

Cytotoxicity of prostaglandin analog eye drops preserved with benzalkonium chloride in multiple corneoconjunctival cell lines

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Purpose: This study evaluated the cytotoxicity of five prostaglandin analog ophthalmic solutions on four ocular surface cell lines, ie, Chang (human conjunctiva), SIRC (rabbit cornea), RC-1 (rabbit cornea), and BCE C/D-1b (bovine cornea).

Methods: Cell viability was measured by neutral red and MTT assays in cells treated for 10, 30, or 60 minutes with various doses of prostaglandins (undiluted, and 2- and 10-fold dilutions). The number of cell lines with viability $\geq 50\%$ in the presence of selected dilution of the drug (CVS_{50}) was used for comparison. In addition, 24 cell viability comparisons (four cell lines, two assays, and three exposure times) were made between latanoprost (Xalatan[®]) and each other solution at each dose. A comparison between the newly introduced tafluprost (Tapros[®]) with 0.01% benzalkonium chloride was also made.

Results: The order of cell viability determined by CVS_{50} was Travatan Z[®] (travoprost with the SofZia system) > Tapros \geq Travatan[®] (travoprost) = Xalatan > Rescula[®] (unoproston). This was consistent with the results of direct comparisons between Xalatan and the other drugs. There was no clear difference in cell viability between Tapros and benzalkonium chloride.

Conclusions: Use of two assays, multiple cell lines, and various dilutions and exposure times provided a unique evaluation of cytotoxicity among ophthalmic solutions. CVS_{50} was useful for comparison of the cell viability of the solutions.

Keywords: glaucoma, cornea, cell viability score

Introduction

Prostaglandin analogs (PGs) form a major part of treatment regimens for glaucoma.¹ PGs successfully reduce ocular pressure,²⁻⁵ with few common side effects including conjunctival hyperemia⁶⁻⁸ and increased iris pigmentation.⁹⁻¹³ Cytopathic evaluations of PGs in conjunctival cells¹⁴⁻¹⁸ have indicated that latanoprost (Xalatan[®]) and travoprost (Travatan[®]) could be cytoprotective against benzalkonium chloride toxicity via an antioxidative effect.¹⁵ Benzalkonium chloride is commonly included as a preservative in ophthalmic solutions,¹⁹⁻²¹ although Travatan Z[®] (also travoprost) is preserved with a SofZia system that has proven to be less cytotoxic.²¹ Among the PGs, Xalatan was introduced early and has become a standard medication. Tafluprost (Tapros[®])^{16,22,23} is a recently introduced PG and the cytotoxicity of preserved tafluprost is untested. We therefore evaluated the cytotoxicity of Tapros compared with that of five other PG ophthalmic agents in four corneoconjunctival cell lines using two cytotoxic assays to compare Xalatan with other available PG drugs, test for intercellular or interassay differences in toxicity, and to determine the optimal dose and exposure time for cytotoxic testing.

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Materials and methods

The cell lines used in the present study were SIRC (rabbit corneal epithelium, CCL-60, American Type Culture Collection [ATCC], Manassas, VA), BCE C/D-1b (bovine corneal epithelial cells, JCRB-9129, Health Science Research Resource Bank, Osaka, Japan), RC-1 (rabbit corneal epithelium, JCRB-0246; Health Science Research Resource Bank), and Chang conjunctiva (human conjunctival cells, ATCC CCL-20.2). All cell lines were cultured according to the standard protocols provided by the suppliers.

Approximately 2×10^4 cells were harvested from 96-well culture plates (NUNC® 167008; Thermo Fisher Scientific, Denmark) and incubated for two days. The culture medium was then replaced with undiluted, twofold, or 10-fold dilutions of the different ophthalmic drugs, and the cells were incubated for a further 10, 30, or 60 minutes. After these drug treatments, the culture medium was replaced with fresh media and cells were incubated for a further 48 hours. Cell viability was determined using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) and neutral red assays. The MTT assay is a quantitative colorimetric assay of mitochondrial activity as a measure of cell viability and proliferation; it detects living cells only, and the signal generated is directly proportional to the number of viable cells. In contrast, neutral red staining lysosomes in living cells and the results of the assay are determined spectrophotometrically (Benchmark Microplate Reader; BIO-RAD, Hercules, CA). Cell viability of treated cells is expressed as a percentage of that in control cells (incubated in culture medium alone). All experiments were repeated 8–16 times

and results are presented as the mean \pm standard deviation (SD). Statistical comparisons were made using the *t*-test by Microsoft Excel (Microsoft, Tokyo, Japan), and $P < 0.05$ was determined as significant.

We compared cell viability using the cell viability score. In a previous study,²⁴ we used CVS_{50} (ie, number of cell lines with viability $\geq 50\%$ in the presence of a selected dose of the drug) to enable easy comparison of the effects of different drugs, and found it was useful. This concept is similar to that of the MIC_{50} (ie, the minimum inhibitory concentration of a drug required to inhibit the growth of 50% of organisms). In addition, 24 comparisons of cell viability (four corneal cell lines, two assays, and three exposure times) were also made between Xalatan, a leading PG and serving as the standard, and each other solution at each dose. A comparison between Tapros and 0.01% benzalkonium chloride was also made.

We used the following solutions for cytotoxicity: 0.0015% Tapros (with 0.01% benzalkonium chloride and polysorbate 80), 0.005% Xalatan (with 0.02% benzalkonium chloride), 0.12% unoproston (Rescula®, with 0.005% benzalkonium chloride), 0.004% travoprost (Travatan, with 0.015% benzalkonium chloride), 0.004% travoprost (Travatan Z), and 0.01% benzalkonium chloride.

Results

Figures 1–4 show the cell viability results for four cell lines, and its order determined by CVS_{50} was Travatan Z \geq Tapros \geq Travatan = Xalatan $>$ Rescula (Table 1). Table 2 details the comparison between Xalatan and the other

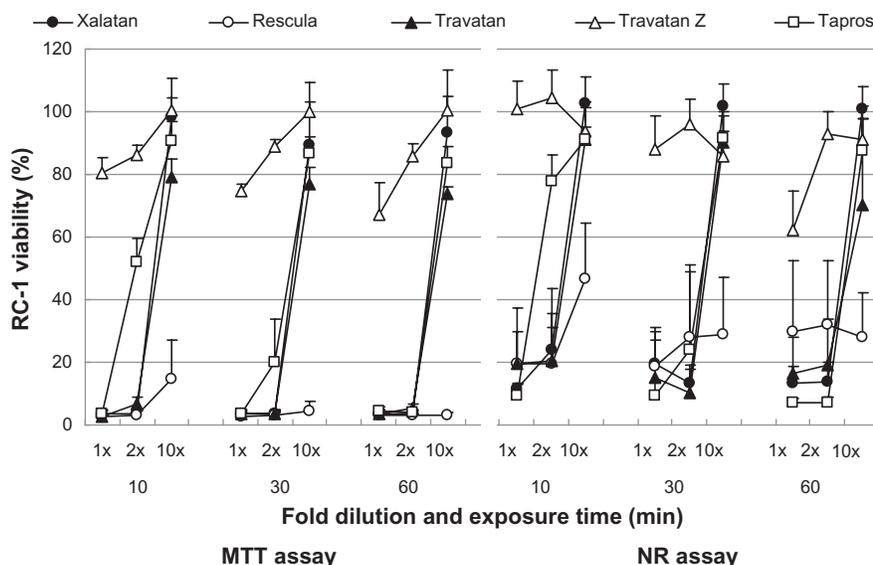


Figure 1 Effects of prostaglandin analog ophthalmic solutions on the viability of cultured rabbit corneal cells (RC-1) using the neutral red (NR) assay and MTT assay. Data are means \pm standard deviations. Note that Travatan Z exhibits the greatest viability and shows minimal differences among exposure times or dilution. The cell viabilities for the other solutions depend on exposure times and dilutions except for Rescula, which shows small cell viabilities even in diluted solutions. These are common findings in all cell lines. Detailed comparisons were made with cell viability score and statistical analysis (Tables 1–3).

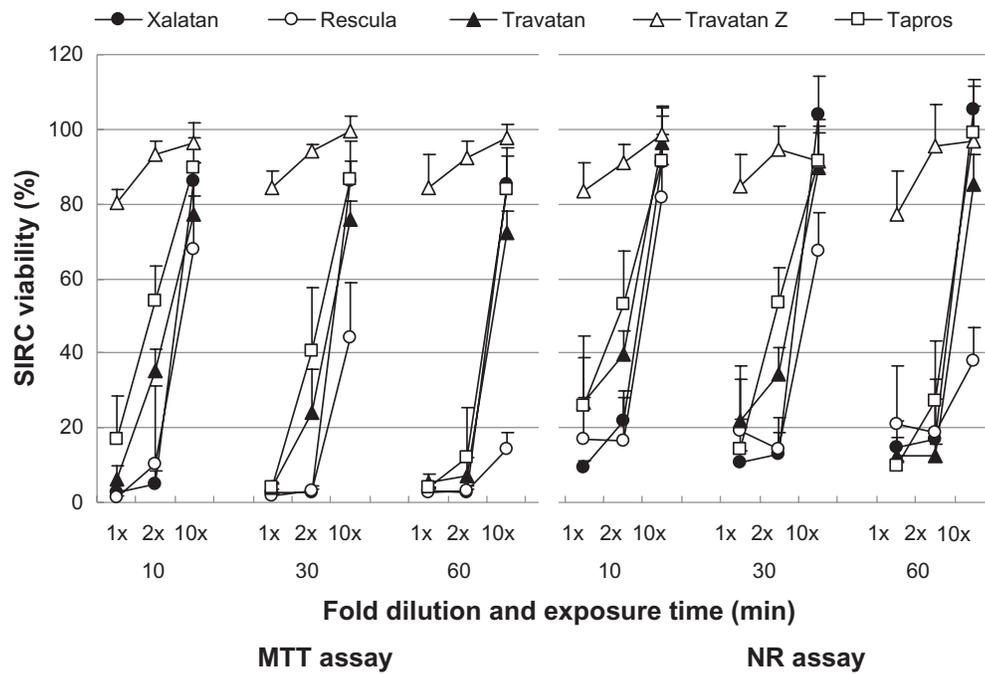


Figure 2 Effects of prostaglandin analog ophthalmic solutions on the viability of cultured rabbit corneal cells (SIRC) using the neutral red (NR) assay and MTT assay. Data are means \pm standard deviations.

drugs for all cell lines. Based on these results, cell viability was clearly the highest for Travatan Z, with only small changes seen among different dilutions and exposure times. In contrast, it was the lowest for Rescula. CVS_{50} expressed general trend of graphic presentation and detailed comparison

was consistent with CVS_{50} . The comparison between Travatan and Xalatan only exhibited conflicting results in three doses, although those two drugs had the same CVS_{50} .

The comparison of Tapros and 0.01% benzalkonium chloride (Table 3) indicated that cell viability in benzalkonium

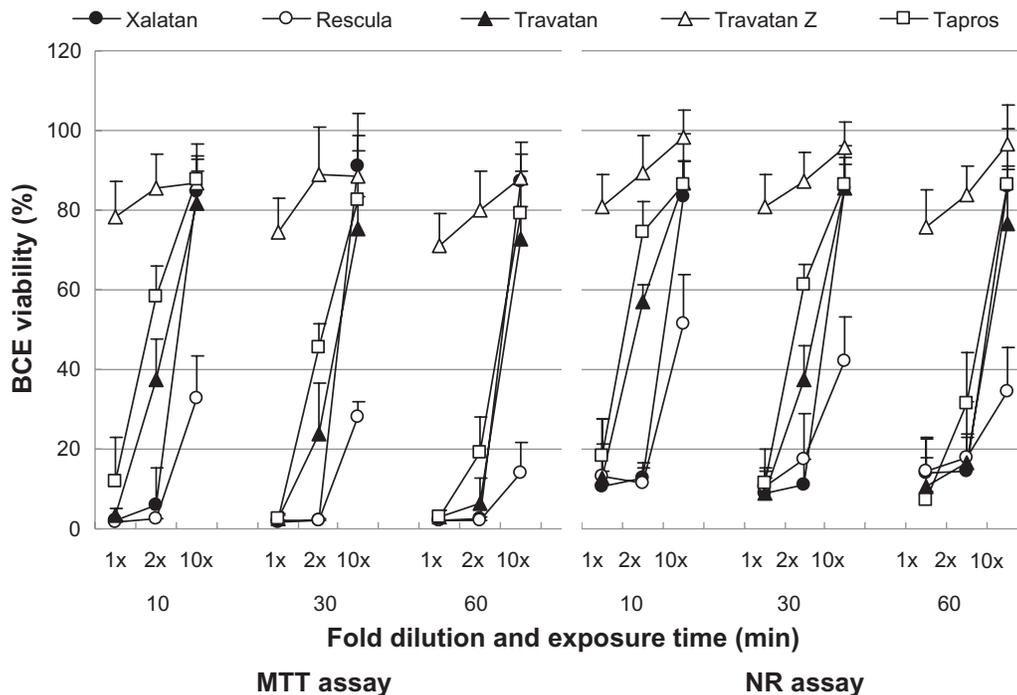


Figure 3 Effects of prostaglandin analog ophthalmic solutions on the viability of cultured bovine corneal cells (BCE C/D I-b) using the neutral red (NR) assay and MTT assay. Data are means \pm standard deviations.

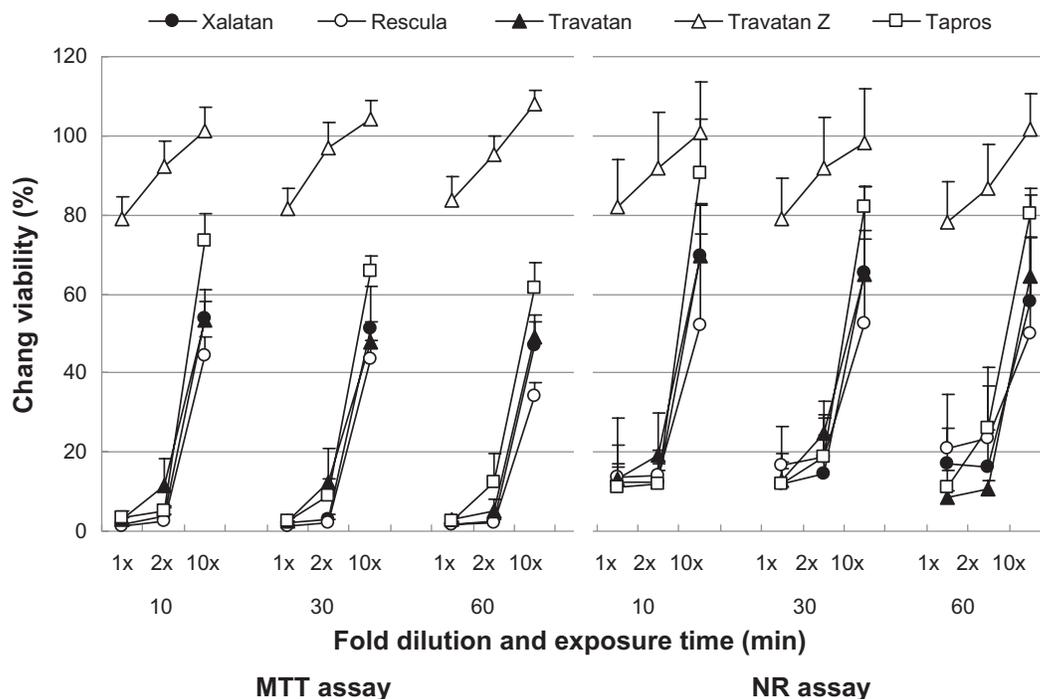


Figure 4 Effects of prostaglandin analog ophthalmic solutions on the viability of cultured human conjunctival cells (Chang) conjunctiva using the neutral red (NR) assay and MTT assay. Data are means \pm standard deviations. Travatan Z exhibited a proliferative effect (cell viability > 100%) with the 10-fold dilution.

chloride was always higher with the 10-minute treatments (8/8 experiments), but was not different (3/8) or was lower (5/8) in benzalkonium chloride after a 30-minute exposure. There was no difference in cell viability between Tapros and benzalkonium chloride with the 60-minute exposure.

The MTT and neutral red assays produced comparable results and there were no conflicting results in any comparisons. The MTT values were generally less than those obtained with the neutral red assay, especially in undiluted and twofold diluted solutions. There were also no major inconsistencies among results from the four cell lines, as shown in Figures 1–4.

Discussion

Benzalkonium chloride concentration in the various ophthalmic solutions tested in this study increased in the following order:

Travatan Z < Rescula < Tapros < Travatan < Xalatan. This increase was inversely in accordance with cell viability, except for Rescula, which was more toxic than Travatan and Tapros with more benzalkonium chloride. Unoproston in Rescula is a metabolized form of natural PG F₂ alpha and is different from the other PGs, although this study did not address whether the difference in cytotoxicity is related to this unique structure. The cytotoxicity of preserved tafluprost was first examined *in vitro* and proved comparable with Xalatan and Travatan. Considering Tapros contains toxic levels of benzalkonium chloride and polysorbate 80, the lower cell viability observed in this study after 10 minutes of exposure compared with benzalkonium chloride alone in all cell lines was not unexpected. Although cell viability in Tapros was greater than or equal to that in 0.01% benzalkonium chloride in some of comparisons, the cytoprotective

Table 1 Cell viability score of the drugs

Drugs	Fold dilution and exposure time (min)									Total of CVS ₅₀
	1 x			2 x			10 x			
	10	30	60	10	30	60	10	30	60	
Xalatan	0	0	0	0	0	0	8	8	7	23
Rescula	0	0	0	0	0	0	4	2	1	7
Travatan	0	0	0	1	0	0	8	7	7	23
Travatan Z	8	8	8	8	8	8	8	8	8	72
Tapros	0	0	0	6	2	0	8	8	8	32

Notes: The full number of assay is eight (four cell lines \times two assays for each time and dilution).

Abbreviations: CVS₅₀, number of cell lines with viability \geq 50% in the presence of the drug; min, minutes.

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Table 2 Comparison of cell viability between Xalatan and other prostaglandins

Drugs and comparison results		Summary of comparisons for each exposure time and fold dilution								
		1 ×			2 ×			10 ×		
		10	30	60	10	30	60	10	30	60
Rescula	>Xalatan*	0	1	0	0	0	0	0	3	0
	NS*	5	6	7	8	8	7	6	2	1
	<Xalatan*	3	1	1	0	0	1	2	3	7
Travatan	>Xalatan	4	2	2	6	6	3	0	0	0
	NS	4	6	5	2	2	5	4	4	3
	<Xalatan	0	0	1	0	0	0	4	4	5
Travatan Z	>Xalatan	8	8	8	8	8	8	4	5	3
	NS	0	0	0	0	0	0	4	2	4
	<Xalatan	0	0	0	0	0	0	0	1	1
Tapros	>Xalatan	4	1	0	7	6	3	3	2	2
	NS	4	6	7	1	2	4	4	5	5
	<Xalatan	0	1	1	0	0	1	1	1	1

Notes: Each comparison includes eight experiments (two assays for four cell lines); * $P < 0.05$ (result of *t*-test).

Abbreviation: NS, not significant.

effect of tafluprost on ocular surface cell lines was unclear in our experiments.

We evaluated cell viability for multiple conditions using two bioassays. The undiluted and twofold diluted solutions seemed suitable for clinically relevant comparisons of cytotoxicity, possibly because these represent levels that are stimulatory immediately after instillation. The 10-fold dilutions were not practical in this experimental setting, but could be useful in testing of long-term cytotoxicity after diluted by tears. The 10-minute exposure time represented actual residence time of the applied solution on an ocular surface and seemed to be the minimum period required for *in vitro* testing. Our results were similar between the 10- and 30-minute exposures, although the latter was deemed more convincing and clinically applicable. The differences between the drugs reduced in significance after 60 minutes of exposure due to the low viability rates. It should be noted that this experiment would not be correlated with pressure

reduction effects that work around the intraocular tissue and last for several hours. As for assays and cells, the MTT assay values were very low in some experiments; however, there were no discrepant comparisons with neutral red assay results. CVS_{50} was simple and useful to compare cell viability in tested solutions and was confirmed by detailed comparisons. We believe bioassay using multiple cell lines is a contributory method to precise results. CVS_{50} might be applicable to such toxicity experiments.

This study had some limitations. Although we used two different assays and tested four cell lines, the bioassay only evaluated cytotoxicity, thus the results should be confirmed using a series of methods. Second, toxicity of the PGs for keratoconjunctival cells was not evaluated directly, although the active component and preservative could potentially interact in the commercial eyedrop mixture. Multiple evaluations should be done to investigate the potential side effects of PGs in glaucoma patients further.

Table 3 Comparison of cell viability between undiluted Tapros and 0.01% BAK

Cell lines		Exposure time for neutral red assay			Exposure time for MTT assay		
		10 min	30 min	60 min	10 min	30 min	60 min
SIRC	Tapros	26.1 ± 12.7	14.4 ± 8.0	9.6 ± 1.4	16.7 ± 11.8	3.9 ± 1.1*	4.0 ± 1.3
	BAK	58.2 ± 19.8**	13.8 ± 7.1	8.1 ± 3.0	37.5 ± 5.8**	2.4 ± 0.7	3.8 ± 3.4
RC-I	Tapros	9.2 ± 3.6	9.1 ± 4.6	7.3 ± 1.2	3.5 ± 0.2	3.5 ± 0.9	4.3 ± 1.4
	BAK	30.2 ± 5.6**	6.8 ± 0.3	7.0 ± 0.4	26.4 ± 14.8**	3.7 ± 0.3	4.3 ± 0.7
BCE	Tapros	18.2 ± 9.2	11.5 ± 8.6	7.3 ± 2.2	11.9 ± 11.1	2.4 ± 1.2	2.8 ± 1.8
	BAK	49.8 ± 20.8**	7.4 ± 0.6	7.5 ± 0.6	33.3 ± 6.9**	1.5 ± 0.5	1.7 ± 0.2
Chang	Tapros	11.2 ± 5.9	12.0 ± 5.6*	11.0 ± 4.6	3.4 ± 1.8	2.6 ± 0.6*	2.7 ± 1.2
Conjunctiva	BAK	20.4 ± 3.6**	7.0 ± 0.7	7.5 ± 1.6	14.0 ± 3.8**	2.0 ± 0.2	2.1 ± 0.3

Notes: *Tapros-treated cells show greater viability ($P < 0.05$, *t*-test); **BAK is greater ($P < 0.05$, *t*-test).

Abbreviations: BAK, benzalkonium chloride; min, minute.

Conclusion

Using two cytotoxicity assays on four ocular surface cell lines with a series of dilutions and exposure times of test PG ophthalmic solutions enabled a unique evaluation of cytotoxicity. The results confirmed that Travatan Z without benzalkonium chloride is the least toxic drug among the ocular surface cells tested. The detailed drug comparison included the newly available preserved Tapros and the order of cell viability rate was determined accordingly. Optimal exposure times and dilutions for cytotoxicity testing of ophthalmic solutions were also discussed. CVS₅₀ was useful for comparison of cell viability in the drugs.

Disclosure

The authors report no conflicts of interest in this work.

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