory Systems, Woburn, MA, USA) (Figure 2). The images in Figure 2 show surface modifications based on the disintegrants used. The morphology of the starch-containing films (F1, F4, and F5) was slightly different from the morphology shown by other formulations. F2 and F3 contained SSG and MCC, respectively, as a disintegrant and show more or less similar morphology. F6 had a unique morphology because of the presence of both SSG and MCC and showed greater efficacy.

The prepared films possessed no cracks on their surfaces and were cut into 2×2 cm<sup>2</sup> pieces. The various parameters evaluated for all the formulated films are represented in



**Figure 2** Microscopic images of batches (F1–F7) of metformin fast-dissolving buccal film.

**Table 3** Results of tensile strength and percentage elongation for all films

Formulation	F1	F2	F3	F4	F5	F6	F7
Tensile strength (N/cm <sup>2</sup> )	0.2452	0.2492	0.2708	0.2502	0.1965	0.2329	0.1884
Percentage elongation	4.5	3.5	2.5	3.5	4.0	2.5	5.0

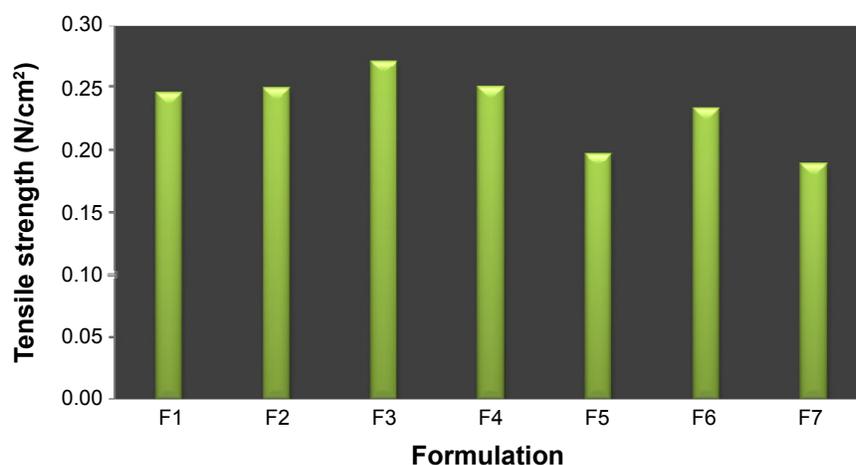
Table 2. The weight of films varied from 70 to 85 mg and they exhibited a thickness of approximately 0.1885 mm. We did not observe any substantial increase in thickness with respect to variations in weight attributed to the addition of disintegrants. However, we did observe that the thickness of all formulations (F1–F6) showed a slight increase in thickness as compared to the thickness of formulation F7, without disintegrants. The folding endurance varied between 285 and 335 times, which is considered the sign of good flexibility, except for film F5, containing starch and MCC, which showed lower flexibility. The surface pH of all the formulations was around neutral (pH 6–7) and hence no irritation would be caused to the buccal cavity. All the formulations contained more than 95% of the drug, as observed by the drug-content study, which indicated that the formulations were satisfactory for further study.

Mucoadhesive formulations have been designed in various forms, like strips,<sup>20</sup> patches,<sup>21</sup> disks,<sup>22</sup> films,<sup>23</sup> etc, and a few potential routes suitable for these dosage forms are via the buccal cavity; sublingually; via the eyes; via the nasal cavity; or via the gastrointestinal, vaginal, or rectal areas.<sup>24</sup> Among these routes, the buccal mucosa, rich in blood supply, is relatively permeable to systemic circulation and facilitates the rapid absorption of drugs due to their inherent flexibility and elasticity.<sup>25,26</sup> This made us want to develop fast-dissolving buccal films of metformin targeted to bring

about immediate therapeutic action. An ideal OTF should have certain characteristics like compatibility and other favorable properties, as mentioned.

In order to determine the flexibility and elasticity of the buccal films, the TS and % elongation were calculated and these showed variations based on the disintegrants used in various ratios represented in Table 3. Higher % elongation and TS were expected for an ideal buccal film.<sup>27</sup> The TS and % elongation varied from 0.1884 to 0.2708 (N/cm<sup>2</sup>) and 2.5% to 5.0%, respectively. Figure 3 represents the comparative TS values. The corresponding % elongation values are given in Table 3. From this, it can be observed that the TS values increase and % elongation decreases with the addition of disintegrants in all formulations as compared to the values obtained for formulation F7, without disintegrants. We did not observe any marked differences between formulations F1–F6.<sup>28</sup>

The results of the in vitro cumulative drug release of metformin from all the formulations (F1–F7) are shown in Table 4. The formulations were designed in such a way that the films showed immediate release, thereby having a shorter contact period in the buccal cavity, providing greater comfort to patients. The in vitro dissolution data confirm that all formulations satisfied the requirement of an immediate drug-release profile. F6 showed 92% of drug release in 6 minutes. All other formulations showed

**Figure 3** Tensile strength of all buccal films (F1–F7).

**Table 4** In vitro dissolution (% cumulative release) profile of metformin buccal film

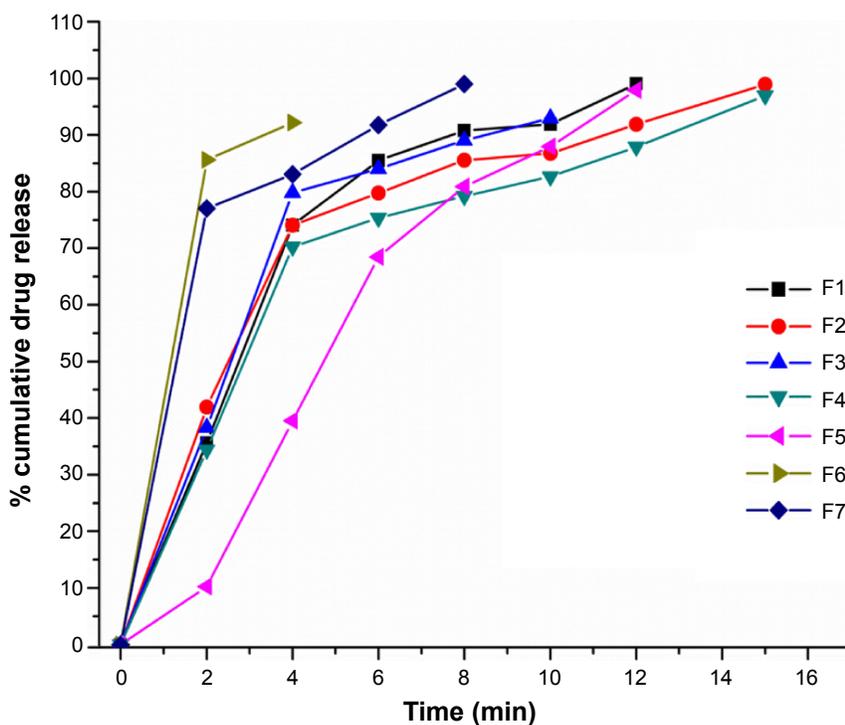
Formulation	2 min	4 min	6 min	8 min	10 min	12 min	15 min
F1	35.59	74.07	85.47	90.82	91.88	99.08	–
F2	41.93	74.07	79.76	85.57	86.76	91.88	98.96
F3	38.32	79.78	84.04	89.02	92.99	–	–
F4	34.39	70.28	75.34	79.21	82.67	87.93	96.95
F5	10.25	39.52	68.44	80.90	87.99	97.92	–
F6	39.27	85.59	92.2	–	–	–	–
F7	37.75	77.05	83.07	91.78	99.00	–	–

more than 90% release within 10 minutes. The difference in release profile was due to the incorporation of different disintegrants in various ratios. Among all the formulations, formula F6, containing SSG and MCC, was useful for immediate-release drug delivery. Table 4 represents the cumulative % drug release and a graphical representation is shown in Figure 4.

## Conclusion

The work presented here indicates the great potential of fast-dissolving buccal films containing metformin for systemic delivery that have the added advantage of circumventing hepatic first-pass metabolism. Although various parameters evaluated show optimum values as per the standard limits, the values seem to vary only slightly for

formulations F1–F6, as compared to those for formulation F7. The prepared buccal films possessed good mechanical properties as indicated by their TS and % elongation values. However, these formulations facilitated the immediate release of metformin and also maintained the therapeutic dosage of the drug administered through the buccal route. But, the dissolution test revealed that the most effective immediate release was demonstrated by formulation F6, due to the combined effect of two different disintegrants, SSG and MCC. It is significant to note that the in vivo bioavailability of a drug can best be predicted based on in vitro dissolution results rather on disintegration tests, as in vivo bioavailability may vary depending upon the nature of the disintegrant or disintegrant combinations used for the specified drug. The types of films discussed here have

**Figure 4** Cumulative drug release of all buccal films (F1–F7).

greater future prospects for their utility value in emergency situations, provided the issues of stability, durability, and efficacy are taken care of.

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## Disclosure

The authors report no conflicts of interest in this work.

## References

- Barnhart S, Vondrak B. Dissolvable films: dissolvable films for flex product format in drug delivery. *Pharmtech.com* [serial online]. 2008 April 1: [about 5 p]. <http://www.pharmtech.com/dissolvable-films-dissolvable-films-flexible-product-format-drug-delivery?id=&pageID=1&sk=&date=>. Accessed March 17, 2015.
- Anders R, Merkle HP. Evaluation of laminated mucoadhesive patches for buccal drug delivery. *Int J Pharm*. 1989;49(3):231–240.
- Chen WG, Hwang GC. Adhesive and in vitro release characteristics of propranolol bioadhesive disc system. *Int J Pharm*. 1992;82(1–2):61–66.
- Alur HH, Pather SI, Mitra AK, Johnston TP. Transmucosal sustained-delivery of chlorpheniramine maleate in rabbits using a novel, natural mucoadhesive gum as an excipient in buccal tablets. *Int J Pharm*. 1999;188(1):1–10.
- Guo JH. Bioadhesive polymer buccal patches for buprenorphine controlled delivery formulation in vitro adhesion and release properties. *Drug Dev Ind Pharm*. 1994;20(18):2809–2821.
- Ahuja A, Dorga M, Agrawal SP. Development of buccal tablets of diltiazem HCL. *Indian J Pharm Sci*. 1995;57:26–30.
- Nagai T, Koshini R. Buccal/gingival drug delivery systems. *J Control Release*. 1987;6:353–360.
- Harris D, Robinson JR. Drug delivery via the mucous membrane of the oral cavity. *J Pharm Sci*. 1992;81(1):1–10.
- Sattar M, Sayed OM, Lane ME. Oral transmucosal drug delivery – current status and future prospects. *Int J Pharm*. 2014;471(1–2):498–506.
- Preis M, Knop K, Breikreutz J. Mechanical strength test for orodispersible and buccal films. *Int J Pharm*. 2014;461(1–2):22–29.
- Mura P, Mennini N, Kosalec I, Furlanetto S, Orlandini S, Jug M. Amidated pectin-based wafers for econazole buccal delivery: Formulation optimization and antimicrobial efficacy estimation. *Carbohydr Polym*. 2015;121:231–240.
- Whitehead L, Fell JT, Collett JH, Sharma HL, Smith AM. Floating dosage forms: an in vivo study demonstrating prolonged gastric retention. *J Control Release*. 1998;55(1):3–12.
- Basak SC, Rahman J, Ramalingam M. Design and in vitro testing of a floatable gastroretentive tablet of metformin hydrochloride. *Pharmazie*. 2007;62(2):145–148.
- Arya A, Chandra A, Sharma V, Pathak K. Fast dissolving oral films: an innovative drug delivery system and dosage form. *International Journal of Chem Tech Research*. 2010;2(1):576–583.
- Mashru RC, Sutariya VB, Sankalia MG, Parikh PP. Development and evaluation of fast-dissolving film of salbutamol sulphate. *Drug Dev Ind Pharm*. 2005;31(1):25–34.
- Khanna R. Development and evaluation of mucoadhesive buccal dosage form of clotrimazole [master's thesis]. New Delhi: Jamia Hamdard; 1995.
- Nafee NA, Boraie NA, Ismail FA, Mortada LM. Design and characterization of mucoadhesive buccal patches containing cetylpyridinium chloride. *Acta Pharm*. 2003;53(3):199–212.
- Mishra R, Amin A. Formulation development of taste masked rapidly dissolving films of cetirizine hydrochloride. *Pharm Tech U S A*. 2009;33(2):48–56.
- Chen MJ, Tirol G, Bass C, Corniello CM, Watson G, Sanchez I. Castable edible pharmaceutical films. *Drug Delivery Tec*. 2008;8:34–41.
- Ilango R, Kavimani S, Mullaicharam AR, Jayakar B. In vitro studies on buccal strips of glibenclamide using chitosan. *Indian J Pharm Sci*. 1997;59(5):232–235.
- Saisivam S, Asheraff MH, Gerald NS, Jayaprakash S, Nagarajan M. Design and evaluation of diltiazem hydrochloride buccal patches. *Indian J Pharm Sci*. 2000;62(3):336–338.
- Krzmar DB, Grcic JF, Zorc B, Zovko M. Dissolution of celecoxib from mucoadhesive disks based on polyaspartamide derivatives. *Acta Pharm*. 2006;56:463–471.
- Kumar GV, Krishna RV, William GJ, Konde A. Formulation and evaluation of buccal films of salbutamol sulphate. *Indian J Pharm Sci*. 2005;67:160–164.
- Ahuja A, Khar RK, Ali J. Mucoadhesive drug delivery systems. *Drug Dev Ind Pharm*. 1997;23:489–515.
- Shojaei AH. Buccal mucosa as a route for systemic drug delivery: a review. *J Pharm Pharm Sci*. 1998;1(1):15–30.
- Raghuraman S, Velrajan G, Ravi R, Jeyabalan B, Benito D. Design and evaluation of propranolol hydrochloride buccal films. *Indian J Pharm Sci*. 2002;64(1):32–36.
- Wong CF, Yuen KH, Peh KK. Formulation and evaluation of controlled release Eudragit buccal patches. *Int J Pharm*. 1999;178(1):11–22.
- Vishnu YV, Chandrasekhar K, Ramesh G, Rao YM. Development of mucoadhesive patches for buccal administration of carvedilol. *Curr Drug Del*. 2007;4(1):27–39.

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