


Modulation of Chronic Inflammation by Quercetin: The Beneficial Effects on Obesity

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Shin Sato¹
Yuuka Mukai² 

¹Department of Nutrition, Aomori University of Health and Welfare, Aomori 030-8505, Japan; ²School of Nutrition and Dietetics, Faculty of Health and Social Work, Kanagawa University of Human Services, Kanagawa 238-8522, Japan

Abstract: Obesity has become a major risk factor for the development of chronic diseases such as insulin resistance, type 2 diabetes mellitus, and cardiovascular disease. Moreover, obesity induces chronic inflammation in adipose tissue, liver, skeletal muscle, and the vascular system. Quercetin is the major representative of the flavonoid subclass of flavonols, which is ubiquitously contained within natural plants such as green tea, and vegetables, including onions and apples. Researchers have focused greater attention to the beneficial physiological roles of quercetin, which has anti-oxidative, anti-inflammatory, and anti-fibrotic effects on insulin resistance and atherosclerosis in obesity-related diseases. Also, the anti-inflammatory effects of quercetin on intestinal microbiota have been demonstrated in obesity. In addition, there is increasing evidence that quercetin is associated with epigenetic activities in cancer, and in maternal undernutrition during gestation and lactation. In this review, we focus on the chemical properties of quercetin, its dietary sources in obesity, and its anti-inflammatory effects on insulin resistance, atherosclerosis, intestinal microbiota, and maternal under-nutrition with epigenetic activity.

Keywords: quercetin, inflammation, obesity, insulin resistance, atherosclerosis

Introduction

Obesity has become one of the most prevalent health problems globally and represents a major risk factor for the development of chronic diseases such as insulin resistance, type 2 diabetes mellitus, and cardiovascular disease.¹ Moreover, obesity induces chronic inflammation in adipose tissue, the liver, skeletal muscle, and the vascular system. Chronic inflammation induces release of proinflammatory cytokines such as tumor necrosis factor- α (TNF- α), monocyte chemoattractant protein-1 (MCP-1), and interleukin-6 (IL-6), and immune cell infiltration is closely associated with the development of insulin resistance through interactions with the insulin signaling pathway in adipose tissue and skeletal muscle^{2,3} and is closely linked to the pathogenesis of atherosclerosis in vessel walls.⁴

Quercetin, a flavonoid compound, is found in vegetables and plants such as onions, apples, and green tea. For instance, the dominant onion flavonoids have been determined to be quercetin, quercetin-3-O- β -glucoside (Q3G), quercetin-4'-O- β -glucoside (Q4'G), and quercetin-3,4'-di-O- β -glucoside (Q3,4'G).⁵ Interestingly, cooking methods affect the final flavonoid content; total quercetin abundance is increased 1.5-fold by microwave heating for 1 min, whereas the levels of Q4'G are decreased by boiling.⁶

To date, researchers have focused more attention on the beneficial physiological roles of quercetin, which has anti-oxidative, anti-inflammatory, and anti-fibrotic

Correspondence: Shin Sato
Department of Nutrition, Aomori University of Health and Welfare, Mase 58-1, Hamadate, Aomori 030-8505, Japan
Tel/Fax +81-17-765-4184
Email s_sato3@auhw.ac.jp

effects,⁷⁻⁹ although quercetin has been reported to have its potential pro-oxidative property in addition to its antioxidative property.^{10,11} More interestingly, there is increasing evidence that quercetin is associated with epigenetic changes in cancer¹² and in maternal under-nutrition during gestation and lactation.¹³ Therefore, studying the physiological roles of quercetin on chronic inflammation would contribute to preventive and therapeutic applications in obesity-related diseases.

In this review, we focused on chemical properties, dietary sources, and anti-inflammatory effects of quercetin on insulin resistance, atherosclerosis, intestinal microbiota, and maternal under-nutrition with epigenetic activity.

Chemical Structure and Dietary Source of Quercetin

Quercetin is the major representative of the flavonoid subclass of flavonols and is ubiquitously present in plants, fruits, and vegetables. The characteristic structure of flavonoids comprises a basic backbone of flavan, which has a C6-C3-C6 structure in which 2 benzene rings (C6) are bonded by 3 carbons (C3). Flavonols are present in plants as flavonoid-sugar compounds, which are referred to as glycosides in general. Figure 1 illustrates the structures of quercetin aglycone, which lacks a sugar moiety, and of quercetin- β -glucoside, which has been reported to be contained mainly in onions.^{14,15}

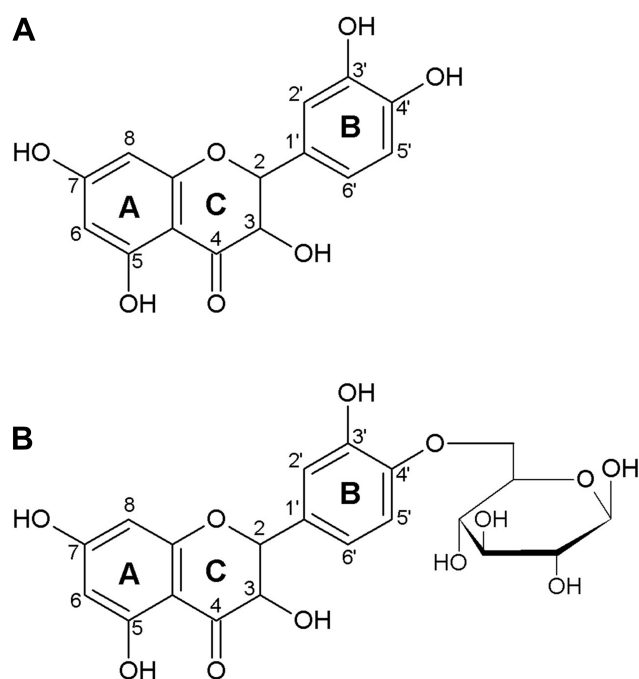


Figure 1 Structure of quercetin aglycone (A), and quercetin-4'-O- β -D-glucoside (B).

The types and amounts of flavonoids are extremely varied among plants. To date, the levels of quercetin in various foodstuffs have been reported in numerous studies.¹⁶⁻²⁰ When foods containing quercetin are consumed, the rate of intestinal absorption is higher for quercetin glycosides than aglycon.²¹ Hollman et al, who evaluated the bioavailability of quercetin, reported detecting increased concentrations of quercetin in plasma immediately after oral administration of onion and apple supplements.¹⁵ Absorbed quercetin is rapidly metabolized in the liver, and circulates as methyl, glucuronide, and sulfate metabolites.²² In human healthy adults, 163 diverse metabolites and quercetin conjugates such as quercetin-3-glucuronide, isorhamnetin-3-glucuronide, quercetin diglucuronide, and quercetin-30-sulphate were measured in the plasma after long-term quercetin-containing supplements; especially the concentrations of conjugates increased at the 1000 mg/day dose for 90 days.²³ Therefore, circulating quercetin and its metabolites in peripheral tissues are expected to exhibit bioavailability, resulting in anti-inflammatory effects.

Anti-Inflammatory Effects of Quercetin on Insulin Resistance

Insulin is well known to induce glucose uptake by binding to its receptors on the cell membrane of target organs such as the liver, skeletal muscle, and adipose tissue. When the insulin receptor (IR) is phosphorylated, the phosphorylation of insulin receptor substances (IRS) is increased. IRS activate the phosphatidylinositol 3-kinase (PI3K)/Akt pathway, which is largely responsible for the metabolic action of insulin, as well as the Ras/mitogen-activated protein kinase (MAPK) pathway, which mediates gene expression for insulin's effect.²⁴ When the PI3K/Akt pathway is activated, glucose transporter 4 (Glut4) expression and translocation are upregulated, promoting the uptake of glucose into cells.^{25,26} Insulin resistance is one of the main factors responsible for the onset and progression of diseases such as obesity, diabetes, and atherosclerosis. Insulin resistance is involved in impairment of the PI3K/Akt pathway in target organs such as adipose tissue and skeletal muscle, leading to the downregulation of Glut4 expression and its translocation. In obesity, excessive accumulation of fat in the visceral adipose tissue induces hypertrophy and dysfunction of adipocytes.^{27,28} The excessive amount of free fatty acids (FFAs) released from visceral adipose tissue is associated with interference in insulin's action.²⁹

Insulin resistance is closely associated with chronic low-grade inflammation through interactions with the insulin signaling pathway in the liver and adipose tissue.^{2,3} For instance, elevated levels of proinflammatory cytokines such as TNF- α , MCP-1, and IL-6, and of proinflammatory enzymes such as cyclooxygenases (COXs) and inducible nitric oxide synthase (iNOS) in adipose tissue, skeletal muscle, and neuronal systems, have been demonstrated to lead to the development of insulin resistance.^{30,31} In particular, TNF- α is one of the most important proinflammatory mediators which is involved in the development of insulin resistance.³²

Regarding the anti-inflammatory effects of quercetin on proinflammatory cytokine production, in macrophages and adipocytes, it decreases the expression levels of the inflammatory genes *TNF- α* , *IL-6*, *IL-1 β* , and *COX-2*, suppressing the activation of nuclear factor (NF- κ B) and c-Jun N-terminal kinase (JNK).³³ Dietary quercetin attenuates adipose tissue expansion and decreases the levels of serum *IL-6* and *MCP-1* mRNA in white adipose tissue in high fat diet (HFD)-induced obese mice.³⁴ In the hypothalamus, quercetin reduces the mRNA expression levels of *TNF- α* , *MCP-1*, and *IL-1 β* in HFD-fed obese mice.³⁵ Moreover, dietary quercetin and the combination of quercetin and catechin reduce metabolic parameters such as insulin concentrations and insulin resistance (HOMA-IR), and also levels of adipocytokine proteins such as TNF- α , visfatin, and resistin in adipose tissue in HFD-fed mice.³⁶ The effects of quercetin on proinflammatory enzymes have been investigated in several studies. For example, quercetin inhibits nitric oxide (NO) production and iNOS expression in PC cells treated with dopaminergic neurotoxin 6-hydroxydopamine, which induces neural damage.³⁷ In lipopolysaccharide-treated colon epithelial cells, the quercetin 3,7-O-dimethyl ether isolated from herbs downregulates the expression of iNOS and COX-2 proteins.³⁸ Intraperitoneal treatment with quercetin suppresses increased COX-2 expression in rats with unilateral ureteral obstruction, which is associated with increased inflammation and oxidative stress.³⁹ Importantly, quercetin may improve insulin resistance through inhibiting the production and expression of proinflammatory cytokines and/or enzymes.

Toll-like receptors (TLRs) are one of the key elements of the immune system, which play central roles in host cell recognition and responses to microbial pathogens.⁴⁰ Also, TLRs are known to regulate obesity-related inflammation and insulin resistance.⁴¹ The activation of TLR pathways encourages the production of proinflammatory cytokines

through the upregulation of transcription factors such as NF- κ B and activated protein 1 (AP-1).^{41,42} FAAs released from adipose tissue increases proinflammatory reactions and polarization of M1 macrophages (classically activated macrophages) through upregulation of the TLR4 pathway.⁴³ In addition, age-associated adipose tissue inflammation is reported to be reduced in TLR4-deficient mice fed HFD.⁴⁴ There is evidence to demonstrate that quercetin has potential to regulate inflammation through the TLR4/NF- κ B signaling pathway. Interestingly, quercetin treatment decreases cortical inflammation by inhibiting the TLR4/NF- κ B signaling pathway in hypoxia-ischemia-induced brain injury in neonatal rats.⁴⁵ In addition, the combined treatment of quercetin and catechin inhibits increased levels of proinflammatory mediators, including TNF- α , IL-1 β , and COX-2 in lipopolysaccharide (LPS)-stimulated macrophage RAW 264.7 cells, suggesting that the combined treatment may suppress the activation of TLR4-mediated NF- κ B and MAPK signaling pathways.⁴⁶ Therefore, quercetin contributes to the suppression of proinflammatory mediators through inhibition of the TLR4/NF- κ B signaling pathway.

Increased activation of AMP-activated protein kinase (AMPK) may play a role in suppression of proinflammatory mediator expression by inhibiting NF- κ B activation.⁴⁷ In addition, metformin, which is widely used as an anti-diabetic drug, ameliorates chronic low-grade inflammation and modulates macrophage polarization through the activation of AMPK in HFD-fed mice.⁴⁸ Moreover, AMPK is closely linked to fibrosis-promoting pathways, as well as inflammation.⁴⁹ AMPK activation reduces glomerular TGF- β , collagen, and fibronectin accumulation in several murine models of diabetic kidney disease.⁵⁰

Several studies have revealed that polyphenolic compounds, such as quercetin, resveratrol, and curcumin, enhance the phosphorylation of AMPK.⁵¹ For example, quercetin treatment showed anti-adipogenesis activity through the upregulation of AMPK and acetyl-CoA carboxylase (ACC) phosphorylation in 3T3-L1 preadipocytes.⁵² The expression of uncoupling protein 1 (UCP1), which plays an important role in increasing energy expenditure, increased in 3T3-L1 adipocytes treated with quercetin.⁵³ The authors suggested that quercetin increased the level of UCP1 in adipose tissues, accompanied by AMPK activation, and may lead to the prevention of obesity.⁵³ In C2C12 myotubes, quercetin ameliorated insulin resistance under inflammatory conditions with activation of AMPK.⁵⁴ Quercetin suppresses inflammation by modulating the AMPK pathway as well as the silent information regulator

(Sirt) 1.^{55–57} In addition, quercetin reduces macrophage infiltration of adipose tissues, lowers the levels of proinflammatory mediators, and upregulates AMPK phosphorylation and Sirt1 expression in adipose tissues of HFD-fed mice.⁵⁵ In a renal ischemia/reperfusion (I/R) animal model, quercetin improves renal I/R injury through upregulation of AMPK phosphorylation and activates autophagy during I/R.⁵⁸ When HFD-fed ApoE^{-/-} mice, an animal atherosclerosis model, received quercetin, the levels of TNF- α , IL-1 β , and IL-18 and ratios of microtubule-associated protein light chain 3A (LC3) II/I were restored,⁵⁹ suggesting that the attenuation of atherosclerosis by quercetin is associated with enhancement of autophagy. Therefore, quercetin may be involved in the resolution of inflammation through upregulation of autophagy activation.

In addition, insulin resistance is closely associated with inflammation and atrophy in skeletal muscle in the obese. In insulin resistance, the production of proinflammatory mediators such as TNF- α , IL-1, and IL-6 is upregulated in skeletal muscle and adipose tissue,^{60,61} which consequently leads to impaired insulin action and muscle fiber metabolism.^{62,63} Treatment with quercetin suppresses the upregulation of TNF- α and iNOS expression levels and restores the reduction of glucose uptake by L6 myotubes treated with palmitate.⁶⁴ In an obese animal model, quercetin suppresses obesity-related skeletal muscle atrophic factors such as MAFbx/atrogen-1 and muscle ring-finger protein 1 (MuRF1), accompanied by increases in heme oxygenase 1 (HO-1) levels, and inactivation of NF- κ B.⁶⁵ In addition, quercetin reduces macrophage accumulation and levels of inflammatory cytokines (eg, TNF- α and MCP-1) and the mRNA levels of MAFbx/atrogen-1 and MuRF1 in skeletal muscle of HFD-fed obese mice, suggesting that quercetin may prevent obesity-induced skeletal muscle atrophy through suppressing inflammatory responses.⁶⁶ Thus, quercetin could contribute to protect against obesity-related skeletal muscle atrophy by suppressing inflammatory mediators and macrophage infiltration in skeletal muscle, leading to improved insulin resistance.

Anti-Inflammatory Effects of Quercetin on Atherosclerosis

Inflammation is closely linked to the pathogenesis of atherosclerosis.⁴ Atherosclerosis is known as an inflammatory condition of vessel walls, characterized by infiltration involving mainly macrophage and T-cells; macrophages play key roles in the onset and development of atherosclerosis. As

stimulated by lipid deposition, hypertension, and oxidative stress during the early stages of atherosclerosis, monocytes and T-cells migrate from the circulation into the vascular intima to differentiate into macrophages. Macrophages phagocytose oxidized low-density lipoprotein (ox-LDL), and the number of cells is increased. Thereafter, macrophages are transformed into foam cells, which take up excessive ox-LDL. These conditions promote the development of atherosclerosis.⁶⁷ Furthermore, macrophages produce pro- and anti-inflammatory cytokines, including IL-1 β , and TNF- α , and emerge as a key mediator in the pathogenesis of atherosclerosis.

Quercetin suppresses the formation of ox-LDL-induced RAW264.7 macrophage foam cells, which is a foam cell model, reducing cellular lipid accumulation.⁶⁸ These phenomena can be interpreted; quercetin increases autophagy and decreases mammalian Ste20-like kinase 1 (MST1) during atherosclerotic progression.⁶⁹ Quercetin improves glucosamine-induced apoptosis and inflammation in human umbilical vein endothelial cells (HUVECs), which represent a model of vascular endothelial injury in the initial stages of atherosclerosis.⁷⁰ The combination of quercetin and docosahexaenoic acid (DHA) suppresses mRNA expression and phosphorylation of NF- κ B subunits p50 and p65, and ERK1/2 and JNK1/2 in LPS-stimulated RAW264.7 macrophage cells.⁷¹ Cao et al revealed that quercetin reduces otherwise elevated levels of TNF- α , IL-1 β , and IL-18, and enhances lowered autophagy activity in HFD-fed ApoE^{-/-} mice.⁵⁹ In addition, quercetin reduces the elevated mRNA expression of *TLRs* and *TNF- α* in high cholesterol diet-fed atherosclerotic rats, and quercetin also inhibits the nuclear translocation of NF- κ B and release of cytokines in ox-LDL-stimulated human peripheral blood mononuclear cells in vitro, suggesting that quercetin may be a promising agent for the prevention of atherosclerosis.⁷² Interestingly, quercetin treatment suppresses the progression of atherosclerosis in ApoE^{-/-} mice by inhibiting dendritic cell activation associated with the development of atherosclerosis through downregulation of CD80, CD86, MHC-II, IL-6, and IL-12 levels.⁷³

Clinical Studies Involving Quercetin Supplementation

In randomized clinical trials, participants who were overweight-to-obese patients with pre- and stage 1 hypertension were randomized to receive a dose of 162 mg/day quercetin from onion skin extract for 6 weeks. As a result, no

significant effects were observed in terms of serum C-reactive protein and TNF- α , as well as the levels of glucose, insulin, and HOMA-IR compared to a placebo group.⁷⁴ When healthy (pre) hypertensive men and women received quercetin-3-glucoside (160 mg/day) for 4 weeks, the levels of soluble endothelial selectin ($p = 0.03$) and IL-1 β ($p = 0.009$), and the z score for inflammation ($p = 0.02$) were lower compared to placebo.⁷⁵ No effects of supplementation of four quercetin capsules per day containing 100 mg quercetin dihydrate (100 mg/day for 10 weeks) were observed for the inflammatory markers, IL-6 and soluble vascular cell adhesion molecule-1, in healthy male smokers, although quercetin reduced the serum levels of total cholesterol and LDL-cholesterol compared to a placebo group.⁷⁶ Women with type 2 diabetes receiving quercetin (500 mg/day for 10 weeks) exhibited reduced systolic blood pressure. However, there were no effects on serum levels of IL-6, TNF- α , and C-reactive protein in comparing the quercetin and placebo groups.⁷⁷ In addition, women with polycystic ovary syndrome were assigned to 2 groups of quercetin treatment: 1 g/day (two 500 mg capsules) daily for 12 weeks, and placebo. Quercetin reduced HOMA-IR levels ($p < 0.001$) and slightly increased serum adiponectin compared to a placebo group, the authors suggesting that quercetin supplementation may improve adiponectin-mediated insulin resistance.⁷⁸ Furthermore, women with rheumatoid arthritis (RA) were assigned into quercetin (500 mg/day) or placebo groups for 8 weeks. The study's results indicated that there were no effects of quercetin on plasma oxidative and inflammatory status, or systolic and diastolic blood pressure in patients with RA.⁷⁹ On the other hand, Javadi et al demonstrated that plasma TNF α levels were significantly decreased in a quercetin group compared to placebo in women with RA allocated into a quercetin (500 mg/day) or placebo group for 8 weeks.⁸⁰ Based on previous clinical studies, the effects of quercetin remain unclear. Further studies with various design and sample sizes, and with different quercetin doses, are needed, considering the beneficial effects of quercetin observed in previous animal and cell investigations.

Anti-Inflammatory Effects of Quercetin via Intestinal Microbiota in Obesity

Intestinal microbiota, which exist with a certain diversity in the human gastrointestinal lumen, are also involved in obesity. For instance, it has been revealed in humans and in animal models that obesity is associated with changes in

the relative abundance of the two dominant bacterial divisions, the Bacteroidetes and the Firmicutes, with increased levels of Actinobacteria.^{81,82} Turnbaugh et al demonstrated that colonization of germ-free mice with microbiota from obese animals resulted in significantly greater increases in total body fat than colonization with a non-obese "lean" microbiota.⁸³ Although there are some studies indicating that the proportions of intestinal microbiota are different among populations⁸⁴ or are not involved in obesity,⁸⁵ imbalances and alterations in composition and/or function of intestinal microbiota, so-called "dysbiosis", have been identified to be related to onset and/or development of obesity. In addition, the gut microbiota may be associated with the onset and/or development of atherosclerosis including inflammation and lipid metabolism.⁸⁶ For example, berberine isolated from various medicinal plants showed the anti-atherosclerotic effect with the changes in composition and functions of gut microbiota, which is associated with anti-inflammatory and glucose and lipid metabolisms.⁸⁷

In recent years, attention has been focused on the effects of food components such as quercetin on intestinal microbiota in obesity. Etxeberria et al found that quercetin administration in rats effectively alleviates intestinal dysbiosis induced by a high-fat sucrose diet.⁸⁸ Quercetin supplementation attenuates the Firmicutes/Bacteroidetes ratio and inhibits the growth of bacterial species associated with diet-induced obesity. In another study, a combination of quercetin and resveratrol was able to ameliorate obesity and reverse the gut microbiota dysbiosis in HFD-fed rats.⁸⁹ Moreover, there are recent reports indicating that the relationship between quercetin and gut microbiota is associated with an anti-inflammatory status. *Citrobacter rodentium*-induced colitis in mice is well documented as an animal model of inflammatory bowel disease (IBD). Interestingly, pre-administered quercetin could alleviate *Citrobacter rodentium*-induced colitis, due to quercetin's ability to suppress pro-inflammatory cytokines such as IL-6 and TNF- α , and/or to modify gut microbiota. That is, pre-administration quercetin may enhance population numbers of *Bifidobacterium*, *Bacteroidetes*, and *Lactobacillus*, and may reduce those of *Fusobacterium* and *Enterococcus*.⁹⁰ In in vitro studies, quercetin reduces the levels of inflammatory mediators in LPS-stimulated macrophages by enhancing secretion of anti-inflammatory substances by *Bifidobacterium adolescentis*.⁹¹ Stearic acid is tentatively identified as the anti-inflammatory molecule from *B. adolescentis* stimulated by quercetin.⁹²

From these findings of the relationships between quercetin and microbiota, quercetin is expected to play a role in moderating intestinal inflammation in obesity. Indeed, a recent study using a mouse model of nonalcoholic fatty liver disease (NAFLD) associated with obesity demonstrates that quercetin can revert gut dysbiosis and related endotoxemia-mediated TLR4 pathway induction, with subsequent inhibition of inflammasome responses.⁹³ In addition, it is reported that the atherosclerotic lesions and size of plaques were reduced in mice fed HFD diets with oral quercetin treatment, by the alternation of the composition of the gut microbiota.⁹⁴ Quercetin treatment may contribute to mitigate the onset and/or development of atherosclerosis by modulating intestinal microbiota balance. In order to elucidate the anti-inflammatory effects of quercetin in obesity, including atherosclerosis, further investigations regarding intestinal microbiota are needed.

Epigenetic Activities of Quercetin During Inflammation

Epigenetics is referred to as heritable phenotypic alterations in gene expression that are independent of DNA sequence changes.^{95,96} Common epigenetic modifications in mammalian cells include changes in DNA methylation, histone modification, and expression of various non-coding microRNAs (miRNAs). DNA methylation is catalyzed by DNA methyltransferases (DNMTs) and is thought to act at promoters so as to induce gene silencing. Histone modifications to specific amino acid residues modulate chromatin structure and gene expression. For example, the histone acetylation state is thought to be adjusted by histone acetyltransferases (HATs) and histone deacetylases (HDACs). miRNAs are known to regulate gene expression post-transcriptionally and function in RNA-silencing.⁹⁷

Studies of epigenetic mechanisms have allowed advances in the understanding of cancer.^{98,99} Interestingly, polyphenolic compounds derived from plants seem to exert anti-tumor effects through epigenetic activities.^{100,101} Treatment with quercetin upregulates miR-503-5p and miR-6867-5p expression and exhibits the potential for anti-proliferative and anti-inflammatory actions in endometriosis implanted mouse models.¹⁰² Treatment with quercetin decreases global DNA methylation levels and the activity of DNMTs, HDACs, and histone methyltransferases (HMTs) in quercetin-treated HeLa cells, a human cervical cancer cell line.¹²

A combination of quercetin and butyrate with chemopreventive activity suppresses human esophageal cancer cell growth and downregulates the expression of DNMT1, NF- κ Bp65, HDAC1, and Cyclin D1.¹⁰³ In addition, quercetin enhances apoptosis by increasing the expression level of Fas ligand through the upregulation of HAT activity in human leukemia HL-60 cells.¹⁰⁴ Thus, quercetin is expected to act as a candidate natural therapeutic agent to prevent cancer through epigenetics activity. The HFD-fed mice showed hypermethylation in the peroxisome proliferator activated-receptor gamma coactivator 1 alpha (PGC-1 α) promoter and Pgc-1 α mRNA expression, which is a transcriptional coactivator, in skeletal muscle. Conversely, quercetin supplementation reduced the increases in DNA methylation and PGC-1 α expression,¹⁰⁵ suggesting that quercetin may regulate PGC-1 α expression through DNA methylation in obesity. Moreover, the treatment of quercetin inhibited inflammation in livers of nickel-treated mice by modulating nuclear factor-E2 related factor 2 (Nrf2) nuclear translocation and HO-1 activity and decreased DNMTs activity and DNA methylation level of the Nrf2 DNA.¹⁰⁶ Therefore, quercetin may epigenetically regulate the obesity and inflammation.

There is growing evidence that nutrients may modify epigenetic programs, thus regulating gene expression. For example, maternal under-nutrition or restriction of dietary protein during pregnancy leads to many diseases, including obesity, diabetes, and renal disease in adult offspring.^{107–110} Such diseases are closely associated with the development of chronic inflammation. Maternal over-nutrition in utero results in developmental programming of genes involved in obesity, inflammation, and pro-fibrogenic genes in the liver of the offspring.¹¹¹ The unfolding pattern of histone H3 lysine 4 trimethylation in children and mothers was associated with human undernutrition.¹¹² Although quercetin exerts anti-inflammatory action with a wide range of mechanisms of actions, few reports have addressed mechanisms by which quercetin modulates inflammation by regulating epigenetic pathways, which is linked to maternal under-nutrition.

We previously reported that the feeding of quercetin to protein-restricted dams during lactation upregulates AMPK activation in the liver of 23 week-old adult offspring.¹¹³ Significant increases in AMPK-associated phosphorylated ACC and endothelial nitric oxide synthase (eNOS) levels are found in the liver of such adult offspring. Quercetin treatment during lactation may lead to long-term alterations to the AMPK pathway in the liver of

adult offspring of protein-restricted dams. However, whether quercetin treatment during lactation directly activated AMPK in adult offspring remain unclear. The decreased levels of histone acetylation and increased levels of promoter methylation of PGC-1 α promoter methylation were correlated with the activity of AMPK in human placenta of diabetic mothers. When diabetic placental explant was treated with metformin, an anti-diabetic drug, AMPK was activated, concomitant with increased H3K27 acetylation and decreased PGC-1 α promoter methylation.¹¹⁴ Thus, because metformin as well as quercetin upregulate the AMPK activation,⁵¹ we hypothesized that quercetin treatment during lactation may activate AMPK via epigenetic regulation.

More interestingly, maternal quercetin intake during lactation may cause long-term alterations to inflammation and autophagy flux in the kidneys of high-fructose-diet fed adult female rat offspring.¹³ Maternal quercetin intake during lactation decreases the number of infiltrating macrophages and depresses *IL-6* mRNA levels in the kidneys of adult female offspring fed a high-fructose diet after birth. After inducing obesity in female rats fed a HFD, maternal quercetin treatment improved glucose metabolism, insulin sensitivity, hepatic inflammation, and adipose tissue deposition in the adult offspring of obese dams.¹¹⁵ On the other hand, maternal quercetin treatment, starting from 3 days before conception until the end of gestation, resulted in increased iron storage and decreased 8-oxo-dG levels in the liver of 12-week old adult murine offspring.¹¹⁶ In addition, the authors indicated that maternal quercetin treatment increases IL-1 β , IL-6, and IL-10 levels in the liver of the adult offspring. Importantly, maternal quercetin intake during gestation and/or lactation may modulate long-term alterations, including inflammatory responses in adult offspring. However, further experiments are required to clarify whether quercetin treatment during gestation and/or lactation contributes to the regulation of epigenetic pathways after birth.

Conclusions and Perspectives

This review provided recent evidence of the anti-inflammatory effects of quercetin. First, quercetin is involved in the attenuation of insulin resistance and atherosclerosis in obesity-related diseases. Insulin resistance is closely associated with the development of chronic low-grade inflammation. Quercetin may improve insulin resistance through inhibiting the production and expression of proinflammatory cytokines and/or enzymes. Importantly,

quercetin is associated with inhibition of the TLR4/NF- κ B signaling pathway. Based on previous cell culture studies and animal experiments, quercetin treatment is clearly required for preventative and therapeutic applications. However, different studies may also show the effects of quercetin in clinical studies. In this regard, the bioavailability of quercetin in organisms may be variable. It is necessary to establish more comprehensive studies that help to guide clinical studies. Second, some studies have shown the anti-inflammatory effects of quercetin on intestinal microbiota in obesity.

Prebiotics alter the intestinal microbiota and reduce serum levels of IL-6 in children with overweight or obesity.¹¹⁷ In addition, quercetin treatment may mitigate the onset and/or development of atherosclerosis.⁹⁴ Not only quercetin but also a combination of quercetin and prebiotic treatment may contribute to improve chronic inflammation in obesity-related diseases. Third, we summarized that maternal quercetin intake during lactation may exert long-term alterations in inflammation. Several studies have investigated epigenetic modulation mediated by quercetin. Quercetin is thought to act as a candidate therapeutic agent to prevent cancer through epigenetic activities. On the other hand, few reports have addressed suggestions that maternal quercetin exerts anti-inflammatory effects in adult offspring programmed by maternal under-nutrition and over-nutrition through modulation of epigenetic pathways. Further investigations are required to advance our understanding of the effects of maternal quercetin intake during gestation and/or lactation on anti-inflammatory activity in obesity- and age-related diseases.

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Disclosure

The authors declare no conflicts of interest.

References

1. Browning JD, Horton JD. Molecular mediators of hepatic steatosis and liver injury. *J Clin Invest*. 2004;114(2):147–152. doi:10.1172/JCI22422
2. Lumeng CN, DelProposto JB, Westcott DJ, Sattler AR. Phenotypic switching of adipose tissue macrophages with obesity is generated by spatiotemporal differences in macrophage subtypes. *Diabetes*. 2008;57(12):3239–3246. doi:10.2337/db08-0872

3. Osborn O, Olefsky JM. The cellular and signaling networks linking the immune system and metabolism in disease. *Nat Med*. 2012;18(3):363–374. doi:10.1038/nm.2627
4. Ross R. Atherosclerosis—an inflammatory disease. *N Engl J Med*. 1999;340(2):115–126. doi:10.1056/NEJM199901143400207
5. Zielinska D, Wiczowski W, Piskula MK. Determination of the relative contribution of quercetin and its glucosides to the antioxidant capacity of onion by cyclic voltammetry and spectrophotometric methods. *J Agric Food Chem*. 2008;56(10):3524–3531. doi:10.1021/jf073521f
6. Ioku K, Aoyama Y, Tokuno A, et al. Various cooking methods and the flavonoid content in onion. *J Nutr Sci Vitaminol*. 2001;47(1):78–83. doi:10.3177/jnsv.47.78
7. Li Y, Yao J, Han C, et al. Quercetin, inflammation and immunity. *Nutrients*. 2016;8(3):167. doi:10.3390/nu8030167
8. Marcolin E, San-Miguel B, Vallejo D, et al. Quercetin treatment ameliorates inflammation and fibrosis in mice with nonalcoholic steatohepatitis. *J Nutr*. 2012;142(10):1821–1828. doi:10.3945/jn.112.165274
9. Boots AW, Haenen GR, Bast A. Health effects of quercetin: from antioxidant to nutraceutical. *Eur J Pharmacol*. 2008;585(2–3):325–337. doi:10.1016/j.ejphar.2008.03.008
10. Yen GC, Duh PD, Tsai HL, Huang SL. Pro-oxidative properties of flavonoids in human lymphocytes. *Biosci Biotechnol Biochem*. 2003;67(6):1215–1222. doi:10.1271/bbb.67.1215
11. Dajas F, Abin-Carriquiry JA, Arredondo F, et al. Quercetin in brain diseases: potential and limits. *Neurochem Int*. 2015;89:140–148. doi:10.1016/j.neuint.2015.07.002
12. Kedhari Sundaram M, Hussain A, Haque S, et al. Quercetin modifies 5' CpG promoter methylation and reactivates various tumor suppressor genes by modulating epigenetic marks in human cervical cancer cells. *J Cell Biochem*. 2019;120(10):18357–18369. doi:10.1002/jcb.29147
13. Sato S, Norikura T, Mukai Y. Maternal quercetin intake during lactation attenuates renal inflammation and modulates autophagy flux in high-fructose-diet-fed female rat offspring exposed to maternal malnutrition. *Food Funct*. 2019;10(8):5018–5031. doi:10.1039/c9fo01134j
14. Khiari Z, Makris DP. Stability and transformation of major flavonols in onion (*Allium cepa*) solid wastes. *J Food Sci Technol*. 2012;49(4):489–494. doi:10.1007/s13197-010-0201-3
15. Hollman PC, van Trijp JM, Buysman MN, et al. Relative bioavailability of the antioxidant flavonoid quercetin from various foods in man. *FEBS Lett*. 1997;418(1–2):152–156. doi:10.1016/s0014-5793(97)01367-7
16. Nishimuro H, Ohnishi H, Sato M, et al. Estimated daily intake and seasonal food sources of quercetin in Japan. *Nutrients*. 2015;7(4):2345–2358. doi:10.3390/nu7042345
17. Lu X, Ross CF, Powers JR, Rasco BA. Determination of quercetins in onion (*Allium cepa*) using infrared spectroscopy. *J Agric Food Chem*. 2011;59(12):6376–6382. doi:10.1021/jf200953z
18. Mian KH, Mohamed S. Flavonoid (myricetin, quercetin, kaempferol, luteolin, and apigenin) content of edible tropical plants. *J Agric Food Chem*. 2001;49(6):3106–3112. doi:10.1021/jf000892m
19. Andarwulan N, Batari R, Sandrasari DA, et al. Flavonoid content and antioxidant activity of vegetables from Indonesia. *Food Chem*. 2010;121(4):1231–1235. doi:10.1016/j.foodchem.2010.01.033
20. Yang RY, Lin S, Kuo G. Content and distribution of flavonoids among 91 edible plant species. *Asia Pac J Clin Nutr*. 2008;17(Suppl 1):275–279.
21. Hollman PC, de Vries JH, van Leeuwen SD, et al. Absorption of dietary quercetin glycosides and quercetin in healthy ileostomy volunteers. *Am J Clin Nutr*. 1995;62(6):1276–1282. doi:10.1093/ajcn/62.6.1276
22. Lotito SB, Zhang WJ, Yang CS, et al. Metabolic conversion of dietary flavonoids alters their anti-inflammatory and antioxidant properties. *Free Radic Biol Med*. 2011;51(2):454–463. doi:10.1016/j.freeradbiomed.2011.04.032
23. Cialdella-Kam L, Nieman DC, Sha W, et al. Dose-response to 3 months of quercetin-containing supplements on metabolite and quercetin conjugate profile in adults. *Br J Nutr*. 2013;109(11):1923–1933. doi:10.1017/S0007114512003972
24. de Luca C, Olefsky JM. Inflammation and insulin resistance. *FEBS Lett*. 2008;582(1):97–105. doi:10.1016/j.febslet.2007.11.057
25. Russell RR 3rd, Bergeron R, Shulman GI, Young LH. Translocation of myocardial GLUT-4 and increased glucose uptake through activation of AMPK by AICAR. *Am J Physiol*. 1999;277(2):H643–H649. doi:10.1152/ajpheart.1999.277.2.H643
26. Mackenzie RW, Elliott BT. Akt/PKB activation and insulin signaling: a novel insulin signaling pathway in the treatment of type 2 diabetes. *Diabetes Metab Syndr Obes*. 2014;7:55–64. doi:10.2147/DMSO.S48260
27. Maury E, Brichard SM. Adipokine dysregulation, adipose tissue inflammation and metabolic syndrome. *Mol Cell Endocrinol*. 2010;314(1):1–16. doi:10.1016/j.mce.2009.07.031
28. Shoelson SE, Herrero L, Naaz A. Obesity, inflammation, and insulin resistance. *Gastroenterology*. 2007;132(6):2169–2180. doi:10.1053/j.gastro.2007.03.059
29. Boden G. Obesity, insulin resistance and free fatty acids. *Curr Opin Endocrinol Diabetes Obes*. 2011;18(2):139–143. doi:10.1097/MED.0b013e3283444b09
30. Glass CK, Olefsky JM. Inflammation and lipid signaling in the etiology of insulin resistance. *Cell Metab*. 2012;15(5):635–645. doi:10.1016/j.cmet.2012.04.001
31. Spagnuolo C, Moccia S, Russo GL. Anti-inflammatory effects of flavonoids in neurodegenerative disorders. *Eur J Med Chem*. 2018;153:105–115. doi:10.1016/j.ejmech.2017.09.001
32. Akash MSH, Rehman K, Liaqat A. Tumor necrosis factor- α : role in development of insulin resistance and pathogenesis of type 2 diabetes mellitus. *J Cell Biochem*. 2018;119(1):105–110. doi:10.1002/jcb.26174
33. Overman A, Chuang CC, McIntosh M. Quercetin attenuates inflammation in human macrophages and adipocytes exposed to macrophage-conditioned media. *Int J Obes*. 2011;35(9):1165–1172. doi:10.1038/ijo.2010.272
34. Forney LA, Lenard NR, Stewart LK, Henagan TM. Dietary quercetin attenuates adipose tissue expansion and inflammation and alters adipocyte morphology in a tissue-specific manner. *Int J Mol Sci*. 2018;19(3):895. doi:10.3390/ijms19030895
35. Yang J, Kim CS, Tu TH, et al. Quercetin protects obesity-induced hypothalamic inflammation by reducing microglia-mediated inflammatory responses via HO-1 induction. *Nutrients*. 2017;9(7):650. doi:10.3390/nu9070650
36. Vazquez Prieto MA, Bettaieb A, Rodriguez Lanzi C, et al. Catechin and quercetin attenuate adipose inflammation in fructose-fed rats and 3T3-L1 adipocytes. *Mol Nutr Food Res*. 2015;59(4):622–633. doi:10.1002/mnfr.201400631
37. Zhang ZJ, Cheang LC, Wang MW, Lee SM. Quercetin exerts a neuroprotective effect through inhibition of the iNOS/NO system and pro-inflammation gene expression in PC12 cells and in zebrafish. *Int J Mol Med*. 2011;27(2):195–203. doi:10.3892/ijmm.2010.571
38. Lee SG, Kim M, Kim CE, et al. Quercetin 3,7-O-dimethyl ether from *Siegesbeckia pubescens* suppress the production of inflammatory mediators in lipopolysaccharide-induced macrophages and colon epithelial cells. *Biosci Biotechnol Biochem*. 2016;80(11):2080–2086. doi:10.1080/09168451.2016.1204219
39. Carlsen I, Frokiaer J, Norregaard R. Quercetin attenuates cyclooxygenase-2 expression in acute ureteral obstruction. *Am J Physiol Renal Physiol*. 2015;308(11):F1297–F1305. doi:10.1152/ajprenal.00514.2014

40. Kawai T, Akira S. Toll-like receptors and their crosstalk with other innate receptors in infection and immunity. *Immunity*. 2011;34(5):637–650. doi:10.1016/j.immuni.2011.05.006
41. Jia L, Vianna CR, Fukuda M, et al. Hepatocyte Toll-like receptor 4 regulates obesity-induced inflammation and insulin resistance. *Nat Commun*. 2014;5:3878. doi:10.1038/ncomms4878
42. Kawasaki T, Kawai T. Toll-like receptor signaling pathways. *Front Immunol*. 2014;5:461. doi:10.3389/fimmu.2014.00461
43. Suganami T, Tanimoto-Koyama K, Nishida J, et al. Role of the Toll-like receptor 4/NF-kappaB pathway in saturated fatty acid-induced inflammatory changes in the interaction between adipocytes and macrophages. *Arterioscler Thromb Vasc Biol*. 2007;27(1):84–91. doi:10.1161/01.ATV.0000251608.09329.9a
44. Ghosh AK, O'Brien M, Mau T, Yung R. Toll-like receptor 4 (TLR4) deficient mice are protected from adipose tissue inflammation in aging. *Aging (Albany NY)*. 2017;9(9):1971–1982. doi:10.18632/aging.101288
45. Wu M, Liu F, Guo Q. Quercetin attenuates hypoxia-ischemia induced brain injury in neonatal rats by inhibiting TLR4/NF-kappaB signaling pathway. *Int Immunopharmacol*. 2019;74:105704. doi:10.1016/j.intimp.2019.105704
46. Li T, Li F, Liu X, et al. Synergistic anti-inflammatory effects of quercetin and catechin via inhibiting activation of TLR4-MyD88-mediated NF-kappaB and MAPK signaling pathways. *Phytother Res*. 2019;33(3):756–767. doi:10.1002/ptr.6268
47. Salminen A, Hyttinen JM, Kaarniranta K. AMP-activated protein kinase inhibits NF-kappaB signaling and inflammation: impact on healthspan and lifespan. *J Mol Med*. 2011;89(7):667–676. doi:10.1007/s00109-011-0748-0
48. Jing Y, Wu F, Li D, et al. Metformin improves obesity-associated inflammation by altering macrophages polarization. *Mol Cell Endocrinol*. 2018;461:256–264. doi:10.1016/j.mce.2017.09.025
49. Sharma K. Obesity, oxidative stress, and fibrosis in chronic kidney disease. *Kidney Int Suppl*. 2014;4(1):113–117. doi:10.1038/kisup.2014.21
50. Dugan LL, You YH, Ali SS, et al. AMPK dysregulation promotes diabetes-related reduction of superoxide and mitochondrial function. *J Clin Invest*. 2013;123(11):4888–4899. doi:10.1172/JCI66218
51. Wang S, Moustaid-Moussa N, Chen L, et al. Novel insights of dietary polyphenols and obesity. *J Nutr Biochem*. 2014;25(1):1–18. doi:10.1016/j.jnutbio.2013.09.001
52. Ahn J, Lee H, Kim S, et al. The anti-obesity effect of quercetin is mediated by the AMPK and MAPK signaling pathways. *Biochem Biophys Res Commun*. 2008;373(4):545–549. doi:10.1016/j.bbrc.2008.06.077
53. Choi H, Kim CS, Yu R. Quercetin upregulates uncoupling protein 1 in white/brown adipose tissues through sympathetic stimulation. *J Obes Metab Syndr*. 2018;27(2):102–109. doi:10.7570/jomes.2018.27.2.102
54. Liu K, Mei F, Wang Y, et al. Quercetin oppositely regulates insulin-mediated glucose disposal in skeletal muscle under normal and inflammatory conditions: the dual roles of AMPK activation. *Mol Nutr Food Res*. 2016;60(3):551–565. doi:10.1002/mnfr.201500509
55. Dong J, Zhang X, Zhang L, et al. Quercetin reduces obesity-associated ATM infiltration and inflammation in mice: a mechanism including AMPKalpha1/SIRT1. *J Lipid Res*. 2014;55(3):363–374. doi:10.1194/jlr.M038786
56. Hung CH, Chan SH, Chu PM, Tsai KL. Quercetin is a potent anti-atherosclerotic compound by activation of SIRT1 signaling under oxLDL stimulation. *Mol Nutr Food Res*. 2015;59(10):1905–1917. doi:10.1002/mnfr.201500144
57. Qiu L, Luo Y, Chen X. Quercetin attenuates mitochondrial dysfunction and biogenesis via upregulated AMPK/SIRT1 signaling pathway in OA rats. *Biomed Pharmacother*. 2018;103:1585–1591. doi:10.1016/j.biopha.2018.05.003
58. Chen BL, Wang LT, Huang KH, et al. Quercetin attenuates renal ischemia/reperfusion injury via an activation of AMP-activated protein kinase-regulated autophagy pathway. *J Nutr Biochem*. 2014;25(11):1226–1234. doi:10.1016/j.jnutbio.2014.05.013
59. Cao H, Jia Q, Shen D, et al. Quercetin has a protective effect on atherosclerosis via enhancement of autophagy in ApoE(-/-) mice. *Exp Ther Med*. 2019;18(4):2451–2458. doi:10.3892/etm.2019.7851
60. Uysal KT, Wiesbrock SM, Marino MW, Hotamisligil GS. Protection from obesity-induced insulin resistance in mice lacking TNF-alpha function. *Nature*. 1997;389(6651):610–614. doi:10.1038/39335
61. Furuya DT, Poletto AC, Favaro RR, et al. Anti-inflammatory effect of atorvastatin ameliorates insulin resistance in monosodium glutamate-treated obese mice. *Metabolism*. 2010;59(3):395–399. doi:10.1016/j.metabol.2009.08.011
62. Thrush AB, Heigenhauser GJ, Mullen KL, et al. Palmitate acutely induces insulin resistance in isolated muscle from obese but not lean humans. *Am J Physiol Regul Integr Comp Physiol*. 2008;294(4):R1205–R1212. doi:10.1152/ajpregu.00909.2007
63. Thrush AB, Antoun G, Nikpay M, et al. Diet-resistant obesity is characterized by a distinct plasma proteomic signature and impaired muscle fiber metabolism. *Int J Obes*. 2018;42(3):353–362. doi:10.1038/ijo.2017.286
64. Anhe GF, Okamoto MM, Kinote A, et al. Quercetin decreases inflammatory response and increases insulin action in skeletal muscle of ob/ob mice and in L6 myotubes. *Eur J Pharmacol*. 2012;689(1–3):285–293. doi:10.1016/j.ejphar.2012.06.007
65. Kim Y, Kim CS, Joe Y, Chung HT, Ha TY, Yu R. Quercetin reduces tumor necrosis factor alpha-induced muscle atrophy by upregulation of heme oxygenase-1. *J Med Food*. 2018;21(6):551–559. doi:10.1089/jmf.2017.4108
66. Le NH, Kim CS, Park T, et al. Quercetin protects against obesity-induced skeletal muscle inflammation and atrophy. *Mediators Inflamm*. 2014;2014:834294. doi:10.1155/2014/834294
67. Taleb S. Inflammation in atherosclerosis. *Arch Cardiovasc Dis*. 2016;109(12):708–715. doi:10.1016/j.acvd.2016.04.002
68. Cao H, Jia Q, Yan L, Chen C, Xing S, Shen D. Quercetin suppresses the progression of atherosclerosis by regulating MST1-mediated autophagy in ox-LDL-induced RAW264.7 macrophage foam cells. *Int J Mol Sci*. 2019;20(23):6093. doi:10.3390/ijms20236093
69. Wang T, Zhang L, Hu J, et al. Mst1 participates in the atherosclerosis progression through macrophage autophagy inhibition and macrophage apoptosis enhancement. *J Mol Cell Cardiol*. 2016;98:108–116. doi:10.1016/j.yjmcc.2016.08.002
70. Cai X, Bao L, Ding Y, et al. Quercetin alleviates cell apoptosis and inflammation via the ER stress pathway in vascular endothelial cells cultured in high concentrations of glucosamine. *Mol Med Rep*. 2017;15(2):825–832. doi:10.3892/mmr.2016.6054
71. Si TL, Liu Q, Ren YF, et al. Enhanced anti-inflammatory effects of DHA and quercetin in lipopolysaccharide-induced RAW264.7 macrophages by inhibiting NF-kappaB and MAPK activation. *Mol Med Rep*. 2016;14(1):499–508. doi:10.3892/mmr.2016.5259
72. Bhaskar S, Helen A. Quercetin modulates toll-like receptor-mediated protein kinase signaling pathways in oxLDL-challenged human PBMCs and regulates TLR-activated atherosclerotic inflammation in hypercholesterolemic rats. *Mol Cell Biochem*. 2016;423(1–2):53–65. doi:10.1007/s11010-016-2824-9
73. Lin W, Wang W, Wang D, Ling W. Quercetin protects against atherosclerosis by inhibiting dendritic cell activation. *Mol Nutr Food Res*. 2017;61(9):1700031. doi:10.1002/mnfr.201700031
74. Brull V, Burak C, Stoffel-Wagner B, et al. No effects of quercetin from onion skin extract on serum leptin and adiponectin concentrations in overweight-to-obese patients with (pre-)hypertension: a randomized double-blinded, placebo-controlled crossover trial. *Eur J Nutr*. 2017;56(7):2265–2275. doi:10.1007/s00394-016-1267-0

75. Dower JI, Geleijnse JM, Gijsbers L, et al. Supplementation of the pure flavonoids epicatechin and quercetin affects some biomarkers of endothelial dysfunction and inflammation in (pre)hypertensive adults: a randomized double-blind, placebo-controlled, crossover trial. *J Nutr.* 2015;145(7):1459–1463. doi:10.3945/jn.115.211888
76. Lee KH, Park E, Lee HJ, et al. Effects of daily quercetin-rich supplementation on cardiometabolic risks in male smokers. *Nutr Res Pract.* 2011;5(1):28–33. doi:10.4162/nrp.2011.5.1.28
77. Zahedi M, Ghiasvand R, Feizi A, et al. Does quercetin improve cardiovascular risk factors and inflammatory biomarkers in women with type 2 diabetes: a double-blind randomized controlled clinical trial. *Int J Prev Med.* 2013;4(7):777–785.
78. Rezvan N, Moini A, Janani L, et al. Effects of quercetin on adiponectin-mediated insulin sensitivity in polycystic ovary syndrome: a randomized placebo-controlled double-blind clinical trial. *Horm Metab Res.* 2017;49(2):115–121. doi:10.1055/s-0042-118705
79. Javadi F, Egtesadi S, Ahmadzadeh A, et al. The effect of quercetin on plasma oxidative status, C-reactive protein and blood pressure in women with rheumatoid arthritis. *Int J Prev Med.* 2014;5(3):293–301.
80. Javadi F, Ahmadzadeh A, Egtesadi S, et al. The effect of quercetin on inflammatory factors and clinical symptoms in women with rheumatoid arthritis: a double-blind, randomized controlled trial. *J Am Coll Nutr.* 2017;36(1):9–15. doi:10.1080/07315724.2016.1140093
81. Ley RE, Turnbaugh PJ, Klein S, Gordon JI. Microbial ecology: human gut microbes associated with obesity. *Nature.* 2006;444(7122):1022–1023. doi:10.1038/4441022a
82. Turnbaugh PJ, Hamady M, Yatsunenko T, et al. A core gut microbiome in obese and lean twins. *Nature.* 2009;457(7228):480–484. doi:10.1038/nature07540
83. Turnbaugh PJ, Ley RE, Mahowald MA, et al. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature.* 2006;444(7122):1027–1031. doi:10.1038/nature05414
84. Andoh A, Nishida A, Takahashi K, et al. Comparison of the gut microbial community between obese and lean peoples using 16S gene sequencing in a Japanese population. *J Clin Biochem Nutr.* 2016;59(1):65–70. doi:10.3164/jcbs.15-152
85. Arumugam M, Raes J, Pelletier E, et al. Enterotypes of the human gut microbiome. *Nature.* 2011;473(7346):174–180. doi:10.1038/nature09944
86. Jonsson AL, Bäckhed F. Role of gut microbiota in atherosclerosis. *Nat Rev Cardiol.* 2017;14(2):79–87. doi:10.1038/nrcardio.2016.183
87. Wu M, Yang S, Wang S, et al. Effect of berberine on atherosclerosis and gut microbiota modulation and their correlation in high-fat diet-fed ApoE^{-/-} mice. *Front Pharmacol.* 2020;11:223. doi:10.3389/fphar.2020.00223
88. Etxeberria U, Arias N, Boque N, et al. Reshaping faecal gut microbiota composition by the intake of trans-resveratrol and quercetin in high-fat sucrose diet-fed rats. *J Nutr Biochem.* 2015;26(6):651–660. doi:10.1016/j.jnutbio.2015.01.002
89. Zhao L, Zhang Q, Ma W, et al. A combination of quercetin and resveratrol reduces obesity in high-fat diet-fed rats by modulation of gut microbiota. *Food Funct.* 2017;8(12):4644–4656. doi:10.1039/c7fo01383c
90. Lin R, Piao M, Song Y. Dietary quercetin increases colonic microbial diversity and attenuates colitis severity in *Citrobacter rodentium*-infected mice. *Front Microbiol.* 2019;10:1092. doi:10.3389/fmicb.2019.01092
91. Kawabata K, Sugiyama Y, Sakano T, Ohigashi H. Flavonols enhanced production of anti-inflammatory substance(s) by *Bifidobacterium adolescentis*: prebiotic actions of galangin, quercetin, and fisetin. *Biofactors.* 2013;39(4):422–429. doi:10.1002/biof.1081
92. Kawabata K, Baba N, Sakano T, et al. Functional properties of anti-inflammatory substances from quercetin-treated *Bifidobacterium adolescentis*. *Biosci Biotechnol Biochem.* 2018;82(4):689–697. doi:10.1080/09168451.2017.1401916
93. Porras D, Nistal E, Martinez-Florez S, et al. Protective effect of quercetin on high-fat diet-induced non-alcoholic fatty liver disease in mice is mediated by modulating intestinal microbiota imbalance and related gut-liver axis activation. *Free Radic Biol Med.* 2017;102:188–202. doi:10.1016/j.freeradbiomed.2016.11.037
94. Nie J, Zhang L, Zhao G, Du X. Quercetin reduces atherosclerotic lesions by altering the gut microbiota and reducing atherogenic lipid metabolites. *J Appl Microbiol.* 2019;127(6):1824–1834. doi:10.1111/jam.14441
95. Bird A. Perceptions of epigenetics. *Nature.* 2007;447(7143):396–398. doi:10.1038/nature05913
96. Nagase H, Ghosh S. Epigenetics: differential DNA methylation in mammalian somatic tissues. *FEBS J.* 2008;275(8):1617–1623. doi:10.1111/j.1742-4658.2008.06330.x
97. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell.* 2004;116(2):281–297. doi:10.1016/s0092-8674(04)00045-5
98. Jones PA, Baylin SB. The fundamental role of epigenetic events in cancer. *Nat Rev Genet.* 2002;3(6):415–428. doi:10.1038/nrg816
99. Sharma S, Kelly TK, Jones PA. Epigenetics in cancer. *Carcinogenesis.* 2010;31(1):27–36. doi:10.1093/carcin/bgp220
100. Busch C, Burkard M, Leischner C, et al. Epigenetic activities of flavonoids in the prevention and treatment of cancer. *Clin Epigenetics.* 2015;7:64. doi:10.1186/s13148-015-0095-z
101. Carlos-Reyes A, Lopez-Gonzalez JS, Meneses-Flores M, et al. Dietary compounds as epigenetic modulating agents in cancer. *Front Genet.* 2019;10:79. doi:10.3389/fgene.2019.00079
102. Park S, Lim W, Bazer FW, et al. Quercetin inhibits proliferation of endometriosis regulating cyclin D1 and its target microRNAs in vitro and in vivo. *J Nutr Biochem.* 2019;63:87–100. doi:10.1016/j.jnutbio.2018.09.024
103. Zheng NG, Wang JL, Yang SL, Wu JL. Aberrant epigenetic alteration in Eca9706 cells modulated by nanoliposomal quercetin combined with butyrate mediated via epigenetic-NF-kappaB signaling. *Asian Pac J Cancer Prev.* 2014;15(11):4539–4543. doi:10.7314/apjcp.2014.15.11.4539
104. Lee WJ, Chen YR, Tseng TH. Quercetin induces FasL-related apoptosis, in part, through promotion of histone H3 acetylation in human leukemia HL-60 cells. *Oncol Rep.* 2011;25(2):583–591. doi:10.3892/or.2010.1097
105. Devarshi PP, Jones AD, Taylor EM, et al. Quercetin and quercetin-rich red onion extract alter Pgc-1alpha promoter methylation and splice variant expression. *PPAR Res.* 2017;2017:3235693. doi:10.1155/2017/3235693
106. Liu CM, Ma JQ, Xie WR, et al. Quercetin protects mouse liver against nickel-induced DNA methylation and inflammation associated with the Nrf2/HO-1 and p38/STAT1/NF-kappaB pathway. *Food Chem Toxicol.* 2015;82:19–26. doi:10.1016/j.fct.2015.05.001
107. Bol VV, Reusens BM, Remacle CA. Postnatal catch-up growth after fetal protein restriction programs proliferation of rat preadipocytes. *Obesity.* 2008;16(12):2760–2763. doi:10.1038/oby.2008.417
108. Victora CG, Adair L, Fall C, et al. Maternal and child undernutrition: consequences for adult health and human capital. *Lancet.* 2008;371(9609):340–357. doi:10.1016/S0140-6736(07)61692-4
109. Sharkey D, Gardner DS, Fainberg HP, et al. Maternal nutrient restriction during pregnancy differentially alters the unfolded protein response in adipose and renal tissue of obese juvenile offspring. *FASEB J.* 2009;23(5):1314–1324. doi:10.1096/fj.08-114330

110. Luyckx VA, Brenner BM. Birth weight, malnutrition and kidney-associated outcomes—a global concern. *Nat Rev Nephrol.* 2015;11(3):135–149. doi:10.1038/nrneph.2014.251
111. Wankhade UD, Zhong Y, Kang P, et al. Enhanced offspring predisposition to steatohepatitis with maternal high-fat diet is associated with epigenetic and microbiome alterations. *PLoS One.* 2017;12(4):e0175675. doi:10.1371/journal.pone.0175675
112. Uchiyama R, Kupkova K, Shetty SJ, et al. Histone H3 lysine 4 methylation signature associated with human undernutrition. *Proc Natl Acad Sci USA.* 2018;115(48):E11264–E11273. doi:10.1073/pnas.1722125115
113. Sato S, Mukai Y, Saito T. Quercetin intake during lactation modulates the AMP-activated protein kinase pathway in the livers of adult male rat offspring programmed by maternal protein restriction. *J Nutr Biochem.* 2013;24(1):118–123. doi:10.1016/j.jnutbio.2012.03.007
114. Jiang S, Teague AM, Tryggestad JB, et al. Role of metformin in epigenetic regulation of placental mitochondrial biogenesis in maternal diabetes. *Sci Rep.* 2020;10(1):8314. doi:10.1038/s41598-020-65415-0
115. Wu Z, Zhao J, Xu H, et al. Maternal quercetin administration during gestation and lactation decrease endoplasmic reticulum stress and related inflammation in the adult offspring of obese female rats. *Eur J Nutr.* 2014;53(8):1669–1683. doi:10.1007/s00394-014-0673-4
116. Vanhees K, Godschalk RW, Sanders A, et al. Maternal quercetin intake during pregnancy results in an adapted iron homeostasis at adulthood. *Toxicology.* 2011;290(2–3):350–358. doi:10.1016/j.tox.2011.10.017
117. Nicolucci AC, Hume MP, Martinez I, et al. Probiotics reduce body fat and alter intestinal microbiota in children who are overweight or with obesity. *Gastroenterology.* 2017;153(3):711–722. doi:10.1053/j.gastro.2017.05.055

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