

# Rift Valley fever virus-infected mosquito ova and associated pathology: possible implications for endemic maintenance

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**Background:** Endemic/Enzootic maintenance mechanisms like vertical transmission (pathogen passage from infected adults to their offspring) are central in the epidemiology of zoonotic pathogens. In Kenya, Rift Valley fever virus (RVFV) may be maintained by vertical transmission in ground-pool mosquitoes such as *Aedes mcintoshi*. RVFV can cause serious morbidity and mortality in humans and livestock. Past epidemics/epizootics have occurred in sub-Saharan Africa but, since the late 1970s, RVFV has also appeared in North Africa and the Middle East. Preliminary results revealed RVFV-infected eggs in *Ae. mcintoshi* after virus injection into the hemocoel after the first of two blood meals, justifying further study.

**Methods:** Mosquitoes were collected from an artificially flooded water-catching depression along a stream in Kenya, shipped live to the USA, and studied using an immunocytochemical method for RVFV-antigen localization in mosquito sections.

**Results and conclusion:** After virus injection into the hemocoel, RVFV-infected reproductive tissues were found, particularly follicular epithelia and oocyte/nurse cells. Ovarian infection from the hemocoel is a crucial step in establishing a vertically transmitting mosquito line. Ovarian follicles originate from germarial cells, primordia located distally in each ovariole, and infection of these cells is expected to be requisite for long-term vertical transmission. However, no germarial cell infection was found, so establishing a new line of vertically transmitting mosquitoes may require two generations. The findings support the hypothesis that *Ae. mcintoshi* is involved in the endemic maintenance of RVFV by vertical transmission. Detection of distinct pathology in infected eggs raises the possibility of virus-laden eggs being deposited among healthy eggs, thereby providing an exogenous source of infection via ingestion by mosquito larvae and other organisms. This has potentially significant epidemiological implications. Possible modes of entry of virus from the hemocoel into the ovaries and routes by which larvae might become infected by ingesting virus are discussed.

**Keywords:** hemocoel, ovary, vertical transmission, *Aedes mcintoshi*, arbovirus, antigen positive

## Introduction

The mechanisms of endemic/enzootic maintenance are central to understanding the epidemiology of zoonotic pathogens. An example among arthropod-borne disease systems is vertical transmission, the passage of a pathogen from infected adults to their eggs (transovarial transmission), through the larval instars (transstadial transmission), and, ultimately, to the next generation of adults. Among arthropods, transovarial/transstadial pathogen transmission was first discovered in ticks relative to the rickettsia associated with Rocky Mountain spotted fever<sup>1</sup> and later in mosquitoes relative to LaCrosse virus.<sup>2</sup> Vertical transmission has subsequently been found in several mosquito/arbovirus combinations.<sup>3–8</sup>

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In this paper, evidence for Rift Valley fever virus (RVFV)-infected eggs (ova) in the African floodwater mosquito *Aedes mcintoshi*<sup>9</sup> after intrathoracic (IT) infection between two blood meals is reported. This finding is consistent with the vertical (transovarial) transmission of RVFV by this species.

Several mosquito-borne arboviruses are active in the USA and, given the introduction and rapid spread of West Nile virus beginning in 1999,<sup>10,11</sup> it is clear that other arboviruses are likewise capable of importation into the USA, for example RVFV.<sup>12,13</sup> RVFV (genus *Phlebovirus*, family Bunyaviridae) can cause serious human and livestock morbidity and mortality and is listed as a Category A potential bioterrorism agent.<sup>14</sup> In the past, RVFV epidemics/epizootics have occurred sporadically throughout much of sub-Saharan Africa, but, from the late 1970s, this virus has appeared in North Africa and the Middle East.<sup>15–20</sup> Cases associated with a recent outbreak in South Africa continue to appear in 2011.<sup>21</sup> In sub-Saharan Africa, RVFV is thought to be maintained by vertical transmission in ground-pool *Aedes* species; for example, one of the hypothetical interepizootic maintenance species in Kenya is the African floodwater mosquito, *Ae. mcintoshi*.<sup>22</sup>

The authors of this paper have been involved for many years in studies designed to elucidate the interaction between mosquitoes and arboviruses, including the use of in situ methods to locate RVFV antigen and genome segments in sections of mosquito tissues.<sup>23–27</sup> In a preliminary study of a Kenyan strain of RVFV in *Ae. mcintoshi*, a single RVFV antigen-positive chorionated egg was found after a second blood meal in a female that had been intrahemocoelically infected after the first blood meal. It was therefore evident that in this species, the reproductive system, particularly the follicular epithelia and oocyte/nurse cells, can become infected with virus originating in the hemocoel. Ovarian infection from the hemocoel after dissemination of virus from the midgut after a viremic blood meal is logically a critical first step in the establishment of a vertically transmitting line of mosquitoes de novo in a previously nonvertically transmitting strain or species. On this basis, the sequence of events that led to infection of follicular epithelia and of an egg was duplicated in a larger sample of mosquitoes.

While the research reported here was carried out several years ago, the ideas presented have developed from discussions, especially between the first and second authors during a study of western equine encephalitis virus infection in *Culex tarsalis* in which the same phenomenon was observed. The study of western equine encephalitis virus/

*Cx. tarsalis* will be reported separately, but the results are fully consistent with our findings in RVFV-infected *Ae. mcintoshi*.

## Materials and methods

### Mosquito collection

Specimens of *Ae. mcintoshi* were collected from an artificially flooded dambo (natural water-catching depression along a stream) at Sakari Ranch near Nairobi, Kenya, and shipped live to the US Army Medical Research Institute of Infectious Diseases (USAMRIID), Fort Detrick, Frederick, Maryland.

### Mosquito infection and analysis

In a preliminary study at USAMRIID, four mosquitoes that had obtained blood from an unknown host in Kenya were inoculated with RVFV (Kenya strain 21445) by injection into the hemocoel through a soft area of cuticle in the thorax (IT infection/innoculation), held at 26°C for several days, allowed to oviposit, and then given a second blood meal from an uninfected hamster. These mosquitoes were killed and fixed for immunocytochemical examination after being held long enough for vitellogenesis to occur.

Based on preliminary results, a larger sample of adult female mosquitoes that had been collected in Kenya and held at USAMRIID were likewise IT inoculated with the 21445 strain of RVFV. After 3 days incubation, mosquitoes were allowed to ingest, digest, and assimilate a blood meal from an uninfected hamster and provided with an opportunity to oviposit. A second blood meal opportunity was then provided and the resulting 35 engorged females incubated for about 3 days. Gravid females were then killed and fixed in 5% formaldehyde for approximately 5 hours and stored in 70% ethyl alcohol. IT-inoculated mosquitoes and similarly treated uninfected virus-diluent-injected female *Culex pipiens* mosquitoes (100% infection rate when IT inoculated) were prepared as positive and negative controls, respectively.

### Immunocytochemistry

To detect viral antigen in serial paraffin sections of mosquitoes, the avidin-biotin-peroxidase complex immunocytochemical technique for light-level microscopy was used.<sup>28,29</sup> The “primary antibody” was a blend of monoclonal antibodies directed against RVFV nucleocapsid protein and two envelope glycoproteins or a monoclonal antibody directed against RVFV nucleocapsid protein, both effective in antigen detection. Dr James Meegan and Dr Jonathan Smith provided

these antibodies. Appropriate reagent elimination studies had been carried out earlier.<sup>29</sup>

## Results

In the preliminary study, one of the four specimens had an antigen-positive egg with a distinct positive signal in all serial sections of that egg as well as an antigen-positive follicular epithelium. The results of the 35 RVFV-infected female *Ae. mcintoshi* study are shown in Table 1 and Figure 1A–H. Representative sections from control specimens are shown in Figure 1I–L.

Among the ultimate follicles with chorionated eggs, RVFV antigen was seen in a single egg in three different specimens; ie, 3/35 or 8.6%. Antigen could be seen in several serial sections of each infected egg along with distinct pathology characterized by the coalescence of yolk granules having a somewhat clumped, smooth appearance (Figure 1A and B). This pathology was in contrast to the granular yolk texture observed in uninfected eggs adjacent to the infected eggs (Figure 1A and B) and in the control (Figure 1L).

Both ovarian (Figure 1E) and ovariole sheaths (Figure 1A and D) were observed to contain antigen in many individuals, 74.3% and 51.4%, respectively. In no case was the germarium (Figure 1B) observed to contain antigen. A single penultimate follicle was observed to be antigen positive. A much higher frequency of antigen presence was observed in the follicular epithelia of ultimate follicles (Figure 1A and C), 62.9% in the cells associated with the anterior (distal) part of an egg,

55.9% in the cells of the central region of an egg, and 84.4% in the cells associated with the posterior (proximal) region of an egg (including the adjacent pedicel and calyx). The oviducts (Figure 1F) displayed high frequencies of antigen presence, as did cells near the spermathecae (Figure 1F and H). In no case was the accessory gland (Figure 1H), or spermatozoa, when present in the spermathecae, seen to contain antigen. Many instances of antigen in the trachea associated with the ovaries were observed (Figure 1G). In one case a chorionated egg contained distinct traces of RVFV antigen (Figure 1C).

## Discussion

The findings of this study are consistent with the hypothesis that *Ae. mcintoshi* is involved in the endemic maintenance of RVFV by vertical transmission.<sup>22</sup> The occurrence of RVFV antigen-positive eggs is evidence that RVFV in *Ae. mcintoshi* can reach and infect eggs from the hemocoel when there is a disseminated infection (virus in the hemocoel beyond the midgut epithelium). The fact that mosquitoes were IT inoculated, given an uninfected blood meal, allowed to oviposit, and then given a second blood meal may have influenced the receptivity of the ovaries to RVFV infection.

The occurrence of strongly antigen-positive eggs with antigen in virtually every section, along with associated pathology, suggests that eggs heavily infected with RVFV can be deposited in the aquatic habitat along with uninfected viable eggs. The observation of distinct traces of antigen in an otherwise normal-appearing egg (Figure 1C) suggests that some eggs could have a mild or even latent viral infection with no pathology and which might remain viable.

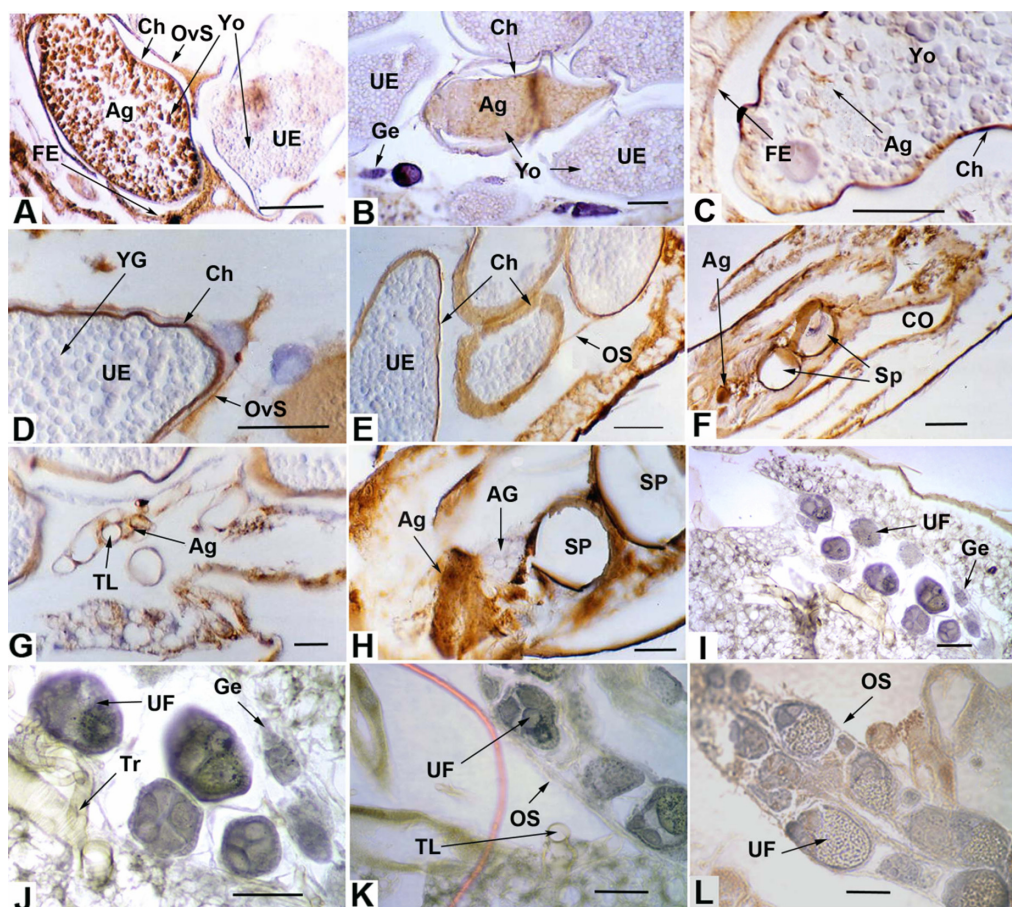
The appearance of distinct antigen in the cytoplasm of smaller tracheae suggests that virus entry into the ovaries could possibly occur via ovarian tracheae. It has been suggested that tracheae could be virus escape conduits from the midgut epithelium into the hemolymph (dissemination).<sup>25,26</sup> Virus could enter the ovaries by direct infection of the ovarian and ovariole sheaths and then the follicular epithelia. Because antigen was observed in all of these tissues, it is possible that both tracheal and sheath routes are involved in infection of the ovaries.

Regardless of mode of ingress into an ovary, infection of a developing egg with virus from the hemocoel can logically be expected to be a critical step in the establishment de novo of a line of vertically transmitting mosquitoes. A permanent vertically transmitting line of mosquitoes would ultimately require infection of the germaria, the undifferentiated

**Table 1** RVFV antigen in *Aedes mcintoshi* female reproductive tissues

Tissue/Structure	N RVFV Antigen +/ N observed (% Ag +)
Ovarian sheath	26/35 (74.3)
Ovariole sheath	18/35 (51.4)
Germarium	0/35 (0)
Penultimate follicle:	
Epithelium	1/35 (2.9)
Oocyte/Nurse cells	1/35 (2.9)
Ultimate follicle:	
Oocyte/Nurse cells	3/35 (8.6)
Epithelium-anterior	22/35 (62.9)
Epithelium-central	19/34 (55.9)
Epithelium-posterior (pedicel/calyx)	27/32 (84.4)
Lateral oviduct	32/35 (91.4)
Common oviduct	31/34 (91.2)
Genital chamber	26/33 (78.8)
Cells near spermathecal pores	20/31 (64.5)
Spermatozoa	0/23 (0)
Accessory gland	0/25 (0)

**Abbreviations:** Ag +, antigen positive; N, number; RVFV, Rift Valley fever virus.



**Figure 1** (A, B) RSVFV antigen in chorionated eggs showing pathology and RSVFV antigen-negative germarium (B). (C) Chorionated egg with distinct traces of RSVFV antigen. (D) Uninfected chorionated egg with antigen-positive ovariole sheath. (E) Uninfected chorionated eggs with RSVFV antigen-positive ovarian sheath. (F) Sagittal section in posterior part of the abdomen showing antigen-positive oviduct epithelium and tissues in proximity to the spermathecae. (G) Antigen-positive small tracheae associated with the ovaries. (H) Section of spermathecae and adjacent antigen-positive tissues, and antigen-negative accessory gland. (I–L) Negative control sections.

**Note:** Scale lines ( $\mu\text{m}$ ): A–E, 150; F, 100; G, 20; H, 50; I–L, 50.

**Abbreviations:** Ag, RSVFV antigen; AG, accessory gland; Ch, chorion; CO, common oviduct; FE, follicular epithelium; Ge, germarium; OS, ovary sheath; OvS, ovariole sheath; RSVFV, Rift Valley fever virus; Sp, spermatheca; TL, tracheal lumen; Tr, trachea; UE, uninfected egg; UF, uninfected follicle; Yo, yolk.

primordial tissue at the apex of each ovariole from which the oocytes/nurse cells and follicular epithelia originate. Virus in germarial cells would have ample opportunity to infect oocytes and follicular epithelia as they develop. It seems safe to assume that many, if not all, follicles originating from an infected germarium would be infected. However, no evidence of antigen in germarial tissue was observed. How, then, could a permanent vertically transmitting line be established?

The finding of what appears to be a lightly infected egg suggests the possibility of other eggs having undetected or latent infections. If such eggs remained viable, a line of a vertically transmitting mosquito generations with a “stabilized” viral infection<sup>30</sup> could be established. This is because infection would be systemic from the very beginning as opposed to the virus from a viremic blood meal being

required to infect several differentiated tissues to spread. Virus that has infected an egg would have opportunity to spread during virtually all of the processes of growth and development, that is, during embryogenesis, during the larval stadia, and during metamorphosis to the adult stage. Virus present during metamorphosis would be in a position to infect any developing adult tissues, including primordial germ cells and adult salivary gland cells. Infected female germ cells would be expected to produce at least some infected germaria and hence ovarioles that would produce infected eggs. Any infected but viable eggs among all eggs deposited would be expected to undergo the same sequence of tissue-infection events during growth and development, thereby leading to infected eggs in the next generation, and so on, a generational repeat of vertical transmission, that is, a stabilized infection.<sup>30</sup>

Likewise, such a systemic infection in a male egg might produce infected male gametes (spermatozoa), raising the possibility of venereal transmission, a phenomenon first described in male *Aedes triseriatus*, tree-hole mosquitoes infected with LaCrosse encephalitis virus<sup>31</sup> and later in two other mosquito-arbovirus combinations.<sup>32,33</sup> Infection of spermatheca-associated tissues could lead to “transovum” transmission, in which case virus might eventually enter chorionated eggs via the micropyle along with spermatozoa as they enter eggs passing along the oviduct.

Though admittedly speculative, another way a permanent vertically transmitting line could be established stems from the fact that heavily infected eggs displayed signs of pathology. This indicates that likely nonviable virus-laden eggs can be deposited along with viable uninfected eggs. Once in the aquatic environment, these eggs could conceivably be a source of viral infection for mosquito larvae or other aquatic organisms. A logical route of egress of virus from an infected egg would be via the micropyle, the opening through which sperm enter and then initiate fertilization. The eventual breakdown of a heavily infected egg could also release virus. Larvae from viable eggs deposited along with one or more infected eggs and larvae from female adults of the same or different species could easily ingest released virus particles. Infected eggs (or larvae) are also likely to be ingested by other animals (vertebrate or invertebrate), offering a putative mechanism for cross-specific viral transmission.<sup>34</sup> Such transmission within the larval habitat could play a very significant role in endemic maintenance and could also enhance virus spread and amplification associated with animal and/or human disease outbreaks. Mosquito larval habitats generally teem with other organisms, including arthropods and other invertebrates, Monera, and Protists,<sup>35</sup> and it is well known that various vertebrates (fish, tadpoles, frogs, toads, birds, and bats) prey on mosquito larvae and/or adults.

The idea of infected eggs releasing virus, which infect larvae when ingested, is consistent with the demonstration that larval ingestion of particles from homogenized, RVFV-infected hamster livers in the rearing medium lead to transstadial virus transmission and produce adults that transmit virus to new hosts during blood feeding.<sup>36</sup> It is unclear how infected hamster livers and infected mosquito eggs compare in regard to the amount of virus present, but the very strong positive reaction of the egg shown in Figure 1A is indicative of widespread infection within the egg and suggests the possibility that the viral titer within an infected egg could be reasonably high.

To establish infection in a mosquito larva, ingested virus has to come into contact with midgut epithelial cell surface-receptor molecules associated with the plasma membranes. However, the larval foregut is lined with an impermeable, noncellular chitinous intima<sup>37,38</sup> and ingested particles in the midgut lumen are completely invested in the cylindrical noncellular peritrophic membrane (PM). The larval PM, while permeable to large molecules such as the mosquitoes’ own proteolytic enzymes,<sup>39,40</sup> is not permeable to virus-sized particles. However, the work of Turell et al<sup>36</sup> demonstrates that ingested RVFV particles must in fact reach plasma membrane and infect epithelial cells. The question remains regarding how virus-sized particles circumvent the foregut intima and larval PM and initiate infection. Relative to the structure of the mosquito alimentary canal and to developmental changes that happen in the fourth instar larval stage, there are at least two possibilities. First, it seems possible that virus could reach the foregut–midgut junction within the larval proventriculus, immediately anterior to where the larval PM forms but posterior to the cells to which the foregut intima attaches and terminates. There is evidence that RVFV can infect the corresponding region in adult mosquitoes and possibly disseminate from there into the hemocoel.<sup>23,27</sup> A second plausible way for ingested virus to reach the plasma membranes of larval midgut epithelial cells is during the pharate pupal stage at the time when the larval PM is destroyed or egested but before the meconial PM forms.<sup>41</sup> In this regard, it is noteworthy that Turell et al used fourth instar larvae in their experiments.<sup>36</sup>

Although four individual eggs infected in four different mosquitoes may seem to be a low frequency of infection, that 4/35 individuals (11.4%) contain any RVFV antigen-positive eggs could be very significant, especially in view of the potentially large populations of floodwater mosquitoes emerging in a short period of time. Arbovirus-infected egg pathology has also been observed in *Cx. tarsalis* in association with an arbovirus in a different family, that is, the Togaviridae,<sup>42</sup> suggesting that our observations of infected eggs associated with apparent pathology are not just random artifacts and may well play a role in the establishment of a line of vertically transmitting mosquitoes.

## Disclosure

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