

# Surgical wound healing using hemostatic gauze scaffold loaded with nanoparticles containing sustained-release granulocyte colony-stimulating factor

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**Background:** The therapeutic strategies for malignant melanoma are still cancer chemotherapy, radiotherapy, and tumor resection. However, these therapeutic strategies often lead to a reduced neutrophilic granulocyte count or loss of more blood after surgical tumor resection. In this study, we developed a formulation of hemostatic gauze impregnated with sustained-release granulocyte colony-stimulating factor (G-CSF) with increasing of the neutrophilic granulocyte count in the blood following chemotherapy and decreasing blood loss after surgical tumor resection.

**Methods:** We designed a formulation with both hemostatic properties and increased neutrophil content to be used in cancer chemotherapy, radiotherapy, and tumor resection, comprising a hemostatic gauze as a scaffold and (G-CSF)-loaded dextran nanoparticles coated with polylactic-co-glycolic acid (PLGA) solution fabricated by direct spray-painting onto the scaffold and then vacuum-dried at room temperature. The performance of this system was evaluated in vitro and in vivo.

**Results:** Nearly zero-order release of G-CSF was recorded for 12–14 days, and the cumulative release of G-CSF retained over 90% of its bioactivity in a NFS-60 cell line proliferation assay when the scaffold was incubated in phosphate-buffered saline (pH 7.4) at 37°C. The in vivo hemostatic efficacy of this formulation was greater than that of native G-CSF, the scaffold directly spray-painted with G-CSF solution or PLGA organic solution as a coating, or when a blank scaffold was covered with the coating.

**Conclusion:** Our results suggest that this formulation has both hemostatic properties and increased neutrophil activity.

**Keywords:** hemostatic gauze scaffold, granulocyte colony-stimulating factor, bioactivity, sustained-release, stability

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## Introduction

Granulocyte colony-stimulating factor (G-CSF)<sup>1</sup> is a member of a family of secreted glycoproteins which act on both neutrophils and granulocyte precursors in the bone marrow to promote their differentiation, proliferation, and maturation, to mobilize neutrophils from the bone marrow pool into the peripheral blood circulation, and to control neutrophil dynamics in the blood.<sup>2</sup> G-CSF has been investigated as a neuroprotective agent in neurological disorders<sup>3–7</sup> and has been used clinically to increase neutrophil levels in bone marrow transplantation,<sup>8</sup> to aid recovery of the neutrophil count in treatment of ritodrine-induced neutropenia,<sup>9</sup> as an adjunct to chemotherapy or radiotherapy in the treatment of cancer,<sup>10–13</sup> and in various conditions,

including neutropenia associated with aplastic anemia or myelodysplastic syndrome.

However, cancer chemotherapy and radiotherapy often lead to a reduced neutrophilic granulocyte count, and G-CSF is used first-line for increasing neutrophilic granulocytes in the blood.<sup>13</sup> Clinical application of G-CSF involves frequent injections, and compliance is poor. In order to enhance compliance, the development of noninvasive dosage routes, including nasal,<sup>14–16</sup> pulmonary,<sup>17–19</sup> and oral<sup>20–22</sup> formulations is expected, and investigations have been performed using various formulations, but the bioavailability of these has been very low, so they have met with little success.<sup>23,24</sup> A long-acting injectable formulation enabling decreased dosing frequency, PEGylated G-CSF, fusion G-CSF protein, and sustained-release formulations have also been studied.<sup>25–27</sup> However, PEGylation and fusion protein often causes G-CSF bioactivity to decrease, and sustained-release formulations suffer from loss of bioactivity and increased immunogenicity due to the methods used in their preparation, eg, a shortage of protective protein, oil-water or water-air interfaces of water-in-oil (w/o),<sup>28</sup> water-in-oil-in-water (w/o/w), or water-in-oil-in-oil (w/o/o) emulsion methods.<sup>29–31</sup> We developed several novel methods for preserving bioactivity of the protein by avoiding creation of oil-water or water-air interfaces, such as a stable aqueous-aqueous phase (“emulsion”) and a low-temperature aqueous-aqueous phase separation method for protein-loaded dextran nanoparticles or microparticles.<sup>32,33</sup> These nanoparticles or microparticles may be used further to prepare sustained-release formulations.<sup>34–36</sup> For example, the particles may be used to coat stents, scaffolds, and medical devices. Because hemostatic gauze is often used following tumor resection or in other diseases to stop bleeding, we wanted to develop a formulation of hemostatic gauze impregnated with sustained-release G-CSF. We hoped that the formulation would increase the neutrophilic granulocyte count in the blood following chemotherapy and decrease blood loss after surgical tumor resection.

The objective of this study was to develop a surgical hemostatic gauze scaffold with high encapsulating efficiency, preserved protein stability, and ability to release G-CSF in a sustained manner, by loading a hemostatic gauze scaffold with G-CSF-dextran nanoparticles and coating the scaffold with a polylactide-co-glycolic acid (PLGA) suspension. The formulation was characterized by scanning electron microscopy, size-exclusion chromatography-high-pressure liquid chromatography (HPLC), in vitro release characteristics, a NSF-60 cell proliferation assay, and in vivo efficacy.

## Materials and methods

### Materials

PLGA (50:50, molecular weight 47 kDa; 50:50, molecular weight 20 kDa) was obtained from Lakeshore BioMaterials Inc (Kent, OH). Hemostatic gauze (oxidized regenerated cellulose) was purchased from Ethicon SARRL (Neuchatel, Switzerland). Ethyl acetate (analytical reagent), dichloromethane (analytical reagent), dextran (molecular weight 64,000–76,000, biochemical reagent) and polyethylene glycol (PEG, molecular weight 6000, biochemical reagent) were purchased from Sigma-Aldrich (St Louis, MO). G-CSF was supplied by Jinsai Yaoye Co Ltd (Changchun, China). A human G-CSF enzyme-linked immunosorbent assay kit was purchased from R&D Systems Inc (Minneapolis, MN). A bipotential murine hemopoietic cell line (NSF-60) was purchased from the American Type Culture Collection (Manassas, VA).

### Animals

Adult male Sprague Dawley rats weighing approximately 200 g were used. The animals were maintained under standardized rodent conditions at a room temperature of  $22^{\circ}\text{C} \pm 1^{\circ}\text{C}$ . The animal experiments complied with the principles of laboratory animal care and were approved by the Institutional Animal Care and Utilization Committee of Shanghai Jiao Tong University.

### G-CSF-loaded dextran nanoparticles

A 3 mL mixture of dextran (1%, w/w), G-CSF (0.25%, w/w), and PEG (5%, w/w) in a ratio of 1:4:20 was prepared,<sup>32</sup> and then frozen in a refrigerator at  $-80^{\circ}\text{C}$  overnight. Samples were lyophilized using a Christ Alpha 1–2 laboratory freeze-dryer at a pressure of  $5.25 \times 10^{-3}$  Pa for 24 hours. The lyophilized powders were washed with 5 mL of dichloromethane, then centrifuged at 12,000 rpm for 5 minutes (Eppendorf 5415D, Hamburg, Germany) to remove the supernatant liquid, ie, the PEG continuous phase (because G-CSF-loaded dextran nanoparticles are insoluble in dichloromethane and would precipitate in the bottom of the centrifuge tube). The washing-centrifugation procedure was repeated three times, and the G-CSF dextran nanoparticles collected were evaporated at 1.33 Pa and room temperature for 24 hours using a vacuum dryer (Fuma DZF-3, Shanghai Fuma Co Ltd, Shanghai, China) to remove the dichloromethane residue. The nanoparticles obtained contained less than 0.5% (w/w) PEG after the washing process.

## Hemostatic gauze scaffold with G-CSF-loaded dextran nanoparticles

The hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles was prepared by spraying and painting a 2.5 cm × 1 cm piece of hemostatic gauze scaffold with a suspension containing G-CSF-loaded dextran nanoparticles and PLGA ethyl acetate solution or G-CSF in aqueous solution and PLGA ethyl acetate solution at a rate of 0.25 mL/minute. The suspension was prepared by vortex mixing 5% PLGA ethyl acetate solution (containing 20 mg PLGA) and 5 mg G-CSF-loaded dextran nanoparticles for 5 minutes. After initial drying, the hemostatic gauze scaffold with G-CSF-loaded dextran nanoparticles was put in a refrigerator at 20°C for at least 10 hours, followed by further evaporation at room temperature in a vacuum below 0.1 mbar for 48 hours to remove any dichloromethane residue. The hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles was then sprayed and painted with additional G-CSF dextran nanoparticle-free PLGA (14 mg), with the outer layer free of dextran nanoparticles to improve the release kinetics of G-CSF.

After the hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles had been refrigerated at -20°C and freeze-dried for 10 hours, the pure hemostatic gauze scaffold was attached and adhered to the surface of the hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles and further evaporated in a vacuum below 0.1 mbar for 48 hours to remove any dichloromethane residue at room temperature.

## Scanning electron microscopy

Scanning electron microscopy (SEM) images of the hemostatic gauze, hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles, and the G-CSF dextran nanoparticles alone were obtained using a FEI Sirion 200 SEM (Hillsboro, OR). Prior to image scanning, the samples were coated with gold vapor under an argon atmosphere and at 5–10 keV.

## Size distribution of G-CSF dextran nanoparticles

The size distribution and average particle size of the G-CSF dextran nanoparticles were detected using a particle size and shape analyzer (Ankersmid CIS-100, Edegem, Belgium). Ten milligrams of dry particles were resuspended in a quartz cell containing isopropyl alcohol with a stirring magnet bar.

## G-CSF encapsulation efficiency

The hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles was washed with 10 mL dichloromethane, and then centrifuged at 12,000 rpm for 5 minutes to remove the PLGA and recover the G-CSF-loaded dextran nanoparticles, which were then redissolved in phosphate-buffered saline (pH 7.4) and diluted using a G-CSF enzyme-linked immunosorbent assay kit. The G-CSF encapsulation efficiency of the hemostatic gauze scaffold with G-CSF-loaded dextran nanoparticles was determined using the following equation:

$$\text{Encapsulation efficiency (\%)} = P/P_i \times 100$$

where P is the actual total weight of G-CSF encapsulated into the coating on the scaffold and  $P_i$  is the theoretical amount of G-CSF (obtained from feeding) encapsulated into the coating. The standard deviation for drug loading was calculated based on the coating experiments which were repeated five times.

## Size exclusion chromatography

Size exclusion chromatography was carried out using a HPLC system equipped with a TSK G2000SWXL size exclusion column (Shimadzu, Tokyo, Japan). Elution was performed using a peristaltic pump with 50 mM phosphate-buffered saline (pH 7.3) and a flow rate of 1.0 mL/minute at room temperature. Absorbance of each sample was measured at a wavelength of 214 nm. The dissolved G-CSF sample was filtered using a 0.22 µm film and then injected into the HPLC system. The flow rate of the mobile phase was 1.0 mL/minute. The amount of monomer or aggregate G-CSF was calculated using the Chromato-Solution-Light Shimadzu software.

## In vitro release profile

The 2.5 cm × 1 cm hemostatic gauze scaffolds containing G-CSF-loaded dextran nanoparticles (18 mg) were individually incubated in vials containing 2 mL of phosphate-buffered saline (100 mM, pH 7.4) at 37°C under constant shaking. For G-CSF release, the release medium was replaced by fresh buffer on a scheduled date and the G-CSF contents were assayed using a G-CSF enzyme-linked immunosorbent assay kit. For each sample, the G-CSF release experiment was repeated five times and release profiles were calculated based on the average of three experiments.

## In vitro release bioactivity

A NFS-60 cell line were cultured in RPMI 1640 (Gibco®, Invitrogen, Carlsbad, CA) with 10% fetal calf serum and 5 ng/mL G-CSF. Prior to the assays, exponentially growing cells were washed free of G-CSF and resuspended in RPMI 1640 with 5% fetal calf serum. Samples containing  $5 \times 10^4$  cells were incubated in 100 mL of RPMI 1640 with 10% fetal calf serum containing solution released from hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles. After 48 hours, growth of NFS-60 cells was determined using the MTT method.<sup>37</sup>

## In vivo release study

Five male Sprague Dawley rats (6 weeks old, average weight 200 g) underwent subcutaneous implantation of a hemostatic gauze scaffold loaded with native G-CSF dextran nanoparticles, a G-CSF-loaded hemostatic gauze scaffold using native G-CSF solution and PLGA suspension, or a pure hemostatic gauze scaffold at 0.15 mg/kg. A 0.4 mL blood sample was collected using a heparinized syringe from each animal at 1, 2, 3, 4, 5, 6, 7, 8, 10, 12, and 15 days after implantation. The blood samples were centrifuged at rate of 12,000 rpm at 4°C for 20 minutes, and then plasma samples were collected. Plasma G-CSF levels were determined using a G-CSF enzyme-linked immunosorbent assay kit.

## In vivo efficacy study

For the in vivo efficacy study, 6-week-old Sprague Dawley rats underwent subcutaneous implantation of a hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles, a hemostatic gauze scaffold loaded with native G-CSF solution and PLGA suspension, a subcutaneous injection of native G-CSF solution at 0.15 mg/kg, or physiological saline. A 0.2 mL blood sample was collected from each rat using a heparinized syringe at 1, 2, 3, 4, 5, 6, 7, 8, 10, 12, and 15 days after administration. Neutrophil counts were determined as described previously.<sup>38</sup> The percentage of neutrophils was determined using a complete blood count (Beckman Coulter LH, Fullerton, CA), and the increasing ratio of neutrophils per  $\mu\text{L}$  of blood was measured using the neutrophil percentage of G-CSF samples divided by neutrophil percentage of physiological saline samples.

To observe the hemostatic efficacy of the hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles, we implanted three pieces of hemostatic gauze scaffold, ie, one piece of pure hemostatic gauze scaffold, one piece containing G-CSF-loaded dextran nanoparticles, and a piece of pure hemostatic gauze scaffold adhered to the surface of a

hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles, and then measured the time to hemostasis.

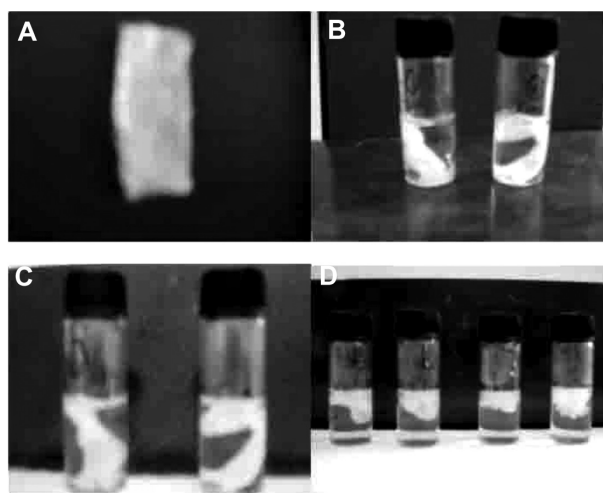
## Statistical analysis

The data were expressed as the mean  $\pm$  standard deviation, and statistically significant differences were sought using the Student's *t*-test.  $P < 0.05$  was considered to be statistically significant. Statistical analyses were carried out using SPSS software (SPSS Inc, Chicago, IL).

## Results

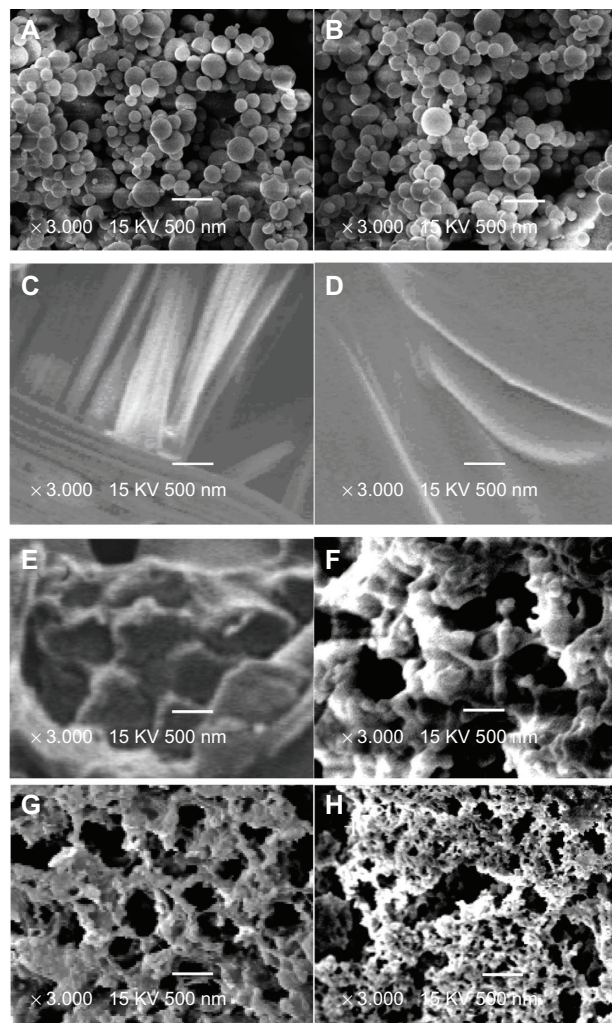
### Morphology and size distribution

The morphology of the samples was characterized using SEM and an Olympus  $\mu\text{710}$  camera. Figure 1 shows the morphology of the hemostatic gauze scaffolds containing different coatings at different release times. We found that the implants started to degrade on day 2 and the hemostatic gauze in the scaffold implants was completely broken down by day 7.<sup>39</sup> Figure 2F, G, and H confirm this finding. Figure 2A and B are images for G-CSF-loaded dextran nanoparticles alone and G-CSF-loaded dextran nanoparticles recovered from the scaffold, respectively. The results show that the surfaces of both G-CSF-loaded dextran nanoparticles alone and those recovered from the scaffold were smooth and nonporous, with the same size and morphology. Figure 2C and D are images for a blank hemostatic gauze scaffold and a scaffold containing G-CSF-loaded dextran nanoparticles, respectively. The hemostatic gauze scaffold had a fibrous structure in which groups of fine fibers formed larger bundles of fibers. The hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles regardless of 5% G-CSF-loaded dextran nanoparticles



**Figure 1** Hemostatic gauze scaffold implants. (A) Implant, (B) release on day 1, (C) release on day 2, and (D) release on day 7.





**Figure 2** Scanning electron micrographs of different samples (A) G-CSF dextran nanoparticles, (B) G-CSF dextran nanoparticles from hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles, (C) blank hemostatic gauze scaffold, (D) hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles, and (E) cross-section from hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles. (F) release on day 1, (G) release on day 2, and (H) release on day 7. **Abbreviation:** G-CSF, granulocyte-colony stimulating factor.

and PLGA lead to complete coverage of the tops of the fiber bundles, and as a result, the fibrous structure became invisible (Figure 2D). A cross-section of a hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles showed that the nanoparticles and PLGA contained gaps (Figure 2E). Figure 2F, G, and H showed that the scaffold itself degraded and became more porous. The mean diameter of the G-CSF-loaded dextran nanoparticles was  $100 \pm 20$  nm, with the majority of nanoparticles ranging from 60 to 100 nm.

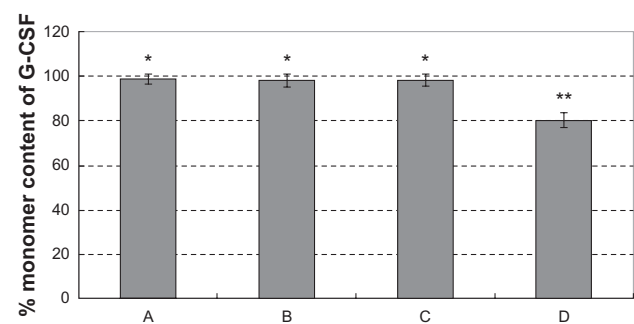
## G-CSF encapsulation efficiency from coating of hemostatic gauze implants

G-CSF encapsulation efficiency was measured using a human G-CSF enzyme-linked immunosorbent assay kit.

The encapsulation efficiency of the first-step particles, ie, loading the proteins into the dextran nanoparticles, was above 95% (w/w),<sup>32</sup> and also for the subsequent two steps. The G-CSF loading yield was 96% for the samples projecting 1% G-CSF loads (ie, G-CSF reached 1% of the total mass of the coating material).

## Aggregation study in hemostatic gauze scaffolds containing G-CSF-loaded dextran nanoparticles

Formation of protein aggregates gives a good indication of protein integrity during the microencapsulation process.<sup>40</sup> Therefore, we assayed the samples using size-exclusion chromatography-HPLC in order to determine any change in G-CSF with the different procedures. Figure 3 shows the size-exclusion chromatography-HPLC results for G-CSF aggregation after each preparation step. Figure 3A, B, C, and D show the monomer contents of the original G-CSF solution, G-CSF from the dextran nanoparticles, recovery from hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles, and from hemostatic gauze scaffold loaded directly with G-CSF solution and PLGA suspension, respectively. The monomer content of G-CSF did not decrease during the coating process compared with the original sample, except that the monomer content of G-CSF in the hemostatic gauze scaffold loaded with G-CSF decreased by 20%. These results strongly suggest that the implant preparation process used in the present study is a useful method for coating hemostatic gauze scaffold implants without aggregation.



**Figure 3** Monomer content of recovery G-CSF from the scaffold by size exclusion chromatography-high pressure liquid chromatography ( $n = 5$ , \* $P > 0.05$ , \*\* $P < 0.05$ ). (A) G-CSF (G-CSF:dextran =  $1.4:5.0 \pm 0.3$  mg) solution; (B) G-CSF from G-CSF-loaded dextran nanoparticles; (C) G-CSF from hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles (PLGA 50/50 3A: first coating =  $20.0 \pm 0.5$  mg PLGA, G-CSF-loaded dextran nanoparticles (G-CSF:dextran, 1:4) =  $5.0 \pm 0.3$  mg, second coating (blank) =  $14.0 \pm 0.6$  mg PLGA); (D) G-CSF from hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles (PLGA 50/50 3A: first coating =  $20.0 \pm 0.5$  mg PLGA, G-CSF solution [G-CSF:dextran = 1:4] =  $5.0 \pm 0.3$  mg, second coating [blank] =  $14.0 \pm 0.6$  mg PLGA).

**Abbreviations:** G-CSF, granulocyte-colony stimulating factor; PLGA, polylactic-co-glycolic acid.

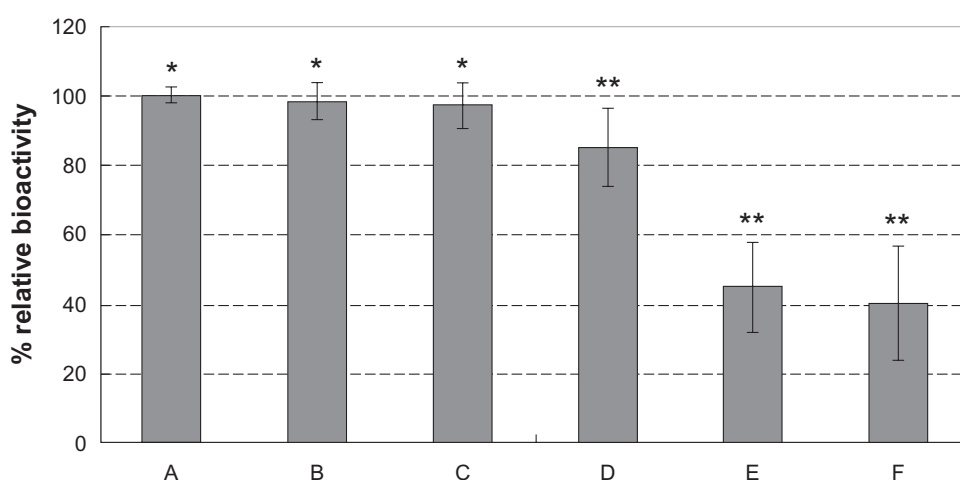
## Bioactivity of hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles

In order to determine the bioactivity of the different samples, G-CSF was recovered from the formulations according to the method used for determination of G-CSF encapsulation efficiency. The bioactivity of G-CSF was assayed using the NFS-60 cell line. Figure 4 shows the bioactivity of G-CSF from the different samples, respectively. Figure 4A, B, and C shows the bioactivity of the original G-CSF solution, G-CSF from G-CSF-loaded dextran nanoparticles, and G-CSF from the hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles (PLGA 50/50 3A), respectively. We found that the bioactivity of the samples was almost the same as for the original G-CSF solution ( $P > 0.05$ ). Figure 4D, E, and F show the bioactivity of G-CSF from a hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles (PLGA 50/50 1A), a hemostatic gauze scaffold loaded with G-CSF solution (PLGA 50/50 3A), and a hemostatic gauze scaffold loaded directly using G-CSF solution and PLGA suspension (PLGA 50/50 1A), respectively. The results show that the recovered bioactivity of G-CSF from a hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles (PLGA 50/50 1A), a hemostatic gauze scaffold loaded with G-CSF solution and PLGA suspension (PLGA 50/50 3A and PLGA 50/50 1A, respectively) was decreased by 17%, 55%, and 60%, respectively ( $P < 0.05$ ). The recovered bioactivity suggested that the G-CSF in

the G-CSF-loaded dextran nanoparticles had been well protected during the preparation process. This is probably because G-CSF-loaded dextran nanoparticles might avoid oil-water and water-gas interfaces which result in loss of G-CSF bioactivity.<sup>31</sup>

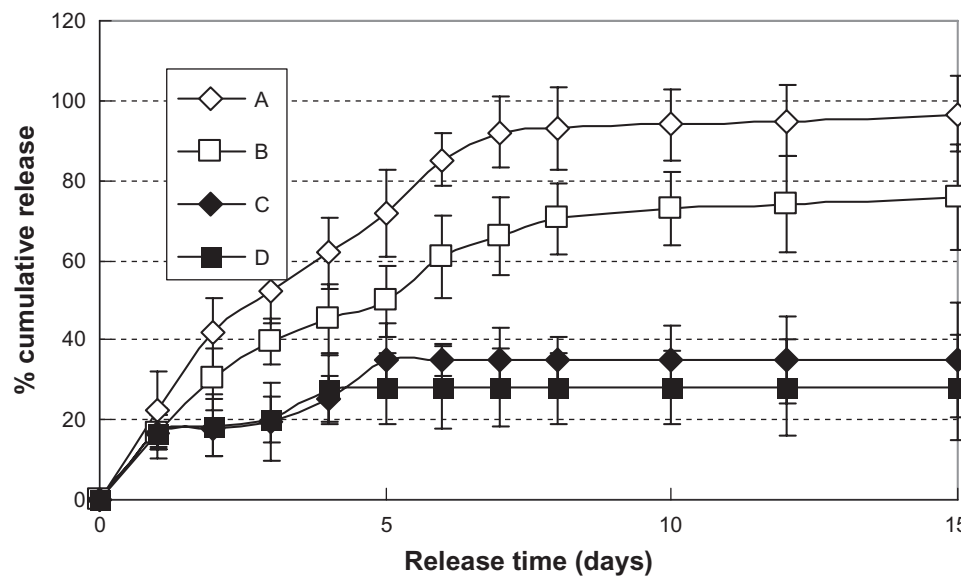
## Release kinetics for hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles

Figure 5A and B summarize the G-CSF release profiles for hemostatic gauze scaffolds containing PLGA coatings of different molecular weights. Interestingly, as shown in Figure 5, the two samples showed a similar zero order G-CSF release profile regardless of molecular weight (release kinetics equations:  $Q_A = 12.574t + 9.4172$ ,  $R^2 = 0.9705$  and  $Q_B = 8.90003t + 7.4446$ ,  $R^2 = 0.962$ , from day 1 to day 7), but the amount of cumulative release was less from the low molecular weight polymer than from the high molecular weight polymer. We reported the result of the degradation study by plotting the weight of each formulation against the release incubation time at 37°C. When a piece of hemostatic gauze scaffold containing no coating (2.5 cm × 1 cm, 18 mg) was incubated in 2 mL of phosphate-buffered saline at 37°C, the total mass completely disappeared within 3 days.<sup>39</sup> As shown in Figures 1, 2F, G, and H, hemostatic gauze scaffolds coated with G-CSF-loaded dextran nanoparticles and a PLGA layer of different molecular weights or with an additional blank PLGA layer all broke down during the same time period, suggesting that the rate of release of G-CSF



**Figure 4** Recovery bioactivity from a hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles ( $n = 5$ ,  $*P > 0.05$ ,  $**P < 0.05$ ). Samples (A), (B), (C), and (E) were the same as for Figure 3A, B, C, and D, respectively ( $n = 5$ ,  $*P > 0.05$ ,  $**P < 0.05$ ). (D) Hemostatic gauze scaffold containing G-CSF from G-CSF-loaded dextran nanoparticles (PLGA 50/50 1A: first coating =  $20.0 \pm 0.6$  mg PLGA, G-CSF loaded-dextran nanoparticles [G-CSF: dextran = 1:4] =  $5.0 \pm 0.2$  mg, second coating [blank] =  $14.0 \pm 0.5$  mg PLGA), (F) G-CSF from hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles (PLGA 50/50 1A: first coating =  $20.0 \pm 0.6$  mg PLGA, G-CSF solution [G-CSF:dextran = 1:4] =  $5.0 \pm 0.2$  mg, second coating [blank] =  $14.0 \pm 0.5$  mg PLGA).

**Abbreviations:** G-CSF, granulocyte-colony stimulating factor; PLGA, polylactic-co-glycolic acid.



**Figure 5** In vitro release profiles of hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles ( $n = 5$ ,  $P < 0.05$ ). Samples (A), (B), (C), and (D) were the same as for Figure 3(C), (D), (E), and (F), respectively.

was due to degradation of the hemostatic gauze scaffold and independent of the PLGA coating. When G-CSF solution was loaded into hemostatic gauze scaffold using PLGA coating, the percentage cumulative release was only 30%–40%. This was probably because the spraying and painting process produced a number of oil-water and gas-water interfaces, resulting in G-CSF aggregation and more hydrophobicity, with no release of G-CSF.<sup>31</sup>

### In vitro G-CSF release bioactivity in hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles

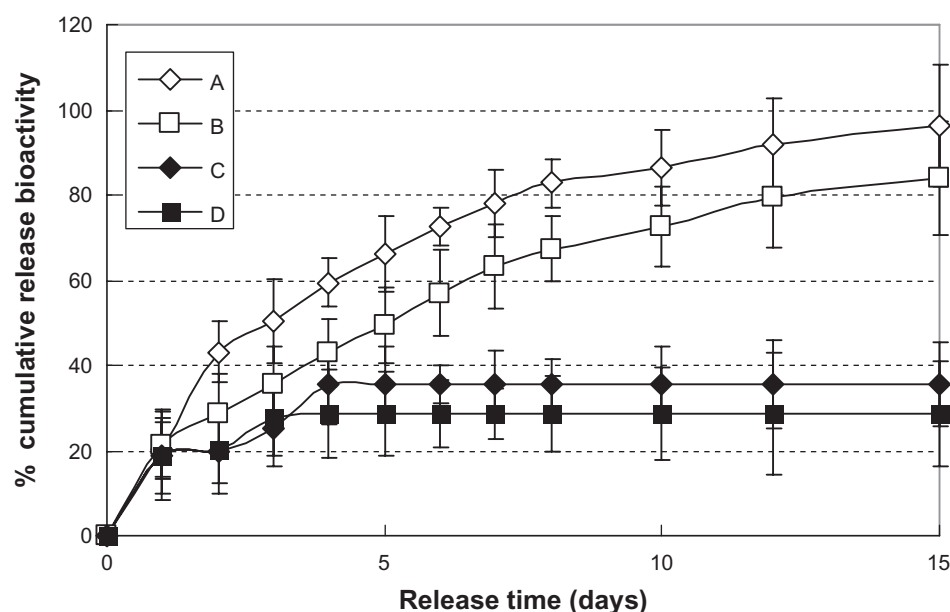
The bioactivity of in vitro release from hemostatic gauze scaffolds containing G-CSF-loaded dextran nanoparticles was investigated using the NFS-60 cell line. Figure 6A and B show that the bioactivity of release of G-CSF from hemostatic gauze scaffolds containing G-CSF-loaded dextran nanoparticles (PLGA 50/50 3A and PLGA 50/50 1A) showed high efficiency. The release bioactivity percent of G-CSF on the first day was 20% of the cumulative release bioactivity of G-CSF, and the percent of cumulative release bioactivity was about 80%–96% of the total bioactivity within the scaffold coating. When the G-CSF solution was loaded into hemostatic gauze scaffolds with PLGA coating, the percent cumulative release bioactivity of G-CSF was only 30%–40%. This was probably because the spraying and painting process produced a number of oil-water and gas-water interfaces resulting in G-CSF aggregation and more hydrophobicity.<sup>31</sup>

### Release of G-CSF in an in vivo rat study

We examined the release of G-CSF from the G-CSF-loaded dextran nanoparticles and from a hemostatic gauze scaffold loaded directly using G-CSF solution and PLGA suspension, because the G-CSF released is only bioactive in vivo. Figure 7  $\diamond$  and  $\blacklozenge$  show the plasma G-CSF concentration-time release profiles for G-CSF-loaded dextran nanoparticles and hemostatic gauze scaffold loaded directly with G-CSF solution and PLGA suspension, respectively. The results showed a stable plasma concentration from days 1 to 7, and then slow release of G-CSF up to day 15 from hemostatic gauze scaffolds containing G-CSF-loaded dextran nanoparticles. However, scaffolds loaded directly with G-CSF solution and PLGA suspension did not show stable plasma concentrations, with fast release of G-CSF from day 1 to day 3, and no G-CSF release thereafter. This confirms that the hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles was more effective than the scaffold containing the directly loaded G-CSF solution.

### In vivo efficacy study

To compare the in vivo efficacy of scaffolds containing G-CSF-loaded dextran nanoparticles, scaffolds containing directly loaded G-CSF solution, pure scaffolds, and native G-CSF alone, male Sprague Dawley rats underwent subcutaneous implantation with scaffolds loaded with dextran nanoparticles, scaffolds loaded with G-CSF, pure scaffolds, or subcutaneous injection of native G-CSF at 0.15 mg/kg. Changes in neutrophil count were determined



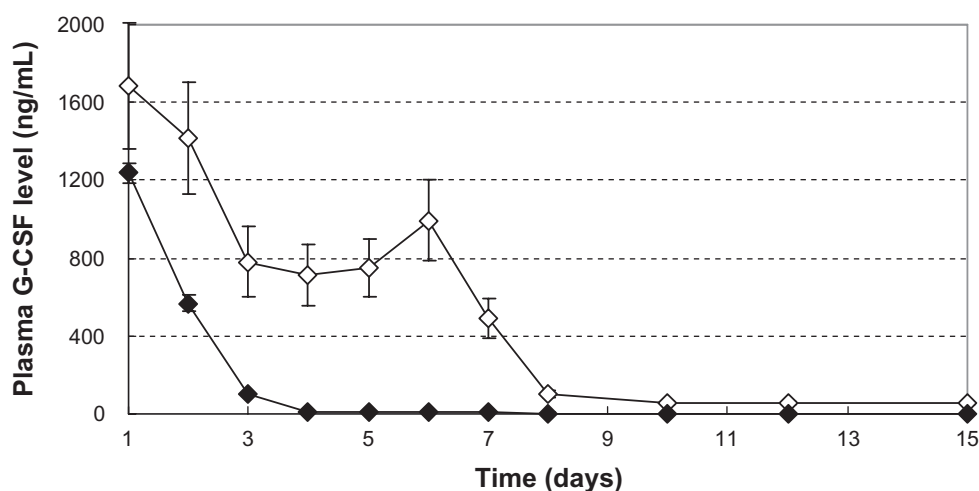
**Figure 6** In vitro release relative bioactivity of hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles ( $n = 5$ ,  $P < 0.05$ ). Samples (A), (B), (C), and (D) were the same as for Figure 3(C), (D), (E), and (F), respectively.

**Abbreviation:** G-CSF, granulocyte-colony stimulating factor.

for 15 days. The time courses in Figure 8 ▲, ◇, and ◆ show that scaffolds containing G-CSF-loaded dextran nanoparticles or G-CSF solution achieved a significant increase in neutrophil count for a prolonged period of time, but the increasing ratio of neutrophil count and prolonged time from G-CSF solution-loaded the scaffold were more than G-CSF solution-loaded the scaffold (the pure scaffold did not show an increase in neutrophil count, data not shown).

We found that the time to hemostasis for the scaffold containing G-CSF-loaded dextran nanoparticles was longer

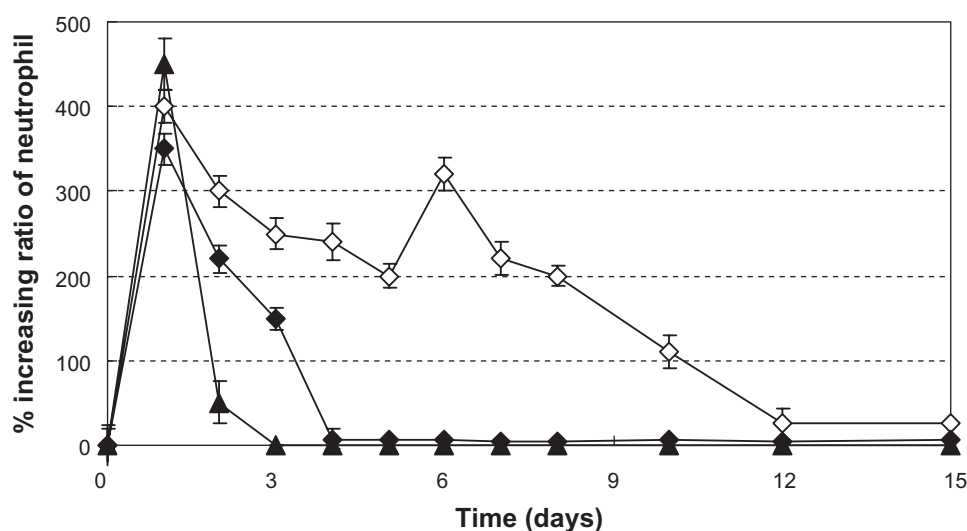
than that for the same-sized pure hemostatic gauze scaffold and pure hemostatic gauze scaffold adhered to a scaffold loaded with dextran nanoparticles. However, the time to hemostasis from both a pure hemostatic gauze scaffold and a pure hemostatic gauze scaffold adhered to the surface of a scaffold loaded with dextran nanoparticles was almost the same ( $P < 0.05$ ). This suggests that the scaffold loaded with dextran nanoparticles might work for both increasing the neutrophil count and stopping bleeding when the pure hemostatic gauze scaffold was attached and adhered to the surface of a scaffold loaded with dextran nanoparticles.



**Figure 7** In vivo release plasma G-CSF level of hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles ( $n = 5$ ,  $P < 0.05$ ). Samples ◇ and ◆ were the same as for Figure 3(C) and (E), respectively.

**Abbreviation:** G-CSF, granulocyte-colony stimulating factor.





**Figure 8** Increasing ratio of in vivo neutrophil levels ( $n = 5$ ,  $P < 0.05$ ). ▲, ◇, and ◆ were the same as for Figure 3(A), (C), and (E), respectively.

## Discussion

A zero order release profile for G-CSF was observed from the scaffolds containing G-CSF-loaded dextran nanoparticles, regardless of the molecular weight of the PLGA coating used from days 1 to 7 (Figures 5A and B). This finding suggests that the rate-limiting step for G-CSF release was not just controlled by PLGA, but by other factors, including the dextran nanoparticles and the scaffold. The pure scaffold is easily degraded, taking 1–2 weeks to break down into molecular pieces (<http://store.k12webstore.com/mkgz24.html>), and dextran nanoparticles also absorb water and swell.<sup>43</sup> These observations suggest that pores or holes in the surface of the PLGA coating can be created when a pure hemostatic gauze scaffold with PLGA is degraded and broken fragments form.<sup>39</sup> Rapid degradation of the hemostatic gauze scaffold creates sufficient penetration of water diffusion channels for G-CSF release of all implants coated with PLGA of different molecular weights.<sup>39</sup> At the same time, the G-CSF diffusion distance from the inner to outer coating decreases due to swelling of the dextran nanoparticles through absorption of water compared with no swelling additives.<sup>42</sup>

These resulting in that at the beginning of the release phase, G-CSF were near the coating surface of the G-CSF-loaded dextran nanoparticle scaffolds, then the hemostatic gauze scaffold started to degrade and formed more and more new holes in the coating that led to water being permeated into the coating and increased diffusion of G-CSF and dextran swelling (of course, finally dextran also was dissolved and diffused, the PLGA also was degraded, but PLGA degraded slower than the pure scaffold during the whole release process). With these diffusion channels, the diffusion rate

of G-CSF became significantly higher than that of high molecular weight dextran, thus the overall release rate was limited by the swelling of dextran. Figure 2F–H also confirmed the process.<sup>39</sup> The linear release from the G-CSF-loaded dextran nanoparticle scaffold appeared attributable to the above effects.<sup>44–46</sup>

Figures 5A and B, 6A and B also show that the degradation rate of PLGA did not affect the release rate or cell proliferation activity of the G-CSF released or the cumulative amount released from the scaffolds containing G-CSF-loaded dextran nanoparticles. The cumulative amount released from scaffolds containing G-CSF-loaded dextran nanoparticles with PLGA of higher molecular weight was more than that of PLGA of lower molecular weight. This was possibly because G-CSF was unstable in a locally acidic environment,<sup>41,43</sup> in which the degradation rate of lower molecular weight PLGA was faster than for higher molecular weight PLGA which, in turn, formed a more acidic local environment.<sup>41,43</sup> Figure 5C and D show further release kinetic profiles which were probably because G-CSF had aggregated and become more hydrophobic, resulting in no release of G-CSF,<sup>43</sup> suggesting that the spray-painted coatings had produced a number of oil-water and gas-water interfaces.<sup>31</sup>

In order to understand the mechanism of the scaffold, a pure hemostatic gauze scaffold was attached and adhered to the surface of a hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles for study of in vitro and in vivo release. We found that the release profile, plasma G-CSF level, and increase in neutrophil count were the same for a hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles (data not shown), but that the time to hemostasis

was shorter. This suggested that the pure hemostatic gauze scaffold does not affect the activity of the G-CSF-loaded dextran nanoparticle scaffold in terms of release of G-CSF, but may improve hemostasis activity.

By coating the gauze scaffold with G-CSF-loaded dextran nanoparticles and PLGA suspension, sustained-release delivery of the drug can be achieved easily. In addition, the desired release kinetics, such as a low level of initial burst release and a zero order release profile can also be easily achieved. With the right thickness, the G-CSF-loaded dextran nanoparticle-free top layer of PLGA may provide additional control of G-CSF release mechanisms. The PLGA coating on the hemostatic gauze scaffold can easily be attached and applied to the wound surface during surgery to stop post-surgical bleeding and to improve the neutrophilic granulocyte count in blood during chemotherapy as well as to staunch bleeding after tumor resection.

## Conclusion

Using a clinically available hemostatic agent as a scaffold, we demonstrated that a hemostatic gauze scaffold containing G-CSF-loaded dextran nanoparticles can achieve controlled delivery of G-CSF. G-CSF were released from this formulation through a mechanism of diffusion channels independent of G-CSF-loaded different molecular weight PLGA that was created due to rapid degradation of the hemostatic gauze scaffold inside the PLGA coating and dextran microparticle swelling. Such diffusion channels could be blocked by an additional coating of G-CSF-loaded dextran nanoparticle-free PLGA to reach the desired G-CSF release rate. In addition to nearly zero order and burst-free release kinetics, this formulation approach was simple and did not require a complicated microencapsulation process, thus could easily be scaled up and potentially used to improve the neutrophilic granulocyte level in blood at the time of chemotherapy and staunch bleeding after tumor resection.

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## Disclosure

The authors declare no competing financial interests in this work.

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