

UVR Promotes Keratinocyte Phagocytosis and Skin Pigmentation Through TRPA1 Channels

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Purpose: Ultraviolet radiation (UVR) enhances skin pigmentation, which involves the production of melanin by melanocytes and subsequent transfer to keratinocytes. In the epidermis, keratinocyte phagocytosis plays a pivotal role in the process of melanosome transfer to protect DNA of epidermal cells against damage from UVR. Previous research suggested that transient receptor potential channels ankyrin 1 (TRPA1) was required for UVR-induced early melanin synthesis in melanocytes. Currently, there is no evidence that supports the detailed mechanism of TRPA1 for UVR-induced phagocytosis by keratinocytes. Here, we investigated the effect and the possible mechanisms of TRPA1 on keratinocyte phagocytosis and skin pigmentation after UVR exposure.

Methods: Flow cytometry was applied to investigate the effect of TRPA1 on intracellular calcium concentration ($[Ca^{2+}]_{ic}$) and fluorescent microspheres uptake was carried out to analyze phagocytosis in HaCaT cells (human immortalized keratinocytes). Western blotting was applied to measure the protein expression of calcium/calmodulin-dependent protein kinase II (CaMKII), phosphorylated CaMKII and β -catenin after UVA/UVB exposure. Masson-Fontana staining was applied to observe the effect of XAV-939 (decreasing the expression of β -catenin) on UVB-induced skin pigmentation in guinea pigs.

Results: TRPA1 channels activated by UVR increased the $[Ca^{2+}]_{ic}$ and phosphorylation of CaMKII in HaCaT cells. The UVR-induced phagocytosis was regulated by TRPA1 in HaCaT cells. TRPA1 promoted the protein expression of β -catenin after UVR exposure in HaCaT cells. XAV-939, inhibiting β -catenin expression, decreased the UVB-induced skin pigmentation on in vivo guinea pig models.

Conclusion: Taken together, TRPA1 activated by UVR led to the increase of intracellular calcium, which promoted the phosphorylation of CaMKII, enhancing keratinocyte phagocytosis. Moreover, TRPA1 regulated the protein expression of β -catenin to exert a lightening effect on skin pigmentation. Our findings suggest that TRPA1 may be a potential therapeutic target for UVR-induced skin pigmentary diseases.

Keywords: transient receptor potential arkyin 1, ultraviolet radiation, phagocytosis, skin pigmentation, CaMKII, β -catenin

Introduction

Ultraviolet radiation (UVR) is known to be the main extrinsic factor that induces skin pigmentation, which is a vital physiological process to protect nuclear DNA of epidermal cells from UV damage.¹ Melanocytes stimulated by UVR synthesize melanin within melanosomes, which are subsequently transferred to adjacent keratinocytes, thus resulting in visually observable skin pigmentation.¹ Proposed mechanisms of melanosome transfer include the following models: cytophagocytosis,² direct membrane fusion,³ shed vesicles,^{4,5} and coupled exo/phagocytosis.^{6,7} Recent observations favor the coupled exo/phagocytosis model, which proposed that the melanosome core was released from melanocytes and then phagocytosed by neighboring keratinocytes.⁸ In this process, keratinocyte phagocytosis serves an important part, however, the detailed mechanism of which has not been fully elucidated.

Transient receptor potential ankyrin 1 (TRPA1), a calcium (Ca^{2+})-permeable non-selective cationic channel, is a unique member of the mammalian TRP ankyrin subfamily which plays key functions in chemo-, thermo-, and mechano-sensing.^{9,10} Previous studies indicated that TRPA1 was mainly expressed in sensory neurons, while recent researches confirmed its non-neuronal expressions in lung, brain and vascular endothelial tissues.⁹ Moreover, TRPA1 was detected to be expressed in human cutaneous cells, including melanocytes and keratinocytes.¹¹ With regard to the biological function of TRPA1 in epidermal cells, it is demonstrated that TRPA1 activated by UVR caused a retinal-dependent current and a rapid calcium influx and was required for the UVR-induced early increase of cellular melanin in melanocytes.¹² In mammalian phototransduction, retinal evoked a UVR-sensitive current in melanocytes, probably due to the conversion of trans-retinal to the cis- conformation catalyzed by the retinoid isomerohydrolase RPE65 (retinal pigment epithelium-specific 65kDa protein), which is also expressed in keratinocytes, as in the rod and cone cells of visual cycle.^{13–15} What is more, TRPA1 was found to be involved in the proliferation and differentiation of keratinocytes.¹¹ However, whether it contributes to keratinocyte phagocytosis for promoting skin pigmentation remains unclear.

Calcium/calmodulin-dependent protein kinase II (CaMKII) is a serine-threonine protein kinase, which plays a crucial role in cell-migration-related cytoskeleton dynamics via auto-phosphorylation mediated mechanism.¹⁶ β -catenin is a core component of the canonical Wnt signaling pathway, and it facilitates the process of gene transcription and cell–cell adhesion.^{17,18} β -catenin binds to the intercellular domain of E-cadherin (Epithelia-cadherin) and links α -catenin, composing a cadherin-catenin complex, which directly anchors the actin cytoskeleton so that mediates the cell–cell adhesion.¹⁹ In the process of phagocytosis, actin cytoskeleton dynamics is required for the localized protrusion of the plasma membrane and the formation of the extended pseudopodia.²⁰ Given the relevance of the CaMKII and β -catenin to the actin cytoskeleton, their involvement in phagocytosis is of great research potential. However, their effect on keratinocyte phagocytosis has not been investigated yet.

Since UVR could activate TRPA1 channels in melanocytes leading to melanin synthesis, the aim of this study was to evaluate whether TRPA1 channels activated by UVR could promote keratinocyte phagocytosis and skin pigmentation in vitro and in vivo, and investigate the possible mechanisms which may involve the phosphorylation of CaMKII and the increased expression of β -catenin in keratinocytes.

Materials and Methods

Main Antibodies and Materials

TRPA1 (19124-1-AP) was purchased from Proteintech Group (Rosemont, IL 60018, USA). CaMKII (4436), pCaMKII (12716), β -catenin (8480) were purchased from Cell Signaling Technology (Danvers, Massachusetts, USA). Retinal (R2500), JT010 (SML1672), HC030031 (H4415) were purchased from Sigma Aldrich (Darmstadt, Germany). Dimethyl sulfoxide (DMSO), RIPA buffer (R0100), BCA protein assay kit (PC0020), poloxamer gel (Polyethylene-polypropylene glycol 407, S7071) and formalin (G2161) were purchased from Solarbio (Beijing, China). Protease inhibitor cocktail (B14011) was purchased from Bimake (Shanghai, China). XAV-939 (S1180) was purchased from Selleck Chemicals (Shanghai, China). Fluo-4 AM (ab142773), DIO Staining Solution (C1038) were purchased from Beyotime (Shanghai, China). Stock solutions were prepared as follows: HC030031, and XAV-939 were dissolved in DMSO, JT010, and retinal in absolute ethanol. Masson-Fontana staining solution (G2032) was purchased from Solarbio (Beijing, China). Opti-MEM medium (31985062), Lipofectamine 2000 (11668030) and chemiluminescence (ECL) were obtained from Thermo Fisher Scientific (Massachusetts, USA). The UVA or UVB source used was a 9W UVA or UVB broadband lamp (Philips, Eindhoven, Netherlands) and radiation energies were measured using a UVX radiometer (UVP, Upland, California, USA).

HaCaT Cell Lines and Transfection

HaCaT cells (human immortalized keratinocytes) were purchased from the CABRI (European Union). Cells were cultured and grown in Minimum Eagle's Medium (MEM, Gibco, USA), supplemented with 10% fetal bovine serum (FBS, Gibco, USA) and 1% penicillin/streptomycin solution (Gibco, USA) at the humidified incubator with 5% CO_2 (v/v) at 37°C. hTRPA1-specific primers were designed based on NM_007332.2: F 5'-GGTTTGGCAGTTGGCGACATTGCTGA3' and R 5'-CTAAGGCTCAAGATGGTGTGTTTTG-3'. The primers amplified a DNA band that was sequenced and found to be identical to hTRPA1.

TRPA1 full-length cDNA was then recombined into pcDNA3.1-HA vector and expressed using a transient transfection system. HaCaT cells (50×10^4 cells/well) were overexpressed TRPA1 plasmid in an Opti-MEM medium containing Lipofectamine 2000 at 37°C. Cells were harvested 48 hours after transfection.

Measurement of Intracellular Calcium Concentration

HaCaT cells (50×10^4 cells/well) were seeded in 6-well plates (Corning Costar, USA) and incubated overnight. Cells were treated with different conditions (Retinal 12 μM , JT010 1 μM , HC-030031 10 μM , XVA-939 10 μM , UVA 225 mJ/cm^2 or UVB 25 mJ/cm^2) for 2 hours, and then loaded with 5 μM Fluo-4 AM calcium probes in PBS for 30 minutes and then washed twice with PBS. Cells were incubated with 0.8 mM Ca^{2+} -containing solution (140 mM NaCl, 3 mM KCl, 0.4 mM Na_2HPO_4 , 10 mM HEPES, 5 mM Glucose, and 1 mM MgCl_2 , and 0.8 mM CaCl_2 with pH 7.4) by incubation for 30 min at room temperature in the dark. To remove the Ca^{2+} -containing solution, cells were washed two times with PBS solution, harvested, resuspended, and then the fluorescence intensity was measured by flow cytometry (BD Aria II software, USA) with excitation of 340 nm and emission at 510 nm. At least 30,000 cells were collected per sample. Intracellular calcium concentration ($[\text{Ca}^{2+}]_{\text{ic}}$) was assessed by the fluorescence intensity ratio of the calcium probe Fluo-4 AM and expressed as relative fluorescence intensity normalized to control cells.

Calcium Imaging

HaCaT cells (20×10^4 cells/dish) were seeded onto culture glass dishes (Nest, China) and incubated overnight. The following day, cells were treated with JT010 (1 μM), HC-030031 (10 μM), XVA-939 (10 μM), UVA exposure (225 mJ/cm^2) or UVB exposure (25 mJ/cm^2), and loaded with 5 μM Fluo-4 AM calcium probes as described in intracellular calcium concentration measurement. After incubation with the Fluo-4 AM calcium probe, calcium imaging was measured by Confocal fluorescence microscopy (Lacia, TCS SP8), and randomly taken from 5 microscopic fields in each experiment. Three independent experiments were conducted for each experiment and the fluorescence areas were measured by Image J software.

Microsphere-Based Phagocytosis Assay

HaCaT cells (30×10^4 cells/well) were seeded onto coverslips in 24-well plates (Corning Costar) and incubated overnight. Cells were serum-starved for 6 hours, then incubated with Fluospheres® carboxylate-modified red fluorescent microspheres (0.5 μm diameter, Invitrogen) for 16 hours, when cells were treated with JT010 (1 μM), HC-030031 (10 μM), XVA-939 (10 μM), UVA exposure (225 mJ/cm^2) or UVB exposure (25 mJ/cm^2). Cells were vigorously washed 3 times with cold PBS to remove microspheres that had not been internalized. Cells were directly harvested and measured the fluorescence intensity by flow cytometry. In addition, cells were fixed in cold 4% paraformaldehyde for 30 min, stained with Dio for 15 min, and observed by confocal fluorescence microscopy. Quantitative analysis was performed by counting the number of internalized microspheres in 100 cells per condition, which were randomly taken from 6 microscopic fields in three independent experiments.

Western Blotting Analysis

Total protein was solubilized from cell lysates using RIPA buffer supplemented with protease inhibitor cocktail. Protein concentration was measured using the BCA protein assay kit. A total of 40 mg proteins were electrophoresed in 8% SDS-PAGE gels and transferred to polyvinylidene difluoride (PVDF) membranes, and then were blocked in 5% bovine serum albumin (BSA) in Tris-buffered saline containing 0.1% Tween-20 (TBST) for 1 h. Subsequently, membranes were incubated with TRPA1, CaMKII, pCaMKII or β -catenin antibodies (all diluted to 1:1000 with 5% BSA solution) overnight at 4°C, followed by horseradish peroxidase-conjugated secondary antibodies (diluted to 1:2000 with 5% BSA solution) for 1 hour at room temperature. Finally, immunopositive bands were analyzed by enhanced chemiluminescence (ECL) and visualized using Tanon 5200 software (China). Band quantification for three experiments was done using Image J software (USA).

Animals

Experimental procedures were approved by the Animal Ethics Committee of Dalian Medical University (AEE22013), and animal care followed the National Institute of Health guidelines on the care and use of laboratory animals. The dorsal skin of brown guinea pigs (7 weeks old, weighing approximately 500–550 g) was divided into 3 areas, Control, Vehicle (33% poloxamer gel), XAV-939 (10 mM in 33% poloxamer gel). The corresponding areas were treated locally for 30 min every other day, and the UVB lamp was placed 15 cm above the guinea pig. The total energy dose of UVB exposure was 500 mJ/cm² for 2 weeks. The L value was measured in each application area (automatically averaging 3 times per point) by the colorimeter (Thermo), which was used to evaluate the lightness of skin. Experimental areas of skin were biopsied, processed, and fixed overnight in neutral-buffered 10% formalin, and then embedded in paraffin. They were then applied to Hematoxylin Eosin (H&E) staining and Masson-Fontana (M&F) staining. M&F staining was performed according to the manufacturer's instructions. The melanin granules were measured by Image J software. Quantitative analysis was performed by counting the positive stained areas of M&F staining in the epidermis, which were randomly taken from 5 microscopic fields in three independent experiments.

Statistical Analysis

All data are presented as means ± standard deviation (SD), and the data are the mean values from at least three independent experiments. Statistical analyses were performed using GraphPad Prism 8.0 software (San Diego, CA). Significant differences between the groups were performed using Brown-Forsythe's test, Student's *t*-test and one-way ANOVA with Tukey's post hoc test. A P-value <0.05 was considered statistically significant.

Results

TRPA1 Channels Contributed to UVA/UVB-Induced Ca²⁺ Responses in HaCaT Cells

To unveil the effect of UVA/UVB on the Ca²⁺ responses in HaCaT cells, the cells were processed with 225 mJ/cm² UVA or 25 mJ/cm² UVB (equivalent to 250 s of full sun exposure).¹² In Figure 1A and B, the data from flow cytometry assay showed that the [Ca²⁺]_{ic} had no change after treatment with retinal (the chromophore required for light activation of opsin G protein-coupled receptors)²¹ or UVA/UVB compared to the control group, respectively. Furthermore, the [Ca²⁺]_{ic} was increased 102%/66%, respectively, after UVA/UVB exposure in comparison with the control group, when HaCaT cells were preincubated with retinal (Figure 1A and B, *p* < 0.001). To investigate the effect of TRPA1 on UVA/UVB-induced Ca²⁺ responses, HaCaT cells were transfected with plasmids to overexpress TRPA1 or treated with JT010 (1 μM, a selective TRPA1 agonist), HC-030031 (10 μM, a selective TRPA1 antagonist). As shown in Figure 1C, the [Ca²⁺]_{ic} was significantly increased 70% after treatment with JT010 (*p* < 0.001), whereas pretreatment with HC-030031 decreased 35%/36% of the UVA/UVB-induced [Ca²⁺]_{ic}, respectively (*p* < 0.01), in HaCaT cells. Besides, UVA/UVB exposure significantly increased 224%/232% [Ca²⁺]_{ic} in HaCaT cells, which were transfected with plasmids overexpressing TRPA1 (Figure 1C, *p* < 0.001). Subsequently, HaCaT cells were loaded with Fluo4-AM, which showed fluorescence on binding with free calcium, and the fluorescence-positive areas were observed using a fluorescence confocal microscopy. The results of the fluorescence-positive area showed that exposure to UVA/UVB, treatment with JT010, or overexpressing TRPA1 enhanced the Ca²⁺ responses, but it was significantly decreased after treatment with HC-030031 in HaCaT cells (Figure 1D). These results indicated that UVR-induced Ca²⁺ responses were regulated by TRPA1 in HaCaT cells.

UVR-Induced Phosphorylation of CaMKII and Keratinocyte Phagocytosis Was Regulated by TRPA1 in HaCaT Cells

To further explore the effect of TRPA1 mediated Ca²⁺ responses on UVR-induced keratinocyte phagocytosis, we detected the expression of phosphorylated CaMKII (pCaMKII) using Western blot, and measured the fluorescent intensity and the fluorescent microspheres uptake by flow cytometry and confocal fluorescence microscopy. As shown in Figure 2A and B, the results showed that the ratio of pCaMKII/CaMKII was significantly increased 1.41-, 1.82- or 1.93-fold after treatment with JT010 (*p* < 0.05) or UVA/UVB (*p* < 0.001), respectively. However, the decline of the ratio

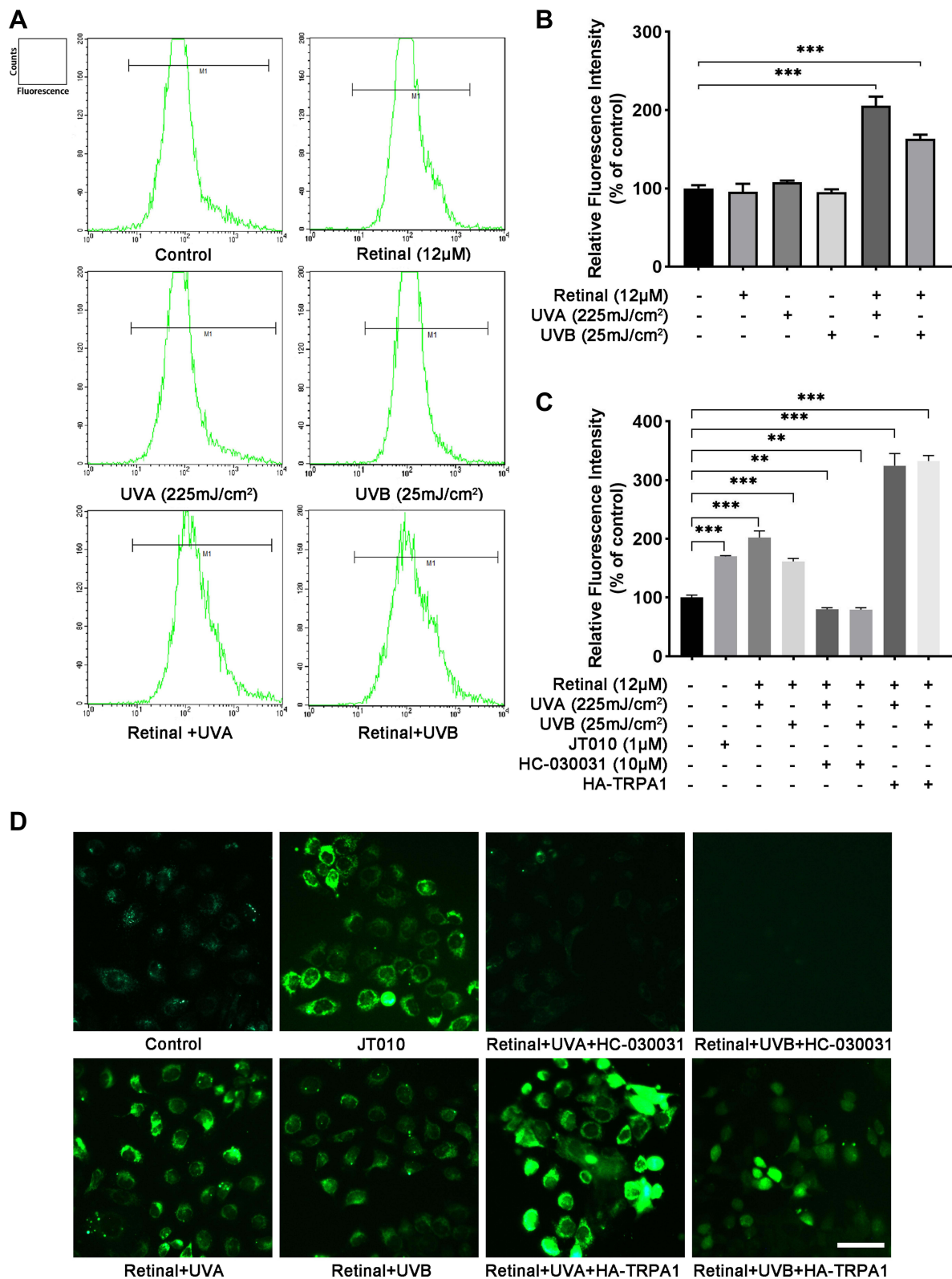


Figure 1 TRPA1 channels regulated UVA/UVB-induced Ca^{2+} responses in HaCaT cells. **(A)** Fluor-4 AM fluorescence intensity indicating calcium concentration was detected by flow cytometry in HaCaT cells after exposure to UVA (225 mJ/cm²) or UVB (25 mJ/cm²) with or without retinal preincubation (12 μM). **(B and C)** Quantification of the fluorescence intensity at different treatments. Values are mean value of relative fluorescence intensity normalized to control from three independent tests $\pm$ SD. **(D)** Calcium imaging was observed by fluorescence confocal microscopy after treatment with UVA or UVB exposure, JT010 (1 μM, TRPA1 agonist), HC-030031 (10 μM, TRPA1 antagonist), or HA-TRPA1. (Fluo-4 AM, green fluorescence). (** $p < 0.01$; *** $p < 0.001$).

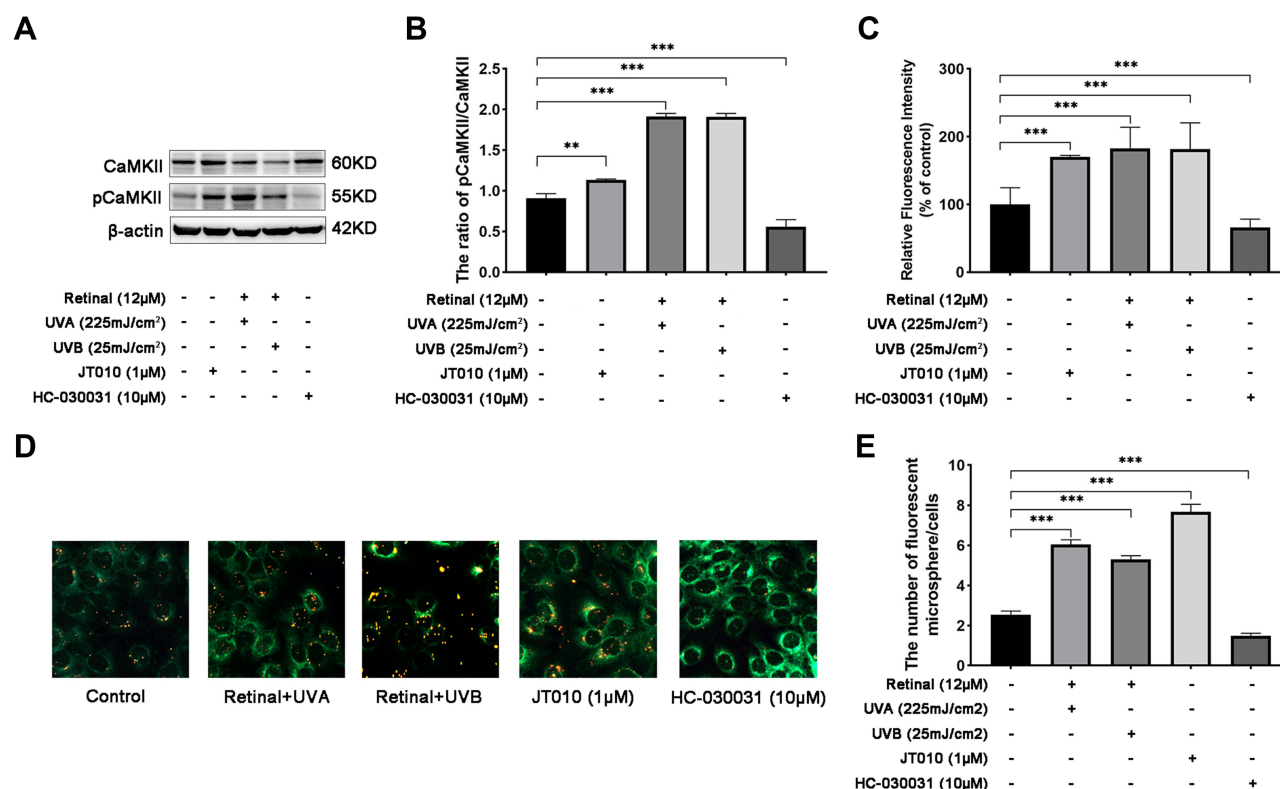


Figure 2 UVR-induced phosphorylation of CaMKII and keratinocyte phagocytosis was regulated by TRPA1 in HaCaT cells. TRPA1 channels promoted phosphorylation of CaMKII (**A** and **B**), (**A**) the protein levels of CaMKII and phosphorylated CaMKII (pCaMKII) were measured by Western blot in HaCaT cells, which were treated with UVA or UVB exposure, JT010 (1 μM, TRPA1 agonist), HC-030031 (10 μM, TRPA1 antagonist). (**B**) Relative protein levels of pCaMKII to CaMKII. Values are mean value of the ratio relative to control from three independent tests ± SD. (**C**) Quantification of fluorescence intensity indicating the uptake of fluorescent microspheres by HaCaT cells detected by Flow cytometry. Values are expressed as mean ± SD from three independent tests. (**D**) Fluorescent microspheres uptake was observed by fluorescence confocal microscopy, with red fluorescence indicating microspheres (0.5 μm microspheres), and green fluorescence indicating cell membrane stained with DiO. HaCaT cells were treated with UVA or UVB exposure, or JT010 (1 μM), HC-030031 (10 μM). Scale bar: 50 μm. (**E**) Quantification of fluorescent microspheres per HaCaT cell. Values are expressed as mean ± SD from three independent tests. (**p* < 0.01; ****p* < 0.001).

of pCaMKII/CaMKII was observed in HaCaT cells after treated with HC-030031, and the fold decreases were 0.5 compared with the control group (*p* < 0.001) (Figure 2A and B). Furthermore, fluorescent microspheres (0.5 μm, red) are often used as pseudo-melanocores to study keratinocyte phagocytosis,^{22,23} and the results showed that the fluorescence intensity was increased 82%/81% after UVA/UVB exposure compared with the control group, respectively (Figure 2C, *p* < 0.001). At the same time, the fluorescent intensity was increased 70% after treatment with JT010, but it was decreased 36% after treatment with HC-030031, in comparison with the control group (Figure 2C, *p* < 0.001). Moreover, the fluorescent microsphere uptake was increased 2.21-/2.00-fold after UVA/UVB exposure, as well as treatment with JT010 (2.92-fold) (Figure 2D and E, *p* < 0.001). The fluorescent microsphere uptake decreased 0.5-fold after treatment with HC-030031 compared with the control group (Figure 2D and E, *p* < 0.05). These discoveries above proved that TRPA1 regulated UVR-induced phosphorylation of CaMKII and the fluorescent microsphere uptake in HaCaT cells.

TRPA1 Enhanced the Expression of β-Catenin Promoting Phagocytosis in HaCaT Cells

To determine the effect of β-catenin on UVR-induced phagocytosis mediated by TRPA1, we measured the expression of β-catenin and observed the fluorescent microsphere uptake in HaCaT cells. The results showed that the expression of TRPA1 increased 1.28-/1.30-fold after UVA/UVB exposure in HaCaT cells, which were transfected with HA-TRPA1 plasmids compared to the control group (Figure 3A and B, *p* < 0.05). Subsequently, compared with the control group, the expression of β-catenin increased 1.64-, 1.72-/1.68-fold after treatment with JT010, UVA/UVB exposure in HaCaT cells (Figure 3A and C, *p* < 0.05). Interestingly, when HaCaT cells were transfected with HA-TRPA1 plasmids, the protein expression of β-catenin significantly increased 2.54-/2.49-fold after UVA/UVB

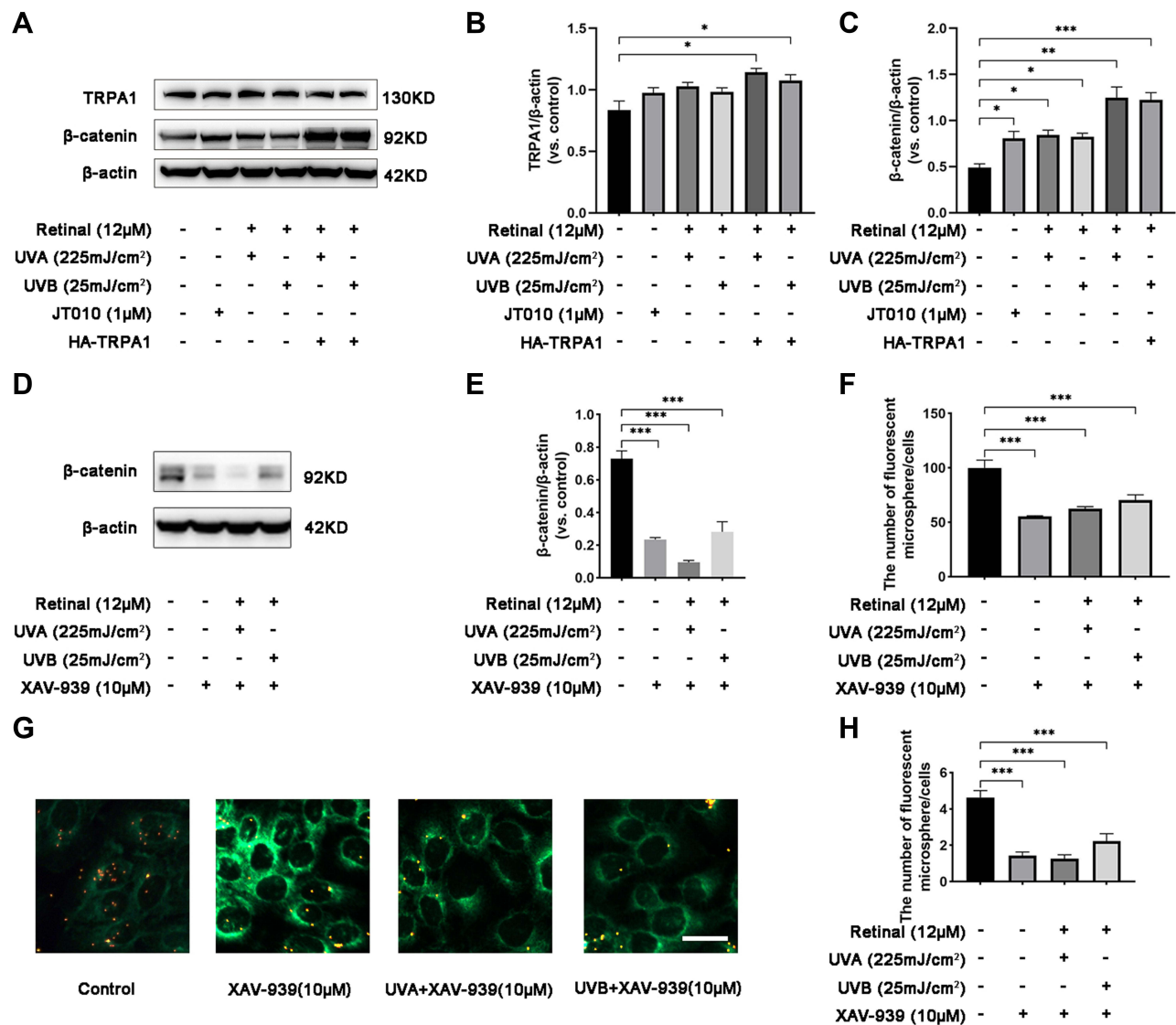


Figure 3 TRPA1 enhanced the expression of β -catenin promoting phagocytosis in HaCaT cells. **(A)** The protein levels of TRPA1 and β -catenin were measured by Western blot in HaCaT cells, which were treated with UVA or UVB exposure, JT010 (1 μ M, TRPA1 agonist), or overexpressed with HA-TRPA1 respectively. **(B and C)** Quantification of TRPA1 and β -catenin protein expression levels. Values are mean value of protein expression normalized to β -actin and relative to control from three independent tests $\pm$ SD. **(D)** The protein levels of β -catenin were measured by Western blot in HaCaT cells, which were treated with XAV-939 (10 μ M, decreasing β -catenin expression) alone or before exposure to UVA/UVB. **(E)** Quantification of β -catenin protein expression levels. Values are mean value of protein expression normalized to β -actin and relative to control from three independent tests $\pm$ SD. **(F)** Quantification of fluorescence intensity indicating the fluorescent microspheres uptake was detected by Flow cytometry in HaCaT cells. Values are expressed as mean $\pm$ SD from three independent tests. **(G)** Fluorescent microspheres uptake was observed by fluorescence confocal microscopy, with red fluorescence indicating microspheres, and green fluorescence indicating cell membrane stained with DiO. HaCaT cells were treated with XAV-939 (10 μ M) alone or before exposure to UVA/UVB respectively. Scale bar: 50 μ m. **(H)** Quantification of fluorescent microspheres per HaCaT cell. Values are expressed as mean $\pm$ SD from three independent tests. (* p < 0.05; ** p < 0.01; *** p < 0.001).

exposure (Figure 3A–C, p < 0.001). Moreover, the expression of β -catenin significantly decreased 0.48-, 0.61-/0.45-fold in HaCaT cells, which were treated with XAV-939 (10 μ M, decreasing β -catenin expression) alone, or before UVA/UVB exposure (Figure 3D and E, p < 0.001). Meanwhile, the fluorescent intensity was decreased 44%, 37%/31% (Figure 3F, p < 0.001), and the fluorescent microsphere uptake decreased 0.32-, 0.27-/0.39-fold compared with the control group (Figure 3G and H, p < 0.001), when HaCaT cells were treated with XAV-939 alone, or before UVA/UVB exposure. These findings indicated that TRPA1 enhanced the protein expression of β -catenin to promote UVR-induced phagocytosis in HaCaT cells.

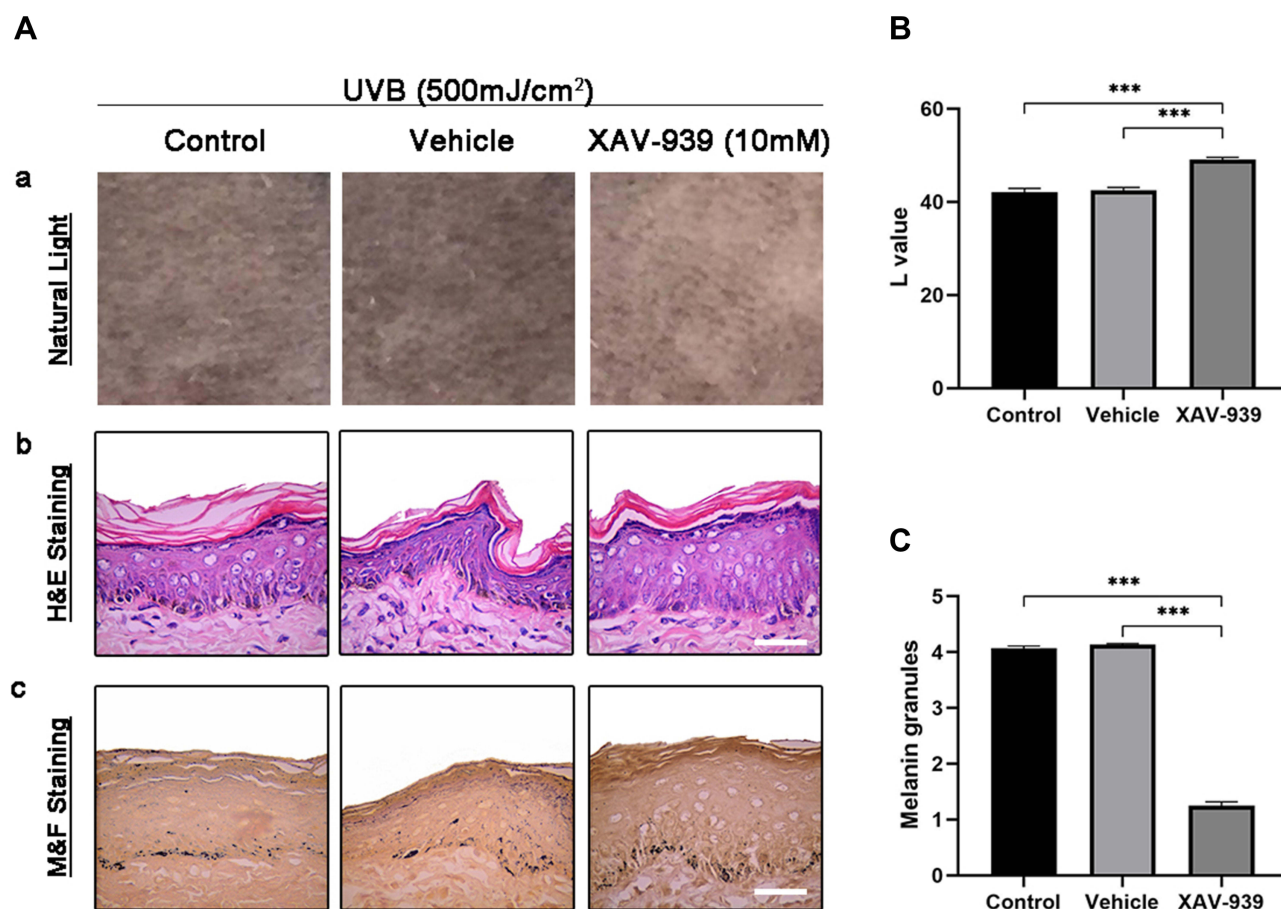


Figure 4 β -catenin regulated the UVB-induced skin pigmentation on Guinea pig models. **(A)** Effect of β -catenin on regulation of UVB-induced skin pigmentation in Guinea pigs. **(A)** Photographs of the lightening effect of XAV-939 (10 mM) on UVB-induced skin pigmentation. **(B-C)** H&E staining and M&F (Masson-Fontana) staining of biopsy specimens from the Control, Vehicle (33% poloxamer gel), or XAV-939 (10 mM in 33% poloxamer gel)-treated dorsal skin areas of brown Guinea pigs (200 $\times$). **(B)** Quantification of L value was measured by colorimeter at the end of the experiment and values are expressed as mean $\pm$ SD from three independent tests. **(C)** Quantitative analysis of melanin granules stained by M&F. Values are expressed as mean $\pm$ SD from three independent tests. (***) $p < 0.001$.

β -Catenin Regulated the UVB-Induced Skin Pigmentation on Guinea Pig Models

To ascertain the effect of β -catenin on pigmentation, we carried out the topical application of XAV-939 in vivo UVB-induced skin pigmentation guinea pig models. In the photograph of dorsal skin, XAV-939 (10 mM) showed depigmenting effects on UVB-induced skin pigmentation after 2 weeks of topical application, compared with the control and vehicle groups (Figure 4A). As for colorimetric measurements, the L value (skin lightness) significantly increased 17% and 16% in XAV-939 areas compared to control and vehicle groups (Figure 4B). The skin biopsy specimens were obtained from treated areas and were processed for light microscopy examination with H&E staining and M&F staining. The results showed no obvious changes of epidermal thickness in the control, vehicle and XAV-939 groups determined by H&E staining (Figure 4A). Furthermore, XAV-939 significantly reduced 69% and 70% the melanin granules revealed by M&F staining, which stains melanin granules as black, comparing with control and vehicle groups (Figure 4A and C). This result indicated that XAV-939 displayed a skin lightening effect on UVB-induced skin pigmentation in guinea pig models.

Discussion

Melanocytes in the epidermis of skin synthesize melanosomes and transfer them to the nuclear area of approximately 36 surrounding keratinocytes under the stimulation of UVR in the sunlight, which causes skin pigmentation, serving as a photo-protecting mechanism.¹ In the process of melanosome transfer, keratinocyte phagocytosis has recently been

considered as a crucial section.²⁴ Notably, a recent study suggested that melanocores (the melanin core devoid of surrounding membrane) were internalized by phagocytosis, whereas melanosomes (melanin core with intact surrounding membrane) were internalized by macropinocytosis in keratinocytes.²⁵ In this study, we confirmed that UVR enhanced the phagocytic ability of keratinocytes, which was regulated by TRPA1.

As a Ca^{2+} -permeable cationic channel, TRPA1 was reported to be activated by UVR and increased the $[\text{Ca}^{2+}]_{\text{ic}}$ in human melanocytes.¹² Our results identified similar findings in keratinocytes. Ca^{2+} is a key second messenger involved in the regulation of numerous cellular functions including adhesion, vesicular trafficking and cytoskeletal rearrangement.²⁰ Calmodulin (CaM), a Ca^{2+} sensor protein, plays an important role in signal transduction pathways by binding with Ca^{2+} .²⁶ CaMKII is activated by the binding of Ca^{2+} saturated CaM ($\text{Ca}^{2+}/\text{CaM}$) in an auto-phosphorylation manner.²⁷ It is reported that CaMKII is a multi-subunit holoenzyme, which contains a kinase domain, an autoinhibitory/regulatory domain, an actin binding domain and an association domain.^{27,28} CaMKII possesses two kinds of functional activities including kinase activity and structural function. At basal status, the autoinhibitory domain masks the kinase domain, which inhibits the kinase activity. Meanwhile, the actin binding domain interacts with more than one filament that is composed of filamentous actin (F-actin), and bundles them together, to stabilize the F-actin of cytoskeleton.²⁷ When $\text{Ca}^{2+}/\text{CaM}$ binds to the regulatory domain, the kinase domain is unmasked and the catalytic activity of the kinase is disinhibited. Once activated, CaMKII not only phosphorylates various substrates but also autophosphorylates itself at the autoinhibitory domain, turning into pCaMKII.^{27,29} In the meantime, autophosphorylated CaMKII dissociates from F-actin, which is subsequently unbundled and remodeled by actin regulators, therefore contributing to the cytoskeleton dynamics.³⁰ In the present study, it was demonstrated that UVR increased the $[\text{Ca}^{2+}]_{\text{ic}}$ in keratinocytes, which was mediated by TRPA1. Moreover, TRPA1 regulated UVR-induced phosphorylation of CaMKII and keratinocyte phagocytosis. Thus, we assumed that UVR enhanced keratinocyte phagocytosis through TRPA1 mediated calcium signaling pathway. The possible mechanism might be that UVR activated TRPA1 and increased the intercellular Ca^{2+} , which phosphorylated CaMKII by binding with CaM, accompanied with the remodeling of F-actin, facilitating phagocytosis in keratinocytes (Figure 5).

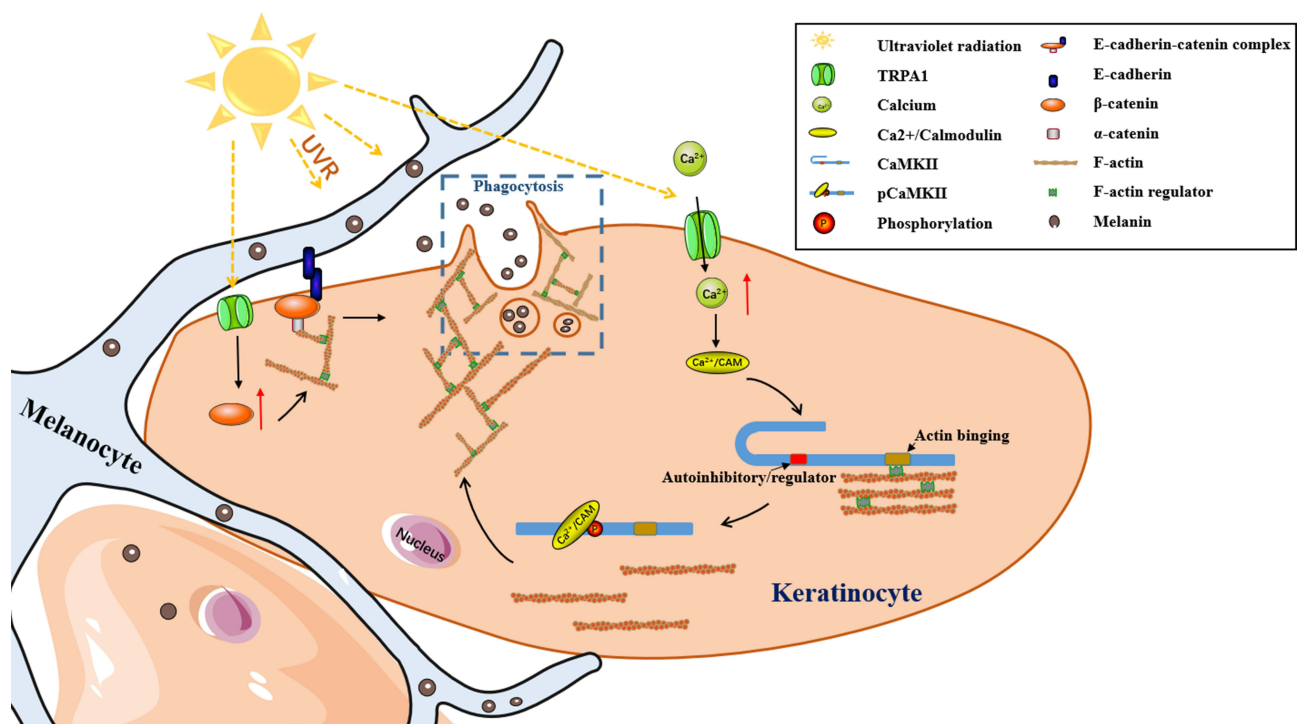


Figure 5 Schematic modulation of UVR-induced phagocytosis by TRPA1 in keratinocytes. TRPA1 is activated by UVR to increase the intracellular calcium, which promotes the phosphorylation of CaMKII, contributing to keratinocyte phagocytosis through the remodeling of F-actin. Moreover, TRPA1 activated by UVR increases the expression of β -catenin to enhance keratinocyte phagocytosis through cell-cell adhesion and cytoskeleton dynamics.

Direct cell–cell contact between MC and KC is a requirement for optimal melanosome transfer, which is accomplished by the adhesion ability of the cadherin-catenin complex, where β -catenin binds to E-cadherin and interacts with α -catenin.^{31,32} And α -catenin bridges these components to actin cytoskeleton, which recruits and organizes actin filaments.³³ Actin remodeling at the cell membrane for melanosome uptake and phagosome vesicular trafficking are key processes in keratinocyte phagocytosis.^{1,34} Previous studies on neuronal cells indicated that β -catenin played a crucial role in the recruitment, localization and distribution of synaptic vesicles in synapses.³⁵ And researches on skeletal muscle cells suggested that β -catenin was involved in regulating glucose transporter 4 containing vesicles recruitment by interacting with cadherin to support cortical actin remodeling at the cell membrane, which provided the physical structure to facilitate the movement of vesicles within the cell.³⁶ Previous work had also identified a role for β -catenin and cadherins in actin remodeling to facilitate insulin vesicle trafficking in pancreatic β -cells.³⁶ Thus, these findings suggested that β -catenin was a regulator of vesicle trafficking and acted as a signaling intermediate controlling actin remodeling in multiple tissues. In keratinocytes, it was demonstrated previously that UVB enhanced the protein expression of β -catenin.³⁷ In the present study, our results confirmed the increased accumulation of β -catenin by UVR in keratinocytes and illustrated that the UVR-induced enhancement of β -catenin was regulated by TRPA1. What is more, the alteration of β -catenin regulated by TRPA1 affected UVR-induced keratinocyte phagocytosis. Therefore, we proposed that UVR interacted with TRPA1 increased the expression of β -catenin which promoted keratinocyte phagocytosis, most likely by enhanced cell–cell adhesion and cytoskeleton dynamics mediated by β -catenin (Figure 5). In addition, our observations on UVB-induced guinea pig skin pigmentation suggested that the inhibition of β -catenin possessed skin lightening effect in vivo, which indicated the therapeutic potential of β -catenin for skin pigmentary diseases (such as melasma).

Conclusion

In conclusion, the data presented herein demonstrated that UVR promoted keratinocyte phagocytosis and skin pigmentation by TRPA1 channels. We speculate that TRPA1 is activated by UVR to promote the increase of intracellular calcium, which causes the activation of CaMKII to affect the remodeling of F-actin, contributing to keratinocyte phagocytosis. Furthermore, TRPA1 activated by UVR upregulates the expression β -catenin to enhance skin pigmentation by cell–cell adhesion and cytoskeleton dynamics. These findings suggest that TRPA1 may be a potential therapeutic target for UVR-induced skin pigmentary diseases.

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Disclosure

The authors report no conflicts of interest in relation to this work.

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