

Association of Complement C3 with Clinical Deterioration Among Hospitalized Patients with COVID-19

Hongbo Jiang^{1,*}, Qiaosen Chen^{1,*}, Shaoping Zheng², Chunxia Guo³, Jinzhuo Luo³, Hua Wang³, Xin Zheng³, Zhihong Weng³

¹Department of Epidemiology and Biostatistics, School of Public Health, Guangdong Pharmaceutical University, Guangzhou, People's Republic of China; ²Department of Ultrasound, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, People's Republic of China; ³Department of Infectious Diseases, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, People's Republic of China

*These authors contributed equally to this work

Correspondence: Zhihong Weng, Department of Infectious Diseases, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, 1277 Jiefang Avenue, Wuhan, 430022, People's Republic of China, Email wengzh@hust.edu.cn

Background: The role of the complement system in coronavirus disease 2019 (COVID-19) remains controversial. This study aimed to evaluate the relationship between serum complement C3 levels, clinical worsening, and risk of death in hospitalized patients with COVID-19.

Methods: Data were collected from 216 adults with COVID-19 admitted to a designated clinical center in Wuhan Union Hospital (China) between February 13, 2020, and February 29, 2020. Their complement C3 levels were measured within 24 h of admission. The primary outcome was a clinical worsening of 2 points on a 6-point ordinal scale. The secondary outcome was all-causes of death. Inverse probability of treatment weighting (IPTW) analysis was conducted to adjust for the baseline confounders.

Results: The median value of C3 was 0.89 (interquartile range, 0.78–1.01) g/L. Clinical worsening occurred in 12.3% (7/57) and 2.5% (4/159) of patients with baseline C3 levels < and ≥ 0.79 g/L, respectively (hazard ratio [HR], 5.22; 95% confidence interval [CI], 1.53–17.86). After IPTW adjustment, the risk for clinical worsening was 4-fold greater (weighted HR, 4.61; 95% CI, 1.16–18.4) in patients with C3 levels less than 0.79 g/L comparatively. The sensitivity analyses revealed the robustness of the results. No significant associations between C3 levels and death were observed on unadjusted (HR, 2.92; 95% CI, 0.73–11.69) and IPTW analyses (weighted HR, 3.78; 95% CI, 0.84–17.04).

Conclusion: Low complement C3 levels are associated with a higher risk for clinical worsening among inpatients with COVID-19. The serum C3 levels may contribute to the identification of patient populations that could benefit from therapeutic complement inhibition.

Keywords: SARS-CoV-2, COVID-19, complement C3, clinical worsening

Introduction

The coronavirus disease 2019 (COVID-19), which is caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), has resulted in a global pandemic and considerable mortality. The fatal outcomes from COVID-19 are attributed mainly to an overactive immune response to the SARS-CoV-2 infection.¹ However, hyperinflammation in patients with COVID-19 remains poorly clarified.

Complement, a critical component of the innate immune system, plays a key role against various pathogens, including bacteria and viruses. However, dysregulated activation of the complement can also lead to severe diseases like thrombotic microangiopathy.² It was indicated that complement activation is associated with the pathogenesis of the Middle East respiratory syndrome (MERS)^{3,4} and severe acute respiratory syndrome (SARS),⁵ which are caused by coronaviruses similar to SARS-CoV-2. A previous study used a mouse model to demonstrate that complement activation

occurred in response to SARS-CoV infection, and mice deficient in C3, a central complement component, had less systemic inflammation.⁵

Evidence from recent studies suggests that complement activation may help the pathogenesis of SARS-CoV-2 infection by triggering a severe cytokine storm.^{6,7} However, studies on the role of the complement system in COVID-19 are limited by controversial results. A study showed that low complement C3 levels were associated with poor prognosis in COVID-19 patients.⁸ However, Qin et al reported that there were no significant differences in the levels of C3 between the mild and severe groups.⁹ Moreover, other data showed that C3 levels cannot predict COVID-19 progression.¹⁰ In this study, our goal was to investigate whether complement C3 level is associated with the risk for clinical worsening or death among inpatients with COVID-19.

Materials and Methods

Study Design

In this retrospective cohort study, we collected data from patients with laboratory-confirmed COVID-19 in the Tumor Center, which is a designated clinical center for COVID-19 in the Union Hospital, Tongji Medical College of Huazhong University of Science and Technology (Wuhan, China), between February 13, 2020, and February 29, 2020. We screened patients aged ≥ 18 y with serum complement C3 levels measured within 24 h of admission. Definite outcomes (discharge or death) were followed up until March 15, 2020.

Patients included in this study were diagnosed using the guidelines of the Diagnosis and Treatment of Novel Coronavirus Pneumonia issued by the Chinese National Health Commission.¹¹ Data on demographic characteristics, clinical features, underlying diseases, laboratory examinations, treatments, and clinical outcomes were extracted from the patients' electronic medical records. Two trained physicians (CXG and JZL) evaluated them independently. This study was approved by the Ethics Commission of Tongji Medical College of Huazhong University of Science and Technology in Wuhan (2020-0058). Written informed consent was waived because of the rapidly emerging infectious disease and requirements for epidemic prevention and control. The patients' personal information was de-identified to ensure confidentiality and comply with the Declaration of Helsinki.

Exposure

The normal values of serum C3 levels may vary slightly among different laboratories. The normal value ranged from 0.80 to 1.78 g/L,¹² 0.88 to 2.01 g/L,¹³ or 0.80 to 1.60 g/L.¹⁴ Some laboratories showed normal values ranging between 0.88 and 2.52 g/L and 0.88 and 2.06 g/L for men and women, respectively.¹⁵ In our hospital, the reference range of serum complement C3 levels was 0.79–1.52 g/L for both men and women. The participants were divided into two groups based on their serum complement C3 levels tested at baseline. Patients with C3 levels less than 0.79 g/L were in the low C3 group and those with C3 levels of 0.79 g/L or greater were in the control group. The study baseline for each patient was 24 h after hospital admission.

Covariates

Potential confounding factors were chosen according to previously published articles and physician advice, including demographics (age and gender), comorbidities (hypertension, coronary heart disease [CHD], and diabetes), laboratory findings (neutrophils, lymphocytes, C-reactive protein [CRP], and D-dimer), and medications (arbidol, interferon α -2b, ribavirin, antibiotics, low molecular weight heparin, and Chinese medicine).

Outcomes

The primary outcome was clinical worsening, which was defined as all-causes of death during the hospital stay, or an increase of at least 2 points from baseline on the 6-point ordinal scale,¹⁶ whichever came first. The 6-point ordinal scale was as follows: 1 = discharge; 2 = hospitalized without supplemental oxygen; 3 = hospitalized, requiring low-flow oxygen; 4 = hospitalized, requiring noninvasive mechanical ventilation or high-flow oxygen; 5 = hospitalized, requiring

invasive ventilation or extracorporeal membrane oxygenation; and 6 = death.¹⁶ The secondary outcome was only all-causes of death.

Statistical Analysis

Categorized variables were summarized using frequency and proportion. Continuous variables were summarized using the mean and standard deviation. We used absolute standardized mean difference (aSMD) for evaluating the balance between the low C3 and control groups.

We conducted the univariate analysis to establish the relationship between low C3 and clinical worsening. In observational studies, univariate analysis is not reliable enough. Therefore, inverse probability treatment weighting (IPTW) was applied for adjusting variables to keep them comparable between the two groups.¹⁷ Meanwhile, two survival curves were generated using the Kaplan–Meier method. One was based on univariate survival analysis and the other was adjusted using IPTW analysis. For examining the robustness of the results, we performed a series of sensitivity analyses: (1) excluding the different combinations of variables that were included in the primary analysis model; (2) trimming the extreme value of weights. Then, we re-conducted the IPTW analysis. Besides clinical worsening, we explored the relationship between overall survival and low C3 with crude and IPTW analyses using the Cox proportional-hazards regression model. The formula for the IPTW modeling is provided in the [Supplementary Methods](#).

Missing data were handled using multiple imputations. Proportional hazards assumption was verified using Schoenfeld residuals test, and no evidence of violation was found. All statistical analyses were performed using the software R version 4.0.2 (<https://www.r-project.org/>).

Results

Characteristics of the Patients

A total of 955 patients with COVID-19 were admitted to this designated clinical center between February 13, 2020, and February 29, 2020. Among them, 728 had no information regarding serum complement C3 levels in their electronic medical records, and 11 were not tested within the 24 h window. Therefore, 216 patients with C3 levels measured within 24 h of admission were included in this study ([Figure 1](#)). The median value of C3 level was 0.89 (interquartile range [IQR], 0.78–1.01) g/L. Of the 216 patients, 57 (26.4%) and 159 (73.6%) had C3 levels < and $\geq$ 0.79 g/L, respectively. The baseline demographics and clinical characteristics of the low C3 and control groups are summarized in [Table 1](#).

Primary Outcome

Clinical outcomes of the 216 patients on 15 March 2020, according to the 6-point ordinal scale, are shown in [Supplementary Table 1](#). In the crude analysis, the cumulative incidences of clinical worsening were 12.3% (7/57) and 2.5% (4/159) among patients with baseline C3 levels < and $\geq$ 0.79 g/L, respectively. Clinical worsening was frequent among patients in the low C3 group compared to the control group (HR, 5.22; 95% CI, 1.53–17.86) ([Figure 2](#), left panel).

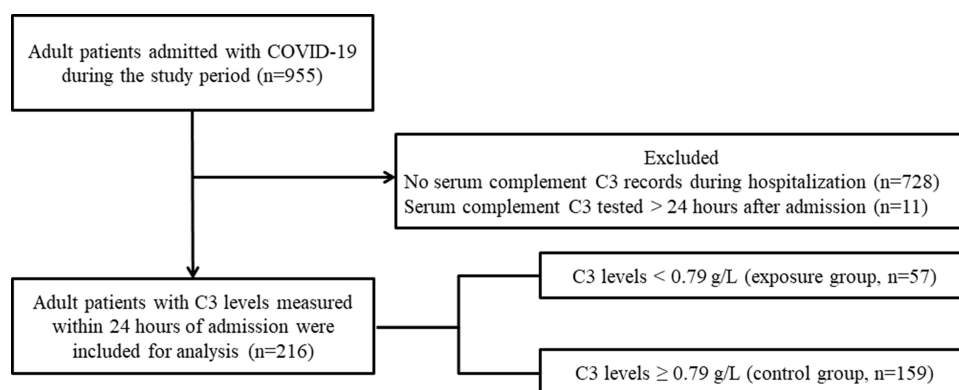


Figure 1 Flow chart of study participants. The study baseline was defined as 24 h after admission. COVID-19 denotes coronavirus disease 2019.

Table 1 Baseline Characteristics of COVID-19 Patients with Low C3 Levels or Not, Before and After Inverse Probability Treatment Weighting

Variable	Before Weighting			aSMD	After Weighting			aSMD
	Low C3 Group	Control Group	Overall		Low C3 Group	Control Group	Overall	
	(N = 57)	(N = 159)	(N = 216)		(N = 57)	(N = 158)	(N = 215)	
Age, yrs, mean (SD)	65.67 (15.00)	61.19 (12.72)	62.38 (13.47)	0.33	62.57 (15.81)	62.12 (12.29)	62.24 (13.27)	0.03
Gender, n (%)	–	–	–	0.01	–	–	–	0.03
Male	26 (45.6)	72 (45.3)	98 (45.4)	–	26.5 (46.8)	71.8 (45.4)	98.3 (45.8)	–
Female	31 (54.4)	87 (54.7)	118 (54.6)	–	30.1 (53.2)	86.2 (54.6)	116.3 (54.2)	–
Comorbidities								
Hypertension, n (%)	7.0 (12.3)	25.0 (15.7)	32.0 (14.8)	0.10	9.4 (16.6)	24.0 (15.2)	33.4 (15.6)	0.04
Diabetes, n (%)	4.0 (7.0)	11.0 (6.9)	15.0 (6.9)	<0.01	3.2 (5.6)	10.2 (6.5)	13.4 (6.2)	0.04
Coronary heart disease, n (%)	2.0 (3.5)	7.0 (4.4)	9.0 (4.2)	0.04	1.8 (3.2)	6.2 (3.9)	8.0 (3.7)	0.04
Laboratory tests								
Neutrophils $\leq 6.3 \times 10^9/L$, n (%)	3.0 (5.3)	17.0 (10.7)	20.0 (9.3)	0.19	4.9 (8.7)	14.6 (9.2)	19.5 (9.1)	0.02
Lymphocytes $\leq 1.1 \times 10^9/L$, n (%)	29.0 (50.9)	109.0 (68.6)	138.0 (63.9)	0.37	33.8 (59.7)	101.2 (64.1)	135.0 (62.9)	0.09
D-dimer $\leq 1\text{mg/L}$, n (%)	16.0 (36.4)	38.0 (29.5)	54.0 (31.2)	0.09	13.6 (30.4)	38.9 (30.2)	52.5 (30.2)	0.02
C-reactive protein $\leq 8\text{mg/L}$, n (%)	24.0 (42.1)	64.0 (40.5)	88.0 (40.9)	0.02	23.8 (42.1)	63.0 (40.1)	86.8 (40.6)	0.03
Treatments								
LMWH, n (%)	47.0 (82.5)	141.0 (88.7)	188.0 (87.0)	0.18	50.4 (89.0)	139.4 (88.2)	189.7 (88.4)	0.02
Arbidol, n (%)	53.0 (93.0)	152.0 (95.6)	205.0 (94.9)	0.12	53.2 (94.0)	149.2 (94.5)	202.5 (94.4)	0.02
Ribavirin, n (%)	7.0 (12.3)	22.0 (13.8)	29.0 (13.4)	0.05	6.7 (11.8)	22.0 (13.9)	28.6 (13.3)	0.06
Interferon α -2b, n (%)	13.0 (22.8)	33.0 (20.8)	46.0 (21.3)	0.05	11.9 (21.0)	33.1 (20.9)	44.9 (20.9)	<0.01
Chinese medicine, n (%)	45.0 (78.9)	148.0 (93.1)	193.0 (89.4)	0.46	50.6 (89.3)	141.7 (89.7)	192.3 (89.6)	0.01
Antibiotics, n (%)	44.0 (77.2)	120.0 (75.5)	164.0 (75.9)	0.04	43.2 (76.3)	119.3 (75.5)	162.5 (75.7)	0.01

Abbreviations: COVID-19, coronavirus disease 2019; aSMD, absolute standardized mean difference; LMWH, low molecular weight heparin.

Similarly, there was a significant difference in the cumulative incidences of clinical worsening between the two groups in the IPTW analysis (weighted HR, 4.61; 95% CI, 1.16–18.4) (Figure 2, right panel). The distribution of weight and aSMD of variables in the IPTW analysis for clinical worsening are shown in [Supplementary Figures 1](#) and [2](#), respectively.

Secondary Outcome

Over a median follow-up of 29.1 d (IQR, 28.6–31.2 d), the secondary outcome developed in eight (3.7%) of the 216 patients. Four patients died in both the low C3 (0.081 case/person-month) and control groups (0.027 case/person-month). At the end of the follow-up on 15 March 2020, 205 (94.9%) patients were discharged alive, and three (1.4%) remained in the hospital.

Figure 3 shows the Kaplan–Meier curves. Both the unadjusted (HR, 2.92; 95% CI, 0.73–11.69) and IPTW analyses (weighted HR, 3.78; 95% CI, 0.84–17.04) showed insignificant differences between the two groups. However, they indicated that the low C3 population might be associated with a higher risk of death.

Sensitivity Analysis

Sensitivity analysis for the primary outcome was robust under different contexts of the model. Some important variables, including D-dimer, CRP, diabetes, CHD, and hypertension, were excluded intentionally and the conclusion remained unchanged (Table 2). Therefore, it is rational to assume that the bias would have been less even if we neglected these variables in the primary analysis. Moreover, we added the weight-trimming analysis to simulate a scenario where the positivity assumption did not hold up and found that the estimate was still robust (Table 2).

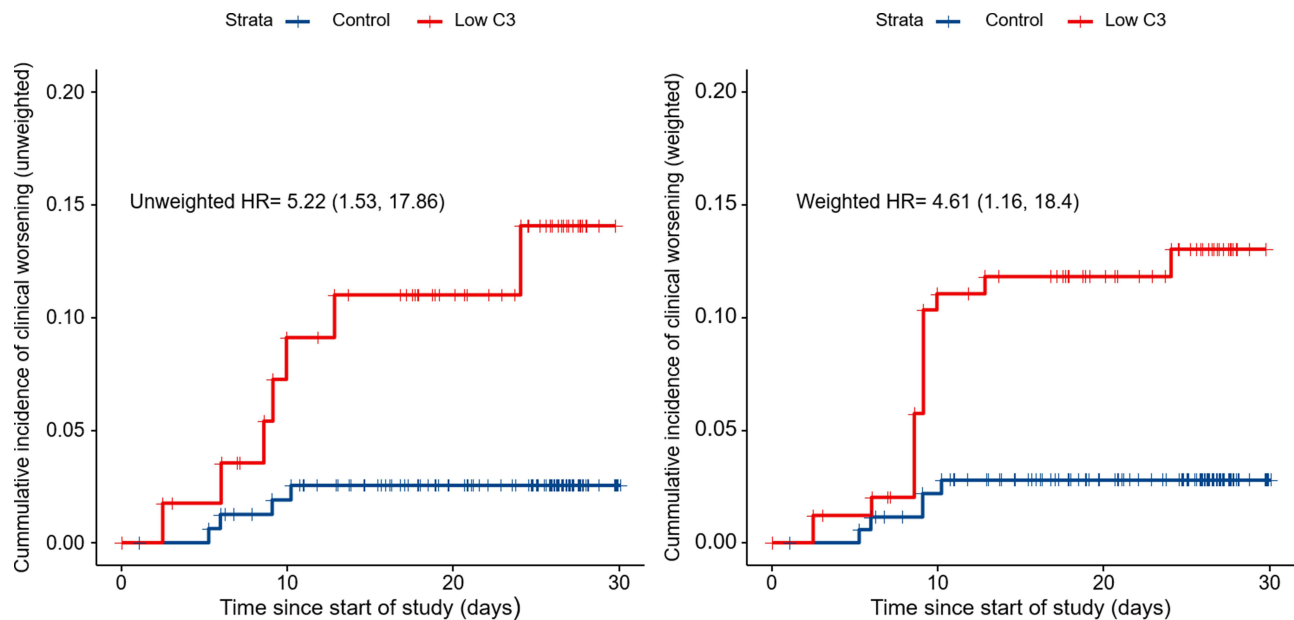


Figure 2 Kaplan–Meier curves for clinical worsening during follow-up, both before and after the inverse probability of treatment weighting (IPTW). There was a significant association between the baseline serum C3 levels and clinical worsening on unadjusted (crude HR, 5.22; 95% CI, 1.53–17.86) and IPTW analyses (weighted HR, 4.61; 95% CI, 1.16–18.4).

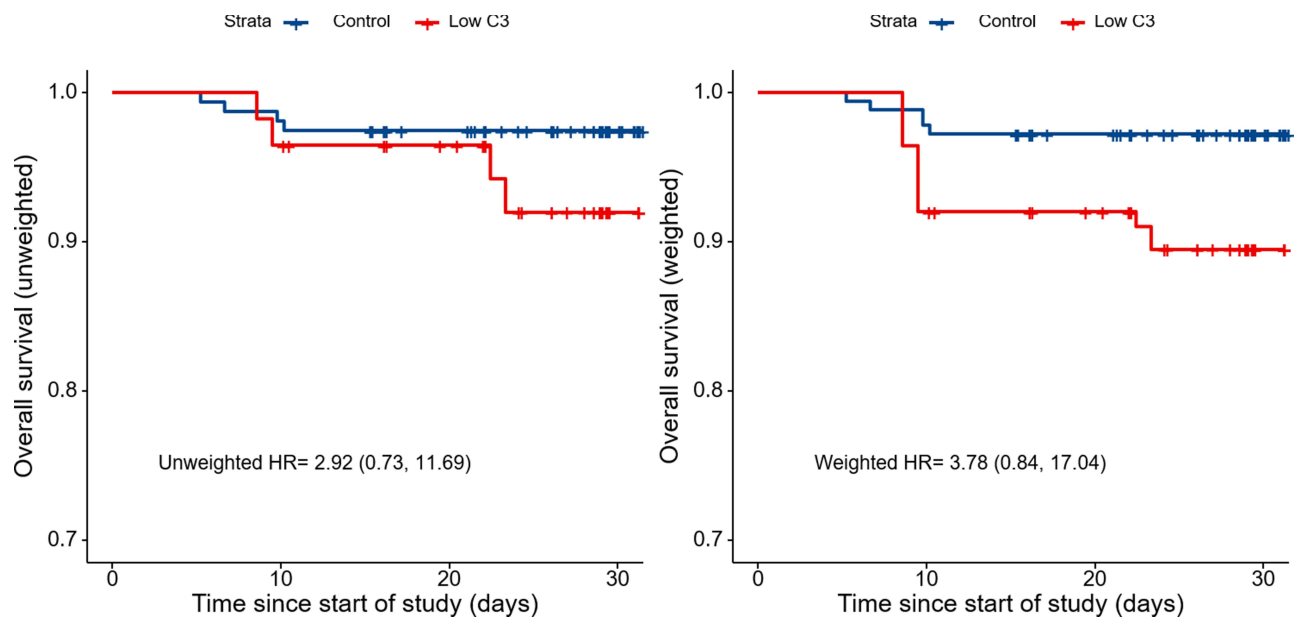


Figure 3 Kaplan–Meier curves for overall survival during follow-up, both before and after the inverse probability of treatment weighting (IPTW). There was no significant association between the baseline serum C3 levels and death on unadjusted (crude HR, 2.92; 95% CI, 0.73–11.69) and IPTW analyses (weighted HR, 3.78; 95% CI, 0.84–17.04).

Discussion

It has been hypothesized that complement activation may contribute to the pathogenesis of COVID-19. Our study provides clinical evidence based on real-world data to support such a theoretical perspective. In this observational study, we investigated the relationship between serum complement C3 levels, clinical worsening, and risk of death in a retrospective cohort of 216 hospitalized patients with confirmed COVID-19. Our results suggested that in COVID-19 patients who have baseline C3 levels <0.79 g/L, there was a 4-fold greater increase in the risk for clinical worsening

Table 2 Summarization of Different Analytical Strategies to Explore the Associations of Low C3 with Outcomes of COVID-19 Patients

Analytical Strategy	HR (95% CI)
Clinical worsening (primary analysis)	
Crude analysis	5.22 (1.53–17.86)
IPTW analysis	4.61 (1.16–18.40)
Clinical worsening (sensitivity analysis-ignoring important variable)	
D-dimer excluded	4.52 (1.14–17.94)
D-dimer and C-reactive protein excluded	4.27 (1.08–16.96)
Diabetes excluded	4.78 (1.21–18.92)
Diabetes and coronary heart disease excluded	4.89 (1.23–19.43)
Diabetes, coronary heart disease and hypertension excluded	4.07 (1.02–16.23)
Clinical worsening (sensitivity analysis-weight-trimming analysis)	
0.5% and 99.5% weights trimming	4.45 (1.12–17.58)
1.0% and 99.0% weights trimming	4.38 (1.11–17.25)
1.5% and 98.5% weights trimming	4.36 (1.11–17.13)
2.0% and 98.0% weights trimming	4.35 (1.11–17.04)
2.5% and 97.5% weights trimming	4.32 (1.11–16.84)
5.0% and 95.0% weights trimming	4.23 (1.13–15.80)
7.5% and 92.5% weights trimming	4.35 (1.20–15.78)
10.0% and 90.0% weights trimming	4.65 (1.31–16.45)
Overall survival (secondary analysis)	
Crude analysis	2.92 (0.73–11.69)
IPTW analysis	3.78 (0.84–17.04)

Abbreviations: COVID-19, coronavirus disease 2019; HR, hazard ratio; CI, confidence interval; IPTW, inverse probability of treatment weighting.

when compared to patients with C3 levels ≥ 0.79 g/L (weighted HR, 4.61; 95% CI, 1.16–18.4). The sensitivity analyses revealed the robustness of the results as well.

The complement system, a bridge between innate and adaptive immunity, can identify and eliminate invading viruses through opsonization, promoting virus-specific immune responses and neutralizing cell-free viruses.^{18,19} After activation, the mature complement C3 is cleaved to generate C3a and C3b. Further cleavage of C3b produces iC3b, C3f, C3c, C3dg, C3d, and C3g.²⁰ C3 acts as a point of convergence among the three complement pathways and plays a central role in the process of complement activation. This results in the amplification of the complement response and helps coordinate downstream immune responses. It has become evident that C3 participates in clearing pathogens and a variety of homeostatic processes, such as tissue regeneration and controlling tumor progression.²¹

Hypocomplementemia was considered to be associated with the consumption of complement components because of complement activation.²² The levels of C3 (not C4) reportedly decreased significantly in patients with septic shock compared to critically ill patients.²³ Similarly, hypocomplementemia was demonstrated in various viral infections, such as parvovirus B19, Ebola virus, and hepatitis C.^{24–26} C3 consumption was described in two children with SARS-CoV-2-induced Kawasaki-like hyperinflammatory syndrome. In Phase 2 of their illness, both patients developed progressive thrombocytopenia, hepatomegaly, and diffuse edema. Fortunately, they improved after corticosteroid treatment.²⁷ Another study reported that very low C3 levels were observed in a child with pediatric inflammatory multisystem syndrome because of SARS-CoV-2 infection. After being treated with high-dose intravenous immunoglobulin and methylprednisolone, the child recovered and the C3 levels returned to normal.²⁸ These studies suggest that complement activation by C3 hypocomplementemia may be implicated in the pathophysiology of COVID-19.

A retrospective study showed that serum complement C3 levels cannot predict COVID-19 progression.¹⁰ In contrast, Fang et al reported that decreased complement C3 was correlated with increased odds of death.⁸ However, in the study by Fang et al, adjustment was only performed for age, sex, laboratory findings, and comorbidities. The lack of adjustment for

treatment regimens could affect the prognosis of the patients, resulting in a confounding bias. It was reported that plasma levels of complement C3 are associated with the development of hypertension,²⁹ diabetes,³⁰ and coronary heart disease.³¹ This may account for the higher comorbidity in the control group with relatively higher C3 levels. Therefore, we adjusted for confounders using the IPTW method, including age, sex, comorbidities, laboratory tests, and medications on admission. According to the Cox regression analysis with IPTW, there was a significant inverse correlation between C3 hypocomplementemia and clinical worsening in patients with COVID-19. A series of sensitivity analyses also generated supportive evidence. Moreover, our results indicate that the low C3 population might be experiencing a higher risk of death. No significant associations were observed between the baseline C3 levels and death in the unadjusted and IPTW analyses.

Emerging evidence suggests that complement C3 activation might be triggered by virus nucleocapsid protein through virus-released pathogen-associated molecular patterns during SARS-CoV-2 infection.^{32,33} Complement and tissue factor (TF)-enriched neutrophil extracellular traps (NETs) are crucial drivers of immunothrombosis. Moreover, C3 blockade can disrupt TF expression in neutrophils, inhibit complement activation, and attenuate platelet-mediated NET-induced thrombogenicity in COVID-19.⁹ Compstatin, a highly selective C3 inhibitor, can inhibit complement activation by binding to C3 and interfering with the convertase formation and C3 cleavage.^{34,35} The latest study reported that treatment with the compstatin was safe and successful in a patient with severe acute respiratory distress syndrome (ARDS) following SARS-CoV-2 infection.³⁶ Another exploratory study in small independent cohorts of severe COVID-19 patients indicated that C3 inhibitor (AMY-101) can abrogate COVID-19 hyperinflammation. This results in the decline of CRP and IL-6 levels, improvement of lung function, and resolution of SARS-CoV-2-associated ARDS.³⁷ These early clinical results support the evaluation of C3-targeting agents as COVID-19 therapeutics in large prospective trials. The results of our study suggest that the complement system is activated by the consumption of C3 during SARS-CoV-2 infection, and COVID-19 patients with C3 hypocomplementemia might benefit from complement inhibition treatment.

There are several limitations to our study. First, this is a single-center observational study with small sample size. Therefore, the results cannot be generalized, and large-scale randomized controlled trials are warranted to assess the associations of C3 hypocomplementemia with clinical worsening and outcomes of COVID-19. Second, all patients with COVID-19 in this designated clinical center were transferred from other hospitals during the outbreak in Wuhan, and the timing of serum C3 level measurement may be inappropriate for indicating the status of complement activation because each patient was at a different stage of the SARS-CoV-2 infection on admission. The longitudinal C3 levels could describe the course of complement activation more accurately compared to the values obtained at baseline. According to the dynamic changes in C3 levels, a reasonable determination can be made regarding the optimal time window for complement intervention. Third, in this designated clinical center, the serum complement C3 level was not routinely measured within 24 h of admission for every COVID-19 patient. Only patients with C3 levels tested at baseline were considered for this study, which might introduce a certain kind of selection bias.

Conclusion

In this preliminary study, our results suggest that low serum complement C3 level is associated with a higher risk for clinical worsening among hospitalized patients with COVID-19. We hypothesize that the C3 level may contribute to the identification of patient populations that could benefit from therapeutic complement inhibition.

Acknowledgments

We thank all health care workers in the Department of Infectious Diseases, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology in Wuhan. We acknowledge all the medical staff on the front line in Wuhan. We also thank Bullet Edits for its linguistic assistance during the preparation of this manuscript.

Funding

There is no funding to report.

Disclosure

The authors report no conflicts of interest in this work.

References

- Wiersinga WJ, Rhodes A, Cheng AC, Peacock SJ, Prescott HC. Pathophysiology, transmission, diagnosis, and treatment of coronavirus disease 2019 (COVID-19): a review. *JAMA*. 2020;324. doi:10.1001/jama.2020.12839.
- Nesargikar PN, Spiller B, Chavez R. The complement system: history, pathways, cascade and inhibitors. *Eur J Microbiol Immunol*. 2012;2:103–111. doi:10.1556/EuJMI.2.2012.2.2
- Jiang Y, Zhao G, Song N, et al. Blockade of the C5a-C5aR axis alleviates lung damage in hDPP4-transgenic mice infected with MERS-CoV. *Emerg Microbes Infect*. 2018;7(1):77. doi:10.1038/s41426-018-0063-8
- Jiang Y, Li J, Teng Y, et al. Complement receptor C5aR1 inhibition reduces pyroptosis in hDPP4-transgenic mice infected with MERS-CoV. *Viruses*. 2019;11(1):39. doi:10.3390/v11010039
- Gralinski LE, Sheahan TP, Morrison TE, et al. Complement activation contributes to severe acute respiratory syndrome coronavirus pathogenesis. *mBio*. 2018;9(5):e01753–18. doi:10.1128/mBio.01753-18
- Polycarpou A, Howard M, Farrar CA, et al. Rationale for targeting complement in COVID-19. *EMBO Mol Med*. 2020;12(8):e12642. doi:10.15252/emmm.202012642
- Skendros P, Mitsios A, Chrysanthopoulou A, et al. Complement and tissue factor-enriched neutrophil extracellular traps are key drivers in COVID-19 immunothrombosis. *J Clin Invest*. 2020;130(11):6151–6157. doi:10.1172/JCI141374
- Fang S, Wang H, Lu L, Jia Y, Xia Z. Decreased complement C3 levels are associated with poor prognosis in patients with COVID-19: a retrospective cohort study. *Int Immunopharmacol*. 2020;89(Pt A):107070. doi:10.1016/j.intimp.2020.107070
- Qin C, Zhou L, Hu Z, et al. Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China. *Clin Infect Dis*. 2020;71(15):762–768. doi:10.1177/0961203310371154
- Zhang Z, Li X, Zhang W, Shi ZL, Zheng Z, Wang T. Clinical features and treatment of 2019-nCoV pneumonia patients in Wuhan: report of a couple cases. *Virol Sin*. 2020;35(3):330–336. doi:10.1007/s12250-020-00203-8
- China NHC. Guidelines of the diagnosis and treatment of novel coronavirus pneumonia (trial version 8). in Chinese. Available from: <http://www.nhc.gov.cn/yzygj/s7653p/202008/0a7bdf12bd4b46e5bd28ca7f9a7f5e5a.shtml>. Accessed January 15, 2022
- Birmingham DJ, Irshaid F, Nagaraja HN, et al. The complex nature of serum C3 and C4 as biomarkers of lupus renal flare. *Lupus*. 2010;19(11):1272–1280. doi:10.1177/0961203310371154
- Complement component 3 (C3) [website on the Internet]. UCSF Benioff Children's Hospitals; 2022. Available from: <https://www.ucsfbenioffchildrens.org/medical-tests/003539>. Accessed January 15, 2022.
- How to Read Complement C3 Test Report [website on the Internet]. MediFee.com; 2022. Available from: <https://www.mediffee.com/how-to-read/complement-c3-test-report>. Accessed January 15, 2022.
- C3 And C4 Blood Test [website on the Internet]. PORTEA – HEAL AT HOME; 2022. Available from: <https://www.portea.com/labs/diagnostic-tests/c3-and-c4-blood-test-204/>. Accessed January 15, 2022.
- Grein J, Ohmagari N, Shin D, et al. Compassionate use of remdesivir for patients with severe COVID-19. *N Engl J Med*. 2020;382(24):2327–2336. doi:10.1056/NEJMoa2007016
- Stuart EA, Lee BK, Leacy FP. Prognostic score-based balance measures can be a useful diagnostic for propensity score methods in comparative effectiveness research. *J Clin Epidemiol*. 2013;66(8 Suppl):S84–S90.e1. doi:10.1016/j.jclinepi.2013.01.013
- West EE, Kolev M, Kemper C. Complement and the regulation of T cell responses. *Annu Rev Immunol*. 2018;36:309–338. doi:10.1146/annurev-immunol-042617-053245
- Nasir A, Caetano-Anolles G. A phylogenomic data-driven exploration of viral origins and evolution. *Sci Adv*. 2015;1(8):e1500527. doi:10.1126/sciadv.1500527
- Janssen BJ, Huizinga EG, Raaijmakers HC, et al. Structures of complement component C3 provide insights into the function and evolution of immunity. *Nature*. 2005;437(7058):505–511. doi:10.1038/nature04005
- Ricklin D, Reis ES, Mastellos DC, Gros P, Lambris JD. Complement component C3 – the “Swiss Army Knife” of innate immunity and host defense. *Immunol Rev*. 2016;274(1):33–58. doi:10.1111/imr.12500
- Atsumi T, Amengual O, Koike T. Antiphospholipid syndrome: pathogenesis. In: *Systemic Lupus Erythematosus*. Fifth ed. Academic Press; 2011:945–965.
- Lin RY, Astiz ME, Saxon JC, Saha DC, Rackow EC. Alterations in C3, C4, factor B, and related metabolites in septic shock. *Clin Immunol Immunopathol*. 1993;69(2):136–142. doi:10.1006/clin.1993.1161
- Hashizume H, Kageyama H. Hypocomplementemia is a diagnostic clue for parvovirus B19 infection in adults. *J Dermatol*. 2017;44(3):e27. doi:10.1111/1346-8138.13469
- Brudner M, Karpel M, Lear C, et al. Lectin-dependent enhancement of Ebola virus infection via soluble and transmembrane C-type lectin receptors. *PLoS One*. 2013;8:e60838. doi:10.1371/journal.pone.0060838
- Itoh K, Tanaka H, Shiga J, et al. Hypocomplementemia associated with hepatitis C viremia in sera from voluntary blood donors. *Am J Gastroenterol*. 1994;89(11):2019–2024.
- Licciardi F, Pruccoli G, Denina M, et al. SARS-CoV-2-induced Kawasaki-like hyperinflammatory syndrome: a novel COVID phenotype in children. *Pediatrics*. 2020;146(2):e20201711. doi:10.1542/peds.2020-1711
- Regev T, Antebi M, Eytan D, et al. Pediatric inflammatory multisystem syndrome with central nervous system involvement and hypocomplementemia following SARS-COV-2 infection. *Pediatr Infect Dis J*. 2020;39(8):e206–e207. doi:10.1097/INF.0000000000002804
- Engström G, Hedblad B, Berglund G, Janzon L, Lindgärde F. Plasma levels of complement C3 is associated with development of hypertension: a longitudinal cohort study. *J Hum Hypertens*. 2007;21(4):276–282. doi:10.1038/sj.jhh.1002129
- Engström G, Hedblad B, Eriksson KF, Janzon L, Lindgärde F. Complement C3 is a risk factor for the development of diabetes: a population-based cohort study. *Diabetes*. 2005;54(2):570–575. doi:10.2337/diabetes.54.2.570

31. Engström G, Hedblad B, Janzon L, Lindgärde F. Complement C3 and C4 in plasma and incidence of myocardial infarction and stroke: a population-based cohort study. *Eur J Cardiovasc Prev Rehabil.* 2007;14(3):392–397. doi:10.1097/01.hjr.0000244582.30421.b2
32. Risitano AM, Mastellos DC, Huber-Lang M, et al. Complement as a target in COVID-19? *Nat Rev Immunol.* 2020;20(6):343–344. doi:10.1038/s41577-020-0320-7
33. Gao T, Hu MD, Zhang XP, Li HZ, Cao C. Highly pathogenic coronavirus N protein aggravates lung injury by MASP-2-mediated complement over-activation. *medRxiv.* 2020. doi:10.1101/2020.03.29.20041962
34. Mastellos DC, Yancopoulou D, Kokkinos P, et al. Compstatin: a C3-targeted complement inhibitor reaching its prime for bedside intervention. *Eur J Clin Invest.* 2015;45(4):423–440. doi:10.1111/eci.12419
35. Mastellos DC, Ricklin D, Lambris JD. Clinical promise of next-generation complement therapeutics. *Nat Rev Drug Discov.* 2018;18:707–729. doi:10.1038/s41573-019-0031-6
36. Mastaglio S, Ruggeri A, Risitano AM, et al. The first case of COVID-19 treated with the complement C3 inhibitor AMY-101. *Clin Immunol.* 2020;215:108450. doi:10.1016/j.clim.2020.108450
37. Mastellos DC, Pires da Silva BGP, Fonseca BAL, et al. Complement C3 vs C5 inhibition in severe COVID-19: early clinical findings reveal differential biological efficacy. *Clin Immunol.* 2020;220:108598. doi:10.1016/j.clim.2020.108598

International Journal of General Medicine

Dovepress

Publish your work in this journal

The International Journal of General Medicine is an international, peer-reviewed open-access journal that focuses on general and internal medicine, pathogenesis, epidemiology, diagnosis, monitoring and treatment protocols. The journal is characterized by the rapid reporting of reviews, original research and clinical studies across all disease areas. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/international-journal-of-general-medicine-journal>