

In Silico, Cytotoxic and Antioxidant Potential of Novel Ester, 3-hydroxyoctyl -5- *trans*-docosenoate Isolated from *Anchusa arvensis* (L.) M.Bieb. Against HepG-2 Cancer Cells

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Background: Cancer is one of the chronic health conditions worldwide. Various therapeutically active compounds from medicinal plants were the current focus of this research in order to uncover a treatment regimen for cancer. *Anchusa arvensis* (*A. anchusa*) (L.) M.Bieb. contains many biologically active compounds.

Methods: In the current study, new ester 3-hydroxyoctyl -5- *trans*-docosenoate (compound-1) was isolated from the chloroform soluble fraction of *A. anchusa* using column chromatography. Using MTT assay, the anticancer effect of the compound was determined in human hepatocellular carcinoma cells (HepG-2) compared with normal epithelial cell line (Vero). DPPH and ABTS radical scavenging assays were performed to assess the antioxidant potential. The Molecular Operating Environment (MOE-2016) tool was used against tyrosine kinase.

Results: The structure of the compound was elucidated based on IR, EI, and NMR spectroscopy technique. It exhibited a considerable cytotoxic effect against HepG-2 cell lines with IC₅₀ value of 6.50 ± 0.70 µg/mL in comparison to positive control (doxorubicin) which showed IC₅₀ value of 1.3±0.21 µg/mL. The compound did not show a cytotoxic effect against normal epithelial cell line (Vero). The compound also exhibited significant DPPH scavenging ability with IC₅₀ value of 12 ± 0.80 µg/mL, whereas ascorbic acid, used as positive control, demonstrated activity with IC₅₀ = 05 ± 0.15 µg/mL. Similarly, it showed ABTS radical scavenging ability (IC₅₀ = 130 ± 0.20 µg/mL) compared with the value obtained for ascorbic acid (06 ± 0.85 µg/mL). In docking studies using MOE-2016 tool, it was observed that compound-1 was highly bound to tyrosine kinase by having two hydrogen bonds at the hinge region. This good bonding network by the compound might be one of the reasons for showing significant activity against this enzyme.

Conclusion: Our findings led to the isolation of a new compound from *A. anchusa* which has significant cytotoxic activity against HepG-2 cell lines with marked antioxidant potential.

Keywords: cancer, *Anchusa arvensis*, 3-hydroxyoctyl-5, *trans*-docosenoate, cytotoxicity against HepG-2 cell lines, antioxidant

Introduction

Cancer is a major public health problem worldwide and is the second leading cause of death after cardiovascular disorders.¹⁻³ Hepatocellular carcinoma is one of the various types of cancer that leads to more than 600,000 deaths worldwide on an annual basis.^{4,5}

Plants have a long history of use in cancer therapy⁶⁻⁸ and several plant-derived anti-cancer drugs are still in clinical use including vincristine, vinblastine, etoposide, camptothecin derivatives, topotecan, irinotecan and paclitaxel, all isolated from plants.^{9,10} However, there is a dire need for development of new and novel cytotoxic drugs, drug combinations, and chemotherapy strategies, by exploration of a pool of synthetic, biological, and natural products.¹¹

Tyrosine kinases (TKs) are beneficial for the study of drug targeting molecules which are used for different types of cancer.^{12,13} TKs play an important role in the variety of growth factor signalling.¹⁴ TKs are vital for different biological procedures like apoptosis, growth and migration, differentiation, and metabolism for both external and internal stimuli. TKs are enzymes which initiate the process of phosphorylation by utilizing ATP for specific tyrosine residues. The covalent and enzymatic modification of protein following biosynthesis is an important integral part for the maintenance of normal cellular communication and homeostasis.¹⁵

A. arvensis which belongs to the family Boraginaceae, has traditionally been used for the treatment of arthritis, rheumatism, gout, cancer, gastroprotection, hypotension and hypoglycemic effects.¹⁶⁻¹⁹ *A. arvensis* contains phytochemical constituents like enantiomeric pigments of naphthoquinone of alkannins and group of shikonins. The enantiomeric alkannins, shikonins and their derivatives comprise a new class of drugs, exhibiting antimicrobial, antitumor, and anti-inflammatory activities.²⁰ Herein we report the chromatographic isolation of a new aliphatic ester from *A. arvensis* which was evaluated against HepG2 cell lines. The results revealed that compound-1 is an antioxidant agent due to its DPHH and ABTS radical scavenging. The in-vitro effect was further evaluated through molecular docking while targeting tyrosine kinase as target enzyme.

Materials and Methods

Chemicals and Equipment

Methanol, n-hexane, chloroform, and ethyl acetate were purchased from Merck, Germany. 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), and dimethyl sulfoxide (DMSO) were purchased from Sigma Chemicals Co. (St. Louis, USA). Silica gel 60 F₂₅₄ card (Merck) was used for thin layer chromatography (TLC) while column chromatography was carried out on silica

gel (70–230 mesh, Scharlau). 2,2-diphenyl-1-picrylhydrazyl (Sigma-Aldrich, USA, CAS 1898-66-4). ABTS (Sigma-Aldrich, USA, CAS 30931-67-0). All chemicals used in experiments were of analytical grade. ELISA reader (ELx800 BioTek) was used for measuring absorbance. The extracts of *A. arvensis* were concentrated by using Rotary evaporator (Rota vapour 210). Nuclear magnetic resonance (NMR) spectra were recorded on Bruker AVANCE-600 MHz NMR spectrometer with 5 mm BBO probe and microcoil-based CapNMR probe.

Extraction and Isolation

The plant material was obtained from district Karak, Pakistan in April 2016. The botanical identity of the plant was determined by the plant taxonomist, Dr. Waheed Murad, Department of Botany, Kohat University of Science and Technology. A voucher specimen with catalog no KUH-1001 was submitted to the herbarium of Department of Botany, Kohat University of Science and Technology. The air-dried plant was chopped and ground to coarse powder and macerated in methanol at room temperature for 14 days.^{21,22} The methanol soluble residue was filtered off and concentrated under vacuum rotary evaporator at 40°C. A crude methanol extract (600 gm) was obtained from the filtrates. The crude extract (500 gm) was suspended in distilled water and sequentially fractionated with *n*-hexane (3 x 500 mL), chloroform (3 x 500 mL), and ethyl acetate (3 x 500 mL), yielding *n*-hexane (140 gm), chloroform (160 gm), ethyl acetate (90 gm), and aqueous (60 gm) fractions respectively.^{23,24} The chloroform extract was subjected to column chromatography over silica gel (70–230 mesh). Elution was started with *n*-hexane/ethyl acetate in increasing order of polarity (95:5, 90:10, 85:15, 75:25, 60:40) resulting in isolation of compound-1. The structure of the compound was determined on the basis of IR, EI, NMR spectroscopy including 2D NMR techniques and chemical methods.²⁵

Cell Culture

Human hepatocellular carcinoma (HepG-2) and normal Vero cell lines were obtained from National Centre of Excellence in Molecular Biology (CEMB), University of the Punjab, Lahore, Pakistan. The cell lines were cultured and preserved in the Department of Biotechnology, Quaid-i-Azam University, Islamabad, Pakistan until they were used in experiments. The cells were cultured in Dulbecco's Modified Eagle Medium (DMEM)

supplemented with 10% fetal bovine serum (FBS), HEPES, 3% glutamine, sodium bicarbonate and antibiotic (streptomycin/penicillin). The cultured cell lines were maintained in a humidified CO₂ incubator at 37°C.

In-vitro Cytotoxicity Assay

We performed MTT assay to evaluate the cytotoxicity of the isolated compound.^{26,27} HepG2 and normal Vero cell lines were seeded in 200 µL culture medium at a density of 5×10^3 cells/well in 96-well plates. After incubation for 24 h, the HepG2 and normal cell lines were treated with different concentrations (200, 100, 50, 10, 5 µg/mL) of the compound and incubated for a further 24 h. Following washing and incubation with 4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide solution (20 µL of 5 mg/mL) at 37°C for 3 h, cells were lysed with dimethyl sulfoxide (DMSO). Doxorubicin and DMSO were used as positive and negative controls respectively. The MTT dye was reduced by succinic dehydrogenase in the mitochondria of viable cells to formazan purple crystals. Absorbance (OD) was recorded at 570 nm using a microplate reader. The percentage of cytotoxicity compared to the untreated cells was determined using the following equation:

$$\text{Cell viability (\%)} = \frac{\text{OD of treated cells}}{\text{OD of control cells}} \times 100$$

The results were generated from three independent experiments; each experiment was performed in triplicate. The IC₅₀ values were calculated from dose-response curve.

DPPH Radical Scavenging Assay

In the present study we determined the antioxidant ability of the compound by using DPPH (2,2-diphenyl-1-picrylhydrazyl) free radical scavenging method.²⁸ Various concentrations (1000, 500, 250, 125, 62.5, 31 and 15.5 µg/mL) of compound-1 (0.1 mL) were added to methanolic solution (0.004%) of DPPH. Absorbance of samples was measured at 517 nm wavelength using UV spectrophotometer after of 30 mins interval. Ascorbic acid was used as positive control; percent scavenging activity of samples was calculated by using the equation,

$$\text{Antioxidant effect (\%)} = \frac{\text{control absorbance} - \text{sample absorbance}}{\text{control absorbance}} \times 100$$

Each experiment was done in triplicate and inhibition graphs were constructed using the GraphPad prism program (GraphPad, San Diego, California, USA) and IC₅₀ values (concentration required for 50% reduction of DPPH free radicals) were determined.

ABTS Free Radical Scavenging Assay

The antioxidant potential of compound-1 was determined using 2, 2-azino bis [3-ethylbenzthiazoline]-6-sulfonic acid (ABTS) method.²⁹ The ABTS free radical scavenging assay is based on the ability of antioxidants to scavenge ABTS radical cation causing a reduction in absorbance. Briefly, solutions of potassium persulphate (K₂S₂O₈) 2.45 mM and ABTS 7 mM were prepared and mixed well. The resultant mixture was kept in the dark at room temperature to get dark colored solution containing radical cations. The antioxidant ability of compound-1 was determined by mixing different concentrations

Table 1 ¹HNMR Data (in CDCl₃, 600 MHz) and ¹³CNMR Data (in CDCl₃, 125 MHz) of Compound-1

Position	δ _H and Multiplicity (J in Hz)	δ _C (DEPT)	Position	δ _H and Multiplicity (J in Hz)	δ _C (DEPT)
1		172.3(C)	19	1.27, <i>m</i>	31.9(CH ₂)
2	2.33, <i>m</i>	34.2(CH ₂)	20	1.27, <i>m</i>	22.7(CH ₂)
3	1.64, <i>m</i>	24.8(CH ₂)	21	0.89, <i>m</i>	14.09(CH ₃)
4	1.27, <i>m</i>	31.9(CH ₂)	1'	4.32, <i>m</i>	62.1(CH ₂)
5	5.28, <i>m</i>	130.0(CH)		4.16, <i>m</i>	
6	5.28, <i>m</i>	129.4(CH)	2'	2.29, <i>m</i>	34.0(CH ₂)
7	1.27, <i>m</i>	31.8(CH ₂)	3'	5.3, <i>m</i>	68.9(CH)
8-14	1.27, <i>m</i>	29.7(CH ₂)	4'	1.27, <i>m</i>	37.1(CH ₂)
15	1.27, <i>m</i>	29.5(CH ₂)	5'	1.65, <i>m</i>	24.6(CH ₂)
16	1.27, <i>m</i>	29.3(CH ₂)	6'	1.27, <i>m</i>	27.3(CH ₂)
17	1.27, <i>m</i>	29.2(CH ₂)	7'	2.05, <i>m</i>	22.3(CH ₂)
18	1.27, <i>m</i>	29.1(CH ₂)	8'	0.86, <i>m</i>	14.1(CH ₃)

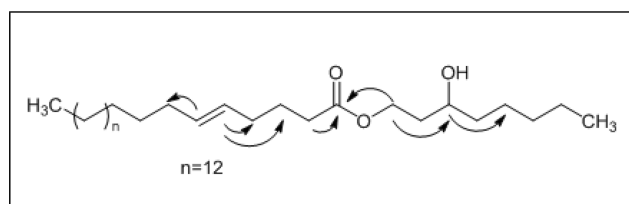


Figure 1 ^1H NMR of isolated compound, 3-hydroxyoctyl -5- *trans*-docosenoate.

(1000, 500, 250, 125, 62.5, 31 and 15.5 $\mu\text{g/mL}$) of compound-1 (300 μL) with ABTS solution (3.0 mL). The reduction in absorbance was measured using UV spectrophotometer after one minute of mixing the solutions and continued for six minutes. Ascorbic acid was used as positive control and percentage inhibition was calculated using the equation:

$$\% \text{ scavenging effect} = \frac{\text{control absorbance} - \text{sample absorbance}}{\text{control absorbance}} \times 100$$

Each experiment was repeated in triplicate. The antioxidant effect was expressed in terms of percent inhibition

and IC_{50} (concentration required for 50% reduction of ABTS radicals) was determined.

Molecular Docking

The three dimensional (3D) structure of the newly isolated compound was modeled by Molecular Operating Environment (MOE-2016) software.³⁰ The hydrogen atoms were added to compound-1 by 3D protonation followed by energy minimization by using MOE. The crystal structure of the tyrosine kinase was retrieved from the protein databank (PDB id: 1M17) [www.rcsb.org/pdb]. Prior to molecular docking, all water molecules were removed from the retrieved crystal structure using the Molecular Operating Environment software (www.chemcomp.com). The 3D protonation and energy minimization of the retrieved protein molecule was carried out by using MOE software with the default parameters. Compound-1 was docked into the active site of prepared tyrosine kinase using MOE by using the default parameters i.e., Placement: Triangle Matcher, Rescoring: London dG. For each ligand ten conformations were

C5_AZHAR_2017.1.fid — HNNMR/C5/CDCI3

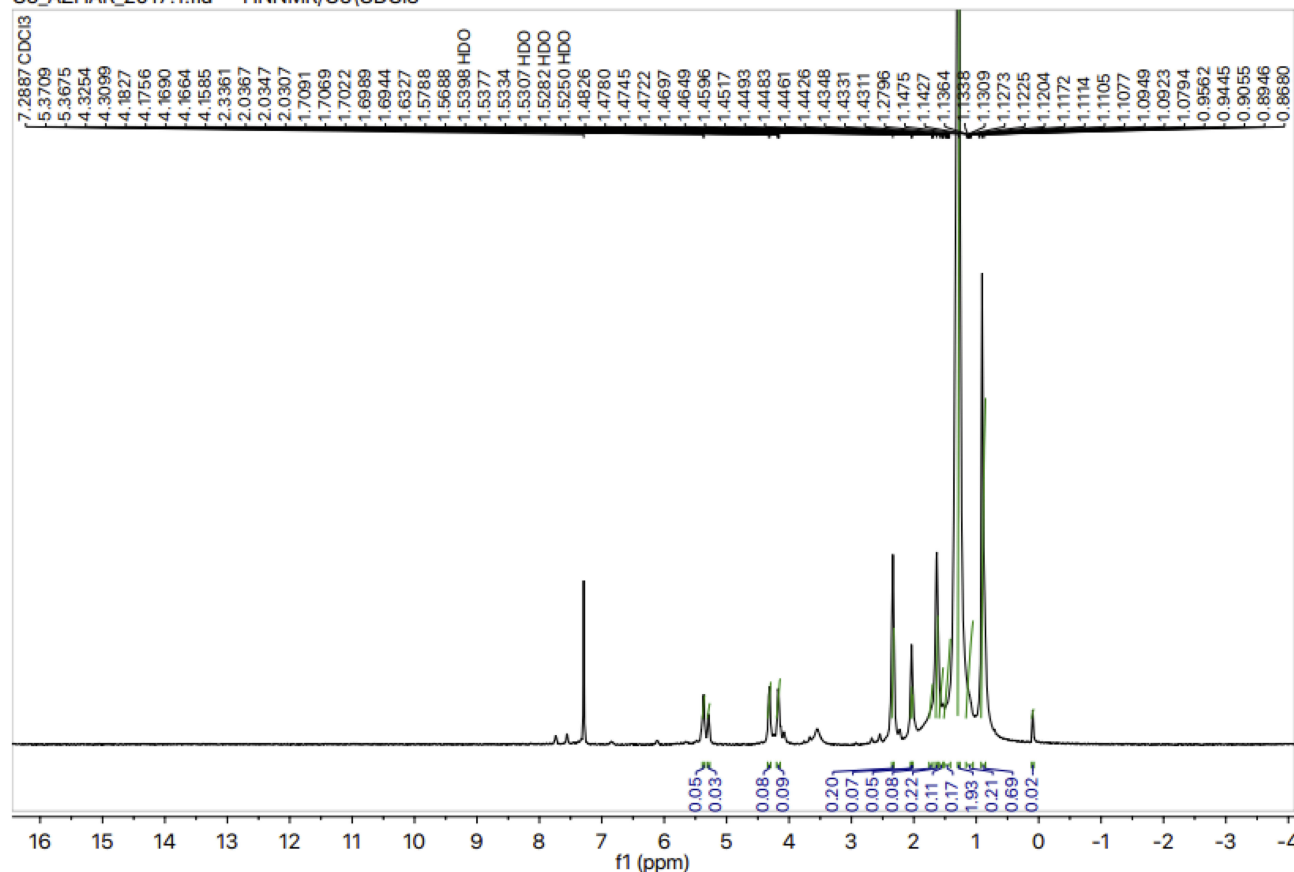


Figure 2 ^1H NMR of isolated compound, 3-hydroxyoctyl -5- *trans*-docosenoate.

C5_AZHAR_2017.1.fid —HNMR/C5\CDCl3

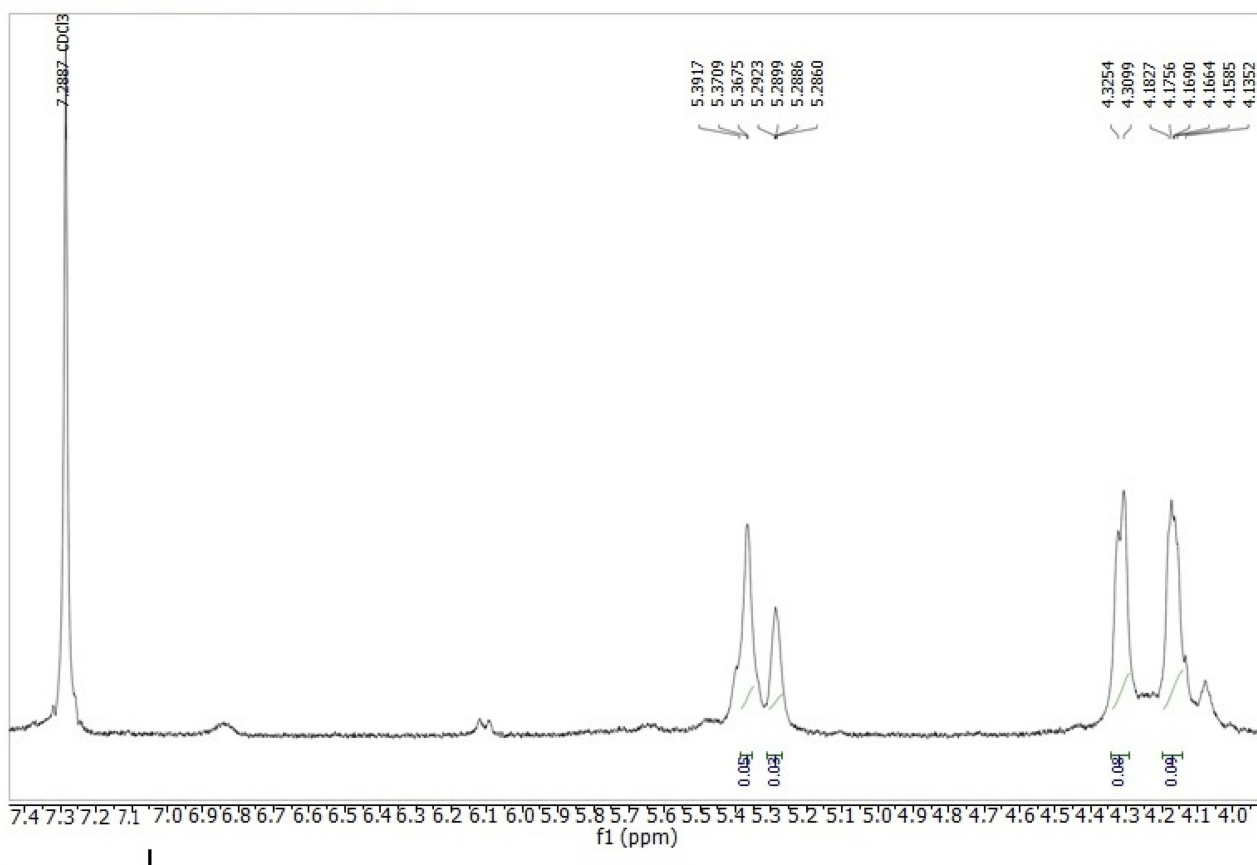


Figure 3 ^{13}C NMR of isolated compound, 3-hydroxyoctyl -5- *trans*-docosenoate.

generated. The top-ranked conformation of each compound was used for further analysis.

Results and Discussion

The current study revealed the isolation of a new aliphatic ester from the chloroform soluble fraction of *A. arvensis*. The newly isolated compound exhibited marked antioxidant and anticancer potential in in-vitro assay that was strongly supported by molecular docking studies.

Compound-1 was obtained as colorless amorphous powder and the structure of the compound was established by IR, EI, NMR spectroscopy including 2D NMR techniques and chemical methods. The molecular formula was determined based on HREIMS and ^{13}C NMR spectra. In HREIMS, the molecular ion peak was found at 466.7778 (calculated for $\text{C}_{30}\text{H}_{58}\text{O}_3$, 466.7785) suggesting the molecular formula $\text{C}_{30}\text{H}_{58}\text{O}_3$. ^1H NMR spectrum displayed a triplet-like peak at δ 0.89 and a broad singlet at δ 1.27 typical of a straight chain hydrocarbon. Other prominent peaks appeared at δ 5.37, 5.28, 4.32, 4.16

that could be assigned to the protons of oxymethine, olefinic bond, and oxymethylene group respectively based on their HSQC spectrum. The ^{13}C NMR spectrum (BB and DEPT) showed signals at δ 172.3, 130 and 129.4 that could be assigned to the carbonyl and olefinic carbons respectively (Table 1 and Figures 1–6). The oxygenated methine and oxygenated methylene resonated at δ 68.9 and 62.1 respectively. The signals observed between 29.7–29.1 revealed the presence of a long chain. The absorption band at 966 cm^{-1} in the IR spectrum showed the stereochemistry of the double to be *trans*. The ^{13}C NMR spectrum showed allylic carbon in the chain at δ 31.8 typical of methylene adjacent to a *trans* double bond.³¹ The position of the double bond was confirmed by oxidative cleavage with osmium tetroxide which produced heptadecanal along with other products. The position of the hydroxyl and double was also confirmed from HMBC correlations as shown in Figure 7. Thus, the structure of compound-1 was elucidated as 3-hydroxyoctyl -5- *trans*-docosenoate.

C5_AZHAR_2017.5.fid — deptNNMR/C5\CDCl3

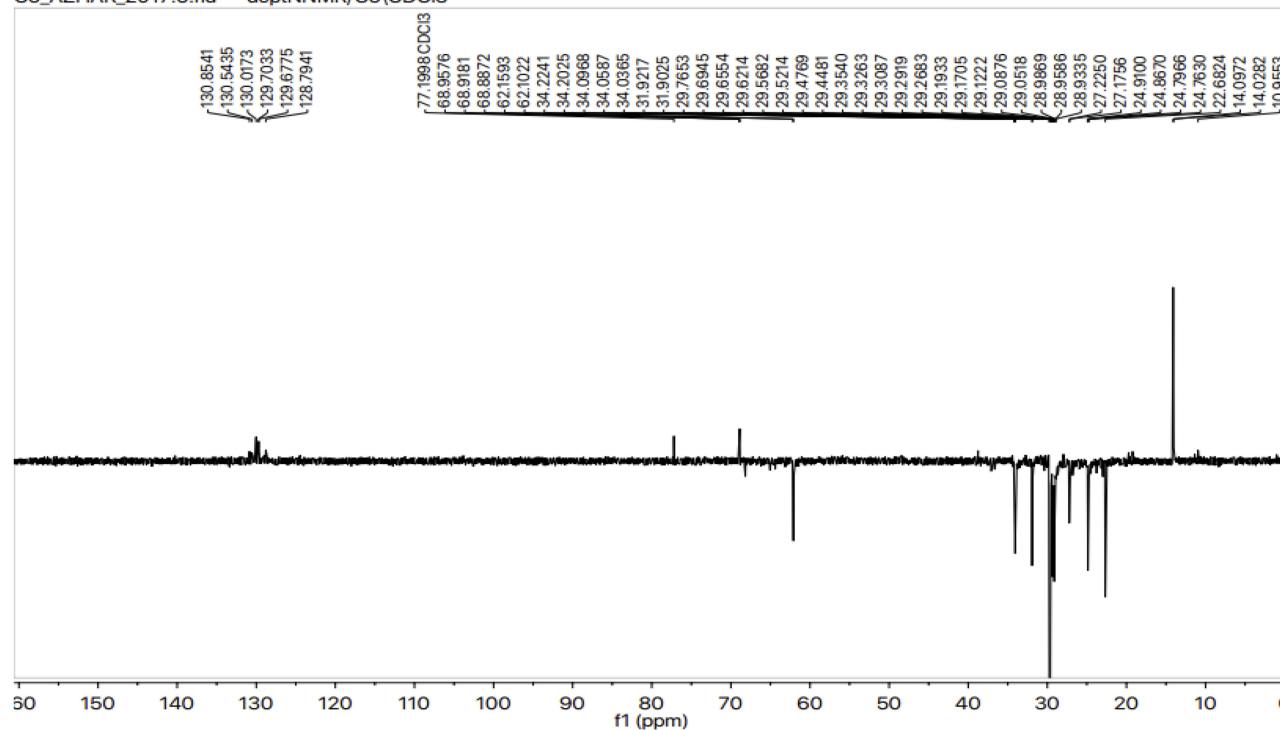


Figure 4 HMBC-NMR of isolated compound, 3-hydroxyoctyl -5- *trans*-docosenoate.

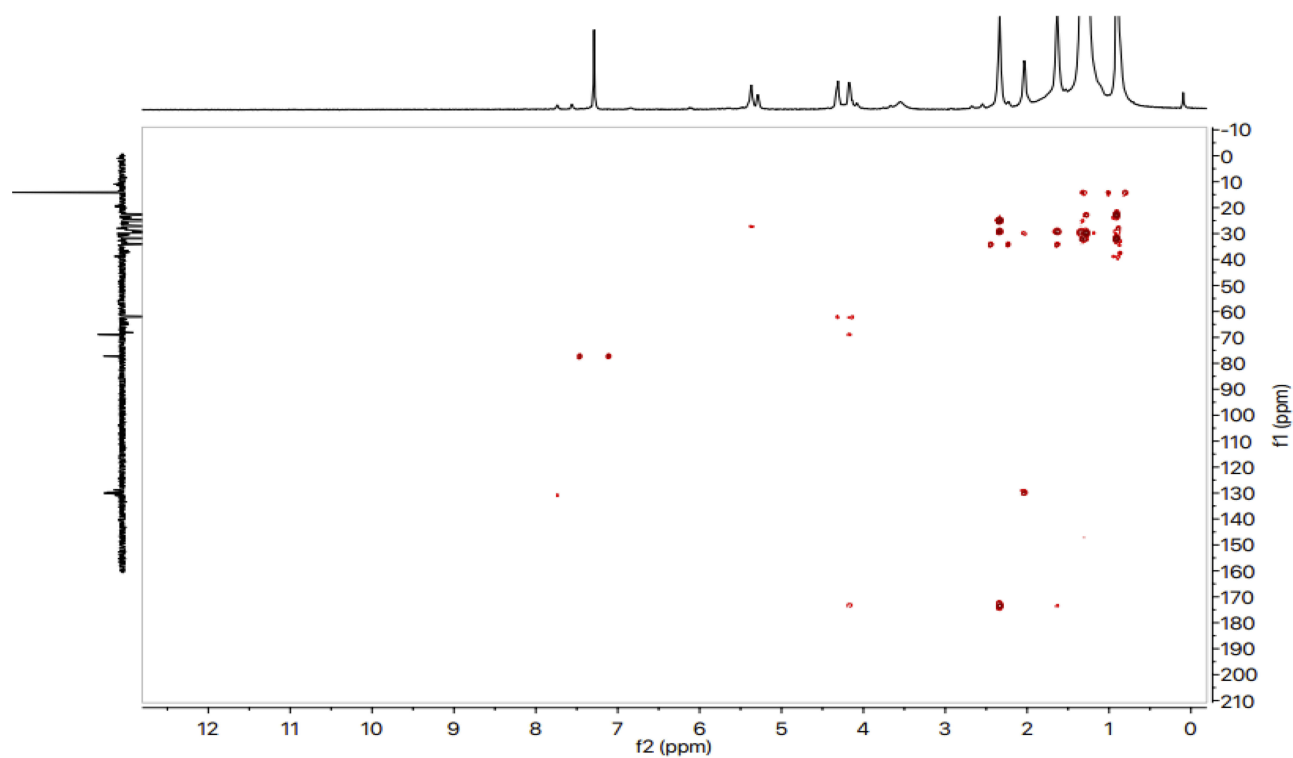


Figure 5 HSQC-NMR of isolated compound, 3-hydroxyoctyl -5- *trans*-docosenoate.

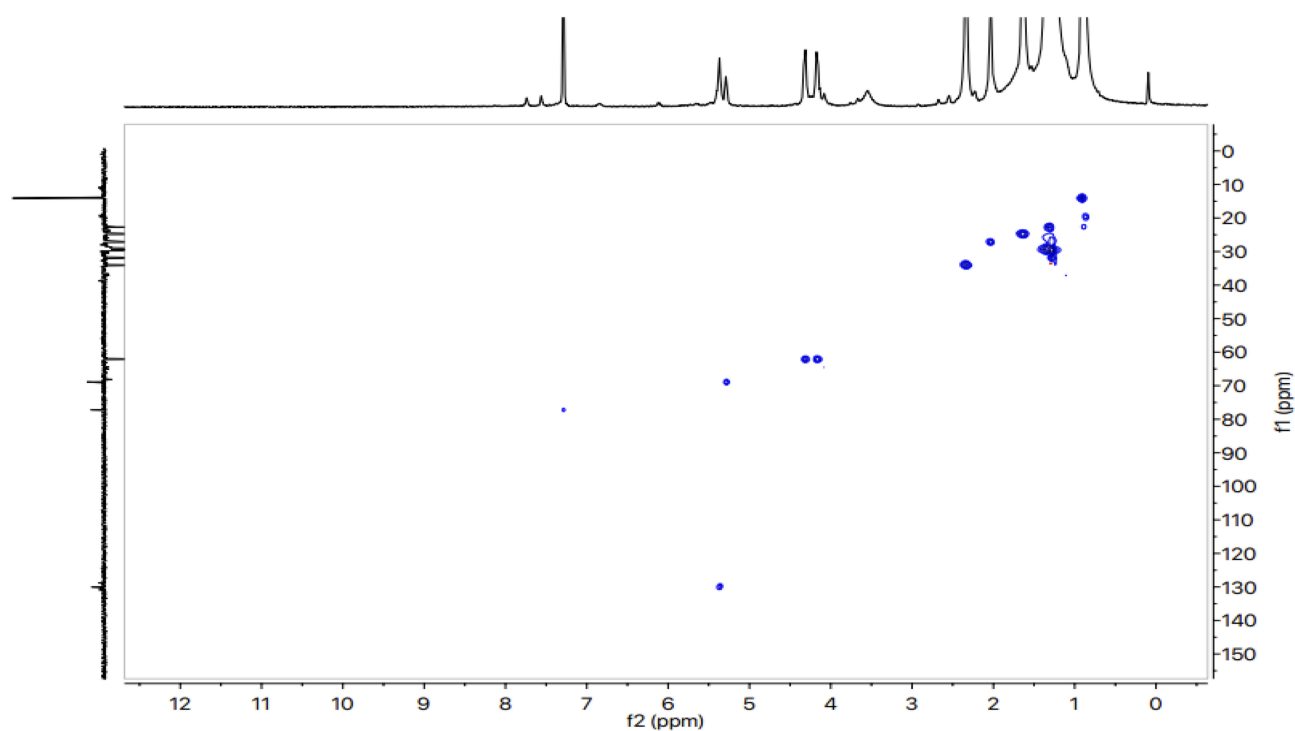


Figure 6 COSY-NMR of isolated compound, 3-hydroxyoctyl -5- *trans*-docosenoate.

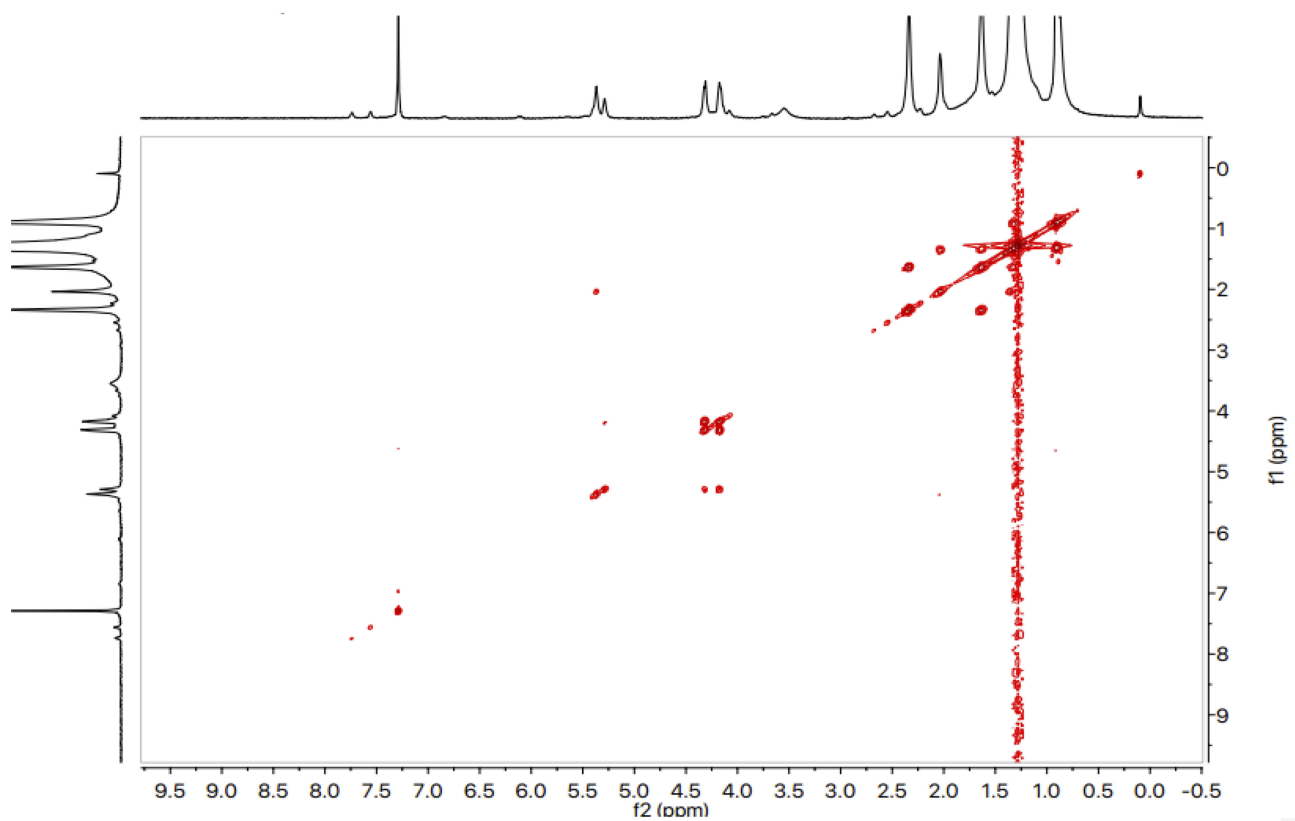


Figure 7 Chemical structure and HMBC correlations of isolated compound.

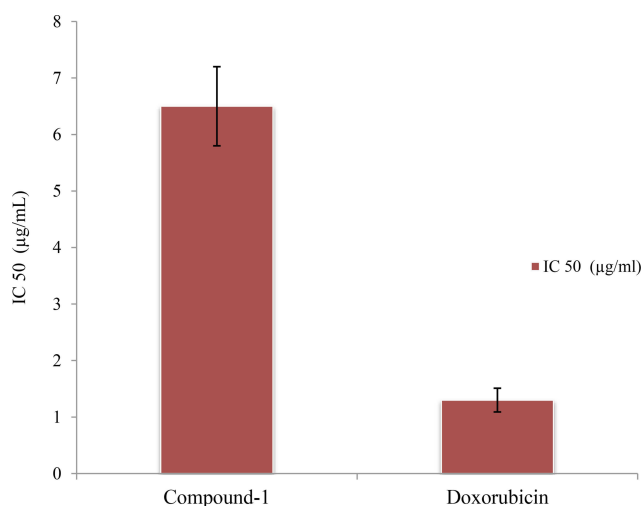


Figure 8 Cytotoxic effect of compound-1 against HepG2 cell line.

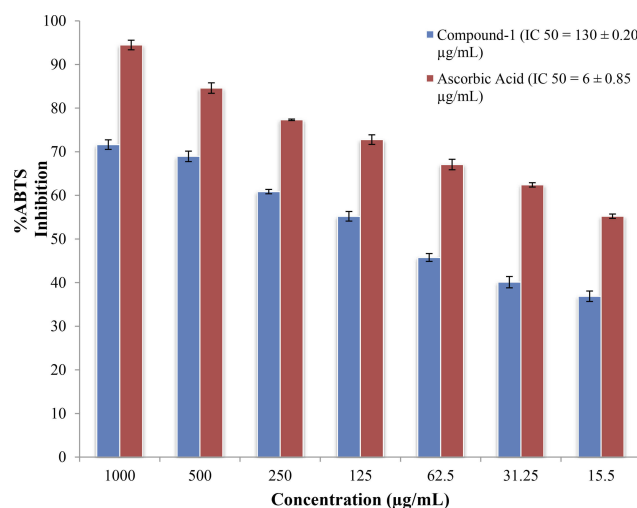


Figure 10 ABTS scavenging ability of compound-1.

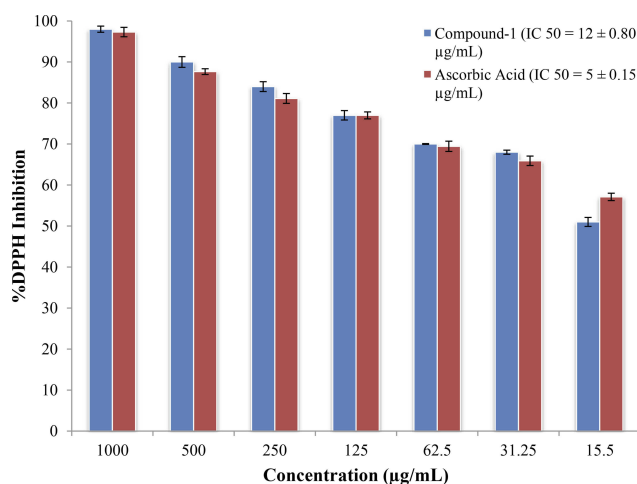


Figure 9 DPHH scavenging ability of compound-1.

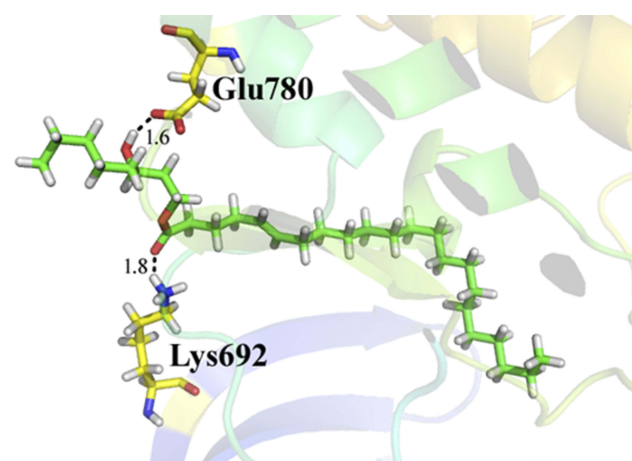


Figure 11 Lowest-energy docked pose of compound 1 with tyrosine kinase (PDB: 1M17) showing key interactions with Glu 780 and Lys 692 active residues of the potential drug target. Hydrogen-bonds are shown as black dashed lines.

Cytotoxicity of the compound isolated from *A. arvensis* was assessed against HepG2 cells by performing MTT assay (Figure 8). This assay is frequently used to investigate the cytotoxicity of test compounds.^{32,33} This study demonstrated that the isolated compound showed promising activity against HepG-2 cell lines with IC₅₀ value of 6.50 ± 0.70 µg/mL in comparison to doxorubicin, a positive cytotoxic drug (IC₅₀ value = 1.3 ± 0.21 µg/mL). DMSO was used as negative control. The compound did not show a cytotoxic effect against normal epithelial cell line (Vero). The other species belonging to the Boraginaceae family including *Arnebia euchroma* (Royle) Jonst,³⁴ *Alkanna*

cappadocica,³⁵ *Heliotropium indicum*,³⁶ *Heliotropium bacciferum*,³⁷ *Glandora rosmarinifolia*,³⁸ have already been reported for their anti-cancer activity.

The antioxidant activity was determined to evaluate the antioxidant potential of the compound. Results of DPHH and ABTS radical scavenging assays of compound-1 are given in Figures 9 and 10. The compound was effective in a concentration-dependant manner. The isolated compound exhibited significant DPHH scavenging ability with IC₅₀ value of 12 ± 0.80 µg/mL whereas ascorbic acid, used as positive control, exhibited antioxidant activity with IC₅₀ = 05 ± 0.15 µg/mL. The compound exhibited ABTS radical scavenging ability (IC₅₀ = 130 ± 0.20 µg/mL) compared with the value obtained

Table 2 Interaction of Ligand with the Active Site Residues

Ligand	Receptor	Interaction	Distance	E (Kcal/mol)
O 21	OE1 GLU 780	H-donor	1.6	-1.8
O 20	NZ LYS 692	H-acceptor	1.8	-4.8

for ascorbic acid ($06 \pm 0.85 \mu\text{g/mL}$). Oxygen is a vital element of life but under certain circumstances it can influence our health by formation of free radicals (reactive oxygen species) leading to some dangerous diseases, like coronary heart disease, diabetes, atherosclerosis, immune-suppression, and cancer.^{39,40} Consequently, agents with free radical scavenging abilities are useful for the prevention and therapy of several dangerous diseases including cancer.⁴¹

To assess the interactions of the newly isolated compound in the active site of tyrosine kinase, molecular docking, using Molecular Operating Environment (MOE-2016) software, was carried out. From the docking study, it was observed that the top ranked conformation of the isolated compound fit well and showed good interactions and docking score (-10.0745) with active site residues of tyrosine kinase. The compound showed two polar and several hydrophobic interactions with active site residues of the enzyme. The hydroxyl oxygen of compound-1 formed hydrogen bond with the carbonyl oxygen atom of active site residue Glu780. Whereas the carbonyl oxygen of the compound showed hydrogen bond with nitrogen atom of active site residue Lys692 (Figure 11). Both hydrogen bonds formed by compound-1 with active site residues were strong hydrogen bonds in terms of distance and energy (Table 2). As in both hydrogen bonds, the bonding atoms became very close to each other and that was the reason why the energy of both bonds was very small. Furthermore, there were hydrophobic interactions between compound-1 and active site residues of tyrosine kinase (Figure 11). This good bonding network shown by compound-1 might be one of the reasons why this compound showed good activity against this enzyme.⁴²

Conclusion

In conclusion, for the first time a new compound, 3-hydroxyoctyl -5- *trans*-docosenoate, was isolated from the chloroform soluble fraction of *A. arvensis*. Our test compound exhibited significant cytotoxicity against human hepatocellular carcinoma cells (HepG-2) with IC_{50} value of $6.50 \pm 0.70 \mu\text{g/mL}$. Also, data from the present results revealed that compound-1 is an antioxidant agent due to its DPHH and ABTS

radical scavenging. In docking studies against tyrosine kinase enzyme, our compound showed top ranked conformation of our compound fit well and showed good interactions and docking score (-10.0745) with active site residues of tyrosine kinase. Further studies are underway in our Laboratory.

Abbreviations

MTT, 3-(4,5-Dimethylthiazol-yl)-diphenyl tetrazolium bromide; *A. arvensis*, *Anchusa arvensis*; TKs, tyrosine kinases; DMSO, dimethyl sulfoxide.

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Author Contributions

SH, FU, WA, and RU performed experimental work and data collection. Data were evaluated by MA, AAS, and SAAS. Computational studies were performed by AW. MA, SH and SMS were involved in literature search and manuscript preparation. MA, SH and refined the manuscript for publication. AAS, RU, and FAN financially supported the research work. All authors contributed to data analysis, drafting and revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

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