

Comparison of oxygenation in peripheral muscle during submaximal aerobic exercise, in persons with COPD and healthy, matched-control persons

Krista G Austin¹
 Larry Mengelkoch²
 Jennifer Hansen¹
 Edward Shahady¹
 Prawee Sirithienthad¹
 Lynn Panton¹

¹Florida State University, Department of Nutrition, Food and Exercise Science, Tallahassee, FL, USA; ²Florida Agricultural and Mechanical University, Division of Physical Therapy, Tallahassee, FL, USA

Objective: The purpose of this study was to compare peripheral muscle oxygenation in persons with chronic obstructive pulmonary disease (COPD) to healthy control persons, during submaximal exercise.

Methods: Eight persons with COPD (forced expiratory volume in one second [FEV₁] = 1.00 ± 0.27 L) and eight healthy control persons (FEV₁ = 1.88 ± 0.55L) performed a submaximal graded exercise test (GXT), and completed 4 min of constant load exercise (CON) at 50% of peak GXT. Measurements included oxygen uptake, heart rate, arterial oxygen saturation and peripheral muscle oxygenation (%StO₂) at rest, during exercise, and recovery.

Results: Significantly greater workloads were attained for controls compared with COPD for peak GXT and CON. No significant differences in %StO₂ were observed between groups at: rest (GXT: 29.5 ± 22.8 vs 30.4 ± 17.3%; CON: 33.3 ± 15.4 vs 35.1 ± 17.2%); peak GXT (29.4 ± 19.4 vs 26.5 ± 15.9%); 4 min of CON (25.9 ± 13.5 vs 34.5 ± 21.8%); and recovery (GXT: 46.6 ± 29.1 vs 44.3 ± 21.7%; CON: 40.9 ± 21.5 vs 44.5 ± 23.2%).

Conclusion: These results suggest that peripheral skeletal muscle oxygenation is not compromised in COPD during submaximal exercise, and limitations in exercise capacity are most likely a result of muscle disuse and poor lung function.

Keywords: Skeletal muscle oxygenation, pulmonary disease, exercise capacity

Introduction

Chronic obstructive pulmonary disease (COPD) is characterized by multiple physiological problems such as impaired ventilation and gas exchange, cardiovascular and peripheral muscle deconditioning, muscular wasting and weakness, and dyspnea (Sala et al 1999; Simon et al 2001; Reusch 2002; Okamoto et al 2003; Saey et al 2003; Richardson et al 2004). Limitations in ventilation (including airflow obstruction, and lung hyperinflation leading to mechanical disadvantage of the ventilatory muscles) and gas exchange have been considered to be the primary causes of exercise intolerance in COPD (ATS and ERS 1999). However, research has recently focused on the limitations in skeletal muscle function that may further contribute to exercise intolerance (Richardson et al 2004). The primary focus of the research on skeletal muscle function in persons with COPD has attempted to determine whether the limitations in skeletal muscle function are a result of muscle dysfunction associated with COPD pathology, or muscle disuse (ATS and ERS 1999).

Studies examining the role of skeletal muscle in the pathophysiology of COPD have primarily compared individuals with COPD to control participants who are regularly active (Maltais et al 1998; Okamoto et al 2003), or have not utilized a

Correspondence: Krista G. Austin
 EIS/L'Boro Performance Centre,
 1st Floor, Loughborough University,
 Loughborough, Leicestershire LE11
 3TU, England
 Tel +44 087 0759 0496
 Fax +44 015 0922 6172
 Email krista.austin@eis2win.co.uk

control group for comparison purposes (Simon et al 2001; Saey et al 2003). Thus, it is difficult to assess the effects COPD is having on skeletal muscle function. Previous research by Simon and colleagues (2001) demonstrated that patients with COPD have limitations in blood flow directed to peripheral muscles and that oxygen extraction during exercise is limited. This was attributed to a redistribution of cardiac output and oxygen from lower limb exercising muscles to ventilatory muscles. Casaburi et al (1991), Maltias et al (1998), Okamoto et al (2003), and Saey et al (2003) have also provided evidence that skeletal muscle dysfunction may be responsible for exercise impairment through a variety of mechanisms including an early onset of lactic acidosis, contractile fatigue, and an impaired re-oxygenation of peripheral muscle. In contrast to these findings, Richardson and colleagues (2004) demonstrated that the muscle metabolic capacity of persons with COPD and healthy, age matched sedentary controls is similar, and suggested that muscular mechanical efficiency is reduced in persons with COPD and may be the primary factor limiting skeletal muscle function.

In summary, it is difficult to assess if the limitations in skeletal muscle function in persons with COPD results from muscle dysfunction associated with COPD pathology or muscle disuse because control subjects have not been closely matched to COPD subjects in previous studies. The aim of this study was to further investigate if the limitations in skeletal muscle function in persons with COPD results from muscle dysfunction associated with COPD pathology or muscle disuse. In the present study we compared peripheral muscle oxygenation, monitored by near infrared spectroscopy (NIRS), during incremental and constant load aerobic exercise, in persons with COPD against healthy control persons, who were matched by age, weight, and gender.

Methods

Participants

Eight persons with COPD and eight healthy age, weight, and gender matched-control persons completed this study. Although a quantitative assessment of activity levels was not done, all subjects were interviewed through personal communications about current activity levels, and we attempted to closely match subjects by their verbal description of their activity levels. Participants' activity levels were matched for type, duration and intensity of exercise performed, as well as activities of daily living that they completed on a day to day basis (ie, laundry, vacuuming,

grocery shopping). The diagnosis of COPD was based on current clinical evaluation by the participant's doctor and from pulmonary function testing during participation in the study. Participants were not suffering from any acute illness at the time of study and were free from any other medical conditions that would limit their ability to perform the exercise testing. Participants were all nonsmokers at the time of the study with the exception of one COPD participant. The participant was asked to abstain from smoking 24hrs prior to any testing and this was confirmed verbally with the investigator upon arrival to the laboratory. Written informed consent, which was approved by the human subjects committee at Florida Agricultural and Mechanical University, was obtained from each individual prior to participation.

Incremental exercise testing

Participants visited the laboratory on two separate occasions. During the first visit, participants performed a submaximal (~70% of estimated maximal heart rate), symptom-limited, graded exercise test (GXT) on an electronically braked cycle ergometer (SensorMedics, Ergoline 800S™, Yorba Linda, CA). Participants rested quietly for 5 min prior to the start of exercise so that baseline resting measurements could be obtained. The GXT utilized a ramp protocol, starting at 0 W for the 1st min and then increased by 7 W every minute thereafter for persons with COPD, and by 15 W for control participants. Participants were constantly monitored to maintain their cadence between 50–60 rpm. Exercise was terminated upon attainment of 70% estimated maximal heart rate and participants remained quietly seated for the next 5 min for the collection of post exercise measurements.

Constant load exercise testing

During a second visit to the lab, participants performed a 6 min steady state exercise test (constant load) at 50% of the workload obtained during the submaximal GXT. Prior to the start of the test, participants rested for 5 min for the determination of resting baseline measurements and then began the exercise test. At the start of exercise, participants warmed up for 1 min at 0 W. The workload then increased to 25% of the workload obtained during the submaximal GXT for 1 min. Following this 2 min warm-up period, the workload was increased to 50% of the workload obtained during the submaximal GXT and participants maintained this workload for 6 min. Participants were constantly monitored to maintain their cadence between 50–60 rpm.

Oxygen uptake, heart rate and dyspnea measures

Oxygen uptake (VO_2) was determined by breath-by-breath gas exchange analysis (SensorMedics, Vmax 229™ Metabolic Cart System, Yorba Linda, CA). Before each exercise test, the flow-volume sensor was calibrated with a 3-L calibration syringe and gas analyzers were calibrated using known gas concentrations. The metabolic data were averaged over the final 20 sec of each minute of collection. A 12-lead electrocardiogram (ECG) (Quniton, Q4500™, Bothall, WA) was utilized to continuously monitor heart rate (HR) and was supervised by a physician during all testing sessions. Dyspnea was subjectively rated during the last 10 seconds of each minute of exercise and recovery, using a modified Borg, 0 to 10 scale (Jones 1988).

Peripheral muscle oxygenation and arterial oxygenation

Peripheral muscle oxygenation (%StO₂) was continuously estimated, noninvasively, through NIRS during rest, exercise, and recovery periods (Inspectra Tissue Spectrometer Model 325 System, Hutchinson Technology Inc., Hutchinson, MN). The principles of NIRS and the reliability and validity of the measurement have been previously described in detail (Mancini et al 1994; Austin et al 2005). Briefly, the NIRS probe was placed on the right vastus lateralis muscle approximately 14–20 cm from the knee joint. The measurement site for each individual was marked with a permanent marker, to locate the same placement of the probe for both exercise tests. Before each exercise test, the NIRS probe was calibrated to known wavelengths equivalent to a StO₂ measurement of $36 \pm 2\%$ and $85 \pm 2\%$, according to the manufacturer's protocol and specifications. The NIRS unit was interfaced with a computer and %StO₂ data were averaged over the final 20 sec of each minute of collection.

Arterial oxygen saturation was continuously estimated (%SpO₂), during rest, exercise, and recovery periods with a pulse oximeter (SensorMedics, StatTrak™, Yorba Linda, CA) and attached to the subject by a finger probe. The pulse oximeter was electronically integrated to the SensorMedics, Vmax 229™ Metabolic Cart System. The %SpO₂ data were averaged over the final 20 sec of each minute of collection.

Pulmonary function testing

During the second visit and prior to exercise, participants completed a pulmonary function test (SensorMedics, Vmax

229™ Metabolic Cart System, Yorba Linda, CA) using standard spirometry procedures (ATS 1995) for the determination of forced vital capacity (FVC), and forced expiratory volume in one second (FEV₁). Before each pulmonary function test, the flow-volume sensor was calibrated with a 3-L calibration syringe. Reference equations from Morris and colleagues (1971) were used to derive the subject's predicted spirometric values.

Statistical analysis

Independent t-tests were utilized to compare differences between groups for measures of the participant physical characteristics, and for the measures of workload during GXT and constant load exercise. A repeated measures analyses of variance (ANOVA; group \times time) was utilized to compare differences between groups and across time for the measures of VO_2 , HR, %StO₂, %SaO₂, and dyspnea during the GXT, and the constant load exercise test. When interactions or main effects were determined to be significantly different, independent t-testing was used to determine which individual means were significantly different. Simple linear regression analysis was utilized to examine possible relationships between %StO₂ and VO_2 , %SaO₂, and measures of pulmonary function (FEV₁, FVC, and %FEV₁/FVC), during GXT and constant load exercise. A statistical significance of $p \leq 0.05$ was used for all analyses.

Results

Physical characteristics of participants

Table 1 presents the descriptive characteristics of the participants. Both groups were closely matched in gender, age, height, weight, and body mass index (BMI). Persons with COPD had significantly lower FEV₁, percent predicted FEV₁, and %FEV₁/FVC values than control participants. Persons with COPD were classified as having moderate to severe COPD, based on percent predicted values (Table 1) (ATS 1986).

Graded exercise testing

Significantly greater workloads were attained for control participants, compared with persons with COPD, at peak GXT ($73.8 \pm 36.2\text{W}$ vs $36.9 \pm 11.9\text{W}$). Table 2 presents data from the GXT. A significant interaction between groups and time was observed for VO_2 and HR. Control participants compared with persons with COPD, had significantly greater peak VO_2 values ($12.3 \pm 3.5 \text{ ml O}_2 \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$ vs $7.9 \pm 2.9 \text{ ml O}_2 \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$) and peak HR values ($122 \pm 11 \text{ beats} \cdot \text{min}^{-1}$

Table 1 Physical characteristics of participants (N = 16)

	Control (N = 8)	COPD (N = 8)
Gender (N Males [M]/ Females [F])	1 M/7 F	1 M/7 F
Age (yr)	67.8 ± 14.5	68.5 ± 11.5
Weight (kg)	75.1 ± 31.8	74.9 ± 31.3
Height (cm)	161.4 ± 5.1	162.1 ± 7.4
BMI (kg*(m ²) ⁻¹)	28.7 ± 12.0	28.2 ± 10.4
FEV ₁ (L)	1.88 ± 0.55*	1.00 ± 0.27
Predicted FEV ₁ %	91 ± 24*	51 ± 22
FVC (L)	2.65 ± 0.54	2.36 ± 0.81
FEV ₁ /FVC%	70 ± 12*	44 ± 12

Notes: Values (other than Gender) are mean ± standard deviation; *Significant difference between groups, control versus COPD, $p \leq 0.05$.

Abbreviations: COPD, chronic obstructive pulmonary disease; BMI, body mass index; FEV₁, forced expiratory volume in one second; FVC, forced vital capacity.

vs 110 ± 7 beats*min⁻¹). No significant differences between groups were found for measures of dyspnea.

No significant differences in %StO₂ were observed between groups (control vs COPD), at rest, peak exercise, or during recovery from exercise. A significant interaction between groups and time was observed for %SaO₂. Significantly higher %SaO₂ values for control participants, compared with persons with COPD, were found at rest ($96.8 \pm 1.0\%$ vs $93.5 \pm 1.7\%$) and during min 1 of recovery ($96.0 \pm 1.3\%$ vs $92.8 \pm 2.4\%$) from exercise.

Constant Load Exercise testing

The eight control participants completed 6 min of constant load exercise. Only six COPD participants completed 6 min

of constant load exercise. Seven COPD participants completed 5 min of constant load exercise and all eight COPD participants completed 4 min of constant load exercise. Significantly greater workloads were attained for control participants, compared with persons with COPD, during constant load exercise (36.9 ± 18.1 W vs 18.1 ± 6.5 W).

Table 3 presents data from the constant load exercise test. A significant interaction between groups and time was observed for VO₂ and HR. Control participants, compared with persons with COPD, had significantly greater VO₂ values at min 4 of constant load exercise (9.3 ± 3.6 ml O₂*kg⁻¹*min⁻¹ vs 6.1 ± 1.7 ml O₂*kg⁻¹*min⁻¹). Control participants, compared with persons with COPD, had significantly lower HR values at rest (67 ± 11 beats*min⁻¹ vs 86 ± 8 beats*min⁻¹), and during recovery from exercise at min 1 (77 ± 13 beats*min⁻¹ vs 93 ± 12 beats*min⁻¹) and min 5 (72 ± 12 beats*min⁻¹ vs 89 ± 11 beats*min⁻¹). No significant differences between groups were found for measures of dyspnea.

No significant differences in %StO₂ were observed between groups (control vs COPD), at rest, constant load exercise, or during recovery from exercise. A significant interaction between groups and time was observed for %SaO₂. Significantly higher %SaO₂ values for control participants, compared with persons with COPD, were found at rest ($96.3 \pm 1.3\%$ vs $93.6 \pm 1.5\%$), at min 4 of constant load exercise ($95.8 \pm 2.2\%$ vs $92.9 \pm 2.2\%$), and during recovery from exercise at min 1 ($95.9 \pm 2.0\%$ vs $93.4 \pm 2.6\%$) and min 5 ($96.8 \pm 0.7\%$ vs $94.9 \pm 1.0\%$).

Table 2 Graded exercise test (GXT) (N = 16)

	Rest	Peak GXT	min 1 Recovery	min 5 Recovery
%StO₂				
Control	29.5 ± 22.8	29.4 ± 19.4	48.0 ± 28.9	46.6 ± 29.1
COPD	30.4 ± 17.3	26.5 ± 15.9	42.6 ± 19.4	44.3 ± 21.7
%SaO₂*				
Control	96.8 ± 1.0 ^δ	94.3 ± 4.0	96.0 ± 1.3 ^δ	96.3 ± 1.3
COPD	93.5 ± 1.7	91.8 ± 2.8	92.8 ± 2.4	95.1 ± 1.1
VO₂ (ml O₂*kg⁻¹*min⁻¹)*				
Control	2.7 ± 0.8	12.3 ± 3.5 ^δ	5.3 ± 1.5	2.5 ± 0.4
COPD	2.9 ± 1.0	7.9 ± 2.9	5.8 ± 1.8	2.5 ± 0.8
HR (beats*min⁻¹)*				
Control	76 ± 14	122 ± 11 ^δ	92 ± 21	79 ± 14
COPD	81 ± 10	110 ± 7	97 ± 13	89 ± 10
Dyspnea (0–10)				
Control	–	2.4 ± 1.3	0.8 ± 1.2	0.1 ± 0.4
COPD	–	2.6 ± 0.9	2.0 ± 1.9	0.3 ± 0.4

Notes: Values are mean ± standard deviation; *Significant interaction between groups and time, $p \leq 0.05$; ^δSignificant difference between groups, control versus COPD, $p \leq 0.05$; Control participants at all time points, N = 8; COPD participants at all time points, N = 8.

Abbreviations: COPD, chronic obstructive pulmonary disease; HR, heart rate; min, minute; %SaO₂, arterial oxygen saturation; %StO₂, peripheral muscle oxygenation; VO₂, oxygen uptake.

Table 3 Constant load exercise

	Rest	min 4 Constant load exercise	min 5 Constant load exercise	min 6 Constant load exercise	min 1 Recovery	min 5 Recovery
%StO₂						
Control	33.3 ± 15.4	25.9 ± 13.5	28.9 ± 13.9	28.1 ± 12.8	39.3 ± 18.4	40.9 ± 21.5
COPD	35.1 ± 17.2	34.5 ± 21.8	35.0 ± 25.5	32.8 ± 27.3	45.4 ± 22.5	44.5 ± 23.2
%SaO₂*						
Control	96.3 ± 1.3 ^δ	95.8 ± 2.2 ^δ	95.4 ± 3.5	95.4 ± 3.4	95.9 ± 2.0 ^δ	96.8 ± 0.7 ^δ
COPD	93.6 ± 1.5	92.9 ± 2.2	93.3 ± 2.4	93.2 ± 2.8	93.4 ± 2.6	94.9 ± 1.0
VO₂ (ml O₂*kg⁻¹*min⁻¹)*						
Control	2.9 ± 1.0	9.3 ± 3.6 ^δ	9.6 ± 3.8	9.2 ± 3.2	5.0 ± 1.5	2.6 ± 1.1
COPD	2.6 ± 0.7	6.1 ± 1.7	6.9 ± 1.8	6.4 ± 1.8	4.5 ± 1.2	2.4 ± 0.4
HR (beats*min⁻¹)*						
Control	67 ± 11 ^δ	97 ± 12	97 ± 12	98 ± 11	77 ± 13 ^δ	72 ± 12 ^δ
COPD	86 ± 8	104 ± 11	102 ± 12	100 ± 10	93 ± 12	89 ± 11
Dyspnea (0–10)						
Control	–	1.9 ± 1.2	2.0 ± 1.2	1.9 ± 1.1	0.5 ± 0.8	0.1 ± 0.2
COPD	–	1.7 ± 1.1	1.6 ± 1.1	1.9 ± 1.3	1.0 ± 1.6	0.0 ± 0.0

Notes: Values are mean ± standard deviation; *Significant interaction between groups and time, $p \leq 0.05$; ^δSignificant difference between groups, control versus COPD, $p \leq 0.05$; Control participants at all time points, N = 9; COPD participants N = 8, at Rest, min 4 Constant Load Exercise, min 1 and min 5 Recovery from exercise; COPD participants N = 7, at min 5 Constant Load Exercise; COPD participants N = 6, at min 6 Constant Load Exercise.

Abbreviations: COPD, chronic obstructive pulmonary disease; HR, heart rate; min, minute; %SaO₂, arterial oxygen saturation; %StO₂, peripheral muscle oxygenation; VO₂, oxygen uptake.

Relationships between %StO₂ and VO₂, %SaO₂ and measures of pulmonary function

Table 4 shows correlation coefficients (r values) between %StO₂ and VO₂, %SaO₂ and pulmonary function measures (FEV₁, FVC, and %FEV₁/FVC) during incremental and constant load exercise, in all subjects, at various time points. The only significant correlations observed were between %StO₂ and VO₂, at rest, during both GXT ($r = 0.60$) and constant load exercise ($r = 0.62$).

Responses of %StO₂ during exercise

Differences in the exercise and post-exercise %StO₂ measures that were demonstrated in both COPD and control participants during the GXT and CON tests can be found in Figures 1 and 2. In most participants (5 COPD, 5 control), no significant change in %StO₂ was found from rest to peak exercise load during the incremental exercise test or during the constant load exercise test (Figure 2). A significant change in %StO₂ was based on the known standard error of the equipment during cycle exercise and placing a 95% confidence interval around the resting %StO₂ value; thus participants had to achieve a greater than 8% change in StO₂ values. In contrast,

six participants (3 COPD, 3 control) demonstrated a steady decline in %StO₂ (Figure 1). The post exercise %StO₂ response was characterized by a rapid re-oxygenation in these 6 participants (3 COPD, 3 controls). However, the other participants (5 COPD, 5 controls) demonstrated almost no change or an increase above resting values in %StO₂ following the incremental and constant load exercise tests.

Discussion

Prior research examining the causes of exercise intolerance in persons with COPD attributed exercise cessation primarily to dyspnea and limitations in ventilation and gas exchange that limit O₂ delivery to the exercising muscle (Maltais et al 1998; ATS and ERS 1999; Richardson et al 2004). However, it has been demonstrated that patients with similar FEV₁ values have a wide range of exercise capacity (Jones and Killian 1991) and that cardiorespiratory fitness is partially dependent on peripheral muscle function (Maltais et al 1998). Previous findings have also suggested that peripheral muscle weakness in persons with COPD is a result of functional changes that have lead to skeletal muscle dysfunction (ATS and ERS 1999). However, these same observed changes are also found in deconditioned muscle and suggest that disuse rather than pathological dysfunction is what causes changes

Table 4 Correlation coefficients (r values) between peripheral muscle oxygenation (%StO₂) and oxygen uptake (VO₂), arterial oxygenation (%SaO₂) and pulmonary function measures (forced expiratory volume in one second [FEV₁], forced vital capacity [FVC], and %FEV₁/FVC) during incremental (GXT) and constant load exercise, in all subjects (N = 16)

GXT	%StO ₂ – Rest	%StO ₂ – Peak exercise	%StO ₂ – min 1 Recovery	%StO ₂ – min 5 Recovery
VO ₂ (at same time points)	0.60*	0.18	0.23	0.16
%SaO ₂ (at same time points)	0.12	0.37	0.07	0.07
FEV ₁	0.27	0.24	0.07	0.03
FVC	0.32	0.45	0.13	0.15
%FEV ₁ /FVC	0.10	0.04	0.04	0.09
Constant load exercise	%StO₂ – Rest	%StO₂ – min 4 Constant load exercise	%StO₂ – min 1 Recovery	%StO₂ – min 5 Recovery
VO ₂ (at same time points)	0.62*	0.20	0.48	0.31
%SaO ₂ (at same time points)	0.11	0.15	0.08	0.03
FEV ₁	0.21	0.39	0.27	0.15
FVC	0.19	0.46	0.20	0.11
%FEV ₁ /FVC	0.15	0.14	0.19	0.13

Notes: *Significant difference between groups, control versus COPD, $p \leq 0.05$.

Abbreviations: COPD, chronic obstructive pulmonary disease.

in muscle structure and function (Booth and Gollnick 1983; Larson and Ansved 1985).

In the present study we attempted to match our participants for age, gender, BMI, and activity levels. Nonetheless, control participants had greater lung function and higher exercise capacities than the COPD participants as was evidenced by the pulmonary function data, VO₂ data,

Muscle deoxygenation response during exercise

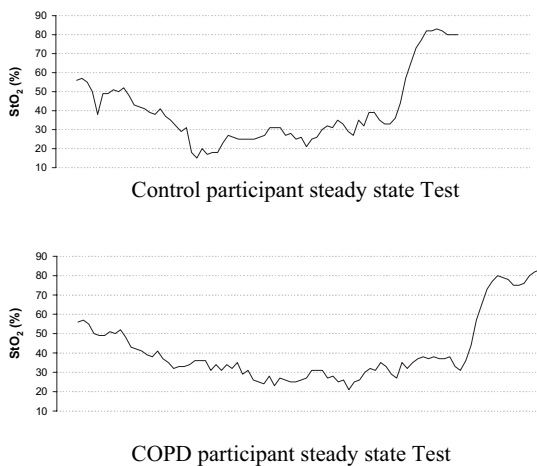


Figure 1 Comparison of deoxygenation in COPD and control participants.
Abbreviations: COPD, chronic obstructive pulmonary disease; StO₂, peripheral muscle oxygenation.

exercise workloads, and the inability of 2 COPD participants to complete 6 min of constant load exercise. Cessation of exercise for the two participants that completed only 4 min and 5 min of exercise, respectively, at their constant workload, was because of muscular fatigue. However, it is interesting to note that during both exercise tests, that there were no significant differences in dyspnea ratings between COPD and control participants. Since the Borg rating scale used in this study is a validated assessment tool for breathlessness (Killian 1985), we believe the results reflect that the exercise intensities were relatively low. Thus, our findings are in agreement with previous studies (Maltais et al 1998; ATS and ERS 1999; Richardson et al 2004) suggesting that exercise intolerance in COPD may be primarily attributed to impaired oxygen delivery associated with COPD pathology causing limitations in ventilation and gas exchange. Furthermore, in the present study we observed that control participants and persons with COPD had similar levels of %StO₂ during exercise. This suggests that COPD pathology does not cause muscle dysfunction in terms of impaired muscle oxygenation in persons with moderate COPD.

The findings of our study are in agreement with Richardson and colleagues (2004) who employed direct measurements of oxygen utilization and blood flow in evaluating metabolic capacity in persons with COPD. Despite the finding of a greater percentage of Type II fibers, structure

No muscle deoxygenation response during exercise

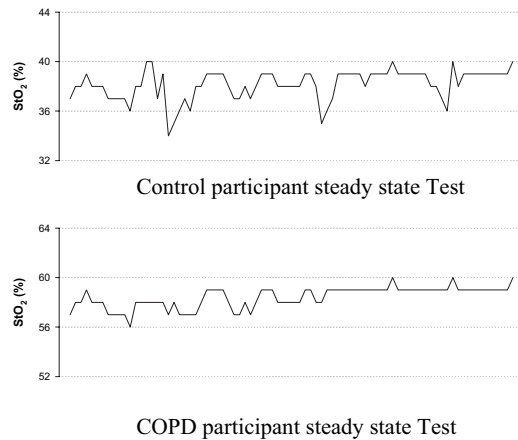


Figure 2 Comparison of no deoxygenation in COPD and control participants. **Abbreviations:** COPD, chronic obstructive pulmonary disease; StO₂, peripheral muscle oxygenation.

in terms of muscle fiber cross sectional area, capillary density and mitochondrial volume were not different between COPD and controls. Thus, Richardson concluded that limitations in exercise capacity were a result of mechanical inefficiency (ie, work performed per unit of oxygen consumed) due to an increase in Type II muscle fibers, and that persons with COPD do not demonstrate a diminishment in muscle O₂ utilization due to dysfunction. While we did not perform assessment of muscle type and structure in the present study, the nonsignificant differences observed between the groups in measures of %StO₂ provide evidence to support the theory that muscle disuse leading to inefficiency rather than muscle dysfunction may further be reducing exercise capacity in persons with COPD and sedentary, healthy adults.

Others have also reported that muscle O₂ delivery is maintained in those with COPD, however impaired O₂ utilization was reported and muscle dysfunction suggested (Maltais et al 1998; Simon et al 2001). Maltais and colleagues have shown that peripheral O₂ delivery is preserved in persons with COPD and that the significantly greater skeletal muscle acidosis observed in these individuals at peak O₂ uptake was potentially a cause for the early cessation of exercise; thus reflecting an altered metabolic function of the skeletal muscle and indicating dysfunction.

Several other explanations for the reported disparity between supply of muscle oxygen and utilization have been proposed; however all have concluded that this indicates muscle dysfunction rather than detraining. Saey and colleagues have suggested that exercise tolerance in COPD

during constant load cycling is dependent on contractile leg fatigue, and that differences in fatigue existed between individuals. When participants were divided based on fatigability during the exercise test, it was found that despite improving FEV₁ through the use of a bronchodilator, that peripheral muscle fatigue limited exercise tolerance in those who demonstrated early fatigue. Unfortunately a control group was not utilized for comparison purposes.

In a study examining limitations in lower limb oxygen consumption of persons with COPD, Simon and colleagues (2001) reported that exercise capacity was significantly greater in participants who did not demonstrate a plateau in leg oxygen consumption, than in those participants who did plateau in leg oxygen consumption despite a continued increase in total body oxygen uptake. This was attributed to a redistribution of cardiac output O₂ from the lower extremities to pulmonary musculature in the plateau group. The rationale provided for redistribution was in part attributed to a limitation in O₂ extraction by peripheral exercising muscle. The results of Simon et al (2001) and Saey et al (2001) indicate that despite ventilatory limitation, some individuals with COPD are additionally limited during whole body cycling exercise by the ability of the peripheral musculature to utilize oxygen.

An additional study by Okamoto and colleagues (2003) which also utilized NIRS to assess peripheral skeletal muscle oxygen utilization, reported a significantly greater time for re-oxygenation following submaximal exercise in persons with COPD when compared with controls. It was additionally found in persons with COPD that a significant positive relationship existed between the time VO₂ remained elevated during exercise and duration needed for complete re-oxygenation of peripheral skeletal muscle. This relationship was not found in the present study. In addition, VO₂ and %StO₂ were not significantly different between the groups at any time point of recovery during the GXT or constant load exercise test; however, %SaO₂ was significantly lower in COPD than controls during recovery. The discrepancy between our study and that of Okamoto and colleagues (2003) might be attributed to differences in the control groups that were utilized for comparison purposes.

Several differences in the exercise and post-exercise %StO₂ measures were observed (Figures 1 and 2) for the COPD and control participants of the present study. Some subjects demonstrated a desaturation response during both types of exercise. As noted above, we believe that the exercise workloads in this study were likely of relatively low intensity,

as indicated by similarities in dyspnea ratings between groups. Thus, it is possible that the intensity was not high enough to stimulate a desaturation response. However, this same phenomena (lack of tissue desaturation) has been observed in well trained triathletes, cyclists, and runners during high intensity exercise (ie, exercise tests to lactate threshold and maximal tests to exhaustion) (Snyder and Parmenter 2002; Snyder et al 2003; Austin et al 2005). Thus, we believe that the workload intensities were sufficient for the study purpose, and this lack of tissue desaturation response, in some individuals, does occur in healthy subjects, athletes, and persons with COPD at various exercise intensities. What other possibilities might explain this lack of tissue desaturation response during exercise? vanBeekvelt and colleagues (2001) have suggested that NIRS measurements may be affected by adipose tissue thickness. These researchers reported muscle oxygen consumption, as measured by NIRS, is negatively correlated ($r = -0.70$, $p < 0.01$) to adipose tissue thickness, indicating that adipose tissue is a confounding factor in obtaining valid NIRS measurements. In the present study we did not measure skinfold thickness, thus it is possible that % StO₂ readings were skewed because of an excess in adipose tissue at the measurement site. However, we found that when we compared BMI values with the %StO₂ response, those with normal BMIs (18.5–24.9) had just as frequent a variation in the %StO₂ response as did those with overweight or obese BMIs (≥ 25.0) (Expert Panel 1998). Austin and colleagues (2005) have also reported the same type of varied response for %StO₂ in well trained athletes who did not appear to have large amounts of body adiposity at the measurement sites; thus, something in addition to the thickness of adipose tissue appears to influence the results obtained by NIRS and warrants further investigation.

Impaired muscle oxygen utilization has also been reported in persons with insulin resistance, hypertension, cardiovascular disease (CVD), diabetes, and peripheral arterial disease (PAD) (Reusch 2002; Bauer et al 2004). Prior literature has reported that COPD causes vasoconstriction in the pulmonary vasculature, resulting in pulmonary hypertension (Hida et al 2002). Additionally, in COPD there is evidence of an impaired endothelial cell release of the vasodilator and nitric oxide in the pulmonary vasculature due to atherosclerosis (Hida et al 2002). This is also reported for persons with cardiovascular disease and PAD in the coronary and peripheral vasculature due to atherosclerosis (Reusch 2002). While none of the participants had known

PAD, CVD, or diabetes, it may be hypothesized that individuals demonstrating impaired muscle oxygen use and without the post-exercise vasodilatory response may additionally be suffering from a lack in vasodilatory capacity due to impairments of endothelial cell release of nitric oxide.

Limitations of the present study

The major limitation of the present study is the small sample size which may have prevented us from determining any significant differences in %StO₂. We had a large and varied response in %StO₂ within the COPD and control groups, which limits the power of the data. This may be a result of the varied activity levels of the COPD and control participants, and should be further examined through a larger population of exercising and nonexercising persons for both of these populations. Additionally, several of our participants were taking medications for both hypertension and hyperlipidemia, which may contribute to decreases in nitric oxide, and thus may have further led to the variance in %StO₂ responses that were seen in the present study. Further work is needed to assess the impact of such medications on peripheral muscle oxygenation, along with the effects which could be solely attributable to hypertension and hyperlipidemia. Future research that examines the role of peripheral oxygenation in persons with COPD should include the assessment of blood flow, hemoglobin concentrations, and muscle fiber type to further elucidate what may cause such a varied response in %StO₂ values.

Summary

Even though we attempted to match our participants for age, gender, BMI, and activity levels, our results indicate that the control participants had higher exercise capacities and greater lung function than the COPD participants as was evidenced by the VO₂ data, exercise workloads, and the inability of 2 COPD participants to complete 6 min of constant load exercise. Control participants and persons with COPD had similar levels of %StO₂ during exercise in our study. This suggests that COPD pathology does not cause muscle dysfunction in terms of impaired muscle oxygenation in persons with moderate COPD. It is concluded that peripheral skeletal muscle oxygenation is not compromised in individuals with moderate COPD during submaximal aerobic exercise, and that limitations in exercise capacity are most likely a result of muscle disuse and poor lung function.

References

- [ATS] American Thoracic Society. 1986. Evaluation of impairment/disability secondary to respiratory disorders. *Am Rev Respir Dis*, 133:1205–9.
- [ATS] American Thoracic Society. 1995. Standardization of spirometry: 1994 update. *Am J Respir Crit Care Med*, 152:1107–36.
- [ATS and ERS] American Thoracic Society and European Respiratory Society. 1999. Skeletal muscle dysfunction in chronic obstructive pulmonary disease: a statement of the American Thoracic Society and European Respiratory Society. *Am J Respir Crit Care Med*, 159:S1–S40.
- Austin KG, Daigle K, Cowman J, et al. 2005. Reliability of near-infrared spectroscopy for determination of muscle oxygen saturation during exercise. *Res Q Exerc Sport*, 76:440–9.
- Bauer T, Brass E, Hiatt W. 2004. Impaired muscle oxygen use at onset of exercise in peripheral arterial disease. *J Vasc Surg*, 40:488–93.
- Booth FW, Gollnick PD. 1983. Effects of disuse on the structure and function of skeletal muscle. *Med Sci Sport Exerc*, 15:415–20.
- Casaburi R, Patessio J, Ioli F, et al. 1991. Reduction in exercise lactic acidosis and ventilation as a result of exercise training in obstructive lung disease. *Am Rev Respir Dis*, 143:9–18.
- Expert Panel. 1998. Executive summary of the clinical guidelines on the identification, evaluation, and treatment of overweight and obesity in adults. *Arch Intern Med*, 158:1855–67.
- Hida W, Tun Y, Kikuchi Y, et al. 2002. Pulmonary hypertension in patients with chronic obstructive pulmonary disease: recent advances in pathophysiology and management. *Respirology*, 7:3–13.
- Jones NL, Killian KJ. 1991. Limitation of exercise in chronic airway obstruction. In: Chermiack NS (ed). *Chronic obstructive pulmonary disease*, 1st ed. Philadelphia: WB Saunders, pp 196–206.
- Jones NL. 1988. *Clinical exercise testing*. 3rd ed. Philadelphia: WB Saunders, p 77.
- Killian KJ. 1985. The objective measurement of breathlessness. *Chest*, 88:885–905.
- Larsson L, Ansved T. 1985. Effects of long term physical training and detraining on enzyme histochemical and functional skeletal muscle characteristics in man. *Muscle Nerve*, 8:714–22.
- Maltais F, Jobin J, Sullivan MJ, et al. 1998. Metabolic and hemodynamic responses of lower limb during exercise in patients with COPD. *J App Physiol*, 84:1573–80.
- Mancini D, Bolinger L, Li H, et al. 1994. Validation of near-infrared spectroscopy in humans. *J App Physiol*, 77:2740–7.
- Morris JF, Koski A, Johnson LC. 1971. Spirometric standards for healthy nonsmoking adults. *Am Rev Respir Dis*, 103:57–67.
- Okamoto T, Kanazawa H, Hirata K, et al. 2003. Evaluation of oxygen uptake kinetics and oxygen kinetics of peripheral skeletal muscle during recovery from exercise in patients with chronic obstructive pulmonary disease. *Clin Physiol Funct Imaging*, 23:257–62.
- Reusch J. 2002. Current concepts in insulin resistance, type 2 diabetes mellitus, and the metabolic syndrome. *Am J Cardiol*, 90(suppl): 19G–26G.
- Richardson R, Leek B, Gavin T, et al. 2004. Reduced mechanical efficiency in chronic obstructive pulmonary disease but normal peak $\dot{V}O_2$ with small muscle mass exercise. *Am J Respir Crit Care Med*, 169:89–96.
- Saey D, Deblgare R, LeBlanc P, et al. 2003. Contractile leg fatigue after cycle exercise: a factor limiting exercise in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*, 168:425–30.
- Sala E, Roca J, Marrades R, et al. 1999. Effect of endurance training on skeletal muscle bioenergetics in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*, 159:1726–34.
- Simon M, LeBlanc P, Jobin J, et al. 2001. Limitation of lower limb $\dot{V}O_2$ during cycling exercise in COPD patients. *J App Physiol*, 90:1013–19.
- Snyder AC, Dorman JC, Parmenter MA. 2003. Muscle oxygen saturation measurements track changes due to training in ironman triathletes. *Med Sci Sport Exerc*, 35(Suppl 1):S37.
- Snyder AC, Parmenter MA. 2002. Use of muscle oxygen saturation in determining maximal steady state exercise intensity. *Med Sci Sport Exerc*, 34(Suppl 1):S78.
- vanBeekvelt M, Borghuis M, vanEngelen B et al. 2001. Adipose tissue thickness affects in vivo quantitative near-IR spectroscopy in human skeletal muscle. *Clin Sci*, 101:21–8.

