

# Endocannabinoid Hydrolase Inhibitors: Potential Novel Anxiolytic Drugs

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**Abstract:** Over the past decade, the idea of targeting the endocannabinoid system to treat anxiety disorders has received increasing attention. Previous studies focused more on developing cannabinoid receptor agonists or supplementing exogenous cannabinoids, which are prone to various adverse effects due to their strong pharmacological activity and poor receptor selectivity, limiting their application in clinical research. Endocannabinoid hydrolase inhibitors are considered to be the most promising development strategies for the treatment of anxiety disorders. More recent efforts have emphasized that inhibition of two major endogenous cannabinoid hydrolases, monoacylglycerol lipase (MAGL) and fatty acid amide hydrolase (FAAH), indirectly activates cannabinoid receptors by increasing endogenous cannabinoid levels in the synaptic gap, circumventing receptor desensitization resulting from direct enhancement of endogenous cannabinoid signaling. In this review, we comprehensively summarize the anxiolytic effects of MAGL and FAAH inhibitors and their potential pharmacological mechanisms, highlight reported novel inhibitors or natural products, and provide an outlook on future directions in this field.

**Keywords:** endocannabinoid hydrolase inhibitors, endocannabinoid system, anxiety disorders, anxiolytic, MAGL, FAAH

## Introduction

Anxiety disorders represent prevalent mental health conditions characterized by persistent psychogenic anxiety, somatic anxiety, and sleep disorders. According to the Diagnostic and Statistical Manual of Mental Disorders, Fifth Edition (DSM-5), the classification of anxiety disorders includes Generalized Anxiety Disorder, Phobias (including specific phobias and social anxiety disorder), Panic Disorder, as well as other anxiety-related conditions such as Separation Anxiety Disorder and Selective Mutism.<sup>1</sup> The global prevalence of anxiety disorders ranges from 7.3% to 28.0%, making them a significant concern worldwide.<sup>2</sup> Specifically, in China, anxiety disorders stand out as the most prevalent mental illness, with a lifetime incidence of 7.6%.<sup>3</sup>

The World Health Organization underscores the significance of anxiety disorders by ranking them as the sixth leading contributor to global disability, establishing it as a pressing health and wellness concern that demands attention.<sup>4</sup> Regrettably, the existing repertoire of anxiolytic medications faces challenges in adequately addressing the needs of the extensive population affected by anxiety disorders.<sup>5</sup> Benzodiazepines and selective serotonin reuptake inhibitors (SSRIs) antidepressants are the primary drugs employed clinically for the treatment of anxiety disorders.<sup>6,7</sup> However, their anxiolytic effects are secondary indications rather than primary, and the drugs, especially benzodiazepines, are subject to stringent controls and possess potential addictive properties.<sup>8</sup> Consequently, the development of novel drugs targeting anxiety disorders remains a focal point in the ongoing research and development of psychotropic medications.

Traditionally, anxiolytic drug development focused on neurotransmitter-based hypotheses, yielding medications like eszopiclone and buspirone.<sup>9,10</sup> Recent attention has turned to novel approaches, particularly targeting the endocannabinoid

system (ECS). The ECS, integral to mood regulation, has been implicated in the pathogenesis of both anxiety and depression, underscoring its potential as a novel therapeutic frontier.<sup>11</sup> Novel drug designs that target the ECS are considered to be the most promising candidates for the treatment of anxiety disorders.<sup>12</sup>

Studies explore ECS modulation for anxiolysis, including increasing endogenous cannabinoids (eCBs) secretion and inhibiting their hydrolase enzymes. However, challenges arise due to the unique characteristics of endocannabinoids. Efforts to develop cannabinoid receptor agonists face obstacles like pharmacological intensity and adverse effects. A more promising strategy involves inhibiting eCB hydrolases, notably monoacylglycerol lipase (MAGL) and fatty acid amide hydrolase (FAAH), indirectly activating cannabinoid receptors. Clinical trials with FAAH inhibitors, such as JNJ-42165279, have shown positive anxiolytic potential.<sup>13</sup> This paper comprehensively summarizes the anxiolytic effects of eCB hydrolase inhibitors, focusing on underlying mechanisms and highlighting novel inhibitors or natural products. The insights from existing studies aim to provide new perspectives for the development of innovative anxiolytic drugs.

## Endocannabinoid System

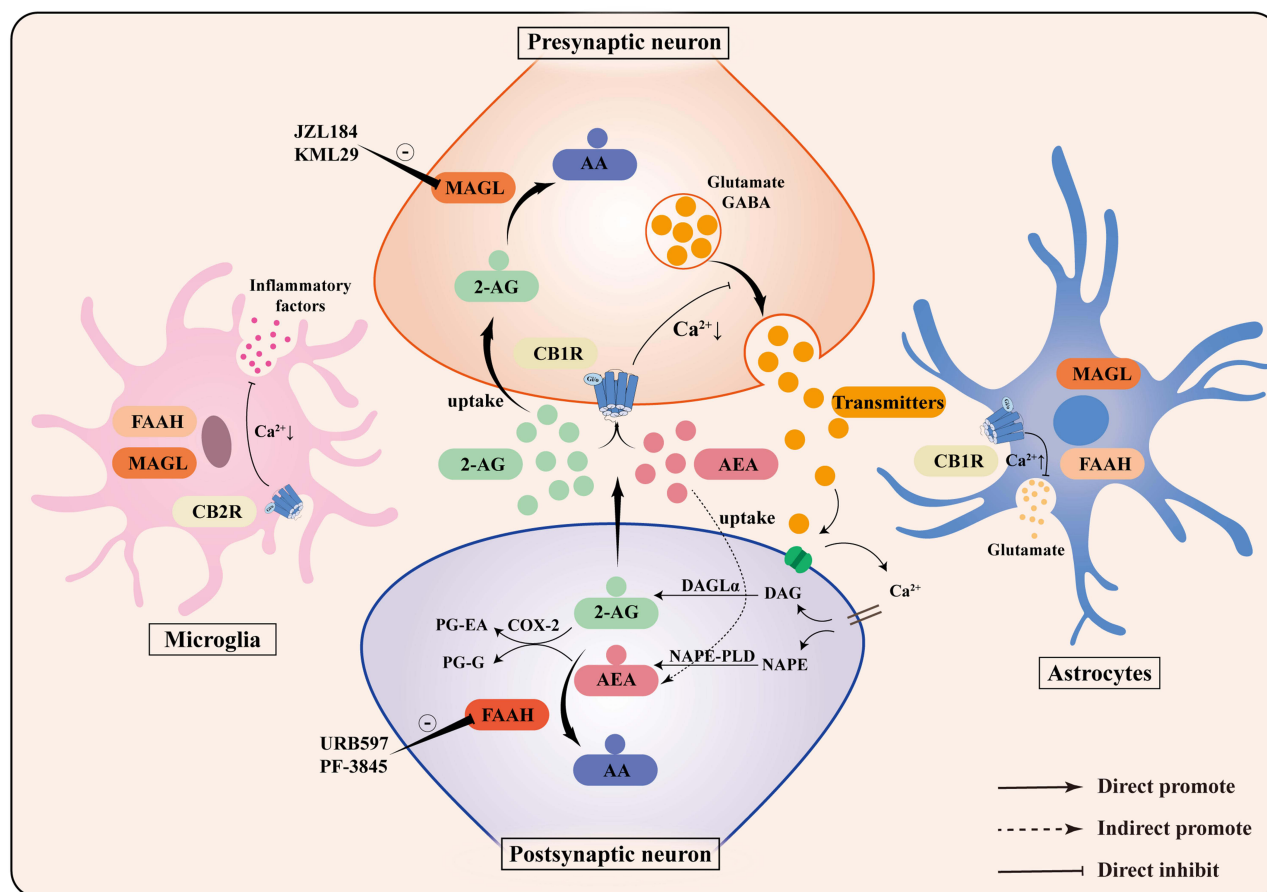
The ECS is one of the crucial regulatory systems in the central nervous system, and consists of cannabinoid receptor 1 (CB1R), cannabinoid receptor 2 (CB2R), eCBs, and their corresponding synthesizing and degrading enzymes. eCBs are classified into three categories according to chemical structure: 1) esters: such as 2-arachidonoylglycerol (2-AG), anandamide (AEA); 2) amides: such as palmitoylethanolamide (PEA), oleoylethanolamide (OEA); 3) ethers: 2-arachidonyl glyceryl ether (noladin ether or 2-AGE).<sup>14,15</sup> Among them, AEA and 2-AG are more abundant and widely distributed in the human body, while specific eCBs such as OEA and 2-AGE can only be detected in particular regions of brain tissue.<sup>16</sup> Notably, 2-AG is agonistic for both CB1R and CB2R, whereas AEA selectively has a high affinity for CB1R but almost no activity on CB2R.<sup>17</sup>

The ways of synthesis, transport and inactivation of 2-AG and AEA in their respective target tissues are also different. 2-AG is mainly produced by 1-oleoyl-2-arachidonoyl-sn-glycerol (OAG) and 1-stearoyl-2-arachidonoylglycerol (SAG) via diacylglycerol lipase  $\alpha$  (DAGL $\alpha$ ), whereas AEA produced by N-acylphosphatidylethanolamine (NAPE) catalysed by N-acylphosphatidylethanolamine-specific phospholipase D (NAPE-PLD).<sup>11</sup> Degradation and inactivation of eCBs mainly involve two pathways: hydrolysis and oxidation. MAGL, a specific hydrolase of 2-AG, is mainly distributed on the axons of presynaptic neurons and is able to terminate retrograde eCB signaling generated by postsynaptic neuronal activity. Approximately 85% of 2-AG in the brain is hydrolyzed and metabolized to arachidonic acid and glycerol by MAGL.<sup>18</sup> AEA was mainly degraded by FAAH into free arachidonic acid and ethanolamine. FAAH, a serine hydrolase mainly present in the endoplasmic membrane, is widely distributed in the central nervous system and hydrolyses various fatty acid amides, including AEA. In another oxidation pathway, the critical enzyme involved is cyclooxygenase-2 (COX-2), catalyzing the synthesis of prostaglandin ethanolamide (PG-EA) and prostaglandin glycerol (PG-G) from 2-AG and AEA.<sup>11</sup> Figure 1 briefly illustrates the composition of the endocannabinoid system, and the synthesis and degradation pathways of endocannabinoids.

Numerous studies have demonstrated that the anxiolytic effects could be exerted by increasing eCB secretion, applying exogenous cannabinoids, agonizing cannabinoid receptors, and decreasing eCB hydrolase activity.<sup>19–21</sup> However, unlike classical neurotransmitters and neuropeptides, eCBs are not stored within synaptic vesicles, but are produced on demand and immediately released from neurons. At the same time, the eCBs synthesis pathways are numerous and complex, and the rate-limiting enzymes are plentiful and poorly defined, leading to insignificant gains through the strategy of increasing the activity of eCB synthetases. In addition, drug development targeting cannabinoid receptor agonists or adding exogenous cannabinoids is susceptible to exclusion from clinical trials or even post-market withdrawal due to stronger pharmacological activity, irreversible binding, poor receptor selectivity, and increased risk of adverse effects.<sup>22,23</sup> Therefore, the most prominent strategies may be to search for inhibitors of eCB hydrolases MAGL and FAAH, especially reversible inhibitors that indirectly activate cannabinoid receptors by increasing synaptic gap levels of 2-AG and AEA, respectively.<sup>24</sup>

## 2-AG Hydrolase MAGL Inhibitors in Treatment for Anxiety

2-AG, an abundant brain eCB, surpasses AEA levels by 200 times.<sup>25</sup> Clinical studies revealed that serum 2-AG levels are significantly reduced in patients with post-traumatic stress disorder (PTSD), which was previously often categorized as an anxiety disorder.<sup>26</sup> Additionally, it has been found that increasing 2-AG levels through exercise can alleviate anxiety



**Figure 1** Composition of the endocannabinoid system. The endocannabinoid system consists of CB1R, CB2R, endocannabinoids, and their corresponding synthesizing and degrading enzymes. 2-AG and AEA, the primary endocannabinoids, are produced on demand and are synthesized from the postsynaptic terminals by DAGL $\alpha$  and NAPE-PLD, respectively, to activate presynaptic cannabinoid receptors. CB1R activation inhibits presynaptic neurotransmitter release and promotes astrocytic glutamate release. CB2R activation reduces microglial inflammatory factor production. 2-AG and AEA are enzymatically degraded to AA by MAGL and FAAH hydrolases, and can also be oxidatively degraded to PG-EA and PG-G by COX-2.

**Abbreviations:** CB1R, cannabinoid receptor 1; CB2R, cannabinoid receptor 2; 2-AG, 2-arachidonoylglycerol; AEA, anandamide; NAPE-PLD, N-acylphosphatidylethanolamine-specific phospholipase D; DAGL $\alpha$ , diacylglycerol lipase  $\alpha$ ; AA, arachidonic acid; MAGL, monoacylglycerol lipase; FAAH, fatty acid amide hydrolase; PG-EA, prostaglandin ethanolamide; PG-G, prostaglandin glycerol; COX-2, cyclooxygenase-2.

symptoms.<sup>27</sup> These findings are consistent with rodent studies,<sup>28,29</sup> where higher MAGL levels have been strongly linked to the production of anxious behaviors.<sup>30</sup> Inhibition of MAGL activity enhances central synaptic 2-AG-mediated phasic and tonic signaling, which has positive implications in anxiety relief, reduction of stress-induced anxiety susceptibility, and fear extinction.<sup>31</sup>

## Anxiolytic Effects of MAGL Inhibitors

Due to MAGL's pivotal role in regulating 2-AG levels and synaptic transmission, efforts to enhance 2-AG signaling for therapeutic purposes have centered on inhibiting MAGL enzyme activity. Numerous pharmacological studies highlight that systemic or local administration of MAGL inhibitors can effectively reduce anxiety-like behaviors induced by acute or chronic stress (Table 1). For example, aberrant excitation of glutamatergic neurons in basolateral amygdala (BLA)-prelimbic prefrontal cortex (plPFC) neural circuit in mice subjected to chronic stress, accompanied by abnormal 2-AG-CB1R signal, and administration of a MAGL inhibitor reversed anxiety-like behavior.<sup>32</sup> Overexpression of MAGL in hippocampal glutamatergic neurons also increases anxiety-like behavior in animals.<sup>33</sup> In addition, anxiety-like behaviors induced by traumatic brain injury and alcohol withdrawal improved after administration with MAGL inhibitor.<sup>34</sup>

However, it is important to note that 2-AG levels do not consistently correlate negatively with anxiety disorders. Acute stress given to healthy people increased circulating concentrations of AEA *in vivo* but had no significant effect on 2-AG.<sup>45</sup> In

**Table 1** Summary of the Anxiolytic Effects of MAGL Inhibitors in Preclinical Studies

Drug	Dose/Administration	Animal	Model	Test	Effects	References
JZL184	2, 10, 40 mg/kg, i.p	C57BL/6j mice, male	–	EPM; LDT	↑ percent time in open-arm; ↑ percent light time	[35]
	2 mg/kg, i.p	Sprague-Dawley rats, male and female	ELS	OFT	↑ activity level	[36]
	1, 3 mg/kg, i.p	C57BL/6j mice, male	ARS	EPM	↑ percent time in open-arm	[34]
	5, 8, 10, 40 mg/kg, i.p	ICR mice, male	ARS and Foot shock stress	LDT; NIH; EZM; OFT	↑ percent light time and distance; ↓ feeding latency and ↑ food consumption; ↑ open arm entries and total distance, ↓ time immobile in open arm and exit latency; ↑ total distance and ↓ number of faeces	[37]
	3, 5, 10, 15 mg/kg, i.p	ICR mice, male	ARS	LDT	↑ percent light time, light distance, and total distance	[38]
	1 μg, intra-NAc microinjection	C57BL/6j mice, male	CSDS	OFT; LDT; EPM	↑ percent center time; ↑ time in light; ↑ percent time in open arm	[39]
	8 mg/kg, i.p	C57BL/6j mice, male	CUS	EPM; LDT	↑ duration and frequency in open arm; ↑ time and frequency in light compartment	[40]
	4, 8, 16 mg/kg, i.p	ICR mice, male	CRS	OFT; NIFS; EPM; MBT	No significance in OFT and EPM test; ↓ feeding latency; ↓ marble burying	[41]
	16 mg/kg, i.p	C57BL/6j mice, male	-	MBT	↓ marble burying	[42]
KML29	200 ng, intra-vmPFC microinjection	Fischer-344 rats, male	Tail shocks	Social exploration	↑ social exploration	[43]
	40 mg/kg, i.p		Tail shocks	Social exploration	↓ social exploration	
MJN110	5, 10 mg/kg, i.p	Wistar rats, male and female	-	NIH	↑ feeding consumption	[44]
	10, 20 mg/kg, i.p	Wistar rats, male	ARS	EPM	↑ percent time in open-arm	[34]

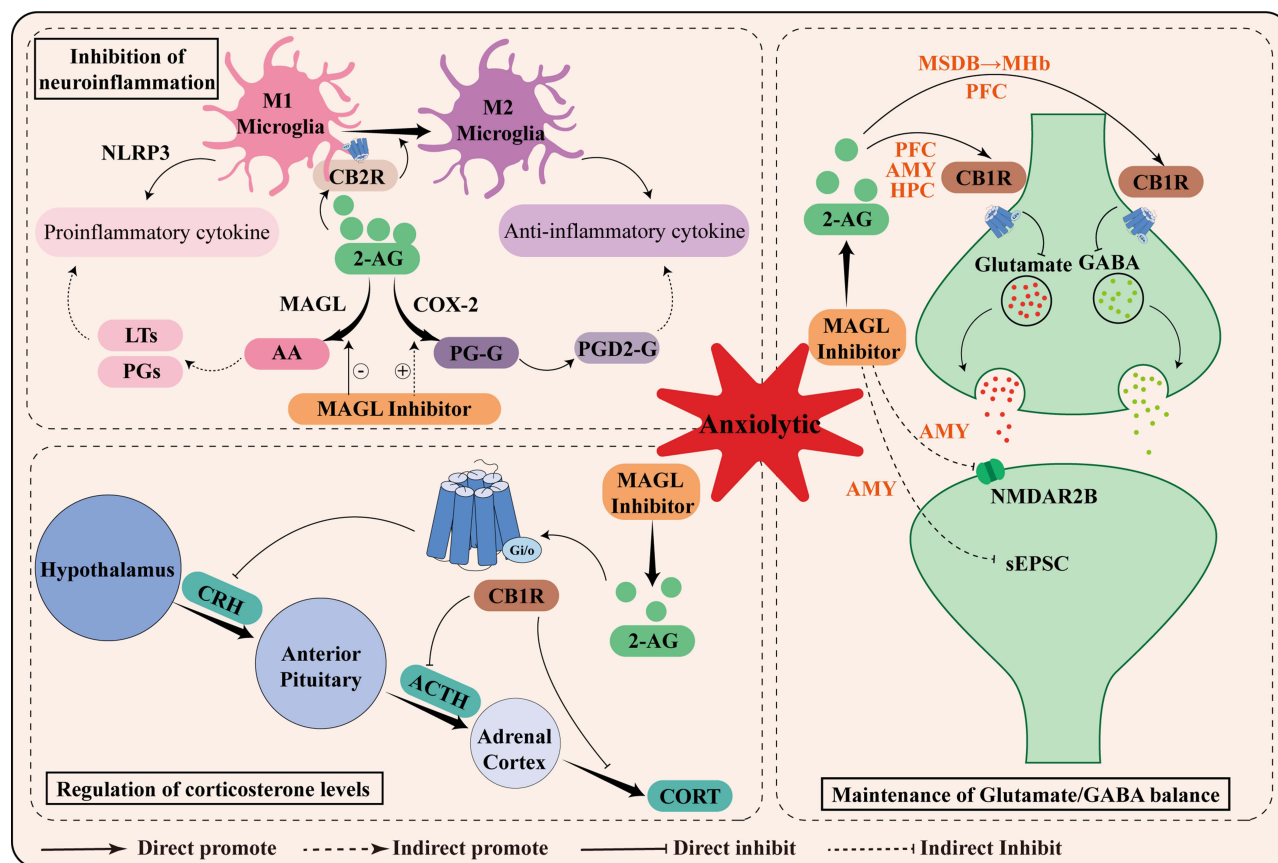
**Abbreviations:** ARS, acute restraint stress; CRS, chronic restraint stress; CSDS, chronic social defeat stress; CUS, chronic unpredictable stress; ELS, early life stress; EPM, elevated plus maze; EZM, elevated zero maze; i.p, intraperitoneal; LDT, light-dark box test; MBT, marble burying test; NAc, nucleus accumbens; NIFS, novelty-induced feeding suppression; NIH, novelty-induced hypophagia; OFT, open-field test.

chronic restraint stress mice, 2-AG levels in the cingulate cortex (ACC), caudate putamen (CP), nucleus accumbens (NAc), and piriform cortex (PIR) all increased.<sup>46</sup> Similarly, 2-AG content increased in the amygdala of chronic stress-induced anxiety mice.<sup>41,47</sup> The researchers explained that the above phenomenon was due to CB1R desensitization, as it did not affect the anxiolytic effect of applying JZL184. Bedse G's research also supports the idea that increased 2-AG is a compensatory response to counteract anxiety-like behaviors induced by stress, and that 2-AG pharmacological enhancers can augment this response to more effectively counteract the adverse effects of stress.<sup>38</sup> Therefore, the benefit of elevating 2-AG pharmacologically through MAGL inhibitors in the treatment of anxiety disorders is definitive.

Interestingly, instead of demonstrating anxiolytic effects, full knockout MAGL significantly reduced the duration in the light box in mice.<sup>48</sup> The study found that the CB1R receptor was significantly downregulated in MAGL knockout mice, possibly due to enhanced levels of innate endogenous cannabinoids leading to CB1R desensitization and the emergence of anxiety-like behavior. Consistent with these findings, CB1R density and functional responses were reduced in chronic MAGL inactivation and MAGL KO mice.<sup>49</sup> In conclusion, congenital and chronic MAGL inactivation may lead to CB1R desensitization and feedback down-regulation, which may inhibit the downstream anti-anxiety effect of CB1R, but more in-depth studies are needed.

## Anxiolytic Mechanisms of MAGL Inhibitors

Although preclinical studies support the anxiolytic effects of MAGL inhibitors, their specific downstream molecular mechanisms remain poorly understood. As shown in Figure 2, we summarized the potential mechanisms of MAGL from the following three aspects based on the above pharmacological studies.



**Figure 2** Schematic representation of the mechanism of anxiolytic action mediated by MAGL inhibitors. Briefly, the anxiolytic effects of MAGL are related to its maintenance of Glutamate/GABA balance, inhibition of neuroinflammation, and regulation of corticosterone levels.

**Abbreviations:** MAGL, monoacylglycerol lipase; CB1R, cannabinoid receptor 1; 2-AG, 2-arachidonoylglycerol; AMY, amygdala; PFC, prefrontal cortex; HPC, hippocampus; MSDB, medial septum and nucleus of the diagonal band; MHb, medial habenula; sEPSC, spontaneous excitatory postsynaptic currents; CB2R, cannabinoid receptor 2; COX-2, cyclooxygenase-2; AA, arachidonic acid; PGs, prostaglandins; LTs, leukotrienes; PG-G, prostaglandin glycerol; PGD2-G, prostaglandin D2-glyceryl ester; CRH, corticotropin-releasing hormone; ACTH, adrenocorticotropic hormone; CORT, corticosterone.

### Maintenance of Glutamate/GABA Balance

Glutamate and GABA are representative excitatory and inhibitory transmitters in the central nervous system, which cause neurons to generate corresponding excitatory and inhibitory currents through inter-synaptic transmitter transmission. Typically, neuronal excitation/inhibition (E/I) is in a dynamic balance. Once this balance is disturbed, especially when it tends to be excitatory, it can lead to the development of anxiety disorders. The E/I balance in the PFC, a brain region responsible for executive function, stress and emotion regulation, seems to play an important role in the anxiolytic effects of MAGL inhibitors.<sup>50</sup> PFC projection neurons are involved in the development of anxiety by modulating neural signaling in downstream brain regions during exposure to stress.<sup>51</sup> Systemic administration of the MAGL inhibitor, KML29, increased vmPFC excitability but could be blocked by CB1R or GABA receptor antagonist, which supports the conclusion that activation of the CB1R leads to transient inhibition of GABA release and long-term inhibition of inhibitory transmission.<sup>43</sup> Consistent with this finding, the CB1R agonist 5F-AMB attenuates glutamatergic and GABAergic synaptic transmission in mPFC L5 pyramidal neurons, leading to an E/I imbalance.<sup>52</sup>

Meanwhile, the amygdala, an essential part of the limbic system that controls emotion, plays a crucial role in the regulation of anxiety, and maintaining its E/I balance is of great significance.<sup>53</sup> MAGL inhibitors inhibit Glutamatergic neurotransmission in the amygdala region. It was found that increased expression of multiple glutamate receptors in the amygdala, including mGluR1, mGluR5, and NMDAR1, was accompanied by elevated expression of MAGL in an alcohol exposure-induced anxiety model,<sup>30</sup> and that JZL184 could exert anxiolytic effects by indirectly activating the CB1R to reduce NMDAR2B expression.<sup>54</sup> Bedse et al found that acute stress increased the frequency of spontaneous

excitatory postsynaptic currents (sEPSC) in basolateral amygdala neurons and was positively correlated with anxiety-like behaviors, with the above reversed after administration with JZL184.<sup>38</sup>

Notably, 2-AG is recognized as a potent agonist of CB1R. CB1R is found in both glutamatergic and GABAergic neurons, but the relative abundance of glutamatergic and GABAergic neurons in various brain regions varies, potentially causing functional alterations in distinct brain regions. Lysine-specific demethylase 1 (LSD1) suppressed hippocampal MAGL transcript levels and activated CB1R to inhibit glutamate release in response to anxiety.<sup>55</sup> JZL184 enhanced the inhibitory effect of 2-AG on the release of GABA from the medial septum and nucleus of the diagonal band (MSDB) axons to the medial habenula (MHb) and produced anxiolytic effects.<sup>56</sup>

### Inhibition of Neuroinflammation

Neuroinflammation is considered to be a trigger for behavioral changes and cognitive deficits in several psychiatric disorders, including anxiety disorders. Prolonged exposure to stress imbalances the central immune system and affects the secretion and function of immune cells and cytokines.<sup>57</sup> Currently, MAGL inhibitors play an important role in the treatment of central and peripheral inflammation.<sup>58,59</sup> The study showed that JZL184 reduced LPS-induced expression of IL-1 $\beta$ , IL-6, TNF- $\alpha$ , and IL-10 in the prefrontal cortex and spleen.<sup>60</sup>

Dual metabolism of 2-AG is closely relevant to the anti-inflammatory effects exerted by MAGL inhibitors. Arachidonic acid (AA), the main metabolite of 2-AG hydrolysis by MAGL, is a crucial precursor of proinflammatory prostaglandins and leukotrienes involved in inflammatory responses and immune initiation. The latest single-cell sequencing found that MAGL KO mice had significantly upregulated genes related to immunity and inflammation in microglia and astrocytes, which enable glial cells to react rapidly to insults.<sup>61</sup> Specifically knocking out MAGL in astrocytes reverses LPS-induced inflammatory activation and is not blocked by the CB1R agonist SR141716.<sup>62</sup> Similarly, CB1R/CB2R antagonists did not block JZL184 from inhibiting LPS-induced neuroinflammation.<sup>63</sup> This suggests that the anti-inflammatory effect produced by MAGL inhibition is a direct result of reduced prostaglandin, rather than a profile result of enhanced endocannabinoid signaling. In addition, enhanced inhibition of 2-AG hydrolysis can promote the oxidation pathway mediated by COX-2 to produce prostaglandin glycerides (PG-Gs), with PGD<sub>2</sub>-G exhibiting anti-inflammatory activation.<sup>64</sup>

CB2R, which can be fully activated by 2-AG, is mainly expressed in brain microglia and associated with neuroinflammation.<sup>65</sup> Many studies have confirmed the anxiolytic effect of CB2R agonists, and found that the use of CB2R agonist AM1241 inhibited the over-activation of PFC microglia by inhibiting the NLRP3 pathway, thereby improving anxiety-like behavior.<sup>66</sup> In addition, CB2R activation also promoted the transformation of microglia to M2 anti-inflammatory phenotype, creating positive feedback by releasing more 2-AG and AEA.<sup>67,68</sup> Thus, the effects of MAGL inhibitors in improving anxiety may be related to increasing the response of glial cells to external stimuli and promoting the polarization transformation of microglia.

### Regulation of Corticosterone Levels

The dysfunction of the hypothalamic-pituitary-adrenal (HPA) axis, the main regulator of stress response, is one of the biological mechanisms of anxiety. Increased corticosterone is an important marker of HPA axis activation. Studies have shown that ECS bi-directionally regulates the function of the HPA axis. MAGL inhibitors reduced high corticosterone levels in mice 30 min after chronic restraint stress but restored higher corticosterone levels after 120 min, whereas these changes were not observed in CB1R knockout mice.<sup>69</sup> The reason for these phenomena was the predominant inhibition of HPA axis activation during the early phase of JZL184 injection, whereas the sustained activation of CB1R elevated circulating corticosterone levels with increasing 2-AG concentrations. Aliczki et al found that metiramone, a cortisol synthesis blocker, reversed the increased open-arm time in the EPM test in mice with JZL184, suggesting that the anxiolytic effect of JZL184 is accompanied by activation of the HPA axis.<sup>70</sup> Glucocorticoids released after exposure to stress activate CB1R signaling in the mPFC, inhibit GABA release, and act on the HPA axis negative feedback process to appropriately reduce corticosterone secretion.<sup>71</sup> Therefore, MAGL inhibitors may exert anxiolytic effects through CB1R-dependent glucocorticoid increases.

## Novel MAGL Inhibitors and Natural Product Development

Given the significant pharmacological effects and higher safety of MAGL inhibitors, there is growing attention towards the development of novel inhibitors, with substantial efforts underway to identify safer, more effective, and reversible MAGL inhibitors. Research has primarily focused on developing novel inhibitors through structural modification of lead compounds or the use of fluorescent probes. Finding MAGL inhibitors from natural products has also been a strategic approach. Encouragingly, the majority of natural products reported to have MAGL inhibitory activity are reversible, enhancing drug safety with their gentle effects.

### Reversible MAGL Inhibitors

A total of 96 patents related to MAGL inhibitors were searched (<https://patentscope.wipo.int/>). Initially developed MAGL inhibitors were eliminated due to poor inhibition and lack of selectivity for MAGL. Later, Cravatt et al discovered JZL184, which was irreversible, inhibitory, and selective, was widely used in experimental studies. JZL184 is a potent tool compound, but it is difficult to use clinically due to the fact that it exhibits some cross-reactivity of FAAH at high doses (40 mg/kg) and some species differences in MAGL inhibitory potency.<sup>72</sup> Subsequently, KML29 was developed with better selectivity and good neuroprotection in a stroke model and good analgesic activity in a migraine model.<sup>73,74</sup>

However, JZL184, KML29, NAM, SAR629 and CK37 are all irreversible inhibitors of MAGL, which are prone to physical dependence, endocannabinoid dependent synaptic plasticity impairment and cross-tolerance to exogenous CB1 agonists.<sup>75</sup> JJKK048 is the first reported reversible, highly selective MAGL inhibitor with powerful analgesic effects and does not cause side effects similar to cannabis.<sup>76</sup> (R)-3t, a synthetic selective and reversible MAGL inhibitor, reduced arachidonic acid levels and increased 2-AG levels in the brain after gavage in mice.<sup>77</sup> Recently discovered M-18c, with an IC<sub>50</sub> value of 662.6 nM for MAGL, attenuates LPS-induced acute kidney injury by inhibiting NLRP3-associated inflammation.<sup>78</sup> Compared with irreversible inhibitors, reversible inhibitors have better pharmacokinetic advantages and offer substantial advantages in drug safety, tolerability and efficacy, with better development prospects and clinical applications.

### Structural Modifications Based on Lead Compounds

Virtual screening based on molecular docking was applied to discover novel reversible MAGL inhibitors. For example, Afzal O performed virtual docking of the ZINC database with 21 million compounds and screened seven potential activities, of which ZINC24092691 showed significant inhibitory activity.<sup>79</sup> DC630-8<sup>80</sup> and CL6a<sup>81</sup> also showed good inhibitory activity based on virtual docking. However, the screened compounds have poor drug-forming properties due to low screening efficacy. Therefore, many studies of structural modification based on lead compounds and functional group replacement have been added to the search for reversible inhibitors. Arylformylpiperidine derivatives developed by Zhi et al exhibit good reversible inhibitory properties and significantly ameliorate rifampicin-induced depressive-like behavior, providing support for MAGL as a potential therapeutic target for depression.<sup>82</sup> In addition, classical MAGL inhibitors such as CAY10499, JZL184, and ABX-1431 (also known as Lu AG06466) are classified as carbamate derivatives. Tiziano Tuccinardi's team discovered a series of new compounds with MAGL-inhibitory properties based on benzylpiperidine derivatives, such as compound 13 with antipancreatic cancer effects,<sup>83</sup> compounds 28 and 29 with low in vivo toxicity and high selectivity,<sup>84</sup> diphenylsulfide-benzoylpiperidine derivatives with anticancer activity.<sup>85</sup>

Looking for potential MAGL inhibitory activity in existing drugs is also a strategy. Disulfiram, an aldehyde dehydrogenase inhibitor used primarily in the treatment of chronic alcoholism, was shown to be a MAGL inhibitor that irreversibly inhibits MAGL through carbamoylation of Cys208 and Cys242 located near the MAGL active site.<sup>86</sup> However, due to the presence of some FAAH inhibitory activity, Omran synthesized compounds targeting MAGL but lacking anti-FAAH activity by replacing the two ethyl groups in the disulfide.<sup>87</sup> Recently, cetirizine and levetiracetam have also been found to have potential MAGL inhibitory activity, with IC<sub>50</sub> values of 9.3931 μM and 3.0095 μM, respectively, and demonstrated some analgesic and anti-inflammatory activity.<sup>88</sup>

Consideration of pharmacokinetic distribution is crucial in developing MAGL inhibitors. LEI-515, a recently discovered peripherally restricted reversible MAGL inhibitor, interestingly increased 2-AG levels only in peripheral organs but not in the mouse brain, hinting at potential applications in peripheral diseases.<sup>89</sup> Additionally, the compound properties can be improved by structural modification or application of nanocarriers if the pharmacokinetics are not ideal. Muhammad Adeel's team

developed the first nano-formulation of a MAGL inhibitor, MAGL23, which showed promising anti-tumour activity by using albumin-complexed nanocrystals that increased its solubility in water from less than 0.01 mg/mL to 0.82 mg/mL.<sup>90</sup>

### Probe Development

Developing MAGL inhibitors based on active molecular probes is also a new strategy. Activity-based protein profiling (ABPP) technology, which uses active site-directed covalent probe molecules to detect the functional state of enzyme activities in complex proteomes, has been applied to various enzyme classes.<sup>91</sup> Cisar et al identified and optimised a highly effective, selective and centrally permeable oral MAGL inhibitor, ABX-1431 (Lu AG06466), from a carbamate library by ABPP technology, which has entered Phase II clinical trials.<sup>92</sup> In addition, some newly screened compounds such as quinoid diterpene and  $\beta$ -carbolines,<sup>93</sup> as well as newly synthesised structural modifiers based on benzylpiperidine and benzylpiperazine,<sup>84</sup> were confirmed for their potential MAGL inhibitory activity by ABPP. The ABPP technique was also applied in discovering and mapping the distribution of eCB hydrolase activity, and MAGL enzyme activity was found to be strongest in the PFC region.<sup>94</sup> In conclusion, the ABPP has made it possible to visualize the spatio-temporal release of eCB hydrolases with high spatial resolution.

In recent years, many target-labeled radioactive probes have been developed based on MAGL inhibitors, some of which have even entered clinical trials. This technology can be used to image the in vivo distribution of MAGL and provide a method for subsequent disease diagnosis and treatment. PET imaging using 18F-T-401 was the first to image and quantify the distribution of MAGL in the human brain and found that MAGL was highest in the cerebral cortex, intermediate in the thalamus and nucleus accumbens, and lowest in the white matter and brainstem.<sup>95</sup> He et al developed a modified compound 7 based on morpholin-3-one derivatives, which may be a potential MAGL PET tracer, and successfully mapped the MAGL distribution pattern on rodent brain in vitro radioautography using the fixation method of direct <sup>11</sup>C<sub>2</sub> synthesis.<sup>96</sup> Based on a unique 4-piperidinylazetidide diamide scaffold, Cheng et al developed a reversible and peripherally specific radiolucinated MAGL PET ligand, [<sup>18</sup>F]FEPAD, which has excellent specificity and selectivity for MAGL in brown adipose tissue, a tissue known to be metabolically active.<sup>97</sup>

### Natural Product Development

Chemically synthesized MAGL inhibitors face challenges in clinical use due to their potent pharmacological effects and safety concerns. The current focus is shifting towards natural products and botanicals to discover milder, safer, and more effective reversible MAGL inhibitors. Pritimerin and euphol were the first identified natural products with reversible MAGL inhibitory activity.<sup>98</sup> Four triterpenoid constituents, including pritimerin and euphol, have been reported to significantly inhibit human recombinant MAGL activity, of which pritimerin ameliorated mechanical pain in mice with a concentration-dependent manner.<sup>99</sup> Protium copal, commonly used as incense by the Maya, displayed significant MAGL inhibitory activity, alleviating anxiety-like behaviors in rats, and this effect was blocked by a CB2R blocker.<sup>100</sup> The compound 8-prenylnaringenin in *Humulus lupulus L.* reversibly inhibited MAGL and reduces neuroinflammation, promising for Alzheimer's disease.<sup>101</sup> Extracts from *Myristica fragrans* exhibited anxiolytic and antidepressant effects, with significant MAGL inhibitory activity.<sup>102</sup>

Taking a computer-aided drug design (CADD) approach, combined with molecular docking, accelerates the discovery of MAGL inhibitors from natural products. Through the establishment of the pharmacophore model Phar-MAGL, combined with molecular docking and Ligplot analysis, NP-2/8-PN (IC<sub>50</sub> = 9.5 ± 1.2 μM), NP-5 (IC<sub>50</sub> = 14.5 ± 1.3 μM), and NP-3 (IC<sub>50</sub> = 15.2 ± 1.4 μM) were successfully screened for their promising in vitro inhibitory activities of MAGL.<sup>101</sup> Interestingly, 8-PN also had a positive metamorphic modulatory effect on GABAA that was not mediated through a high-affinity benzodiazepine binding site, and its potential anxiolytic effect could be further investigated.<sup>103</sup> Phenylethanoid glycosides from *C. phelypeae*<sup>104</sup> and Jewenol A from *S. pseudorosmarinus*<sup>105</sup> also demonstrated good MAGL inhibitory activity through enzymatic assays and molecular docking. Screening based on existing efficacy can enhance hit rates, as demonstrated in the study by Mei et al, who screened the MAGL inhibitory activity of 12 Chinese herbal medicines commonly used for analgesia, identifying *Corydalis yanhusuo* as the most effective.<sup>106</sup> Forsythiaside, a phenolic acid glycoside in *Forsythia suspensa*, inhibits COX-2 and MAGL, demonstrating neuroprotective effects in Alzheimer's disease by increasing hippocampal 2-AG content.<sup>107</sup> Table 2 summarizes some reported natural products with MAGL inhibiting activity.

**Table 2** Summary of MAGL Inhibitors Derived from Natural Products

Compounds	Category	Source	Pharmacological Effects	Experiment Type	References
Betulinic acid, Cucurbitacin B, Euphol, Pristimerin	Triterpenes	–	Analgesic	In vitro and ex vivo	[99]
Dehydrocorydaline	Alkaloid	<i>Corydalis yanhusuo</i>	Analgesic	In vitro or in vivo	[106]
8-prenylnaringenin	Flavonoid	<i>Humulus lupulus</i> L.	Promotion of neurogenesis and neurodifferentiation	In vitro	[101]
Lepidine B&E	Alkaloid	<i>Lepidium sativum</i> L. seeds	Inhibition of $\beta$ -amyloid production and accumulation	In vitro	[108]
Forsythiaside	Phenolic acid glycosides	<i>Forsythia suspensa</i>	Inhibition of $\beta$ -amyloid production and accumulation	Ex vivo	[107]
$\alpha$ -amyrins, $\beta$ -amyrins, Lupeol, Protium copal resin	Triterpenes	<i>Protium copal</i>	Analgesic, anti-inflammatory, anxiolytic	In vivo and in vitro	[100,109]
Jewenol A	Diterpenes	<i>S.pseudorosmarinus</i> aerial parts	Anticancer	In vitro	[105]
–	Phenylethanoid glycosides	<i>C. phelypaea</i> aerial parts	Anticancer	In vitro	[104]
–	–	<i>Myristica fragrans</i> methanol extracts	Anxiolytic, Antidepressant	In vitro	[102]

## AEA Hydrolase FAAH Inhibitors in Treatment for Anxiety

Human and rodent studies consistently show a negative correlation between anxiety levels and AEA concentrations.<sup>45,110</sup> Reduced AEA levels are strongly associated with PTSD severity, and moderate aerobic exercise has been found to increase AEA levels.<sup>111</sup> Stress-induced anxiety-like behaviors in rodents coincide with significant reductions in brain AEA levels, while microinjection of methanandamide (an AEA analogue) into the rat prefrontal cortex produces anxiolytic effects.<sup>112</sup> Alternatively, inhibition of AEA synthase promotes anxiety production. In NAPE-PLD-deficient mice, dorsal hippocampal AEA is significantly reduced and induces anxiety-like behavior.<sup>113</sup> FAAH is widely distributed in major neurons, including pyramidal cells in the BLA and hippocampus. Increased FAAH activity and decreased AEA levels have been found in mice subject to chronic restraint stress<sup>114</sup> and in Marchigian Sardinian alcohol-preferring rats with innate anxiety.<sup>115</sup> Therefore, enhancing AEA signaling by inhibiting FAAH activity is a potential strategy for the treatment of anxiety disorders.

## Anxiolytic Effects of FAAH Inhibitors

Several new compounds developed for FAAH inhibitors have entered clinical trials with promising prospects (<https://classic.clinicaltrials.gov/ct2/home>). JNJ-42165279 produces central and peripheral FAAH inhibition, significantly increases AEA levels in cerebrospinal fluid and plasma, and has not been found to have any safety concerns.<sup>116</sup> In the latest clinical pilot study, attenuation of amygdala, bilateral anterior cingulate gyrus, and bilateral insula activation during an emotional face processing task was found after 4 days of administration of JNJ-42165279 (100 mg) to 43 subjects, which is in line with the effects previously observed with anxiolytics.<sup>117</sup> Meanwhile, in another clinical study, JNJ-42165279 (25 mg/d) was effective in improving anxiety-like symptoms after 12 weeks of administration to patients with social anxiety disorder.<sup>13</sup> For PF-04457845, it was well tolerated,<sup>118</sup> attenuated anxiety effects in healthy subjects facing stress,<sup>119</sup> and weakened cannabis withdrawal symptoms.<sup>120</sup>

Several preclinical studies have shown that pharmacological inhibition of FAAH activity exhibited remarkable anxiolytic effects in different animal models of anxiety (Table 3). Meanwhile, genetic evidence also supports the above results. It was found that FAAH whole genome knockout C57BL/6J mice are not induced with anxiety-like behavior by chronic restraint stress.<sup>114</sup> In contrast, increased FAAH expression leads to the development of anxious behaviors in animals. Specific overexpression of FAAH in hippocampal glutamatergic neurons using AAV vectors significantly reduced AEA and PEA levels and increased anxiety-like behaviors, which may be related to enhanced LTP in glutamatergic neurons, leading to increased glutamate release.<sup>121</sup> Similarly, specific overexpression of FAAH in PFC significantly reduced AEA levels and had anxiogenic effects.<sup>112</sup>

**Table 3** Summary of the Anxiolytic Effects of FAAH Inhibitors in Preclinical Studies

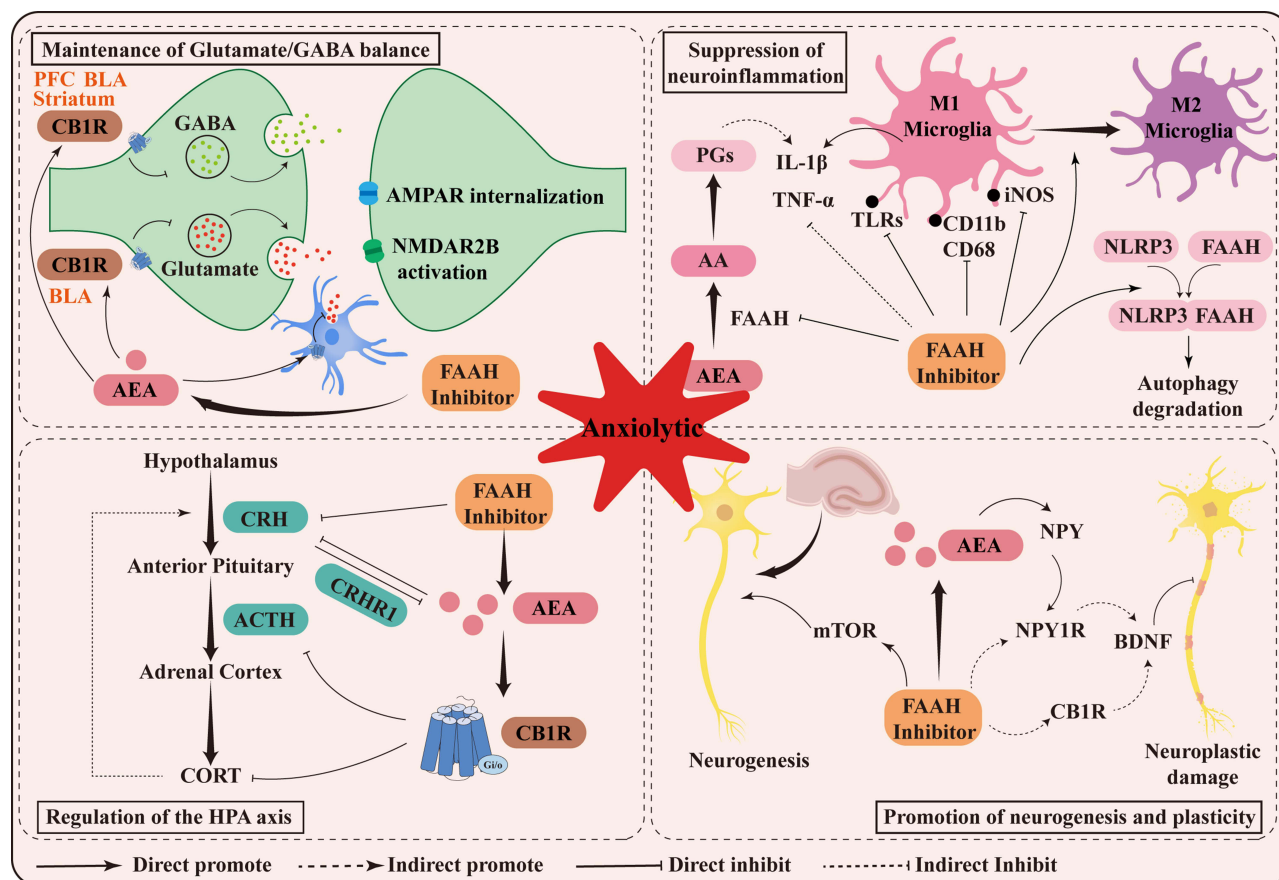
Drug	Dose/ Administration	Animal	Model	Test	Effects	References
URB597	0.1, 0.3, 1 mg/kg, i.p	C57BL/6J mice, male	SDS	NSF	↓ latency to feed	[122]
	0.3384 ng, 3.384 ng, 33.84 ng, intra-BNST microinjection	Wistar rats, male	ARS	EPM; CFC	No significance in EPM test; ↓ percentage of freezing behavior	[123]
	0.3 mg/kg, i.p	Sprague-Dawley rats, male	TMT exposure	EPM	↑ open arm time and entries, ↓ anxiety index	[124]
	3 mg/kg, i.p	Sprague-Dawley rats, female	Induced by poly I:C	OFT; EPM	↑ number of transition to inner arena; ↑ duration in open arm	[125]
	1 μg, intra-CeA microinjection	Wistar rats, male	CRS	EPM	↑ open arm time and entries	[115]
PF-3845	1 mg/kg, i.p	C57BL/6J mice, male	CUS	EPM; LDT	↑ duration and frequency in open arm; ↑ time and frequency in light compartment	[40]
	10 mg/kg, i.p	Wistar rats, male	Cafeteria diet exposure	OFT; EPM	↑ center zone entries and distance; ↑ open arm time and ↓ closed arm time	[126]
	1, 3, 10 mg/kg, i.p	C57BL/6J mice, male	–	EPM; LDT	↑ percent time in open-arm; ↑ percent light time	[35]
URB937	0.1, 1, 10 mg/kg, i.p	ICR mice, female	ARS and Foot-shock stress	LDT; NIH; EZM; OFT	↑ percent light time and distance, no significance in NIH test, EZM test and OFT	[37]
	10 mg/kg, i.p	ICR mice, male	Foot-shock stress	LDT	↑ percent light time and light entries	[127]
	1, 3 mg/kg, i.p	Wistar rats and Sprague-Dawley rats, male	SDS, TMT exposure	EPM; SAA	↑ open arm time and entries, ↓ closed arm time; ↓ time spent in non-social compartment and latency to access the social compartment, ↑ time spent in social compartment	[128]
PF-04457845	100 ng, 1 μg, icv	Sprague-Dawley rats, male	Colitis-induced anxiety	EPM	↑ open arm time	[129]
ST4070	3, 10, 30 mg/kg, p.o	CD1 mice, male	–	EPM	↑ open arm time	[130]
JNJ5003	10, 30 mg/kg, p.o	Wistar rats, male	–	LDT	↑ light time	
	50 mg/kg, p.o	C57BL/6J mice, male	CRS	EPM	↑ time spent in the open arm, ↓ latency to enter the open arm	[114]

**Abbreviations:** ARS, acute restraint stress; BNST, bed nucleus of the stria terminalis; CeA, central amygdala; CFC, contextual fear conditioning; CRS, chronic restraint stress, CUS-chronic unpredictable stress; EPM, elevated plus maze; EZM, Elevated zero maze; icv, intracerebroventricular; i.p, intraperitoneal; LDT, light-dark box test; NIH, novelty-induced hypophagia; NSF, novelty suppressed feeding; OFT, open-field test; SAA, social avoidance/approach; SDS, social defeat stress; TMT, 2,5-dihydro-2,4,5-trimethylthiazoline.

Single nucleotide polymorphism (SNP) in the FAAH gene, particularly the C385A (rs324420) allele, is noteworthy due to its significant association with FAAH expression. The polymorphism consists of the replacement of cytosine (C) by adenine (A) at nucleotide position 385, which translates into an amino acid exchange in which proline (Pro) replaces threonine (Thr) in codon 129. Mutations in the A allele result in decreased FAAH activity and increased AEA levels.<sup>131</sup> Healthy male adults carrying AC heterozygotes have higher levels of AEA and lower anxiety scores than CC homozygotes,<sup>132</sup> and carriers of the A allele have been found to have more robust neural network connectivity and lower anxiety levels in adolescents.<sup>133</sup> Similarly, knock-in of the FAAH C385A gene in mice significantly ameliorated anxiety-like behavior, which was associated with reduced FAAH expression, elevated AEA levels, and enhanced eCB signaling.<sup>134</sup> However, additional studies have shown that the FAAH C385A genetic variant in children increased the risk of anxiety.<sup>135</sup> Therefore, when FAAH inhibitor therapy is taken in the future, it may be necessary to consider the genotype of the patient's C385A to adopt a safer and more effective therapeutic strategy.

## Anxiolytic Mechanisms of FAAH Inhibitors

As shown in [Figure 3](#), FAAH inhibitors have partly the same anxiolytic mechanism as MAGL inhibitors due to similar functions. Except for maintaining Glutamate/GABA balance, suppressing neuroinflammation, and regulating the HPA



**Figure 3** Schematic representation of the mechanism of anxiolytic action mediated by FAAH inhibitors. Briefly, the anxiolytic effects of FAAH are related to its maintenance of Glutamate/GABA balance, suppression of neuroinflammation, regulation of HPA axis, and promotion of neurogenesis and plasticity.

**Abbreviations:** FAAH, fatty acid amide hydrolase; CB1R, cannabinoid receptor 1; AEA, anandamide; AMY, amygdala; PFC, prefrontal cortex; AA, arachidonic acid; PGs, prostaglandins; IL-1 $\beta$ , Interleukin-1 $\beta$ ; TNF- $\alpha$ , Tumor Necrosis Factor- $\alpha$ ; iNOS, inducible nitric oxide synthase; TLRs, Toll-like receptors; CRH, corticotropin-releasing hormone; ACTH, adrenocorticotropic hormone; CORT, corticosterone; corticotropin-releasing hormone receptor 1; NPY, neuropeptide Y; BDNF, brain-derived neurotrophic factor.

axis, the promotion of neurogenesis and plasticity has also been suggested to be related to the anxiolytic effects of FAAH inhibitors.

### Maintenance of Glutamate/GABA Balance

Similar to MAGL inhibitors, FAAH inhibitors also exert anxiolytic effects by regulating the E/I balance in various brain regions. As we mentioned, AEA selectively exhibits a high affinity for CB1R. CB1R exists in high density in the axonal terminals of Glutamatergic neurons and GABAergic neurons. When activated by AEA signals, G protein-mediated signal cascades are activated, thereby inhibiting the opening of voltage-gated calcium channels and membrane hyperpolarization caused by increased potassium channel opening.<sup>136</sup> Thus, FAAH inhibition restores the E/I balance and exerts anxiolytic effects primarily through CB1R alteration of brain Glutamatergic and GABAergic signaling.

Within the amygdala, constitutive signaling of AEA is present at the glutamatergic terminal CB1R, which limits excitatory transmission in the BLA and central amygdala (CeA).<sup>137</sup> Expression of FAAH in the postsynaptic terminal is higher in the BLA compared to the CeA.<sup>138</sup> The loss of AEA signaling in the amygdala leads to increased glutamate release, which increases the activity of postsynaptic output neurons, triggering anxious behaviors and stress responses.<sup>139</sup> Inhibition of FAAH prevented the rapid loss of AEA signalling caused by stress, which countered the effects of different types of stress, including elevated anxiety.<sup>124</sup> PF-04457845 similarly reversed the dysregulation of amygdala E/I balance in rats, reducing anxiety-like behaviors and increasing social behaviors.<sup>140,141</sup> Interestingly, in contrast to activating neuronal CB1R to inhibit glutamate release, AEA could activate astrocytic CB1R to promote glutamate release, leading

to NMDAR2B activation and AMPAR internalization, which induced long-term depression (LTD) and inhibited BLA pyramidal neurons to produce anxiolytic effects.<sup>142</sup> Therefore, the excitatory balance of BLA may depend on the level of glutamate release in neurons and astrocytes.

The striatum is also rich in CB1R and is involved in the regulation of anxiety. FAAH inhibition of accumulated AEA modulates excitatory and inhibitory neurotransmission in the striatum.<sup>143</sup> Social defeat stress induction decreased the sensitivity of striatal GABAergic synapses, but not glutamatergic, to CB1R activation and was restored by URB597.<sup>144</sup> URB597 also prevented quinolinic acid-induced neuroexcitotoxic damage and preserved striatal structural integrity.<sup>145</sup>

### Suppression of Neuroinflammation

FAAH inhibitors have also been reported to have robust anti-inflammatory effects. For example, URB937 reversed increased plasma inflammatory factor levels in social failure stress rats.<sup>128</sup> URB597 also reversed the expression of peripheral and cerebral proinflammatory cytokines under stress induced by LPS.<sup>146</sup> Studies have additionally shown that increased FAAH activity in LPS induced social behavioral deficits in adolescent rats and that imposition of PF04457845 reversed social behavioral changes.<sup>147</sup>

The central anti-inflammatory effects of FAAH inhibitors have been linked to the modulation of microglia polarisation phenotype.<sup>148</sup> AEA is an important signalling molecule in regulating microglia function, promoting their anti-inflammatory gene expression and inhibiting pro-inflammatory cytokine release.<sup>67,149</sup> The FAAH inhibitors PF3845 and URB597 both inhibited the production of prostaglandin E2 and pro-inflammatory gene expression in the BV2 microglial cell line, and the inhibitory effect of PF3845 was more pronounced.<sup>150</sup> URB597 improved the morphological characteristics of rat hippocampal microglia and promoted the transformation of microglia into an anti-inflammatory phenotype.<sup>151</sup> In addition, PF3845 also inhibits the expression of inducible nitric oxide synthase and COX-2 and promotes the shift of M1 pro-inflammatory phenotype to M2 anti-inflammatory phenotype.<sup>152</sup>

Modulation of Toll-like receptors (TLRs) may be another pathway. TLRs are intimately involved in the nervous system's immune response and are thought to underlie and exacerbate neurological disorders. Systemic administration of URB597 or PF3845 both inhibited TLR3/4-induced associated neuroinflammation in the PFC and hippocampus.<sup>153</sup> URB597 also reduced the TLR3-mediated increase in the expression of microglia/macrophage activation markers CD11b/CD68 and significantly alleviated anxiety-like behavior in rats.<sup>125</sup> Furthermore, recent studies revealed that FAAH has membrane anchoring and stabilising effects on NLRP3, and that URB597 and PF-04457845 inhibit NLRP3-FAAH interactions and induce autophagic NLRP3 degradation, suppressing the inflammatory phenotype.<sup>154</sup>

### Regulation of the HPA Axis

Patients with anxiety disorders were accompanied by high levels of adrenocorticotrophic hormone (ACTH), glucocorticoids in the blood, and overall hyperactivity of the HPA axis.<sup>155</sup> The level of AEA is negatively correlated with the activation of the HPA axis. It has been found that reduced homeostasis of AEA after repeated stressor exposure leads to basal overproduction of corticosterone.<sup>156</sup> The use of the hydrolase inhibitor URB937 reversed high plasma corticosterone levels 24 hours after social failure.<sup>128</sup>

The underlying mechanisms of FAAH inhibitors suppress HPA axis activation may be mediated by AEA/CB1R. In stressed rats, AEA levels in the amygdala were negatively correlated with serum corticosterone concentrations, and repeated corticosterone injections also resulted in a stress response. Meanwhile, injection of URB597 into the BLA reduced stress-induced corticosterone secretion, and this effect was blocked by the CB1R antagonist AM251.<sup>157</sup> The administration of AM251 to rats similarly increased their plasma concentrations of ACTH and CORT.<sup>158</sup>

Alternatively, corticotropin-releasing hormone (CRH) may be a candidate for linking the AEA to the HPA axis. The FAAH inhibitor URB597 reduced HPA axis hyperactivation and anxiety-like responses to stress<sup>159</sup> and dose-dependently down-regulated stress-induced CRH mRNA expression in the paraventricular nucleus of the hypothalamus.<sup>160</sup> CRH administration rapidly reduced AEA levels in the amygdala but not 2-AG, and induced anxiety-like behavior and HPA activation, both of which were reversed by URB597.<sup>161</sup> Interestingly, the increase in FAAH activity induced by the administration of CRH was not accompanied by an increase in FAAH protein levels or mRNA levels, which may be related to dynamic changes in enzyme activity or consistent with the specific coupling of the CRHR1 and FAAH regions.

Up-regulation of CRH was associated with CORT, with CRH mRNA significantly elevated in the rat PFC after CORT administration and dependent on CRHR1 signaling to regulate amygdala AEA content but not 2-AG.<sup>162</sup> Therefore, the relationship between CRH, AEA and HPA axis is not completely linear, indicating a potential crosstalk that requires further exploration.

### Promotion of Neurogenesis and Plasticity

Impaired neurogenesis as well as altered neuroplasticity are commonly seen in anxiety disorders caused by chronic stress. Studies have shown that neural progenitor cell division is affected by eCB signaling<sup>163</sup> and enhanced neurogenesis is always associated with elevated eCBs.<sup>164</sup> Repeated injections of cannabidiol, an exogenous cannabinoid, attenuated anxiety-like behavior by inhibiting FAAH activity and also promoted hippocampal neurogenesis and dendritic remodeling.<sup>165</sup> Moreover, HIV-1 Gp120-mediated impaired neurogenesis was rescued by FAAH gene deletion.<sup>166</sup> URB597 increased neuroplasticity by modulating long-term potentiation in the hippocampal CA1 region and the amygdala, attenuating fear memory.<sup>167</sup> JNJ5003 significantly reversed chronic restraint stress-induced dendritic expansion and increased spine density in BLA, promoting synaptic remodeling and reducing anxiety-like behavior.<sup>114,168</sup> URB532 and URB597 similarly prevented the reduction in AEA, the dendritic hypertrophy of the BLA, and the increase in anxiety-like behavior induced by stress.<sup>168</sup> The above studies illustrate that the use of FAAH inhibitors enhances brain neurogenesis as well as interneuronal transmission.

mTOR and neuropeptide Y (NPY) signaling may be associated with enhanced neurogenesis and neuroplasticity by FAAH inhibitors. mTOR signaling is essential for maintaining hippocampal neurogenesis and protecting against stress-induced impairment of neuroplasticity, and inhibition of this signaling increased anxiety-like behaviors in mice.<sup>169</sup> Recent studies have found that URB597 has an inverted U-shaped anxiolytic quantity-effect relationship in mice exposed to social defeat stress, and that its anxiolytic effect can be blocked by rapamycin (an mTOR antagonist), while its anxiogenic dose reduces the number of newborn neurons.<sup>122</sup> NPY is widely distributed in the nervous system and is particularly highly expressed in NAc and BLA.<sup>170</sup> Numerous studies have demonstrated that NPY levels are negatively correlated with anxiety,<sup>171</sup> which may be related to neuroprotective effects by promoting neurogenesis and neuroplasticity,<sup>172</sup> and decreasing amygdala excitability.<sup>173</sup> NPY has been reported to be involved in the neuroplastic protective effects of AEA, and inhibition of its expression antagonizes the effects of URB597 in suppressing PTSD behavior.<sup>174</sup>

### Novel FAAH Inhibitors and Natural Product Development

Unlike the urgent need to find reversible inhibitors of MAGL, irreversible FAAH inhibitors, such as URB597, PF3845, and PF04457845, do not exhibit significant toxic side effects and are well tolerated clinically. Therefore, the structural modification and optimization of FAAH inhibitors are more directed towards improving the drug-forming properties like solubility and central permeability for better bioavailability. ARN14633 and ARN14280 are novel analogues of URB597 with improved solubility and bioavailability effectively alleviating anxiety-like behavior in rats exposed to predator-evoked fear models.<sup>175,176</sup> Structural modifications based on lead compounds, as well as extractions from natural products, are equally pivotal in the development of novel FAAH inhibitors.

### Structural Modifications Based on Lead Compounds

In recent years, the development of novel FAAH inhibitors is mainly based on the skeletons of piperazine, isatin, oxazole and carbamate for structural modification and performance optimisation. Among them, piperazine-based FAAH inhibitors account for a relatively large proportion. Compound 4i (IC<sub>50</sub>= 0.12 μM) developed on the basis of indole-2-carbonyl piperazinourea derivatives possessed desirable antidepressant, analgesic and anti-inflammatory effects.<sup>177</sup> Among the piperazinourea derivatives with thiaziazole portion, the compounds with 4-chlorobenzyl (19) and 4-fluorobenzyl (20) tails on the piperazine side were found to be the most effective in inhibiting FAAH, with IC<sub>50</sub> of 0.13 and 0.22 μM, respectively.<sup>178</sup> Heteroaryl ureas with a thickened bicyclic diamine core exhibited better FAAH inhibitory activity compared to compounds constructed with a piperazine core.<sup>179</sup>

Jaiswal et al designed a series of isatin derivatives using the contemporary scaffold hopping approach in which compound 8c possessed antidepressant and anxiolytic effects without any neurotoxicity<sup>180</sup> and used the Dihydroindole-

2,3-dione derivatives as lead compounds to further search for FAAH inhibitors with good pharmacological properties.<sup>181</sup> JZP327A ( $IC_{50} = 11$  nM), synthesized using 1,3,4-oxadiazol-2-ones as a scaffold, is a highly selective, slowly reversible FAAH inhibitor<sup>182</sup> with good analgesic effects.<sup>183</sup> FAAH inhibitors developed on the basis of carbamates also exhibit good selectivity, reversibility, water solubility and some neuroprotective effects.<sup>184</sup>

Three-dimensional quantitative structure-activity relationship (3D-QSAR) model, which can better represent the structure-activity relationship of ligand-enzyme interaction and help to develop more effective compounds, has been widely used in FAAH inhibitor development. Zięba et al constructed two 3D-QSAR models based on 31 FAAH inhibitors containing the 1,3,4-oxadiazol-2-one structure, which contribute to the design of novel, more potent, and more indicative FAAH inhibitors.<sup>185</sup> Lorca et al also constructed a similar model based on piperazine-carboxamide scaffold and designed 10 new compounds with highly predicted FAAH inhibitory activity.<sup>186</sup> Application of QSAR model and molecular docking technology to screen potential FAAH inhibitors in current clinical drugs also deserves attention. Montelukast, Repaglinide, Refenacin, Raloxifene and Buclizine are considered to have potential FAAH inhibitory activity, but further in vivo and in vitro validation is required.<sup>187</sup>

### Dual-Target FAAH Inhibitors

Dual-target FAAH inhibition increases disease specificity. For example, Ibu-AM68<sup>188</sup> and Flu-AM4<sup>189</sup> both have dual FAAH and COX-2 inhibition, which circumvents the gastrointestinal response to NSAIDs, and have anti-inflammatory or analgesic effects. Development of dual FAAH/ChE inhibitors targeting the neuroprotective effects of FAAH inhibitors as promising candidates for the treatment of Alzheimer's disease.<sup>190,191</sup> Dual FAAH/sEH inhibitors with the piperidinyl-sulfonamide portion as the pharmacophore have been shown to have good inhibitory effects on neuropathic pain and inflammation.<sup>192</sup> In addition to dual inhibition, inhibition/excitation can also exist simultaneously. UCM1341 inhibits FAAH while activating melatonin receptors, producing anti-inflammatory effects to provide neuroprotection.<sup>193</sup> Compounds designed to target FAAH inhibition/activation of CB2R also have potent neuroinflammatory inhibitory effects.<sup>194</sup>

Furthermore, FAAH inhibitors also have central and peripheral inhibition selectivity. For example, ASP3652, which entered clinical trials, is a well-tolerated peripheral reversible FAAH inhibitor that reduces lower urinary tract symptoms but no efficacy in the improvement of patients' pain symptoms, possibly related to its lack of central inhibition.<sup>195</sup> Surprisingly, URB937, a peripherally restricted FAAH inhibitor, was unable to cross the blood-brain barrier but also possessed central activities such as anxiolysis<sup>128</sup> and analgesia.<sup>196</sup> This result suggests the presence of peripheral and central crosstalk, with anxiolysis possibly related to sympathetic efferents and analgesia possibly related to reduced afferents for injurious pain, requiring further experimental verification.

### Off-Target Effects of FAAH Inhibitors

However, although FAAH inhibitors show relatively excellent pharmacological activity, their off-target effects should not be overlooked. FAAH is a serine hydrolase that uses highly conserved serine residues in its active site as nucleophilic reagents to catalyse the hydrolysis of its substrate. Most FAAH inhibitors exert their inhibitory effects by binding and modifying catalytic serine residues, which can also inhibit other serine hydrolases.<sup>197</sup> For example, URB597, BMS-1, OL-135 and LY2077855 all have low selectivity and show a variety of off-target effects, the main off-target being carboxylesterases.<sup>197</sup> Unfortunately, the off-target effects of FAAH inhibitors have been disregarded and even carried over into clinical trials, resulting in a tragic lesson. BIA 10-2474, an irreversible FAAH inhibitor entered Phase I clinical trials in 2016, was urgently called off due to resulting in the death of one volunteer and mild to severe neurological symptoms in four volunteers.<sup>198</sup> Subsequent studies showed that the cause of the clinical incident of BIA 10-2474 may be that its off-target protein PNPLA6 is strongly associated with organophosphorus neurotoxicity.<sup>199</sup> Another irreversible FAAH inhibitor, PF04457845, has entered a Phase 2 trial with no serious adverse events.<sup>120</sup> Therefore, the off-target effects must be evaluated to ensure clinical safety when developing novel FAAH inhibitors.

As mentioned above, ABPP technology can also be used to screen FAAH enzyme inhibitors. Otrubova et al assessed the FAAH inhibition performance of a series of N-acyl pyrazole derivatives by ABPP, minimizing off-target activity.<sup>200</sup> Lamani et al demonstrated that FAAH inhibitors 9 and 31 were highly selective for brain FAAH and protective against

kainic acid-induced excitotoxicity by ABPP method.<sup>201</sup> JZP-327A, a slowly reversible FAAH inhibitor with over 900-fold selectivity for MAGL and COX isozymes, was shown by ABPP to have higher FAAH selectivity compared to other serine hydrolases.<sup>182</sup> Generally, evaluating the potential pharmacological activity of novel inhibitors by ABPP technology enables timely circumvention of off-target effects and minimization of compound side effects.

## Natural Product Development

Various active ingredients with FAAH inhibitory activity were also found in natural products (Table 4). Among them, flavonoids inhibited the activity of FAAH significantly. Daidzein, silybin and chickpea bractein A showed high FAAH inhibitory activity both in vivo and in vitro.<sup>202</sup> Kaempferol inhibited FAAH activity in a concentration-dependent manner in vitro, while in vivo experiments further confirmed that acting on FAAH reduced anxiety-like behavior.<sup>203</sup> In addition, isoflavonoids are thought to target the ECS by modulating eCB metabolism. Three isoflavonoids, 7-hydroxyflavone, biochanin-A, and genistein, all bind to the FAAH active site and dose-dependently inhibit FAAH activity, increasing AEA levels in the PFC and decreasing blood corticosterone concentrations.<sup>204</sup>

Terpenoids and phenolic compounds also showed positive FAAH inhibitory activity. All 17 triterpenoids isolated from *Ganoderma lucidum* exhibited some inhibitory activity against FAAH without cytotoxicity, and FAAH may be a potential target for anti-neuroinflammation.<sup>209,220</sup>  $\beta$ -stigmaterol and eugenol extracted from *Harpagophytum procumbens*, a sesquiterpene and monosterane constituent, respectively, exerted an anti-arthritis effects by inhibiting FAAH expression,<sup>214</sup> with  $\beta$ -stigmaterol also being a selective CB2R agonist.<sup>221,222</sup> Some phenolic compounds, such as cannabidiol extracted from *Cannabis sativa* L. and 5'-methoxylicarin A extracted from *Myristica fragrans* Houtt., have also been reported to inhibit FAAH activity and have anxiolytic efficacy.<sup>216</sup>

Some other types of natural products also have FAAH inhibition effects. Macamides, a unique series of non-polar long-chain fatty acids N-benzamide isolated from Maca, are mainly alkaloids with neuroprotective properties,<sup>223</sup> of which N-Benzyl octadeca-9Z,12Z-dienamide exhibits the best FAAH inhibitory activity and attenuates ischaemic stroke injury,<sup>224</sup> but this inhibition is an irreversible inhibitor that exhibits time-dependent inhibition.<sup>205</sup> Lavender essential oil, an over-the-counter herbal medicine approved by the European Medicines Agency for the relief of anxiety, displayed

**Table 4** Summary of FAAH Inhibitors Derived from Natural Products

Compounds	Category	Source	Pharmacological Effects	Experiment Type	References
Macamides	Alkaloid	<i>Lepidium meyenii</i> Walp	Relief of ischemic injury, analgesic, anti-inflammatory, neuroprotective	In vitro	[205, 206]
Silybin, isosilybin	Flavonolignans	<i>Silybum marianum</i>	Alleviate peripheral neuropathy	In vitro	[207]
–	–	Ashwagandha	Antioxidant	In vivo	[208]
–	Lanostane triterpenoids	<i>Ganoderma lucidum</i>	Suppress neuroinflammation	In vitro	[209, 210]
7-Hydroxyflavone, biochanin A, genistein	Isoflavone	<i>Glycine max</i> Merrill. germ, <i>Trifolium pratense</i> L.	Antidepressant, analgesic	In vivo and in vitro	[204, 211]
Isorhamnetin, kaempferol, quercetin	Flavonoid	<i>Moricandia sinaica</i> aerial parts	Analgesic, antipyretic, anti-inflammatory	In vivo and in vitro	[212, 213]
Kaempferol	Flavonoid	–	Anxiolytic	In vivo and in vitro	[203]
Eugenol, $\beta$ -caryophyllene	Terpenoid	<i>Harpagophytum procumbens</i> root	Anti-arthritis	In vitro	[214]
Citral	Monoterpene	<i>Cymbopogon citratus</i>	Anti-inflammatory, analgesic	In vivo and in vitro	[215]
Licarin A, 5'-methoxylicarin A, malabaricone C	Phenol	<i>Myristica fragrans</i> Houtt.	Anxiolytic	In vivo and in vitro	[216]
Linalool, Linalyl acetate	Essential oil	<i>Lavandula angustifolia</i> Mill.	Relief of neuropathic pain, anxiolytic, antidepressant	In vivo and in vitro	[217]
Cannabidiol, tetrahydrocannabinol, cannabigerol	Phenol	<i>Cannabis sativa</i> L.	Antioxidant, inhibition of inflammatory bowel disease-associated hypermobility, anxiolytic	In vivo and in vitro	[218, 219]

FAAH inhibitory activity in vitro and produced effects comparable to diazepam in mice.<sup>217</sup> In addition, Xiaoyao Pills, a proprietary Chinese medicine widely used in China for the treatment of depression, was shown to alleviate depression-like behavior in rats by inhibiting FAAH levels in the brain.<sup>225</sup>

Techniques such as fluorescent probes and molecular docking have been used in recent years to screen potential FAAH inhibitors from natural products. Ginkgolide, the main active ingredient in *Ginkgo biloba*, was confirmed to bind well to FAAH by molecular docking.<sup>226</sup> The aqueous extracts and fatty oils of *Platycladi Semen* are also predicted to exert anxiolytic effects through FAAH.<sup>227</sup> The FAAH-activated fluorescent probe named THPO developed by Tian et al identified a natural inhibitor, neobavaisoflavone, from 68 traditional herbs.<sup>228</sup> Similarly, DAND, a FAAH-activated near-infrared fluorescent probe, screened piperine as a novel inhibitor of FAAH and presented excellent anti-inflammatory activity in the vitro experiments.<sup>229</sup>

## Summary and Prospect

The high prevalence of anxiety disorders and socio-economic burden highlights the pressing need for effective treatments, as current options remain limited. Although named with a plant possessing psychoactive properties, ECS is actually one of the crucial regulatory systems of the organism. In recent years, the idea of targeting the ECS for the treatment of anxiety disorders has received increasing attention. Inhibition of hydrolase activity, particularly targeting the dominant enzymes MAGL and FAAH, offers a promising avenue by indirectly increasing eCB levels, thereby avoiding the addictive properties and adverse effects linked to direct exogenous cannabinoid supplementation.

The anxiolytic efficacy of FAAH and MAGL inhibitors has been supported by both clinical and preclinical studies, indicating their potential as promising treatments for anxiety disorders. The pharmacological mechanisms of these inhibitors are primarily associated with maintaining Glutamate/GABA balance, suppressing neuroinflammation, modulating the HPA axis, and promoting neurogenesis. Given the crucial role of neuronal excitatory/inhibitory (E/I) imbalance in the pathology of anxiety disorders, the maintenance of Glutamate/GABA homeostasis is particularly vital. CB1R, the primary mediator of the biological effects of 2-AG and AEA, is extensively expressed in glutamatergic and GABAergic terminals, offering the potential to reverse this imbalance. The clarification of the above mechanisms is not only crucial for drug development and clinical application of endocannabinoid hydrolase inhibitors, but also aids in achieving precision treatment for anxiety disorders. Similarly, more research is necessary to support or further clarify the precise mechanisms of FAAH and MAGL inhibitors in treating anxiety disorders.

Numerous biochemical techniques, including ABPP, fluorescence probes, molecular docking, and 3D-QSAR, have been used to generate novel MAGL and FAAH inhibitors. These techniques have greatly improved the screening efficiency and facilitated the discovery of new compounds. Notably, most of these new compounds are mainly developed for anticancer, analgesic, and anti-inflammatory purposes, and their potential anxiolytic activity remains to be further evaluated. Meanwhile, while significantly reducing the adverse effects associated with direct supplementation of cannabinoids or activation of cannabinoid receptors, the potential cardiotoxicity of some MAGL inhibitors and off-target effects of FAAH inhibitors should not be overlooked. In addition, given the polymorphism in the FAAH C385A gene, the genotype of patients must be thoroughly considered when applying FAAH inhibitors for treatment to achieve precision medicine. In contrast, some natural products isolated from plants have been shown to exert anxiolytic activity by inhibiting FAAH or MAGL, mostly reversibly, which greatly improves the tolerability and safety of the drugs, and thus active compounds derived from natural products would be a good source of novel anxiolytic drugs.

The development of dual MAGL/FAAH inhibitors has also come into the limelight. The newly developed AKU-005 significantly enhanced 2-AG and AEA levels and inhibited neural excitability in rat and human meninges, which is expected to be a new treatment for migraine.<sup>230</sup> JZL195, a classical dual MAGL/FAAH inhibitor, embodies an antidepressant activity,<sup>231</sup> potent neuroleptic activity,<sup>232</sup> weak antihypertensive effect<sup>233</sup> and anti-inflammatory activity.<sup>234</sup> Regrettably, the anxiolytic effects of JZL195 remain a subject of controversy. Some studies suggested an improvement in anxiety-like behavior in EPM test in mice.<sup>35</sup> On the contrary, other research have shown that it fails to reverse restraint stress-induced anxiety-like behaviors or even promotes anxiety-like behaviors.<sup>37,235</sup> These discrepancies may stem from variations in experimental conditions and animal models. However, it's crucial to acknowledge that the potential anxiolytic effects of MAGL/FAAH dual inhibition cannot be conclusively denied.

Furthermore, consistent with the inhibition of hydrolase activity, the levels of synaptic interstitial 2-AG and AEA can also be increased by inhibiting the activity of eCB transporter proteins, and intracellular transport is necessary for eCB hydrolysis. Current studies focus on AEA transporter proteins, which mainly include FABP5, HSP70, and FLAT1.<sup>236,237</sup> Recent studies have found that the use of a FABP5 inhibitor, SBFI-103, in either the amygdala or the PFC produces significant anxiolytic effects.<sup>238</sup> WOBE437, a natural product-derived inhibitor of AEA reuptake, irreversibly blocks 2-AG, AEA membrane transport, producing anxiolytic effects.<sup>239</sup> Similarly, AM404, also an AEA reuptake inhibitor, has been reported to exhibit reliable anxiolytic activity.<sup>240,241</sup> Although the above studies demonstrated the anxiolytic promise of targeted transporter proteins, it should not be neglected that they are still in their infancy, and need to be confirmed by further experimental studies.

In summary, there exists great potential to develop a clinically effective, safe, and well-tolerated novel anxiolytic drug from MAGL and FAAH inhibitors compared to direct cannabinoid receptor agonists. This can be achieved by screening and synthesizing new hydrolase inhibitors using advanced technologies, or by exploring active ingredients with robust inhibitory activity from natural products. Structural optimization and modification may further enhance the selectivity and biological activity of these potential drugs.

## Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

## Funding

This work was supported by National Natural Science Foundation of China (No. 82104836), Natural Science Foundation of Hunan Province (No. 2023JJ60482), Scientific Research Project of Hunan Provincial Department of Education (No. 23B0391), and Science & Technology talent promotion Project of Hunan Province (No. 2023TJ-N22).

## Disclosure

The authors declare no conflicts of interest in this work.

## References

1. American Psychiatric Association. *Diagnostic and Statistical Manual of Mental Disorders (DSM-5®)*. American Psychiatric Publisher; 2013.
2. Szuhany KL, Simon NM. Anxiety disorders: a review. *JAMA*. 2022;328(24):2431–2445. doi:10.1001/jama.2022.22744
3. Huang Y, Wang Y, Wang H, et al. Prevalence of mental disorders in China: a cross-sectional epidemiological study. *Lancet Psychiatry*. 2019;6(3):211–224. doi:10.1016/S2215-0366(18)30511-X
4. Inserra A, Piot A, De Gregorio D, Gobbi G. Lysergic Acid Diethylamide (LSD) for the treatment of anxiety disorders: preclinical and clinical evidence. *CNS Drugs*. 2023;37(9):733–754. doi:10.1007/s40263-023-01008-5
5. Skolnick P. Anxiolytic: on a quest for the Holy Grail. *Trends Pharmacol Sci*. 2012;33(11):611–620. doi:10.1016/j.tips.2012.08.003
6. Gosmann NP, de Abreu Costa M, de Barros Jaeger M, et al. Selective serotonin reuptake inhibitors, and serotonin and norepinephrine reuptake inhibitors for anxiety, obsessive-compulsive, and stress disorders: a 3-level network meta-analysis. *PLoS Med*. 2021;18(6):e1003664. doi:10.1371/journal.pmed.1003664
7. Murphy SE, Capitão LP, Giles SLC, Cowen PJ, Stringaris A, Harmer CJ. The knowns and unknowns of SSRI treatment in young people with depression and anxiety: efficacy, predictors, and mechanisms of action. *Lancet Psychiatry*. 2021;8(9):824–835. doi:10.1016/S2215-0366(21)00154-1
8. Gosmann NP, de Abreu Costa M, de Barros Jaeger M, et al. Incidence of adverse events and comparative tolerability of selective serotonin reuptake inhibitors, and serotonin and norepinephrine reuptake inhibitors for the treatment of anxiety, obsessive-compulsive, and stress disorders: a systematic review and network meta-analysis. *Psychol Med*. 2023;53(9):3783–3792. doi:10.1017/S0033291723001630
9. Joffe H, Petrillo L, Viguera A, et al. Eszopiclone improves insomnia and depressive and anxious symptoms in perimenopausal and postmenopausal women with hot flashes: a randomized, double-blinded, placebo-controlled crossover trial. *Am J Obstet Gynecol*. 2010;202(2):171.e1–171.e11. doi:10.1016/j.ajog.2009.10.868
10. Mokhber N, Azarpazhooh MR, Khajehdaloue M, Velayati A, Hopwood M. Randomized, single-blind, trial of sertraline and buspirone for treatment of elderly patients with generalized anxiety disorder. *Psychiatry Clin Neurosci*. 2010;64(2):128–133. doi:10.1111/j.1440-1819.2009.02055.x
11. Lowe H, Toyang N, Steele B, Bryant J, Ngwa W. The endocannabinoid system: a potential target for the treatment of various diseases. *Int J Mol Sci*. 2021;22(17):9472. doi:10.3390/ijms22179472

12. Alcaraz-Silva J, Feingold D, Viana-Torre G, et al. The endocannabinoid system as a biomarker for diagnostic and therapeutic applications in depression and anxiety. *CNS Neurol Disord Drug Targets*. 2023;22(3):417–430. doi:10.2174/1871527321666220405114402
13. Schmidt ME, Liebowitz MR, Stein MB, et al. The effects of inhibition of fatty acid amide hydrolase (FAAH) by JNJ-42165279 in social anxiety disorder: a double-blind, randomized, placebo-controlled proof-of-concept study. *Neuropsychopharmacology*. 2021;46(5):1004–1010. doi:10.1038/s41386-020-00888-1
14. Cristino L, Bisogno T, Di Marzo V. Cannabinoids and the expanded endocannabinoid system in neurological disorders. *Nat Rev Neurol*. 2020;16(1):9–29. doi:10.1038/s41582-019-0284-z
15. Murkar A, De Koninck J, Merali Z. Cannabinoids: revealing their complexity and role in central networks of fear and anxiety. *Neurosci Biobehav Rev*. 2021;131:30–46. doi:10.1016/j.neubiorev.2021.09.002
16. Kaczocha M, Haj-Dahmane S. Mechanisms of endocannabinoid transport in the brain. *Br J Pharmacol*. 2022;179(17):4300–4310. doi:10.1111/bph.15469
17. Petrie GN, Nastase AS, Aukema RJ, Hill MN. Endocannabinoids, cannabinoids and the regulation of anxiety. *Neuropharmacology*. 2021;195:108626. doi:10.1016/j.neuropharm.2021.108626
18. Morgan AJ, Kingsley PJ, Mitchener MM, et al. Detection of cyclooxygenase-2-derived oxygenation products of the endogenous cannabinoid 2-arachidonoylglycerol in mouse brain. *ACS Chem Neurosci*. 2018;9(7):1552–1559. doi:10.1021/acscchemneuro.7b00499
19. Patel S, Hill MN, Cheer JF, Wotjak CT, Holmes A. The endocannabinoid system as a target for novel anxiolytic drugs. *Neurosci Biobehav Rev*. 2017;76(Pt A):56–66. doi:10.1016/j.neubiorev.2016.12.033
20. Maldonado R, Cabañero D, Martín-García E. The endocannabinoid system in modulating fear, anxiety, and stress. *Dialogues Clin Neurosci*. 2020;22(3):229–239. doi:10.31887/DCNS.2020.22.3/maldonado
21. Lama A, Pirozzi C, Severi I, et al. Palmitoylethanolamide dampens neuroinflammation and anxiety-like behavior in obese mice. *Brain Behav Immun*. 2022;102:110–123. doi:10.1016/j.bbi.2022.02.008
22. Fraguas-Sánchez AI, Torres-Suárez AI. Medical Use of Cannabinoids. *Drugs*. 2018;78(16):1665–1703. doi:10.1007/s40265-018-0996-1
23. Cohen K, Weizman A, Weinstein A. Positive and negative effects of cannabis and cannabinoids on health. *Clin Pharmacol Ther*. 2019;105(5):1139–1147. doi:10.1002/cpt.1381
24. Stasiulewicz A, Znajdek K, Grudzień M, Pawiński T, Sulkowska AJ. A Guide to Targeting the Endocannabinoid System in Drug Design. *Int J Mol Sci*. 2020;21(8):2778. doi:10.3390/ijms21082778
25. Gipson CD, Kupchik YM, Kalivas PW. Rapid, transient synaptic plasticity in addiction. *Neuropharmacology*. 2014;76(Pt B):276–286. doi:10.1016/j.neuropharm.2013.04.032
26. Hill MN, Bierer LM, Makotkine I, et al. Reductions in circulating endocannabinoid levels in individuals with post-traumatic stress disorder following exposure to the World Trade Center attacks. *Psychoneuroendocrinology*. 2013;38(12):2952–2961. doi:10.1016/j.psyneuen.2013.08.004
27. Siebers M, Biedermann SV, Bindila L, Lutz B, Fuss J. Exercise-induced euphoria and anxiolysis do not depend on endogenous opioids in humans. *Psychoneuroendocrinology*. 2021;126:105173. doi:10.1016/j.psyneuen.2021.105173
28. Shonesy BC, Bluett RJ, Ramikie TS, et al. Genetic disruption of 2-arachidonoylglycerol synthesis reveals a key role for endocannabinoid signaling in anxiety modulation. *Cell Rep*. 2014;9(5):1644–1653. doi:10.1016/j.celrep.2014.11.001
29. Jenniches I, Ternes S, Albayram O, et al. Anxiety, stress, and fear response in mice with reduced endocannabinoid levels. *Biol Psychiatry*. 2016;79(10):858–868. doi:10.1016/j.biopsych.2015.03.033
30. Sánchez-Marín L, Flores-López M, Pastor A, et al. Acute stress and alcohol exposure during adolescence result in an anxious phenotype in adulthood: role of altered glutamate/endocannabinoid transmission mechanisms. *Prog Neuropsychopharmacol Biol Psychiatry*. 2022;113:110460. doi:10.1016/j.pnpbp.2021.110460
31. Hill MN, Haney M, Hillard CJ, Karhson DS, Vecchiarelli HA. The endocannabinoid system as a putative target for the development of novel drugs for the treatment of psychiatric illnesses. *Psychol Med*. 2023;53(15):7006–7024. doi:10.1017/S0033291723002465
32. Marcus DJ, Bedse G, Gauden AD, et al. Endocannabinoid signaling collapse mediates stress-induced amygdalo-cortical strengthening. *Neuron*. 2020;105(6):1062–1076.e6. doi:10.1016/j.neuron.2019.12.024
33. Guggenhuber S, Romo-Parra H, Bindila L, et al. Impaired 2-AG signaling in hippocampal glutamatergic neurons: aggravation of anxiety-like behavior and unaltered seizure susceptibility. *Int J Neuropsychopharmacol*. 2015;19(2):pyv091. doi:10.1093/ijnp/pyv091
34. Serrano A, Pavon FJ, Buczynski MW, et al. Deficient endocannabinoid signaling in the central amygdala contributes to alcohol dependence-related anxiety-like behavior and excessive alcohol intake. *Neuropsychopharmacology*. 2018;43(9):1840–1850. doi:10.1038/s41386-018-0055-3
35. Pavón FJ, Polis IY, Stouffer DG, et al. Selective inhibition of monoacylglycerol lipase is associated with passive coping behavior and attenuation of stress-induced dopamine release in the medial prefrontal cortex. *Neurobiol Stress*. 2021;14:100293. doi:10.1016/j.ynstr.2021.100293
36. Alteba S, Mizrachi Zer-Aviv T, Tenenhaus A, et al. Antidepressant-like effects of URB597 and JZL184 in male and female rats exposed to early life stress. *Eur Neuropsychopharmacol*. 2020;39:70–86. doi:10.1016/j.euroneuro.2020.08.005
37. Bedse G, Bluett RJ, Patrick TA, et al. Therapeutic endocannabinoid augmentation for mood and anxiety disorders: comparative profiling of FAAH, MAGL and dual inhibitors. *Transl Psychiatry*. 2018;8(1):92. doi:10.1038/s41398-018-0141-7
38. Bedse G, Hartley ND, Neale E, et al. Functional redundancy between canonical endocannabinoid signaling systems in the modulation of anxiety. *Biol Psychiatry*. 2017;82(7):488–499. doi:10.1016/j.biopsych.2017.03.002
39. Bosch-Bouju C, Larrieu T, Linders L, Manzoni OJ, Layé S. Endocannabinoid-mediated plasticity in nucleus accumbens controls vulnerability to anxiety after social defeat stress. *Cell Rep*. 2016;16(5):1237–1242. doi:10.1016/j.celrep.2016.06.082
40. Lomazzo E, Bindila L, Remmers F, et al. Therapeutic potential of inhibitors of endocannabinoid degradation for the treatment of stress-related hyperalgesia in an animal model of chronic pain. *Neuropsychopharmacology*. 2015;40(2):488–501. doi:10.1038/npp.2014.198
41. Sumislawski JJ, Ramikie TS, Patel S. Reversible gating of endocannabinoid plasticity in the amygdala by chronic stress: a potential role for monoacylglycerol lipase inhibition in the prevention of stress-induced behavioral adaptation. *Neuropsychopharmacology*. 2011;36(13):2750–2761. doi:10.1038/npp.2011.166
42. Kinsey SG, O'Neal ST, Long JZ, Cravatt BF, Lichtman AH. Inhibition of endocannabinoid catabolic enzymes elicits anxiolytic-like effects in the marble burying assay. *Pharmacol Biochem Behav*. 2011;98(1):21–27. doi:10.1016/j.pbb.2010.12.002
43. Worley NB, Varela JA, Gaillardetz GP, Hill MN, Christianson JP. Monoacylglycerol lipase alpha inhibition alters prefrontal cortex excitability and blunts the consequences of traumatic stress in rat. *Neuropharmacology*. 2020;166:107964. doi:10.1016/j.neuropharm.2020.107964

44. Vozella V, Cruz B, Feldman HC, et al. Sexually dimorphic effects of monoacylglycerol lipase inhibitor MJN110 on stress-related behavior and drinking in Marchigian Sardinian alcohol-preferring rats. *Br J Pharmacol.* 2023;180(24):3130–3145. doi:10.1111/bph.16197
45. Dlugos A, Childs E, Stuhr KL, Hillard CJ, de Wit H. Acute stress increases circulating anandamide and other N-acyl ethanolamines in healthy humans. *Neuropsychopharmacology.* 2012;37(11):2416–2427. doi:10.1038/npp.2012.100
46. Zhai Q, Islam A, Chen B, et al. Endocannabinoid 2-arachidonoylglycerol levels in the anterior cingulate cortex, caudate putamen, nucleus accumbens, and piriform cortex were upregulated by chronic restraint stress. *Cells.* 2023;12(3):393. doi:10.3390/cells12030393
47. Di S, Itoga CA, Fisher MO, et al. Acute stress suppresses synaptic inhibition and increases anxiety via endocannabinoid release in the basolateral amygdala. *J Neurosci.* 2016;36(32):8461–8470. doi:10.1523/JNEUROSCI.2279-15.2016
48. Imperatore R, Morello G, Luongo L, et al. Genetic deletion of monoacylglycerol lipase leads to impaired cannabinoid receptor CB<sub>1</sub>R signaling and anxiety-like behavior. *J Neurochem.* 2015;135(4):799–813. doi:10.1111/jnc.13267
49. Schlosburg JE, Blankman JL, Long JZ, et al. Chronic monoacylglycerol lipase blockade causes functional antagonism of the endocannabinoid system. *Nat Neurosci.* 2010;13(9):1113–1119. doi:10.1038/nn.2616
50. Kenwood MM, Kalin NH, Barbas H. The prefrontal cortex, pathological anxiety, and anxiety disorders. *Neuropsychopharmacology.* 2022;47(1):260–275. doi:10.1038/s41386-021-01109-z
51. Liu WZ, Zhang WH, Zheng ZH, et al. Identification of a prefrontal cortex-to-amygdala pathway for chronic stress-induced anxiety. *Nat Commun.* 2020;11(1):2221. doi:10.1038/s41467-020-15920-7
52. Domoto M, Sasase H, Wada S, et al. The synthetic cannabinoid 5F-AMB changes the balance between excitation and inhibition of layer V pyramidal neurons in the mouse medial prefrontal cortex. *Psychopharmacology.* 2018;235(8):2367–2376. doi:10.1007/s00213-018-4933-5
53. Zhang WH, Zhang JY, Holmes A, Pan BX. Amygdala Circuit Substrates for Stress Adaptation and Adversity. *Biol Psychiatry.* 2021;89(9):847–856. doi:10.1016/j.biopsych.2020.12.026
54. Jiang S, Zheng C, Wen G, Bu B, Zhao S, Xu X. Down-regulation of NR2B receptors contributes to the analgesic and antianxiety effects of enriched environment mediated by endocannabinoid system in the inflammatory pain mice. *Behav Brain Res.* 2022;435:114062. doi:10.1016/j.bbr.2022.114062
55. Longaretti A, Forastieri C, Gabaglio M, Rubino T, Battaglioli E, Rusconi F. Termination of acute stress response by the endocannabinoid system is regulated through lysine-specific demethylase 1-mediated transcriptional repression of 2-AG hydrolases ABHD6 and MAGL. *J Neurochem.* 2020;155(1):98–110. doi:10.1111/jnc.15000
56. Vickstrom CR, Liu X, Liu S, et al. Role of endocannabinoid signaling in a septohabenular pathway in the regulation of anxiety- and depressive-like behavior. *Mol Psychiatry.* 2021;26(7):3178–3191. doi:10.1038/s41380-020-00905-1
57. Biltz RG, Sawicki CM, Sheridan JF, Godbout JP. The neuroimmunology of social-stress-induced sensitization. *Nat Immunol.* 2022;23(11):1527–1535. doi:10.1038/s41590-022-01321-z
58. Pasquarelli N, Porazik C, Hanselmann J, Weydt P, Ferger B, Witting A. Comparative biochemical characterization of the monoacylglycerol lipase inhibitor KML29 in brain, spinal cord, liver, spleen, fat and muscle tissue. *Neuropharmacology.* 2015;91:148–156. doi:10.1016/j.neuropharm.2014.12.001
59. Wenzel TJ, Klegeris A. Novel multi-target directed ligand-based strategies for reducing neuroinflammation in Alzheimer's disease. *Life Sci.* 2018;207:314–322. doi:10.1016/j.lfs.2018.06.025
60. Kerr DM, Harhen B, Okine BN, Egan LJ, Finn DP, Roche M. The monoacylglycerol lipase inhibitor JZL184 attenuates LPS-induced increases in cytokine expression in the rat frontal cortex and plasma: differential mechanisms of action. *Br J Pharmacol.* 2013;169(4):808–819. doi:10.1111/j.1476-5381.2012.02237.x
61. Zhu D, Zhang J, Hashem J, Gao F, Chen C. Inhibition of 2-arachidonoylglycerol degradation enhances glial immunity by single-cell transcriptomic analysis. *J Neuroinflammation.* 2023;20(1):17. doi:10.1186/s12974-023-02701-4
62. Grabner GF, Eichmann TO, Wagner B, et al. Deletion of monoglyceride lipase in astrocytes attenuates lipopolysaccharide-induced neuroinflammation. *J Biol Chem.* 2016;291(2):913–923. doi:10.1074/jbc.M115.683615
63. Nomura DK, Morrison BE, Blankman JL, et al. Endocannabinoid hydrolysis generates brain prostaglandins that promote neuroinflammation. *Science.* 2011;334(6057):809–813. doi:10.1126/science.1209200
64. Alhouayek M, Masquelier J, Cani PD, Lambert DM, Muccioli GG. Implication of the anti-inflammatory bioactive lipid prostaglandin D2-glycerol ester in the control of macrophage activation and inflammation by ABHD6. *Proc Natl Acad Sci U S A.* 2013;110(43):17558–17563. doi:10.1073/pnas.1314017110
65. Kibret BG, Ishiguro H, Horiuchi Y, Onaivi ES. New insights and potential therapeutic targeting of CB2 cannabinoid receptors in CNS disorders. *Int J Mol Sci.* 2022;23(2):975. doi:10.3390/ijms23020975
66. Li J, Wang H, Liu D, et al. CB2R activation ameliorates late adolescent chronic alcohol exposure-induced anxiety-like behaviors during withdrawal by preventing morphological changes and suppressing NLRP3 inflammasome activation in prefrontal cortex microglia in mice. *Brain Behav Immun.* 2023;110:60–79. doi:10.1016/j.bbi.2023.02.001
67. Mecha M, Feliú A, Carrillo-Salinas FJ, et al. Endocannabinoids drive the acquisition of an alternative phenotype in microglia. *Brain Behav Immun.* 2015;49:233–245. doi:10.1016/j.bbi.2015.06.002
68. Young AP, Denovan-Wright EM. The dynamic role of microglia and the endocannabinoid system in neuroinflammation. *Front Pharmacol.* 2021;12:806417. doi:10.3389/fphar.2021.806417
69. Roberts CJ, Stuhr KL, Hutz MJ, Raff H, Hillard CJ. Endocannabinoid signaling in hypothalamic-pituitary-adrenocortical axis recovery following stress: effects of indirect agonists and comparison of male and female mice. *Pharmacol Biochem Behav.* 2014;117:17–24. doi:10.1016/j.pbb.2013.11.026
70. Aliczki M, Zelena D, Mikics E, et al. Monoacylglycerol lipase inhibition-induced changes in plasma corticosterone levels, anxiety and locomotor activity in male CD1 mice. *Horm Behav.* 2013;63(5):752–758. doi:10.1016/j.yhbeh.2013.03.017
71. Hill MN, McLaughlin RJ, Pan B, et al. Recruitment of prefrontal cortical endocannabinoid signaling by glucocorticoids contributes to termination of the stress response. *J Neurosci.* 2011;31(29):10506–10515. doi:10.1523/JNEUROSCI.0496-11.2011
72. Long JZ, Li W, Booker L, et al. Selective blockade of 2-arachidonoylglycerol hydrolysis produces cannabinoid behavioral effects. *Nat Chem Biol.* 2009;5(1):37–44. doi:10.1038/nchembio.129

73. Yamasaki T, Hatori A, Zhang Y, et al. Neuroprotective effects of minocycline and KML29, a potent inhibitor of monoacylglycerol lipase, in an experimental stroke model: a small-animal positron emission tomography study. *Theranostics*. 2021;11(19):9492–9502. doi:10.7150/thno.64320
74. Della Pietra A, Giniatullin R, Savinainen JR. Distinct activity of endocannabinoid-hydrolyzing enzymes MAGL and FAAH in key regions of peripheral and central nervous system implicated in migraine. *Int J Mol Sci*. 2021;22(3):1204. doi:10.3390/ijms22031204
75. Chanda PK, Gao Y, Mark L, et al. Monoacylglycerol lipase activity is a critical modulator of the tone and integrity of the endocannabinoid system. *Mol Pharmacol*. 2010;78(6):996–1003. doi:10.1124/mol.110.068304
76. Aaltonen N, Kedzierska E, Orzelska-Górka J, et al. In Vivo Characterization of the Ultrapotent Monoacylglycerol Lipase Inhibitor {4-[bis-(benzo[d][1,3]dioxol-5-yl)methyl]-piperidin-1-yl}(1H-1,2,4-triazol-1-yl)methanone (JJKK-048). *J Pharmacol Exp Ther*. 2016;359(1):62–72. doi:10.1124/jpet.116.233114
77. Aida J, Fushimi M, Kusumoto T, et al. Design, synthesis, and evaluation of piperazinyl pyrrolidin-2-ones as a novel series of reversible monoacylglycerol lipase inhibitors. *J Med Chem*. 2018;61(20):9205–9217. doi:10.1021/acs.jmedchem.8b00824
78. Xiang H, Wang Y, Yang L, et al. Novel MAGL inhibitors alleviate LPS-induced acute kidney injury by inhibiting NLRP3 inflammatory vesicles, modulating intestinal flora, repairing the intestinal barrier, and interfering with serum metabolism. *Molecules*. 2023;28(21):7245. doi:10.3390/molecules28217245
79. Afzal O, Kumar S, Kumar R, Firoz A, Jaggi M, Bawa S. Docking based virtual screening and molecular dynamics study to identify potential monoacylglycerol lipase inhibitors. *Bioorg Med Chem Lett*. 2014;24(16):3986–3996. doi:10.1016/j.bmcl.2014.06.029
80. Xiong F, Ding X, Zhang H, et al. Discovery of novel reversible monoacylglycerol lipase inhibitors via docking-based virtual screening. *Bioorg Med Chem Lett*. 2021;41:127986. doi:10.1016/j.bmcl.2021.127986
81. Tuccinardi T, Granchi C, Rizzolio F, et al. Identification and characterization of a new reversible MAGL inhibitor. *Bioorg Med Chem*. 2014;22(13):3285–3291. doi:10.1016/j.bmc.2014.04.057
82. Zhi Z, Zhang W, Yao J, et al. Discovery of Aryl formyl piperidine derivatives as potent, reversible, and selective monoacylglycerol lipase inhibitors. *J Med Chem*. 2020;63(11):5783–5796. doi:10.1021/acs.jmedchem.9b02137
83. Bononi G, Di Stefano M, Poli G, et al. Reversible monoacylglycerol lipase inhibitors: discovery of a new class of benzylpiperidine derivatives. *J Med Chem*. 2022;65(10):7118–7140. doi:10.1021/acs.jmedchem.1c01806
84. Di Stefano M, Masoni S, Bononi G, et al. Design, synthesis, ADME and biological evaluation of benzylpiperidine and benzylpiperazine derivatives as novel reversible monoacylglycerol lipase (MAGL) inhibitors. *Eur J Med Chem*. 2024;263:115916. doi:10.1016/j.ejmech.2023.115916
85. Bononi G, Tonarini G, Poli G, et al. Monoacylglycerol lipase (MAGL) inhibitors based on a diphenylsulfide-benzoylpiperidine scaffold. *Eur J Med Chem*. 2021;223:113679. doi:10.1016/j.ejmech.2021.113679
86. Kapanda CN, Muccioli GG, Labar G, Poupaert JH, Lambert DM. Bis(dialkylaminethiocarbonyl)disulfides as potent and selective monoacylglycerol lipase inhibitors. *J Med Chem*. 2009;52(22):7310–7314. doi:10.1021/jm901323s
87. Omran Z. New Disulfiram Derivatives as MAGL-Selective Inhibitors. *Molecules*. 2021;26(11):3296. doi:10.3390/molecules26113296
88. Andrei C, Mihai DP, Nitulescu G, et al. Cetirizine and levetiracetam as inhibitors of monoacylglycerol lipase: investigating their repurposing potential as novel osteoarthritic pain therapies. *Pharmaceuticals*. 2023;16(11):1563. doi:10.3390/ph16111563
89. Jiang M, Huizenga MCW, Wirt JL, et al. A monoacylglycerol lipase inhibitor showing therapeutic efficacy in mice without central side effects or dependence. *Nat Commun*. 2023;14(1):8039. doi:10.1038/s41467-023-43606-3
90. Adeel M, Saorin G, Boccalon G, et al. A carrier free delivery system of a monoacylglycerol lipase hydrophobic inhibitor. *Int J Pharm*. 2022;613:121374. doi:10.1016/j.ijpharm.2021.121374
91. Deng H, Li W. Monoacylglycerol lipase inhibitors: modulators for lipid metabolism in cancer malignancy, neurological and metabolic disorders. *Acta Pharm Sin B*. 2020;10(4):582–602. doi:10.1016/j.apsb.2019.10.006
92. Cisar JS, Weber OD, Clapper JR, et al. Identification of ABX-1431, a selective inhibitor of monoacylglycerol lipase and clinical candidate for treatment of neurological disorders. *J Med Chem*. 2018;61(20):9062–9084. doi:10.1021/acs.jmedchem.8b00951
93. Deng H, Zhang Q, Lei Q, et al. Discovering monoacylglycerol lipase inhibitors by a combination of fluorogenic substrate assay and activity-based protein profiling. *Front Pharmacol*. 2022;13:941522. doi:10.3389/fphar.2022.941522
94. Punt JM, van der Vliet D, van der Stelt M. Chemical probes to control and visualize lipid metabolism in the brain. *Acc Chem Res*. 2022;55(22):3205–3217. doi:10.1021/acs.accounts.2c00521
95. Takahata K, Seki C, Kimura Y, et al. First-in-human in vivo imaging and quantification of monoacylglycerol lipase in the brain: a PET study with 18F-T-401. *Eur J Nucl Med Mol Imaging*. 2022;49(9):3150–3161. doi:10.1007/s00259-021-05671-y
96. He Y, Grether U, Taddio MF, et al. Multi-parameter optimization: development of a morpholin-3-one derivative with an improved kinetic profile for imaging monoacylglycerol lipase in the brain. *Eur J Med Chem*. 2022;243:114750. doi:10.1016/j.ejmech.2022.114750
97. Cheng R, Fujinaga M, Yang J, et al. A novel monoacylglycerol lipase-targeted 18F-labeled probe for positron emission tomography imaging of brown adipose tissue in the energy network. *Acta Pharmacol Sin*. 2022;43(11):3002–3010. doi:10.1038/s41401-022-00912-8
98. King AR, Dotsey EY, Lodola A, et al. Discovery of potent and reversible monoacylglycerol lipase inhibitors. *Chem Biol*. 2009;16(10):1045–1052. doi:10.1016/j.chembiol.2009.09.012
99. Al-Romaiyan A, Masocha W. Pristimerin, a triterpene that inhibits monoacylglycerol lipase activity, prevents the development of paclitaxel-induced allodynia in mice. *Front Pharmacol*. 2022;13:944502. doi:10.3389/fphar.2022.944502
100. Merali Z, Cayer C, Kent P, et al. Sacred Maya incense, copal (Protium copal - Burseraceae), has antianxiety effects in animal models. *J Ethnopharmacol*. 2018;216:63–70. doi:10.1016/j.jep.2018.01.027
101. Tung M-C, Fung K-M, Hsu H-M, Tseng T-S. Discovery of 8-prenylnaringenin from hop (*Humulus lupulus* L.) as a potent monoacylglycerol lipase inhibitor for treatments of neuroinflammation and Alzheimer's disease. *RSC Adv*. 2021;11(49):31062–31072. doi:10.1039/d1ra05311f
102. El-Alfy AT, Joseph S, Brahmabhatt A, Akati S, Abourashed EA. Indirect modulation of the endocannabinoid system by specific fractions of nutmeg total extract. *Pharm Biol*. 2016;54(12):2933–2938. doi:10.1080/13880209.2016.1194864
103. Benkherouf AY, Soini SL, Stompor M, Uusi-Oukari M. Positive allosteric modulation of native and recombinant GABAA receptors by hops prenylflavonoids. *Eur J Pharmacol*. 2019;852:34–41. doi:10.1016/j.ejphar.2019.02.034
104. Beladjila KA, Berrehal D, De Tommasi N, et al. New phenylethanoid glycosides from cistanche phelypaea and their activity as inhibitors of Monoacylglycerol Lipase (MAGL). *Planta Med*. 2018;84(9–10):710–715. doi:10.1055/s-0044-100187

105. De Leo M, Huallpa CG, Alvarado B, et al. New diterpenes from *Salvia pseudorosmarinus* and their activity as inhibitors of monoacylglycerol lipase (MAGL). *Fitoterapia*. 2018;130:251–258. doi:10.1016/j.fitote.2018.09.010
106. Mei J, Guo R, Zhang F, et al. Identification of bioactive natural products using yeast: application to monoacylglycerol lipase inhibitor extraction from *Corydalis Rhizoma*. *Biomed Pharmacother*. 2022;149:112798. doi:10.1016/j.biopha.2022.112798
107. Chen L, Yan Y, Chen T, et al. Forsythiaside prevents  $\beta$ -amyloid-induced hippocampal slice injury by upregulating 2-arachidonoylglycerol via cannabinoid receptor 1-dependent NF- $\kappa$ B pathway. *Neurochem Int*. 2019;125:57–66. doi:10.1016/j.neuint.2019.02.008
108. Talia S, Benarous K, Lamrani M, Yousfi M. Lepidine B from *Lepidium sativum* seeds as multi-functional anti- Alzheimer's disease agent: in vitro and in silico studies. *Curr Comput Aided Drug Des*. 2021;17(3):360–377. doi:10.2174/1573409916666200302120305
109. Chicca A, Marazzi J, Gertsch J. The antinociceptive triterpene  $\beta$ -amyryn inhibits 2-arachidonoylglycerol (2-AG) hydrolysis without directly targeting cannabinoid receptors. *Br J Pharmacol*. 2012;167(8):1596–1608. doi:10.1111/j.1476-5381.2012.02059.x
110. Vecchiarelli HA, Morena M, Lee TTY, et al. Sex and stressor modality influence acute stress-induced dynamic changes in corticolimbic endocannabinoid levels in adult Sprague Dawley rats. *Neurobiol Stress*. 2022;20:100470. doi:10.1016/j.ynstr.2022.100470
111. Botsford C, Brellenthin AG, Cisler JM, Hillard CJ, Koltyn KF, Crombie KM. Circulating endocannabinoids and psychological outcomes in women with PTSD. *J Anxiety Disord*. 2023;93:102656. doi:10.1016/j.janxdis.2022.102656
112. Rubino T, Realini N, Castiglioni C, et al. Role in anxiety behavior of the endocannabinoid system in the prefrontal cortex. *Cereb Cortex*. 2008;18(6):1292–1301. doi:10.1093/cercor/bhm161
113. Tevosian M, Todorov H, Lomazzo E, et al. NAPE-PLD deletion in stress-TRAPed neurons results in an anxiogenic phenotype. *Transl Psychiatry*. 2023;13(1):152. doi:10.1038/s41398-023-02448-9
114. Hill MN, Kumar SA, Filipinski SB, et al. Disruption of fatty acid amide hydrolase activity prevents the effects of chronic stress on anxiety and amygdalar microstructure. *Mol Psychiatry*. 2013;18(10):1125–1135. doi:10.1038/mp.2012.90
115. Stopponi S, Fotio Y, Domi A, et al. Inhibition of fatty acid amide hydrolase in the central amygdala alleviates co-morbid expression of innate anxiety and excessive alcohol intake. *Addict Biol*. 2018;23(6):1223–1232. doi:10.1111/adb.12573
116. Postnov A, Schmidt ME, Pemberton DJ, et al. Fatty acid amide hydrolase inhibition by JNJ-42165279: a multiple-ascending dose and a positron emission tomography study in healthy volunteers. *Clin Transl Sci*. 2018;11(4):397–404. doi:10.1111/cts.12548
117. Paulus MP, Stein MB, Simmons AN, Risbrough VB, Halter R, Chaplan SR. The effects of FAAH inhibition on the neural basis of anxiety-related processing in healthy male subjects: a randomized clinical trial. *Neuropsychopharmacol*. 2021;46(5):1011–1019. doi:10.1038/s41386-020-00936-w
118. Li GL, Winter H, Arends R, et al. Assessment of the pharmacology and tolerability of PF-04457845, an irreversible inhibitor of fatty acid amide hydrolase-1, in healthy subjects. *Br J Clin Pharmacol*. 2012;73(5):706–716. doi:10.1111/j.1365-2125.2011.04137.x
119. Mayo LM, Asratian A, Lindé J, et al. Elevated anandamide, enhanced recall of fear extinction, and attenuated stress responses following inhibition of fatty acid amide hydrolase: a randomized, controlled experimental medicine trial. *Biol Psychiatry*. 2020;87(6):538–547. doi:10.1016/j.biopsych.2019.07.034
120. D'Souza DC, Cortes-Briones J, Creatura G, et al. Efficacy and safety of a fatty acid amide hydrolase inhibitor (PF-04457845) in the treatment of cannabis withdrawal and dependence in men: a double-blind, placebo-controlled, parallel group, phase 2a single-site randomised controlled trial. *Lancet Psychiatry*. 2019;6(1):35–45. doi:10.1016/S2215-0366(18)30427-9
121. Zimmermann T, Bartsch JC, Beer A, et al. Impaired anandamide/palmitoylethanolamide signaling in hippocampal glutamatergic neurons alters synaptic plasticity, learning, and emotional responses. *Neuropsychopharmacology*. 2019;44(8):1377–1388. doi:10.1038/s41386-018-0274-7
122. Fusse EJ, Scarante FF, Vicente MA, et al. Anxiogenic doses of rapamycin prevent URB597-induced anti-stress effects in socially defeated mice. *Neurosci Lett*. 2024;818:137519. doi:10.1016/j.neulet.2023.137519
123. Borges-Assis AB, Uliana DL, Hott SC, Guimarães FS, Lisboa SF, Resstel LBM. Bed nucleus of the stria terminalis CB1 receptors and the FAAH enzyme modulate anxiety behavior depending on previous stress exposure. *Prog Neuropsychopharmacol Biol Psychiatry*. 2023;125:110739. doi:10.1016/j.pnpbp.2023.110739
124. Danandeh A, Vozella V, Lim J, et al. Effects of fatty acid amide hydrolase inhibitor URB597 in a rat model of trauma-induced long-term anxiety. *Psychopharmacology*. 2018;235(11):3211–3221. doi:10.1007/s00213-018-5020-7
125. Flannery LE, Kerr DM, Finn DP, Roche M. FAAH inhibition attenuates TLR3-mediated hyperthermia, nociceptive- and anxiety-like behavior in female rats. *Behav Brain Res*. 2018;353:11–20. doi:10.1016/j.bbr.2018.06.030
126. de Ceglia M, Micioni Di Bonaventura MV, Romano A, et al. Anxiety associated with palatable food withdrawal is reversed by the selective FAAH inhibitor PF-3845: a regional analysis of the contribution of endocannabinoid signaling machinery. *Int J Eat Disord*. 2023;56(6):1098–1113. doi:10.1002/eat.23917
127. Bluett RJ, Gamble-George JC, Hermanson DJ, Hartley ND, Marnett LJ, Patel S. Central anandamide deficiency predicts stress-induced anxiety: behavioral reversal through endocannabinoid augmentation. *Transl Psychiatry*. 2014;4(7):e408. doi:10.1038/tp.2014.53
128. Carnevali L, Barbetti M, Fotio Y, et al. Enhancement of peripheral fatty acyl ethanolamide signaling prevents stress-induced social avoidance and anxiety-like behaviors in male rats. *Psychopharmacology*. 2023. doi:10.1007/s00213-023-06473-w
129. Vecchiarelli HA, Morena M, Keenan CM, et al. Comorbid anxiety-like behavior in a rat model of colitis is mediated by an upregulation of corticolimbic fatty acid amide hydrolase. *Neuropsychopharmacology*. 2021;46(5):992–1003. doi:10.1038/s41386-020-00939-7
130. Marco EM, Rapano KM, Caprioli A, Borsini F, Laviola G, Maccarrone M. Potential therapeutic value of a novel FAAH inhibitor for the treatment of anxiety. *PLoS One*. 2015;10(9):e0137034. doi:10.1371/journal.pone.0137034
131. Silva HH, Tavares V, Neto BV, Cerqueira F, Medeiros R, Silva MRG. FAAH rs324420 polymorphism: biological pathways, impact on elite athletic performance and insights for sport medicine. *Genes*. 2023;14(10):1946. doi:10.3390/genes14101946
132. Spohrs J, Ulrich M, Grön G, Plener PL, Abler B. FAAH polymorphism (rs324420) modulates extinction recall in healthy humans: an fMRI study. *Eur Arch Psychiatry Clin Neurosci*. 2022;272(8):1495–1504. doi:10.1007/s00406-021-01367-4
133. Sisk LM, Rapuano KM, Conley MI, et al. Genetic variation in endocannabinoid signaling is associated with differential network-level functional connectivity in youth. *J Neurosci Res*. 2022;100(3):731–743. doi:10.1002/jnr.24946
134. Dincheva I, Drysdale AT, Hartley CA, et al. FAAH genetic variation enhances fronto-amygdala function in mouse and human. *Nat Commun*. 2015;6:6395. doi:10.1038/ncomms7395

135. Marusak HA, Evanski J, Desai S, Rabinak CA. Impact of childhood trauma exposure, genetic variation in endocannabinoid signaling, and anxiety on frontolimbic pathways in children. *Cannabis Cannabinoid Res.* 2023;8(6):1079–1089. doi:10.1089/can.2022.0144
136. Hillard CJ. Chapter one - the endocannabinoid signaling system in the CNS: a primer. In: Parsons L, Hill M editors. *International Review of Neurobiology. Vol 125. Endocannabinoids.* Academic Press; 2015:1–47. doi:10.1016/bs.irm.2015.10.001
137. Yasmin F, Colangeli R, Morena M, et al. Stress-induced modulation of endocannabinoid signaling leads to delayed strengthening of synaptic connectivity in the amygdala. *Proc Natl Acad Sci.* 2020;117(1):650–655. doi:10.1073/pnas.1910322116
138. Gulyas AI, Cravatt BF, Bracey MH, et al. Segregation of two endocannabinoid-hydrolyzing enzymes into pre- and postsynaptic compartments in the rat hippocampus, cerebellum and amygdala. *Eur J Neurosci.* 2004;20(2):441–458. doi:10.1111/j.1460-9568.2004.03428.x
139. Morena M, Patel S, Bains JS, Hill MN. Neurobiological Interactions Between Stress and the Endocannabinoid System. *Neuropsychopharmacol.* 2016;41(1):80–102. doi:10.1038/npp.2015.166
140. Alugubelly N, Mohammed AN, Carr RL. Persistent proteomic changes in glutamatergic and GABAergic signaling in the amygdala of adolescent rats exposed to chlorpyrifos as juveniles. *Neurotoxicology.* 2021;85:234–244. doi:10.1016/j.neuro.2021.05.012
141. Ri C, A N, de L K, et al.. Inhibition of fatty acid amide hydrolase by chlorpyrifos in juvenile rats results in altered exploratory and social behavior as adolescents. *Neurotoxicology.* 2020;77. doi:10.1016/j.neuro.2020.01.002
142. Duan T, Gu N, Wang Y, et al. Fatty acid amide hydrolase inhibitors produce rapid anti-anxiety responses through amygdala long-term depression in male rodents. *J Psychiatry Neurosci.* 2017;42(4):230–241. doi:10.1503/jpn.160116
143. Pavón FJ, Serrano A, Stouffer DG, et al. Ethanol-induced alterations in endocannabinoids and relevant neurotransmitters in the nucleus accumbens of fatty acid amide hydrolase knockout mice. *Addict Biol.* 2019;24(6):1204–1215. doi:10.1111/adb.12695
144. Rossi S, De Chiara V, Musella A, et al. Preservation of striatal cannabinoid CB1 receptor function correlates with the antianxiety effects of fatty acid amide hydrolase inhibition. *Mol Pharmacol.* 2010;78(2):260–268. doi:10.1124/mol.110.064196
145. Aguilera-Portillo G, Rangel-López E, Villeda-Hernández J, et al. The pharmacological inhibition of fatty acid amide hydrolase prevents excitotoxic damage in the rat striatum: possible involvement of CB1 receptors regulation. *Mol Neurobiol.* 2019;56(2):844–856. doi:10.1007/s12035-018-1129-2
146. Kerr DM, Burke NN, Ford GK, et al. Pharmacological inhibition of endocannabinoid degradation modulates the expression of inflammatory mediators in the hypothalamus following an immunological stressor. *Neuroscience.* 2012;204:53–63. doi:10.1016/j.neuroscience.2011.09.032
147. Doenni VM, Gray JM, Song CM, Patel S, Hill MN, Pittman QJ. Deficient adolescent social behavior following early-life inflammation is ameliorated by augmentation of anandamide signaling. *Brain Behav Immun.* 2016;58:237–247. doi:10.1016/j.bbi.2016.07.152
148. Grieco M, De Caris MG, Maggi E, et al. Fatty Acid Amide Hydrolase (FAAH) inhibition modulates amyloid-beta-induced microglia polarization. *Int J Mol Sci.* 2021;22(14):7711. doi:10.3390/ijms22147711
149. Malek N, Popiolek-Barczyk K, Mika J, Przewlocka B, Starowicz K. Anandamide, acting via CB2 receptors, alleviates LPS-induced neuroinflammation in rat primary microglial cultures. *Neural Plast.* 2015;2015:130639. doi:10.1155/2015/130639
150. Tanaka M, Yagyu K, Sackett S, Zhang Y. Anti-inflammatory effects by pharmacological inhibition or knockdown of fatty acid amide hydrolase in BV2 microglial cells. *Cells.* 2019;8(5):491. doi:10.3390/cells8050491
151. Rivera P, Del Mar Fernández-Arjona M, Silva-Peña D, et al. Pharmacological blockade of fatty acid amide hydrolase (FAAH) by URB597 improves memory and changes the phenotype of hippocampal microglia despite ethanol exposure. *Biochem. Pharmacol.* 2018;157:244–257. doi:10.1016/j.bcp.2018.08.005
152. Tchanchou F, Tucker LB, Fu AH, et al. The fatty acid amide hydrolase inhibitor PF-3845 promotes neuronal survival, attenuates inflammation and improves functional recovery in mice with traumatic brain injury. *Neuropharmacology.* 2014;85:427–439. doi:10.1016/j.neuropharm.2014.06.006
153. Henry RJ, Kerr DM, Finn DP, Roche M. FAAH-mediated modulation of TLR3-induced neuroinflammation in the rat hippocampus. *J Neuroimmunol.* 2014;276(1–2):126–134. doi:10.1016/j.jneuroim.2014.09.002
154. Zhu Y, Zhang H, Mao H, et al. FAAH served a key membrane-anchoring and stabilizing role for NLRP3 protein independently of the endocannabinoid system. *Cell Death Differ.* 2023;30(1):168–183. doi:10.1038/s41418-022-01054-4
155. Abelson JL, Liberzon I, Young EA, Khan S. Cognitive modulation of the endocrine stress response to a pharmacological challenge in normal and panic disorder subjects. *Arch Gen Psychiatry.* 2005;62(6):668–675. doi:10.1001/archpsyc.62.6.668
156. Hill MN, McLaughlin RJ, Bingham B, et al. Endogenous cannabinoid signaling is essential for stress adaptation. *Proc Natl Acad Sci U S A.* 2010;107(20):9406–9411. doi:10.1073/pnas.0914661107
157. Hill MN, McLaughlin RJ, Morrish AC, et al. Suppression of amygdalar endocannabinoid signaling by stress contributes to activation of the hypothalamic-pituitary-adrenal axis. *Neuropsychopharmacology.* 2009;34(13):2733–2745. doi:10.1038/npp.2009.114
158. Newsom RJ, Osterlund C, Masini CV, Day HE, Spencer RL, Campeau S. Cannabinoid receptor type 1 antagonism significantly modulates basal and loud noise induced neural and hypothalamic-pituitary-adrenal axis responses in male Sprague-Dawley rats. *Neuroscience.* 2012;204:64–73. doi:10.1016/j.neuroscience.2011.11.043
159. Haller J, Barna I, Barsvari B, et al. Interactions between environmental aversiveness and the anxiolytic effects of enhanced cannabinoid signaling by FAAH inhibition in rats. *Psychopharmacology.* 2009;204(4):607–616. doi:10.1007/s00213-009-1494-7
160. Bedse G, Colangeli R, Lavecchia AM, et al. Role of the basolateral amygdala in mediating the effects of the fatty acid amide hydrolase inhibitor URB597 on HPA axis response to stress. *Eur Neuropsychopharmacol.* 2014;24(9):1511–1523. doi:10.1016/j.euroneuro.2014.07.005
161. Gray JM, Vecchiarelli HA, Morena M, et al. Corticotropin-releasing hormone drives anandamide hydrolysis in the amygdala to promote anxiety. *J Neurosci.* 2015;35(9):3879–3892. doi:10.1523/JNEUROSCI.2737-14.2015
162. Gray JM, Wilson CD, Lee TTY, et al. Sustained glucocorticoid exposure recruits cortico-limbic CRH signaling to modulate endocannabinoid function. *Psychoneuroendocrinology.* 2016;66:151–158. doi:10.1016/j.psyneuen.2016.01.004
163. Duranti A, Beldarrain G, Álvarez A, et al. The endocannabinoid system as a target for neuroprotection/neuroregeneration in perinatal hypoxic-ischemic brain injury. *Biomedicines.* 2022;11(1):28. doi:10.3390/biomedicines11010028
164. Oddi S, Fiorenza MT, Maccarrone M. Endocannabinoid signaling in adult hippocampal neurogenesis: a mechanistic and integrated perspective. *Prog Lipid Res.* 2023;91:101239. doi:10.1016/j.plipres.2023.101239
165. Fogaça MV, Campos AC, Coelho LD, Duman RS, Guimarães FS. The anxiolytic effects of cannabidiol in chronically stressed mice are mediated by the endocannabinoid system: role of neurogenesis and dendritic remodeling. *Neuropharmacology.* 2018;135:22–33. doi:10.1016/j.neuropharm.2018.03.001

166. Avraham HK, Jiang S, Fu Y, et al. Impaired neurogenesis by HIV-1-Gp120 is rescued by genetic deletion of fatty acid amide hydrolase enzyme. *Br J Pharmacol.* 2015;172(19):4603–4614. doi:10.1111/bph.12657
167. Segev A, Korem N, Mizrahi Zer-Aviv T, et al. Role of endocannabinoids in the hippocampus and amygdala in emotional memory and plasticity. *Neuropsychopharmacology.* 2018;43(10):2017–2027. doi:10.1038/s41386-018-0135-4
168. Kathuria S, Gaetani S, Fegley D, et al. Modulation of anxiety through blockade of anandamide hydrolysis. *Nat Med.* 2003;9(1):76–81. doi:10.1038/nm803
169. Koehl M, Ladevèze E, Catania C, Cota D, Abrous DN. Inhibition of mTOR signaling by genetic removal of p70 S6 kinase 1 increases anxiety-like behavior in mice. *Transl Psychiatry.* 2021;11(1):165. doi:10.1038/s41398-020-01187-5
170. Tanaka M, Yamada S, Watanabe Y. The Role of Neuropeptide Y in the Nucleus Accumbens. *Int J Mol Sci.* 2021;22(14):7287. doi:10.3390/ijms22147287
171. Amstadter AB, Koenen KC, Ruggiero KJ, et al. NPY moderates the relation between hurricane exposure and generalized anxiety disorder in an epidemiologic sample of hurricane-exposed adults. *Depress Anxiety.* 2010;27(3):270–275. doi:10.1002/da.20648
172. Decressac M, Barker RA. Neuropeptide Y and its role in CNS disease and repair. *Exp Neurol.* 2012;238(2):265–272. doi:10.1016/j.expneurol.2012.09.004
173. Molosh AI, Sajdyk TJ, Truitt WA, Zhu W, Oxford GS, Shekhar A. NPY Y1 receptors differentially modulate GABAA and NMDA receptors via divergent signal-transduction pathways to reduce excitability of amygdala neurons. *Neuropsychopharmacology.* 2013;38(7):1352–1364. doi:10.1038/npp.2013.33
174. Maymon N, Mizrahi Zer-Aviv T, Sabban EL, Akirav I. Neuropeptide Y and cannabinoids interaction in the amygdala after exposure to shock and reminders model of PTSD. *Neuropharmacology.* 2020;162:107804. doi:10.1016/j.neuropharm.2019.107804
175. Greco R, Demartini C, Zanaboni AM, et al. Potentiation of endocannabinoids and other lipid amides prevents hyperalgesia and inflammation in a pre-clinical model of migraine. *J Headache Pain.* 2022;23(1):79. doi:10.1186/s10194-022-01449-1
176. Fotio Y, Mabou Tagne A, Jung KM, Piomelli D. Fatty acid amide hydrolase inhibition alleviates anxiety-like symptoms in a rat model used to study post-traumatic stress disorder. *Psychopharmacology.* 2023. doi:10.1007/s00213-023-06358-y
177. Shang Y, Wang M, Hao Q, et al. Development of indole-2-carbonyl piperazine urea derivatives as selective FAAH inhibitors for efficient treatment of depression and pain. *Bioorg Chem.* 2022;128:106031. doi:10.1016/j.bioorg.2022.106031
178. Gur Maz T, Turanlı S, Caliskan HB, Çalışkan B, Banoglu E. Development and molecular modeling studies of new thiaziazole piperazine urea derivatives as potential fatty acid amide hydrolase inhibitors. *Arch Pharm.* 2022;355(8):e2200082. doi:10.1002/ardp.202200082
179. Keith JM, Jones W, Pierce JM, et al. Heteroaryureas with fused bicyclic diamine cores as inhibitors of fatty acid amide hydrolase. *Bioorg Med Chem Lett.* 2020;30(20):127463. doi:10.1016/j.bmcl.2020.127463
180. Jaiswal S, Tripathi RKP, Ayyannan SR. Scaffold hopping-guided design of some isatin based rigid analogs as fatty acid amide hydrolase inhibitors: synthesis and evaluation. *Biomed Pharmacother.* 2018;107:1611–1623. doi:10.1016/j.biopha.2018.08.125
181. Jaiswal S, Ayyannan SR. Lead optimization study on indoline-2,3-dione derivatives as potential fatty acid amide hydrolase inhibitors. *J Biomol Struct Dyn.* 2023;41(19):9632–9650. doi:10.1080/07391102.2022.2145372
182. Patel JZ, Parkkari T, Laitinen T, et al. Chiral 1,3,4-oxadiazol-2-ones as highly selective FAAH inhibitors. *J Med Chem.* 2013;56(21):8484–8496. doi:10.1021/jm400923s
183. Greco R, Francavilla M, Demartini C, et al. Activity of FAAH-Inhibitor JZP327A in an Experimental Rat Model of Migraine. *Int J Mol Sci.* 2023;24(12):10102. doi:10.3390/ijms241210102
184. Papa A, Pasquini S, Galvani F, et al. Development of potent and selective FAAH inhibitors with improved drug-like properties as potential tools to treat neuroinflammatory conditions. *Eur J Med Chem.* 2023;246:114952. doi:10.1016/j.ejmech.2022.114952
185. Zięba A, Laitinen T, Patel JZ, Poso A, Kaczor AA. Docking-based 3D-QSAR studies for 1,3,4-oxadiazol-2-one derivatives as FAAH inhibitors. *Int J Mol Sci.* 2021;22(11):6108. doi:10.3390/ijms22116108
186. Lorca M, Valdes Y, Chung H, Romero-Parra J, Pessoa-Mahana CD, Mella J. Three-Dimensional Quantitative Structure-Activity Relationships (3D-QSAR) on a Series of Piperazine-Carboxamides Fatty Acid Amide Hydrolase (FAAH) inhibitors as a useful tool for the design of new cannabinoid ligands. *Int J Mol Sci.* 2019;20(10):2510. doi:10.3390/ijms20102510
187. Zanfrescu A, Nitulescu G, Mihai DP, Nitulescu GM. Identifying FAAH inhibitors as new therapeutic options for the treatment of chronic pain through drug repurposing. *Pharmaceuticals.* 2021;15(1):38. doi:10.3390/ph15010038
188. Deplano A, Karlsson J, Svensson M, et al. Exploring the fatty acid amide hydrolase and cyclooxygenase inhibitory properties of novel amide derivatives of ibuprofen. *J Enzyme Inhib Med Chem.* 2020;35(1):815–823. doi:10.1080/14756366.2020.1743283
189. Deplano A, Karlsson J, Moraca F, et al. Design, synthesis and in vitro and in vivo biological evaluation of flurbiprofen amides as new fatty acid amide hydrolase/cyclooxygenase-2 dual inhibitory potential analgesic agents. *J Enzyme Inhib Med Chem.* 2021;36(1):940–953. doi:10.1080/14756366.2021.1875459
190. Montanari S, Scalvini L, Bartolini M, et al. Fatty Acid Amide Hydrolase (FAAH), Acetylcholinesterase (AChE), and Butyrylcholinesterase (BuChE): networked targets for the development of carbamates as potential anti-Alzheimer's Disease agents. *J Med Chem.* 2016;59(13):6387–6406. doi:10.1021/acs.jmedchem.6b00609
191. Maleki MF, Nadri H, Kianfar M, et al. Design and synthesis of new carbamates as inhibitors for fatty acid amide hydrolase and cholinesterases: molecular dynamic, in vitro and in vivo studies. *Bioorg Chem.* 2021;109:104684. doi:10.1016/j.bioorg.2021.104684
192. Wilt S, Kodani S, Le TNH, et al. Development of multitarget inhibitors for the treatment of pain: design, synthesis, biological evaluation and molecular modeling studies. *Bioorg Chem.* 2020;103:104165. doi:10.1016/j.bioorg.2020.104165
193. Cammarota M, Ferlenghi F, Vacondio F, et al. Combined targeting of fatty acid amide hydrolase and melatonin receptors promotes neuroprotection and stimulates inflammation resolution in rats. *Br J Pharmacol.* 2023;180(10):1316–1338. doi:10.1111/bph.16014
194. Intranuovo F, Brunetti L, DelRe P, et al. Development of N-(1-Adamantyl)benzamides as novel anti-inflammatory multitarget agents acting as dual modulators of the cannabinoid CB2 receptor and fatty acid amide hydrolase. *J Med Chem.* 2023;66(1):235–250. doi:10.1021/acs.jmedchem.2c01084
195. Wagenlehner FME, van Till JWO, Houbiers JGA, et al. Fatty acid amide hydrolase inhibitor treatment in men with chronic prostatitis/chronic pelvic pain syndrome: an adaptive double-blind, randomized controlled trial. *Urology.* 2017;103:191–197. doi:10.1016/j.urology.2017.02.029

196. Demartini C, Greco R, Zanaboni AM, Francavilla M, Facchetti S, Tassorelli C. URB937 prevents the development of mechanical allodynia in male rats with trigeminal neuralgia. *Pharmaceuticals*. 2023;16(11):1626. doi:10.3390/ph16111626
197. Zhang D, Saraf A, Kolasa T, et al. Fatty acid amide hydrolase inhibitors display broad selectivity and inhibit multiple carboxylesterases as off-targets. *Neuropharmacology*. 2007;52(4):1095–1105. doi:10.1016/j.neuropharm.2006.11.009
198. Kerbrat A, Ferré JC, Fillatre P, et al. Acute neurologic disorder from an inhibitor of fatty acid amide hydrolase. *N Engl J Med*. 2016;375(18):1717–1725. doi:10.1056/NEJMoa1604221
199. van Esbroeck ACM, Janssen APA, Cognetta AB, et al. Activity-based protein profiling reveals off-target proteins of the FAAH inhibitor BIA 10-2474. *Science*. 2017;356(6342):1084–1087. doi:10.1126/science.aaf7497
200. Otrubova K, Chatterjee S, Ghimire S, Cravatt BF, Boger DL. N-Acyl pyrazoles: effective and tunable inhibitors of serine hydrolases. *Bioorg Med Chem*. 2019;27(8):1693–1703. doi:10.1016/j.bmc.2019.03.020
201. Lamani M, Malamas MS, Farah SI, et al. Piperidine and piperazine inhibitors of fatty acid amide hydrolase targeting excitotoxic pathology. *Bioorg Med Chem*. 2019;27(23):115096. doi:10.1016/j.bmc.2019.115096
202. Thors L, Eriksson J, Amide C. Inhibition of the cellular uptake of anandamide by genistein and its analogue daidzein in cells with different levels of fatty acid amide hydrolase-driven uptake. *Br J Pharmacol*. 2007;152(5):744–750. doi:10.1038/sj.bjp.0707401
203. Ahmad H, Rauf K, Zada W, et al. Kaempferol facilitated extinction learning in contextual fear conditioned rats via inhibition of fatty-acid amide hydrolase. *Molecules*. 2020;25(20):4683. doi:10.3390/molecules25204683
204. Zada W, VanRyzin JW, Perez-Pouchoulen M, et al. Fatty acid amide hydrolase inhibition and N-arachidonylethanolamine modulation by isoflavonoids: a novel target for upcoming antidepressants. *Pharmacol Res Perspect*. 2022;10(5):e00999. doi:10.1002/prp2.999
205. Alasmari M, Böhlke M, Kelley C, Maher T, Pino-Figueroa A. Inhibition of Fatty Acid Amide Hydrolase (FAAH) by macamides. *Mol Neurobiol*. 2019;56(3):1770–1781. doi:10.1007/s12035-018-1115-8
206. Almukadi H, Wu H, Böhlke M, Kelley CJ, Maher TJ, Pino-Figueroa A. The macamide N-3-methoxybenzyl-linoleamide is a time-dependent fatty acid amide hydrolase (FAAH) inhibitor. *Mol Neurobiol*. 2013;48(2):333–339. doi:10.1007/s12035-013-8499-2
207. Police A, Shankar VK, Pandey P, Rangappa S, Doerksen RJ, Narasimha Murthy S. Novel topical anandamide formulation for alleviating peripheral neuropathic pain. *Int J Pharm*. 2023;641:123085. doi:10.1016/j.ijpharm.2023.123085
208. Shanmugasundaram D, Roza JM. Effect of broad-spectrum hemp extract on neurobehavioral activity on the immobilization stress-induced model in Sprague Dawley rats. *ScientificWorldJournal*. 2023;2023:3425576. doi:10.1155/2023/3425576
209. Lin YX, Sun JT, Liao ZZ, et al. Triterpenoids from the fruiting bodies of *Ganoderma lucidum* and their inhibitory activity against FAAH. *Fitoterapia*. 2022;158:105161. doi:10.1016/j.fitote.2022.105161
210. Li DW, Liu M, Leng YQ, et al. Lanostane triterpenoids from *Ganoderma lucidum* and their inhibitory effects against FAAH. *Phytochemistry*. 2022;203:113339. doi:10.1016/j.phytochem.2022.113339
211. Thors L, Alajakku K, Fowler CJ. The “specific” tyrosine kinase inhibitor genistein inhibits the enzymic hydrolysis of anandamide: implications for anandamide uptake. *Br J Pharmacol*. 2007;150(7):951–960. doi:10.1038/sj.bjp.0707172
212. El-Mekkawy S, Shahat AA, Alqahtani AS, et al. A polyphenols-rich extract from *Moricandia sinaica* Boiss. Exhibits analgesic, anti-inflammatory and antipyretic activities in vivo. *Molecules*. 2020;25(21):5049. doi:10.3390/molecules25215049
213. Thors L, Belghiti M, Fowler CJ. Inhibition of fatty acid amide hydrolase by kaempferol and related naturally occurring flavonoids. *Br J Pharmacol*. 2008;155(2):244–252. doi:10.1038/bjp.2008.237
214. Mariano A, Di Sotto A, Leopizzi M, et al. Antiarthritic effects of a root extract from *Harpagophytum procumbens* DC: novel insights into the molecular mechanisms and possible bioactive phytochemicals. *Nutrients*. 2020;12(9):2545. doi:10.3390/nu12092545
215. Gonçalves ECD, Assis PM, Junqueira LA, et al. Citral inhibits the inflammatory response and hyperalgesia in mice: the role of TLR4, TLR2/Dectin-1, and CB2 cannabinoid receptor/ATP-sensitive K<sup>+</sup> channel pathways. *J Nat Prod*. 2020;83(4):1190–1200. doi:10.1021/acs.jnatprod.9b01134
216. El-Alfy AT, Abourashed EA, Patel C, Mazhari N, An H, Jeon A. Phenolic compounds from nutmeg (*Myristica fragrans* Houtt.) inhibit the endocannabinoid-modulating enzyme fatty acid amide hydrolase. *J Pharm Pharmacol*. 2019;71(12):1879–1889. doi:10.1111/jphp.13174
217. Sanna MD, Les F, Lopez V, Galeotti N. Lavender (*Lavandula angustifolia* Mill.) essential oil alleviates neuropathic pain in mice with spared nerve injury. *Front Pharmacol*. 2019;10:472. doi:10.3389/fphar.2019.00472
218. De Petrocellis L, Ligresti A, Moriello AS, et al. Effects of cannabinoids and cannabinoid-enriched Cannabis extracts on TRP channels and endocannabinoid metabolic enzymes. *Br J Pharmacol*. 2011;163(7):1479–1494. doi:10.1111/j.1476-5381.2010.01166.x
219. Capasso R, Borrelli F, Aviello G, et al. Cannabidiol, extracted from *Cannabis sativa*, selectively inhibits inflammatory hypermotility in mice. *Br J Pharmacol*. 2008;154(5):1001–1008. doi:10.1038/bjp.2008.177
220. Li D, Leng Y, Liao Z, et al. Nor-triterpenoids from the fruiting bodies of *Ganoderma lucidum* and their inhibitory activity against FAAH. *Nat Prod Res*. 2023;37(4):579–585. doi:10.1080/14786419.2022.2078817
221. Chicca A, Caprioglio D, Minassi A, et al. Functionalization of  $\beta$ -caryophyllene generates novel polypharmacology in the endocannabinoid system. *ACS Chem Biol*. 2014;9(7):1499–1507. doi:10.1021/cb500177c
222. Irrera N, D’Ascola A, Pallio G, et al.  $\beta$ -Caryophyllene Mitigates Collagen Antibody Induced Arthritis (CAIA) in mice through a cross-talk between CB2 and PPAR- $\gamma$  receptors. *Biomolecules*. 2019;9(8):326. doi:10.3390/biom9080326
223. Wu H, Kelley CJ, Pino-Figueroa A, Vu HD, Maher TJ. Macamides and their synthetic analogs: evaluation of in vitro FAAH inhibition. *Bioorg Med Chem*. 2013;21(17):5188–5197. doi:10.1016/j.bmc.2013.06.034
224. Taboada-Rosell K, Castro-García FA, Medina-Saldivar C, Cruz-Visalaya SR, Pacheco-Otalora LF. The novel FAAH inhibitor, MCH1, reduces the infarction area in the motor cortex-related region but does not affect the sensorimotor function or memory and spatial learning in rats exposed to transient middle cerebral artery occlusion. *Brain Res*. 2024;1822:148636. doi:10.1016/j.brainres.2023.148636
225. Zhang ZW, Han P, Fu J, et al. Gut microbiota-based metabolites of Xiaoyao Pills (a typical Traditional Chinese medicine) ameliorate depression by inhibiting fatty acid amide hydrolase levels in brain. *J Ethnopharmacol*. 2023;313:116555. doi:10.1016/j.jep.2023.116555
226. Samandar F, Tehranizadeh ZA, Saberi MR, Chamani J. CB1 as a novel target for Ginkgo biloba’s terpene trilactone for controlling chemotherapy-induced peripheral neuropathy (CIPN). *J Mol Model*. 2022;28(9):283. doi:10.1007/s00894-022-05284-8
227. Xie J, Li Y, Liang Y, Kui H, Wang C, Huang J. Integration of non-targeted metabolomics with network pharmacology deciphers the anxiolytic mechanisms of Platycladi Semen extracts in CUMS mice. *J Ethnopharmacol*. 2023;315:116571. doi:10.1016/j.jep.2023.116571

228. Tian X, Liu T, Li L, et al. Visual high-throughput screening for developing a fatty acid amide hydrolase natural inhibitor based on an enzyme-activated fluorescent probe. *Anal Chem*. 2020;92(14):9493–9500. doi:10.1021/acs.analchem.9b05826
229. Tian M, Tian Z, Yao D, et al. A NIR fluorescent probe for fatty acid amide hydrolase bioimaging and its application in development of inhibitors. *J Mater Chem B*. 2021;9(32):6460–6465. doi:10.1039/d1tb01054a
230. Della Pietra A, Krivoshein G, Ivanov K, et al. Potent dual MAGL/FAAH inhibitor AKU-005 engages endocannabinoids to diminish meningeal nociception implicated in migraine pain. *J Headache Pain*. 2023;24(1):38. doi:10.1186/s10194-023-01568-3
231. Dong B, Shilpa BM, Shah R, et al. Dual pharmacological inhibitor of endocannabinoid degrading enzymes reduces depressive-like behavior in female rats. *J Psychiatr Res*. 2020;120:103–112. doi:10.1016/j.jpsychires.2019.10.010
232. Adamson Barnes NS, Mitchell VA, Kazantzis NP, Vaughan CW. Actions of the dual FAAH/MAGL inhibitor JZL195 in a murine neuropathic pain model. *Br J Pharmacol*. 2016;173(1):77–87. doi:10.1111/bph.13337
233. Toczek M, Ryszkiewicz P, Remiszewski P, et al. Weak hypotensive effect of chronic administration of the dual FAAH/MAGL inhibitor JZL195 in spontaneously hypertensive rats as revealed by area under the curve analysis. *Int J Mol Sci*. 2023;24(13):10942. doi:10.3390/ijms241310942
234. Abohalaka R, Karaman Y, Recber T, Onder SC, Nemutlu E, Bozkurt TE. Endocannabinoid metabolism inhibition ameliorates ovalbumin-induced allergic airway inflammation and hyperreactivity in Guinea pigs. *Life Sci*. 2022;306:120808. doi:10.1016/j.lfs.2022.120808
235. Manduca A, Morena M, Campolongo P, et al. Distinct roles of the endocannabinoids anandamide and 2-arachidonoylglycerol in social behavior and emotionality at different developmental ages in rats. *Eur Neuropsychopharmacol*. 2015;25(8):1362–1374. doi:10.1016/j.euroneuro.2015.04.005
236. Fowler CJ. Transport of endocannabinoids across the plasma membrane and within the cell. *FEBS J*. 2013;280(9):1895–1904. doi:10.1111/febs.12212
237. Nicolussi S, Gertsch J. Endocannabinoid transport revisited. *Vitam Horm*. 2015;98:441–485. doi:10.1016/bs.vh.2014.12.011
238. Jones MJ, Uzuneser TC, Clement T, et al. Inhibition of fatty acid binding protein-5 in the basolateral amygdala induces anxiolytic effects and accelerates fear memory extinction. *Psychopharmacology*. 2023. doi:10.1007/s00213-023-06468-7
239. Chicca A, Nicolussi S, Bartholomäus R, et al. Chemical probes to potently and selectively inhibit endocannabinoid cellular reuptake. *Proc Natl Acad Sci U S A*. 2017;114(25):E5006–E5015. doi:10.1073/pnas.1704065114
240. Bortolato M, Campolongo P, Mangieri RA, et al. Anxiolytic-like properties of the anandamide transport inhibitor AM404. *Neuropsychopharmacology*. 2006;31(12):2652–2659. doi:10.1038/sj.npp.1301061
241. Kwee CMB, Leen NA, Van der Kamp RC, et al. Anxiolytic effects of endocannabinoid enhancing compounds: a systematic review and meta-analysis. *Eur Neuropsychopharmacol*. 2023;72:79–94. doi:10.1016/j.euroneuro.2023.04.001

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