

Guanidinylated bioresponsive poly(amido amine)s designed for intranuclear gene delivery

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Abstract: Guanidinylated poly(amido amine)s with multiple disulfide linkages (Gua-SS-PAA)s were designed and constructed as nonviral gene carriers. The main chains of these novel carriers were synthesized based on monomers containing guanidino groups (guanidine hydrochloride and chlorhexidine), which could avoid complicated side-chain-modification reactions while introducing the guanidino groups. The synthesized Gua-SS-PAA)s polymers were characterized by ¹H nuclear magnetic resonance, molecular weight, and polydispersity. Furthermore, Gua-SS-PAA)s polymers were complexed with pDNA, and the properties of the complexes were determined, including entrapment efficiency, particle size, ζ-potential, atomic force microscopy images, stability, DNA complexation ability, reduction sensitivity, cytotoxicity, and transfection efficiency. The new Gua-SS-PAA)s carriers exhibited higher transfection efficiency and lower cytotoxicity compared with two widely used gene delivery carriers, polyethylenimine and lipofectamine 2000. Furthermore, the relationship between the side-chain structure and morphological/biological properties was extrapolated, and the results showed that guanidine in the side chain aids in the improvement of transfection efficiency. In addition, the introduction of guanidino group might confer the new carriers with nuclear localization function compared to carriers without it.

Keywords: gene carrier, transfection efficiency, cytotoxicity, nuclear localization, guanidine

Introduction

The main challenge in gene therapy is the design of a suitable carrier for safe and efficient delivery of therapeutic genes.¹ Carriers for gene delivery can be divided into two classes: viral and nonviral. Viral carriers are modified viruses that can deliver exogenous genes to the cells. Although viral carriers exhibit high transfection efficiency, they possess some defects that are difficult to overcome, such as low gene loading capacity, difficulty in production and quality control, and, especially, immune responses leading to serious drug safety issues.² In contrast, nonviral carriers have high gene loading capacity with no or lower host immunogenicity. At the same time, nonviral carriers can be obtained through chemical synthesis with controllable quality, although they normally possess lower transfection efficiency compared to viral carriers.³

Nonviral carriers are mainly defined as various polymers that can connect to or complex with exogenous genes, deliver them to cytoplasm, and release the exogenous gene fragments through the reactions in cell membranes or organelles mediated by the complexes.⁴ Currently, the two most popular nonviral gene delivery systems have been widely used in laboratory and clinical studies: 1) using electroporation to transfect cells,^{5–7} and 2) cationic polymers, such as cationic liposomes and polyethylenimines (PEIs).⁸ Poly(amido amine)s (PAAs) with multiple disulfide linkages (SS-PAA)s are the most promising alternatives for liposomes and PEIs.⁹ The multi-amino structure

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confers the complexes with good cell membrane permeability and facilitates them to escape from subcellular structure degradation (such as endosomes and lysosomes) due to the amino proton sponge effect.¹⁰ The multiple disulfide linkages in the main chains lead to reduction in sensitivity due to the reductive environment in cytoplasm including lysosomes and endosomes, resulting in the rapid release of gene fragments in the intracellular environment.¹¹ In addition, both quality control and industrial production of SS-PAA nonviral carriers are easy to achieve because of the concise synthetic routes and mild reaction conditions.

Lin et al¹² reported that, compared to linear polymers, branched SS-PAA show higher transfection efficiency and lower cytotoxicity. Their studies further showed that increased transfection efficiency and decreased cytotoxicity were related to the buffering capacity and charge density in the polymer side chains. Martello et al¹³ synthesized a series of SS-PAA using various amino-containing monomers and disulfide-containing cross-linking agents and investigated the relationship between different end groups of SS-PAA and in vitro transfection efficiency. Zhang et al¹¹ found that the number of disulfide bonds in the main chains of SS-PAA had an impact on their transfection efficiency. Merdan et al¹⁵ and Lin et al¹⁶ reported that PEGylated SS-PAA enhanced the colloidal stability, reduced the interactions with blood components, and decreased the cytotoxicity of the complexes compared with SS-PAA. However, PEGylated SS-PAA also showed lower transfection efficiency than SS-PAA. Salmaso et al¹⁷ and Li et al¹⁴ investigated the effect of cell membrane thiols and reduction-triggered disassembly on the transfection activity of bioreducible polyplexes, which revealed that the plasma membrane protein thiols played a key role in the observed enhancement of DNA transfection.

Guanidine is one of the strongest organic bases with some biological activities.¹⁸ Nitrogen and hydrogen atoms in the guanidino group have a strong affinity to carbonic acid esters, phosphate esters, and peptides, leading to the formation of hydrogen bonds with certain geometric configurations. The amino moiety of guanidine has the ability to react with various acids in the human body to form water-soluble salts.¹⁹ As a result, a drug containing guanidino groups could be easily transported with selectivity in vivo absorption and/or permeation. With good permeability and low toxicity, guanidine and the polymer containing guanidino groups have been widely used as bacteriostatic agents, antibacterial finishing agents, and food additives in pharmaceutical settings, and also in textile industries.¹⁷ Compared to cationic

liposomes, the guanidinylation of PEIs and poly(β -amino ester)s showed superior transfection efficiency and reduced cytotoxicity, as reported by Nimesh and Chandra²⁰ and Kim et al.²¹ Kim et al²² further showed that the improved transfection efficiency of guanidylated gene carriers was due to, 1) the good cell membrane permeability mediated by the charged nature of guanidino groups, and 2) the peptidomimetic nuclear localization effect of guanidylated polymeric gene carriers.

To better address these issues in the search for nonviral gene carriers, guanidylated SS-PAA (Gua-SS-PAA) gene carriers were designed and constructed in our laboratory. The main chains of the novel carriers were constructed based on monomers containing guanidino groups, which were quite different from the existing carriers without guanidino groups. This approach could avoid complicated side-chain modification reactions while introducing the guanidino groups.²³ These newly synthesized Gua-SS-PAA showed high transfection efficiency and low cytotoxicity, suggesting that these polymers could serve as potential delivery carriers in gene therapy.

Materials and methods

Materials

Branched PEI (water free) with a molecular weight of 25 kDa, L-glutathione reduced (GSH), and thiazolyl blue tetrazolium bromide (MTT) were purchased from Sigma-Aldrich Co. (St Louis, MO, USA). Lipofectamine 2000 (Lipo) reagent, Dulbecco's Modified Eagle's Medium, fetal bovine serum (FBS), penicillin, streptomycin, and trypsin were purchased from Thermo Fisher Scientific (Waltham, MA, USA). *N,N'*-Cystamine bisacrylamide (CBA) was purchased from Alfa Aesar (Ward Hill, MA, USA). TIANpure Midi Plasmid Kit was obtained from TIANGEN Biotech (Beijing) Co., Ltd. (Beijing, People's Republic of China). Agarose was purchased from Thermo Fisher Scientific. Hoechst 33342 and 3,3'-diocetadecyloxycarbocyanine perchlorate (DiO) were purchased from Beyotime Institute of Biotechnology (Shanghai, People's Republic of China). *pEGFP-N3* vector strain was kindly provided by the Department of Pharmacology, China Medical University (CLONTECH Laboratories, Palo Alto, CA, USA). Lyso-ID Red detection kit and TOTAL NUCLEAR-ID Green/Red nucleolar/nuclear detection kits were purchased from Enzo Life Sciences (East Farmingdale, NY, USA). The 2,2,4,6,7-Pentamethyldihydrobenzofuran-5-sulfonyl chloride (Pbf-Cl), guanidine hydrochloride (CAR), and chlorhexidine (CHL) were purchased from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, People's Republic

of China). Acetone, *N,N*-dimethylformamide, and other common reagents were of analytical grade.

Synthesis of Gua-SS-PAA

Gua-SS-PAA were synthesized by the following three reactions: 1) CAR or CHL acetate (10.46/1.60 mmol, 1.00/1.00 g) was dissolved in water (20 mL) and added to a three-necked flask. After the solution was cooled to 0°C–3°C, a Pbf-Cl/acetone solution (10 mL, 10.46/6.58 mmol, 3.02/1.90 g) was added dropwise to the flask. After the solution addition was completed, the reaction mixture was stirred continuously at room temperature for 3 hours. Throughout the course of the reaction, the pH of the system was maintained at 11–12 with an NaOH solution (4 M). Then, the resulting white precipitate was collected through filtration and washed three times with water; 2) Gua-SS-PAA-Pbf-Cl was synthesized by Michael addition between CBA and CAR/CHL–Pbf-Cl according to a previous report.²⁴ After oven-drying overnight, the white precipitate (CAR–Pbf-Cl/CHL–Pbf-Cl, 0.76 mmol, and 0.24/0.60 g) and CBA (0.38 mmol, 0.1 g) were added to a brown flask and dissolved with *N,N*-dimethylformamide (5 mL). The reaction solution was then placed in a temperature-controlled water bath at 60°C under nitrogen atmosphere and was stirred continuously in the dark for at least 6 days. Subsequently, 10% excess CAR/CHL–Pbf-Cl was added to consume any unreacted acrylamide groups, and the reaction was allowed to continue for another 2 days; 3) Ten milliliters of trifluoroacetic acid–triisopropylsilane–water was added to the flask with the volume ratio of 95:2.5:2.5 and then stirred at room temperature for 3 hours.²⁵ After that, the solution was diluted with water to 30 mL and then dialyzed against distilled water (MWCO: 1,000) for 2 days to remove the unreacted monomers. Finally, the solution was lyophilized for 36 hours to obtain the product. The synthesis route is shown in Figure 1.

Polymer characterization

¹H nuclear magnetic resonance (NMR) spectra of the prepared Gua-SS-PAA were recorded using a Varian Unity 600 MHz NMR spectrometer (Bruker Corporation, Billerica, MA, USA) with deuterated dimethyl sulfoxide (DMSO).

Molecular weights and polydispersity of the Gua-SS-PAA were measured by gel permeation chromatography with PEG as standard using a Waters 2690D HPLC system equipped with Ultrahydrogel 120 and 250 columns (Waters Corporation, Milford, MA, USA). An HAc–NaAc buffer solution (0.25 M, pH 4.5) was used as the eluent at a flow rate of 0.7 mL/min.

Preparation of Gua-SS-PAA–pEGFP nanocomplexes

The plasmid DNA (*pEGFP-N3*) was extracted from the *pEGFP-N3* vector strain using the TIANpure Midi plasmid extraction kit and diluted to a final concentration of 20 µg/mL in HEPES buffer solution (HBS; 50 mM HEPES, 750 mM NaCl, pH 7.4). The polymer complexes (Lipo, PEI, and Gua-SS-PAA–*pEGFP*) at varying w/w ratios ranging from 1/3 to 1/96 were prepared by adding a certain amount of the polymer (10 µg/µL in HBS) to 1 mL plasmid solution (20 µg/mL in HBS buffer pH 7.4), followed by vortexing for 10 seconds, and the solution was incubated for 20 minutes at room temperature prior to use.

Determination of encapsulation efficiency through Hoechst 33342 intercalation

The *pDNA* (0.5 µg, *pEGFP-N3*) condensations with Lipo, PEI, and Gua-SS-PAA (CAR/CHL–CBA) at different nucleic acid/polymer (N/P, weight ratio) ratios from 1/0.5 to 1/96 were determined by Hoechst 33342 intercalation assay. The *pDNA* was mixed with 100 µL Hoechst 33342 (0.10 µg/mL) and then incubated with different polymers for 20 minutes at room temperature. The fluorescence of free DNA was used as control. The fluorescence of all samples was measured by a microplate reader (SpectraMax M3; Molecular Devices LLC, Sunnyvale, CA, USA). The encapsulation efficiency (EE) was determined by the following equation:²⁶

$$EE (\%) = \frac{\text{Fluorescence}_{\text{control}} - \text{Fluorescence}_{\text{sample}}}{\text{Fluorescence}_{\text{control}}} \times 100 \quad (1)$$

Measurements of particle size and ζ-potential

The size and ζ-potential of the complexes formed by *pDNA* and PEI or Gua-SS-PAA (CBA–CAR/CBA–CHL) at varying N/P ratios from 1/6 to 1/96 were measured at 25°C on a Zetasizer Nano system (Malvern Instruments, Malvern, UK) using the dynamic light scattering (DLS) technique. The results were processed using the Dispersion Technology Software V7.01 (Malvern Instruments).

Atomic force microscopy analysis of complex particle formation

Samples were prepared on freshly cleaved mica using a mixed solution of *pDNA* (final concentration: 20 µg/mL) and different polymers (final concentration: 0.48 mg/mL),

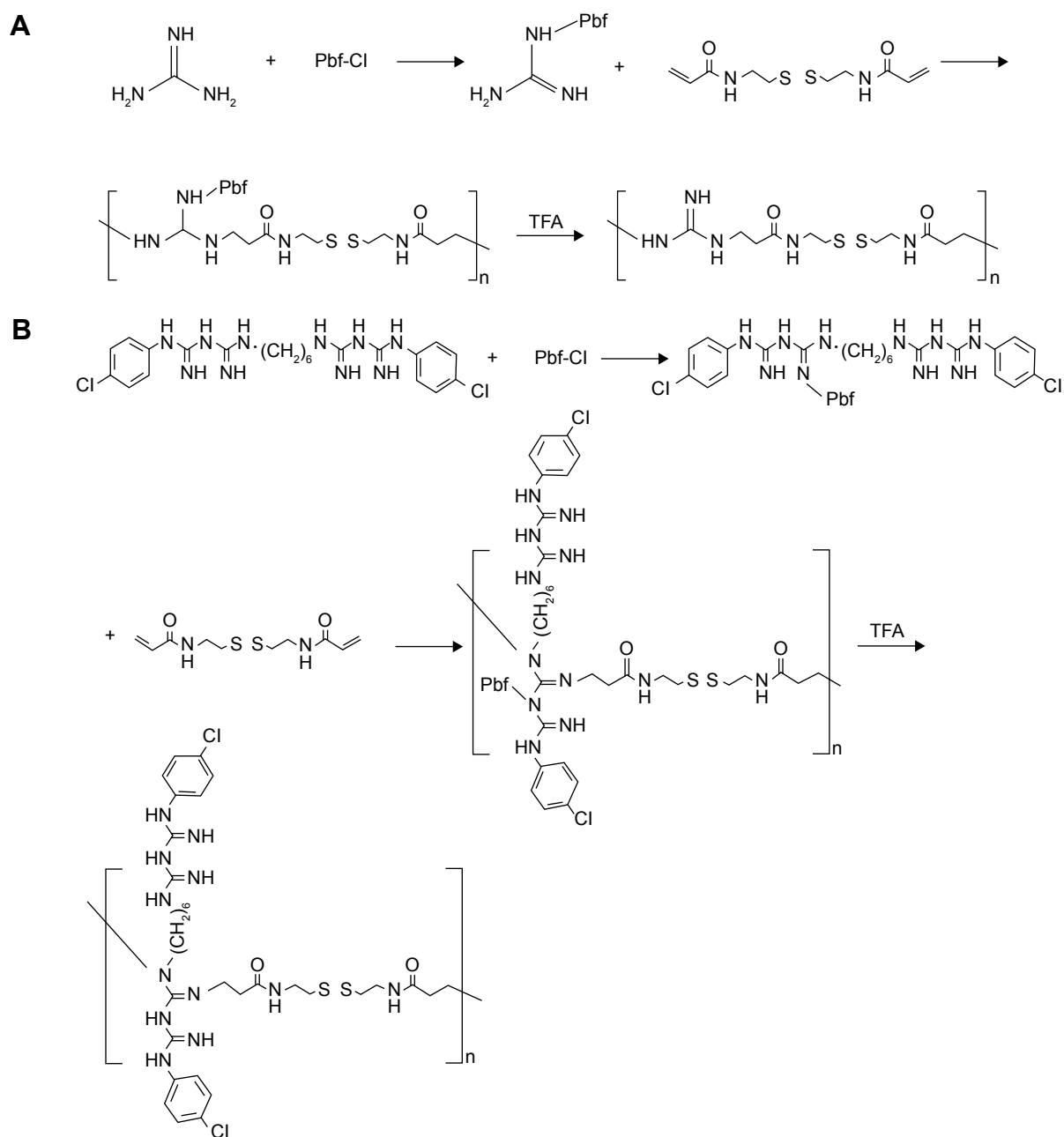


Figure 1 Schematic synthesis routes of CAR-CBA (A) and CHL-CBA (B).

Abbreviations: CAR, guanidine hydrochloride; CHL, chlorhexidine; CBA, *N,N'*-cystamine bisacrylamide; TFA, trifluoroacetic acid.

resulting in complexes with an N/P weight ratio of 1/48 in HBS. In addition, the Lipo and branched PEI were used at concentrations recommended by their respective providers' instructions. A total volume of 1 μ L of the sample was coated on the mica surface for measuring the images of complexes in the buffer solution. Complex formation was studied using a Picoforce Multimode Nanoscope IIIa (Veeco Instruments, Inc., Town of Oyster Bay, NY, USA) in tapping mode. Tapping mode imaging was performed in situ in HBS using a standard liquid cell and a standard Veeco NP cantilever. Similarly, in order to study different complexes in the solid

state, nitrogen was used to blow-dry the samples after they were prepared on the mica surface. Atomic force microscopy (AFM) imaging of the dry samples was then carried out in the same way as described earlier.

Agarose gel electrophoresis assay

Agarose gel retardation assay

The polymer complexes (Lipo, PEI, or Gua-SS-PAAs-*pEGFP*) at varying N/P ratios from 1/3 to 1/96 were prepared by adding a certain amount of the polymer (10 μ g/ μ L in HBS) to 100 μ L plasmid solution (20 μ g/mL in HBS buffer pH 7.4),

followed by vortexing for 10 seconds and incubating the solution for 20 minutes at room temperature. After the addition of 2 μ L of 6 \times loading buffer to 10 μ L of plasmid solution, Lipo-*pEGFP*, PEI-*pEGFP*, or CBA-CAR/CBA-CHL-*pEGFP*, 12 μ L of this mixture was loaded onto a 1% agarose gel containing 0.5 μ g/mL GoodviewTM and run at 100 V for 30 minutes in TAE buffer (40 mM, tris-acetate and 1 mM EDTA). After development of the gel, DNA was visualized with a UV lamp using a Biospectrum AC system W/cooled Gel Camera P/N 97-0154-05 (Upland, CA, USA).

Reduction sensitivity assay

The polymer complexes at varying N/P ratios from 1/6 to 1/48 were prepared by the same method as described in the “Agarose gel retardation assay” section. After the preparation, GSH was added to 10 μ L of the plasmid solution (200 ng of *pEGFP*), including Lipo-*pEGFP*, PEI-*pEGFP*, and CBA-CAR/CBA-CHL-*pEGFP*, at a final concentration of 20 mM. The dispersions were then incubated for 20 minutes. The same polymer complex solutions were also prepared without GSH as negative controls. After mixing with 2 μ L of 6 \times loading buffer, 12 μ L of this mixture was loaded onto a 1% agarose gel containing 0.5 μ g/mL GoodviewTM, and electrophoresis was carried out. The electrophoresis conditions were the same as described in the “Agarose gel retardation assay” section.

Heparin replacement assay

The polymer complexes at an N/P ratio of 1/9 were prepared using the same method as described in the “Agarose gel retardation assay” section. Heparin (10 U/ μ L) was diluted to different concentrations and added to 10 μ L of plasmid solution (200 ng of *pEGFP*), including Lipo-*pEGFP*, PEI-*pEGFP*, and CBA-CAR/CBA-CHL-*pEGFP*. The dispersions with different contents of heparin were then incubated for 30 minutes at 37°C. After the addition of 2 μ L of 6 \times loading buffer, 12 μ L of this mixture was loaded onto a 1% agarose gel containing 0.5 μ g/mL GoodviewTM, and electrophoresis was carried out. The electrophoresis conditions were the same as described in the “Agarose gel retardation assay” section.

DNase I degradation assay

Polymer complexes at different N/P ratios (from 1/0.5 to 1/16) were prepared by the same method as described in the “Agarose gel retardation assay” section and incubated with 10 μ L DNase I digestion system (DNase I 10 U, 50 mM KCl, 10 mM Tris-HCl, 10 mM MgCl₂, 0.1% Triton X-100,

pH 9.0) at 37°C for 20 minutes. The mixtures were then incubated with 5 μ L termination solution (400 mM NaCl, 100 mM EDTA, pH 8.0) for 5 minutes at room temperature. The DNA integrity was assessed by agarose gel electrophoresis after replacement with heparin solution (4 U/ μ L). The electrophoresis conditions were the same as described in the “Agarose gel retardation assay” section.

Cell viability assay

MCF-7 (human breast cancer cells) were maintained in RPMI 1640 growth medium supplemented with 10% FBS and penicillin (100 U/mL)-streptomycin (100 μ g/mL) at 37°C in a humidified atmosphere containing 5% CO₂. MCF-7 cells were seeded in 96-well plates (4 \times 10³ cells/well) and cultured overnight. Different N/P weight ratios, ranging from 1/6 to 1/96, were used to prepare the complexes (PEI-*pEGFP* and CBA-CAR/CBA-CHL-*pEGFP*). Briefly, polymers and *pDNA* were dissolved in the serum-containing culture medium. Complexes were prepared by adding 200 μ L of polymers at concentrations of 60–960 μ g/mL to 50 μ L of *pDNA* at a concentration of 40 μ g/mL, followed by vortexing for 10 seconds and incubation at room temperature for 20 minutes. The MCF-7 cells in each well were incubated with the desired amount of complexes for 12 hours at the same cell culture conditions as described earlier, and then the complex solution in each well was removed. After that, 20 μ L MTT (5 mg/mL) solution was added to each well and incubated for 4 hours. The medium was removed, and 150 μ L DMSO was added. The absorbance was measured at 570 nm using a microplate reader (Spectra Max M3; Molecular Devices LLC), and the absorbance of untreated cells (with cell media only) was taken as 100% cell viability.²⁷ The relative cell viability was calculated as follows: cell viability (%) = (OD_{570 (samples)}/OD_{570 (control)}) \times 100, where OD_{570 (control)} was obtained with the medium alone and OD_{570 (samples)} was obtained in the presence of polymers.

In vitro transfection efficiency assay

Transfection experiments were also performed with MCF-7 cells using the plasmid *pEGFP-N3*. The cell culture and complex preparation were carried out by the same methods as described in the “Cell viability assay” section. Two parallel transfection series – one for the qualitative analysis of transfection efficiency using fluorescence microscopy (FM) and the other for the quantitative analysis using flow cytometry (FCM) – were established. MCF-7 cells were seeded in six-well plates at a density of 1.0 \times 10⁵ cells/well. The cells were cultured in 2 mL RPMI 1640 complete culture

medium until the cell confluence reached ~60%. They were then washed with fresh 1× PBS buffer and incubated with 1.5 mL complete medium containing complexes with N/P ratios from 1/6 to 1/96 (the preparation of complexes was the same as described in the “Cell viability assay” section) for 4 hours at 37°C in a humidified atmosphere containing 5% CO₂. The medium was then replaced with 2 mL of fresh RPMI 1640 complete medium, and the cells were incubated for another 44 hours. Cells in all wells of the two parallel series were collected, and one series of cells was collected for FCM assay by determining the transfection efficiency using a FACScan flow cytometer (Becton, Dickinson and Company, Franklin Lakes, NJ, USA), with an excitation wavelength of 488 nm. The cells in other series were observed under a Nikon TE2000U inverted fluorescence microscope (Nikon Corporation, Tokyo, Japan) with an excitation wavelength of 488 nm and a magnification of 200×. The untreated cells were used as a negative control, and Lipo-*pEGFP* and PEI-*pEGFP* served as positive controls.

Dynamic uptake assay

The dynamic uptake process of PEI-*pEGFP* and CBA-CAR/CBA-CHL-*pEGFP* in MCF-7 cells was recorded at different intervals by laser scanning confocal microscopy. In order to display the intracellular localization of *pDNA*, Total NUCLEAR-ID Green/Red nucleolar/nuclear detection kit, Hoechst 33342, and DiO were used to stain the nucleolus, nucleus, *pDNA*, and cell membrane, respectively, according to the protocols provided by the manufacturers. In brief, *pEGFP* (2 µg/well) and Hoechst 33342 (33.3 µg/mL) were incubated at 37°C for 1 hour. The stained *pEGFP* was then precipitated with 70% ethanol and resuspended in 1× PBS. MCF-7 cells were seeded at a density of 5×10⁴ cells/well (glass bottom dish, *d*=35 mm) and cultured overnight in normal cell culture conditions as previously mentioned. After incubation, the cells in each well were washed with PBS thrice and stained with the Total NUCLEAR-ID Green/Red nucleolar/nuclear detection kit and DiO sequentially. Simultaneously, the resuspended *pEGFP* was incubated with polymers to prepare different complexes at the N/P weight ratio of 1/24 following the method described earlier. After staining, the cells were incubated in complete culture medium containing different complexes, and the localization of complexes was recorded at 5 minutes, 30 minutes, 1 hour, 2 hours, 3 hours, and 4 hours using laser scanning confocal microscopy (Olympus FV1000S-SIM/IX81; Olympus Corporation, Tokyo, Japan) with a 60× objective lens. The intracellular *pDNA*/polymer, nucleus, nucleolus, and cell membrane appeared in blue, red, green and green, respectively.

Statistics

Statistical comparisons were performed using a one-way analysis of variance, followed by the Dunnett test. Pairwise comparisons between treatments were achieved by using Student's *t*-test. A *P*-value of <0.05 was considered statistically significant.

Results and discussion

Synthesis of CAR-CBA and CHL-CBA

In previous studies involving SS-PAAAs as gene delivery carriers, the presence of bioreducible disulfide linkages in these polymers resulted in significant increases in transfection efficiency along with low cytotoxicity.²⁸ In this study, two amino monomers, CHL and CAR, were used to introduce guanidino groups into SS-PAAAs, leading to the synthesis of CAR-CBA and CHL-CBA following the route shown in Figure 1. CHL is well known and widely used as a bacteriostatic agent in the pharmaceutical field or as a finishing agent in the textile industry with good bioactivity, biocompatibility, and low cytotoxicity. Another guanidine donor, CAR, was chosen in our study to compare it with CHL.

Due to low reactivity of guanidine, Pbf-Cl was used to activate the guanidine group before the Michael addition polymerization occurred between CBA and CHL/CAR. Trifluoroacetic acid was used for deprotecting Pbf-Cl after the Michael addition polymerization. The introduction of the guanidine group was achieved by the polymerization of CBA with the guanidine-based monomer, without any other modification after polymerization happening on the side chains. Compared to traditional methods, which introduce guanidine groups utilizing 1*H*-pyrazole-1-carboxamide hydrochloride and *N,N*-diisopropylethylamine, this new synthetic route requires milder reaction conditions, facilitates product purification and collection, simplifies the entire preparation process, and, not surprisingly, leads to a reduction of the total production cost.

The ¹H NMR spectra of CBA in deuterated DMSO solvent confirmed that CAR/CHL-CBA polymers were successfully connected to CBA because the characteristic olefin peak of CBA disappeared (data shown in [Figures S1–S5](#)). The molecular weights and polydispersity of two Gua-SS-PAAAs were measured by gel permeation chromatography, and the results are listed in Table 1.

Hoechst 33342 intercalation assay

The DNA EE of two polymers (CAR/CHL-CBA) at different N/P ratios was determined by Hoechst 33342 intercalation and agarose gel assay (Figure 2). As shown in the figure, the EEs of CAR-CBA were all >70% at N/P ratios ranging

Table 1 Molecular weight and polydispersity of CAR–CBA and CHL–CBA polymers measured by GPC

Polymer	Retention time (minutes)	M_n (g/mol)	M_w (g/mol)	M_p (g/mol)	M_z (g/mol)	PD (M_w/M_n)
PEI	17.292	12,894	23,610	17,658	46,230	1.83
CAR–CBA	18.017	7,198	10,595	8,203	18,302	1.47
CHL–CBA	18.250	6,139	7,609	6,989	9,908	1.24

Abbreviations: CAR, guanidine hydrochloride; CBA, *N,N'*-cystamine bisacrylamide; CHL, chlorhexidine; PEI, polyethylenimine; GPC, gel permeation chromatography; M_n , number average molecular weight; M_p , peak molecular weight; M_w , weight average molecular weight; PD, polydispersity.

from 1/1 to 1/96, except at the ratio of 1/0.5, while the EEs of CHL–CBA were ~80%, except at the ratio of 1/96. The EEs of PEI and Lipo used at concentrations recommended by manufacturers (Lipo at 1 μ L/ μ g *p*DNA; PEI at the N/P ratio of 1/12) were 87.31% \pm 7.21% and 75.23% \pm 6.50%, respectively. These results show that there was no significant difference in EEs between the two polymers, and both of them exhibited equal or even higher EEs than PEI or Lipo. Moreover, *p*DNA was fully condensed at N/P ratios from 1/1 to 1/48. Notably, CHL–CBA showed a much higher EE (91.58% \pm 5.98%) than CAR–CBA (45.59% \pm 6.63%) at the ratio of 1/0.5, indicating that the former had an enhanced encapsulation capability due to its complicated side-chain structure. In addition, the EE of both polymers reached the peak value at the N/P ratio of 1/24 and did not increase, while the amount of polymers was increased. Instead, a slight downward trend was observed when the ratio was over 1/24, especially for CHL–CBA (falling to 63.66% \pm 6.03% at the ratio of 1/96).

AFM analysis of complex particle formation

The DNA condensation capabilities of two polymers (CAR/CHL–CBA) were further studied by AFM (Figure 3). Based

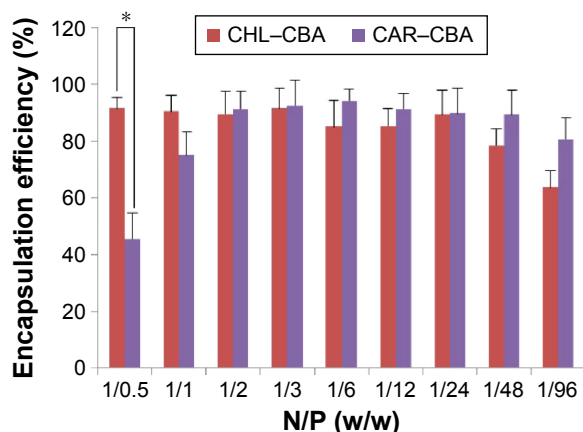


Figure 2 Encapsulation efficiency of CAR–CBA and CHL–CBA at N/P ratios varying from 1/0.5 to 1/96.

Note: Student's *t*-test, **P*<0.05.

Abbreviations: CAR, guanidine hydrochloride; CBA, *N,N'*-cystamine bisacrylamide; CHL, chlorhexidine; N/P, nucleic acid/polymer weight ratio.

on the results of a pretest, inadequate polymer dosage (PAAs) could affect the adhesion between the complexes and mica, leading to difficulties in the imaging process of AFM analysis, which was consistent with Piest and Engbersen's²⁶ report. Therefore, the N/P ratio of the two complexes (CAR/CHL–CBA–*p*DNA) was set as 1/48 based on that report.²⁶ As the top row in Figure 3 shows, a typical plectonemic conformation of supercoiled plasmids is observed after naked *p*DNA was deposited onto freshly cleaved mica in the absence or presence of the buffer. Interestingly, we found that there was no particle-like images in CAR–CBA and CHL–CBA groups in the absence or presence of buffer when the N/P ratio was below 1/6 (data not shown), which implied that, though an interaction between *p*DNA and the polymer was observed in Hoechst 33342 intercalation assay at N/P ratios ranging from 1/0.5 to 1/6, no visible particle-like form condensed by two polymers was actually formed. The images of complexes on dried mica at an N/P ratio of 1/48 (CHL/CAR–CBA) revealed that sizes of these particles were as follows: Lipo (400 \leq 500 nm), CHL–CBA (\leq 250 nm), CAR–CBA (\leq 150 nm), and PEI (\leq 100 nm). The morphological images of complexes on mica in the buffer system were quite different from those of the dried complexes, and the sizes of these particles were smaller than those in dried forms and arranged in a different order: Lipo (\leq 100 nm), PEI (\leq 50 nm), CHL–CBA (\leq 40 nm), and CAR–CBA (\leq 30 nm). At the same time, in the buffer system, Lipo–*pEGFP* and PEI–*pEGFP* exhibited a higher particle density than the other two types of complexes at an N/P ratio of 1/48. This could be caused by the blow-dry process, which might have blown away a part of these particles. The AFM analysis of particle formation also demonstrated that significant morphological difference existed between complexes in the dry form and in the buffer system. However, the complexes in the buffer system were settling on the mica surface, which could make a difference in the results. Since the suspended complexes might exhibit a difference in the particle size and other properties, DLS assay was performed for a quantitative assessment of the complexes.

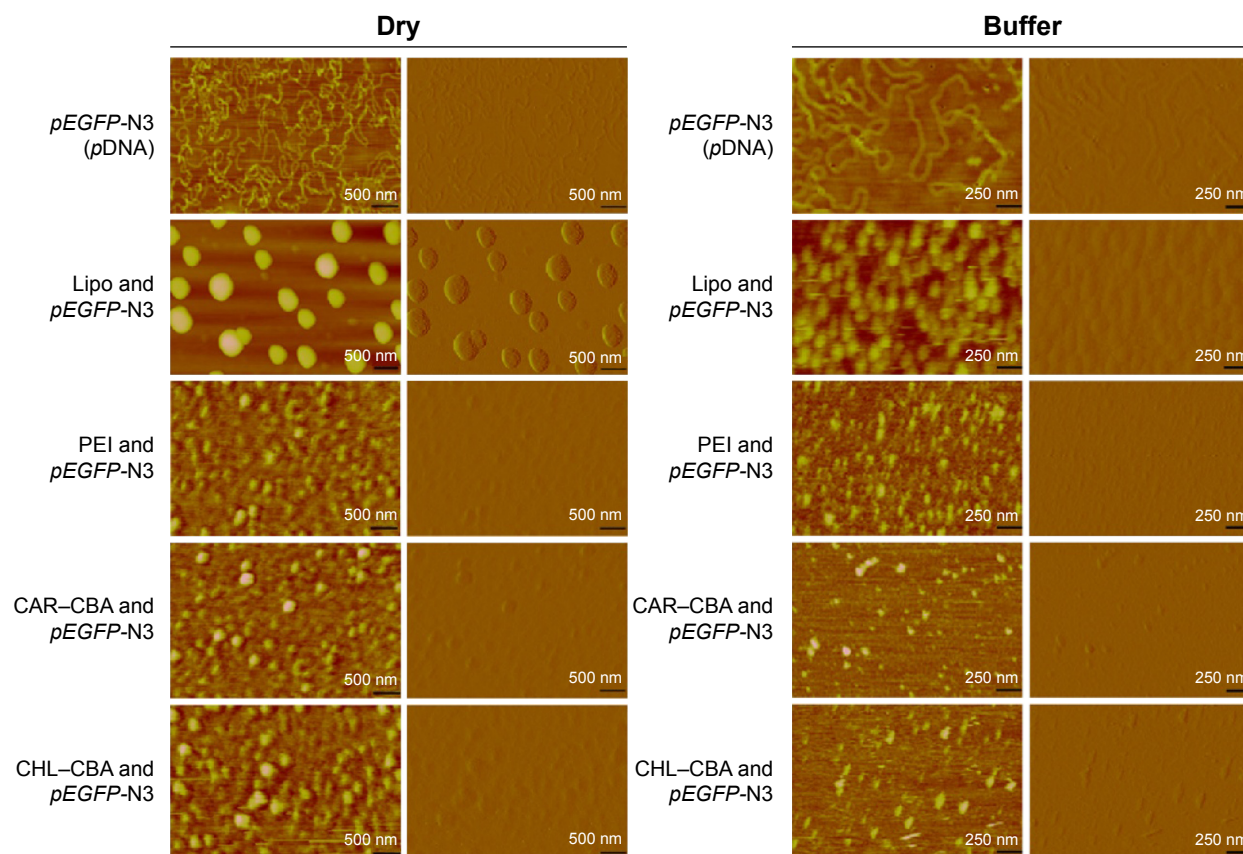


Figure 3 AFM images of Gua-SS-PAA complexes and naked pDNA in dry form or in buffer system.

Notes: CAR-CBA-pDNA and CHL-CBA-pDNA were complexed with pDNA at an N/P ratio of 1/48. Each image represents a $2 \times 2 \mu\text{m}$ scan. Dry, complexes measured in a dry state; buffer, complexes measured in buffer solution.

Abbreviations: AFM, atomic force microscopy; CAR, guanidine hydrochloride; CBA, *N,N'*-cystamine bisacrylamide; CHL, chlorhexidine; Gua-SS-PAA, guanidylated poly(amido amine) with multiple disulfide linkages; Lipo, lipofectamine 2000; N/P, nucleic acid/polymer weight ratio; PEI, polyethylenimine.

Particle size and ζ -potential measurement

The DNA-condensing ability of Gua-SS-PAA (CAR-CBA and CHL-CBA) was evaluated by particle size and ζ -potential measurements. Figure 4 shows that the particle size of CAR-CBA-pDNA complexes at N/P ratios ranging from 1/6 to 1/96 was consistently ~ 50 nm, and the maximum was 62.45 ± 5.82 nm. The ζ -potential of CAR-CBA-pDNA was ~ 20 mV and reached a maximum value (29.60 ± 1.22 mV) at the N/P ratio of 1/24. The particle size of CHL-CBA-pDNA complexes was < 90 nm, which was much larger than that of CAR-CBA-pDNA. When the N/P ratio was varied from 1/12 to 1/96, the particle size increased and reached a maximum of 87.14 ± 3.07 nm. The ζ -potential of CHL-CBA-pDNA showed a similar trend as observed from the measurement of particle size. However, there was no obvious difference between these two types of complexes. Compared to PEI, the average particle sizes of both complexes were larger, but their average ζ -potentials were

lower. Along with an increase in the amount of the polymers, the particle size of CAR/CHL-CBA complexes did not continuously grow, which might be a result of using the optimal N/P ratio. When the amount of polymer reached the limit, the repulsive force between the polymers could interfere with the interaction between polymers and DNA, thereby hindering the complexation process. The large particle size at a low N/P ratio indicated that DNA might not have been condensed compactly. Interestingly, the ζ -potential results of CAR/CHL-CBA demonstrated a similar trend, which confirmed that the particulate structure was a prerequisite for generating the ζ -potential. Based on these observations, the particle size and ζ -potential of the two complexes might be affected not only by the N/P ratio but also by the complex structure and/or some other factors.

Agarose gel electrophoresis assay

The agarose gel electrophoresis assay was applied to evaluate the complexation ability and stability of Gua-SS-PAA.

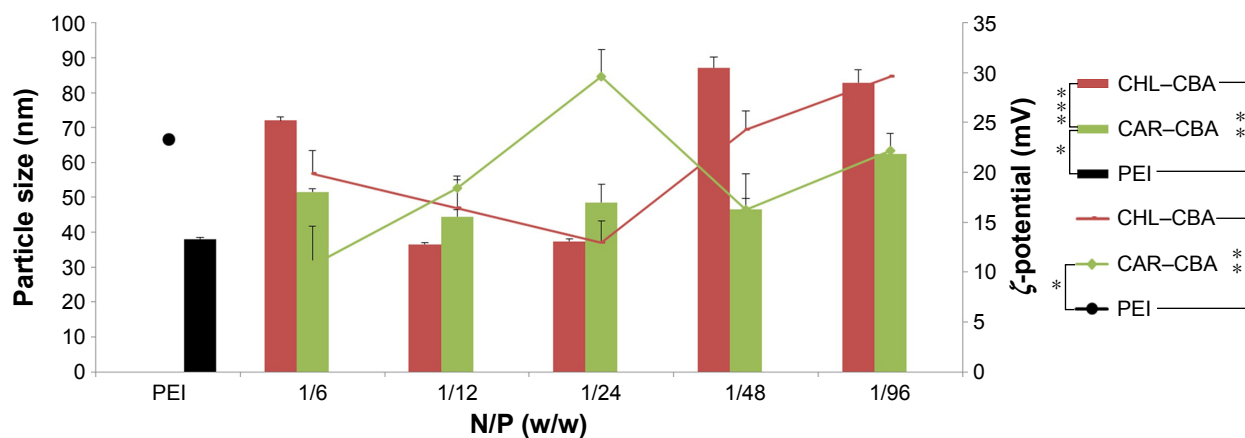


Figure 4 Particle size (in columns) and ζ -potential (in lines) measurements of Gua-SS-PAA complexes.

Notes: CAR-CBA-*p*DNA and CHL-CBA-*p*DNA were complexed with *p*DNA at N/P ratios varying from 1/6 to 1/96. Results are reported as mean \pm SD for three individual measurements. Student's *t*-test, **P*<0.05, ***P*<0.05 and ****P*<0.05.

Abbreviations: CAR, guanidine hydrochloride; CBA, *N,N'*-cystamine bisacrylamide; CHL, chlorhexidine; Gua-SS-PAA, guanidylated poly(amido amine)s with multiple disulfide linkages; N/P, nucleic acid/polymer weight ratio; PEI, polyethylenimine.

As shown in Figure 5A, no retardation was found with *p*DNA alone. Retardation mediated by Lipo at the recommended concentration was not complete since a light band was observed. In contrast, PEI showed complete retardation at the recommended concentration. Similarly, complete retardation was observed for CAR/CHL-CBA at N/P ratios from 1/6 to 1/96. There was a slightly higher amount of free *p*DNA in the channel of CAR-CBA than in CHL-CBA, which indicated that the complexation of CHL-CBA was more capable than CAR-CBA. As shown in Figure 5B, *p*DNA was not replaced by heparin until the concentration of heparin was > 18 U/mL in both CAR-CBA and CHL-CBA complexes. However, in PEI complexes, *p*DNA was replaced until the concentration of heparin reached 36 U/mL. The *p*DNA replaced by heparin was more in CAR-CBA complexes than in CHL-CBA complexes. Furthermore, CHL-CBA/CAR-CBA complexes at the N/P ratio above 1/3 were resistant to DNase I digestion, and CHL-CBA showed better resistance compared to CAR-CBA (Figure 5C). Figure 5C also demonstrates the protection of Lipo and PEI against DNase I-mediated DNA degradation.

Based on the Hoechst 33342, AFM, DLS, and agarose gel assays, we concluded that *p*DNA was well encapsulated in CAR/CHL-CBA, and the encapsulation abilities of CAR/CHL-CBA were greater than that of the two positive controls (Lipo and PEI). Furthermore, the complexation mediated by CHL-CBA was more complete and stable than that by CAR-CBA. This might be due to the difference of functional structures and more nitrogen atoms of the guanidino groups in the side chains of CHL-CBA. In addition, different methods were used to evaluate the condensation ability of polymers,

and these studies showed diverse results, which should not be simply discarded and will be explored in the future.

Reduction sensitivity of the Gua-SS-PAA

Reduction-triggered disassembly of Gua-SS-PAA was evaluated under reducing conditions using agarose gel electrophoresis (Figure 6). Li et al¹⁴ reported that 0.75 M NaCl in HBS was indispensable for *p*DNA release. Incubation with 20 mM GSH resulted in the dissociation of the complexes (CAR/CHL-CBA) and a release of *p*DNA at N/P ratios from 1/6 to 1/48. In contrast, there was no reduction-triggered disassembly observed in Lipo and PEI complexes under the same conditions. As shown in Figure 6, CAR-CBA was more sensitive to GSH than CHL-CBA since more *p*DNA was released from the CAR-CBA-*p*DNA complexes. This might be a result of longer side chains and more guanidine groups in CHL-CBA. In addition, the vast majority of released *p*DNA was in the close-loop state (with faster electrophoretic speed and higher transfection capability), although a small open-loop *p*DNA also existed, indicating that plasmid with high transfection capability could be entrapped, protected, and released reduction-triggering from the Gua-SS-PAA.

Cell viability assay

Cytotoxicity is an important parameter in the development of safe nonviral carriers. High cytotoxicity of a carrier will negatively affect the transfection efficiency. According to the particle size and stability results of complexes from the agarose assay, polymeric carriers were capable of forming stable complexes at N/P ratios from 1/6 to 1/96. In order to investigate the cytotoxicity of Gua-SS-PAA, MCF-7 cells were treated

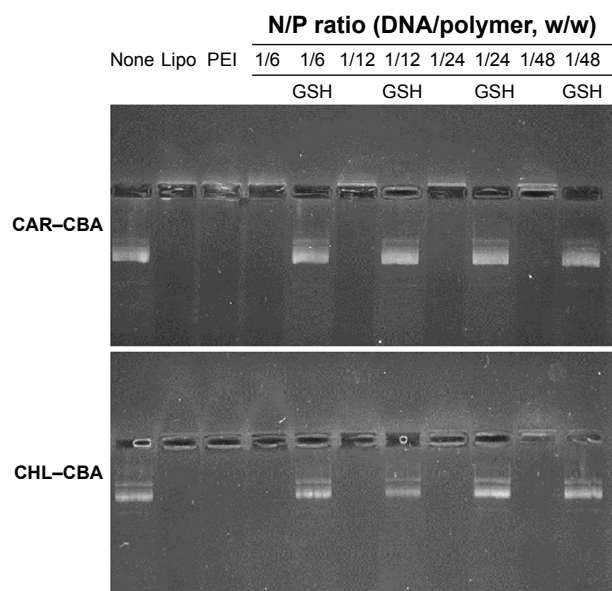


Figure 6 Reduction sensitivity of the Gua-SS-PAA complexes with or without GSH treatment.

Notes: CAR-CBA-pDNA and CHL-CBA-pDNA were complexed with pDNA at N/P ratios varying from 1/6 to 1/48.

Abbreviations: CAR, guanidine hydrochloride; CBA, *N,N'*-cystamine bisacrylamide; CHL, chlorhexidine; GSH, L-glutathione reduced; Gua-SS-PAA, guanidylated poly(amido amine) with multiple disulfide linkages; Lipo, lipofectamine 2000; N/P, nucleic acid/polymer weight ratio; PEI, polyethylenimine.

with the complexes (CAR/CHL-CBA-pDNA) prepared at N/P ratios from 1/6 to 1/96 in the presence of 10% FBS. Cell viability was determined by MTT assays, and Lipo and PEI complexes were used as positive controls. Compared to Lipo and PEI, CAR-CBA and CHL-CBA confer the complexes with significantly lower cytotoxicity, as shown in Figure 7. The cytotoxicity of CAR-CBA complexes was higher than that of

CHL-CBA complexes. The stronger cytotoxicity mediated by CAR-CBA could be the result of a strong interaction between the complex and cell membrane due to the high density of guanidine in the main chain of CAR-CBA.²⁹ In contrast, though CHL-CBA possesses more guanidine groups, these groups are located in the side chains of CHL-CBA, leading to less cytotoxicity. As reported by Martello et al,¹³ cytotoxicity is determined by many factors, including the molecular weight, charge density, type of amines, polymer structure (linear, branched), intracellular degradability, and chain flexibility. In our case, this was a typical example in which the difference in polymer structure caused a discrepancy in the charge density, but not in the ζ -potential, resulting in the difference in cytotoxicity. In addition, though the presence of serum in cell culture medium might result in the enhancement of cell viability,³⁰ this condition was closer to the in vivo working environment of the carriers. Furthermore, the incubation time of complexes was extended to 12 hours, which tripled the in vitro transfection time (4 hours) and further confirmed the low cytotoxicity of these two polymers.

Determination of in vitro transfection efficiency

The transfection efficiencies of the complexes from CAR-CBA and CHL-CBA at different N/P ratios from 1/6 to 1/96 were investigated in MCF-7 cells in the presence of serum. Lipo and PEI served as controls. The fluorescence intensity of green fluorescent protein was analyzed by FM combined with FCM to determine the transfection efficiencies of the carriers (Figure 8). It was found that

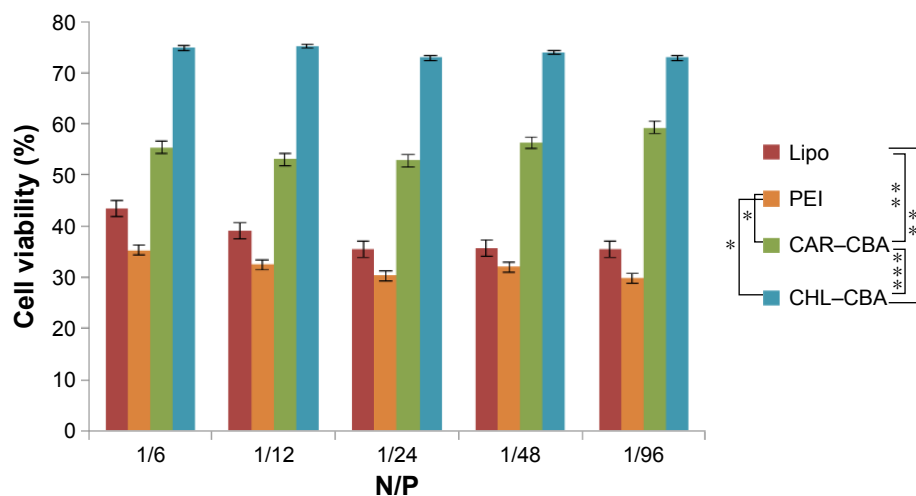


Figure 7 In vitro cytotoxicity of Gua-SS-PAA, CAR-CBA-pDNA and CHL-CBA-pDNA complexed with pDNA at N/P ratios varying from 1/6 to 1/96.

Notes: MCF-7 cells were incubated with the desired amount of complexes for 12 hours. Results are reported as mean \pm SD for three individual measurements. Student's *t*-test, * $P < 0.05$, ** $P < 0.05$ and *** $P < 0.05$.

Abbreviations: CAR, guanidine hydrochloride; CBA, *N,N'*-cystamine bisacrylamide; CHL, chlorhexidine; Gua-SS-PAA, guanidylated poly(amido amine)s with multiple disulfide linkages; Lipo, lipofectamine 2000; N/P, nucleic acid/polymer weight ratio; PEI, polyethylenimine.

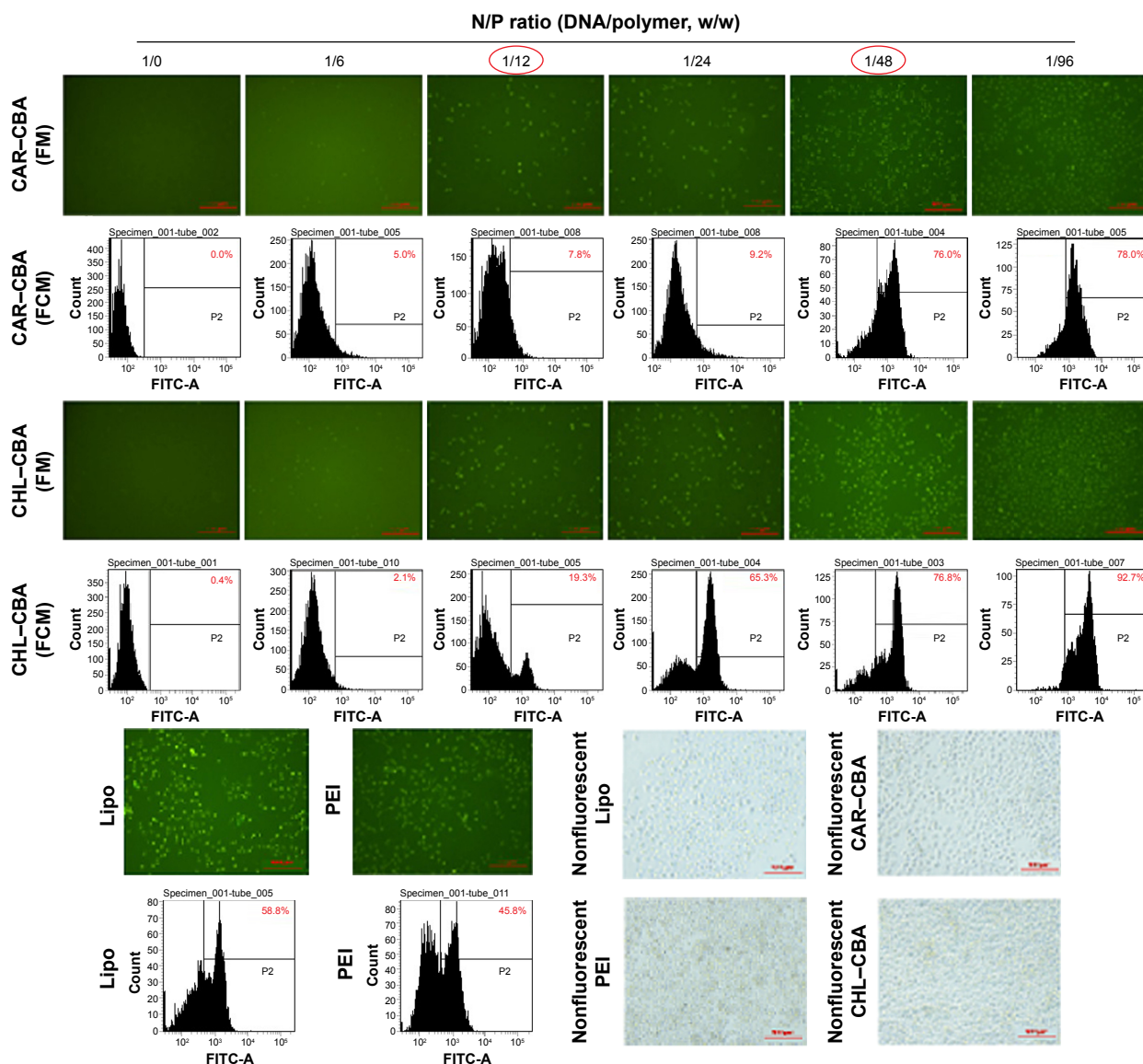


Figure 8 In vitro transfection and expression results of Gua-SS-PAs at N/P ratios varying from 1/6 to 1/96.

Notes: The fluorescence intensity of EGFP was observed by FM (100 \times , scale bar 100 μ m), and transfection efficiency was determined by FCM after 48 hours gene transfection. Nonfluorescent means MCF-7 cell images recorded under natural light as cell density control.

Abbreviations: CAR, guanidine hydrochloride; CBA, *N,N'*-cystamine bisacrylamide; CHL, chlorhexidine; FCM, flow cytometry; FM, fluorescence microscopy; Gua-SS-PAs, guanidylated poly(amido amine)s with multiple disulfide linkages; N/P, nucleic acid/polymer weight ratio; PEI, polyethylenimine; Lipo, lipofectamine 2000; EGFP, enhanced Green Fluorescent Protein.

the transfection efficiencies of CAR-CBA and CHL-CBA increased with the increase in dosage of the polymers and reached a maximum of 78.0% and 92.7%, respectively, at the N/P ratio of 1/96. Compared to those of the two controls, the transfection efficiencies of the two polymers were considerably enhanced when the N/P ratio was $>1/48$. At the same time, the transfection efficiency of CHL-CBA was much higher than that of CAR-CBA when the N/P ratio was $>1/12$. These data were consistent with the results from DLS assays, indicating that the transfection efficiency was not related to ζ -potential but affected by

other factors. For instance, the transfection efficiency of the two complexes (CHL/CAR-CBA-pDNA) increased with the particle size especially when the N/P ratio was varied from 1/48 to 1/96. Combining the results from cell viability and transfection efficiency, we conclude that the higher cytotoxicity of CAR-CBA might lead to a negative effect on transfection efficiency. In addition, the main reason might be due to the nuclear localization effect (NLE) mediated by the guanidine groups in the side chains of CHL-CBA. This hypothesis was further tested in the following in vitro cellular uptake study.

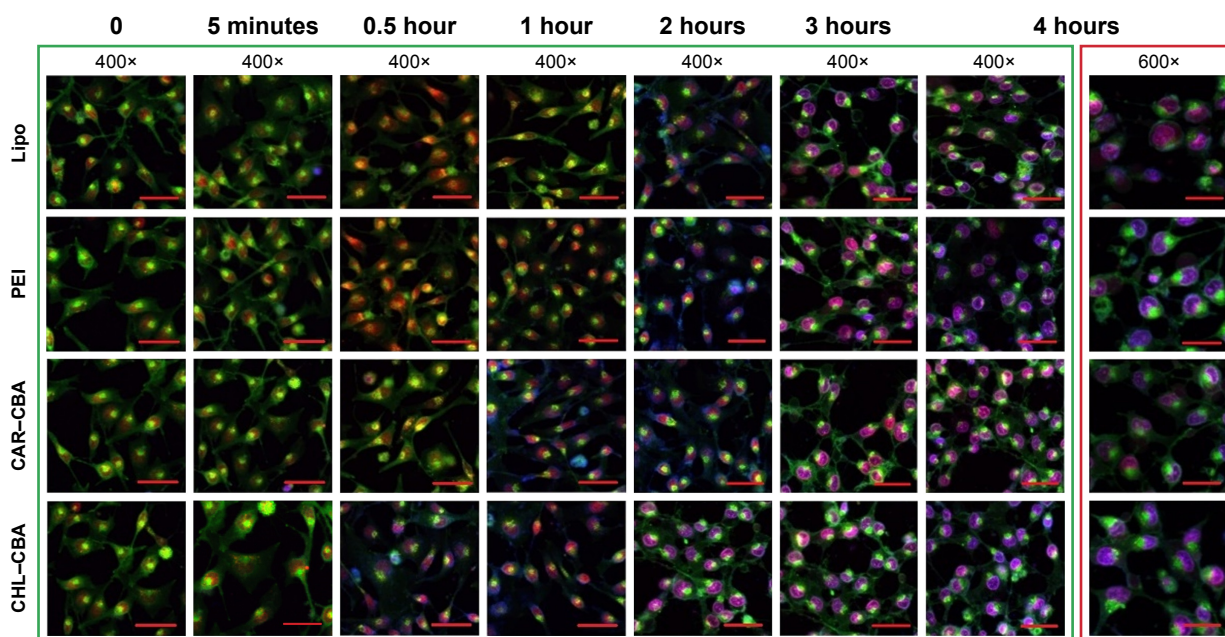


Figure 9 Intracellular trafficking of CAR-CBA-*p*DNA and CHL-CBA-*p*DNA at an N/P ratio of 1/48 at 5-minute, 30-minute, 1-hour, 2-hour, 3-hour, and 4-hour time intervals.

Notes: Lipo and PEI were used at the recommended concentration. Scale bar 100 μ m. The fluorescence signals were collected by LSCM with three channels: *p*DNA stained with Hoechst 33342, blue; DiO-labeled cell membrane, green; nucleolus stained with the nucleolus tracker, green; nuclear stained with the nuclear tracker, red. Only merged images of three channels are shown.

Abbreviations: CAR, guanidine hydrochloride; CBA, *N,N'*-cystamine bisacrylamide; CHL, chlorhexidine; DiO, 3,3'-dioctadecyloxacarbocyanine perchlorate; Lipo, lipofectamine 2000; LSCM, laser-scanning confocal microscopy; N/P, nucleic acid/polymer weight ratio; PEI, polyethylenimine.

In vitro cellular uptake

In order to further understand the effects of polymer/*p*DNA complexes on the process of transfection, the uptake and subcellular localization of the complexes were investigated. In this study, naked Hoechst 33342-labeled *p*DNA was used as control. As shown in Figure 9, the *p*DNA stained with blue could be hardly found at 0 minutes or 5 minutes. CHL-CBA-*p*DNA complexes were found in the nucleus and cytoplasm of MCF-7 cells, corresponding to the purple and light blue colors, respectively, but no significant color change was observed in the other three groups 30 minutes after the transfection process started. CAR-CBA-*p*DNA appeared in the nucleus 1 hour after the transfection process started, while Lipo-*p*DNA and PEI-*p*DNA just started entering the nuclear region at that time. Four hours after the transfection process started (ie, the ending time point), all treatment groups demonstrated that many of the complexes had already entered the nuclear regions since the red-stained nucleus had turned into dark purple or blue. Moreover, there was more *p*DNA appearing in the nucleus in the PEI and CHL-CBA groups than in the Lipo and CAR-CBA groups. In addition, more intact cells were observed in the CHL/CAR-CBA groups compared with other two groups, which was consistent with

cell viability results, confirming the lower cytotoxicity of CHL/CAR-CBA.

Conclusion

Two different Gua-SS-PAAs carriers were successfully prepared. Both of them showed higher transfection efficiency and lower cytotoxicity than two widely used carriers (PEI and Lipo). Furthermore, the reasons for the biological difference between these two carriers related to their structural factors were discussed. The results of physical/chemical analysis and biological studies confirmed that both Gua-SS-PAAs carriers possessed promising encapsulation, cell membrane permeability, and nuclear localization effect. In addition, the heparin replacement and DNase I degradation assay suggested that the new carriers might have potential applications as delivery carriers in gene therapy. Certainly, the systemic circulation stability, distribution, and safety issues such as hemolysis or immunogenicity, need to be further investigated through comprehensive in vivo experiments.

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Disclosure

The authors report no conflicts of interest in this work.

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