

Polydopamine-Based Biomaterials in Orthopedic Therapeutics: Properties, Applications, and Future Perspectives

Min Zhang^{1-3,*}, Man Mi^{1,2,4,*}, Zilong Hu^{3,4,*}, Lixian Li^{3,4}, Zhiping Chen¹⁻³, Xiang Gao⁵, Di Liu^{3,4}, Bilian Xu³, Yanzhi Liu¹⁻³

¹Zhanjiang Key Laboratory of Orthopaedic Technology and Trauma Treatment, Zhanjiang Central Hospital, Guangdong Medical University, Zhanjiang, 524037, People's Republic of China; ²Key Laboratory of Traditional Chinese Medicine for the Prevention and Treatment of Infectious Diseases, Guangdong Provincial Administration of Traditional Chinese Medicine (Central People's Hospital of Zhanjiang), Zhanjiang, 524037, People's Republic of China; ³Marine Medical Research Institute of Zhanjiang, School of Ocean and Tropical Medicine, Guangdong Medical University, Zhanjiang, 524023, People's Republic of China; ⁴Guangdong Provincial Key Laboratory for Research and Development of Natural Drug, School of Pharmacy, Guangdong Medical University, Zhanjiang, 524023, People's Republic of China; ⁵Stem Cell Research and Cellular Therapy Center, The Affiliated Hospital of Guangdong Medical University, Zhanjiang, Guangdong, 524001, People's Republic of China

*These authors contributed equally to this work

Correspondence: Bilian Xu, Marine Medical Research Institute of Zhanjiang, School of Marine and Tropical Medicine, Guangdong Medical University, Zhanjiang, 524023, People's Republic of China, Tel +86-0759-2388588, Email l24879846@qq.com; Yanzhi Liu, Zhanjiang Key Laboratory of Orthopaedic Technology and Trauma Treatment, Zhanjiang Central Hospital, Guangdong Medical University, Zhanjiang, 524037, People's Republic of China, Tel +86-0759-3157875, Email liuyanzhi02@163.com

Abstract: Polydopamine is a versatile and modifiable polymer, known for its excellent biocompatibility and adhesiveness. It can also be engineered into a variety of nanoparticles and biomaterials for drug delivery, functional modification, making it an excellent choice to enhance the prevention and treatment of orthopedic diseases. Currently, the application of polydopamine biomaterials in orthopedic disease prevention and treatment is in its early stages, despite some initial achievements. This article aims to review these applications to encourage further development of polydopamine for orthopedic therapeutic needs. We detail the properties of polydopamine and its biomaterial types, highlighting its superior performance in functional modification on nanoparticles and materials. Additionally, we also explore the challenges and future prospects in developing optimal polydopamine biomaterials for clinical use in orthopedic disease prevention and treatment.

Keywords: polydopamine, biomaterials, nanoparticles, bone, orthopaedics

Introduction

Polydopamine (PDA), a polymer synthesized from dopamine monomers, is a key component found in human melanin. It has demonstrated that PDA exhibits exceptional adhesiveness, numerous covalently modifiable functional groups, significant near-infrared (NIR) absorption capacity,¹ and favorable biodegradability. Due to these advantages, PDA is commonly used for surface modification of materials, attracting interest from the scientific community. PDA has been thoroughly investigated across a spectrum of biomedical fields, including cell culture, drug delivery, biomaterial coatings, biomedical imaging, tissue engineering, and biosensing applications.^{2,3} PDA has emerged as a versatile biomaterial with immense potential in orthopedic therapeutics. Studies have demonstrated that PDA nanoparticles exhibit effective performance in the delivery and sustained release of anti-osteoarthritis (OA) drugs. Additionally, applying PDA nanoparticle coatings to biomaterial surfaces enhances their adhesiveness and antibacterial properties,⁴ while PDA's antioxidative properties provide protection to biomaterials within tissues. Research on bone reconstruction and osteogenic differentiation has shown that PDA nanoparticles are capable of downregulating pro-inflammatory factors in chondrocytes, creating a favorable microenvironment, regulating cell behavior, and promoting the vitality, adhesion,

migration, and osteogenic differentiation of mesenchymal stem cells (MSCs).⁵ Polydopamine-based biomaterials, despite being at a rapid development stage, offer promising avenues for the prevention and treatment of orthopedic diseases. This review aims to comprehensively explore the current state of polydopamine-based orthopedic therapeutics, highlighting its properties, applications, achievements, challenges, and future directions.

Synthesis and Properties of Polydopamine: Methods and Applications

PDA forms from dopamine (DA) under alkaline conditions through oxidation, cyclization, and rearrangement to form indolequinone, followed by further reactions with DA molecules,⁶ as illustrated in Figure 1. The polymerization of PDA is typically achieved through three methods, including solution oxidation, enzyme oxidation, or electro-polymerization.⁷ Among these methods, solution oxidation remains the most widely used due to its simplicity. Although this method functions under mild conditions and does not necessitate complex equipment, it exhibits sensitivity to various factors, including DA concentration,⁸ oxidant content, temperature, pH, and stirring speed. PDA is usually polymerized at room temperature (20–25°C). Although higher temperatures, up to 60°C, can accelerate dopamine polymerization, they may also lead to non-specific aggregation.⁹ Oxygen plays a vital role and is sufficient for PDA polymerization. The addition of ozone accelerates the dopamine polymerization process within the pH range of 4.0 to 10.0 and extends the temperature range, allowing polymerization even at low temperatures, such as 5°C.¹⁰ The optimal pH range for dopamine (DA) polymerization is between 8.5 and 9.5. Previous study revealed that DA molecules first undergo a pH-induced auto-oxidation reaction to form dopamine quinone (DAQ).¹¹ DAQ molecules then crosslink through the formation of biphenyl bonds, leading to the formation of polycatecholamine intermediates, which further grow into the PDA film. During the polymerization of PDA, π - π interactions and hydrogen bonding play crucial roles. Dopamine (DA) oxidizes to quinone, which can cyclize into 5,6-dihydroxyindole (DHI). These three monomers can coexist in solution, each contributing to stacking and polymerization. Between monomer layers, π - π stacking and hydrogen bonding occur, while covalent bonds form between monomers at multiple sites.¹¹ Monomers bond through benzene rings to form lengthy polymer backbone chains, with the PDA backbone containing multiple types of constructing monomers. Indole groups may crosslink, lengthening the side chain or stretching the backbone. Studies show that two cross-plane dopamine monomers physically interact during polymerization, clasping one DHI and generating a physical trimer.^{12,13} DA forms hydrogen bonds with molecules and other functional groups, contributing to its high adhesion strength to both organic and inorganic surfaces.^{14–16} In conclusion, π - π interactions and hydrogen bonding are fundamental to the properties and applications of PDA materials, significantly influencing their structural integrity and adhesion capabilities.

On the other hand, the laccase-catalyzed method is suitable for alkaline-sensitive materials, as it allows PDA polymerization to be completed at a pH of 5.5.¹⁷ Usually, the polymerization reaction can take several hours or even a day.¹⁸ During the polymerization process, dopamine molecules gradually polymerize and deposit on surfaces or form nanoparticles. Ultrasound can increase the deposition rate of PDA coatings as polymer films.¹⁹ Additionally, adding a Fe catalyst can accelerate PDA polymerization at pH 7.0 and even facilitate it at lower pH levels down to 4.0.²⁰ Typically, the rate of PDA polymerization of this method is notably slow, and the resulting PDA often settles at the bottom of the container, posing challenges for researchers. Enzyme oxidation, while more complex than solution oxidation, is considered relatively environmentally friendly and typically involves enzymes such as tyrosinase, polyphenol oxidase (also known as laccase), and urase.²¹ Electro-polymerization can produce PDA more rapidly than solution/enzyme

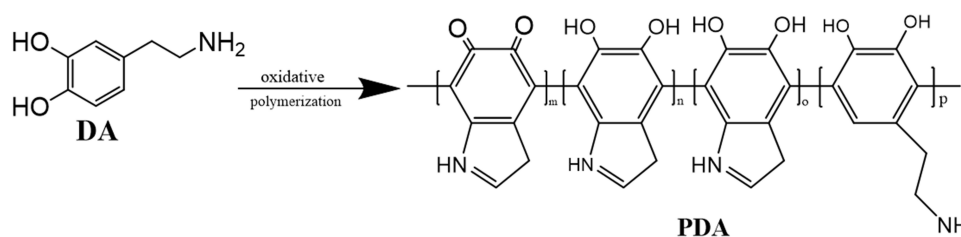


Figure 1 The Self-Polymerization Process of Polydopamine.

oxidation and avoids employing environmentally unfriendly chemical oxidants. This method can be readily applied to various metal substrates, rendering PDA suitable for surface modification of metal implants.²² Structurally, PDA resembles the adhesive proteins secreted by invertebrate mussels (3, 4-dihydroxyphenylalanine). It is broadly recognized that multiple functional groups in PDA, comprising catechol groups, amino groups, carboxyl groups, indole units, and quinone functionalities, play a key role in its multifunctionality, resembling the adhesive properties of mussels. The abundance of primary and secondary amines and ortho-quinones in PDA's structure is instrumental in its strong adhesion, enabling it to adhere to nearly any material surface. The thickness and stability of the PDA adhesive film depend on the concentration of monomeric dopamine during self-polymerization. Prior research indicated that the elongation of the alkyl chain connecting the ortho-quinone and amine groups in PDA does not impact its adhesion strength.²³ Primary amines of PDA can be readily covalently modified, its excellent modifiability allows for tailored designs to suit specific therapeutic needs. Consequently, PDA represents an ideal adhesive material capable of modifying nearly all material surfaces and offers biocompatibility, presenting significant safety advantages.

Polydopamine Biomaterials: Properties and Types

PDA is derived from the polymerization of dopamine (DA), a critical neurotransmitter exist in the human body. DA plays a significant role in motor control, emotion regulation, endocrine regulation, cognitive function, and reward pathways. Consequently, polydopamine (PDA) degrades into DA, which contributes to its excellent biocompatibility and low cytotoxicity. Furthermore, PDA does not provoke significant immune responses, making it ideal for implant fabrication and drug delivery systems. Inspired by the adhesive proteins found in mussels, PDA exhibits strong adhesive properties. It contains a high density of primary amines, secondary amines, and catechol groups, which allow it to adhere to nearly any material surface. This versatility makes PDA suitable for coating a wide variety of substrates. PDA contains numerous functional groups that facilitate chemical modifications. This allows for the easy conjugation of various molecules, including drugs, proteins, and other biomolecules, enabling PDA to serve as a versatile platform for targeted drug delivery and other therapeutic applications. PDA's unique molecular structure and specific functional groups enable it to absorb light and convert it into heat, imparting special photothermal properties. The catechol groups in PDA contain aromatic rings with hydroxyl groups that can absorb light, particularly in the near-infrared (NIR) region. When exposed to NIR light, these groups undergo electronic transitions, generating heat. The π -conjugated system within the PDA polymer further enhances light absorption across a wide range of wavelengths, including the NIR region, allowing for efficient conversion of absorbed light into thermal energy. This makes PDA suitable for photothermal therapy applications.^{24–26} Polydopamine can be engineered into various biomaterial forms, including nanospheres, nanocapsules, nanofilms, and nanocomposites. Some representative electron microscope images of the PDA biomaterials are shown in [Figure 2](#), including PDA porous particles, PDA/Cu₂O coating, Ag/TiO₂/PDA-bamboo, and PU-PDA, among others. These forms offer distinct advantages in terms of drug loading and release kinetics, providing precise control over therapeutic interventions. PDA has been successfully applied in coating or modifying various nanostructures, enhancing their specific surface areas and increasing the surface available for cell and biomolecular interactions. Based on existing studies, PDA nanostructures can be classified into several categories, as illustrated in [Figure 3](#). These include: (1) organic PDA nanoparticles formed by self-oxidation and polymerization in alkaline solutions without templates; (2) hollow PDA nanoparticles, nanocapsules, nanotubes (NTs), and nanorods, which result from deposition on movable templates; and (3) PDA nanoparticles co-assembled with other biologically active components, such as PDA core/shell nanoparticles and PDA nanofilms that adhere to the surface of other nanostructures.^{25,27,28} During the preparation and application of PDA nanoparticles (PDA NPs), an increase in the pH of the alkali solution may reduce the diameter of the nanoparticles.²⁹

Application of Polydopamine Biomaterials in Orthopedic Disease Prevention and Treatment

Currently, extensive research has led to the development of various organic and inorganic nano-drug delivery systems, including silica nanoparticles, titanium nanotubes, gold nanoparticles, calcium phosphate nanoparticles, chitosan nanoparticles, liposomal nanoparticles, and polymer nanoparticles, along with micelles, and dendritic macromolecules, aimed

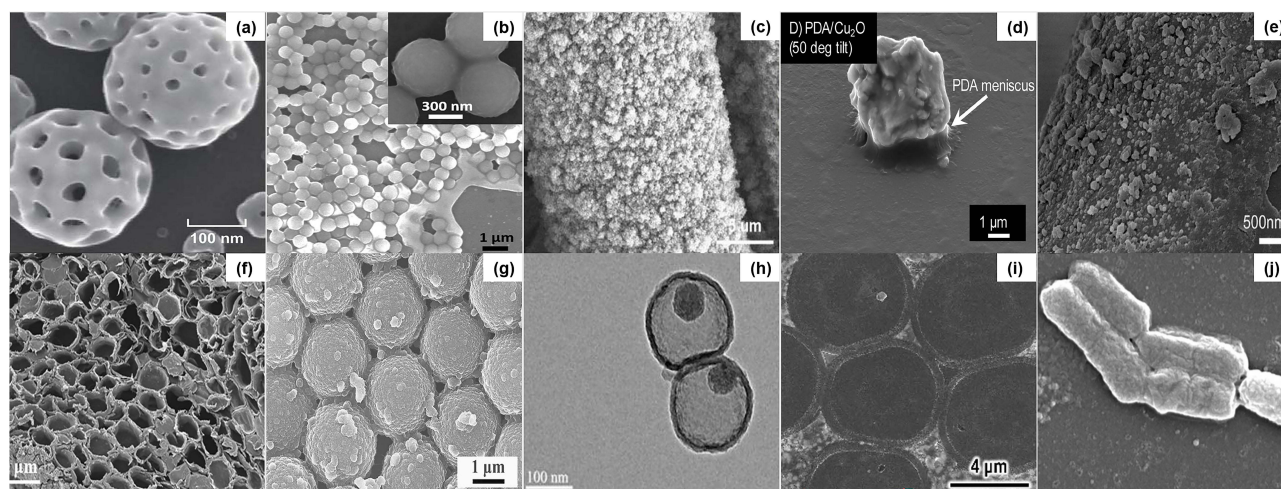


Figure 2 Overview of different SEM structures of polydopamine materials. (a) SEM images of the PDA porous particles. Adapted from Mei S, Kochovski Z, Roa R, et al. Enhanced catalytic activity of Gold@Polydopamine nanoreactors with multi-compartment structure under NIR irradiation. *Nanomicro Lett.* 2019;11(1):83. Creative Commons.³⁰ (b) SEM images of the PDA@SiO₂. Adapted from Khan MZH, Daizy M, Tarafder C, Liu X. Au-PDA@SiO₂ core-shell nanospheres decorated rGO modified electrode for electrochemical sensing of cefotaxime. *Sci Rep.* 2019;9(1):19041. Creative Commons.³¹ (c) SEM images of cotton/pDA/MnO₂. Adapted with permission from Zhang Y, Zhao Z, Li D, et al. In situ growth of MnO₂ on pDA-templated cotton fabric for degradation of formaldehyde. *Cellulose.* 2022;29(13):7353–7363.³² (d) SEM images of the PDA/Cu₂O coating. Adapted from Behzadinasab S, Williams MD, Hosseini M, et al. Transparent and sprayable surface coatings that kill drug-resistant bacteria within minutes and inactivate SARS-CoV-2 virus. *ACS Appl Mater Interfaces.* 2021;13(46):54706–54714. Copyright © 2021 American Chemical Society.³³ (e) SEM images of the non-woven polydopamine (NW@PDA) fabrics. Adapted from Zhang Z, Si T, Liu J, Zhou G. In-situ grown silver nanoparticles on nonwoven fabrics based on mussel-inspired polydopamine for highly sensitive SERS carbaryl pesticides detection. *Nanomaterials.* 2019;9(3). Creative Commons.³⁴ (f) SEM images of the Ag/TiO₂/PDA-bamboo surface. Adapted from Liu G, Lu Z, Zhu X, et al. Facile in-situ growth of Ag/TiO₂ nanoparticles on polydopamine modified bamboo with excellent mildew-proofing. *Sci Rep.* 2019;9(1):16496. Creative Commons.³⁵ (g) SPS-MS with PDA-PPy coatings. Adapted from Xie C, Li P, Han L, et al. Electroresponsive and cell-affinitive polydopamine/polypyrrole composite microcapsules with a dual-function of on-demand drug delivery and cell stimulation for electrical therapy. *NPG Asia Materials.* 2017;9(3):e358–e358. Creative Commons.³⁶ (h) TEM images of MSN@PDA. Adapted from Tran HQ, Bhavne M, Xu G, Sun C, Yu A. Synthesis of polydopamine hollow capsules via a polydopamine mediated silica water dissolution process and its application for enzyme encapsulation. *Front Chem.* 2019;7:468. Creative Commons.³⁷ (i) SEM images of C-PDA coated carbon fiber reinforced high-temperature composite. Adapted from Liu Y, Su C, Zu Y, Chen X, Sha J, Dai J. Ultrafast deposition of polydopamine for high-performance fiber-reinforced high-temperature ceramic composites. *Sci Rep.* 2022;12(1):20489. Creative Commons.³⁸ (j) SEM images of the PU-PDA. Adapted from Wang P, Zhang Y-L, K-L F, et al. Zinc-coordinated polydopamine surface with a nanostructure and superhydrophilicity for antibiofouling and antibacterial applications. *Mater Adv.* 2022;3(13):5476–5487. Creative Commons.³⁹

at treating bone diseases. However, these systems encounter limitations such as low loading capacity, limited physiological compatibility, degradability, and the propensity for rapid drug release by the systems. In recent years, numerous investigations have incorporated PDA as a composite component in biomaterials/drug delivery vehicles to augment drug loading capacities, bolster the trapping efficiency of cells in vivo, and address pathogen resistance. PDA-modified biomaterials have developed into various forms of applications, including nanotubes, nanospheres, and nanofilms, etc. (Table 1). Within the realm of orthopedic disease management, PDA-modified biomaterials are gradually becoming a major focus of research and application due to their multifunctionality and biocompatibility. The involvement of PDA can significantly enhance the anti-inflammatory, antibacterial, and antioxidant capabilities of orthopedic treatment strategies, and they notably improve the effectiveness of these strategies in promoting osteogenic differentiation and enhancing the activity of bone cells (Figure 4). The research and application of PDA span a diverse array of conditions, including bone injury repair,⁴⁰ bone regeneration,⁴¹ osteoporosis,⁴² and osteoarthritis.⁴³ Irrespective of the type of orthopedic condition, the onset age for bone diseases is increasingly younger,^{44–46} with a concurrently diversifying patient demographic. The development of innovative drug systems utilizing PDA is poised to significantly enhance personalized treatment approaches for orthopedic diseases due to their high loading capacity and controlled release mechanisms. In the application of PDA coatings on implants for bone injury repair, the self-healing property is crucial. PDA holds significant promise in the field of self-healing materials due to its exceptional adhesion and intrinsic chemical structure, which enables the automatic formation of new cross-links. PDA can be combined with polymer matrices to create self-healing polymer materials used in various applications, such as electronic equipment and automotive parts. For instance, PDA particles and hindered urea bonds can be utilized as functional nanofillers and dynamic motifs, respectively, to produce dynamic cross-linked polyurea/PDA(DCPU/PDA) nanocomposites via facile in situ

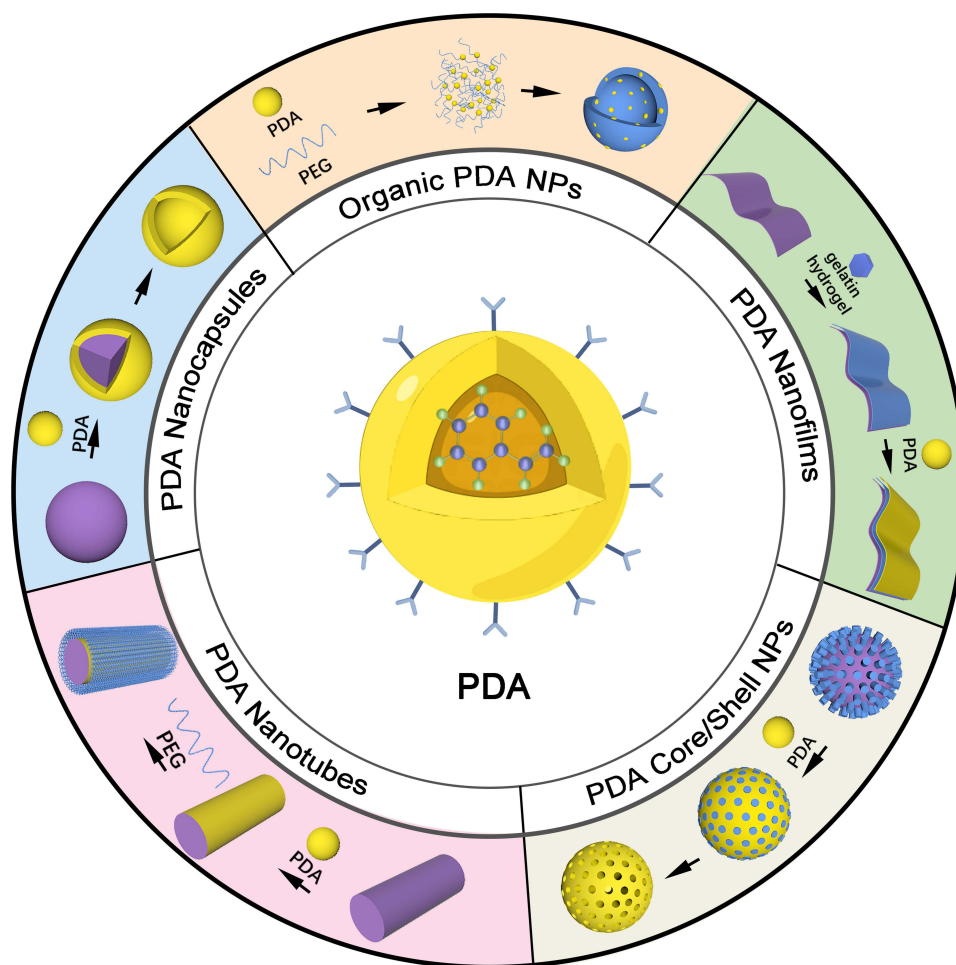


Figure 3 Classification of Polydopamine Nanoparticles. The diagram categorizes the various forms of polydopamine nanoparticles, including: organic PDA nanoparticles, PDA nanocapsules, PDA nanotubes, PDA core/shell nanoparticles and PDA nanofilms.

photoinitiated copolymerization.⁴⁷ These nanocomposites exhibit higher toughness and stretchability compared to pure DCPU. They self-heal rapidly and effectively in response to near-infrared light. Traditional thermo-responsive self-healing elastomers face challenges, including long repair times and limited self-healing sites. To address these issues, fast NIR light photo-responsive self-healing nanocomposites were fabricated by blending PDA particles into cross-linked polyurea containing sextuple H-bonds (SHBs) and hindered urea bonds (HUBs).⁴⁸ These composites show enhanced

Table I Advantages of Different Polydopamine-Based Applications for Orthopedic Diseases

Types	Advantages
PDA nanosphere shell PDA nanocapsules	Characteristics of High-Loading, Sustained-Release Drug Carriers The thickness of the material is controllable, enabling regulated drug release. Upon conjugation with branched chains, the system acquires enhanced bone-targeting capabilities, alongside notable antibacterial and antioxidant properties
PDA carbon/titanium nanotubes/nanorods PDA nanohydrogel	Regulates cell behavior to enhance osteogenic activity, thereby promoting the viability, adhesion, migration, and osteogenic differentiation of mesenchymal stem cells (MSCs) in vitro Enhances the mechanical properties, antibacterial activity, tissue adhesion, and osteogenic differentiation capabilities of hydrogels
PDA nanofilm coating	The microenvironment enhances MSCs' adhesion, proliferation, and osteogenesis, while concurrently downregulating pro-inflammatory cytokines in chondrocytes

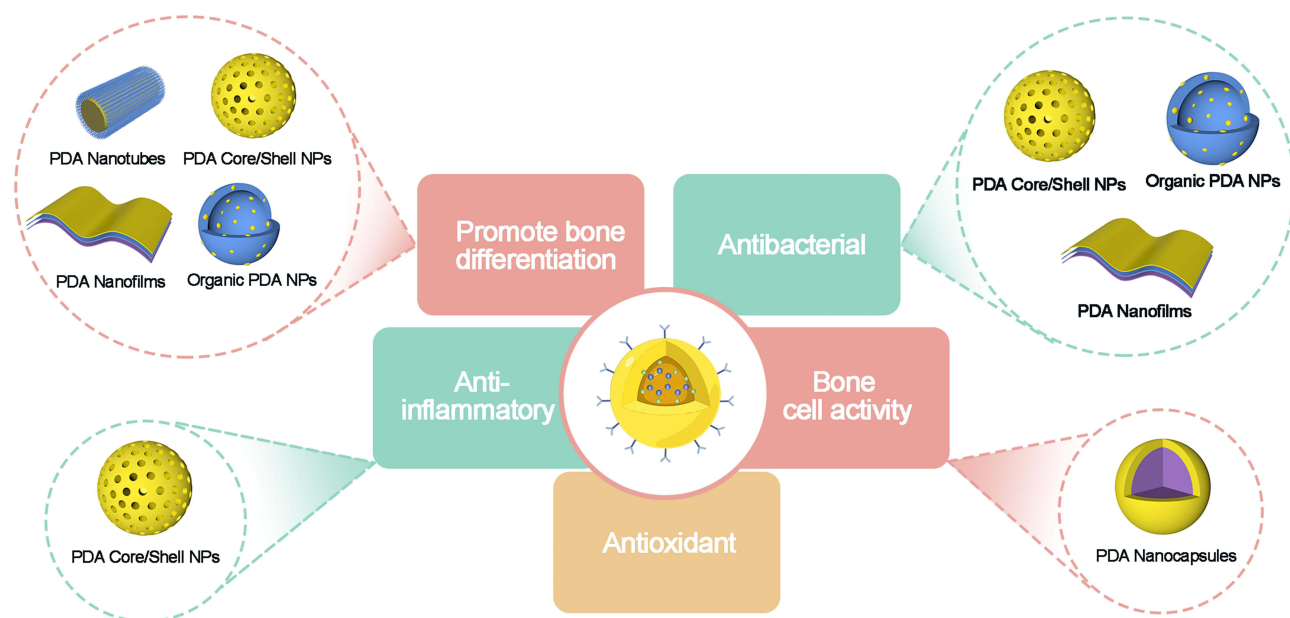


Figure 4 Application Spectrum of Polydopamine (PDA) in Orthopedic Disease Management: Material-Specific Utilizations and Therapeutic Directions.

mechanical properties and rapid photoresponsive self-healing capabilities due to the extensive H-bonds between PDA and polyurea fragments. In orthopedic applications, self-healing coating can alleviate corrosion, excessive degradation, and accidental scratching of metal-based implants. A previous study demonstrated the construction of a self-healing polymeric coating on Mg alloy, incorporating a stimuli-responsive drug delivery nanopatform via spin-spray layer-by-layer (SSLbL) assembly. This nanocontainer system, based on simvastatin (SIM)-encapsulated hollow mesoporous silica nanoparticles (S@HMSs) modified with PDA and polycaprolactone diacrylate (PCL-DA) bilayer,⁴⁹ exhibited dynamic reversible reactions, fast and stable self-healing capacity, and excellent antibacterial properties under NIR irradiation. Additionally, it promoted cytocompatibility, osteogenesis, and angiogenesis. Collectively, PDA-based materials exhibit significant potential as self-healing materials and can play crucial roles across various fields. With technological advancements and increasing application demands, the prospects for PDA-based self-healing materials will continue to expand and deepen.

Polydopamine Core/Shell Drug Delivery Systems for Bone Disease Treatment

PDA can encapsulate a diverse range of drugs, enabling therapeutic efficacy at lower doses and minimizing side effects. The PDA spherical shell exhibits a porous structure, extensive specific surface area, and compact volume. Its interconnected pore architecture not only mimics bone composition but also aligns with the microscopic size and shape requisites for bone repair. Recent study demonstrated the Chitosan/Polydopamine/Octacalcium Phosphate (CS/PDA/OCP) microcarrier, which combines porous chitosan for cell adhesion and proliferation with octacalcium phosphate to mimic natural bone components.²⁴ Enhanced with PDA for superior adhesion, this microcarrier facilitates the formation of microstructures conducive to bone repair, establishing a three-dimensional microenvironment aimed at precisely repairing irregular bone defects. The PDA shell surface offers numerous active sites for additional modification,⁵⁰ This affects the mechanical properties, stability, and even optical properties of the material. In PDA-NPs, π - π interactions contribute to the formation of hierarchical structures and affect the ability of the material to adhere to various surfaces.⁵¹ Thus this π interaction anchors the peptide or protein to the nanoparticle.⁵² Zhang et al encapsulates the NO precursor N, N'-disec-butyl-N,N'- dinitroso-1, 4- phenylenediamine (BNN6) within a β -CD hydrophobic cavity on PDA-coated BG NPs.⁵³ This setup is designed to enable NIR-triggered NO release, promoting antibacterial effects and osteogenic differentiation of MSCs. PDA-coated inorganic nanoparticles, including gold nanoparticles, magnetic iron oxide, and non-metallic nanoparticles, can be tailored for sustained and controlled release by incorporating active ingredients. PDA itself is not a therapeutic agent but must be conjugated with drugs, photosensitizers, metals, or metallic nanoparticles to

enhance their functionality. The combination of PDA with metallic nanoparticles, such as gold, platinum, silver, iron oxide, and zinc oxide, synergizes cancer treatment and imaging of cancer cells, which are valued for their magnetic properties, chemical stability, tunable morphology, and ease of surface functionalization.⁵⁴ The functional groups on PDA's surface (catechol, carboxylic, amine, and imine) enable the binding of specific molecules or the loading of transition metal ions. For example, iron oxide nanoparticles can serve as contrast agents in magnetic resonance imaging (MRI), while zinc oxide enhances cancer cell killing, and gold nanoparticles improve cancer cell death.^{54–57} Numerous *in vitro* studies have demonstrated that iron oxide nanoparticles promote osteoblast differentiation and inhibit osteoclast formation, while *in vivo* studies show that they accelerate bone defect repair and prevent bone loss.⁵⁵ For instance, previous studies reported that iron oxide nanoparticles coated with a PDA film acquire superior biocompatibility and multifunctionality.⁵⁶ A magnetic iron oxide/polydopamine coating significantly improves osteogenesis in 3D-printed porous titanium scaffolds under a static magnetic field.⁵⁷ Moreover, PDA combined with iron oxide nanoparticles can provide precise targeting to specific areas under the influence of a magnetic field. Researchers used Fe₃O₄@PDA to label MSCs, guiding them to pain-related response sites in spinal cord segments under magnetic control, thereby enhancing MSC homing and gathering for effective repair.⁵⁸ Most applications of PDA-coated iron-oxide NPs focus on cancer treatment, also for bone tumor diseases. For instance, to improve the efficacy of cisplatin in treating cisplatin-insensitive osteosarcoma, a study reported using iron-polydopamine coated multifunctional nanoparticles (SiO₂@PDA/Fe³⁺-FA).⁵⁹ These PDA-modified NPs have demonstrated high drug delivery efficiency, precise pH-responsive drug release, good biocompatibility, effective tumor targeting, and satisfactory photothermal efficiency, making them an effective tool for synergistic therapy in combination with NIR irradiation for drug-resistant tumors.

On the other hand, near-infrared (NIR) light-triggered shape memory materials show significant potential in biomedical applications compared to traditional heat-triggered shape memory polymers. For instance, a study reported the synthesis of polycaprolactone (PCL)–PDA polyurethanes using PCL, PDA nanoparticles (PDA NPs), hexamethylene diisocyanate (HDI), and 1,4-butanediol (BDO). These polyurethanes exhibited rapid shape recovery under NIR light due to the photothermal effect of PDA. The PCL–PDA polyurethanes demonstrated excellent *in vivo* NIR light-triggered shape memory performance under an 808 nm laser with low intensity, highlighting their potential for biomedical implant applications.⁶⁰ Another study reported the preparation of HNTs@PDA through *in-situ* free radical polymerization of acrylamide in a mixture of Laponite-RD, HNTs@PDA, and gelatin. HNTs@PDA, synthesized via oxidative polymerization of DA on the surfaces of HNTs, served as a superior photothermal agent for light-responsive hydrogels. The NIR-triggered shape recovery process of the HNTs@PDA hydrogel was notably rapid. These reinforced hydrogels, with superior mechanical properties, NIR-triggered shape memory, and self-healing ability, exhibit promising applications in biomedical materials.⁶¹ Smart self-healing coatings have also garnered significant interest due to their switchable and desirable functionalities in response to external environmental changes. An eco-friendly smart self-healing coating with NIR and pH dual-responsive superhydrophobic and anti-biofouling properties was fabricated by mixing biomimetic stimuli-responsive mesoporous PDA microspheres (polydimethylsiloxane-loaded mesoporous polydopamine microspheres, abbreviated as P-PDMS@MPDA MSPs) with waterborne resin and hydrophobic nanoparticles.⁶² This coating demonstrated self-healing of its superhydrophobicity and active anti-biofouling properties under NIR or pH stimuli due to the release of low-surface-energy PDMS from the P-PDMS@MPDA MSPs, showing excellent self-healing and biological properties. By adjusting the NIR light intensity, PDA microspheres can achieve precise temperature control of shape memory materials, enabling the material to undergo shape changes within the desired temperature range.^{63,64} Not only the toughness and strength of PDA particle-filled polyurethane composites (SMPU-PDAPs) have been significantly improved with the addition of PDAPs.⁶⁵ The tensile stress of damaged nanocomposites can also be recovered using the heat energy generated by near-infrared lasers.⁶⁶ This capability holds significant promise for biomedical applications necessitating precise manipulation, such as minimally invasive surgical instruments and implantable medical devices. For instance, strontium ions, which promote osteogenesis and angiogenesis, are stably incorporated into PDA microspheres due to PDA's coordination reaction,⁴⁰ optimizing the osteogenic potential of strontium ions while mitigating the adverse effects of high concentrations. Xiong Wei et al introduced a composite microsphere that can release both the antibacterial agent berberine (BBR) and the bone-strengthening drug naringin (NG), utilizing the simple synthesis of PDA coatings.⁴¹ This microsphere demonstrates excellent biocompatibility and degradability, making it

a promising option for treating bone defects at infected sites. Additionally, Leveraging the photothermal conversion capabilities of PDA particles and the concurrent release of adjuvants from mesoporous silica shells, researchers have developed PDA (core)-mesoporous silica (shell) nanocapsules aimed at tumor photothermal therapy.⁶⁷ PDA serves as a biodegradable photothermal agent with excellent biocompatibility, playing a key role in anti-tumor activity. PDA is renowned for its exceptional near-infrared (NIR) absorption properties, which are crucial for biomedical applications involving irradiation-sensitive materials. Here are the fundamental reasons for its effectiveness. PDA possesses a high concentration of aromatic rings, creating an extensive π - π conjugated system. This conjugation enables the material to absorb light energy across a broad wavelength range, including the NIR region.¹¹ The π - π system facilitates electron delocalization within the molecule, resulting in more efficient light absorption and energy transfer. PDA is abundant in radicals and semiquinone structures, which exhibit strong NIR absorption. These structures provide additional energy levels and electron transition pathways, significantly enhancing PDA's NIR absorption capabilities. The high degree of polymerization and tight molecular packing in PDA amplify intermolecular interactions. This packing not only supports efficient light absorption and conduction, but also enhances the material's stability and thermodynamic properties, thereby improving NIR absorption efficiency. The electronic structure of PDA can be optimized by adjusting synthesis conditions such as pH, temperature, and reaction time. For instance, doping PDA with metal ions or other functional groups can modify its electronic energy levels and optical properties, optimizing NIR absorption.⁶⁸ Conjugated π -electronic systems in PDA molecule-coated gold nanoparticles exemplify how such modifications can effectively enhance NIR light absorption.

PDA is highly multifunctional and tunable. Chemical modifications can introduce various functional groups, altering its optical properties. This tunability ensures that PDA performs exceptionally well in diverse NIR applications. As a biomimetic material, PDA offers excellent biocompatibility and environmental friendliness. Its safety and effectiveness in NIR absorption make PDA an ideal choice for biomedical applications involving irradiation-sensitive materials.

PDA Nanocapsules

PDA-based nanocapsules represent a prevalent form of drug delivery system. Previous report demonstrated PDA nanocapsules exhibit a high loading capacity, leveraging curcumin (CCM) and monoconcosaccharide interactions to create supramolecular structures. Upon modification with polydopamine and incorporation of drugs similar in polarity to CCM, the capsules maintain integrity while facilitating the formation of dual-drug nanoparticles for bidirectional drug delivery.⁶⁹ The capsule's drug release rate can be precisely adjusted through the manipulation of PDA thickness, enabling controlled drug delivery. This feature is particularly useful for administering anti-inflammatory agents like curcumin in the treatment of osteoarthritis.⁷⁰ Furthermore, the synthesis of PDA nanocapsules were through ammonia- induced PDA polymerization for coating silica mesoporous nanoparticles, as well as the formation of PDA nanocapsules by template removal following water dispersion,⁷¹ have been effectively employed as nanoparticles in drug delivery. Several studies have coated $Gd_2(CO_3)_3$ cores with PDA to create nanoparticles, to which a cartilage-targeting peptide was modified, and hesperetin was incorporated to establish a cartilage-specific functional drug delivery system. Experimental results have shown high affinity of this system for cartilage and its potential applications in magnetic resonance imaging (MRI).⁷² Research has shown that folic acid (FA) is an effective targeting agent for arthritis. By conjugating FA to PDA, a targeted nanodrug delivery system can be developed to target the arthritis site. Utilizing the pH-responsive mechanism, the NH_2 -PEG-FA ligand is synthesized to actively target OA, with size optimization ensuring full utilization of the EPR effect instigated by inflammation, thereby facilitating both effective passive accumulation and active targeting in OA treatment.⁷³ Yun Wang et al developed a mesopore-based microsphere encapsulating a drug clearance delay system (RCGD423@MPDA), employing MPDA loaded with the small molecule RCGD423 for modulating the inflammatory response. This system demonstrates superior drug loading capacity, exhibiting longer drug retention and more stable, prolonged drug release compared to traditional poly (lactate-co-glycolic acid).⁷⁴ In a rat model of osteoarthritis, administration of this system proved more effective in inhibiting cartilage damage and proteoglycan loss compared to treatments without the drug delivery system, suggesting its potential in mitigating the degenerative changes associated with arthritic cartilage. Mesoporous polydopamine nanoparticles are capable of extending the drug release cycle, thereby diminishing the need for frequent intra-articular injections. This presents significant benefits compared to the regular

administration of short-acting hyaluronic acid. Moreover, the ability to control drug release minimizes adverse drug reactions and mitigates the potential for drug resistance.

PDA Nanotubes

Recently, nanotubes formed by PDA coating (PDA@CNTs) have garnered considerable interest. Studies have demonstrated that PDA coating on carbon nanotubes significantly enhances cytocompatibility.⁷⁵ These PDA-coated carbon nanotubes have shown promise in various applications such as drug delivery, antimicrobial and antioxidant agents, and biological drugs.⁷⁶ Their porous and adjustable interfaces make PDA@CNTs a promising biocomposite with extensive biological, diagnostic, and therapeutic potential. For example, the integration of osteogenic growth peptide (OGP) with TiO₂ nanotubes via PDA has been found to promote cell proliferation and differentiation.⁷⁷ Moreover, PDA-coated TiO₂ nanotubes-MoS₂/PDA-LL-37 demonstrate potent antimicrobial activity and enhance bone regeneration when exposed to near-infrared irradiation.⁷⁸ PDA-modified titanium (Ti) substrates enhance the viability, adhesion, migration, and osteogenic differentiation of MSCs.⁷⁹ Additionally, biofunctionalized TiO₂/MoS₂/PDA nanotube coatings on Ti implants have been developed to stimulate osteogenic activity, notably through the simulation of MC3T3-E1 osteoblasts' differentiation and the upregulation of Runx2.⁸⁰ Recent clinical studies indicate that the majority of Ti implant failures are due to bacterial adhesion. Introducing a PDA bioactive coating on nanotubes can significantly mitigate implant infection issues associated with Ti implantation, due to the inherent antibacterial properties of PDA. Furthermore, PDA is capable of immobilizing growth factor proteins, for example vascular endothelial growth factor (VEGF),⁸¹ a property utilized in the PDA coating of TiO/MoS/PDA nanotubes to augment osteoinductivity.⁸² Strontium nanotubes/PDA arginine-glycine-aspartic acid coatings have also been employed on Ti₆Al₄V materials, significantly enhancing MSC adhesion and promoting MSC differentiation, thereby demonstrating outstanding bone-regeneration potential.⁸³ The recent proposition of composite artificial periosteum offers an innovative solution to the significant clinical challenge posed by periosteal defects.⁸⁴ Leveraging PDA's high tissue adhesion properties, the composite artificial periosteum features ends cross-linked with a PDA hydrogel layer of carbon nanotubes (CNTs), embodying a design that mimics the directional arrangement of periosteal collagen fibers. This construction ensures adequate mechanical strength and the desired directional nanotopological surface. Highlighting the potential of PDA nanotubes in orthopedic applications, their role in promoting bone regeneration demonstrates promising prospects.

Advancements in PDA Nanoparticles Crosslinked / Integrated with Other Materials

PDA nanoparticles can be cross-linked with acrylamide and additional polymers to fabricate hydrogels,⁸⁵ PDA nanoparticles crosslinked with acrylamide and other polymers leverage the high adhesion properties of polydopamine to enhance mechanical properties and biocompatibility. One notable example is the PAM/BA-Ag@PDA hydrogel, created by in situ polymerizing acrylamide (AM) with N,N'-bis(acryloyl)cystamine (BA), dynamically crosslinked with silver-modified polydopamine (PDA) nanoparticles.⁸⁶ This multi-functional hydrogel sensor exhibits significantly enhanced tensile and compressive strength, reduced hysteresis, improved conductivity, and excellent near-infrared (NIR) light-triggered self-healing abilities compared to traditional polyacrylamide (PAM) hydrogels. As a strain sensor, PAM/BA-Ag@PDA hydrogel demonstrates good sensitivity, rapid response time, and high stability. Previous studies have reported the fabrication of self-healing and adhesive, electrically conductive, and biocompatible PAM nanocomposite hydrogels. These are produced via in situ polymerization of acrylamide in the presence of polydopamine-modified carbon nanotubes (PDA@CNTs). Such hydrogels function as flexible strain or pressure sensors, effectively detecting human motions, identifying materials and their shapes, and transmitting health information,^{87,88} This strategy was also been extensively explored in the development of conductive hydrogels for biosensors. Additionally, PDA's photothermal effect significantly improves antimicrobial applications. One study demonstrated that PDA's photothermal properties can induce heat production to cause bacterial death under near-infrared light radiation. This research involved the uniform coating of PDA and polypyrrole (PPy) onto poly(l-lactide) (PLLA) nanofibers via in situ polymerization, resulting in a novel PPy/PDA/PLLA three-layer core-shell structure. The homogeneously coated PPy and PDA layers significantly increased hydrophilicity, conductivity, and near-infrared photothermal antibacterial properties, while also providing antioxidant capacity and reactive oxygen species (ROS) scavenging ability.⁸⁹ Given their high biosafety, numerous covalent modification sites, and superior adhesion, hydrogels derived from cross-linked PDAs are ideally suited for biomedical applications. Hydroxyapatite

Table 2 Effects of PDA Combined with Other Materials in Preventing and Treating Orthopedic Diseases

PDA+Other Materials	Effect
PDA+NO Precursors	Enhance antibacterial, promote the osteogenic differentiation of mesenchymal stem cells (MSCs). ⁵³
PDA+Mesoporous silica (shell)	Sustained-release drug carriers. ^{67,71}
PDA+PEG-FA	Target osteoarthritis. ⁷³
PDA+TiO ₂	Promote new bone formation. ^{35,77,78,80}
PDA+Ti ₆ Al ₄ V	Enhance the attachment and differentiation of MSCs, and promote bone reconstruction and regeneration. ^{83,104,105}
PDA+CNTs	Enhance mechanicality, fix the artificial periosteum, and promote bone regeneration. ^{75,88}
PDA+Hap	Increase biocompatibility and promote osteogenic differentiation, ^{26,90,99,100,106–108}
PDA+Ti	Enhance resistance to MRSA infection, ^{79,94}
PDA+HA	Enhance mechanical properties, cell affinity, tissue viscosity. ^{98–100}
PDA+PVDF+BaTiO ₃	Promote pre-osteoblast viability. ¹⁰⁹
PDA+mPEG-NH ₂	Down-regulation of cartilage pro-inflammatory factor. ¹¹⁰
PDA+PCL	Promote stem cell osteogenic differentiation. ^{60,111–113}

(HAP), a critical component of human bone, significantly affects cell behavior, including adhesion, proliferation, and differentiation, and is commonly employed in tissue regeneration. However, HAP fragments may trigger inflammation, impeding osteoblast growth.²⁶ PDA-modified HAP nanoparticles can surmount the inherent limitations of HAP-based materials in tissue repair. Specifically, PDA-mediated incorporation of bioactive peptides or proteins (such as bone morphogenetic protein-2) onto HAP nanoparticles has been shown to enhance their biocompatibility and promote osteogenic differentiation.⁹⁰

Numerous studies have utilized PDA NPs in combination with various active compounds to promote osteogenic differentiation,^{91–95} and exhibit antimicrobial properties.^{96,97} Additionally, PDA can produce diverse effects in the prevention and treatment of orthopedic diseases when combined with other materials. (Table 2). In clinical surgery, researchers have developed a multifunctional material, Ti-PDA@SNP-OGP, to address the antimicrobial deficiencies of titanium implants.⁹⁴ This composite integrates mesoporous polydopamine nanoparticles, the nitric oxide-releasing donor sodium nitroprusside (SNP), and osteogenic growth peptide (OGP) onto titanium implants. The goal is to effectively combat methicillin-resistant *Staphylococcus aureus* (MRSA) infections during implant replacement procedures. Furthermore, PDA acts as a functional modifier for cell-loaded hydrogels, enhancing the properties of the base materials. Pure hyaluronic acid (HA) hydrogels exhibit suboptimal mechanical properties. However, when HA hydrogels are modified by cross-linking with PDA, their mechanical properties are enhanced, and the critical gelation concentration is lowered.⁹⁸ This modification improves cellular affinity and tissue adhesion, while also imparting free radical scavenging and antibacterial capabilities to the hydrogel. Additionally, incorporating HAP into PDA-HA further boosts the hydrogels' osteogenic differentiation capabilities and exhibits significant antimicrobial potential.^{99,100} Synthetic polymers, such as polylactic acid (PLLA), polyglycolic acid (PGA), and polycaprolactone (PCL), serve as versatile substitutes for natural bone. These polymers exhibit notable biocompatibility, flexibility, and stability within the human body. However, these materials face challenges, including biologically inert surfaces and insufficient mechanical strength. To address these issues, recent studies have focused on functionalizing synthetic polymers notably through employing PDA as a nanoparticle coating.^{101,102} This approach leverages PDA's excellent adhesion and biocompatibility to enhance the surface activity of polymers and improve their mechanical strength, including tensile properties. Nanoparticles, integrated with PDA to create "building blocks" are rapidly advancing across various domains, including pharmaceuticals, inorganic compounds, proteins, and metal-organic frameworks (MOFs).¹⁰³

Evaluating the Impact of PDA Nanofilm Coatings on the Bone Reconstruction and Regeneration Microenvironment

The significant adhesive properties of PDA allow it to be applied to a wide range of material surfaces. To date, utilizing PDA has been recognized as a straightforward and efficacious strategy for modulating interfaces in biological tissue engineering. This coating can directly transmit biological signals to cells,^{114,115} enhance the hydrophilicity of the interface, and promote cell diffusion and attachment.^{116,117} Moreover, a specific surface roughness of the coating augments protein absorption, consequently enhancing cell adhesion.¹¹⁸ Furthermore, early studies have demonstrated that surface roughness at the micron and submicron levels significantly facilitates osteoblast differentiation.¹¹⁹ PDA nanofilm coatings have been extensively studied in

the treatment of orthopedic diseases,^{49,120–124} and for antimicrobial purposes,^{125–128} with strategies being progressively refined. The surface hydrophilicity of bone repair materials is closely related to their biocompatibility and cell adhesion. PDA contains substantial numbers of hydrophilic groups, such as amino and hydroxyl groups, which can be bound to hydrophobic surfaces, thereby effectively improving the hydrophilic properties of bone repair materials.¹²⁹ For example, polycaprolactone (PCL) is a commonly investigated material in the field of bone tissue engineering; however, its application is limited by its bioactivity. Numerous studies have reported using PDA to coat PCL surfaces, either in powder form or on scaffolds. PDA-coated PCL powder used for scaffold fabrication provides more hydrophilic surfaces for cell adhesion and growth than pure PCL scaffolds due to the presence of amino and hydroxyl functional groups.¹³⁰ A study demonstrated the effects of PDA coating under different conditions, including titanium (Ti), PDA-coated Ti samples, and PDA-coated Ti samples either stored for up to two weeks at room temperature or heated at 121 °C for 24 hours. The results emphasized that PDA coating heated at 121 °C for 24 hours did not impair the water contact angle and increased cell proliferation for both hDFs, HaCaTs, and MC3T3-E1 cells compared to pristine PDA. This underscores the importance of post-treatment and shelf-time for PDA coatings.¹³¹ Additionally, moderately hydrophilic materials can adsorb an optimal layer of proteins, fostering a conducive microenvironment for cell adhesion at the cell-material interface. Based on the rapid formation and accumulation of PDA nanoparticles, some researchers have used superhydrophilic PDA coatings synthesized in situ by Ag NPs in the presence of sodium periodate, which effectively reduced the non-specific adsorption of proteins.¹³² By using PDA materials, can regulate cell behavior at the interface of biomaterials, which can be better used for wound healing, including bone trauma treatment.¹³³ A novel Mn₃O₄@PDA@Pd-SS31 nanozyme targeting mitochondria was designed to reverse mitochondrial dysfunction and inhibit inflammation.¹³⁴ PDA has the ability to bind to various serum proteins, resulting in substrates that enhance cell attachment. On this hydrophilic matrix, cells exhibit excellent attachment, expansion, proliferation, and differentiation properties.¹³⁵ This is especially important in tissue engineering, regenerative medicine, and biosensor applications. Polydopamine nanopreparations when coated with polyvinylidene fluoride (PVDF) and integrated into nanocomposites with BaTiO₃, have been found to significantly promote cell viability and pre-osteoblast migration in vitro, as well as accelerate the formation of biomineralized apatite layers.¹⁰⁹ The study demonstrated that mPVDF-BT coatings, leveraging the adhesion and biocompatibility of PDA, were applied to rough Ti₆Al₄V biomaterial surfaces. PDA addresses the bio-inertness and enhances the long-term stability of Ti₆Al₄V, with the coated biomaterial showing potential in promoting bone reconstruction and regeneration. Ko et al developed a functional electrospun silk fibroin (SF) nanofiber scaffold that underwent conversion into two-stage HAP particles through PDA coating. Subsequent studies validated that HAP coated with PDA facilitates osteogenic differentiation and boosts bone formation both in vivo and in vitro.^{106,107} Furthermore, a coating on Hap/polyamide-66 (HA/P66) substrates was devised through a PDA-assisted biomimetic process, enhancing HAP dispersion and accelerating osseointegration.¹⁰⁸ The surface roughness and bioactive properties of the HAP coating are likely contributors to the osteogenic differentiation supported by Hap-PDA-HA/P66 substrates in mouse MSCs. Numerous studies have highlighted the ability of PDA to enhance cell adhesion and osteocyte differentiation.^{136–138} The primary mechanism underlying osteogenesis promotion by PDA is attributed to its enhancement of the material's hydrophilicity and surface roughness, thereby improving cellular access to the biomaterial surface. Moreover, PDA facilitates the integration of bioactive factors, including bone morphogenesis protein-2 (BMP-2) and vascular endothelial growth factor (VEGF), into its multilayer coatings.¹³⁹ This approach enhances osteoconductivity for cellular protein binding, further encouraging osteoblast adhesion, proliferation, and differentiation.¹⁴⁰ Research has revealed that PDA nanocoatings on biomaterials not only facilitate mesenchymal stem cell attachment but also trigger human cell reprogramming and sustain the long-term self-renewal of human pluripotent stem cells (hPSCs).¹⁴¹ Investigations have demonstrated that methoxypolyethyleneamine (mPEG-NH₂)-modified PDA NPs effectively downregulate pro-inflammatory cytokines in chondrocytes, thereby mitigating cartilage and subchondral bone inflammation in rat OA models.¹¹⁰ Moreover, the PDA NPs coating has been optimized for application on electrospun PCL fiber membranes, creating an optimal microenvironment for directing local stem cells towards osteoblast differentiation. Additionally, this fiber membrane acquires degradability from PDA NPs,¹¹¹ enhancing hydrophilicity and cytocompatibility alongside its inherent biodegradability. The findings indicate that PDA/PCL fiber membranes facilitate the diffusion, proliferation, and osteogenic activity of human mesenchymal stem cells (hMSCs) in vitro, exhibiting dose-dependent effectiveness. Significantly, in a mouse model presenting a critical-sized skull defect, the osteogenic differentiation of hMSCs was notably enhanced by PDA-modified biomaterials. PDA-based materials have demonstrated the capability to

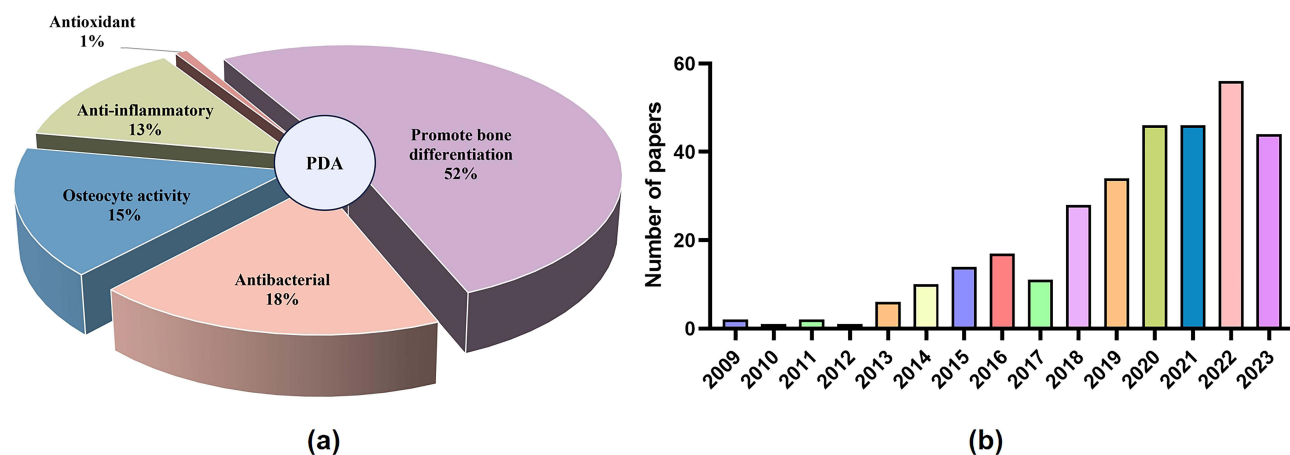


Figure 5 Distribution of PDA Applications and Research Trends. (a) Proportional representation of PDA application directions. (b) Annual distribution of orthopedic-related publications of PDA, including a total of 4 articles in 2024 currently.

eliminate reactive oxygen species (ROS) both in vitro and in vivo,⁵ with prior research delving into the mechanism of PDA NPs as ROS scavengers in dental applications, yielding promising outcomes. In vitro experiments revealed that PDA NPs act as scavengers for hydroxyl (HO) and superoxide radicals, achieving a scavenging rate of up to 90% for HO at a concentration of 0.1 mg/mL, with superoxide being entirely eliminated from the system as anticipated. Fluorescence imaging and antioxidant therapy experiments for in vivo ROS clearance showed that PDA NPs, at a dosage of 0.2 mg/site through subgingival injection, could efficiently eradicate local ROS and significantly mitigate periodontal inflammation. Furthermore, spectroscopy and additional in vitro experimental findings offer robust evidence that PDA NPs can eliminate various ROS and suppress ROS-induced inflammatory reactions,¹⁴² suggesting that PDA-NPs create an anti-inflammatory milieu conducive to bone reconstruction and regeneration.

Summary and Outlook

Bone, a critical tissue and organ for normal motor function in humans, has made the treatment of bone diseases a significant focus in medical field. Currently, biomaterials for bone disease treatment face challenges such as instability of drug-loaded materials, insufficient cell adhesion, reduced biocompatibility, and potential human toxicity from degradation products. Extensive research has demonstrated that biomaterials those modified with PDA surface engineering, possess antibacterial, anti-inflammatory, and antioxidant properties, can induce osteogenesis, enhance hMSC bone differentiation, promote osseointegration, and accelerate new bone formation. Since 2009, polydopamine nanoparticles have gained popularity among orthopedic treatment researches, with 52% of studies concentrating on osteogenic differentiation. This is followed by 18% on antimicrobial effects, 15% on enhancing cell activity, 13% on anti-inflammatory effects, and the rest on antioxidant properties (Figure 5). Research publications involved PDA on bone diseases treatment have significantly increased post-2017, peaking in 2022 with the highest number in recent years, predominantly focusing on promoting osteogenic differentiation (Table 3 and Figure 5). Polydopamine nanopreparations are becoming one of the key biomaterials for the treatment of bone diseases. As a drug delivery system, nanopreparations

Table 3 Summary of Research Publication on the Application of Polydopamine (PDA)

PDA Applications	Total Publications
Enhances Osteogenic Differentiation ^{5,24,26,40–42,49,57,72,77,78,80,82–84,90–95,99,100,104–106,108,112,119–124,126,129,138,140,143–292}	188
Enhances Antimicrobial Activity ^{53,76,80,81,90,96,97,99,100,112,113,125–128,211,223,278,293–340}	66
Stimulates Relevant Cellular Functions ^{24,79,106,136–141,146,160,169,183,188,189,193,210,215,221,222,226,227,240,264,265,285,287,341–370}	57
Enhances Anti-inflammatory Properties ^{43,70,72,73,94,97,110,146,228,249,269,371–406}	47
Enhances Antioxidant Activity ^{76,407,408}	3

incorporating PDA exhibit high drug loading capacity, effective sustained-release properties, and delayed drug clearance, offering significant potential to address current challenges in the clinical treatment of bone diseases. Furthermore, PDA can functionalize a diverse array of nanomaterials, facilitating the development of a PDA-based multifunctional platform for targeted or synergistic therapy. Despite the limited clinical translation of many biomaterials in the realm of bone disease treatment, PDA preparations, whether employed as biologically functional additives or drug carriers, are expected to play a more significant role in managing and preventing orthopedic disorders.

Data Sharing Statement

The data presented in this study are available in this article.

Funding

This research was funded by grants from the National Natural Science Foundation of China (No. 81703584), The regional joint fund of natural science foundation of Guangdong province (No. 2020B1515120052), Guangdong Province Natural Science Foundation of China (No. 2022A1515220166, 2023A1515011091, and 2021A1515010975), Discipline construction project of Guangdong Medical University (No. 4SG23002G, and CLP2021B012), the Science and Technology Foundation of Zhanjiang (No. 2022A01099, 2022A01163, 2022A01170), the Discipline Construction Fund of Central People's Hospital of Zhanjiang (No. 2022A09), Special Funds for Scientific Technological Innovation of Undergraduates in Guangdong Province (No. pdjh2022a0214), Guangdong medical university research fund (No. FZZM05, FYZM001).

Disclosure

Min Zhang, Man Mi, and Zilong Hu are co-first authors for this study. The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

1. Wang C, Bai J, Liu Y, Jia X, Jiang X. Polydopamine coated selenide molybdenum: a new photothermal nanocarrier for highly effective chemo-photothermal synergistic therapy. *ACS Biomater Sci Eng*. 2016;2(11):2011–2017. doi:10.1021/acsbomaterials.6b00416
2. Hao M, Kong C, Jiang C, et al. Polydopamine-coated Au-Ag nanoparticle-guided photothermal colorectal cancer therapy through multiple cell death pathways. *Acta Biomater*. 2019;83:414–424. doi:10.1016/j.actbio.2018.10.032
3. Wu Q, Niu M, Chen X, et al. Biocompatible and biodegradable zeolitic imidazolate framework/polydopamine nanocarriers for dual stimulus triggered tumor thermo-chemotherapy. *Biomaterials*. 2018;162:132–143. doi:10.1016/j.biomaterials.2018.02.022
4. Lin LS, Cong ZX, Cao JB, et al. Multifunctional Fe₃O₄@polydopamine core-shell nanocomposites for intracellular mRNA detection and imaging-guided photothermal therapy. *ACS nano*. 2014;8(4):3876–3883. doi:10.1021/nn500722y
5. Sharma V, Chowdhury S, Bose S, Basu B. Polydopamine Codoped BaTiO(3)-Functionalized polyvinylidene fluoride coating as a piezo-biomaterial platform for an enhanced cellular response and bioactivity. *ACS Biomater Sci Eng*. 2022;8(1):170–184. doi:10.1021/acsbomaterials.1c00879
6. Cheng W, Zeng X, Chen H, et al. Versatile polydopamine platforms: synthesis and promising applications for surface modification and advanced nanomedicine. *ACS nano*. 2019;13(8):8537–8565. doi:10.1021/acsnano.9b04436
7. Deng Z, Shang B, Peng B. Polydopamine based colloidal materials: synthesis and applications. *Chem Rec*. 2018;18(4):410–432. doi:10.1002/tcr.201700051
8. Alfieri ML, Panzella L, Oscurato SL, et al. The chemistry of polydopamine film formation: the amine-quinone interplay. *Biomimetics*. 2018;3(3). doi:10.3390/biomimetics3030026
9. García-Mayorga JC, Rosu HC, Jasso-Salcedo AB, Escobar-Barríos VA. Kinetic study of polydopamine sphere synthesis using TRIS: relationship between synthesis conditions and final properties. *RSC Adv*. 2023;13(8):5081–5095. doi:10.1039/d2ra06669f
10. Tan L, Zhu T, Huang Y, et al. Ozone-induced rapid and green synthesis of polydopamine coatings with high uniformity and enhanced stability. *Adv Sci*. 2024;11(10):e2308153. doi:10.1002/adv.202308153
11. Hemmatpour H, De Luca O, Crestani D, et al. New insights in polydopamine formation via surface adsorption. *Nature Commun*. 2023;14(1):664. doi:10.1038/s41467-023-36303-8
12. Ryu JH, Messersmith PB, Lee H. Polydopamine surface chemistry: a decade of discovery. *ACS Appl Mater Interfaces*. 2018;10(9):7523–7540. doi:10.1021/acsmi.7b19865
13. Chinchulkar SA, Patra P, Dehariya D, Yu A, Rengan AK. Polydopamine nanocomposites and their biomedical applications: a review. *Polym Adv Technol*. 2022;33(12):3935–3956. doi:10.1002/pat.5863
14. Alfieri ML, Weil T, Ng DYW, Ball V. Polydopamine at biological interfaces. *Adv Colloid Interface Sci*. 2022;305:102689. doi:10.1016/j.cis.2022.102689

15. Han X, Gao W, Zhou Z, et al. Curcumin-loaded mesoporous polydopamine nanoparticles modified by quaternized chitosan against bacterial infection through synergistic effect. *Int J Biol Macromol*. 2024;267:131372. doi:10.1016/j.ijbiomac.2024.131372
16. Hu J, Ding Y, Tao B, et al. Surface modification of titanium substrate via combining photothermal therapy and quorum-sensing-inhibition strategy for improving osseointegration and treating biofilm-associated bacterial infection. *Bioact. Mater*. 2022;18:228–241. doi:10.1016/j.bioactmat.2022.03.011
17. Li F, Yu Y, Wang Q, Yuan J, Wang P, Fan X. Polymerization of dopamine catalyzed by laccase: comparison of enzymatic and conventional methods. *Enzyme Microb Technol*. 2018;119:58–64. doi:10.1016/j.enzmictec.2018.09.003
18. Lee H, Dellatore SM, Miller WM, Messersmith PB. Mussel-inspired surface chemistry for multifunctional coatings. *Science*. 2007;318:426–30. doi:10.1126/science.1147241
19. Cihanoglu A, Schiffman JD, Altinkaya SA. Ultrasound-assisted dopamine polymerization: rapid and oxidizing agent-free polydopamine coatings on membrane surfaces. *Chem Commun*. 2021;57(100):13740–13743. doi:10.1039/d1cc05960b
20. Zhou X, Gao S, Huang D, et al. Bioinspired, ultra-fast polymerization of dopamine under mild conditions. *Macromol Rapid Commun*. 2022;43(23):e2200581. doi:10.1002/marc.202200581
21. Tan Y, Deng W, Li Y, et al. Polymeric bionanocomposite cast thin films with in situ laccase-catalyzed polymerization of dopamine for biosensing and biofuel cell applications. *J Phys Chem A*. 2010;114(15):5016–5024. doi:10.1021/jp100922t
22. Li H, Yin D, Li W, Tang Q, Zou L, Peng Q. Polydopamine-based nanomaterials and their potentials in advanced drug delivery and therapy. *Colloids Surf B*. 2021;199:111502. doi:10.1016/j.colsurfb.2020.111502
23. Hu H, Dyke JC, Bowman BA, Ko CC, You W. Investigation of dopamine analogues: synthesis, mechanistic understanding, and structure-property relationship. *Langmuir*. 2016;32(38):9873–9882. doi:10.1021/acs.langmuir.6b02141
24. Liu G, Ma M, Yang H, et al. Chitosan/polydopamine/octacalcium phosphate composite microcarrier simulates natural bone components to induce osteogenic differentiation of stem cells. *Biomater Adv*. 2023;154:213642. doi:10.1016/j.bioadv.2023.213642
25. Tan L, Tang W, Liu T, et al. Biocompatible hollow polydopamine nanoparticles loaded ionic liquid enhanced tumor microwave thermal ablation in vivo. *ACS Appl Mater Interfaces*. 2016;8(18):11237–11245. doi:10.1021/acsami.5b12329
26. Treccani L, Yvonne Klein T, Meder F, Pardun K, Rezwani K. Functionalized ceramics for biomedical, biotechnological and environmental applications. *Acta Biomater*. 2013;9(7):7115–7150. doi:10.1016/j.actbio.2013.03.036
27. Cho S, Park W, Kim DH. Silica-coated metal chelating-melanin nanoparticles as a dual-modal contrast enhancement imaging and therapeutic agent. *ACS Appl Mater Interfaces*. 2017;9(1):101–111. doi:10.1021/acsami.6b11304
28. Hashemi-Moghaddam H, Kazemi-Bagsangani S, Jamili M, Zavareh S. Evaluation of magnetic nanoparticles coated by 5-fluorouracil imprinted polymer for controlled drug delivery in mouse breast cancer model. *Int J Pharm*. 2016;497(1–2):228–238. doi:10.1016/j.ijpharm.2015.11.040
29. Ho CC, Ding SJ. The pH-controlled nanoparticles size of polydopamine for anti-cancer drug delivery. *J Mater Sci Mater Med*. 2013;24(10):2381–2390. doi:10.1007/s10856-013-4994-2
30. Mei S, Kochovsky Z, Roa R, et al. Enhanced catalytic activity of Gold@Polydopamine nanoreactors with multi-compartment structure under NIR irradiation. *Nanomicro Lett*. 2019;11(1):83. doi:10.1007/s40820-019-0314-9
31. Khan MZH, Daizy M, Tarafder C, Liu X. Au-PDA@SiO₂ core-shell nanospheres decorated rGO modified electrode for electrochemical sensing of cefotaxime. *Sci Rep*. 2019;9(1):19041. doi:10.1038/s41598-019-55517-9
32. Zhang Y, Zhao Z, Li D, et al. In situ growth of MnO₂ on pDA-templated cotton fabric for degradation of formaldehyde. *Cellulose*. 2022;29(13):7353–7363. doi:10.1007/s10570-022-04734-z
33. Behzadinasab S, Williams MD, Hosseini M, et al. Transparent and sprayable surface coatings that kill drug-resistant bacteria within minutes and inactivate SARS-CoV-2 virus. *ACS Appl Mater Interfaces*. 2021;13(46):54706–54714. doi:10.1021/acsami.1c15505
34. Zhang Z, Si T, Liu J, Zhou G. In-situ grown silver nanoparticles on nonwoven fabrics based on mussel-inspired polydopamine for highly sensitive SERS carbaryl pesticides detection. *Nanomaterials*. 2019;9(3). doi:10.3390/nano9030384
35. Liu G, Lu Z, Zhu X, et al. Facile in-situ growth of Ag/TiO₂ nanoparticles on polydopamine modified bamboo with excellent mildew-proofing. *Sci Rep*. 2019;9(1):16496. doi:10.1038/s41598-019-53001-y
36. Xie C, Li P, Han L, et al. Electroresponsive and cell-affinitive polydopamine/polypyrrole composite microcapsules with a dual-function of on-demand drug delivery and cell stimulation for electrical therapy. *NPG Asia Materials*. 2017;9(3):e358–e358. doi:10.1038/am.2017.16
37. Tran HQ, Bhave M, Xu G, Sun C, Yu A. Synthesis of polydopamine hollow capsules via a polydopamine mediated silica water dissolution process and its application for enzyme encapsulation. *Front Chem*. 2019;7:468. doi:10.3389/fchem.2019.00468
38. Liu Y, Su C, Zu Y, Chen X, Sha J, Dai J. Ultrafast deposition of polydopamine for high-performance fiber-reinforced high-temperature ceramic composites. *Sci Rep*. 2022;12(1):20489. doi:10.1038/s41598-022-24971-3
39. Wang P, Zhang Y-L, K-L F, et al. Zinc-coordinated polydopamine surface with a nanostructure and superhydrophilicity for antibiofouling and antibacterial applications. *Mater Adv*. 2022;3(13):5476–5487. doi:10.1039/D2MA00482H
40. Cheng D, Ding R, Jin X, et al. Strontium Ion-functionalized nano-hydroxyapatite/chitosan composite microspheres promote osteogenesis and angiogenesis for bone regeneration. *ACS Appl Mater Interfaces*. 2023;15(16):19951–19965. doi:10.1021/acsami.3c00655
41. Xiong W, Yuan L, Wang L, et al. Preparation of berberine-naringin dual drug-loaded composite microspheres and evaluation of their antibacterial-osteogenic properties. *Zhongguo Xiu Fu Chong Jian Wai Ke Za Zhi*. 2023;37(12):1505–1513. doi:10.7507/1002-1892.202308054
42. Wang Y, Hu Y, Lan S, et al. A recombinant parathyroid hormone-related peptide locally applied in osteoporotic bone defect. *Adv Sci*. 2023;10(22):e2300516. doi:10.1002/advs.202300516
43. Gan D, Huang Z, Wang X, et al. Bioadhesive and electroactive hydrogels for flexible bioelectronics and supercapacitors enabled by a redox-active core-shell PEDOT@PZIF-71 system. *Mater Horiz*. 2023;10(6):2169–2180. doi:10.1039/d2mh01234k
44. Baccaro LF, Conde DM, Costa-Paiva L, Pinto-Neto AM. The epidemiology and management of postmenopausal osteoporosis: a viewpoint from Brazil. *Clin Interv Aging*. 2015;10:583–591. doi:10.2147/cia.S54614
45. Vina ER, Kwok CK. Epidemiology of osteoarthritis: literature update. *Curr Opin Rheumatol*. 2018;30(2):160–167. doi:10.1097/bor.0000000000000479
46. Lv H, Chen W, Yao M, Hou Z, Zhang Y. Collecting data on fractures: a review of epidemiological studies on orthopaedic traumatology and the Chinese experience in large volume databases. *Int Orthop*. 2022;46(5):945–951. doi:10.1007/s00264-022-05299-z

47. Zhou Z, Wang X, Yu H, Yu C, Zhang F. Dynamic cross-linked polyurea/polydopamine nanocomposites for photoresponsive self-healing and photoactuation. *Macromolecules*. 2022;55(6):2193–2201. doi:10.1021/acs.macromol.1c02534
48. Lu L, Niu W, Li J, et al. Rapid photo-responsive self-healing cross-linked polyurea/ polydopamine nanocomposites with multiple dynamic bonds and bio-based rosin. *Compos Sci Technol*. 2024;254:110693. doi:10.1016/j.compscitech.2024.110693
49. Zhao Y, He P, Yao J, et al. pH/NIR-responsive and self-healing coatings with bacteria killing, osteogenesis, and angiogenesis performances on magnesium alloy. *Biomaterials*. 2023;301:122237. doi:10.1016/j.biomaterials.2023.122237
50. Dong Z, Gong H, Gao M, et al. Polydopamine nanoparticles as a versatile molecular loading platform to enable imaging-guided cancer combination therapy. *Theranostics*. 2016;6(7):1031–1042. doi:10.7150/thno.14431
51. Deng JH, Luo J, Mao YL, et al. π - π stacking interactions: non-negligible forces for stabilizing porous supramolecular frameworks. *Sci Adv*. 2020;6(2):eaax9976. doi:10.1126/sciadv.aax9976
52. Li JY, Long XY, Yin HX, Qiao JQ, Lian HZ. Magnetic solid-phase extraction based on a polydopamine-coated Fe₃O₄ nanoparticles absorbent for the determination of bisphenol A, tetrabromobisphenol A, 2,4,6-tribromophenol, and (S)-1,1'-bi-2-naphthol in environmental waters by HPLC. *J Separat Sci*. 2016;39(13):2562–2572. doi:10.1002/jssc.201600231
53. Zhang M, Fan Z, Zhang J, et al. Multifunctional chitosan/alginate hydrogel incorporated with bioactive glass nanocomposites enabling photothermal and nitric oxide release activities for bacteria-infected wound healing. *Int J Biol Macromol*. 2023;232:123445. doi:10.1016/j.ijbiomac.2023.123445
54. Shlapakova LE, Botvin VV, Mukhortova YR, et al. Magnetoactive composite conduits based on Poly(3-hydroxybutyrate) and magnetite nanoparticles for repair of peripheral nerve injury. *ACS Appl Bio Mater*. 2024;7(2):1095–1114. doi:10.1021/acsabm.3c01032
55. Yang J, Wu J, Guo Z, Zhang G, Zhang H. Iron oxide nanoparticles combined with static magnetic fields in bone remodeling. *Cells*. 2022;11;20. doi:10.3390/cells11203298.
56. Siciliano G, Monteduro AG, Turco A, et al. Polydopamine-coated magnetic iron oxide nanoparticles: from design to applications. *Nanomaterials*. 2022;12(7). doi:10.3390/nano12071145
57. Huang Z, He Y, Chang X, et al. A magnetic iron oxide/polydopamine coating can improve osteogenesis of 3D-printed porous titanium scaffolds with a static magnetic field by upregulating the TGF β -smads pathway. *Adv Healthc Mater*. 2020;9(14):e2000318. doi:10.1002/adhm.202000318
58. Liu M, Yu W, Zhang F, et al. Fe₃O₄@Polydopamine-Labeled MSCs targeting the spinal cord to treat neuropathic pain under the guidance of a magnetic field. *Int J Nanomed*. 2021;16:3275–3292. doi:10.2147/ijn.S296398
59. Yang W, Hu H, Pan Q, Deng X, Zhang Y, Shao Z. Iron-polydopamine coated multifunctional nanoparticle SiO₂@PDA/Fe₃+FA mediated low temperature photothermal for chemodynamic therapy of cisplatin-insensitive osteosarcoma. *Mater Des*. 2023;227:111785. doi:10.1016/j.matdes.2023.111785
60. Dai S, Yue S, Ning Z, Jiang N, Gan Z. Polydopamine nanoparticle-reinforced near-infrared light-triggered shape memory polycaprolactone-polydopamine polyurethane for biomedical implant applications. *ACS Appl Mater Interfaces*. 2022;14(12):14668–14676. doi:10.1021/acsami.2c03172
61. Cao X, Liu H, Yang X, Tian J, Luo B, Liu M. Halloysite nanotubes@polydopamine reinforced polyacrylamide-gelatin hydrogels with NIR light triggered shape memory and self-healing capability. *Compos Sci Technol*. 2020;191:108071. doi:10.1016/j.compscitech.2020.108071
62. Ni X, Gao Y, Zhang X, Lei Y, Sun G, You B. An eco-friendly smart self-healing coating with NIR and pH dual-responsive superhydrophobic properties based on biomimetic stimuli-responsive mesoporous polydopamine microspheres. *Chem Eng J*. 2021;406:126725. doi:10.1016/j.cej.2020.126725
63. Wang Q, Yan X, Liu P, Xu Y, Guan Q, You Z. Near-Infrared Light Triggered the Shape Memory Behavior of Polydopamine-Nanoparticle-Filled Epoxy Acrylate. *Polymers*. 2023;15(16). doi:10.3390/polym15163394
64. Wei Y, Qi X, He S, Deng S, Liu D, Fu Q. Gradient polydopamine coating: a simple and general strategy toward multishape memory effects. *ACS Appl Mater Interfaces*. 2018;10(38):32922–32934. doi:10.1021/acsami.8b13134
65. Yang L, Tong R, Wang Z, Xia H. Polydopamine particle-filled shape-memory polyurethane composites with fast near-infrared light responsibility. *Chemphyschem*. 2018;19(16):2052–2057. doi:10.1002/cphc.201800022
66. Ha YM, Kim YN, Jung YC. Rapid and local self-healing ability of polyurethane nanocomposites using photothermal polydopamine-coated graphene oxide triggered by near-infrared laser. *Polymers*. 2021;13(8). doi:10.3390/polym13081274
67. Seth A, Gholami Derami H, Gupta P, et al. Polydopamine-mesoporous silica core-shell nanoparticles for combined photothermal immunotherapy. *ACS Appl Mater Interfaces*. 2020;12(38):42499–42510. doi:10.1021/acsami.0c10781
68. Liu J, Peng Q. Protein-gold nanoparticle interactions and their possible impact on biomedical applications. *Acta Biomater*. 2017;55:13–27. doi:10.1016/j.actbio.2017.03.055
69. Wong S, Cao C, Lessio M, Stenzel MH. Sugar-induced self-assembly of curcumin-based polydopamine nanocapsules with high loading capacity for dual drug delivery. *Nanoscale*. 2022;14(26):9448–9458. doi:10.1039/d2nr01795d
70. Chin KY. The spice for joint inflammation: anti-inflammatory role of curcumin in treating osteoarthritis. *Drug Des Devel Ther*. 2016;10:3029–3042. doi:10.2147/dddt.S117432
71. Nador F, Guisasaola E, Baeza A, Villaeija MA, Vallet-Regí M, Ruiz-Molina D. Synthesis of polydopamine-like nanocapsules via removal of a sacrificial mesoporous silica template with water. *Chemistry*. 2017;23(12):2753–2758. doi:10.1002/chem.201604631
72. Ouyang Z, Tan T, Liu C, et al. Targeted delivery of hesperetin to cartilage attenuates osteoarthritis by bimodal imaging with Gd(2)(CO(3))(3)@PDA nanoparticles via TLR-2/NF- κ B/Akt signaling. *Biomaterials*. 2019;205:50–63. doi:10.1016/j.biomaterials.2019.03.018
73. Nie J, Cheng W, Peng Y, et al. Co-delivery of docetaxel and bortezomib based on a targeting nanoplatform for enhancing cancer chemotherapy effects. *Drug Delivery*. 2017;24(1):1124–1138. doi:10.1080/10717544.2017.1362677
74. Wang Y, Ge W, Ma Z, et al. Use of mesoporous polydopamine nanoparticles as a stable drug-release system alleviates inflammation in knee osteoarthritis. *APL Bioeng*. 2022;6(2):026101. doi:10.1063/5.0088447
75. Wang JL, Ren KF, Chang H, Zhang SM, Jin LJ, Ji J. Facile fabrication of robust superhydrophobic multilayered film based on bioinspired poly(dopamine)-modified carbon nanotubes. *Phys Chemist Chem Phys*. 2014;16(7):2936–2943. doi:10.1039/c3cp54354d
76. Demirci S, Sahiner M, Suner SS, Sahiner N. Improved biomedical properties of polydopamine-coated carbon nanotubes. *Micromachines*. 2021;12(11). doi:10.3390/mi12111280

77. Lai M, Jin Z, Su Z. Surface modification of TiO₂ nanotubes with osteogenic growth peptide to enhance osteoblast differentiation. *Mater Sci Eng C Mater Biol Appl.* 2017;73:490–497. doi:10.1016/j.msec.2016.12.083
78. Jin M, Zhu J, Meng Z, et al. TiO₂ nanotubes-MoS₂/PDA-LL-37 exhibits efficient anti-bacterial activity and facilitates new bone formation under near-infrared laser irradiation. *Biom Mater.* 2022;17(4). doi:10.1088/1748-605X/ac6470
79. He Y, Mu C, Shen X, et al. Peptide LL-37 coating on micro-structured titanium implants to facilitate bone formation in vivo via mesenchymal stem cell recruitment. *Acta Biomater.* 2018;80:412–424. doi:10.1016/j.actbio.2018.09.036
80. Zhang G, Zhang X, Yang Y, et al. Dual light-induced in situ antibacterial activities of biocompatible TiO₂/MoS₂/PDA/RGD nanorod arrays on titanium. *Biomater Sci.* 2020;8(1):391–404. doi:10.1039/c9bm01507h
81. Luo R, Tang L, Wang J, et al. Improved immobilization of biomolecules to quinone-rich polydopamine for efficient surface functionalization. *Colloids Surf B.* 2013;106:66–73. doi:10.1016/j.colsurfb.2013.01.033
82. Godoy-Gallardo M, Portolés-Gil N, López-Periago AM, Domingo C, Hosta-Rigau L. Multi-layered polydopamine coatings for the immobilization of growth factors onto highly-interconnected and bimodal PCL/HA-based scaffolds. *Mater Sci Eng C Mater Biol Appl.* 2020;117:111245. doi:10.1016/j.msec.2020.111245
83. Hong L, Yuan L, Xu X, et al. Biocompatible Nanotube-Strontium/polydopamine-arginine-glycine-aspartic acid coating on Ti6Al4V enhances osteogenic properties for biomedical applications. *Microsc Res Techn.* 2022;85(4):1518–1526. doi:10.1002/jemt.24014
84. Sun H, Shang Y, Guo J, et al. Artificial periosteum with oriented surface nanotopography and high tissue adherent property. *ACS Appl Mater Interfaces.* 2023;15(39):45549–45560. doi:10.1021/acsami.3c07561
85. Gan D, Xing W, Jiang L, et al. Plant-inspired adhesive and tough hydrogel based on Ag-Lignin nanoparticles-triggered dynamic redox catechol chemistry. *Nat Commun.* 2019;10(1):1487. doi:10.1038/s41467-019-09351-2
86. Shi W, Li H, Chen J, et al. Stretchable, self-healing, and bioactive hydrogel with high-functionality N,N-bis(acryloyl)cystamine dynamically bonded Ag@polydopamine crosslinkers for wearable sensors. *Adv Sci.* 2024:e2404451. doi:10.1002/advs.202404451
87. Chen C, Zheng N, Wu W, et al. Self-adhesive and conductive dual-network polyacrylamide hydrogels reinforced by aminated lignin, dopamine, and biomass carbon aerogel for ultrasensitive pressure sensor. *ACS Appl Mater Interfaces.* 2022;14(48):54127–54140. doi:10.1021/acsami.2c12914
88. Li Y, Yang D, Wu Z, et al. Self-adhesive, self-healing, biocompatible and conductive polyacrylamide nanocomposite hydrogels for reliable strain and pressure sensors. *Nano Energy.* 2023;109:108324. doi:10.1016/j.nanoen.2023.108324
89. Xiong F, Wei S, Sheng H, et al. Three-layer core-shell structure of polypyrrole/polydopamine/poly(L-lactide) nanofibers for wound healing application. *Int J Biol Macromol.* 2022;222(Pt B):1948–1962. doi:10.1016/j.ijbiomac.2022.09.284
90. Xu X, Liu X, Tan L, et al. Controlled-temperature photothermal and oxidative bacteria killing and acceleration of wound healing by polydopamine-assisted Au-hydroxyapatite nanorods. *Acta Biomater.* 2018;77:352–364. doi:10.1016/j.actbio.2018.07.030
91. Chen M, Chen Y, Wei C. Nanoparticles based composite coatings with tunable vascular endothelial growth factor and bone morphogenetic protein-2 release for bone regeneration. *J Biomed Mater Res A.* 2023;111(7):1044–1053. doi:10.1002/jbm.a.37489
92. Han R, Min Y, Li G, Chen S, Xie M, Zhao Z. Supercritical CO₂-assisted fabrication of CM-PDA/SF/nHA nanofibrous scaffolds for bone regeneration and chemo-photothermal therapy against osteosarcoma. *Biomater Sci.* 2023;11(15):5218–5231. doi:10.1039/d3bm00532a
93. Wu Y, Huo S, Liu S, Hong Q, Wang Y, Lyu Z. Cu-Sr bilayer bioactive glass nanoparticles/polydopamine functionalized polyetheretherketone enhances osteogenic activity and prevents implant-associated infections through spatiotemporal immunomodulation. *Adv Healthc Mater.* 2023;12(32):e2301772. doi:10.1002/adhm.202301772
94. Yu YL, Wu JJ, Lin CC, et al. Elimination of methicillin-resistant *Staphylococcus aureus* biofilms on titanium implants via photothermally-triggered nitric oxide and immunotherapy for enhanced osseointegration. *Mil Med Res.* 2023;10(1):21. doi:10.1186/s40779-023-00454-y
95. Jiang X, Lei L, Sun W, et al. [Bioceramic scaffolds with two-step internal/external modification of copper-containing polydopamine enhance antibacterial and alveolar bone regeneration capability]. 含铜聚多巴胺内外双修饰法构建抗菌促骨再生生物陶瓷支架. *J Zhejiang Univ Sci B.* 2024;25(1):65–82. doi:10.1631/jzus.B23d0004
96. Equy E, Hirtzel J, Hellé S, et al. Fluorescent bioinspired albumin/polydopamine nanoparticles and their interactions with *Escherichia coli* cells. *Beilstein J Nanotechnol.* 2023;14:1208–1224. doi:10.3762/bjnano.14.100
97. Langeder J, Döring K, Schmietendorf H, Grienke U, Schmidtke M, Rollinger JM. (1)H NMR-based biochemometric analysis of morus alba extracts toward a multipotent herbal anti-infective. *J Nat Prod.* 2023;86(1):8–17. doi:10.1021/acs.jnatprod.2c00481
98. Peng L, Liang Y, Yue J, et al. Dramatic improvement in the mechanical properties of polydopamine/polyacrylamide hydrogel mediated human amniotic membrane. *RSC Adv.* 2023;13(6):3635–3642. doi:10.1039/d2ra07622e
99. Ma W, Chen H, Cheng S, Wu C, Wang L, Du M. Gelatin hydrogel reinforced with mussel-inspired polydopamine-functionalized nanohydroxyapatite for bone regeneration. *Int J Biol Macromol.* 2023;240:124287. doi:10.1016/j.ijbiomac.2023.124287
100. Zheng P, Ding B, Li G. Polydopamine-incorporated nanoformulations for biomedical applications. *Macromol biosci.* 2020;20(12):e2000228. doi:10.1002/mabi.202000228
101. Teo AJT, Mishra A, Park I, Kim YJ, Park WT, Yoon YJ. Polymeric biomaterials for medical implants and devices. *ACS Biomater Sci Eng.* 2016;2(4):454–472. doi:10.1021/acsbiomaterials.5b00429
102. Liu Y, Ai K, Lu L. Polydopamine and its derivative materials: synthesis and promising applications in energy, environmental, and biomedical fields. *Chem. Rev.* 2014;114(9):5057–5115. doi:10.1021/cr400407a
103. Jin A, Wang Y, Lin K, Jiang L. Nanoparticles modified by polydopamine: working as “drug” carriers. *Bioact. Mater.* 2020;5(3):522–541. doi:10.1016/j.bioactmat.2020.04.003
104. Wu HY, Lin YH, Lee AK, Kuo TY, Tsai CH, Shie MY. Combined effects of polydopamine-assisted copper immobilization on 3D-Printed Porous Ti6Al4V scaffold for angiogenic and osteogenic bone regeneration. *Cells.* 2022;11(18). doi:10.3390/cells11182824
105. Wang H, Yuan C, Lin K, Zhu R, Zhang S. Modifying a 3D-printed Ti6Al4V implant with polydopamine coating to improve BMSCs growth, osteogenic differentiation, and in situ osseointegration in vivo. *Front Bioeng Biotechnol.* 2021;9:761911. doi:10.3389/fbioe.2021.761911
106. Ko E, Lee JS, Kim H, et al. Electrospun silk fibroin nanofibrous scaffolds with two-stage hydroxyapatite functionalization for enhancing the osteogenic differentiation of human adipose-derived mesenchymal stem cells. *ACS Appl Mater Interfaces.* 2018;10(9):7614–7625. doi:10.1021/acsami.7b03328

107. Guzman RE, Evans MG, Bove S, Morenko B, Kilgore K. Mono-iodoacetate-induced histologic changes in subchondral bone and articular cartilage of rat femorotibial joints: an animal model of osteoarthritis. *Toxicol Pathol.* 2003;31(6):619–624. doi:10.1080/01926230390241800
108. Xu Y, Li H, Wu J, Yang Q, Jiang D, Qiao B. Polydopamine-induced hydroxyapatite coating facilitates hydroxyapatite/polyamide 66 implant osteogenesis: an in vitro and in vivo evaluation. *Int j Nanomed.* 2018;13:8179–8193. doi:10.2147/ijn.S181137
109. Yan S, Huang Q, Chen J, et al. Tumor-targeting photodynamic therapy based on folate-modified polydopamine nanoparticles. *Int j Nanomed.* 2019;14:6799–6812. doi:10.2147/ijn.S216194
110. Wu Z, Yuan K, Zhang Q, Guo JJ, Yang H, Zhou F. Antioxidant PDA-PEG nanoparticles alleviate early osteoarthritis by inhibiting osteoclastogenesis and angiogenesis in subchondral bone. *J Nanobiotechnology.* 2022;20(1):479. doi:10.1186/s12951-022-01697-y
111. Xie M, Ge J, Lei B, Zhang Q, Chen X, Star-Shaped MPX. Biodegradable, and Elastomeric PLLA-PEG-POSS hybrid membrane with biomineralization activity for guiding bone tissue regeneration. *Macromol biosci.* 2015;15(12):1656–1662. doi:10.1002/mabi.201500237
112. Qian Y, Zhou X, Zhang F, Diekwisch TGH, Luan X, Yang J. Triple PLGA/PCL scaffold modification including silver impregnation, collagen coating, and electrospinning significantly improve biocompatibility, antimicrobial, and osteogenic properties for orofacial tissue regeneration. *ACS Appl Mater Interfaces.* 2019;11(41):37381–37396. doi:10.1021/acsami.9b07053
113. Zhou Z, Yao Q, Li L, et al. Antimicrobial Activity of 3D-Printed Poly(ϵ -Caprolactone) (PCL) Composite Scaffolds Presenting Vancomycin-Loaded Polylactic Acid-Glycolic Acid (PLGA) Microspheres. *Med Sci Monit.* 2018;24:6934–6945. doi:10.12659/msm.911770
114. Zhong S, Luo R, Wang X, et al. Effects of polydopamine functionalized titanium dioxide nanotubes on endothelial cell and smooth muscle cell. *Colloids Surf B.* 2014;116:553–560. doi:10.1016/j.colsurfb.2014.01.030
115. Chien CY, Tsai WB. Poly(dopamine)-assisted immobilization of Arg-Gly-Asp peptides, hydroxyapatite, and bone morphogenic protein-2 on titanium to improve the osteogenesis of bone marrow stem cells. *ACS Appl Mater Interfaces.* 2013;5(15):6975–6983. doi:10.1021/am401071f
116. Zhao MH, Chen XP, Wang Q. Wetting failure of hydrophilic surfaces promoted by surface roughness. *Sci Rep.* 2014;4:5376. doi:10.1038/srep05376
117. Wang W, Tang Z, Zhang Y, Wang Q, Liang Z, Zeng X. Mussel-inspired polydopamine: the bridge for targeting drug delivery system and synergistic cancer treatment. *Macromol biosci.* 2020;20(10):e2000222. doi:10.1002/mabi.202000222
118. Al Qahtani WM, Schille C, Spintzyk S, et al. Effect of surface modification of zirconia on cell adhesion, metabolic activity and proliferation of human osteoblasts. *Biomed Tech.* 2017;62(1):75–87. doi:10.1515/bmt-2015-0139
119. Gittens RA, McLachlan T, Olivares-Navarrete R, et al. The effects of combined micron-/submicron-scale surface roughness and nanoscale features on cell proliferation and differentiation. *Biomaterials.* 2011;32(13):3395–3403. doi:10.1016/j.biomaterials.2011.01.029
120. Chen M, Li M, Ren X, et al. DNAzyme Nanoconstruct-Integrated Autonomously-Adaptive Coatings Enhance Titanium-Implant Osteointegration by Cooperative Angiogenesis and Vessel Remodeling. *ACS Nano.* 2023;17(16):15942–15961. doi:10.1021/acsnano.3c04049
121. Cheng X, Yang X, Liu C, et al. Stabilization of apatite coatings on PPENK surfaces by mechanical interlocking to promote bioactivity and osseointegration in vivo. *ACS Appl Mater Interfaces.* 2023;15(1):697–710. doi:10.1021/acsami.2c20633
122. Davaie S, Hooshmand T, Najafi F, Haghbin Nazarpak M, Pirmoradian M. Synthesis, characterization, and induced osteogenic differentiation effect of collagen membranes functionalized by polydopamine/graphene oxide for bone tissue engineering. *ACS Appl Bio Mater.* 2023;6(11):4629–4644. doi:10.1021/acsbm.3c00400
123. Li Y, Liu C, Cheng X, et al. PDA-BPs integrated mussel-inspired multifunctional hydrogel coating on PPENK implants for anti-tumor therapy, antibacterial infection and bone regeneration. *Bioact. Mater.* 2023;27:546–559. doi:10.1016/j.bioactmat.2023.04.020
124. Liu Q, Chen M, Gu P, et al. Covalently grafted biomimetic matrix reconstructs the regenerative microenvironment of the porous gradient polycaprolactone scaffold to accelerate bone remodeling. *Small.* 2023;19(19):e2206960. doi:10.1002/smll.202206960
125. Bedhiafi T, Idoudi S, Alhams AA, et al. Applications of polydopaminic nanomaterials in mucosal drug delivery. *J Cont Rel.* 2023;353:842–849. doi:10.1016/j.jconrel.2022.12.037
126. Wang S, Wu Z, Wang Y, et al. A homogeneous dopamine-silver nanocomposite coating: striking a balance between the antibacterial ability and cytocompatibility of dental implants. *Regen Biomater.* 2023;10:rbac082. doi:10.1093/rb/rbac082
127. Xing L, Song H, Wei J, et al. Influence of a composite polylysine-polydopamine-quaternary ammonium salt coating on titanium on its osteogenic and antibacterial performance. *Molecules.* 2023;28:10. doi:10.3390/molecules28104120
128. Jin X, Xie D, Zhang Z, et al. In vitro and in vivo studies on biodegradable Zn porous scaffolds with a drug-loaded coating for the treatment of infected bone defect. *Mater Today Bio.* 2024;24:100885. doi:10.1016/j.mtbio.2023.100885
129. Du J, Zhou Y, Bao X, et al. Surface polydopamine modification of bone defect repair materials: characteristics and applications. *Front Bioeng Biotechnol.* 2022;10:974533. doi:10.3389/fbioe.2022.974533
130. Feng P, Liu M, Peng S, Bin S, Zhao Z, Shuai C. Polydopamine modified polycaprolactone powder for fabrication bone scaffold owing intrinsic bioactivity. *J Mater Res Technol.* 2021;15:3375–3385. doi:10.1016/j.jmrt.2021.09.137
131. Davidsen MB, Teixeira JFL, Dehli J, et al. Post-treatments of polydopamine coatings influence cellular response. *Colloids Surf B.* 2021;207:111972. doi:10.1016/j.colsurfb.2021.111972
132. Li L, Yang L, Liao Y, et al. Superhydrophilic versus normal polydopamine coating: a superior and robust platform for synergistic antibacterial and antithrombotic properties. *Chem Eng J.* 2020;402:126196. doi:10.1016/j.cej.2020.126196
133. Yazdi MK, Zare M, Khodadadi A, et al. Polydopamine biomaterials for skin regeneration. *ACS Biomater Sci Eng.* 2022;8(6):2196–2219. doi:10.1021/acsbm.1c01436
134. Li Y, Yang J, Chen X, et al. Mitochondrial-targeting and NIR-responsive Mn(3)O(4)@PDA@Pd-SS31 nanozymes reduce oxidative stress and reverse mitochondrial dysfunction to alleviate osteoarthritis. *Biomaterials.* 2024;305:122449. doi:10.1016/j.biomaterials.2023.122449
135. Musilkova J, Kotelnikov I, Novotna K, et al. Cell adhesion and growth enabled by biomimetic oligopeptide modification of a polydopamine-poly(ethylene oxide) protein repulsive surface. *J Mater Sci Mater Med.* 2015;26(11):253. doi:10.1007/s10856-015-5583-3
136. Yang K, Lee JS, Kim J, et al. Polydopamine-mediated surface modification of scaffold materials for human neural stem cell engineering. *Biomaterials.* 2012;33(29):6952–6964. doi:10.1016/j.biomaterials.2012.06.067
137. Ge L, Li Q, Huang Y, et al. Polydopamine-coated paper-stack nanofibrous membranes enhancing adipose stem cells' adhesion and osteogenic differentiation. *J Mat Chem B.* 2014;2(40):6917–6923. doi:10.1039/c4tb00570h
138. Lin CC, Fu SJ. Osteogenesis of human adipose-derived stem cells on poly(dopamine)-coated electrospun poly(lactic acid) fiber mats. *Mater Sci Eng C Mater Biol Appl.* 2016;58:254–263. doi:10.1016/j.msec.2015.08.009

139. Godoy-Gallardo M, Portolés-Gil N, López-Periago AM, Domingo C, Hosta-Rigau L. Immobilization of BMP-2 and VEGF within multilayered polydopamine-coated scaffolds and the resulting osteogenic and angiogenic synergy of co-cultured human mesenchymal stem cells and human endothelial progenitor cells. *Int J Mol Sci.* 2020;21(17). doi:10.3390/ijms21176418
140. Kaushik N, Nhat Nguyen L, Kim JH, Choi EH, Kumar kaushik N. Strategies for using polydopamine to induce biomineralization of hydroxyapatite on implant materials for bone tissue engineering. *Int J Mol Sci.* 2020;21(18). doi:10.3390/ijms21186544
141. Zhou P, Wu F, Zhou T, et al. Simple and versatile synthetic polydopamine-based surface supports reprogramming of human somatic cells and long-term self-renewal of human pluripotent stem cells under defined conditions. *Biomaterials.* 2016;87:1–17. doi:10.1016/j.biomaterials.2016.02.012
142. Bao X, Zhao J, Sun J, Hu M, Yang X. Polydopamine nanoparticles as efficient scavengers for reactive oxygen species in periodontal disease. *ACS nano.* 2018;12(9):8882–8892. doi:10.1021/acsnano.8b04022
143. Abdallah HM, Farag MA, Algandaby MM, et al. Osteoprotective Activity and metabolite fingerprint via UPLC/MS and GC/MS of lepidium sativum in ovariectomized rats. *Nutrients.* 2020;12(7). doi:10.3390/nu12072075
144. Chai H, Sang S, Luo Y, He R, Yuan X, Zhang X. Icaritin-loaded sulfonated polyetheretherketone with osteogenesis promotion and osteoclastogenesis inhibition properties via immunomodulation for advanced osseointegration. *J Mat Chem B.* 2022;10(18):3531–3540. doi:10.1039/d1tb02802b
145. Chakka JL, Acri T, Laird NZ, et al. Polydopamine functionalized VEGF gene-activated 3D printed scaffolds for bone regeneration. *RSC Adv.* 2021;11(22):13282–13291. doi:10.1039/d1ra01193f
146. Chen D, Yu C, Ying Y, et al. Study of the osteoimmunomodulatory properties of curcumin-modified copper-bearing titanium. *Molecules.* 2022;27:10. doi:10.3390/molecules27103205
147. Chen L, Wang B, Ren H, et al. Arg-Gly-Asp peptide functionalized poly-amino acid/ poly (p-benzamide) copolymer with enhanced mechanical properties and osteogenicity. *Biomater Adv.* 2022;133:112627. doi:10.1016/j.msec.2021.112627
148. Chen T, Zou Q, Du C, Wang C, Li Y, Fu B. Biodegradable 3D printed HA/CMCS/PDA scaffold for repairing lacunar bone defect. *Mater Sci Eng C Mater Biol Appl.* 2020;116:111148. doi:10.1016/j.msec.2020.111148
149. Chen X, Zhu L, Liu H, et al. Biomineralization guided by polydopamine-modified poly(L-lactide) fibrous membrane for promoted osteoconductive activity. *Biomater.* 2019;14(5):055005. doi:10.1088/1748-605X/ab2f2d
150. Cheng CH, Chen YW, Kai-Xing Lee A, Yao CH, Shie MY. Development of mussel-inspired 3D-printed poly (lactic acid) scaffold grafted with bone morphogenetic protein-2 for stimulating osteogenesis. *J Mater Sci Mater Med.* 2019;30(7):78. doi:10.1007/s10856-019-6279-x
151. Cheng CH, Shie MY, Lai YH, Foo NP, Lee MJ, Yao CH. Fabrication of 3D Printed Poly(lactic acid)/Polycaprolactone Scaffolds Using TGF- β 1 for promoting bone regeneration. *Polymers.* 2021;13(21). doi:10.3390/polym13213731
152. Das EC, Dhawan S, Babu J, et al. Self-assembling polymeric dendritic peptide as functional osteogenic matrix for periodontal regeneration scaffolds-an in vitro study. *J Periodontol Res.* 2019;54(5):468–480. doi:10.1111/jre.12647
153. Dashtimoghdam E, Fahimipour F, Tongas N, Tayebi L. Microfluidic fabrication of microcarriers with sequential delivery of VEGF and BMP-2 for bone regeneration. *Sci Rep.* 2020;10(1):11764. doi:10.1038/s41598-020-68221-w
154. Deng Y, Shi J, Chan YK, et al. Heterostructured metal-organic frameworks/polydopamine coating endows polyetheretherketone implants with multimodal osteogenicity and photoswitchable disinfection. *Adv Healthc Mater.* 2022;11(14):e2200641. doi:10.1002/adhm.202200641
155. Deng Y, Sun Y, Bai Y, et al. In vitro biocompatibility/osteogenesis and in vivo bone formation evaluation of peptide-decorated apatite nanocomposites assisted via polydopamine. *J Biom Nanotechnol.* 2016;12(4):602–618. doi:10.1166/jbn.2016.2096
156. Dimassi S, Tabary N, Chai F, et al. Polydopamine treatment of chitosan nanofibers for the conception of osteoinductive scaffolds for bone reconstruction. *Carbohydr Polym.* 2022;276:118774. doi:10.1016/j.carbpol.2021.118774
157. Douglas TE, Wlodarczyk M, Pamula E, et al. Enzymatic mineralization of gellan gum hydrogel for bone tissue-engineering applications and its enhancement by polydopamine. *J Tissue Eng Regen Med.* 2014;8(11):906–918. doi:10.1002/term.1616
158. Du T, Zhao S, Dong W, et al. Surface modification of carbon fiber-reinforced polyetheretherketone with MXene nanosheets for enhanced photothermal antibacterial activity and osteogenicity. *ACS Biomater. Sci. Eng.* 2022;8(6):2375–2389. doi:10.1021/acsbmaterials.2c00095
159. Duan L, Zuo J, Zhang F, et al. Magnetic Targeting of HU-MSCs in the Treatment of glucocorticoid-associated osteonecrosis of the femoral head through Akt/Bcl2/Bad/caspase-3 pathway. *Int j Nanomed.* 2020;15:3605–3620. doi:10.2147/ijn.S244453
160. Fardjahromi MA, Ejeian F, Razmjou A, et al. Enhancing osteoregenerative potential of biphasic calcium phosphates by using bioinspired ZIF8 coating. *Mater Sci Eng C Mater Biol Appl.* 2021;123:111972. doi:10.1016/j.msec.2021.111972
161. Fu C, Jiang Y, Yang X, Wang Y, Ji W, Jia G. Mussel-inspired gold nanoparticle and PLGA/L-lysine-g-graphene oxide composite scaffolds for bone defect repair. *Int j Nanomed.* 2021;16:6693–6718. doi:10.2147/ijn.S328390
162. Gao T, Zhang N, Wang Z, et al. Biodegradable Microcarriers of Poly(Lactide-co-Glycolide) and nano-hydroxyapatite decorated with IGF-1 via polydopamine coating for enhancing cell proliferation and osteogenic differentiation. *Macromol Biosci.* 2015;15(8):1070–1080. doi:10.1002/mabi.201500069
163. Gao X, Song J, Ji P, et al. Polydopamine-Templated Hydroxyapatite Reinforced Polycaprolactone Composite Nanofibers with Enhanced Cytocompatibility and Osteogenesis for Bone Tissue Engineering. *ACS Appl Mater Interfaces.* 2016;8(5):3499–3515. doi:10.1021/acscami.5b12413
164. Gao X, Zhang X, Song J, et al. Osteoinductive peptide-functionalized nanofibers with highly ordered structure as biomimetic scaffolds for bone tissue engineering. *Int j Nanomed.* 2015;10:7109–7128. doi:10.2147/ijn.S94045
165. Gao Y, Yuan Z, Yuan X, et al. Bioinspired porous microspheres for sustained hypoxic exosomes release and vascularized bone regeneration. *Bioact. Mater.* 2022;14:377–388. doi:10.1016/j.bioactmat.2022.01.041
166. Ge L, Liu L, Wei H, et al. Preparation of a small intestinal submucosa modified polypropylene hybrid mesh via a mussel-inspired polydopamine coating for pelvic reconstruction. *J Biomater Appl.* 2016;30(9):1385–1391. doi:10.1177/0885328216628469
167. Ghorai SK, Dutta A, Roy T, et al. Metal ion augmented mussel inspired polydopamine immobilized 3D printed osteoconductive scaffolds for accelerated bone tissue regeneration. *ACS Appl Mater Interfaces.* 2022;14(25):28455–28475. doi:10.1021/acscami.2c01657
168. Ghorbani F, Ghalandari B, Khan AL, Li D, Zamanian A, Yu B. Decoration of electrical conductive polyurethane-polyaniline/polyvinyl alcohol matrixes with mussel-inspired polydopamine for bone tissue engineering. *Biotechnol Prog.* 2020;36(6):e3043. doi:10.1002/btpr.3043

169. Ghorbani F, Kim M, Monavari M, Ghalandari B, Boccaccini AR. Mussel-inspired polydopamine decorated alginate dialdehyde-gelatin 3D printed scaffolds for bone tissue engineering application. *Front Bioeng Biotechnol.* 2022;10:940070. doi:10.3389/fbioe.2022.940070
170. Ghorbani F, Zamanian A, Sahranavard M. Mussel-inspired polydopamine-mediated surface modification of freeze-cast poly (ϵ -caprolactone) scaffolds for bone tissue engineering applications. *Biomed Tech.* 2020;65(3):273–287. doi:10.1515/bmt-2019-0061
171. Han L, Jiang Y, Lv C, et al. Mussel-inspired hybrid coating functionalized porous hydroxyapatite scaffolds for bone tissue regeneration. *Colloids Surf B.* 2019;179:470–478. doi:10.1016/j.colsurfb.2019.04.024
172. Han L, Sun H, Tang P, et al. Mussel-inspired graphene oxide nanosheet-enwrapped Ti scaffolds with drug-encapsulated gelatin microspheres for bone regeneration. *Biomater Sci.* 2018;6(3):538–549. doi:10.1039/c7bm01060e
173. He F, Li J, Wang Y, et al. Design of cefotaxime sodium-loaded polydopamine coatings with controlled surface roughness for titanium implants. *ACS Biomater. Sci. Eng.* 2022;8(11):4751–4763. doi:10.1021/acsbmaterials.2c00702
174. Huang B, Chen M, Tian J, et al. Oxygen-carrying and antibacterial fluorinated nano-hydroxyapatite incorporated hydrogels for enhanced bone regeneration. *Adv Healthc Mater.* 2022;11(12):e2102540. doi:10.1002/adhm.202102540
175. Huang J, Lu J, Liu Z, et al. Covalent immobilization of VEGF on allogeneic bone through polydopamine coating to improve bone regeneration. *Front Bioeng Biotechnol.* 2022;10:1003677. doi:10.3389/fbioe.2022.1003677
176. Huang S, Liang N, Hu Y, Zhou X, Abidi N. Polydopamine-assisted surface modification for bone biosubstitutes. *Biomed Res Int.* 2016;2016:2389895. doi:10.1155/2016/2389895
177. Huang Y, Du Z, Zheng T, et al. Antibacterial, conductive, and osteocompatible polyorganophosphazene microscaffolds for the repair of infectious calvarial defect. *J Biomed Mater Res A.* 2021;109(12):2580–2596. doi:10.1002/jbm.a.37252
178. Huang Z, Wu Z, Ma B, et al. Enhanced in vitro biocompatibility and osteogenesis of titanium substrates immobilized with dopamine-assisted superparamagnetic Fe(3)O(4) nanoparticles for hBMSCs. *R Soc Open Sci.* 2018;5(8):172033. doi:10.1098/rsos.172033
179. Ji S, Dube K, Chesterman JP, et al. Polyester-based ink platform with tunable bioactivity for 3D printing of tissue engineering scaffolds. *Biomater Sci.* 2019;7(2):560–570. doi:10.1039/c8bm01269e
180. Jiang BB, Li SH, Zheng W. Preparation of Antibiotic-loaded copper-doped hydroxyapatite microspheres and evaluation of their antibacterial and osteogenic effect. *Sichuan Da Xue Xue Bao Yi Xue Ban.* 2021;52(5):799–806. doi:10.12182/20210960209
181. Kao CT, Chen YJ, Ng HY, et al. Surface modification of calcium silicate via mussel-inspired polydopamine and effective adsorption of extracellular matrix to promote osteogenesis differentiation for bone tissue engineering. *Materials.* 2018;11(9). doi:10.3390/ma11091664
182. Kao CT, Lin CC, Chen YW, Yeh CH, Fang HY, Shie MY. Poly(dopamine) coating of 3D printed poly(lactic acid) scaffolds for bone tissue engineering. *Mater Sci Eng C Mater Biol Appl.* 2015;56:165–173. doi:10.1016/j.msec.2015.06.028
183. Ko E, Yang K, Shin J, Cho SW. Polydopamine-assisted osteoinductive peptide immobilization of polymer scaffolds for enhanced bone regeneration by human adipose-derived stem cells. *Biomacromolecules.* 2013;14(9):3202–3213. doi:10.1021/bm4008343
184. Kong L, Han Y, Lu Q, et al. Polydopamine coating with static magnetic field promotes the osteogenic differentiation of human bone-derived mesenchymal stem cells on three-dimensional printed porous titanium scaffolds by upregulation of the BMP-Smads signaling pathway. *Am J Transl Res.* 2020;12(12):7812–7825.
185. Kwack KH, Ji JY, Park B, Heo JS. Fucoidan (Undaria pinnatifida)/polydopamine composite-modified surface promotes osteogenic potential of periodontal ligament stem cells. *Mar Drugs.* 2022;20(3). doi:10.3390/md20030181
186. Lee DJ, Lee YT, Zou R, Daniel R, Ko CC. Polydopamine-laced biomimetic material stimulation of bone marrow derived mesenchymal stem cells to promote osteogenic effects. *Sci Rep.* 2017;7(1):12984. doi:10.1038/s41598-017-13326-y
187. Lee DJ, Tseng HC, Wong SW, Wang Z, Deng M, Ko CC. Dopaminergic effects on in vitro osteogenesis. *Bone Res.* 2015;3:15020. doi:10.1038/boneres.2015.20
188. Lee JS, Jin Y, Park HJ, et al. In situ bone tissue engineering with an endogenous stem cell mobilizer and osteoinductive nanofibrous polymeric scaffolds. *Biotechnol J.* 2017;12(12). doi:10.1002/biot.201700062
189. Lee JS, Lee JC, Heo JS. Polydopamine-assisted BMP-2 immobilization on titanium surface enhances the osteogenic potential of periodontal ligament stem cells via integrin-mediated cell-matrix adhesion. *J Cell Commun Signal.* 2018;12(4):661–672. doi:10.1007/s12079-018-0468-0
190. Lee JS, Yi JK, An SY, Heo JS. Increased osteogenic differentiation of periodontal ligament stem cells on polydopamine film occurs via activation of integrin and PI3K signaling pathways. *Cell Physiol Biochem.* 2014;34(5):1824–1834. doi:10.1159/000366381
191. Lee SJ, Lee D, Yoon TR, et al. Surface modification of 3D-printed porous scaffolds via mussel-inspired polydopamine and effective immobilization of rhBMP-2 to promote osteogenic differentiation for bone tissue engineering. *Acta Biomater.* 2016;40:182–191. doi:10.1016/j.actbio.2016.02.006
192. Lee SJ, Lee HJ, Kim SY, et al. In situ gold nanoparticle growth on polydopamine-coated 3D-printed scaffolds improves osteogenic differentiation for bone tissue engineering applications: in vitro and in vivo studies. *Nanoscale.* 2018;10(33):15447–15453. doi:10.1039/c8nr04037k
193. Li B, Liu F, Ye J, et al. Regulation of macrophage polarization through periodic photo-thermal treatment to facilitate osteogenesis. *Small.* 2022;18(38):e2202691. doi:10.1002/sml.202202691
194. Li H, Chen S, Chen J, et al. Mussel-inspired artificial grafts for functional ligament reconstruction. *ACS Appl Mater Interfaces.* 2015;7(27):14708–14719. doi:10.1021/acsami.5b05109
195. Li H, Wang H, Pan J, et al. Nanoscaled bionic periosteum orchestrating the osteogenic microenvironment for sequential bone regeneration. *ACS Appl Mater Interfaces.* 2020;12(33):36823–36836. doi:10.1021/acsami.0c06906
196. Li H, Wang X, Shen Y, Tang H, Tang X, Zhang Y. Chondrogenic and ameliorated inflammatory effects of chitosan-based biomimetic scaffold loaded with icariin. *Sheng Wu Gong Cheng Xue Bao.* 2022;38(6):2308–2321. doi:10.13345/j.cjb.210838
197. Li H, Zheng L, Wang M. Biofunctionalized nanofibrous bilayer scaffolds for enhancing cell adhesion, proliferation and osteogenesis. *ACS Appl Bio Mater.* 2021;4(6):5276–5294. doi:10.1021/acsabm.1c00414
198. Li J, Yao Q, Xu Y, Zhang H, Li LL, Wang L. Lithium chloride-releasing 3D printed scaffold for enhanced cartilage regeneration. *Med Sci Monit.* 2019;25:4041–4050. doi:10.12659/msm.916918
199. Li L, Li Y, Yang L, et al. Polydopamine coating promotes early osteogenesis in 3D printing porous Ti6Al4V scaffolds. *Ann Transl Med.* 2019;7(11):240. doi:10.21037/atm.2019.04.79

200. Li M, Liu X, Xu Z, Yeung KW, Wu S. Dopamine modified organic-inorganic hybrid coating for antimicrobial and osteogenesis. *ACS Appl Mater Interfaces*. 2016;8(49):33972–33981. doi:10.1021/acsami.6b09457
201. Li N, Liu L, Wei C, et al. Immunomodulatory blood-derived hybrid hydrogels as multichannel microenvironment modulators for augmented bone regeneration. *ACS Appl Mater Interfaces*. 2022;14(48):53523–53534. doi:10.1021/acsami.2c16774
202. Li W, Liu Y, Zhang P, et al. Tissue-engineered bone immobilized with human adipose stem cells-derived exosomes promotes bone regeneration. *ACS Appl Mater Interfaces*. 2018;10(6):5240–5254. doi:10.1021/acsami.7b17620
203. Li Y, Shi Y, Duan S, et al. Electrospun biodegradable polyorganophosphazene fibrous matrix with poly(dopamine) coating for bone regeneration. *J Biomed Mater Res A*. 2014;102(11):3894–3902. doi:10.1002/jbm.a.35065
204. Li Y, Yang W, Li X, et al. Improving osteointegration and osteogenesis of three-dimensional porous Ti6Al4V scaffolds by polydopamine-assisted biomimetic hydroxyapatite coating. *ACS Appl Mater Interfaces*. 2015;7(10):5715–5724. doi:10.1021/acsami.5b00331
205. Lin H, Shi S, Lan X, et al. Scaffold 3D-printed from metallic nanoparticles-containing ink simultaneously eradicates tumor and repairs tumor-associated bone defects. *Small Methods*. 2021;5(9):e2100536. doi:10.1002/smt.202100536
206. Lin Y, Zhang L, Liu NQ, et al. In vitro behavior of tendon stem/progenitor cells on bioactive electrospun nanofiber membranes for tendon-bone tissue engineering applications. *Int J Nanomed*. 2019;14:5831–5848. doi:10.2147/ijn.S210509
207. Liu C, Wu J, Gan D, et al. The characteristics of mussel-inspired nHA/OSA injectable hydrogel and repaired bone defect in rabbit. *J Biomed Mater Res B Appl Biomater*. 2020;108(5):1814–1825. doi:10.1002/jbm.b.34524
208. Liu F, Cheng X, Xiao L, et al. Inside-outside Ag nanoparticles-loaded polylactic acid electrospun fiber for long-term antibacterial and bone regeneration. *Int J Biol Macromol*. 2021;167:1338–1348. doi:10.1016/j.ijbiomac.2020.11.088
209. Liu H, Li W, Luo B, Chen X, Wen W, Zhou C. Icarin immobilized electrospinning poly(L-lactide) fibrous membranes via polydopamine adhesive coating with enhanced cytocompatibility and osteogenic activity. *Mater Sci Eng C Mater Biol Appl*. 2017;79:399–409. doi:10.1016/j.msec.2017.05.077
210. Liu H, Wen W, Chen S, Zhou C, Luo B. Preparation of icaritin and deferoxamine functionalized Poly(L-lactide)/chitosan Micro/nanofibrous membranes with synergistic enhanced osteogenesis and angiogenesis. *ACS Appl Bio Mater*. 2018;1(2):389–402. doi:10.1021/acsabm.8b00129
211. Liu M, Zhou J, Yang Y, Zheng M, Yang J, Tan J. Surface modification of zirconia with polydopamine to enhance fibroblast response and decrease bacterial activity in vitro: a potential technique for soft tissue engineering applications. *Colloids Surf B*. 2015;136:74–83. doi:10.1016/j.colsurfb.2015.06.047
212. Liu T, Li B, Chen G, Ye X, Zhang Y. Nano tantalum-coated 3D printed porous polylactic acid/beta-tricalcium phosphate scaffolds with enhanced biological properties for guided bone regeneration. *Int J Biol Macromol*. 2022;221:371–380. doi:10.1016/j.ijbiomac.2022.09.003
213. Liu W, Zhu L, Ma Y, et al. Well-ordered chitin whiskers layer with high stability on the surface of poly(D,L-lactide) film for enhancing mechanical and osteogenic properties. *Carbohydr Polym*. 2019;212:277–288. doi:10.1016/j.carbpol.2019.02.060
214. Liu X, Chen W, Shao B, et al. Mussel patterned with 4D biodegrading elastomer durably recruits regenerative macrophages to promote regeneration of craniofacial bone. *Biomaterials*. 2021;276:120998. doi:10.1016/j.biomaterials.2021.120998
215. Lu L, Wang H, Yang M, Wang L, Gan K. Three-dimensional-printed MPBI@ β -TCP scaffold promotes bone regeneration and impedes osteosarcoma under near-infrared laser irradiation. *FASEB J*. 2023;37(5):e22924. doi:10.1096/fj.202201991R
216. Lu Y, Wan Y, Gan D, et al. Enwrapping polydopamine on doxorubicin-loaded lamellar hydroxyapatite/Poly(lactic-co-glycolic acid) composite fibers for inhibiting bone tumor recurrence and enhancing bone regeneration. *ACS Appl Bio Mater*. 2021;4(8):6036–6045. doi:10.1021/acsabm.1c00297
217. Luo S, Wu J, Jia Z, et al. An injectable, bifunctional hydrogel with photothermal effects for tumor therapy and bone regeneration. *Macromol biosci*. 2019;19(9):e1900047. doi:10.1002/mabi.201900047
218. Ma H, Han H, Zhao X, et al. Engineering multifunctional polyether ether ketone implant: mechanics-adaptability, biomineralization, immunoregulation, anti-infection, osteointegration, and osteogenesis. *Adv Healthc Mater*. 2023;12(12):e2202799. doi:10.1002/adhm.202202799
219. Ma L, Cheng S, Ji X, et al. Immobilizing magnesium ions on 3D printed porous tantalum scaffolds with polydopamine for improved vascularization and osteogenesis. *Mater Sci Eng C Mater Biol Appl*. 2020;117:111303. doi:10.1016/j.msec.2020.111303
220. Ma L, Li G, Lei J, et al. Nanotopography sequentially mediates human mesenchymal stem cell-derived small extracellular vesicles for enhancing osteogenesis. *ACS nano*. 2022;16(1):415–430. doi:10.1021/acsnano.1c07150
221. Ma T, Wang CX, Ge XY, Zhang Y. Applications of polydopamine in implant surface modification. *Macromol biosci*. 2023;23(10):e2300067. doi:10.1002/mabi.202300067
222. Mahnavi A, Shahriari-Khalaji M, Hosseinpour B, et al. Evaluation of cell adhesion and osteoconductivity in bone substitutes modified by polydopamine. *Front Bioeng Biotechnol*. 2022;10:1057699. doi:10.3389/fbioe.2022.1057699
223. Meng X, Zhang J, Chen J, et al. KR-12 coating of polyetheretherketone (PEEK) surface via polydopamine improves osteointegration and antibacterial activity in vivo. *J Mat Chem B*. 2020;8(44):10190–10204. doi:10.1039/d0tb01899f
224. Moazami S, Kharaziha M, Emadi R, Dinari M. Multifunctional bioinspired bredigite-modified adhesive for bone fracture healing. *ACS Appl Mater Interfaces*. 2023;15(5):6499–6513. doi:10.1021/acsami.2c20038
225. Ou Q, Zhang S, Fu C, et al. More natural more better: triple natural anti-oxidant puerarin/ferulic acid/polydopamine incorporated hydrogel for wound healing. *J Nanobiotechnology*. 2021;19(1):237. doi:10.1186/s12951-021-00973-7
226. Pan H, Zheng Q, Guo X, Wu Y, Wu B. Polydopamine-assisted BMP-2-derived peptides immobilization on biomimetic copolymer scaffold for enhanced bone induction in vitro and in vivo. *Colloids Surf B*. 2016;142:1–9. doi:10.1016/j.colsurfb.2016.01.060
227. Park J, Lee SJ, Jung TG, et al. Surface modification of a three-dimensional polycaprolactone scaffold by polydopamine, biomineralization, and BMP-2 immobilization for potential bone tissue applications. *Colloids Surf B*. 2021;199:111528. doi:10.1016/j.colsurfb.2020.111528
228. Peng F, Cheng S, Zhang R, et al. Zn-contained mussel-inspired film on Mg alloy for inhibiting bacterial infection and promoting bone regeneration. *Regen Biomater*. 2021;8(1):rbaa044. doi:10.1093/rb/rbaa044
229. Qi J, Wang Y, Chen L, et al. 3D-printed porous functional composite scaffolds with polydopamine decoration for bone regeneration. *Regen Biomater*. 2023;10:rbad062. doi:10.1093/rb/rbad062
230. Qin S, Lu Z, Gan K, et al. Construction of a BMP-2 gene delivery system for polyetheretherketone bone implant material and its effect on bone formation in vitro. *J Biomed Mater Res B Appl Biomater*. 2022;110(9):2075–2088. doi:10.1002/jbm.b.35062

231. Ren S, Tang X, Liu L, et al. Reinforced blood-derived protein hydrogels enable dual-level regulation of bio-physiochemical microenvironments for personalized bone regeneration with remarkable enhanced efficacy. *Nano Lett.* 2022;22(10):3904–3913. doi:10.1021/acs.nanolett.2c00057
232. Rezaei H, Shahrezaei M, Jalali Monfared M, Ghorbani F, Zamanian A, Sahebzamani M. Mussel-inspired polydopamine induced the osteoinductivity to ice-templating PLGA-gelatin matrix for bone tissue engineering application. *Biotechnol Appl Biochem.* 2021;68(1):185–196. doi:10.1002/bab.1911
233. Shim NY, Heo JS. Performance of the polydopamine-graphene oxide composite substrate in the osteogenic differentiation of mouse embryonic stem cells. *Int J Mol Sci.* 2021;22(14). doi:10.3390/ijms22147323
234. Si Y, Liu H, Li M, Jiang X, Yu H, Sun D. An efficient metal-organic framework-based drug delivery platform for synergistic antibacterial activity and osteogenesis. *J Colloid Interface Sci.* 2023;640:521–539. doi:10.1016/j.jcis.2023.02.149
235. Si Y, Liu H, Yu H, Jiang X, Sun D. MOF-derived CuO@ZnO modified titanium implant for synergistic antibacterial ability, osteogenesis and angiogenesis. *Colloids Surf B.* 2022;219:112840. doi:10.1016/j.colsurfb.2022.112840
236. Sun H, Dong J, Wang Y, et al. Polydopamine-Coated Poly(L-lactide) Nanofibers with Controlled Release of VEGF and BMP-2 as a regenerative periosteum. *ACS Biomater. Sci. Eng.* 2021;7(10):4883–4897. doi:10.1021/acsbmaterials.1c00246
237. Sun X, Jiao X, Wang Z, et al. Polydopamine-coated 3D-printed β -tricalcium phosphate scaffolds to promote the adhesion and osteogenesis of BMSCs for bone-defect repair: mRNA transcriptomic sequencing analysis. *J Mat Chem B.* 2023;11(8):1725–1738. doi:10.1039/d2tb02280j
238. Sun Y, Deng Y, Ye Z, Liang S, Tang Z, Wei S. Peptide decorated nano-hydroxyapatite with enhanced bioactivity and osteogenic differentiation via polydopamine coating. *Colloids Surf B.* 2013;111:107–116. doi:10.1016/j.colsurfb.2013.05.037
239. Sun Y, Li Y, Zhang Y, Wang T, Lin K, Liu J. A polydopamine-assisted strontium-substituted apatite coating for titanium promotes osteogenesis and angiogenesis via FAK/MAPK and PI3K/AKT signaling pathways. *Mater Sci Eng C Mater Biol Appl.* 2021;131:112482. doi:10.1016/j.msec.2021.112482
240. Suttiat K, Wattanuchariya W, Manaspon C. Preparation and Characterization of Porous Poly(Lactic Acid)/Poly(Butylene Adipate-Co-Terephthalate) (PLA/PBAT) scaffold with polydopamine-assisted biomineralization for bone regeneration. *Materials.* 2022;15(21). doi:10.3390/ma15217756
241. Wang B, Lan J, Qiao H, et al. Porous surface with fusion peptides embedded in strontium titanate nanotubes elevates osteogenic and antibacterial activity of additively manufactured titanium alloy. *Colloids Surf B.* 2023;224:113188. doi:10.1016/j.colsurfb.2023.113188
242. Wang B, Li Y, Wang S, et al. Electrodeposited dopamine/strontium-doped hydroxyapatite composite coating on pure zinc for anti-corrosion, antimicrobial and osteogenesis. *Mater Sci Eng C Mater Biol Appl.* 2021;129:112387. doi:10.1016/j.msec.2021.112387
243. Wang DX, He Y, Bi L, et al. Enhancing the bioactivity of Poly(lactic-co-glycolic acid) scaffold with a nano-hydroxyapatite coating for the treatment of segmental bone defect in a rabbit model. *Int j Nanomed.* 2013;8:1855–1865. doi:10.2147/ijn.S43706
244. Wang H, Lin C, Zhang X, Lin K, Wang X, Shen SG. Mussel-inspired polydopamine coating: a general strategy to enhance osteogenic differentiation and osseointegration for diverse implants. *ACS Appl Mater Interfaces.* 2019;11(7):7615–7625. doi:10.1021/acsami.8b21558
245. Wang L, Shang X, Hao Y, et al. Bi-functional titanium-polydopamine-zinc coatings for infection inhibition and enhanced osseointegration. *RSC Adv.* 2019;9(6):2892–2905. doi:10.1039/c8ra09112a
246. Wang M, Deng Y, Zhou P, et al. In vitro culture and directed osteogenic differentiation of human pluripotent stem cells on peptides-decorated two-dimensional microenvironment. *ACS Appl Mater Interfaces.* 2015;7(8):4560–4572. doi:10.1021/acsami.5b00188
247. Wang P, Yin HM, Li X, et al. Simultaneously constructing nanotopographical and chemical cues in 3D-printed polylactic acid scaffolds to promote bone regeneration. *Mater Sci Eng C Mater Biol Appl.* 2021;118:111457. doi:10.1016/j.msec.2020.111457
248. Wang S, Duan C, Yang W, et al. Two-dimensional nanocoating-enabled orthopedic implants for bimodal therapeutic applications. *Nanoscale.* 2020;12(22):11936–11946. doi:10.1039/d0nr02327b
249. Wang X, Ao J, Lu H, et al. Osteoimmune modulation and guided osteogenesis promoted by barrier membranes incorporated with S-xNitrosoglutathione (GSNO) and mesenchymal stem cell-derived exosomes. *Int j Nanomed.* 2020;15:3483–3496. doi:10.2147/ijn.S248741
250. Wang Z, Chen T, Wu Z, et al. The dual-effects of PLGA@MT electrospun nanofiber coatings on promoting osteogenesis at the titanium-bone interface under diabetic conditions. *J Mat Chem B.* 2022;10(21):4020–4030. doi:10.1039/d2tb00120a
251. Wang Z, Mei L, Liu X, Zhou Q. Hierarchically hybrid biocoatings on Ti implants for enhanced antibacterial activity and osteogenesis. *Colloids Surf B.* 2021;204:111802. doi:10.1016/j.colsurfb.2021.111802
252. Wei X, Zhou W, Tang Z, et al. Magnesium surface-activated 3D printed porous PEEK scaffolds for in vivo osseointegration by promoting angiogenesis and osteogenesis. *Bioact Mater.* 2023;20:16–28. doi:10.1016/j.bioactmat.2022.05.011
253. Wu H, Zhao C, Lin K, Wang X. Mussel-inspired polydopamine-based multilayered coatings for enhanced bone formation. *Front Bioeng Biotechnol.* 2022;10:952500. doi:10.3389/fbioe.2022.952500
254. Wu J, Cao L, Liu Y, et al. Functionalization of silk fibroin electrospun scaffolds via BMSC affinity peptide grafting through oxidative self-polymerization of dopamine for bone regeneration. *ACS Appl Mater Interfaces.* 2019;11(9):8878–8895. doi:10.1021/acsami.8b22123
255. Wu M, Chen F, Liu H, et al. Bioinspired sandwich-like hybrid surface functionalized scaffold capable of regulating osteogenesis, angiogenesis, and osteoclastogenesis for robust bone regeneration. *Mater Today Bio.* 2022;17:100458. doi:10.1016/j.mtbio.2022.100458
256. Wu M, Liu H, Zhu Y, et al. Mild photothermal-stimulation based on injectable and photocurable hydrogels orchestrates immunomodulation and osteogenesis for high-performance bone regeneration. *Small.* 2023;19(28):e2300111. doi:10.1002/sml.202300111
257. Wu M, Zhang Y, Wu P, et al. Mussel-inspired multifunctional surface through promoting osteogenesis and inhibiting osteoclastogenesis to facilitate bone regeneration. *NPJ Regen Med.* 2022;7(1):29. doi:10.1038/s41536-022-00224-9
258. Wu P, Shen L, Liu HF, et al. The marriage of immunomodulatory, angiogenic, and osteogenic capabilities in a piezoelectric hydrogel tissue engineering scaffold for military medicine. *Mil Med Res.* 2023;10(1):35. doi:10.1186/s40779-023-00469-5
259. Wu Z, Tian Q, Wang J, et al. A bone implant with NIR-responsiveness for eliminating osteosarcoma cells and promoting osteogenic differentiation of BMSCs. *Colloids Surf B.* 2022;211:112296. doi:10.1016/j.colsurfb.2021.112296
260. Xian M, Fang L, Liu Y, et al. Electrical field induce mBMSCs differentiation to osteoblast via protein adsorption enhancement. *Colloids Surf B.* 2022;209(Pt 2):112158. doi:10.1016/j.colsurfb.2021.112158
261. Xie C, Lu X, Wang K, et al. Pulse electrochemical driven rapid layer-by-layer assembly of polydopamine and hydroxyapatite nanofilms via alternative redox in situ synthesis for bone regeneration. *ACS Biomater. Sci. Eng.* 2016;2(6):920–928. doi:10.1021/acsbmaterials.6b00015

262. Xie K, Zhou Z, Guo Y, et al. Long-term prevention of bacterial infection and enhanced osteoinductivity of a hybrid coating with selective silver toxicity. *Adv Healthc Mater.* 2019;8(5):e1801465. doi:10.1002/adhm.201801465
263. Xu Q, Li Y, Zhu Y, Zhao K, Gu R, Zhu Q. Recombinant human BMP-7 grafted poly(lactide-co-glycolide)/hydroxyapatite scaffolds via polydopamine for enhanced calvarial repair. *RSC Adv.* 2018;8(48):27191–27200. doi:10.1039/c8ra05606d
264. Xu Y, Zhao S, Weng Z, et al. Jelly-inspired injectable guided tissue regeneration strategy with shape auto-matched and dual-light-defined antibacterial/osteogenic pattern switch properties. *ACS Appl Mater Interfaces.* 2020;12(49):54497–54506. doi:10.1021/acsami.0c18070
265. Xu Z, Wang N, Liu P, et al. Poly(Dopamine) Coating on 3D-printed poly-lactic-co-glycolic acid/ β -tricalcium phosphate scaffolds for bone tissue engineering. *Molecules.* 2019;24(23). doi:10.3390/molecules24234397
266. Xue H, Zhang Z, Lin Z, et al. Enhanced tissue regeneration through immunomodulation of angiogenesis and osteogenesis with a multifaceted nanohybrid modified bioactive scaffold. *Bioact. Mater.* 2022;18:552–568. doi:10.1016/j.bioactmat.2022.05.023
267. Xue Y, Niu W, Wang M, Chen M, Guo Y, Lei B. Engineering a biodegradable multifunctional antibacterial bioactive nanosystem for enhancing tumor photothermo-chemotherapy and bone regeneration. *ACS nano.* 2020;14(1):442–453. doi:10.1021/acsnano.9b06145
268. Yang T, Dong Y, Wan J, et al. Sustained Release of BMSC-EVs from 3D Printing Gel/HA/nHAP scaffolds for promoting bone regeneration in diabetic rats. *Adv Healthc Mater.* 2023;12(18):e2203131. doi:10.1002/adhm.202203131
269. Yang X, Wang Q, Yan C, et al. A dual-functional strontium-decorated titanium implants that guides the immune response for osseointegration of osteoporotic rats. *Colloids Surf B.* 2024;233:113643. doi:10.1016/j.colsurfb.2023.113643
270. Yang X, Wang Q, Zhang Y, et al. A dual-functional PEEK implant coating for anti-bacterial and accelerated osseointegration. *Colloids Surf B Biointerf.* 2023;224:113196. doi:10.1016/j.colsurfb.2023.113196
271. Yang Y, Zhang B, Yang Y, Peng B, Ye R. PLGA containing human adipose-derived stem cell-derived extracellular vesicles accelerates the repair of alveolar bone defects via transfer of CGRP. *Oxid Med Cell Longev.* 2022;2022:4815284. doi:10.1155/2022/4815284
272. Yang Z, Xie L, Zhang B, et al. Preparation of BMP-2/PDA-BCP Bioceramic Scaffold by DLP 3D printing and its ability for inducing continuous bone formation. *Front Bioeng Biotechnol.* 2022;10:854693. doi:10.3389/fbioe.2022.854693
273. Yao H, Wang J, Deng Y, Li Z, Wei J. Osteogenic and antibacterial PLLA membrane for bone tissue engineering. *Int J Biol Macromol.* 2023;247:125671. doi:10.1016/j.ijbiomac.2023.125671
274. Yao M, Zou Q, Zou W, et al. Bifunctional scaffolds of hydroxyapatite/poly(dopamine)/carboxymethyl chitosan with osteogenesis and anti-osteosarcoma effect. *Biomater Sci.* 2021;9(9):3319–3333. doi:10.1039/d0bm01785j
275. Yi Q, Liang P, Liang D, et al. Improvement of polydopamine-loaded salidroside on osseointegration of titanium implants. *Chin Med.* 2022;17(1):26. doi:10.1186/s13020-022-00569-9
276. Yu Y, Li X, Li J, Li D, Wang Q, Teng W. Dopamine-assisted co-deposition of hydroxyapatite-functionalised nanoparticles of polydopamine on implant surfaces to promote osteogenesis in environments with high ROS levels. *Mater Sci Eng C Mater Biol Appl.* 2021;131:112473. doi:10.1016/j.msec.2021.112473
277. Yu Y, Sun Y, Zhou X, et al. Ag and peptide co-decorate polyetheretherketone to enhance antibacterial property and osteogenic differentiation. *Colloids Surf B.* 2021;198:111492. doi:10.1016/j.colsurfb.2020.111492
278. Zeng J, Gu C, Geng X, Lin K, Xie Y, Chen X. Combined photothermal and sonodynamic therapy using a 2D black phosphorus nanosheets loaded coating for efficient bacterial inhibition and bone-implant integration. *Biomaterials.* 2023;297:122122. doi:10.1016/j.biomaterials.2023.122122
279. Zeng J, Gu C, Zeng F, Xie Y. 2D silicene nanosheets-loaded coating for combating implant-associated infection. *Int J Biol Macromol.* 2023;253(Pt 8):127585. doi:10.1016/j.ijbiomac.2023.127585
280. Zhang B, Li J, He L, Huang H, Weng J. Bio-surface coated titanium scaffolds with cancellous bone-like biomimetic structure for enhanced bone tissue regeneration. *Acta Biomater.* 2020;114:431–448. doi:10.1016/j.actbio.2020.07.024
281. Zhang D, Deng T, Luo Z, et al. Surface modification of titanium implant with hBMP-2/hIGF-1 for promoting biocompatibility and osteogenesis. *Nan Fang Yi Ke Da Xue Xue Bao.* 2021;41(8):1277–1282. doi:10.12122/j.issn.1673-4254.2021.08.22
282. Zhang J, Cai L, Wang T, et al. Lithium doped silica nanospheres/poly(dopamine) composite coating on polyetheretherketone to stimulate cell responses, improve bone formation and osseointegration. *Nanomedicine.* 2018;14(3):965–976. doi:10.1016/j.nano.2018.01.017
283. Zhang L, Zhang H, Zhou H, et al. A Ti(3)C(2) MXene-integrated near-infrared-responsive multifunctional porous scaffold for infected bone defect repair. *J Mat Chem B.* 2023;12(1):79–96. doi:10.1039/d3tb01578e
284. Zhang M, Zhang J, Ban L, et al. Polydopamine regulated hydroxyapatite microspheres grown in the three-dimensional honeycomb-like mollusk shell-derived organic template for osteogenesis. *Biofabrication.* 2020;12(3):035022. doi:10.1088/1758-5090/ab8f20
285. Zhang R, Jo JI, Kanda R, Nishiura A, Hashimoto Y, Matsumoto N. Bioactive polyetheretherketone with gelatin hydrogel leads to sustained release of bone morphogenetic protein-2 and promotes osteogenic differentiation. *Int J Mol Sci.* 2023;24(16). doi:10.3390/ijms241612741
286. Zhang X, Li J, Chen J, et al. Enhanced bone regeneration via PHA scaffolds coated with polydopamine-captured BMP2. *J Mat Chem B.* 2022;10(32):6214–6227. doi:10.1039/d2tb01122k
287. Zhao X, Han Y, Li J, et al. BMP-2 immobilized PLGA/hydroxyapatite fibrous scaffold via polydopamine stimulates osteoblast growth. *Mater Sci Eng C Mater Biol Appl.* 2017;78:658–666. doi:10.1016/j.msec.2017.03.186
288. Zhao Y, He P, Wang B, Bai J, Xue F, Chu C. Incorporating pH/NIR responsive nanocontainers into a smart self-healing coating for a magnesium alloy with controlled drug release, bacteria killing and osteogenesis properties. *Acta Biomater.* 2024;174:463–481. doi:10.1016/j.actbio.2023.12.004
289. Zhong W, Li J, Hu C, et al. 3D-printed titanium implant-coated polydopamine for repairing femoral condyle defects in rabbits. *J Orthop Surg Res.* 2020;15(1):102. doi:10.1186/s13018-020-01593-x
290. Zhou J, Guo X, Zheng Q, Wu Y, Cui F, Wu B. Improving osteogenesis of three-dimensional porous scaffold based on mineralized recombinant human-like collagen via mussel-inspired polydopamine and effective immobilization of BMP-2-derived peptide. *Colloids Surf B.* 2017;152:124–132. doi:10.1016/j.colsurfb.2016.12.041
291. Zhou Y, Wang G, Wang T, et al. Multidynamic osteogenic differentiation by effective polydopamine micro-arc oxide manipulations. *Int j Nanomed.* 2022;17:4773–4790. doi:10.2147/ijn.S378387

292. Zhu Y, Cao Z, Peng Y, Hu L, Guney T, Tang B. Facile surface modification method for synergistically enhancing the biocompatibility and bioactivity of poly(ether ether ketone) that induced osteodifferentiation. *ACS Appl Mater Interfaces*. 2019;11(31):27503–27511. doi:10.1021/acsami.9b03030
293. Ai L, Wang Y, Tao G, et al. Polydopamine-based surface modification of ZnO nanoparticles on sericin/polyvinyl alcohol composite film for antibacterial application. *Molecules*. 2019;24(3). doi:10.3390/molecules24030503
294. Cai R, Tao G, He H, et al. One-step synthesis of silver nanoparticles on polydopamine-coated sericin/polyvinyl alcohol composite films for potential antimicrobial applications. *Molecules*. 2017;22(5). doi:10.3390/molecules22050721
295. Ding X, Zhang Y, Ling J, Lin C. Rapid mussel-inspired synthesis of PDA-Zn-Ag nanofilms on TiO(2) nanotubes for optimizing the antibacterial activity and biocompatibility by doping polydopamine with zinc at a higher temperature. *Colloids Surf B*. 2018;171:101–109. doi:10.1016/j.colsurfb.2018.07.014
296. Fan L, Xie J, Zheng Y, et al. Antibacterial, self-adhesive, recyclable, and tough conductive composite hydrogels for ultrasensitive strain sensing. *ACS Appl Mater Interfaces*. 2020;12(19):22225–22236. doi:10.1021/acsami.0c06091
297. Fan S, Lin W, Huang Y, et al. Advances and potentials of polydopamine nanosystem in photothermal-based antibacterial infection therapies. *Front Pharmacol*. 2022;13:829712. doi:10.3389/fphar.2022.829712
298. Fan XL, Li HY, Ye WY, et al. Magainin-modified polydopamine nanoparticles for photothermal killing of bacteria at low temperature. *Colloids Surf B*. 2019;183:110423. doi:10.1016/j.colsurfb.2019.110423
299. Fang Y, Xing C, Zhan S, et al. Multifunctional magnetic-fluorescent nanoparticle: fabrication, bioimaging, and potential antibacterial applications. *ACS Biomater Sci Eng*. 2019;5(12):6779–6793. doi:10.1021/acsbiomaterials.9b01332
300. Fu Y, Yang L, Zhang J, et al. Polydopamine antibacterial materials. *Mater Horiz*. 2021;8(6):1618–1633. doi:10.1039/d0mh01985b
301. Gao C, Wang Y, Han F, et al. Antibacterial activity and osseointegration of silver-coated poly(ether ether ketone) prepared using the polydopamine-assisted deposition technique. *J Mat Chem B*. 2017;5(47):9326–9336. doi:10.1039/c7tb02436c
302. Guan M, Chen Y, Wei Y, et al. Long-lasting bactericidal activity through selective physical puncture and controlled ions release of polydopamine and silver nanoparticles-loaded TiO(2) nanorods in vitro and in vivo. *Int j Nanomed*. 2019;14:2903–2914. doi:10.2147/ijn.S202625
303. He X, Obeng E, Sun X, Kwon N, Shen J, Yoon J. Polydopamine, harness of the antibacterial potentials-A review. *Mater Today Bio*. 2022;15:100329. doi:10.1016/j.mtbio.2022.100329
304. Li F, Huang K, Chang H, et al. A polydopamine coated nanoscale FeS theranostic platform for the elimination of drug-resistant bacteria via photothermal-enhanced Fenton reaction. *Acta Biomater*. 2022;150:380–390. doi:10.1016/j.actbio.2022.07.046
305. Liang Y, Zhao X, Hu T, et al. Adhesive hemostatic conducting injectable composite hydrogels with sustained drug release and photothermal antibacterial activity to promote full-thickness skin regeneration during wound healing. *Small*. 2019;15(12):e1900046. doi:10.1002/sml.201900046
306. Lin Z, Liu L, Wang W, et al. The role and mechanism of polydopamine and cuttlefish ink melanin carrying copper ion nanoparticles in antibacterial properties and promoting wound healing. *Biomater Sci*. 2021;9(17):5951–5964. doi:10.1039/d1bm00622c
307. Liu C, Yao W, Tian M, Wei J, Song Q, Qiao W. Mussel-inspired degradable antibacterial polydopamine/silica nanoparticle for rapid hemostasis. *Biomaterials*. 2018;179:83–95. doi:10.1016/j.biomaterials.2018.06.037
308. Liu CY, Huang CJ. Functionalization of polydopamine via the aza-michael reaction for antimicrobial interfaces. *Langmuir*. 2016;32(19):5019–5028. doi:10.1021/acs.langmuir.6b00990
309. Liu G, Wang L, He Y, et al. Polydopamine nanosheets doped injectable hydrogel with nitric oxide release and photothermal effects for bacterial ablation and wound healing. *Adv Healthc Mater*. 2021;10(23):e2101476. doi:10.1002/adhm.202101476
310. Liu H, Qu X, Tan H, et al. Role of polydopamine's redox-activity on its pro-oxidant, radical-scavenging, and antimicrobial activities. *Acta Biomater*. 2019;88:181–196. doi:10.1016/j.actbio.2019.02.032
311. Lu Z, Xiao J, Wang Y, Meng M. In situ synthesis of silver nanoparticles uniformly distributed on polydopamine-coated silk fibers for antibacterial application. *J Colloid Interface Sci*. 2015;452:8–14. doi:10.1016/j.jcis.2015.04.015
312. Mohd Daud N, Saeful Bahri IF, Nik Malek NAN, Hermawan H, Saidin S. Immobilization of antibacterial chlorhexidine on stainless steel using crosslinking polydopamine film: towards infection resistant medical devices. *Colloids Surf B*. 2016;145:130–139. doi:10.1016/j.colsurfb.2016.04.046
313. Muller C, Berber E, Lutzweiler G, et al. Polyarginine decorated polydopamine nanoparticles with antimicrobial properties for functionalization of hydrogels. *Front Bioeng Biotechnol*. 2020;8:982. doi:10.3389/fbioe.2020.00982
314. Nazi N, Humblot V, Debieuvre-Chouvy C. A new antibacterial n-halamine coating based on polydopamine. *Langmuir*. 2020;36(37):11005–11014. doi:10.1021/acs.langmuir.0c01856
315. Niyonshuti II, Krishnamurthi VR, Okyere D, et al. Polydopamine surface coating synergizes the antimicrobial activity of silver nanoparticles. *ACS Appl Mater Interfaces*. 2020;12(36):40067–40077. doi:10.1021/acsami.0c10517
316. Peng D, Liu G, He Y, et al. Fabrication of a pH-responsive core-shell nanosystem with a low-temperature photothermal therapy effect for treating bacterial biofilm infection. *Biomater Sci*. 2021;9(22):7483–7491. doi:10.1039/d1bm01329g
317. Qi X, Huang Y, You S, et al. Engineering robust ag-decorated polydopamine nano-photothermal platforms to combat bacterial infection and prompt wound healing. *Adv Sci*. 2022;9(11):e2106015. doi:10.1002/advs.202106015
318. Ren L, He G, Zhou Y, et al. A hydrogel based on nanocellulose/polydopamine/gelatin used for the treatment of MRSA infected wounds with broad-spectrum antibacterial and antioxidant properties and tissue suitability. *Biomater Sci*. 2022;10(12):3174–3187. doi:10.1039/d2bm00157h
319. Sileika TS, Kim HD, Maniak P, Messersmith PB. Antibacterial performance of polydopamine-modified polymer surfaces containing passive and active components. *ACS Appl Mater Interfaces*. 2011;3(12):4602–4610. doi:10.1021/am200978h
320. Song J, Liu H, Lei M, et al. Redox-channeling polydopamine-ferrocene (PDA-Fc) coating to confer context-dependent and photothermal antimicrobial activities. *ACS Appl Mater Interfaces*. 2020;12(7):8915–8928. doi:10.1021/acsami.9b22339
321. Su L, Yu Y, Zhao Y, Liang F, Zhang X. Strong antibacterial polydopamine coatings prepared by a shaking-assisted method. *Sci Rep*. 2016;6:24420. doi:10.1038/srep24420
322. Su R, Yan H, Li P, Zhang B, Zhang Y, Su W. Photo-enhanced antibacterial activity of polydopamine-curcumin nanocomposites with excellent photodynamic and photothermal abilities. *Photodiagnosis Photodyn Ther*. 2021;35:102417. doi:10.1016/j.pdpdt.2021.102417

323. Sun A, Lin X, Xue Z, et al. Facile surface functional polyetheretherketone with antibacterial and immunoregulatory activities for enhanced regeneration toward bacterium-infected bone destruction. *Drug Delivery*. 2021;28(1):1649–1663. doi:10.1080/10717544.2021.1960924
324. Sun J, Tan H, Liu H, et al. A reduced polydopamine nanoparticle-coupled sprayable PEG hydrogel adhesive with anti-infection activity for rapid wound sealing. *Biomater Sci*. 2020;8(24):6946–6956. doi:10.1039/d0bm01213k
325. Wang K, Liu Y, Wang H, Liu Y, Yang X, Sun S. Multi-functional nanofilms capable of angiogenesis, near-infrared-triggered anti-bacterial activity and inflammatory regulation for infected wound healing. *Biomater Adv*. 2022;142:213154. doi:10.1016/j.bioadv.2022.213154
326. Wang LS, Xu S, Gopal S, et al. Facile fabrication of antibacterial and antiviral perhydrolase-polydopamine composite coatings. *Sci Rep*. 2021;11(1):12410. doi:10.1038/s41598-021-91925-6
327. Wei Z, Li K, Wang S, et al. Controllable AgNPs encapsulation to construct biocompatible and antibacterial titanium implant. *Front Bioeng Biotechnol*. 2022;10:1056419. doi:10.3389/fbioe.2022.1056419
328. Xie X, Mao C, Liu X, et al. Tuning the bandgap of photo-sensitive polydopamine/Ag(3)PO(4)/graphene oxide coating for rapid, noninvasive disinfection of implants. *ACS Cent Sci*. 2018;4(6):724–738. doi:10.1021/acscentsci.8b00177
329. Xiong Q, Fang Q, Xu K, et al. Near-infrared light-responsive photothermal α -Fe(2)O(3)@Au/PDA core/shell nanostructure with on-off controllable anti-bacterial effects. *Dalton Trans*. 2021;50(40):14235–14243. doi:10.1039/d1dt02251b
330. Xu Z, Wang T, Liu J. Recent development of polydopamine anti-bacterial nanomaterials. *Int J Mol Sci*. 2022;23(13). doi:10.3390/ijms23137278
331. Yeroslavsky G, Lavi R, Alishaev A, Rahimpour S. Sonochemically-produced metal-containing polydopamine nanoparticles and their antibacterial and antibiofilm activity. *Langmuir*. 2016;32(20):5201–5212. doi:10.1021/acs.langmuir.6b00576
332. Yin J, Han Q, Zhang J, et al. MXene-based hydrogels endow polyetheretherketone with effective osteogenicity and combined treatment of osteosarcoma and bacterial infection. *ACS Appl Mater Interfaces*. 2020;12(41):45891–45903. doi:10.1021/acsami.0c14752
333. Yu QH, Zhang CM, Jiang ZW, Qin SY, Zhang AQ. Mussel-inspired adhesive polydopamine-functionalized hyaluronic acid hydrogel with potential bacterial inhibition. *Glob Chall*. 2020;4(2):1900068. doi:10.1002/gch2.201900068
334. Yuan Z, Tao B, He Y, et al. Biocompatible MoS(2)/PDA-RGD coating on titanium implant with antibacterial property via intrinsic ROS-independent oxidative stress and NIR irradiation. *Biomaterials*. 2019;217:119290. doi:10.1016/j.biomaterials.2019.119290
335. Zhang J, He X, Yu S, et al. A novel dental adhesive containing Ag/polydopamine-modified HA fillers with both antibacterial and mineralization properties. *J Dent*. 2021;111:103710. doi:10.1016/j.jdent.2021.103710
336. Zhang Q, Wang Y, Zhang W, et al. In situ assembly of well-dispersed Ag nanoparticles on the surface of polylactic acid-Au@polydopamine nanofibers for antimicrobial applications. *Colloids Surf B*. 2019;184:110506. doi:10.1016/j.colsurfb.2019.110506
337. Zhang WJ, Li S, Vijayan V, et al. ROS- and pH-responsive polydopamine functionalized Ti(3)C(2)T(x) MXene-based nanoparticles as drug delivery nanocarriers with high antibacterial activity. *Nanomaterials (Basel)*. 2022;12(24). doi:10.3390/nano12244392
338. Zhang Y, Wang F, Huang Q, et al. Layer-by-layer immobilizing of polydopamine-assisted ϵ -polylysine and gum Arabic on titanium: tailoring of antibacterial and osteogenic properties. *Mater Sci Eng C Mater Biol Appl*. 2020;110:110690. doi:10.1016/j.msec.2020.110690
339. Zhang Z, Zhang J, Zhang B, Tang J. Mussel-inspired functionalization of graphene for synthesizing Ag-polydopamine-graphene nanosheets as antibacterial materials. *Nanoscale*. 2013;5(1):118–123. doi:10.1039/c2nr32092d
340. Zhou W, Peng X, Ma Y, et al. Two-staged time-dependent materials for the prevention of implant-related infections. *Acta Biomater*. 2020;101:128–140. doi:10.1016/j.actbio.2019.10.023
341. Bian Y, Wang H, Xu J, et al. Polydopamine-Ag composite surface guides HBMSCs adhesion and proliferation. *Biom Mater*. 2021;16(2):025003. doi:10.1088/1748-605X/abdd6f
342. Cheng YL, Chen YW, Wang K, Shie MY. Enhanced adhesion and differentiation of human mesenchymal stem cell inside apatite-mineralized/poly(dopamine)-coated poly(ϵ -caprolactone) scaffolds by stereolithography. *J Mat Chem B*. 2016;4(38):6307–6315. doi:10.1039/c6tb01377e
343. Choi CKK, Choi CHJ, Bian L. A Gold@Polydopamine Core-shell nanoprobe for long-term intracellular detection of MicroRNAs in differentiating stem cells. *Methods Mol Biol*. 2017;1570:155–164. doi:10.1007/978-1-4939-6840-4_10
344. Ejeian F, Razmjou A, Nasr-Esfahani MH, et al. ZIF-8 modified polypropylene membrane: a biomimetic cell culture platform with a view to the improvement of guided bone regeneration. *Int j Nanomed*. 2020;15:10029–10043. doi:10.2147/ijn.S269169
345. Ho CC, Ding SJ. Novel SiO(2)/PDA hybrid coatings to promote osteoblast-like cell expression on titanium implants. *J Mat Chem B*. 2015;3(13):2698–2707. doi:10.1039/c4tb01841a
346. Huang Y, Jing W, Li Y, Cai Q, Yang X. Composites made of polyorganophosphazene and carbon nanotube up-regulating osteogenic activity of BMSCs under electrical stimulation. *Colloids Surf B*. 2021;204:111785. doi:10.1016/j.colsurfb.2021.111785
347. Jia L, Han F, Wang H, et al. Polydopamine-assisted surface modification for orthopaedic implants. *J Orthop Translat*. 2019;17:82–95. doi:10.1016/j.jot.2019.04.001
348. Jin L, Yuan F, Chen C, et al. Degradation products of polydopamine restrained inflammatory response of LPS-stimulated macrophages through mediation TLR-4-MYD88 dependent signaling pathways by antioxidant. *Inflammation*. 2019;42(2):658–671. doi:10.1007/s10753-018-0923-3
349. Kim M, Kim JS, Lee H, Jang JH. Polydopamine-decorated sticky, water-friendly, biodegradable polycaprolactone cell carriers. *Macromol biosci*. 2016;16(5):738–747. doi:10.1002/mabi.201500432
350. Kim S, Jang LK, Jang M, Lee S, Hardy JG, Lee JY. Electrically conductive polydopamine-polypyrrole as high performance biomaterials for cell stimulation in vitro and electrical signal recording in vivo. *ACS Appl Mater Interfaces*. 2018;10(39):33032–33042. doi:10.1021/acsami.8b11546
351. Lee EJ, Ahmad K, Pathak S, et al. Identification of Novel FNIN2 and FNIN3 fibronectin-derived peptides that promote cell adhesion, proliferation and differentiation in primary cells and stem cells. *Int J Mol Sci*. 2021;22(6). doi:10.3390/ijms22063042
352. Li J, Tan L, Liu X, et al. Balancing bacteria-osteoblast competition through selective physical puncture and biofunctionalization of ZnO/polydopamine/arginine-glycine-aspartic acid-cysteine nanorods. *ACS nano*. 2017;11(11):11250–11263. doi:10.1021/acsnano.7b05620
353. Li Q, Sun L, Zhang L, Xu Z, Kang Y, Xue P. Polydopamine-collagen complex to enhance the biocompatibility of polydimethylsiloxane substrates for sustaining long-term culture of L929 fibroblasts and tendon stem cells. *J Biomed Mater Res A*. 2018;106(2):408–418. doi:10.1002/jbm.a.36254
354. Li W, Zheng Y, Zhao X, et al. Osteoinductive effects of free and immobilized bone forming peptide-1 on human adipose-derived stem cells. *PLoS One*. 2016;11(3):e0150294. doi:10.1371/journal.pone.0150294
355. Li X, Wei Z, Lv H, et al. Iron oxide nanoparticles promote the migration of mesenchymal stem cells to injury sites. *Int j Nanomed*. 2019;14:573–589. doi:10.2147/ijn.S184920

356. Liu C, Li Y, Wang J, Liu C, Liu W, Jian X. Improving hydrophilicity and inducing bone-like apatite formation on PPBES by polydopamine coating for biomedical application. *Molecules*. 2018;23(7). doi:10.3390/molecules23071643
357. Liu W, Morschauser A, Zhang X, et al. Human placenta-derived adherent cells induce tolerogenic immune responses. *Clin Transl Immunology*. 2014;3(5):e14. doi:10.1038/cti.2014.5
358. Meng Z, Liu Y, Xu K, et al. Biomimetic Polydopamine-Modified Silk Fibroin/Curcumin Nanofibrous Scaffolds for Chemo-photothermal Therapy of Bone Tumor. *ACS Omega*. 2021;6(34):22213–22223. doi:10.1021/acsomega.1c02903
359. Mertgen AS, Trossmann VT, Guex AG, Maniura-Weber K, Scheibel T, Rottmar M. Multifunctional biomaterials: combining material modification strategies for engineering of cell-contacting surfaces. *ACS Appl Mater Interfaces*. 2020;12(19):21342–21367. doi:10.1021/acscami.0c01893
360. Pacelli S, Rampetsreiter K, Modaresi S, et al. Fabrication of a double-cross-linked interpenetrating polymeric network (IPN) hydrogel surface modified with polydopamine to modulate the osteogenic differentiation of adipose-derived stem cells. *ACS Appl Mater Interfaces*. 2018;10(30):24955–24962. doi:10.1021/acscami.8b05200
361. Shi X, Li L, Ostrovidov S, Shu Y, Khademhosseini A, Wu H. Stretchable and micropatterned membrane for osteogenic differentiation of stem cells. *ACS Appl Mater Interfaces*. 2014;6(15):11915–11923. doi:10.1021/am5029236
362. Shin J, Cho JH, Jin Y, et al. Mussel adhesion-inspired reverse transfection platform enhances osteogenic differentiation and bone formation of human adipose-derived stem cells. *Small*. 2016;12(45):6266–6278. doi:10.1002/sml.201601868
363. Wu C, Han P, Liu X, et al. Mussel-inspired bioceramics with self-assembled Ca-P/polydopamine composite nanolayer: preparation, formation mechanism, improved cellular bioactivity and osteogenic differentiation of bone marrow stromal cells. *Acta Biomater*. 2014;10(1):428–438. doi:10.1016/j.actbio.2013.10.013
364. Xing X, Han S, Ni Y, et al. Mussel-inspired functionalization of electrospun scaffolds with polydopamine-assisted immobilization of mesenchymal stem cells-derived small extracellular vesicles for enhanced bone regeneration. *Int J Pharm*. 2021;609:121136. doi:10.1016/j.ijpharm.2021.121136
365. Xu Q, Bai Y, Misra RDK, et al. Improving biological functions of three-dimensional printed Ti2448 scaffolds by decoration with polydopamine and extracellular matrices. *ACS Appl Bio Mater*. 2022;5(8):3982–3990. doi:10.1021/acscabm.2c00521
366. Ye K, Liu D, Kuang H, et al. Three-dimensional electrospun nanofibrous scaffolds displaying bone morphogenetic protein-2-derived peptides for the promotion of osteogenic differentiation of stem cells and bone regeneration. *J Colloid Interface Sci*. 2019;534:625–636. doi:10.1016/j.jcis.2018.09.071
367. Yeh CH, Chen YW, Shie MY, Fang HY. Poly(Dopamine)-Assisted Immobilization of Xu Duan on 3D Printed Poly(Lactic Acid) scaffolds to up-regulate osteogenic and angiogenic markers of bone marrow stem cells. *Materials*. 2015;8(7):4299–4315. doi:10.3390/ma8074299
368. Yin X, Ran S, Cheng H, et al. Polydopamine-modified ZIF-8 nanoparticles as a drug carrier for combined chemo-photothermal osteosarcoma therapy. *Colloids Surf B*. 2022;216:112507. doi:10.1016/j.colsurfb.2022.112507
369. Yu Y, Wang X, Zhu Y, He Y, Xue H, Ding J. Is polydopamine beneficial for cells on the modified surface? *Regen Biomater*. 2022;9:rbac078. doi:10.1093/rb/rbac078
370. Zhang M, Zhang F, Liu T, et al. Polydopamine nanoparticles camouflaged by stem cell membranes for synergistic chemo-photothermal therapy of malignant bone tumors. *Int J Nanomed*. 2020;15:10183–10197. doi:10.2147/ijn.S282931
371. Aikio O, Härmä A, Härkin P, et al. Inflammatory biomarkers in very preterm infants during early intravenous paracetamol administration. *Early Hum Dev*. 2021;161:105464. doi:10.1016/j.earlhumdev.2021.105464
372. Al-Sayed E, El-Naga RN. Protective role of ellagitannins from *Eucalyptus citriodora* against ethanol-induced gastric ulcer in rats: impact on oxidative stress, inflammation and calcitonin-gene related peptide. *Phytomedicine*. 2015;22(1):5–15. doi:10.1016/j.phymed.2014.10.002
373. Augimeri G, Plastina P, Gionfriddo G, et al. N-eicosapentaenoyl dopamine, A Conjugate of Dopamine and Eicosapentaenoic Acid (EPA), exerts anti-inflammatory properties in mouse and human macrophages. *Nutrients*. 2019;11(9). doi:10.3390/nu11092247
374. Aursnes M, Tungen JE, Vik A, et al. Total synthesis of the lipid mediator PD1n-3 DPA: configurational assignments and anti-inflammatory and pro-resolving actions. *J Nat Prod*. 2014;77(4):910–916. doi:10.1021/np4009865
375. Bedair TM, Lee CK, Kim DS, et al. Magnesium hydroxide-incorporated PLGA composite attenuates inflammation and promotes BMP2-induced bone formation in spinal fusion. *J Tissue Eng*. 2020;11:2041731420967591. doi:10.1177/2041731420967591
376. Chehl N, Gong Q, Chipitsyna G, Aziz T, Yeo CJ, Arafat HA. Angiotensin II regulates the expression of monocyte chemoattractant protein-1 in pancreatic cancer cells. *J Gastrointest Surg*. 2009;13(12):2189–2200. doi:10.1007/s11605-009-1055-8
377. Chlif N, Bouymajane A, Oulad El Majdoub Y, et al. Phenolic compounds, in vivo anti-inflammatory, analgesic and antipyretic activities of the aqueous extracts from fresh and dry aerial parts of *Brocchia cinerea* (Vis.). *J Pharm Biomed Anal*. 2022;213:114695. doi:10.1016/j.jpba.2022.114695
378. Costa G, Ferreira JP, Vitorino C, et al. Polyphenols from *Cymbopogon citratus* leaves as topical anti-inflammatory agents. *J Ethnopharmacol*. 2016;178:222–228. doi:10.1016/j.jep.2015.12.016
379. Dangi B, Obeng M, Nauroth JM, et al. Biogenic synthesis, purification, and chemical characterization of anti-inflammatory resolvins derived from docosapentaenoic acid (DPA_n-6). *J Biol Chem*. 2009;284(22):14744–14759. doi:10.1074/jbc.M809014200
380. Do Santos RC, de Souza AV, Andrade-Silva M, et al. Antioxidant, anti-rheumatic and anti-inflammatory investigation of extract and dicentrinone from *Duguetia furfuracea* (A. St.-Hil.) Benth. & Hook. f. *J Ethnopharmacol*. 2018;211:9–16. doi:10.1016/j.jep.2017.09.019
381. Du Preez CI, Gründemann C, Reinhardt JK, Mumbengegwi DR, Huber R. Immunomodulatory effects of some Namibian plants traditionally used for treating inflammatory diseases. *J Ethnopharmacol*. 2020;254:112683. doi:10.1016/j.jep.2020.112683
382. El-Mekkawy S, Shahat AA, Alqahtani AS, et al. A polyphenols-rich extract from *Moricandia sinaica* Boiss. Exhibits analgesic, anti-inflammatory and antipyretic activities in vivo. *Molecules*. 2020;25(21). doi:10.3390/molecules25215049
383. Faqueti LG, Briudes V, Halabalaki M, et al. Antinociceptive and anti-inflammatory activities of standardized extract of polymethoxyflavones from *Ageratum conyzoides*. *J Ethnopharmacol*. 2016;194:369–377. doi:10.1016/j.jep.2016.09.025
384. Fatima H, Shahid M, Pruitt C, et al. Chemical Fingerprinting, antioxidant, and anti-inflammatory potential of hydroethanolic extract of *Trigonella foenum-graecum*. *Antioxidants*. 2022;11(2). doi:10.3390/antiox11020364
385. Golabchi A, Wu B, Cao B, Bettinger CJ, Cui XT. Zwitterionic polymer/polydopamine coating reduce acute inflammatory tissue responses to neural implants. *Biomaterials*. 2019;225:119519. doi:10.1016/j.biomaterials.2019.119519
386. Horton SC, Tan AL, Freeston JE, Wakefield RJ, Buch MH, Emery P. Discordance between the predictors of clinical and imaging remission in patients with early rheumatoid arthritis in clinical practice: implications for the use of ultrasound within a treatment-to-target strategy. *Rheumatology (Oxford)*. 2016;55(7):1177–1187. doi:10.1093/rheumatology/kew037

387. Lei J, Zhou Y, Zhao H, et al. Dabigatran activates inflammation resolution by promoting fibrinogen-like protein 2 shedding and RvD5(n-3 DPA) production. *Theranostics*. 2021;11(9):4251–4261. doi:10.7150/thno.50182
388. Lima TC, Matos SS, Carvalho TF, et al. Evidence for the involvement of IL-1 β and TNF- α in anti-inflammatory effect and antioxidative stress profile of the standardized dried extract from *Miconia albicans* Sw. (Triana) Leaves (Melastomataceae). *J Ethnopharmacol*. 2020;259:112908. doi:10.1016/j.jep.2020.112908
389. Magiera A, Marchelak A, Michel P, Owczarek A, Olszewska MA. Lipophilic extracts from leaves, inflorescences and fruits of *Prunus padus* L. as potential sources of corosolic, ursolic and oleanolic acids with anti-inflammatory activity. *Nat Prod Res*. 2021;35(13):2263–2268. doi:10.1080/14786419.2019.1666385
390. Misra A, Chaudhary MK, Tripathi D, et al. Nutritional potential of an edible terrestrial orchid *Eulophia nuda* LINDL and validation of its traditional claim in arthritis. *J Ethnopharmacol*. 2023;306:116123. doi:10.1016/j.jep.2022.116123
391. Nauroth JM, Liu YC, Van Elswyk M, et al. Docosahexaenoic acid (DHA) and docosapentaenoic acid (DPA-n-6) algal oils reduce inflammatory mediators in human peripheral mononuclear cells in vitro and paw edema in vivo. *Lipids*. 2010;45(5):375–384. doi:10.1007/s11745-010-3406-3
392. Nesman JI, Gangestad Primdahl K, Tungen JE, Palmas F, Dalli J, Hansen TV. Synthesis, Structural Confirmation, and Biosynthesis of 22-OH-PD1(n-3 DPA). *Molecules*. 2019;24(18). doi:10.3390/molecules24183228
393. Olszewska MA, Granica S, Kolodziejczyk-Czepas J, et al. Variability of sinapic acid derivatives during germination and their contribution to antioxidant and anti-inflammatory effects of broccoli sprouts on human plasma and human peripheral blood mononuclear cells. *Food Funct*. 2020;11(8):7231–7244. doi:10.1039/d0fo01387k
394. Pablos JL, Jover JA, Roman-Ivorra JA, et al. Patient Decision Aid (PDA) for patients with rheumatoid arthritis reduces decisional conflict and improves readiness for treatment decision making. *Patient*. 2020;13(1):57–69. doi:10.1007/s40271-019-00381-y
395. Panagiotidou C, Burgers LD, Tsadila C, et al. HPLC- and NMR-based chemical profiling, wound-healing potential, anti-inflammatory and antibacterial activities of *Satureja pilosa* (Lamiaceae), a neglected medicinal-aromatic herb. *Plants*. 2023;12(24). doi:10.3390/plants12244114
396. Pangopoulos MK, Nolsøe JMN, Antonsen SG, et al. Enzymatic studies with 3-oxa n-3 DPA. *Bioorg Chem*. 2020;96:103653. doi:10.1016/j.bioorg.2020.103653
397. Park JU, Kim SJ, Na CS, et al. Chondroprotective and anti-inflammatory effects of ChondroT, a new complex herbal medication. *BMC Complement Altern Med*. 2016;16:213. doi:10.1186/s12906-016-1211-0
398. Perestrelo R, Silva CL, Câmara JS. Determination of urinary levels of leukotriene B(4) using a highly specific and sensitive methodology based on automatic MEPS combined with UHPLC-PDA analysis. *Talanta*. 2015;144:382–389. doi:10.1016/j.talanta.2015.06.054
399. Popov AM, Kozlovskaya EP, Klimovich AA, et al. Carotenoids from starfish *patiria pectinifera*: therapeutic activity in models of inflammatory diseases. *Mar Drugs*. 2023;21(9). doi:10.3390/md21090470
400. Ragab AE, Al-Madboly LA, Al-Ashmawy GM, Saber-Ayad M, Abo-Saif MA. Unravelling the in vitro and in vivo anti-helicobacter pylori effect of delphinidin-3-O-glucoside rich extract from pomegranate exocarp: enhancing autophagy and downregulating TNF- α and COX2. *Antioxidants*. 2022;11(9). doi:10.3390/antiox11091752
401. Si Y, Li X, Guo T, et al. Isolation and characterization of phellodendronoside A, a new isoquinoline alkaloid glycoside with anti-inflammatory activity from *Phellodendron chinense* Schneid. *Fitoterapia*. 2021;154:105021. doi:10.1016/j.fitote.2021.105021
402. Toupin-April K, Huber AM, Duffy CM, et al. Development and acceptability of a patient decision aid for pain management in juvenile idiopathic arthritis: the JIA option map. *Patient*. 2020;13(6):719–728. doi:10.1007/s40271-020-00458-z
403. Tungen JE, Aursnes M, Dalli J, Arnardottir H, Serhan CN, Hansen TV. Total synthesis of the anti-inflammatory and pro-resolving lipid mediator MaR1n-3 DPA utilizing an sp(3)-sp(3) Negishi cross-coupling reaction. *Chemistry*. 2014;20(45):14575–14578. doi:10.1002/chem.201404721
404. Tungen JE, Primdahl KG, Hansen TV. The first total synthesis of the lipid mediator PD2(n-3 DPA). *J Nat Prod*. 2020;83(7):2255–2260. doi:10.1021/acs.jnatprod.0c00385
405. Washington K, Shacklady C. Patients' experience of shared decision making using an online patient decision aid for osteoarthritis of the knee—A service evaluation. *Musculoskeletal Care*. 2015;13(2):116–126. doi:10.1002/msc.1086
406. Wu D, Chen J, Zhu H, et al. UPLC-PDA determination of paeoniflorin in rat plasma following the oral administration of *Radix Paeoniae Alba* and its effects on rats with collagen-induced arthritis. *Exp Ther Med*. 2014;7(1):209–217. doi:10.3892/etm.2013.1358
407. Ashraf A, Sarfraz RA, Rashid MA, Mahmood A, Shahid M, Noor N. Chemical composition, antioxidant, antitumor, anticancer and cytotoxic effects of *Psidium guajava* leaf extracts. *Pharm Biol*. 2016;54(10):1971–1981. doi:10.3109/13880209.2015.1137604
408. Assar DH, Elhabashi N, Mokhatly AA, et al. Wound healing potential of licorice extract in rat model: antioxidants, histopathological, immunohistochemical and gene expression evidences. *Biomed Pharmacother*. 2021;143:112151. doi:10.1016/j.biopha.2021.112151

Drug Design, Development and Therapy

Dovepress

Publish your work in this journal

Drug Design, Development and Therapy is an international, peer-reviewed open-access journal that spans the spectrum of drug design and development through to clinical applications. Clinical outcomes, patient safety, and programs for the development and effective, safe, and sustained use of medicines are a feature of the journal, which has also been accepted for indexing on PubMed Central. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/drug-design-development-and-therapy-journal>