

Integrated Systems Pharmacology and Surface Plasmon Resonance Approaches to Reveal the Synergistic Effect of Multiple Components of Gu-Ben-Ke-Chuan Decoction on Chronic Bronchitis

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Introduction: Gu-Ben-Ke-Chuan (GBKC) decoction, a well-known prescription composed of seven herbs, has been widely used for treating chronic bronchitis (CB). However, the pharmacological constituents of GBKC and the underlying mechanisms by which these components act on CB remain unclear.

Methods: Ultra-high-pressure liquid chromatography coupled with linear ion trap–Orbitrap tandem mass spectrometry (UHPLC-LTQ-Orbitrap) was first employed to rapidly identify compounds from GBKC. Thereafter, network pharmacology and molecular docking analyses were performed to identify the potential active constituents, candidate targets, and major pathways. Finally, the affinities between the key compounds and targets were verified via surface plasmon resonance (SPR) analysis. In addition, the anti-inflammatory effect of GBKC was verified using an LPS-induced inflammatory cell model based on the predicted results.

Results: A total of 53 major compounds were identified in the GBKC decoction. After network pharmacology-based virtual screening, 141 major targets and 39 main compounds were identified to be effective in the treatment of CB. The major targets were highly enriched in the tumor necrosis factor (TNF) signaling pathway, suggesting that GBKC could attenuate the inflammatory response in patients with CB. Furthermore, molecular docking results indicated that 20 pairs of components and target proteins relevant to the TNF pathway exhibited notable interactions. Among them, eight compound-target pairs exhibited good affinity as per SPR analysis. In addition, the production of interleukin 6 and TNF- α in LPS-induced MH-S cells was suppressed after GBKC treatment.

Conclusion: This study successfully clarified the mechanism of action of GBKC against CB, which demonstrated that the integrated strategy described above is reliable for identifying the active compounds and mechanisms responsible for the pharmacological activities of GBKC decoction.

Keywords: Gu-Ben-Ke-Chuan decoction, chronic bronchitis, UHPLC-LTQ-Orbitrap, network pharmacology, surface plasmon resonance

Introduction

Chronic bronchitis (CB), a type of chronic obstructive pulmonary disease, is defined as chronic cough and sputum production for 3 months/year for two consecutive years.¹ CB remains the fourth largest cause of hospitalizations and death worldwide, and has brought a heavy health burden and soaring healthcare costs to

individuals, families, and society.² Many environmental and host factors may contribute to CB progression or exacerbation, such as tobacco smoke, infection, occupational hazards, and poor lung development during childhood.³ Although the number of patients with CB keeps increasing, its pathophysiological mechanism is not yet clearly understood. Current pharmacological treatments for CB, including antibiotics and corticosteroids, may compromise the efficacy and safety characteristics of these agents, resulting in unacceptable long-term adverse reactions, such as diarrhea, nausea, osteoporosis, pneumonia, and psychiatric disturbances;⁴⁻⁶ therefore, new anti-CB medications with good efficacy and safety are urgently required.

Gu-Ben-Ke-Chuan (GBKC) decoction, a well-known traditional Chinese herbal formula, contains seven herbs such as *Radix Codonopsis* (“Dang-Shen” in Chinese, DS), *Rhizoma Atractylodis Macrocephalae* (“Bai-Zhu” in Chinese, BZ), *Poria cocos* (“Fu-Ling” in Chinese, FL), *Radix Ophiopogonis* (“Mai-Dong” in Chinese, MD), *Fructus Psoraleae* (“Bu-Gu-Zhi” in Chinese, BGZ), *Glycyrrhizae Radix et Rhizoma* (“Gan-Cao” in Chinese, GC), and *Schisandrae Chinensis Fructus* (“Wu-Wei-Zi” in Chinese, WWZ) at the weight ratio of 6:6:4:6:6:3:3. This traditional Chinese medicine (TCM) prescription has been recorded in the Chinese Pharmacopeia and approved by the China Food and Drug Administration for the treatment of airway diseases, such as asthma, CB, and bronchiectasis.⁷ In recent years, accumulating evidence has demonstrated that GBKC exerts substantial therapeutic effects on CB.⁸⁻¹⁰ However, its underlying mechanisms of action are still largely unknown.

As a relatively new discipline in modern pharmacological studies, network pharmacology has been proposed as a promising approach to obtain a systematic understanding of the complex relationships between drug and disease, which adheres to the holistic philosophy of traditional Chinese medicine (TCM).¹¹ To date, this method has been successfully used to reveal the synergistic effects and molecular mechanisms of TCM in many complicated diseases, such as Alzheimer’s disease,¹¹ cardiovascular disease,^{12,13} diabetes,¹⁴ uveitis,¹⁵ cancer,¹⁶ and gastrointestinal diseases.¹⁷

Surface plasmon resonance (SPR) biosensor serves as a powerful tool for label-free and real-time analysis of the interaction between drug and protein;^{18,19} it can quickly and easily monitor the association and

disassociation of molecules.²⁰ In addition, SPR can directly screen active compounds from complex matrices.²¹ Consequently, optical biosensor methods based on SPR have been extensively used in TCM research, such as for lead compound discovery and target identification.²²

In this study, the main chemical compounds were first detected and identified using UHPLC-LTQ-Orbitrap. Second, a network pharmacology-based strategy was used to explore the active components and major targets of GBKC for treating CB. Third, high-throughput molecular docking studies using LibDock were performed to confirm the binding activities between the active components and the related targets. Fourth, the specific interactions between the targets and key components were investigated and verified using SPR methodologies. Finally, the anti-inflammatory function of GBKC was further verified via in vitro cell experiments based on the predicted results. We hope that this study will not only reveal the anti-CB mechanism of GBKC, but also provide a rapid and reliable strategy for screening active compounds from medicinal plants and prescriptions. The workflow of this study is illustrated in Figure 1.

Materials and Methods

Materials and Reagents

The seven herbs, including *Radix Codonopsis*, *Rhizoma Atractylodis Macrocephalae*, *Poria cocos*, *Radix Ophiopogonis*, *Fructus Psoraleae*, *Glycyrrhizae Radix et Rhizoma*, and *Schisandrae Chinensis Fructus* were purchased from Beijing Tongrentang and authenticated by Professor Jingjuan Wang (Beijing University of Chinese Medicine). Chromatographic-grade acetonitrile was supplied by Fisher Scientific. Ultrapure water was prepared using the Millipore Milli Q plus purification system. All other reagents used were of analytical grade and commercially available.

The mouse alveolar macrophage cell line MH-S was purchased from Peking Union Medical College (Beijing, China). RPMI-1640 culture medium and trypsin were provided by Corning Inc. (New York, NY, USA). Fetal bovine serum (FBS) and a mixture of penicillin and streptomycin were purchased from Gibco (Grand Island, NY, USA). Lipopolysaccharide (*Escherichia coli* 055:B5, LPS) was purchased from Yuanye Bio-Technology Co., Ltd. (Shanghai, China). Cell Counting

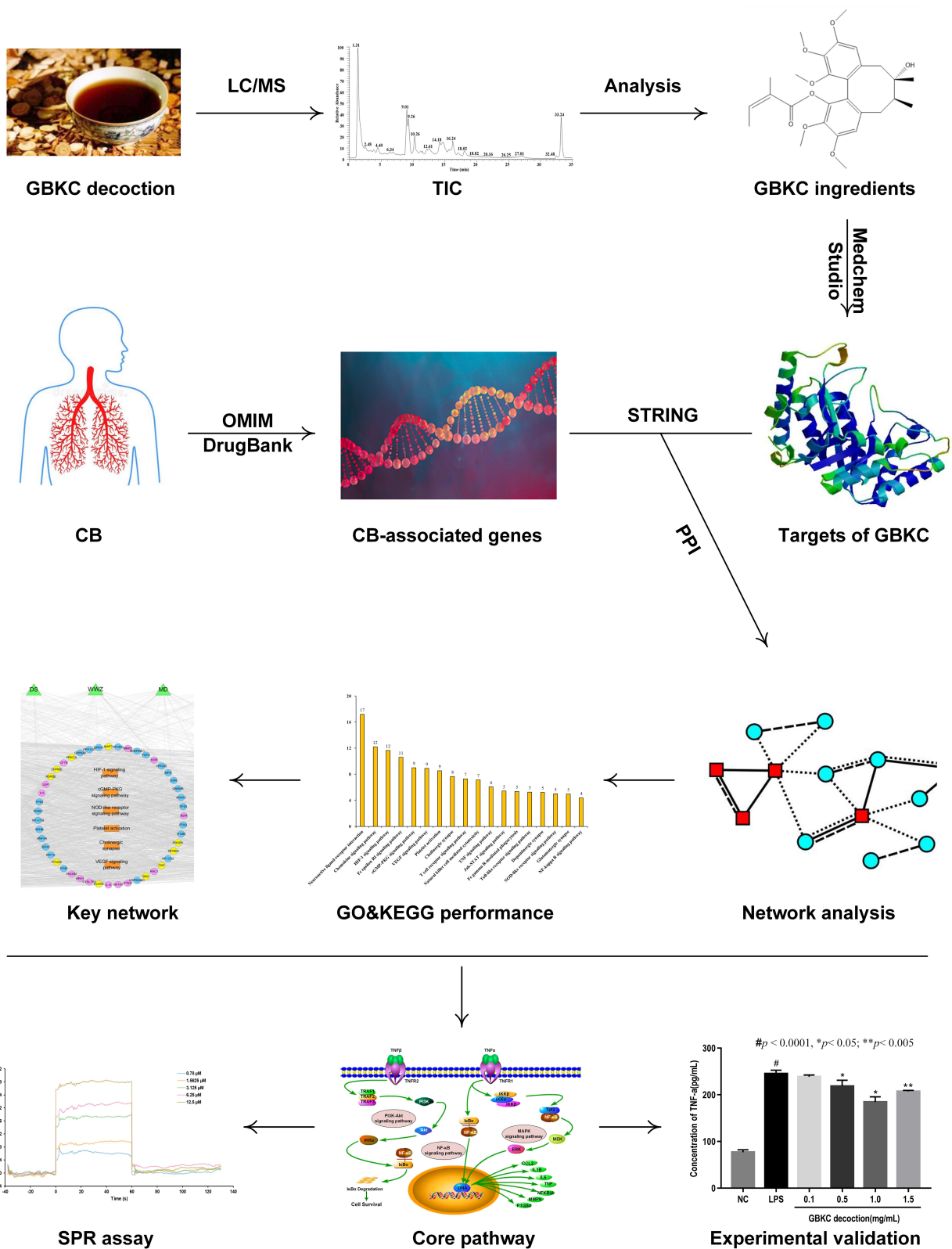


Figure 1 Experimental design and schematic diagram of the workflow of this study.

Kit-8 (CCK-8) was purchased from KeyGEN Bio-Technology Co., Ltd. (Nanjing, China). Enzyme-linked immunosorbent assay (ELISA) kit for TNF- α and IL-6 were purchased from Proteintech Group, Inc. (Chicago, IL, USA).

Preparation of GBKC Decoction

DS (30g), BZ (30g), FL (20g), MD (30g), BGZ (30g), GC (15g) and WWZ (15g) were mixed and immersed with six-fold mass of water for 1 h. The mixtures were boiled twice, 1 h for each time. The extracted water solutions were combined, filtered, and concentrated to 2 g/mL, and stored at -20 °C until use. For chemical analysis, the GBKC decoction (2 g/mL) was diluted to 10 mg/mL (w/v, crude drug/water) and filtered using a 0.22 μ m membrane filter before UHPLC-LTQ-Orbitrap analysis. For cell experiments, the GBKC decoction (2 g/mL) was sterilized and diluted to appointed concentrations (0.1, 0.5, 1.0, 1.5 mg/mL) with cell culture medium.

LC/MS Analysis

For LC/MS experiments, an LTQ-Orbitrap Velos mass spectrometer equipped with an electrospray ionization (ESI) source was connected to a Thermo Dionex Ultimate 3000 UHPLC system (including an auto-sampler, a thermostatically controlled column compartment and a binary pump coupled to a vacuum degasser). The Xcalibur, Networks and Mass Frontier 6.0 software packages (Thermo Fisher Scientific) were employed for data recording and processing.

LC analysis was conducted on a Thermo Scientific Hypersil BDS C₁₈ (2.1 mm \times 150 mm, 2.4 μ m) column maintained at 35°C. The mobile phase system was composed of water with 0.1% formic acid (A) and acetonitrile (B). The flow rate was set to 0.3 mL/min with a linear gradient as follows: 0–6.0 min (3–3% B), 6.0–13.0 min (3–15% B), 13.0–23.0 min (15–40% B), 23.0–25.0 min (40–65% B), 25.0–30.0 min (65–97% B), 30.0–30.5 (97–3% B) and 30.5–35.0 (3–3% B). The injection volume was 3 μ L.

The MS analysis was conducted on both the negative and positive ion modes. The capillary temperature was 350 °C, source voltage was set at 3.0 kV, sheath gas (N₂) flow was 40 Arb and aux gas flow rate was 20 Arb. Full MS scans were acquired in the range of *m/z* 50–1500 and the MS/MS experiments were set as data-dependent scans.

Predicting the Targets of the Identified Components Within GBKC

MedChem Studio (MedChem Studio, 3.0; Simulations Plus, Inc., Lancaster, CA, USA, 2012), a powerful tool for predicting targets based on structure similarity, was applied to retrieve the targets of candidate ingredients of GBKC, with the similarity threshold set at 0.6.²³

Known Therapeutic Targets Associated with CB

The target proteins/genes associated with FDA-approved drugs for CB were retrieved from DrugBank database (<http://www.drugbank.ca/>, version 5.1.1) and Online Mendelian Inheritance in Man (OMIM) database (<http://www.omim.org/>, updated on May 4, 2018) using the keyword “Chronic bronchitis” and confining the species to “Homo sapiens.” DrugBank is a high quality database containing comprehensive information on drugs and their targets.²⁴ OMIM is a comprehensive, freely accessible database of genetic diseases and their related human genes.²⁵

Protein-Protein Interaction (PPI) Data

PPI data were extracted from Search Tool for the Retrieval of Interacting Genes/Proteins (STRING) database (<http://string-db.org/>) which integrated almost all publicly available sources of PPI information.²⁶ The cut-off value was set at 0.4 for the screening of high confidence edges.

Network Construction and Analysis

For understanding the relationships between the predicted targets of GBKC and CB-related target genes/proteins, Cytoscape (version 3.6.0, Boston, MA, USA) was adopted to visualize the “bioactive components-targets-disease” network and draw the network diagram. Using the Cytoscape option network analyzer, the degree values of nodes in the network were assessed, and a node was represented as a hub when its degree value was two-fold above the median value of all nodes in the network. Next, the hub network was constructed by employing direct interactions among the hubs. Moreover, to estimate the topological importance of the hub nodes, “degree,” “betweenness,” and “closeness” were calculated. According to our previous studies,¹¹ if the values of the three topological

properties exceeded the corresponding median values, these hub nodes would be recognized as the key GBKC targets for CB.

Pathway Enrichment Analysis

To determine the significance of the major hubs, the Database for Annotation, Visualization and Integrated Discovery Bioinformatics Resources 6.7 online tool (<http://david.abcc.ncifcrf.gov/home.jsp/>, version 6.7) was employed to analyze the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway (<http://www.genome.jp/kegg/>) and gene ontology (GO) enrichment of GBKC on the direct or indirect targets of CB. The threshold value of confident gene enrichment was set at $P < 0.05$.

Molecular Docking Stimulation

Molecular docking studies were carried out to validate the credibility of the compound-target interactions using the Discovery Studio 2016 option LibDock. The high-resolution crystal structures of the key targets were extracted from the RCSB protein database (<http://www.pdb.org/>) and prepared by deleting the co-crystallized ligand and water molecules, adding hydrogen atoms, and fixing the side chains. The binding site was defined by the ligand atoms, and the cutoff radius was set to 10 Å. Chem3D Pro 12.0 was used to generate the three-dimensional structures of the candidate components that were saved in PDB format. The docking scores were used to evaluate the binding affinities between the hub targets and their corresponding compounds. A docked conformation with the best fitness score was selected. The other parameters used in calculation were default except for explained.

SPR Analysis

Based on the virtual screening results, four target proteins were selected for SPR analysis: nuclear factor kappa B (NFκB1, Novoprotein), matrix metalloproteinase-9 (MMP-9, Novoprotein), tumor necrosis factor (TNF- α , Novoprotein), and mitogen-activated protein kinase (MAPK1, Sino Biological). SPR experiments were performed using a Biacore 8 K (GE Healthcare, Sweden). The surface of the CM5 sensor chip was activated by injecting a freshly prepared mixture of NHS and EDC (1:1, v/v). NFκB1, MMP-9,

TNF- α , and MAPK1 were immobilized on CM5 chips via a standard amine coupling method with immobilization levels of 11,800, 14,874, 11,000, and 11,000 response units, respectively. Detection was conducted according to the protocol supplied by GE Healthcare. For each sample analysis, a reference channel without the conjugated protein was activated and blocked to eliminate non-specific binding to the sensor surface. The analytes were injected at a flow rate of 30 μ L/min. The association and dissociation times were set at 60 s and 120 s, respectively. The SPR curves were fitted using Biacore Insight Evaluation Software according to the 1:1 Langmuir binding model and the steady-state affinity model, from which the kinetics (the association rate constant, K_a ; the dissociation rate constant, K_d) and affinity constants (K_D) were calculated.

Cell Culture and Treatment

The MH-S cells were maintained in RPMI-1640 medium containing 10% FBS and 1% penicillin-streptomycin at 37 °C in 5% CO₂ humidified air. For each experiment, the cells were stimulated with LPS (1 μ g/mL) in the presence or absence of GBKC (0.1, 0.5, 1.0, and 1.5 mg/mL) for 24 h.

Cell Viability Assay

A cell counting Kit-8 (CCK-8) assay was performed to assess the viability of MH-S cells after GBKC treatment. Briefly, the cells were seeded into 96-well plates at a density of 3×10^4 cells/well and then treated with or without GBKC (0.1, 0.5, 1.0, or 1.5 mg/mL) for 24 h. Next, cell-free supernatants were removed and 200 μ L of 10% CCK-8 solution was added to each well. The plates were then incubated at 37 °C for another 0.5–2 h. Finally, the optical density of each well was measured at 490 nm using a microplate reader (Molecular Devices, Sunnyvale, CA, USA).

Enzyme-Linked Immunosorbent Assay (ELISA)

The levels of TNF- α and interleukin (IL)-6, two vital cytokines in the early stage of inflammation,²⁷ in the cell incubation medium were determined using ELISA kits following the manufacturer's instructions.

Table I Analysis of the Chemical Constituents of GBKC by UHPLC-LTQ-Orbitrap in Positive Ion Mode

No.	RT (Min)	[M+H] ⁺ / [M+Na] ⁺	MS/MS	Molecular Formula	Error (ppm)	Compound	Herb
3	5.81	268.1534	88.0755[C ₄ H ₁₀ ON] ⁺	C ₁₄ H ₂₂ O ₄ N	-0.955	Codonopsine	DS
4	8.79	565.1530	547.1432[M+H-H ₂ O] ⁺ , 529.1332[M+H-2H ₂ O] ⁺	C ₂₆ H ₂₈ O ₁₄	-3.949	Schaftoside/Isoschaftoside	GC
5	9.01	389.0830	345.0941[M+Na-CO ₂] ⁺ , 227.0311[M+Na-C ₆ H ₁₀ O ₅] ⁺ , 185.0417[C ₆ H ₁₀ O ₅ Na] ⁺	C ₁₇ H ₁₈ O ₉	-3.401	Psoralenoxide/Isopsoralenoxide	BGZ
6	9.26	389.0828	345.0940[M+Na-CO ₂] ⁺ , 227.0311[M+Na-C ₆ H ₁₀ O ₅] ⁺ , 185.0417[C ₆ H ₁₀ O ₅ Na] ⁺	C ₁₇ H ₁₈ O ₉	-3.966	Psoralenoxide/Isopsoralenoxide	BGZ
7	9.54	565.1535	547.1439[M+H-H ₂ O] ⁺ , 529.1339[M+H-2H ₂ O] ⁺	C ₂₆ H ₂₈ O ₁₄	-2.976	Schaftoside/Isoschaftoside	GC
9	10.32	257.0801	242.0570[M+H-CH ₃] ⁺ , 239.0701[M+H-H ₂ O] ⁺ , 211.0751[M+H-H ₂ O-CO] ⁺ , 163.0387[M+H-H ₂ O-C ₆ H ₄] ⁺ , 147.0437[M+H-C ₆ H ₆ O ₂] ⁺ , 137.0230[M+H-C ₈ H ₈ O] ⁺ , 119.0489[C ₈ H ₇ O] ⁺	C ₁₅ H ₁₂ O ₄	-2.938	Liquiritigenin	GC
10	10.39	551.1738	419.1330[M+H-C ₅ H ₈ O ₄] ⁺ , 389.1225[M+H-C ₆ H ₁₀ O ₅] ⁺ , 257.0806[M+H-C ₅ H ₈ O ₄ -C ₄ H ₆ O ₄ -C ₂ H ₂ O] ⁺	C ₂₆ H ₃₀ O ₁₃	-3.787	Liquiritin apioside	GC
11	10.42	257.0801	242.0577[M+H-CH ₃] ⁺ , 239.0707[M+H-H ₂ O] ⁺ , 211.0757[M+H-CH ₂ O ₂] ⁺ , 147.0442[M+H-C ₆ H ₆ O ₂] ⁺ , 137.0233[M+H-C ₈ H ₈ O] ⁺	C ₁₅ H ₁₂ O ₄	-2.938	Isoliquiritigenin	GC
14	12.11	551.1738	419.1330[M+H-C ₅ H ₈ O ₄] ⁺ , 389.1225[M+H-C ₆ H ₁₀ O ₅] ⁺ , 257.0806[M+H-C ₅ H ₈ O ₄ -C ₄ H ₆ O ₄ -C ₂ H ₂ O] ⁺	C ₂₆ H ₃₀ O ₁₃	-3.787	Liquiritin apioside isomer	GC
15	12.44	419.1331	257.0805[M+H-C ₆ H ₁₀ O ₅] ⁺	C ₂₁ H ₂₂ O ₉	-1.285	Isoliquiritin	GC
16	12.59	431.132	NA	C ₂₂ H ₂₂ O ₉	-3.801	Ononin	GC
17	12.59	269.0800	254.0569[M+H-CH ₃] ⁺ , 237.0544[M+H-CH ₃ OH] ⁺ , 213.0907[M+H-2CO] ⁺	C ₁₆ H ₁₂ O ₄	-3.476	Formononetin	GC
18	14.10	350.1950	250.1435[M-C ₅ H ₈ O ₂] ⁺ , 220.1329[M-C ₆ H ₁₀ O ₃] ⁺ , 182.1174[M-C ₅ H ₈ O ₂ -C ₄ H ₄ O] ⁺ , 205.0856[M-C ₅ H ₈ O ₂ -C ₂ H ₇ N] ⁺ , 161.0595[C ₁₀ H ₉ O ₂] ⁺	C ₁₉ H ₂₈ O ₅ N	-0.669	Codonopyrrolidinium A	DS
19	14.25	187.0382	159.0438[M+H-H ₂ O] ⁺ , 143.0487[M+H-CO ₂] ⁺ , 131.0488[M+H-C ₂ O ₂] ⁺ , 115.0539[M+H-C ₂ O ₃] ⁺	C ₁₁ H ₆ O ₃	-4.013	Psoralen/Angelicin	BGZ
20	14.55	187.0382	159.0439[M+H-H ₂ O] ⁺ , 143.0488[M+H-CO ₂] ⁺ , 131.0489[M+H-C ₂ O ₂] ⁺ , 115.0540[M+H-C ₂ O ₃] ⁺	C ₁₁ H ₆ O ₃	-4.013	Psoralen/Angelicin	BGZ
21	14.71	469.3296	451.3203[M+H-H ₂ O] ⁺ , 433.3100[M+H-2H ₂ O] ⁺	C ₃₀ H ₄₄ O ₄	-3.444	16-deoxyporicoic acid B	FL
22	14.77	839.4041	NA	C ₄₂ H ₆₂ O ₁₇	-2.224	Licoricesaponin G2/isomer	GC
23	15.58	839.4041	NA	C ₄₂ H ₆₂ O ₁₇	-2.224	Licoricesaponin G2/isomer	GC
25	15.60	337.1062	319.0961[M+H-H ₂ O] ⁺ , 279.0650[M+H-C ₃ H ₆ O] ⁺	C ₂₀ H ₁₆ O ₅	-2.433	Psoralidin	BGZ
28	16.93	777.4008	NA	C ₃₉ H ₆₂ O ₁₄	-3.110	(25R)-spirost-5-ene-3β,14α,17α-triol-3-O-α-L-rhamnopyranosyl-(1→2)-β-D-glucopyranoside	MD

(Continued)

Table I (Continued).

No.	RT (Min)	[M+H] ⁺ / [M+Na] ⁺	MS/MS	Molecular Formula	Error (ppm)	Compound	Herb
30	17.30	985.4611	NA	C ₄₈ H ₇₂ O ₂₁	-2.806	Licoricesaponine A3	GC
31	17.32	325.1427	269.0805[M+H-C ₄ H ₇] ⁺	C ₂₀ H ₂₀ O ₄	-2.355	Bavachin	BGZ
32	18.00	455.2024	440.1799[M+Na-CH ₃] ⁺ , 409.1617[M+H-CH ₃ -OCH ₃] ⁺	C ₂₄ H ₃₂ O ₇	-3.525	Schisandrol A	WWZ
33	18.14	323.1268	267.0646[M+H-C ₄ H ₈] ⁺ , 255.0652[M+H-C ₅ H ₈] ⁺	C ₂₀ H ₁₈ O ₄	-3.050	Neobavaisoflavone	BGZ
35	18.77	249.1478	231.1379[M+H-H ₂] ⁺	C ₁₅ H ₂₀ O ₃	-2.974	Atractylenolide III	BZ
36	18.82	553.2030	NA	C ₂₈ H ₃₄ O ₁₀	-2.564	Gomisin D	WWZ
37	18.82	231.1374	213.1274[M+H-OH] ⁺ , 203.1067[M+H-CO] ⁺ , 185.1325[M+H-CO-H ₂ O] ⁺ , 163.0753[M+H-C ₅ H ₈] ⁺	C ₁₅ H ₁₈ O ₂	-2.537	Atractylenolide I	BZ
38	18.82	325.1426	269.0805[M+H-C ₄ H ₇] ⁺	C ₂₀ H ₂₀ O ₄	-2.539	Bavachalcone/ Isobavachalcone	BGZ
40	19.28	439.1711	424.1485[M+Na-CH ₃] ⁺ , 393.1303[M+Na-CH ₃ -OCH ₃] ⁺	C ₂₃ H ₂₈ O ₇	-3.744	Schisandrol B	WWZ
43	20.57	389.2535	371.2438[M+H-H ₂ O] ⁺	C ₂₀ H ₃₆ O ₇	0.257	Tetradecylcitric acid	BZ
45	20.91	523.2281	508.2060[M+Na-H ₂ O] ⁺ , 477.1876[M+Na-C ₂ H ₆ O] ⁺ , 441.1879[M+Na-C ₅ H ₆ O] ⁺ , 409.1617[M+Na-C ₆ H ₁₀ O ₂] ⁺ , 386.1719[M+Na-C ₈ H ₉ O ₆] ⁺	C ₂₈ H ₃₆ O ₈	-4.108	Tigloylgomisin H/ Angeloylgomisin H	WWZ
46	21.21	523.2280	508.2060[M+Na-H ₂ O] ⁺ , 477.1876[M+Na-C ₂ H ₆ O] ⁺ , 441.1879[M+Na-C ₅ H ₆ O] ⁺ , 409.1617[M+Na-C ₆ H ₁₀ O ₂] ⁺ , 386.1719[M+Na-C ₈ H ₉ O ₆] ⁺	C ₂₈ H ₃₆ O ₈	-4.337	Tigloylgomisin H/ Angeloylgomisin H	WWZ
48	21.67	233.1528	215.1429[M+H-H ₂ O] ⁺ , 187.1480[M+H-H ₂ O-CO] ⁺ , 151.0752[M+H-C ₆ H ₁₀] ⁺	C ₁₅ H ₂₀ O ₂	-3.330	Atractylenolide II	BZ
49	21.97	537.2079	437.1566[M+Na-C ₅ H ₈ O ₂] ⁺ , 415.1751[M+Na-C ₇ H ₆ O ₂] ⁺	C ₂₈ H ₃₄ O ₉	-3.004	Tigloylgomisin P/Gomisin E	WWZ
50	22.74	339.1580	321.1483[M+H-H ₂ O] ⁺ , 283.0961[M+H-C ₄ H ₈] ⁺ , 271.0962[M+H-C ₅ H ₈] ⁺	C ₂₁ H ₂₂ O ₄	-3.850	Bavachinin A	BGZ
51	22.89	537.2073	437.1566[M+Na-C ₅ H ₈ O ₂] ⁺ , 415.1751[M+Na-C ₇ H ₆ O ₂] ⁺	C ₂₈ H ₃₄ O ₉	-4.139	Tigloylgomisin P/Gomisin E	WWZ
53	25.31	339.1579	321.1482[M+H-H ₂ O] ⁺ , 283.0962[M+H-C ₄ H ₈] ⁺ , 271.0962[M+H-C ₅ H ₈] ⁺	C ₂₁ H ₂₂ O ₄	-3.584	4'-O-methylbavachalcone	BGZ

Abbreviations: NA, not available.

Statistical Analysis

All experiments were performed in triplicate, and the data are presented as mean ± standard deviation. Using GraphPad Prism version 7.0 (GraphPad Software, CA, USA) with Student's *t*-test, statistical analysis was conducted to evaluate the significance of differences between groups. Statistical significance was set at *P* < 0.05.

Results and Discussion

Identification of the Main Constituents of GBKC

The chemical composition of GBKC was detected and identified by UHPLC-LTQ-Orbitrap (Figure S1 and S2). In total, 53 major components, including flavonoids and their glycosides, coumarins, lignans, steroids, alkaloids

Table 2 Analysis of the Chemical Constituents of GBKC by UHPLC-LTQ-Orbitrap in Negative Ion Mode

No.	RT (Min)	[M-H] ⁻ / [M+HCOO] ⁻	MS/MS	Molecular Formula	Error (ppm)	Compound	Herb
1	1.91	191.0190	173.0088[M-H-H ₂ O] ⁻ , 111.0088[M-H-CH ₄ O ₄] ⁻	C ₆ H ₈ O ₇	1.995	Citric acid/Isocitric acid	WWZ
2	2.26	191.0188	173.0088[M-H-H ₂ O] ⁻ , 111.0088[M-H-CH ₄ O ₄] ⁻	C ₆ H ₈ O ₇	1.995	Citric acid/Isocitric acid	WWZ
4	8.79	563.1379	545.1290[M-H-H ₂ O] ⁻ , 503.1184 [M-H-C ₂ H ₂ O ₂] ⁻ , 473.1080[M-H-C ₃ H ₆ O ₃] ⁻ , 443.0974[M-H-C ₄ H ₈ O ₄] ⁻	C ₂₆ H ₂₈ O ₁₄	-1.652	Schaftoside/Isoschaftoside	GC
5	9.01	365.0859	203.0347[M-H-C ₆ H ₁₀ O ₅] ⁻ , 159.0450 [M-H-C ₆ H ₁₀ O ₅ -CO ₂] ⁻	C ₁₇ H ₁₈ O ₉	-2.269	Psoralenoside/ Isopsoralenoside	BGZ
6	9.26	365.0859	203.0347[M-H-C ₆ H ₁₀ O ₅] ⁻ , 159.0450 [M-H-C ₆ H ₁₀ O ₅ -CO ₂] ⁻	C ₁₇ H ₁₈ O ₉	-2.352	Psoralenoside/ Isopsoralenoside	BGZ
7	9.54	563.1384	545.1290[M-H-H ₂ O] ⁻ , 503.1184 [M-H-C ₂ H ₂ O ₂] ⁻ , 473.1080[M-H-C ₃ H ₆ O ₃] ⁻ , 443.0974[M-H-C ₄ H ₈ O ₄] ⁻	C ₂₆ H ₂₈ O ₁₄	-1.102	Schaftoside/Isoschaftoside	GC
8	10.30	417.1173	255.0656[M-H-C ₆ H ₁₀ O ₅] ⁻	C ₂₁ H ₂₂ O ₉	-0.729	Liquiritin	GC
9	10.32	255.0650	135.0088[M-H-C ₈ H ₈ O] ⁻	C ₁₅ H ₁₂ O ₄	-0.135	Liquiritigenin	GC
10	10.39	549.1582	417.1190[M-H-C ₅ H ₈ O ₄] ⁻ , 297.0764 [M-H-C ₅ H ₈ O ₄ -C ₄ H ₆ O ₄] ⁻ , 255.0661 [M-H-C ₅ H ₈ O ₄ -C ₄ H ₆ O ₄ -C ₂ H ₂ O] ⁻	C ₂₆ H ₃₀ O ₁₃	-2.067	Liquiritin apioside	GC
11	10.42	255.0650	135.0088[M-H-C ₈ H ₈ O] ⁻	C ₁₅ H ₁₂ O ₄	-0.844	Isoliquiritigenin	GC
12	10.42	425.2007	263.1493[M-H-C ₆ H ₁₀ O ₅] ⁻ , 161.0453[C ₆ H ₉ O ₅] ⁻	C ₁₈ H ₃₄ O ₁₁	-2.559	Hexyl-β-D-maltoside	DS
13	10.89	395.1903	263.1493[M-H-C ₅ H ₈ O ₄] ⁻ , 161.0453[C ₆ H ₉ O ₅] ⁻	C ₁₇ H ₃₂ O ₁₀	-0.834	Dextran	DS
14	12.11	549.1586	417.1190[M-H-C ₅ H ₈ O ₄] ⁻ , 297.0764 [M-H-C ₅ H ₈ O ₄ -C ₄ H ₆ O ₄] ⁻ , 255.0661 [M-H-C ₅ H ₈ O ₄ -C ₄ H ₆ O ₄ -C ₂ H ₂ O] ⁻	C ₂₆ H ₃₀ O ₁₃	-1.637	Liquiritin apioside isomer	GC
15	12.44	417.1173	255.0656[M-H-C ₆ H ₁₀ O ₅] ⁻	C ₂₁ H ₂₂ O ₉	-0.729	Isoliquiritin	GC
16	12.59	475.1227	NA	C ₂₂ H ₂₂ O ₉	-1.764	Ononin	GC
17	12.59	267.0649	NA	C ₁₆ H ₁₂ O ₄	-3.084	Formononetin	GC
22	14.77	837.3875	819.3779[M-H-H ₂ O] ⁻ , 661.3566 [M-H-C ₆ H ₈ O ₆] ⁻ , 351.0552[M-H-C ₃₀ H ₄₆ O ₄] ⁻	C ₄₂ H ₆₂ O ₁₇	-4.090	Licoricesaponin G2/isomer	GC
23	15.58	837.3909	819.3771[M-H-H ₂ O] ⁻ , 661.3569 [M-H-C ₆ H ₈ O ₆] ⁻ , 351.0553[M-H-C ₃₀ H ₄₆ O ₅] ⁻	C ₄₂ H ₆₂ O ₁₇	0.649	Licoricesaponin G2/isomer	GC
24	15.58	329.2321	311.2220[M-H-H ₂ O] ⁻ , 293.2117[M-H-2H ₂ O] ⁻ , 229.1438[M-H-C ₆ H ₁₂ O] ⁻ , 211.1335 [M-H-C ₆ H ₁₂ O ₂] ⁻ , 171.1024[C ₉ H ₁₅ O ₃] ⁻	C ₁₈ H ₃₄ O ₅	-0.488	9(S),12(S),13(S)-trihydroxy-10 (E)-octadecenoic Acid	DS
26	16.42	821.3919	803.3821[M-H-H ₂ O] ⁻ , 759.3926 [M-H-H ₂ O-CO ₂] ⁻ , 645.3611[M-H-C ₆ H ₈ O ₆] ⁻ , 627.3506[M-H-C ₆ H ₁₀ O ₇] ⁻ , 351.0548 [C ₁₂ H ₁₅ O ₁₂] ⁻	C ₄₂ H ₆₂ O ₁₆	-4.931	Glycyrrhizic acid	GC
27	16.44	327.2167	309.2063[M-H-H ₂ O] ⁻ , 291.1958[M-H-2H ₂ O] ⁻ , 229.1438[M-H-C ₆ H ₁₀ O ₇] ⁻ , 211.1334 [M-H-C ₆ H ₁₀ O ₇ -H ₂ O] ⁻ , 171.1023[C ₉ H ₁₅ O ₃] ⁻	C ₁₉ H ₂₀ O ₅	0.335	Methylpogonone B	MD

(Continued)

Table 2 (Continued).

No.	RT (Min)	[M-H] ⁻ / [M+HCOO] ⁻	MS/MS	Molecular Formula	Error (ppm)	Compound	Herb
29	17.27	821.3919	803.3820[M-H-H ₂ O] ⁻ , 645.3611 [M-H-C ₆ H ₈ O ₆] ⁻	C ₄₂ H ₆₂ O ₁₆	-4.931	Uralsaponin B	GC
30	17.30	983.4451	965.4338[M-H-H ₂ O] ⁻ , 893.4147 [M-H-C ₃ H ₇ O ₃] ⁻ , 821.3914[M-H-C ₆ H ₁₀ O ₅] ⁻ , 803.3823[M-H-C ₆ H ₁₀ O ₅ -H ₂ O] ⁻	C ₄₈ H ₇₂ O ₂₁	-3.776	Licoricesaponine A3	GC
31	17.32	323.1275	203.0709[M-H-C ₈ H ₈ O] ⁻	C ₂₀ H ₂₀ O ₄	-0.977	Bavachin	BGZ
33	18.14	321.1117	265.0504[M-H-C ₄ H ₈] ⁻	C ₂₀ H ₁₈ O ₄	-1.481	Neobavaisoflavone	BGZ
34	18.64	359.1124	344.0891 [M-H-CH ₃] ⁻	C ₁₉ H ₂₀ O ₇	-0.159	5,7-dihydroxy-3-(2-hydroxy-4-methoxybenzyl)-8-methoxy-6-methylchroman-4-one	MD
38	18.82	323.1269	203.0709[M-H-C ₈ H ₈ O] ⁻	C ₂₀ H ₂₀ O ₄	-2.679	Bavachalcone/ Isobavachalcone	BGZ
39	18.94	783.4	NA	C ₃₉ H ₆₂ O ₁₃	-0.252	Dracaenoside F	MD
41	19.89	319.0965	304.0742[M-H-CH ₃] ⁻ , 301.0863[M-H-H ₂ O] ⁻	C ₂₀ H ₁₆ O ₄	0.046	Corylin	BGZ
42	20.48	335.0913	280.0371 [M-H-C ₄ H ₇] ⁻	C ₂₀ H ₁₆ O ₅	-0.179	Derrone/Isoderrone	GC
44	20.88	313.2374	295.2271 [M-H-H ₂ O] ⁻ , 277.2166[M-H-2H ₂ O] ⁻ , 201.1128[M-H-C ₈ H ₁₆] ⁻ , 171.1024[C ₉ H ₁₅ O ₃] ⁻	C ₁₈ H ₃₄ O ₄	0.109	9,10-dihydroxy-12-octadecenoic acid	DS
47	21.58	351.0864	283.0969[M-H-CH ₃ O] ⁻ , 307.0967 [M-H-CO ₂] ⁻ , 265.0862[M-H-C ₃ H ₂ O ₃] ⁻	C ₂₀ H ₁₆ O ₆	-1.319	l-methoxyphaseollidin	GC
52	23.49	389.1749	277.0499[M-H-C ₈ H ₆] ⁻ , 265.0500 [M-H-C ₉ H ₆] ⁻	C ₂₅ H ₂₆ O ₄	0.345	Corylifol A	BGZ

Abbreviation: NA, not available.

and phenolic acids, were confirmed or tentatively identified based on comparison with the published data, according to retention times, molecular ions, fragmentation data, and fragmentation rules. As depicted in [Figure S3–S12](#), liquiritigenin (peak 9), codonopyrrolidinium A (peak 18), psoralen (peak 19), glycyrrhizic acid (peak 26) and schisandrol A (peak 32) were chosen as examples to illustrate the mass fragmentation patterns of the components with the same basic skeletons. The detailed information of the identified compounds was summarized in [Tables 1 and 2](#).

Putative Targets for GBKC

As given in [Table S1](#), 594 GBKC-related targets were predicted by MedChem Studio.

CB-Related Targets

After eliminating the repetitive targets, 414 predictive targets associated with CB were reserved. Among them, 138 targets

were from DrugBank database and 286 genes were from OMIM database. The details were shown in [Table S2](#).

Network and Pathway Analysis

To achieve a systematic understanding of the complex relationships, a “compound-target-disease” network containing the potential active components, GBKC-related targets, and CB-related targets was established based on the PPI information in the STRING database. There were 851 nodes and 12,548 edges in the network ([Table S3](#)).

The degree value of each node in the network was calculated to identify the central nodes. Ultimately, 423 nodes with an average degree value of >23 were identified as hubs. The interaction network was then established based on direct connections among the hubs. After conditional screening with topological coefficients (“degree” > 33, “betweenness” > 0.0011, and

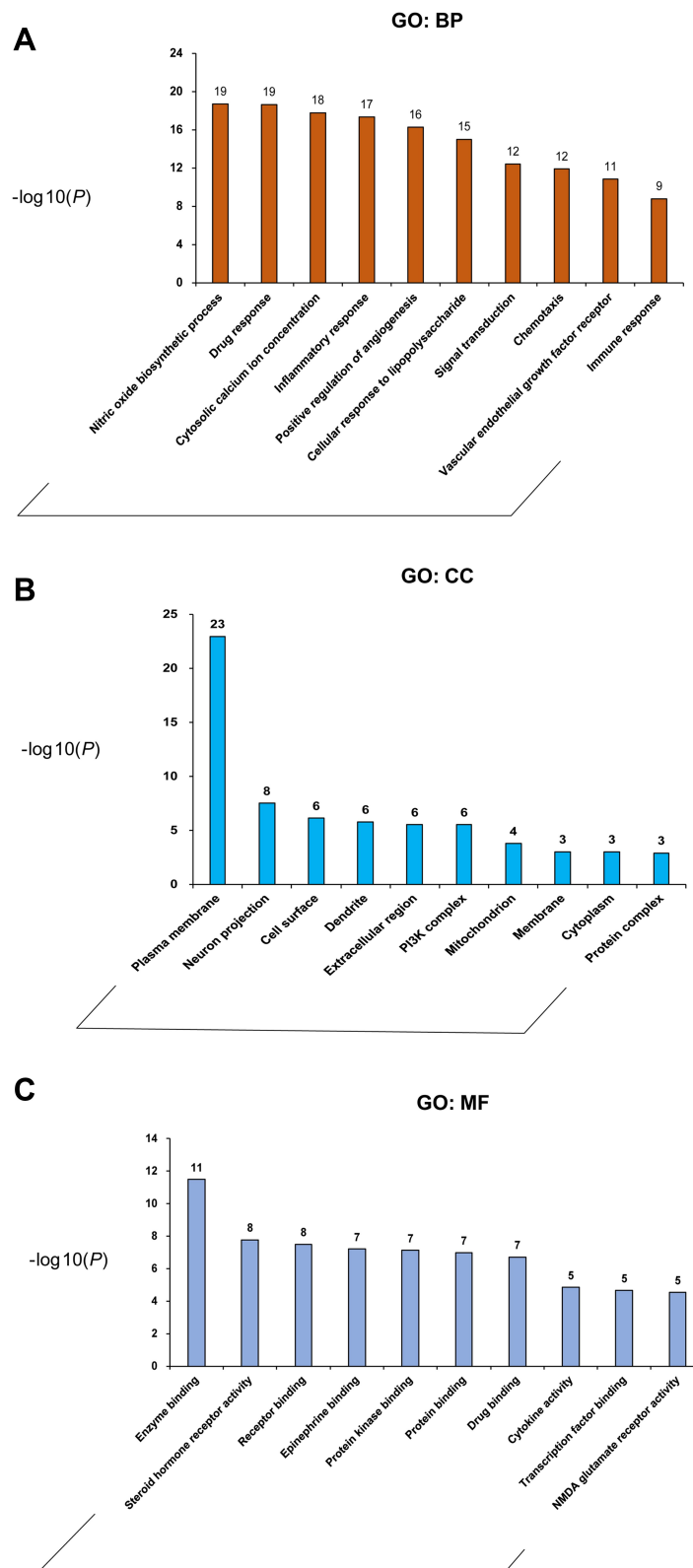


Figure 2 GO term performance of the major hubs. **(A)** GO: BP; **(B)** GO: CC; **(C)** GO: MF. The abscissa stands for GO terms, the ordinate stands for minus log₁₀(P).

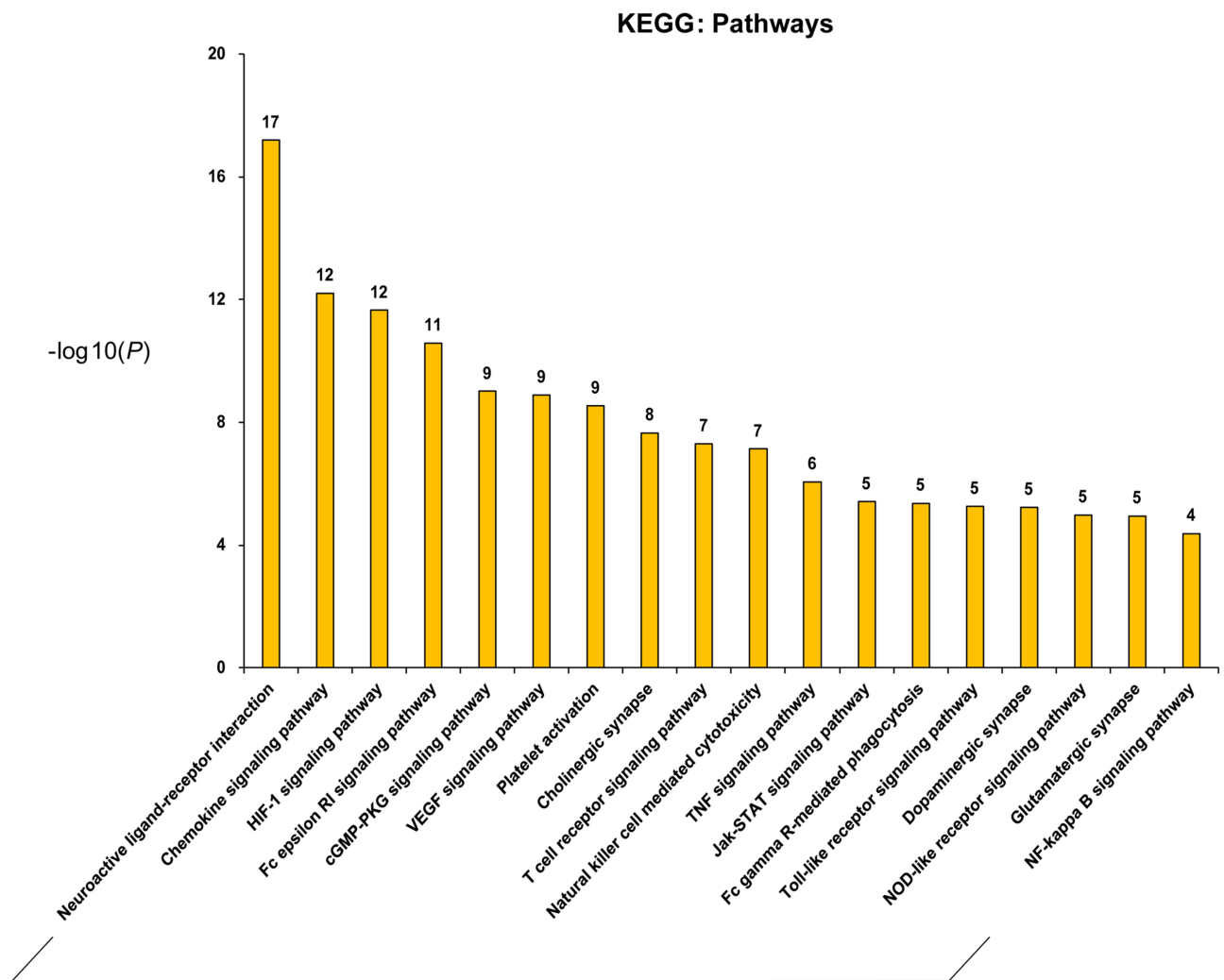


Figure 3 Main KEGG pathways composed of the major hubs. The abscissa stands for KEGG pathways, the ordinate stands for minus log 10(P).

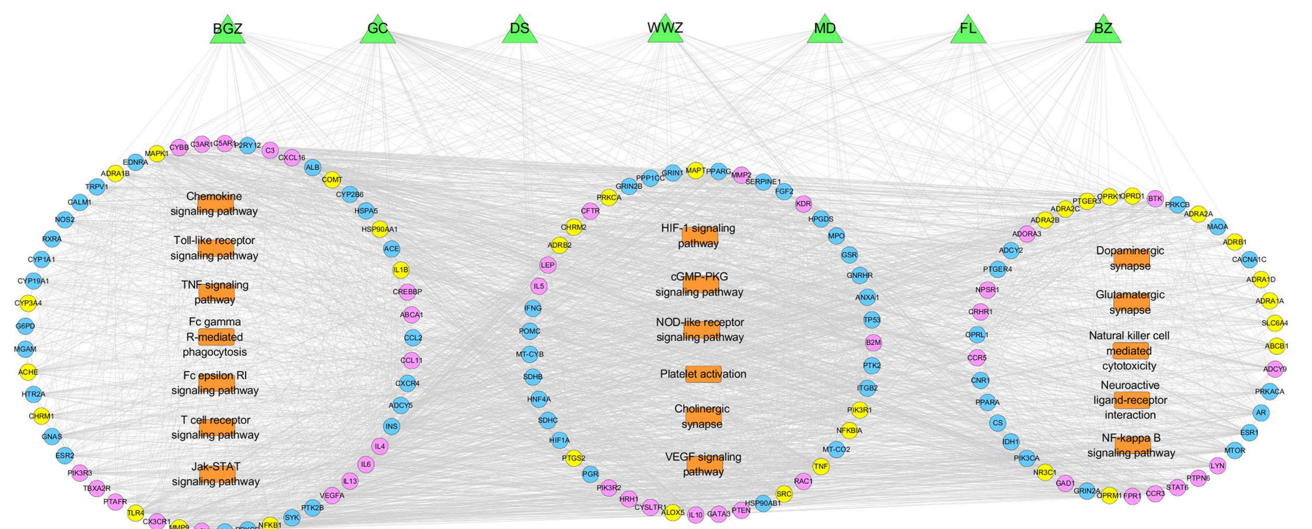


Figure 4 GBKC-major hubs-main pathway network. Green triangles represent each herbs contained in GBKC; round blue nodes represent putative targets of GBKC; round red nodes represent CB associated targets; round yellow nodes represent both GBKC targets and CB associated targets; orange rectangles represent top 18 pathways from enrichment analysis of major targets; edges represent interactions among herbs in GBKC, putative targets, CB associated targets, and main pathways.

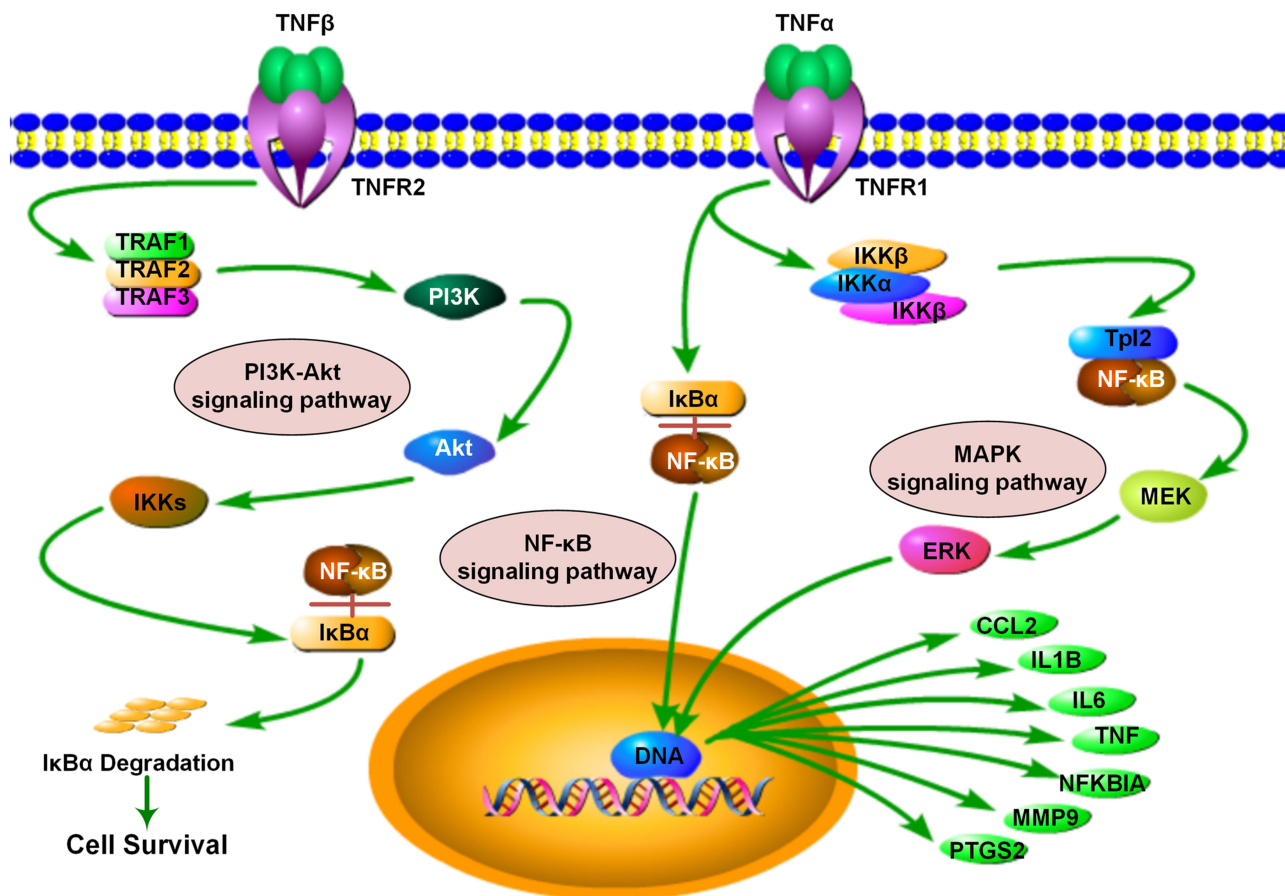


Figure 5 Illustration of TNF signaling pathway influenced by major putative targets of GBKC.

“closeness” > 0.4626), 141 hubs were considered as pivotal targets. Among them, 65 hubs were relevant to GBKC, 42 were relevant to CB, and 34 were both GBKC and CB-associated targets. The details are presented in [Table S4](#).

To clarify the pathways involved in the major hubs, GO enrichment analysis was conducted, which was composed of biological process (BP), cell component (CC), and molecular function (MF). As described in [Figure 2A–C](#), the top 10 significant terms of BP, CC, and MF were obtained. GBKC mainly participates in regulating the following BPs: nitric oxide biosynthetic process, drug response, cytosolic calcium ion concentration, inflammatory response, positive regulation of angiogenesis, cellular response to lipopolysaccharide, signal transduction, chemotaxis, vascular endothelial growth factor receptor, and immune response. GBKC affected CB by regulating the following MFs: enzyme binding, steroid hormone receptor activity, receptor binding, epinephrine binding, protein kinase binding,

protein binding, drug binding, cytokine activity, transcription factor binding, and NMDA glutamate receptor activity. For CCs, the major hubs were enriched in the plasma membrane, neuron projection, cell surface, dendrite, extracellular region, PI3K complex, mitochondrion, membrane, cytoplasm, and protein complex.

To illustrate the signaling pathways that have remarkably changed in the course of CB, KEGG pathway enrichment analysis was performed. A total of 134 significant pathways were identified, and the top 18 signal pathways were depicted in [Figure 3](#). These pathways were divided into the following three functional modules: immune system (such as Toll-like receptor signaling pathway, JAK-STAT signaling pathway, T cell receptor signaling pathway, Fc epsilon RI signaling pathway, Fc gamma R-mediated phagocytosis, chemokine signaling pathway, and TNF signaling pathway), blood circulation system (such as NOD-like receptor signaling pathway, cholinergic synapse, cGMP-PKG

signaling pathway, platelet activation, VEGF signaling pathway, and HIF-1 signaling pathway), and nervous system (such as neuroactive ligand-receptor interaction, dopaminergic synapse, NF-kappa B signaling pathway, natural killer cell mediated cytotoxicity, and glutamatergic synapse).

To facilitate a clear interpretation of the complex connections among GBKC components, key hubs of GBKC, and meaningful pathways, the compound-target-pathway network was graphed, as shown in Figure 4. Interestingly, the TNF signaling pathway was highly enriched in the main KEGG pathways. Previous studies have shown that TNF- α levels increase markedly in peripheral blood, bronchoalveolar lavage fluid, bronchial biopsies, and induced sputum of patients with CB.^{28,29} Therefore, GBKC may ameliorate airway inflammation in patients with CB.

However, the network pharmacology method used in this study had some limitations. First, the components in GBKC were treated equally, without considering their contents. Second, some identified constituents may not be absorbed into the blood circulatory system, and in many situations, it is the metabolites, not the parent compounds, which have therapeutic effects. Third, the GBKC decoction is not suitable for CB with all TCM syndromes. Therefore, using targets related to the specific syndrome of CB to construct a network model would be more accurate.

GBKC Attenuates the Inflammatory Response of CB Patients by Regulating the TNF Signaling Pathway

As shown in Table S5, the GBKC-related targets associated with TNF signaling pathway include TNF- α , NF- κ B, NF- κ B inhibitor alpha (I κ B α), MAPK1, phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha isoform (PIK3CA), phosphoinositide-3-kinase regulatory subunit 1 (PIK3R1), PIK3R2, PIK3R3, MMP-9, prostaglandin-endoperoxide synthase 2 (PTGS2), IL-1 β , IL-6, and C-C motif chemokine 2 (CCL2). Figure 5 shows a graphical overview of the TNF pathway influenced by the major putative targets of GBKC.

Molecular Docking

Table 3 depicts the docking scores (cut-off value of 80) of GBKC hit components against the active sites of the

Table 3 Molecular Docking Results

Targets	Compound	LibDockscore
MMP9	Liquiritin	155.244
MMP9	Isoliquiritin	125.357
TNF	Isoliquiritin	120.423
MAPK1	Ononin	120.082
NFKBIA	Isoliquiritigenin	117.096
MAPK1	Liquiritin	112.194
PIK3CA	1-Methoxyphaseollidin	109.393
NFKB1	Isoliquiritigenin	109.047
MAPK1	Isopsoralenoside	107.812
MAPK1	Psoralenoside	107.07
MAPK1	Bavachin	105.343
PIK3CA	Corylifol A	104.358
MAPK1	Bavachinin A	103.145
PTGS2	9,10-Dihydroxy-12-octadecenoic acid	101.172
NFKB1	Formononetin	100.175
MAPK1	Isoliquiritin	99.9182
NFKBIA	Formononetin	95.6731
MAPK1	5,7-Dihydroxy-3-(2-hydroxy-4-methoxybenzyl)-8-methoxy-6-methylchroman-4-one	92.7798
MAPK1	Neobavaisoflavone	88.7266
MAPK1	5,7-Dihydroxy-3-(2-hydroxy-4-methoxybenzyl)-8-methoxy-6-methylchroman-4-one	83.3577

identified protein targets in the TNF pathway, including MMP-9, TNF- α , MAPK1, NFKBIA, NFKB1, and PTGS2. The major binding interactions between the above active compounds and target proteins include hydrogen bonding and amide- π stacked, π - π stacked, π -alkyl, and alkyl interactions. Herein, the molecular simulation of isoliquiritin in the active site of TNF- α was selected as an example to demonstrate how the active compounds bound to their targets (Figure 6). Isoliquiritin forms six hydrogen bonds, two of them were phenolic hydroxyl with LEU 157 and LEU 55. The other four, namely hydroxyl groups on glucose were with LEU 126, GLN 125 and SER 95. GLU 53 and GLY 54 were bound with other interactions, including amide- π stacked and π -sigma interactions. The molecular docking results described above are helpful for a basic understanding of the mechanism of drug action.

SPR Assays for Affinity

SPR assay was used to confirm the direct binding efficacy of the targets and their corresponding ligands based on the molecular docking results, including isoliquiritigenin-NFKB1, formononetin-NFKB1, liquiritin-MMP-9, isoliquiritin-MMP-9, liquiritin-TNF- α , isoliquiritin-TNF- α , ononin-MAPK1,

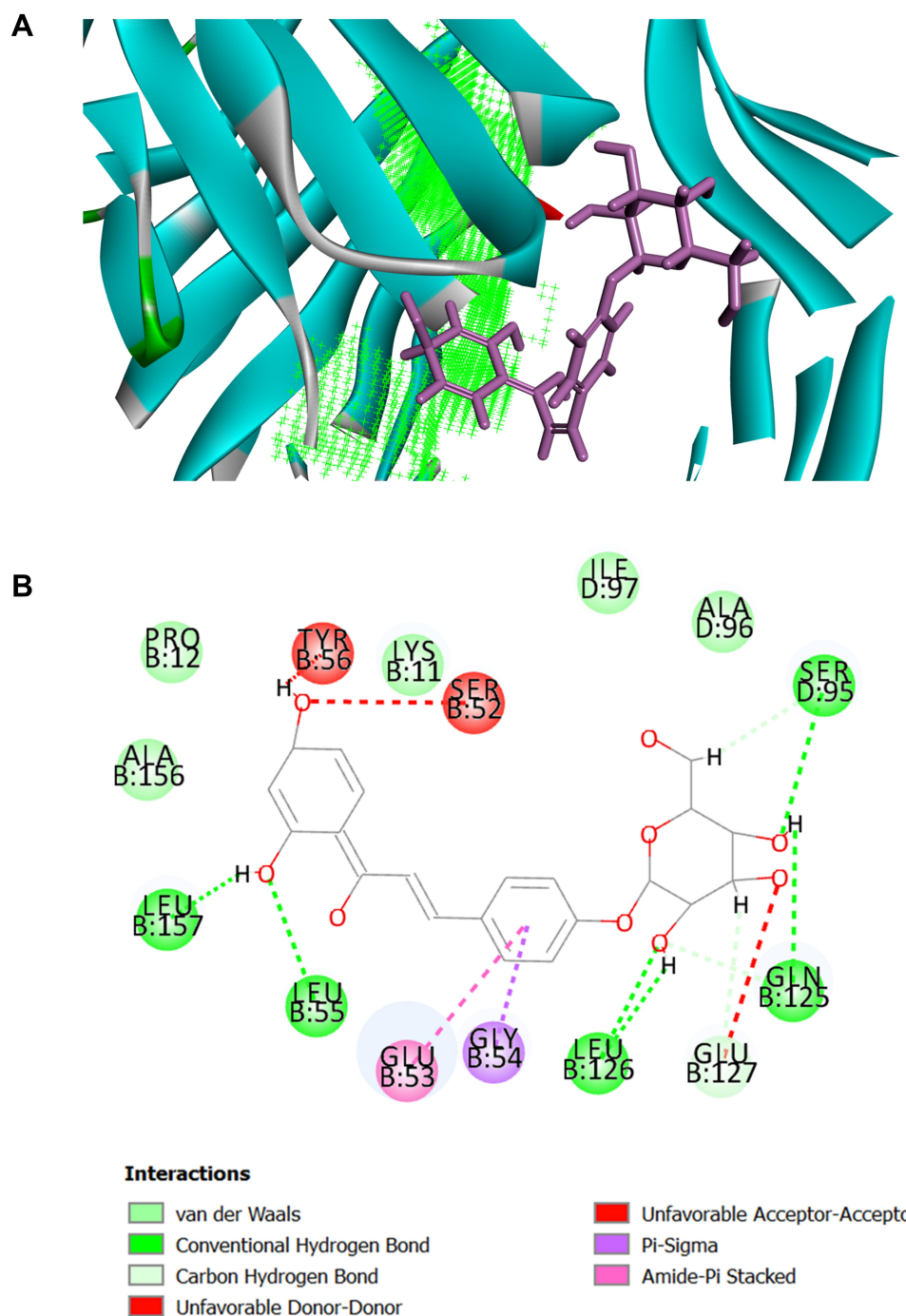


Figure 6 The binding modes of Isoliquiritin and protein TNF- α . **(A)** Schematic (3D) representation and **(B)** Schematic (2D) representation of the interplay between isoliquiritin and TNF- α (PDB IDchimeric 2AZ5). The compounds were shown as stick model with purple colored and the others were active site amino acid residues.

liquiritin-MAPK1, isopsoralenoside-MAPK1, psoralenoside-MAPK1, bavachin-MAPK1, bavachinin A-MAPK1, and isoliquiritin-MAPK1. As shown in [Figure 7](#), we have found some ideal molecule-protein pairs that have relatively better affinities, including formononetin-NFKB1 ($K_D = 32.5 \mu\text{M}$), isoliquiritin-TNF- α ($K_D = 3.6 \mu\text{M}$), isoliquiritin-MMP-9 ($K_D = 2.7 \mu\text{M}$), liquiritin-MMP-9 ($K_D = 105 \mu\text{M}$), bavachin-MAPK1

($K_D = 26.6 \mu\text{M}$), bavachinin A-MAPK1 ($K_D = 6.05 \mu\text{M}$), ononin-MAPK1 ($K_D = 41.4 \mu\text{M}$), and liquiritin-MAPK1 ($K_D = 382 \mu\text{M}$). It was interesting to note that liquiritin-MMP-9 ($K_a = 251 \text{ M}^{-1}\text{s}^{-1}$; $K_d = 2.64 \times 10^{-2} \text{ s}^{-1}$), bavachin-MAPK1 ($K_a = 9.29 \times 10^3 \text{ M}^{-1}\text{s}^{-1}$; $K_d = 2.47 \times 10^{-1} \text{ s}^{-1}$), and liquiritin-MAPK1 ($K_a = 258 \text{ M}^{-1}\text{s}^{-1}$; $K_d = 9.84 \times 10^{-2} \text{ s}^{-1}$) exhibit binding and/or release kinetics characteristics, which

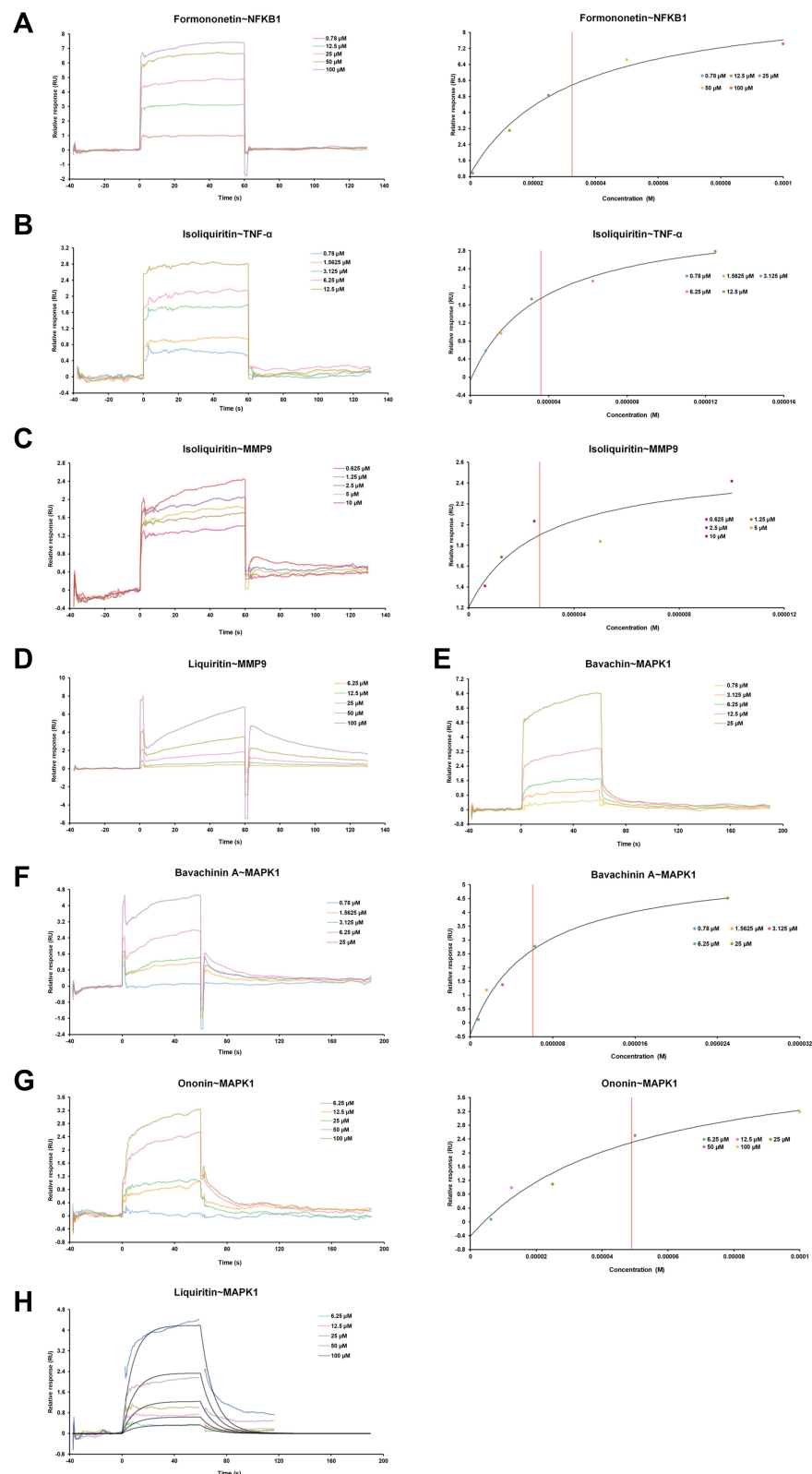


Figure 7 The surface plasmon resonance (SPR) assay of the interaction of formononetin with NFKB1, isoliquiritin with TNF- α , MMP9 proteins, liquiritin with MMP9, and bavachin, bavachinin A, liquiritin, ononin with MAPK I. **(A)** The SPR curves of formononetin binding to NFKB1 (left) and the representative binding curve (right). **(B)** The SPR curves of isoliquiritin binding to TNF- α (left) and the representative binding curve (right). **(C)** The SPR curves of isoliquiritin binding to MMP9 (left) and the representative binding curve (right). **(D)** The SPR curves of liquiritin binding to MMP9. **(E)** The SPR curves of bavachin binding to MAPK I. **(F)** The SPR curves of bavachinin A binding to MAPK I (left) and the representative binding curve (right). **(G)** The SPR curves of ononin binding to MAPK I (left) and the representative binding curve (right). **(H)** The SPR curves of liquiritin binding to MAPK I.

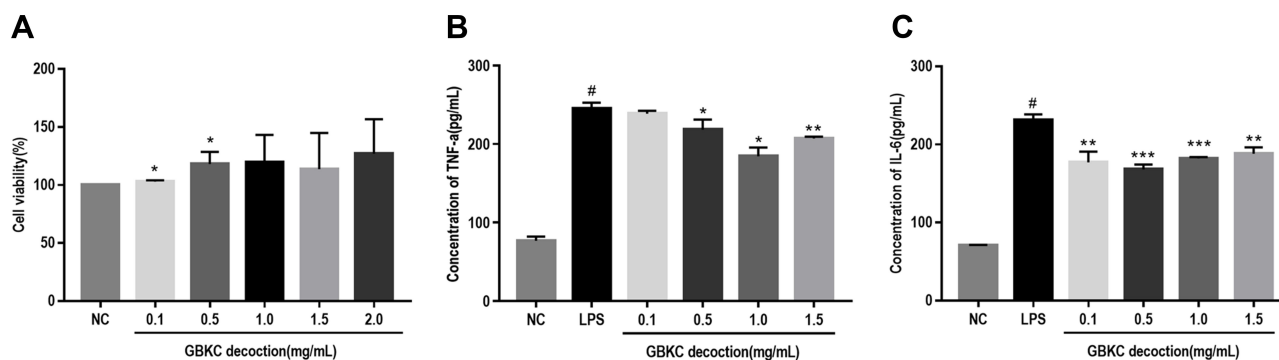


Figure 8 GBKC alleviates the secretions of IL-6 and TNF- α in LPS-induced MH-S cells. (A) Cell viability. Data are presented as the mean \pm SD (n=3) * p < 0.05, compared with normal control (NC) group. (B) TNF- α . (C) IL-6. Data are presented as the mean \pm SD (n=3) # p < 0.0001 compared with NC group. * p < 0.05; ** p < 0.005; *** p < 0.0005, compared with the LPS group.

may provide valuable information for further lead compound discovery. Collectively, these results demonstrate the validity of network pharmacology and molecular docking approaches, and the in-depth biological functions of these ligand-target pairs are worth exploring.

Effect of GBKC on TNF- α and IL-6 Protein Release After LPS Induction

CCK-8 assay indicated that no obvious cytotoxicity to MH-S cells was noted after GBKC treatment at the chosen concentrations (Figure 8A). As shown in Figure 8B and C, the levels of TNF- α and IL-6 were markedly increased in MH-S cells after LPS induction. However, GBKC administration caused statistically significant reductions in TNF- α and IL-6 secretion in a dose-dependent manner. The results also indicated that GBKC exhibited the best inhibitory effects on TNF- α at 1 mg/mL and IL-6 at 0.5 mg/mL.

Conclusions

TCM has accumulated valuable information regarding alleviation of CB progression. In the present study, 53 main components were identified from GBKC using UHPLC-LTQ-Orbitrap. According to the identified compounds, network and pathway analyses indicated that the TNF signaling pathway was closely related to 13 key targets (TNF- α , NF κ B, I κ B α , MAPK1, PIK3CA, PIK3R1, PIK3R2, PIK3R3, MMP-9, PTGS2, IL-1 β , IL-6, and CCL2) and the anti-CB effect of GBKC. In vitro experiments demonstrated that GBKC inhibited TNF- α and IL-6 release in a dose-dependent manner. Furthermore, molecular docking and SPR assays showed that several constituents exhibited good affinity to specific targets (formononetin-NFKB1, K_D = 32.5 μ M; isoliquiritin-TNF- α , K_D = 3.6 μ M; isoliquiritin-MMP-9, K_D = 2.7 μ M; bavachinin A-MAPK1, K_D = 6.05 μ M). This study provides a systematic

perspective to determine the anti-CB mechanisms of GBKC and also offers a novel approach to illustrate the chemical and pharmacological basis of other herbal medicines. Further experiments are required to validate these hypotheses to enhance the reliability of the results.

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Disclosure

The authors declared no conflict of interests for this work.

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