

Assessment of the presence and anti-tumor potential of tumor-infiltrating lymphocytes in patients with acute myeloid leukemia

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Purpose: Assessing the possibility of finding tumor-infiltrating lymphocytes (TIL) in bone marrow of acute myeloid leukemia (AML) patients and evaluating the anti-tumor activity of these TIL against autologous AML cells.

Patients and methods: TIL were immunomagnetically isolated by using anti-CD3 from bone marrow samples of 20 patients at the presentation of AML or four weeks upon completion of chemotherapy. TIL were ex vivo expanded for two weeks and immunophenotyped. Functionality in terms of cytokine secretion and cytotoxicity was assessed by γ -interferon quantitation and Elispot assay, respectively.

Results: TIL were detected in bone marrow samples of 50% (10/20) of the patient cohort. They were noted to highly express CD137 and PD-1 and display a significantly higher anti-tumor reactivity compared to that of autologous peripheral blood lymphocytes. TIL could be expanded in co-cultures with irradiated feeder cells supplemented with interleukin (IL)-7 and IL-15.

Conclusion: Data suggested the presence of reactive γ -interferon-secreting TIL in AML patients. They are expandable and possess anti-tumor activity, which might have a great potential in the development of adoptive cellular therapy for AML.

Keywords: tumor-infiltrating lymphocytes, acute myeloid leukemia, adoptive cell therapy, immunotherapy

Introduction

Acute myeloid leukemia (AML) is the most common hematological malignancies in adults, with an incidence rate ranging from three to five cases per 1,00,000 population.^{1,2} AML is characterized by monoclonal expansion of abnormal myeloid precursors, resulting in impaired hematopoiesis and bone marrow failure.³ With the contemporary chemotherapy, the mortality rate is high especially among patients above 60 years of age, being approximately 70% within one year of diagnosis.^{4,5} Besides, less than 50% of AML patients achieved a complete response, and in most cases, the disease relapsed quickly and became refractory to standard treatment.⁶ Allogeneic hematopoietic stem cell transplantation (allo-HSCT) is the effective therapy available to refractory and relapsed AML patients.⁷⁻⁹ However, serious adverse events and graft-versus-host disease are often associated with allo-HSCT.¹⁰ Many AML patients are also not eligible for allo-HSCT. Undoubtedly, new treatment modalities are urgently needed.

Immunotherapies might be good alternatives, but reports of the clinical outcomes available in the literature were not very encouraging.¹¹⁻¹⁵ The human immune system can recognize and eliminate autologous tumor cells expressing

“nonself” antigens.¹⁶ Malignant tumors are known to have genetic mutations, some of which mediate neo-epitopes, provoking an immune response of host defense.¹⁷ Neo-epitopes are “non-self” so they may trigger T cell responses. Indeed, neoantigen-reactive T cells which infiltrated human malignant tumors were detected.^{18–21} Adoptive cell therapy (ACT) with tumor-infiltrating lymphocytes (TIL) was noted to be effective in the treatment of metastatic melanoma patients, leading to response rates to 50%.^{22,23} In addition, it was reported that adoptive transfer of mutation-reactive CD4⁺ T cells mediated a durable and ongoing regression of metastatic diseases of more than two years in a patient with metastatic cholangiocarcinoma.²⁴ However, it is unclear whether TIL are present in AML patients and how the naturally occurring and mutation-specific T cells interact with autologous leukemic counterparts.

Like solid tumors, AML cells may have gene mutations. Some of which may possess neo-antigens and become “non-self” that can trigger T cell responses. In this study, we assessed TIL in AML patients and examined the potential significance in the ACT of AML.

Patients and methods

Patient samples

The study was approved by the Institutional Review Board of Harbin Medical University and conducted according to the Declaration of Helsinki. Patients were at least 18 years of age at presentation of AML or four weeks upon completion of chemotherapy. Patients with a life expectancy of less than six months, impaired organ function, having undergone allo-HSCT were excluded. Bone marrow and peripheral blood samples were taken with written informed consents of the patients. All patients had disease-relevant gene mutations examined by next-generation sequencing at the time of enrollment. Mutations in the following 127 genes were detected in the indicated number of patients (Supplementary materials, Methods).

Enrichment and expansion of TIL

Bone marrow mononuclear cells (BMMC) and peripheral blood mononuclear cells (PBMC) were enriched by density-gradient centrifugation using Ficoll-Hypaque (Supplier, Country). Cells were then labeled with CD3 microbeads (Miltenyi Biotech, Germany) for 15 mins at 4°C–6°C. Immunomagnetic device VarioMACS (Miltenyi Biotech, Germany) was used for cell selection according

to the manufacturer’s protocol. CD3⁺ cells were considered AML TIL, whereas CD3⁻ cells were considered autologous AML cells.

BMMC and PBMC were seeded at 1×10^6 cells/mL in X-VIVO 15 medium (Lonza, Switzerland) supplemented with 5% heat-inactivated human serum, 100 ng/mL anti-CD3, 1000 IU/mL of interleukin (IL)-2, 10 ng/mL of IL-7 and 10 ng/mL of IL-15 (PeproTech, USA) and co-cultured with 50 Gy-irradiated PBMC derived from unrelated donors as feeder cells at the ratio (10:1) of feeder cells to BMMC or PBMC for 14 days at 37°C. Fresh X-VIVO 15 medium with cytokines was added whenever the medium turned yellow.

AML TIL were cultured with autologous AML cells at the ratio of 1:1 in X-VIVO 15 medium with 5% heat-inactivated human serum, 10 ng/mL of IL-7 and 10 ng/mL of IL-15 overnight at 37°C. Cultured cells were washed with phosphate-buffered saline (PBS) and labeled with CD137 microbeads. Immunomagnetic sorting was conducted using the selection kit according to the manufacturer’s instructions (Miltenyi Biotech, Germany).

Immunophenotyping

Cells were stained with anti-human CD3-APC, anti-human CD4-PE, anti-human CD8-FITC, anti-human CD56-PE, anti-human CD279-APC, and anti-human CD137-PE (BD Biosciences, USA) for 15 mins at 4°C. They were then washed with and re-suspended in PBS. Events were acquired using a flow cytometer (Calibur, BD Bioscience) and analyzed by Cell Quest software (BD Biosciences).

Elispot assay

Assays were performed according to the manufacturer’s protocol (Mabtech, Nacka, Sweden). In brief, AML TIL of 1×10^5 were mixed with equal numbers of autologous AML cells in 200 μ L of X-VIVO 15 medium (Lonza, Switzerland) and seeded onto wells of Interferon- γ (IFN- γ) Elispot plate pre-coated with monoclonal antibody IFN- γ . Having cultured overnight at 37°C in a 5% CO₂ incubator, the plate was washed. Spots were detected and visualized by sequential incubation with the antibody 7-B6-1-biotin, streptavidin-ALP and the substrate BCIP/NBT-plus. Spots were counted using an ELISPOT Plate Reader (ImmunoSpot, CTL-Europe GmbH). Assays were run in duplicate.

IFN- γ release assay

AML TIL or AML activated peripheral blood lymphocytes (aPBL) of 1×10^5 were co-cultured with equal numbers of autologous AML cells in 200 μ L of X-VIVO 15 medium

on 96-well plates overnight at 37°C. Cultures in triplicate were harvested and centrifuged. Supernatant was collected. Secreted IFN- γ were assayed in triplicate by enzyme-linked immunosorbent assay according to the manufacturer's instructions (R&D, USA).

Cell-mediated cytotoxicity assay

Leukemic cell line K562 and NB4 were obtained from America Type Culture Collection (Manassas, VA, USA). Cytosine arabinoside (Actavis S.P.A, Italia) was used as a positive control. AML TIL or AML aPBL of 2×10^5 and K562 cells or NB4 cells of 2×10^4 at the ratio of 10:1 were co-cultured for 4 hrs at 37°C in 200 μ L of X-VIVO 15 medium. Cultures were harvested. K562 cells and NB4 cells (at a density of 2×10^5 cells/mL) were treated with cytosine arabinoside (Ara-C) at a final concentration of 20 μ M in 100 μ L of X-VIVO 15 medium. Supernatant was collected upon centrifugation. The cytosolic enzyme, lactate dehydrogenase, was assayed in triplicate using CytoTox 96R kit according to the manufacturer's instructions (Promega, USA).

Statistical analysis

Data are presented as mean and SD, unless stated otherwise. The non-parametric two-tailed Student's *t*-test was used to analyze variables. Differences in values between tested groups were regarded as significant at a value of $p \leq 0.05$.

Results

Patient characteristics

Twenty AML patients (13 patients with newly diagnosed AML and seven patients with relapsed AML) of a median year of age of 38 (range: 19–56) were enrolled to the study between March 2017 and July 2018 (Table 1). Blast counts of bone marrow samples at disease presentation ranged from 20% to 97% of nucleated cells. Cytogenetic analysis of bone marrow of nine patients exhibited a normal karyotype accounting to 45% (9/20) of the patient cohort and gene mutations were detected in 80% (16/20) of the patients. None had undergone transplantation.

Presence and frequency of TIL

AML TIL, which were CD3+ and capable to produce IFN- γ , were detected in bone marrow samples of 50% (10/20) of the patient cohort (Figure 1A and B). Positive Elispot responses were identified in Pt. 1, Pt. 2, Pt. 3, Pt. 6, Pt. 7,

Pt. 10, Pt. 13, Pt. 14, Pt. 15, and Pt. 16. Negative Elispot responses were identified in Pt. 4, Pt. 5, Pt. 8, Pt. 9, Pt. 11, Pt. 12, Pt. 17, Pt. 18, Pt. 19, and Pt. 20. The spot counts of AML TIL were shown in Table S1. The median spot counts were 152 (range: 70–250).

Expansion of TIL

We tested the expandability of AML TIL in 10 bone marrow samples and compared to that of circulating lymphocytes from 10 peripheral blood samples collected from the patient cohort in 14-day co-culture system using irradiated feeder cells of unrelated PBMC supplemented with anti-CD3, IL-2, IL-7, and IL-15. Bone marrow group yielded a mean \pm SD of $8.7 \times 10^9 \pm 1.0 \times 10^9$ cells being similar to $7.1 \times 10^9 \pm 0.8 \times 10^9$ derived from the peripheral blood cell group ($p=0.093$). The folds of increase in cell numbers of 875 ± 102 were noted among bone marrow samples being comparable to 713 ± 80 of the circulating lymphocytes in the peripheral blood samples ($p=0.098$) (Figure 2).

Phenotypic characterization of expanded TIL

The immunophenotypings of ex vivo expanded TIL and circulating lymphocytes in peripheral blood are shown in Table 2. There were no significant differences of the relative frequencies of the cell subsets, namely, CD3+CD8+ T cells, CD3+CD4+ T cells, and CD3+CD56+ T cells derived from bone marrow and peripheral blood of AML patients, suggesting the culture system could expand AML TIL equally well. However, expressions of CD137 (4-1BB) and CD279 (PD-1) on TIL were significantly up-regulated compared to those expressed on circulating lymphocytes.

Functionality of expanded TIL

Figure 3A shows the quantitation of secretory IFN- γ in the culture supernatants of the 10 bone marrow samples evident of detectable TIL. A mean of 2228.5 pg/mL (range: 670–3900 pg/mL) was found. All readouts were far beyond the cut-off threshold of 200 pg/mL being widely acknowledged in the field.²⁰ CD137 is a costimulatory marker that is induced upon the specific recognition of T cells by their target cell, and reactivity of the TILs was also evaluated by the upregulation of CD137. The baseline CD137 expression in the cultures of AML TILs before and following coincubation with autologous AML cells was determined in the 10 bone marrow samples, supporting the understanding that CD137 serves as a marker of the reactivity of AML (Figure 3B).

Table 1 Patient demography

Patient	Sex/ Age	FAB Subtype	Karyotype	Previous treatment (Yes/No)	Marrow blast (%)	Gene mutations
1	F/31	AML M2	Multiple abnormalities	No	56	KIT, SMC3, RAD21
2	M/19	AML M4	Multiple abnormalities	No	22	TET2, KIT, RAS, FLT3-TKD
3	M/44	AML M2	45,X,-Y,t(8;21)(q22;q22)[15]/46,XY[5]	No	22	TET2, KMT2A
4	M/28	AML M1	Multiple abnormalities	Yes	87	No mutation detected
5	F/48	AML M5	46,XX,t(3;5)(q25;q34)[9]/46,XX[1]	Yes	20	No mutation detected
6	F/34	AML M2	46,XX	No	21	IDH1
7	F/36	AML M2	45,X,-X,t(8;21)(q22;q22)[10]	Yes	60	GATA2, IDH1
8	F/56	AML M2	46,XX	No	60	No mutation detected
9	M/31	AML M5	46,XY	No	26	No mutation detected
10	M/28	AML M2	47,XY,+21[4]/46,XY[16]	No	70	CEBPA
11	M/54	AML M4	Multiple abnormalities	Yes	97	NRAS, KRAS, RUX1, TET2
12	F/27	AML M4	46,XX	Yes	32	WT1, NRAS, CEBPA, PTPN11
13	F/42	AML M2	46,XX	Yes	63	CEBPA, TET2
14	F/18	AML M2	46,XX	No	70	FLT3-ITD, RUNX1
15	M/24	AML M2	46,XY	Yes	21	CEBPA
16	F/33	AML M2	46,XX	No	42	GATA2, TET2
17	M/55	AML M2	45,X,-Y,t(8;21)(q22;q22)[5]	Yes	80	DNMT3A, FLT3-ITD
18	M/38	AML M2	46,XY,t(6;11)(q27;q23)[10]	No	21	GATA2, TET2
19	M/40	AML M2	46,XY	No	82	TET2, FLT3
20	F/53	AML M2	Multiple abnormalities	No	76	FLT3, NPML1, IDH1, TET2

Abbreviations: AML, acute myeloid leukemia.

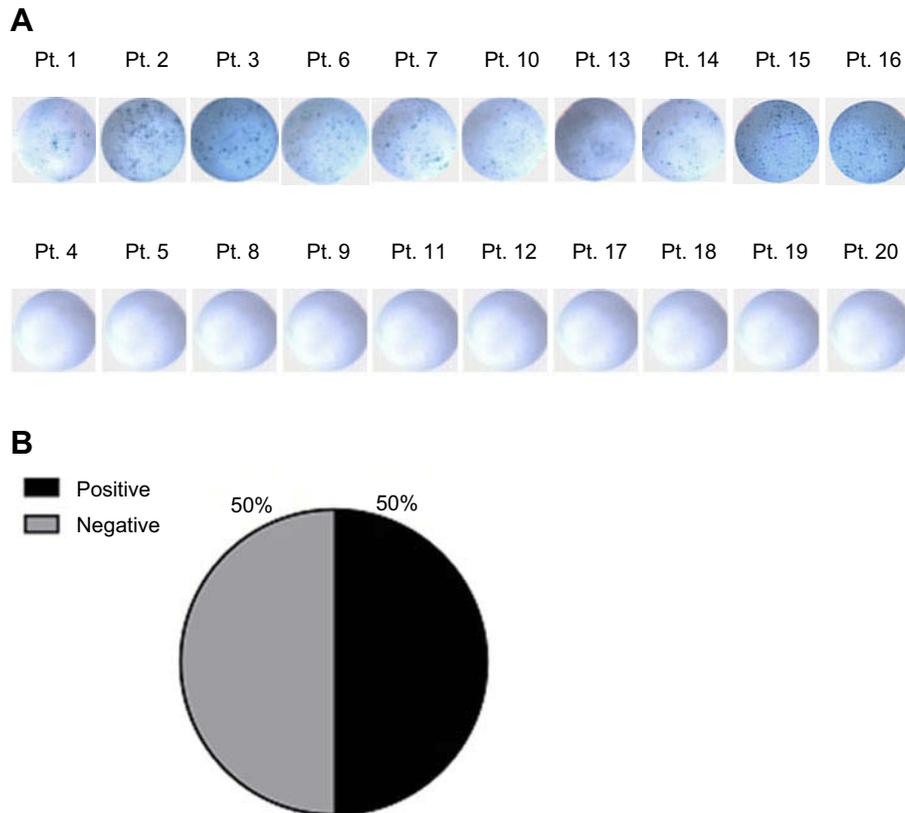


Figure 1 IFN- γ Elispot. **(A)** AML TIL for each patient were co-cultured overnight with autologous AML cells. Representative images of duplicate experiments are shown. **(B)** Distribution chart of AML patients with and without detectable TIL.

Abbreviations: AML, acute myeloid leukemia; Pt, patient; TIL, tumor-infiltrating lymphocytes.

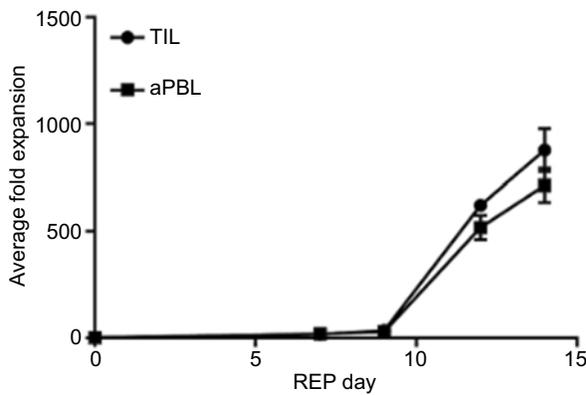


Figure 2 Growth kinetics of AML TIL (n=10) and AML aPBL (n=10) in triplicate. **Abbreviations:** aPBL, activated peripheral blood lymphocytes; AML, acute myeloid leukemia; TIL, tumor-infiltrating lymphocytes.

Table 2 Immunophenotypes of ex vivo expanded TIL and aPBL of AML patients

Phenotype	Frequency in percentage (mean \pm SD)		p-value
	TIL (n=10)	aPBL (n=10)	
CD8 ⁺ CD3 ⁺	53.5 \pm 31.67	57.67 \pm 14.64	0.843
CD4 ⁺ CD3 ⁺	38.75 \pm 31.94	28.33 \pm 10.07	0.613
CD56 ⁺ CD3 ⁺	14.75 \pm 14.31	20.67 \pm 7.57	0.550
CD137 ⁺ CD3 ⁺	4.3 \pm 2.50	1.125 \pm 0.46	0.020
CD279 ⁺ CD3 ⁺	2.07 \pm 0.43	0.37 \pm 0.18	0.003

Abbreviations: aPBL, activated peripheral blood lymphocytes; AML, acute myeloid leukemia; TIL, tumor-infiltrating lymphocytes.

In the comparative study of expanded TIL and aPBL of three AML patients, the secretory IFN- γ levels derived from co-cultures of TIL and autologous AML cells were much higher than those from co-cultures of aPBL and autologous AML cells suggesting greater anti-leukemia reactivity (Figure 4A). The anti-leukemia propensity of TIL was further attested in cytotoxicity

assays using K562 cells which numbers were normalized to the same numbers of TIL and aPBL in the assays (Figure 4B, Patient 1: TIL-mediated cytotoxicity vs aPBL-mediated; 36.8% \pm 2.2% vs 18.7% \pm 2.0%; $p=0.001$, Patient 2: 41.5% \pm 8.5% vs 21.0% \pm 4.5%; $p=0.021$, Patient 3: 38.1% \pm 5.0% vs 21.6% \pm 3.1%; $p=0.008$). (Ara-C)-mediated cytotoxicity was 14.93% \pm 0.64% in K562 cells. The anti-leukemia propensity of TIL was also attested in cytotoxicity assays using NB4 cells which numbers were normalized to the same numbers of TIL and aPBL in the assays (Figure 4C, Patient 1: TIL-mediated cytotoxicity vs aPBL-mediated; 28.9% \pm 1.5% vs 10.0% \pm 1.1%; $p=0.001$, Patient 2: 31.6% \pm 2.5% vs 17.0% \pm 2.5%; $p=0.002$, Patient 3: 30.0% \pm 3.5% vs 14.0% \pm 2.1%; $p=0.025$). (Ara-C)-mediated cytotoxicity was 7.34% \pm 0.26% in NB4 cells.

Magnetic bead separation of CD137-expressing cells

Immunomagnetic selection of CD137-expressing cells was performed on expanded TIL of Patient 3. The expression of CD137 on TIL before expansion culture was low being 2.66%, but CD137⁺ cells increased significantly to 6.24% after co-culture with autologous AML cells and was further increased to 42.13% in the CD137⁺ fraction after the magnetic separation (Figure 5A–C). The immunomagnetic selection enables the enrichment of IFN- γ -producing TIL as evidence of an increase of Elispot counts (Figure S1, pre-selection vs post-selection; 66 \pm 3 vs 661 \pm 20).

Discussion

The treatment of AML is challenging. Although a majority of AML patients achieve remission after chemotherapy, only a small portion of patients especially those >60 years

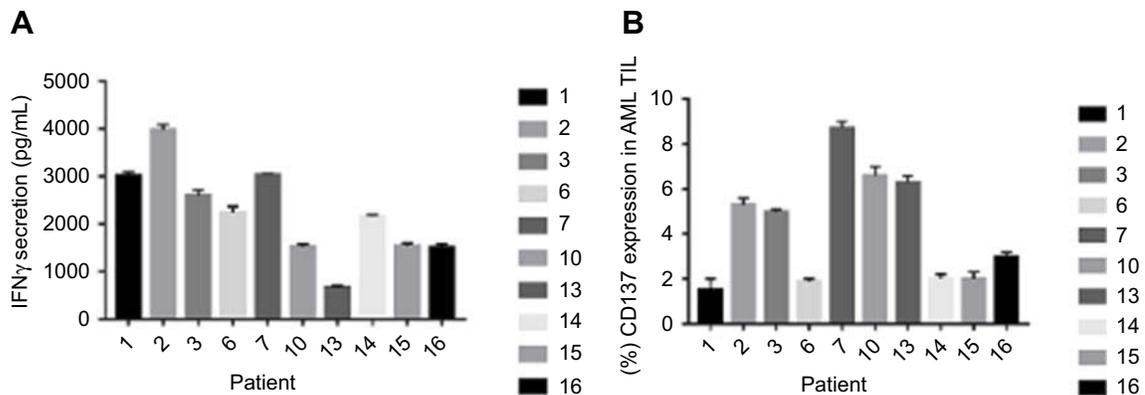


Figure 3 Overnight co-cultures of AML TIL and autologous AML cells in triplicate. **(A)** IFN- γ secretion. **(B)** CD137 expression. **Abbreviations:** AML, acute myeloid leukemia; TIL, tumor-infiltrating lymphocytes.

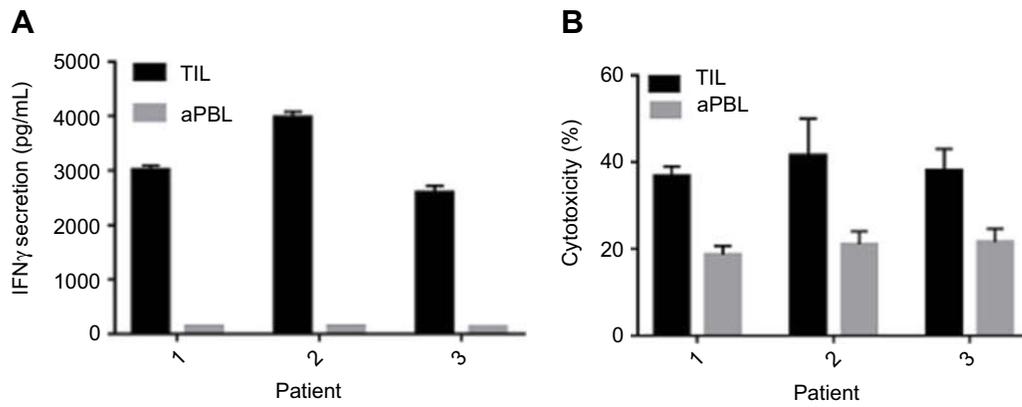


Figure 4 Comparison of TIL and aPBL on IFN- γ secretion and anti-leukemia reactivity. **(A)** AML TIL and AML aPBL of patients 1, 2, and 3 were co-cultured with autologous AML cells in triplicate. **(B)** AML TIL and AML aPBL were co-cultured with K562 cell line in triplicate. **Abbreviations:** aPBL, activated peripheral blood lymphocytes; AML, acute myeloid leukemia; TIL, tumor-infiltrating lymphocytes.

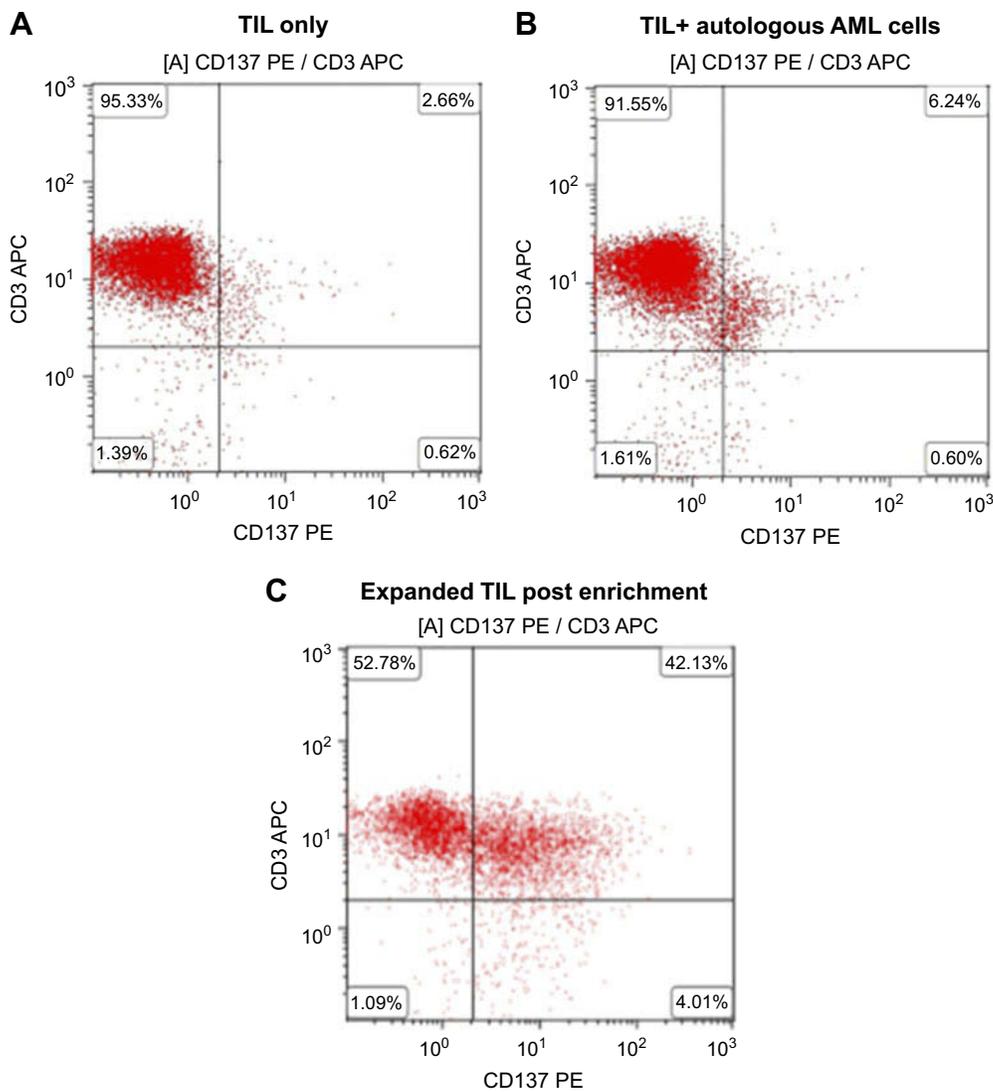


Figure 5 CD137 separation with magnetic beads enriches TIL from AML Patient 3. **(A)** TIL. **(B)** Overnight co-culture of TIL and autologous AML cells. **(C)** Expanded TIL post-CD137-selection. **Abbreviations:** AML, acute myeloid leukemia; TIL, tumor-infiltrating lymphocytes.

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of age might experience durable responses. Besides, the remission is relatively short. Approximately 20% of AML patients can achieve disease remission at two years. The five-year survival rate of AML patients of <60 years of age is approximately 35%.²⁵ The high rate of relapse after chemotherapy is thought to be related to the intrinsically resistant clonal populations to cytotoxic injury. Allo-HSCT is potentially curative for a subset of patients because of the capacity of allo-reactive lymphocytes to eradicate malignant clonal populations.²⁶ However, the treatment modality is often limited by treatment-related mortality and morbidity.

There is a need for novel therapies for AML including immune-based therapies. To our knowledge, there is little promising immunotherapy available to the treatment of AML. Striking responses have been observed in patients with refractory acute lymphocytic leukemia undergoing immunotherapy with chimeric antigen receptor-T (CAR-T) cell therapy, in which persistent leukemia-specific T cells has led to durable response.²⁷ Unfortunately, identifying a target that is widely expressed on AML cells is challenging. As a result, the application of CAR-T cell therapy in AML patients has proven to be limited. Cellular immunotherapy may be an alternative.

“Neo-epitopes” derived from the tumor are ideal targets. They are unique in tumor cells but not normal tissue. T cells are able to recognize differences between mutated epitopes and non-mutated epitopes because of peptides on the surface of tumor cells presented by MHC molecules. They are “non-self”. They are not subject to central tolerance and may potentially be immunogenic.²⁸ They can elicit T cell response and are often found infiltrating solid tumors and likely play an important role in ACT.

Reports in the literature suggest that ACT using autologous TIL was promising in patients with solid tumors. T cells that are reactive to tumor-specific antigens might mediate tumor regression.²⁹ Twenty percent of metastatic melanoma patients were reported to experience durable complete regression after treatment with autologous TIL in the light of reactivity with autologous tumor cells.³⁰ In the postoperative treatment of non-small cell lung cancer, the three-year survival rate was significantly higher for patients who underwent TIL ACT compared to that of untreated patients.³¹ In patients with pancreatic adenocarcinomas, infiltration of CD4⁺ T and CD8⁺ cells was noted to associate with an improved prognosis and a markedly better five-year survival rate.³² A recent study demonstrated that the infiltration of CD3⁺ T cells correlated with an improved clinical outcome in patients with epithelial ovarian cancer.³³

T cell immune response in patients with solid tumors suggests the potential application of TIL.

Whether AML harbor TIL has been debated for decades, but no direct evidence has so far been reported. In this study, we demonstrate, for the first time, that the presence of AML TIL and they are able to specifically recognize autologous AML cells. The major observation on a small cohort of AML patients is: 1) TIL could be identified in bone marrow of 50% of AML patient cohort as revealed by immunophenotyping and IFN- γ Elispot assay, 2) AML TIL exerted anti-AML reactivity and anti-leukemia cytotoxicity, 3) AML TIL exhibited a higher anti-AML reactivity and anti-leukemia cytotoxicity compared with those of aPBL, 4) AML TIL were expandable, and 5) enrichment of expandable AML-reactive TIL is feasible. More works are needed to explore the potential application of AML TIL in ACT.

The recognition of neo-epitopes is important in the understanding of tumor immunity and host immune defense such as the generation of TIL. For a tumor-specific antigen to be recognized by the human immune system, several conditions must be met: 1) production of mutant peptides by the antigen processing machinery, 2) high affinity of mutant peptides to human leukocyte antigen (HLA), and 3) recognition of mutant peptide-HLA complex by host T cells.³⁴ Studies on solid tumors demonstrated that 0.5–4% of mutations spontaneously trigger T-cell response.¹⁸ AML TIL could recognize autologous AML cells as neo-epitopes on AML cells were presented by HLA molecules. Among 10 AML patients whom TIL were not detected, disease-related gene mutations were demonstrated in six patients. Mutant proteins may be very low in quantity or poorly expressed on AML cells. Mutant epitopes may not be properly presented to the HLA molecules. The affinity of the epitopes to the host HLA molecules is very low leading to poor and unstable formation of mutant peptide-HLA complex binding to T cell receptors.

A recent study demonstrated that TIL reactivity represents neo-epitope-driven T-cell responses in solid tumor.³⁵ Three patients with metastatic cervical cancer experienced durable complete regression after treatment with autologous TIL. The reactivity of autologous TIL in terms of IFN- γ release and CD137 upregulation was noted.³⁶ In a study of patient with metastatic triple negative breast cancer, the T cell response against a special mutation was characterized.³⁷ Increases of IFN- γ secretion and CD137 expression on TIL were reported. In this report, we identified anti-leukemia

reactivity in AML TIL which could secrete IFN- γ . The expression of CD137 on AML TIL was up-regulated in co-cultures with autologous AML cells. The AML TIL demonstrated a profound anti-leukemia reactivity compared to that of AML aPBL in terms of IFN- γ secretion and cell-mediated cytotoxicity. Our observation attests the findings in solid tumors.

Upon antigen recognition, T-cells evolve significantly with functional and phenotypic changes. The upregulation of co-inhibitory molecule CD279 (PD-1) and the co-stimulatory molecule CD137 (4-1BB) provides the opportunity to use these biomarkers to identify tumor-specific T cells in solid tumors.³⁸ CD137 (4-1BB), a TNFR-family member, is an important regulator of immune responses with specific antigen expression.³⁹ CD279 (PD-1) was used to recognize neo-antigen-reactive T cells, but it was noted that self-antigen-reactive T cells also express CD279 (PD-1).⁴⁰ A previous study reported that the expression of CD137 (4-1BB), but not CD279 (PD-1), on lymphocytes defines a population of tumor-reactive cells in solid human cancers.³⁰ In this study, the AML TIL have a higher expression of CD137 (4-1BB) and CD279 (PD-1) than those of AML aPBL. The numbers of CD137 (4-1BB)-expressing cells were upregulated following co-cultures with autologous AML cells. Data of the study are in accordance with previous reports suggesting that CD137 (4-1BB) may be a potential biomarker for identification and enrichment of AML-reactive TIL. Tumor-reactive TIL numbers are low in solid tumors. The ex-vivo expansion of TIL may be amenable to yield a therapeutic dose of functional TIL for ACT. An enrichment of CD137+ cells further enhances the purity of TIL from IL-7- and IL-15-supplemented expansion cultures. More works are needed to confirm the preliminary findings in this study.

Conclusion

We first demonstrate TIL in 50% of AML patient cohort. AML TIL express CD137 (4-1BB) and secrete IFN- γ . They exhibit profound anti-AML reactivity and cytotoxicity. Besides, they are expandable in cultures, which might be translated to ACT for AML in the future.

Ethical approval

The study was conducted subject to the approval of the Institutional Review Board of Harbin Medical University and in accordance with the Declaration of Helsinki. Bone

marrow and peripheral blood samples were taken with written informed consent of patients and donors.

Abbreviation list

ACT, Adoptive cell therapy; allo-HSCT, Allogeneic hematopoietic stem cell transplantation; AML, Acute myeloid leukemia; aPBL, Activated peripheral blood lymphocytes; APL, Acute promyelocytic leukemia; BMMC, Bone marrow mononuclear cells; ELISA, Enzyme-linked immunosorbent assay; IL, Interleukin; IFN- γ , Interferon- γ ; LDH, Lactate dehydrogenase; PBMC, Peripheral blood mononuclear cells; TIL, Tumor-infiltrating lymphocytes.

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Disclosure

The authors report no conflicts of interest in this work.

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Supplementary materials

Methods

Mutations in the following 127 genes were used in the indicated number of patients: CEBPA,NPM1,FLT3-ITD, FLT3-TKD,TP53,KIT,DNMT3A,IDH1,IDH2,MLL,TET2, RUNX1,KRAS,NRAS,ASXL1,WT1,GATA1,SF3B1, SRSF2,U2AF1,ZRSR2,EZH2,CBL,JAK2,SETBP1,ETV6, MPL,CALR,PTPN11,CSF3R,GATA2,STAG2,JKA3, BRAF,TERC,IKZF1,ABL,SH2B3,NOTCH1,FBXW7, CDKN2A,TPMT,IL7R,JAK1,JAK3,RB1,MYD88, ABCB1,ABCC3,AKT2,AKT3,AMER1,APC,ATM,ATRX, BCL2,BCOR,BCORL1,CACNA1E,CARD11,CBLB, CBLC,CCND1,CD79B,CDA,CREBBP,CRLF2,CSF1R, CTLA4,CUX1,CYP2C19,CYP3A4,DIS3,DNAH9,E2A, EGFR,ERCC1,ERG,FAM46C,GNAS,GSTM1,GSTP1, HRAS,ID3,KDM6A,KMT2C,KMT3A,MAP2K4, MAP3K7,MDM2,MEF2B,MLH1,MTHFR,NF2, NOTCH2,NQO1,NT5C2,NTRK1,NTRK2,PDGFRA,

PIGA,PIK3CA,PTEN,RAD21,SMAD4,SMC1A,SMC3, STAT5A,STAT5B,SYK,TRAF3,XRCC1,XO,GATA3, EP300,NF1,ETNK1,ANKRD26,DDX41,DKC1,PRPF8, STAT3,TERT,SRP72,ELA2,HAX1,GFI1,BLM,ATG2B, GSKIP,ATM,BCOR,PIGA.

Table S1 IFN- γ Elispot counts

Patient	1	2	3	6	7	10	13	14	15	16
Spot counts	150	250	136	180	125	100	70	160	170	180

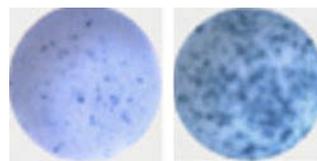


Figure S1 IFN γ release of CD137 separated TIL compared with unseparated TIL. **Abbreviation:** TIL, tumor-infiltrating lymphocytes.

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