

Hydrogen sulfide, a potential novel drug, attenuates concanavalin A-induced hepatitis

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Background: Hydrogen sulfide (H₂S) is known to exert anti-inflammatory properties. Apoptosis and autophagy play important roles in concanavalin A (Con A)-induced acute hepatitis. The purpose of this study was to explore both the effect and mechanism of H₂S on Con A-induced acute hepatitis.

Methods: BALB/c mice were randomized into sham group, Con A-injection group, and 14 μmol/kg of sodium hydrosulfide (NaHS, an H₂S donor) pretreatment group.

Results: Aspartate aminotransferase, alanine aminotransferase, and pathological damage were significantly ameliorated by NaHS pretreatment. NaHS pretreatment significantly reduced the levels of interleukin-6 and tumor necrosis factor-α compared with those of the Con A group. The expression of Bcl-2, Bax, Beclin-1, and LC3-2, which play important roles in the apoptosis and autophagy pathways, were also clearly affected by NaHS. Furthermore, NaHS affected the p-mTOR and p-AKT.

Conclusion: H₂S attenuates Con A-induced acute hepatitis by inhibiting apoptosis and autophagy, in part, through activation of the PtdIns3K-AKT1 signaling pathway.

Keywords: NaHS, apoptosis, PtdIns3K-AKT, autophagy

Introduction

Hepatitis seriously threatens human health and daily life. It is caused by various factors such as viruses, drugs, chemicals, alcohol, genetic factors, or a patient's own immune system. Currently, effective drugs in the treatment of hepatitis are lacking, so effective therapies must be explored to provide new applications in the clinic. Concanavalin A (Con A) can induce necrosis of hepatocytes; this causes hepatic dysfunction and also attracts and activates immune cells. Pathological studies have shown infiltration and a large number of lymphocyte accumulation of mainly the CD4+ T cells in the liver parenchyma.^{1,2} Therefore, Con A is often applied in the animal model of acute hepatitis.³

There are complicated mechanisms in the evolution of Con A-induced hepatitis. Recent studies show that blocking MAPK and activating AKT cell death pathways can both significantly reduce Con A-induced hepatitis.^{3,4} Apoptosis, named type I programmed cell death, may be a major cell death mechanism in Con A-induced hepatitis. The Bcl-2 family is considered to have an important role in the apoptosis pathway. In the Bcl-2 family, the representative antiapoptotic genes are *Bcl-2* and *Bcl-xl*, and the proapoptotic genes are *Bax* and *Bad*. It has been reported that the balance between Bax and Bcl-2 proteins determines the survival status of cells after particular kinds of stimulus or damage.^{1,5}

Autophagy, named type 2 programmed cell death, is a newly described type of programmed cell death. Autophagy is a protective cell behavior for surviving harsh environments.^{6,7} However, beyond a certain range, it eventually leads to cell death, with the excessive accumulation of autophagosomes.⁸ Damaged organelles are surrounded by isolation membrane to form autophagosomes, and these autophagosome fuse with lysosomes to form autophagy-lysosomes. It has been confirmed that many autophagy-related genes are linked with the formation of autophagy. Autophagy-related gene protein Beclin-1 and the diaphragm combine to generate the production of the Atg12-Atg5-Atg16 complex. Further, raised LC3-2 binds to the diaphragm to promote the extension of autophagy membrane. Mature autophagosomes eventually form while the Atg12-Atg5-Atg16 complex is deactivated. Therefore, restraining the expression of the Atg family can effectively block the formation of autophagosome.

Hydrogen sulfide (H₂S), a third common endogenous signaling molecule after nitric oxide and carbon monoxide, is known to exert anti-inflammatory properties.⁹ H₂S is naturally synthesized in the liver from L-cysteine by two main enzymes, cystathionine-β-synthetase and cystathionine-γ-lyase (CSE).¹⁰ DL-propargylglycine (PAG), an irreversible inhibitor of CSE, penetrates cell membranes and decreases H₂S production in many rat tissues.^{11,12} Generally, two-thirds of H₂S molecules dissociate into bisulfide ions and hydrogen ions. Therefore, sodium hydrosulfide (NaHS) can be seen as a water-soluble H₂S donor.¹³ In this study, we explored both the effect of H₂S on Con A-induced acute hepatitis and the regulation of apoptosis and autophagy.

Materials and methods

Reagents and drug administration

Con A and dimethyl sulfoxide were purchased from Sigma-Aldrich (St Louis, MO, USA) and stored at 4°C. PAG was purchased from Sigma-Aldrich, and the RNA polymerase chain reaction (PCR) kit was obtained from Takara Biotechnology (Dalian, People's Republic of China). Fetal bovine serum and Dulbecco's Modified Eagle's Medium were obtained from Thermo Fisher Scientific (Waltham, MA, USA). LC3-2 and Beclin-1 were purchased from Abcam (Cambridge, UK). The antibodies used for immunoblotting and immunohistochemical staining were anti-tumor necrosis factor (TNF)-α, anti-interleukin (IL)-6, anti-mTOR, anti-p-mTOR, anti-p-AKT, and anti-AKT (Santa Cruz Biotechnology, Dallas, TX, USA). Con A was dissolved in pyrogen-free physiological saline and intravenously injected at a dose of 20 mg/kg body weight to induce hepatitis as previously

described.¹ NaHS was dissolved in saline and intravenously injected at a dose of 14 μmol/kg body weight. PAG was dissolved in pyrogen-free physiological saline and intraperitoneally injected at a dose of 50 mg/kg body weight.

Animals and ethics statement

Male BALB/c mice (23±2 g, 6–8 weeks old) were obtained from Shanghai SLAC Laboratory Animal Co, Ltd (Shanghai, People's Republic of China). This study conscientiously implemented principles from the Guide for the National Science Council of the Republic of China and the Animal Care and Use Committee of The Tenth People's Hospital of Shanghai (permit number 2011-0111).

Model establishment and experimental design

Based on previous experiments by our team, time points at 8, 12, and 24 hours were used. Con A was dissolved in pyrogen-free physiological saline at a concentration of 2.5 mg/mL as previously described. Next, 72 mice were randomly divided into four groups of 18 mice each.

Group 1 (normal control, n=18) mice were intravenously injected in the tail with physiological saline only; group 2 (model group, n=18) mice were intravenously injected in the tail with 20 mg/kg Con A; group 3 (PAG group, n=18) mice were intravenously injected in the tail with 50 mg/kg PAG 1 hour prior to Con A injection;¹⁴ group 4 (protected group, n=18) mice were intravenously injected in the tail with NaHS (14 μmol/kg) 1 hour prior to Con A injection. Eighteen mice from each group were randomly killed at 8, 12, and 24 hours. All liver tissue (median and left lobes) and serum were collected and stored at -80°C.

Biochemical analysis

After blood collection, serum was separated by centrifugation at 2,000 rpm for 10 minutes at room temperature. Serum aspartate aminotransferase (AST) and alanine aminotransferase (ALT) were tested using an automated chemistry analyzer (Olympus AU1000; Olympus Corporation, Tokyo, Japan).

Histopathology

When mice were killed, their liver tissues (median and left lobes) were collected, incubated in 4% paraformaldehyde for more than 24 hours, and prepared in paraffin blocks according to the traditional method. The degree of inflammation and tissue damage was observed in paraffin sections (3 μm thick) stained with hematoxylin and eosin using an optical microscope.

Measurement of plasma H₂S

Plasma H₂S was measured as described by Chunyu et al.¹⁵

Assay of liver H₂S-synthesizing activity

H₂S-synthesizing activity in liver was detected as described by Zhang et al¹⁶ and Labarca and Paigen.¹⁷

Immunohistochemistry

Immunohistochemistry using the antibodies against Beclin-1 (1:2,000), LC3-2 (1:500), p-AKT (1:1,000), p-mTOR (1:1,000), and rabbit anti-mouse was performed according to standard procedures.¹⁸ Slides were then observed by optical microscopy.

Western blot analysis

Total protein was prepared using standard procedures. Protein samples were separated by electrophoresis in a sodium dodecyl sulfate polyacrylamide gel and transferred to polyvinylidene fluoride membrane. After blocking, membranes were incubated overnight with different primary antibodies and β-actin at 4°C. After incubation with peroxidase-conjugated secondary antibodies for 1 hour at 37°C, membranes were developed with the Odyssey two-color infrared laser imaging system (LI-COR Biosciences, Lincoln, NE, USA).¹⁸

Reverse transcription-PCR and real-time PCR

The mRNA transcripts from liver tissues were detected and analyzed via quantitative reverse transcription-PCR. Total RNA was extracted using TRIzol[®] reagent (Thermo Fisher Scientific) as described by the manufacturer's instructions. Reverse transcription-PCR was performed in a 7900HT Fast Real-time PCR system (Thermo Fisher Scientific) according to the manufacturer's guidelines. The primers used in the PCR reactions are listed in Table 1.

Transmission electron microscopy

Male BALB/c mice were treated as described above, and laparotomy was performed under ketamine/xylazine anesthesia. The liver was perfused with 2 mL of 2.5% glutaraldehyde in phosphate buffered saline and was then treated as previously described.¹⁹

Statistical analysis

Statistical significance was assessed using the Student's *t*-test or Tukey's post hoc tests. All statistical analyses were performed using SPSS 17.0 (SPSS, Inc., Chicago, IL, USA), and *P*<0.05 was considered significant.

Table 1 Primers used in the polymerase chain reactions

Gene	Primer sequence (5' → 3')
TNF-α	
Forward	CAGGCGGTGCCTATGTCTC
Reverse	CGATCACCCCGAAGTTCAGTAG
IL-6	
Forward	CTGCAAGAGACTTCCATCCAG
Reverse	AGTGGTATAGACAGGTCTGTTGG
β-actin	
Forward	GGCTGTATTCCCCTCCATCG
Reverse	CCAGTTGGTAACAATGCCATGT
Bax	
Forward	AGACAGGGGCCTTTTTGCTAC
Reverse	AATTCGCCGGAGACACTCG
Bcl-2	
Forward	GCTACCGTCGTGACTTCGC
Reverse	CCCCACCGAACTCAAAGAAGG
LC3-2	
Forward	GACCGCTGAAGGAGGTGC
Reverse	AGAAGCCGAAGGTTTCTTGCG
Beclin-1	
Forward	ATGGAGGGGTCTAAGGCCTC
Reverse	TGGGCTGTGGTAAGTAATGGA
CSE	
Forward	GCAGCGATTACACCACAAACC
Reverse	AATATCAGCACCCAGAGCCAAAG

Abbreviations: CSE, cystathionine-γ-lyase; IL-6, interleukin-6; TNF-α, tumor necrosis factor-α.

Results

H₂S pretreatment ameliorates Con A-induced hepatitis

We performed an assay using a Con A-induced hepatitis model. The expression level of CSE mRNA in the liver was significantly increased in the Con A group compared with that in the saline group. Con A resulted in a significant increase in the plasma H₂S level and the amount of H₂S formed in liver compared with the saline group, and NaHS preconditioning further increased them compared with the Con A group. In addition, PAG pretreatment significantly decreased the expression level of CSE mRNA, reduced the plasma H₂S level and the amounts of H₂S formed in liver compared with the Con A group (*P*<0.05) (Figure 1A). Liver function was assessed by measuring the serum level of ALT and AST. As shown in Figure 1B, the level of ALT and AST clearly increased after Con A injection compared with the saline group at 8, 12, and 24 hours (*P*<0.05), and NaHS pretreatment significantly attenuated the Con A injection-induced elevation of serum ALT and AST (*P*<0.05). Conversely, PAG pretreatment aggravated serum ALT and AST at all three time points compared to the Con A group (*P*<0.05) (Figure 1B). Histopathological changes in the liver tissues from the three groups were examined

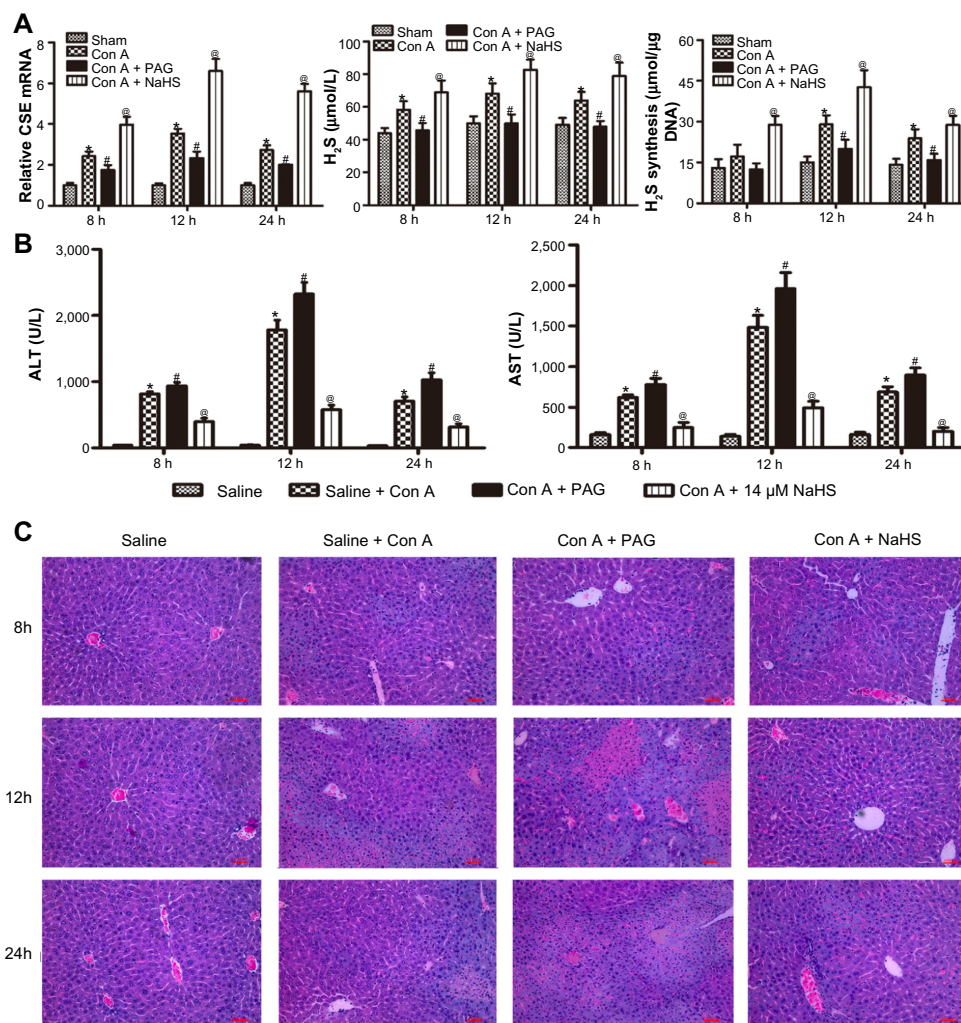


Figure 1 H₂S pretreatment ameliorates Con A-induced hepatitis.

Notes: (A) (Left) The mRNA expression of CSE was detected by real-time PCR $^{*}P < 0.05$ for saline vs Con A, $^{*}P < 0.05$ for Con A vs Con A + PAG, $^{@}P < 0.05$ for Con A vs Con A + NaHS (14 μm/kg). (Middle) Plasma hydrogen sulfide (H₂S) concentration was expressed as the mean ± SD of 6 animals per group. $^{*}P < 0.05$ for saline vs Con A, $^{*}P < 0.05$ for Con A vs Con A + PAG, $^{@}P < 0.05$ for Con A vs Con A + NaHS (14 μm/kg). (Right) Liver H₂S-synthesizing activity was expressed as the mean ± SD of 6 animals per group. $^{*}P < 0.05$ for saline vs Con A, $^{*}P < 0.05$ for Con A vs Con A + PAG, $^{@}P < 0.05$ for Con A vs Con A + NaHS (14 μm/kg). (B) Liver function was assessed in the mice subjected to saline, Con A, Con A + PAG and NaHS + Con A at 8 h, 12 h, 24 h; the results are expressed as the mean ± SD of 6 animals per group. $^{*}P < 0.05$ for saline vs Con A, $^{*}P < 0.05$ for Con A vs Con A + PAG, $^{@}P < 0.05$ for Con A vs Con A + NaHS (14 μm/kg). (C) Representative photographs (200x) of H&E-stained liver sections were taken from mice at 8 h.

Abbreviations: CSE, cystathionine-γ-lyase; PAG, DL-propargylglycine; NaHS, sodium hydrosulfide; Con A, concanavalin A; ALT, alanine aminotransferase; AST, aspartate aminotransferase; H₂S, hydrogen sulfide; h, hours; SD, standard deviation; H&E, hematoxylin and eosin; vs, versus.

after hematoxylin and eosin staining (Figure 1C). The structure of the liver tissues was completely maintained and ordered in the saline group, whereas a disordered lobular structure with some hepatocyte necrosis and polymorphonuclear cell infiltration were observed in the model group at 12 hours and 24 hours. Pretreatment with NaHS at 12 hours attenuated these pathological changes. It is worth noting that at all three time points, the administration of PAG clearly accentuated these pathological features, which included significant congestion of liver, periportal inflammatory cell infiltration, and large fragments of liver tissue necrosis.

H₂S pretreatment inhibits the release of cytokines during Con A-induced hepatitis

Con A-induced hepatitis is associated with changes in the levels of inflammatory cytokines. We demonstrated with real-time PCR and immunoblotting that the expression of TNF-α and IL-6 was significantly increased in the model group compared to that in the saline group (Figure 2A and B). In addition, the location and expression of TNF-α and IL-6 by immunohistochemical staining were stronger in the model group than in the saline group at 12 hours ($P < 0.05$) (Figure 2C). Furthermore, NaHS pretreatment significantly attenuated the expression of

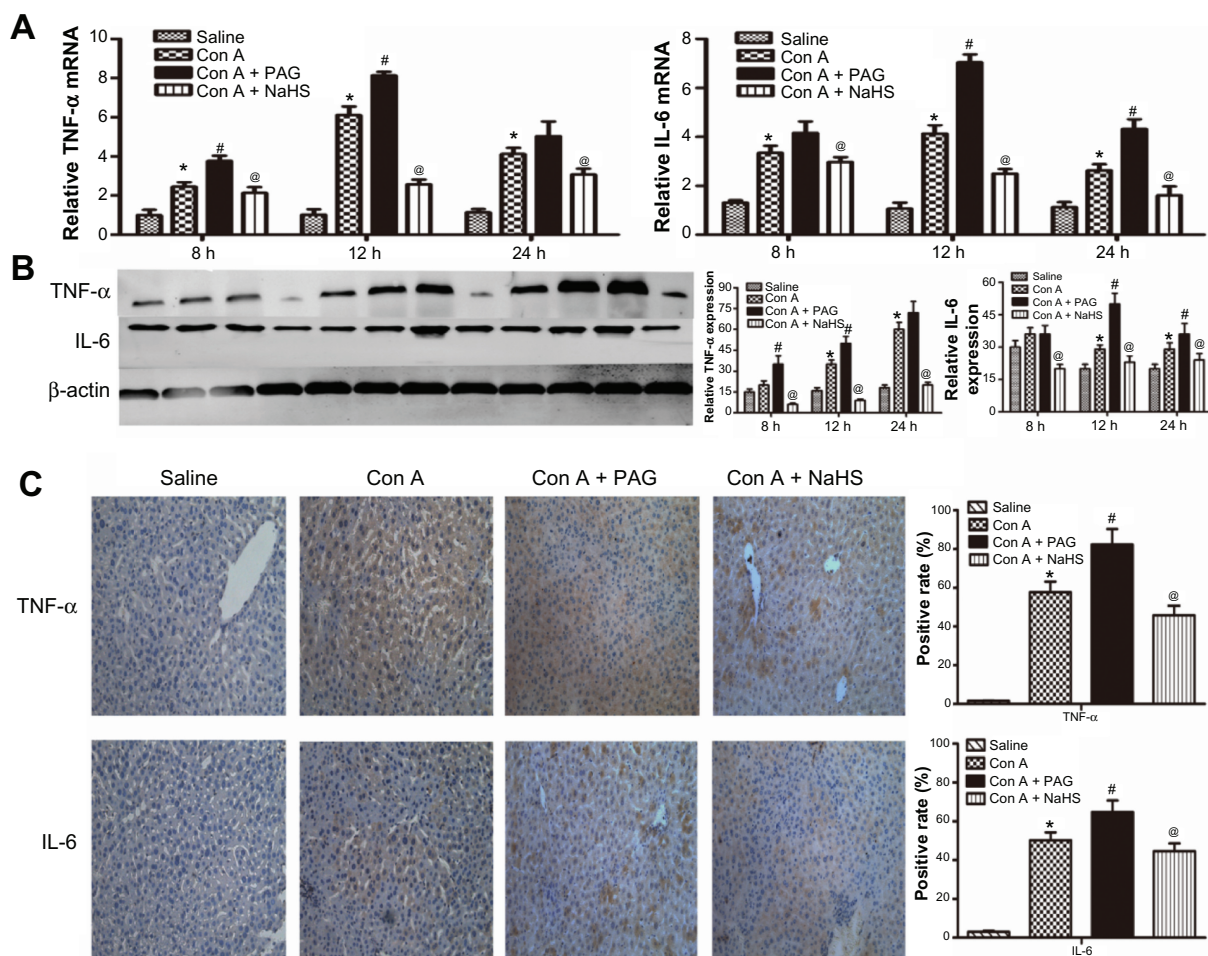


Figure 2 H₂S pretreatment inhibits the release of cytokines during Con A-induced hepatitis.

Notes: (A) Real-time PCR evaluation of mRNA expression of IL-6 and TNF- α in liver tissues. * $P < 0.05$ for saline vs Con A, # $P < 0.05$ for Con A vs Con A + PAG, @ $P < 0.05$ for Con A vs Con A + NaHS (14 $\mu\text{m}/\text{kg}$). (B) Western blots and quantitative evaluation of the expression of IL-6 and TNF- α in liver tissues. * $P < 0.05$ for saline vs Con A, # $P < 0.05$ for Con A vs Con A + PAG, @ $P < 0.05$ for Con A vs Con A + NaHS (14 $\mu\text{m}/\text{kg}$). (C) Immunohistochemistry staining (200 \times) showed the expression of TNF- α and IL6 protein in liver tissue at 12 h. * $P < 0.05$ for saline vs Con A, # $P < 0.05$ for Con A vs Con A + PAG, @ $P < 0.05$ for Con A vs Con A + NaHS (14 $\mu\text{m}/\text{kg}$).

Abbreviations: PAG, DL-propargylglycine; NaHS, sodium hydrosulfide; Con A, concanavalin A; H₂S, hydrogen sulfide; h, hours; IL, interleukin; TNF, tumor necrosis factor, vs, versus.

TNF- α and IL-6 compared to the model group ($P < 0.05$) (Figure 2C). In contrast, with PAG treatment, there were opposite experimental results compared to the NaHS group ($P < 0.05$) (Figure 2C).

H₂S attenuates hepatocyte apoptosis in Con A-induced hepatitis in mice

We next investigated the effect of H₂S on apoptosis in Con A-induced hepatitis in mice. The expression of Bcl-2 and Bax cDNAs was detected with real-time PCR, as expected. NaHS pretreatment significantly increased the expression of Bcl-2 at all three time points and reduced the expression of Bax at 12 hours and 24 hours. PAG preconditioning significantly elicited the opposite effect of NaHS (Figure 3A). NaHS also increased the expression of Bcl-2 at the protein

level at 8 hours and 24 hours, and the expression of Bax was mainly reduced at all three time points with NaHS pretreatment (Figure 3B). However, PAG preconditioning did not displayed the opposite effect of NaHS, it only reduced the role of NaHS.

H₂S attenuates autophagy in Con A-induced hepatitis in mice

It is well known that Con A-induced hepatitis can involve autophagy. LC3 is an important marker of autophagy, and Beclin-1 plays an important role in autophagy. Therefore, to further assess activation of autophagy by NaHS and PAG pretreatment in the Con A model, the expression of LC3 and Beclin-1 in liver tissues were investigated with real-time PCR and immunoblotting (Figure 4A and B). These results

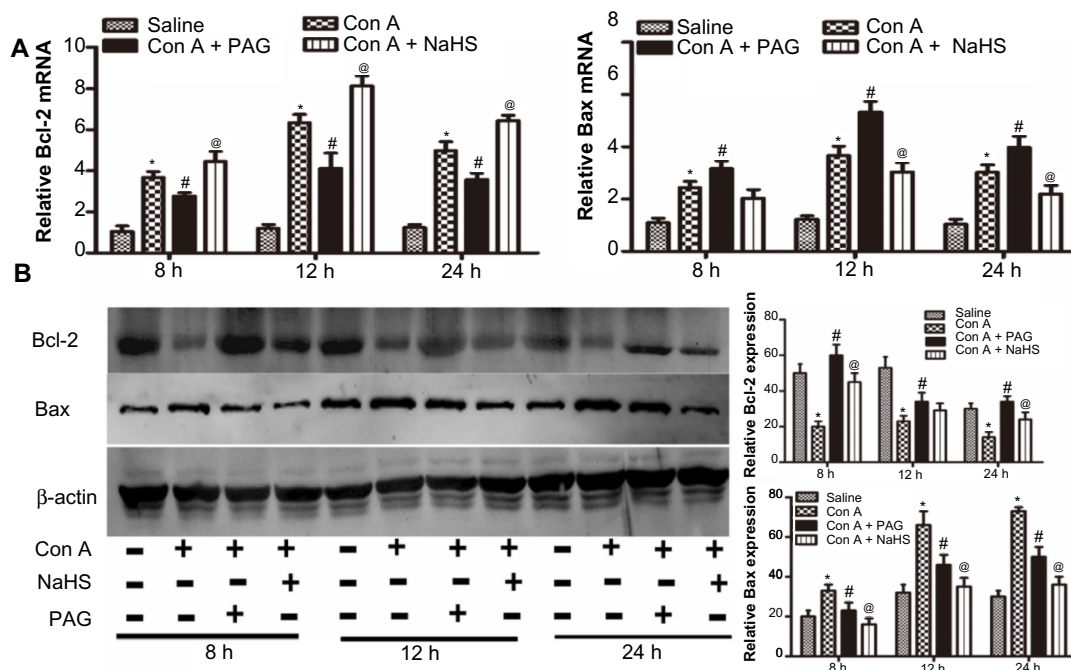


Figure 3 H₂S attenuates hepatocyte apoptosis in Con A-induced hepatitis in mice.

Notes: (A) Real-time PCR evaluation of mRNA expression of Bcl-2 and Bax in liver tissues. * $P < 0.05$ for saline vs Con A, # $P < 0.05$ for Con A vs Con A + PAG, @ $P < 0.05$ for Con A vs Con A + NaHS (14 $\mu\text{m}/\text{kg}$). (B) Western blots and quantitative evaluation of the expression of Bcl-2 and Bax in liver tissues. * $P < 0.05$ for saline vs Con A, # $P < 0.05$ for Con A vs Con A + PAG, @ $P < 0.05$ for Con A vs Con A + NaHS (14 $\mu\text{m}/\text{kg}$).

Abbreviations: PAG, DL-propargylglycine; NaHS, sodium hydrosulfide; Con A, concanavalin A; H₂S, hydrogen sulfide; h, hours; vs, versus.

indicate that the levels of LC3 were significantly reduced after NaHS preconditioning compared to the model group. In contrast, the influence of PAG on autophagy in Con A-induced hepatitis exhibited an increased conversion of LC3. However, Beclin-1 did not appear to have a role in the Con A model and NaHS/PAG models (Figure 4A and B). Analysis of the immunohistochemical changes in the mouse livers confirmed these results (Figure 4C). In addition, the formation of autophagosomes is a pivotal process in autophagy; hence, electron microscopy was applied to observe the ultrastructure of hepatic cells (Figure 4D). Compared with basal levels in the saline group, autophagic vacuoles were dramatically increased in the model group. However, after NaHS treatment, the liver nuclear chromatin was more homogeneous, and the integrity of the cell structure was still intact. Meanwhile, there were more autophagic vacuoles in PAG-treated murine liver tissues.

H₂S activates PtdIns3K-AKT1 signaling in Con A-induced hepatitis in mice

As PtdIns3K-AKT1 signaling plays a key role in regulating inflammatory responses, to further explore the mechanism and effect of H₂S, the activation of AKT and mTOR were analyzed through Western blot analysis of Con A-induced

acute hepatitis in mice (Figure 5A). Con A actually enhanced p-AKT1 and p-mTOR levels in the mouse liver ($P < 0.05$) (Figure 5A), which is further increased by NaHS pretreatment compared to the Con A model group at 12 hours ($P < 0.05$) (Figure 5A). Pretreatment with PAG decreased the activation of AKT1 and mTOR compared to the NaHS group ($P < 0.05$) (Figure 5A). To confirm the results of Western blot analysis, immunohistochemical changes were further detected (Figure 5B).

Discussion

Hepatitis, as a transmitted disease of high incidence, seriously threatens human health and daily life.¹⁹ Currently, effective drugs in the treatment of hepatitis are not readily available, so we must explore new therapies to provide effective applications in the clinic. As previously discussed, Con A is often applied in animal models of acute hepatitis.

The gas H₂S is a new regulator of important physiologic functions. Zhang et al²⁰ reported that H₂S protects liver against ischemia/reperfusion injury, exhibiting anti-inflammatory and antiapoptotic activities. As an irreversible inhibitor of CSE, PAG decreases H₂S production in many rat tissues.²¹ Based on the multiple physiologic functions of H₂S, we speculate that it might be an effective therapy for Con A-induced

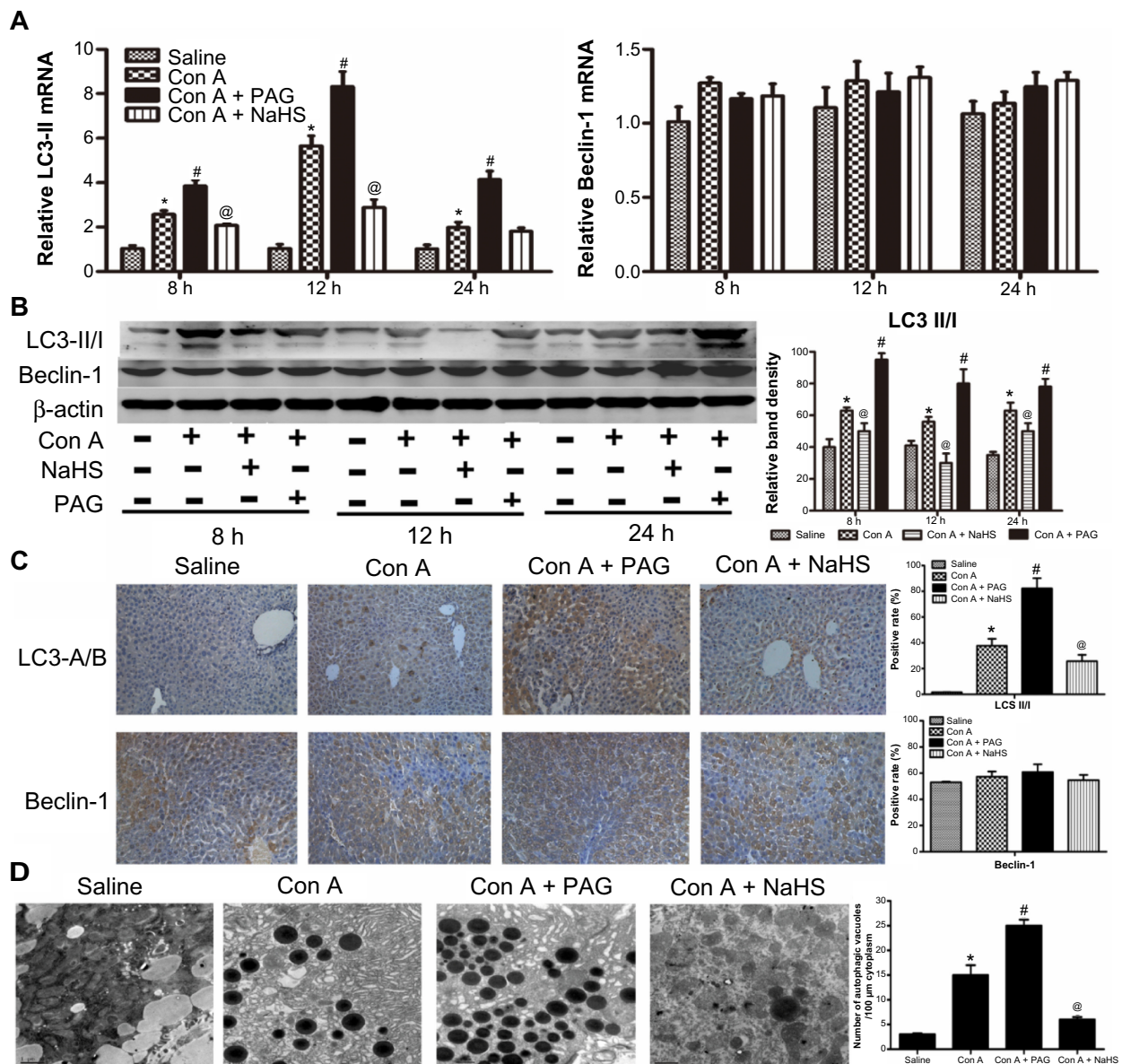


Figure 4 H₂S attenuates autophagy in Con A-induced hepatitis in mice.

Notes: (A) Real-time PCR evaluation of mRNA expression of LC3-II and Beclin-1 in liver tissues. **P*<0.05 for saline vs Con A, #*P*<0.05 for Con A vs Con A + PAG, @*P*<0.05 for Con A vs Con A + NaHS (14 μm/kg). (B) Western blots and quantitative evaluation of the expression of LC3-II and Beclin-1 in liver tissues **P*<0.05 for saline vs Con A, #*P*<0.05 for Con A vs Con A + PAG, @*P*<0.05 for Con A vs Con A + NaHS (14 μm/kg). (C) Immunohistochemistry staining (200×) showed the expression of Beclin-1 and LC3 protein in liver tissue at 12 h. (D) Representative electron micrographs showing autophagic vacuoles in liver sections from mice and the quantification of the number of autophagic vacuoles per 100 μm cytoplasm. Data are expressed as mean ± SD of 6 animals per group. **P*<0.05 for saline vs Con A, #*P*<0.05 for Con A vs Con A + PAG, @*P*<0.05 for Con A vs Con A + NaHS (14 μm/kg).

Abbreviations: PAG, DL-propargylglycine; NaHS, sodium hydrosulfide; Con A, concanavalin A; H₂S, hydrogen sulfide; h, hours; vs, versus; SD, standard deviation.

hepatitis. Our results, indeed, confirmed that H₂S pretreatment ameliorates Con A-induced hepatitis and inhibits the release of cytokines (Figures 1 and 2). It is worth noting that the administration of PAG, which can decrease H₂S production by inhibiting CSE, clearly accentuated Con A-induced hepatitis. This confirmed the protective effect of H₂S from another angle.

PtdIns3K-AKT1 is one of the most important cell survival pathways and is known to play an essential role

in hepatitis. Mayoral et al²² have reported that tissue-specific COX-2-dependent prostaglandins exert efficient protection against Con A-induced hepatitis in mice through an antiapoptotic effect by activating AKT and AMP kinase. Therefore, the PtdIns3K signaling pathway may contribute to multiple endogenous protective pathways to reduce liver injury. AKT can inhibit cytochrome c release through blocking the channel formed by Bcl-2-associated X protein (Bax) and phosphorylating Bad.²³ Therefore, we

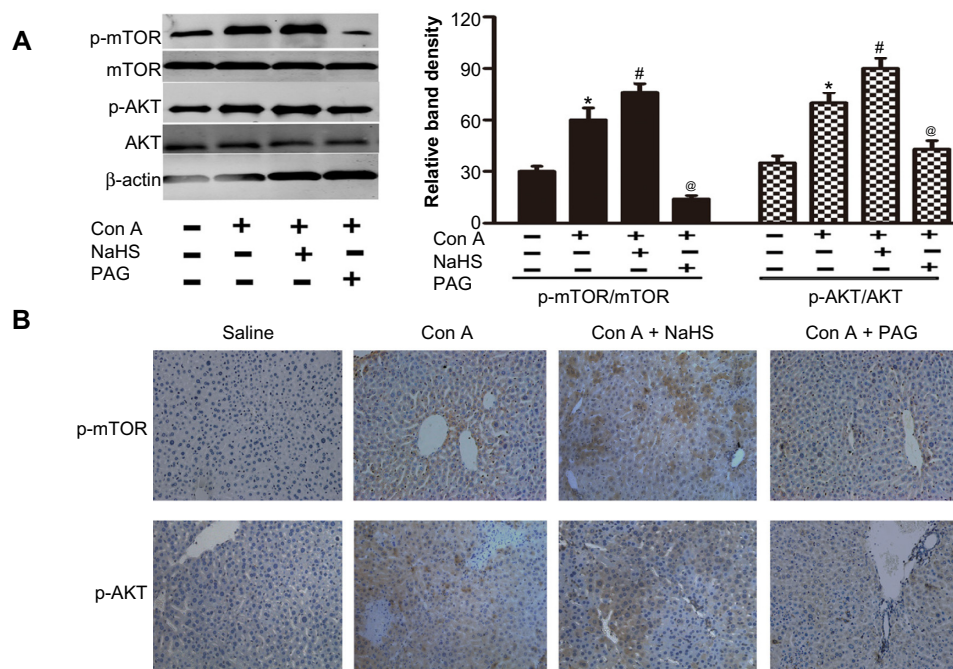


Figure 5 H₂S activates PtdIns3K-AKT1 signaling in Con A-induced hepatitis in mice.

Notes: (A) Western blots and quantitative evaluation of P-AKT1 and p-mTOR expression in liver tissues * $P < 0.05$ for saline vs Con A, # $P < 0.05$ for Con A vs Con A + PAG, @ $P < 0.05$ for Con A vs Con A + NaHS (14 $\mu\text{m}/\text{kg}$). (B) Immunohistochemistry staining (200 \times) showed the expression of Beclin-1 and LC3 protein in liver tissue at 12 h.

Abbreviations: PAG, DL-propargylglycine; NaHS, sodium hydrosulfide; Con A, concanavalin A; H₂S, hydrogen sulfide; h, hours; vs, versus.

attempted to examine the AKT signaling pathway to illuminate how H₂S pretreatment ameliorates Con A-induced hepatitis. Wang et al²⁴ previously reported that H₂S preconditioning produces liver protective effects against hepatic ischemia/reperfusion injury in mice through activation of the PtdIns3K-AKT1 pathway. We have also observed in the present study that NaHS pretreatment significantly increased p-AKT compared to the Con A group (Figure 5). Hence, we supposed that NaHS ameliorated cell death in Con A-induced hepatitis by activating the PI3K/AKT signaling pathway. It is well known that *Bax* is the representative proapoptotic gene, *Bcl-2* is the representative antiapoptotic gene, and the balance between *Bax* and *Bcl-2* proteins has been linked with induction of apoptosis in cell death.¹⁹ Also, our results showed that the Con A increase of *Bax* and decrease of *Bcl-2* finally resulted in the cell death. However, with NaHS treatment, the balance between *Bax* and *Bcl-2* trended to normal, with the upregulation of *Bcl-2* and downregulation of *Bax* (Figure 3B). Hence, we supposed that NaHS ameliorated cell death in Con A-induced hepatitis by inhibiting the intrinsic pathway of apoptosis.

A recent paper has shown that activation of the PtdIns3K-AKT1 signaling molecules link receptor tyrosine kinases to mTOR activation (PI3K-AKT-mTOR), which is an important

signal that inhibits autophagy.²⁵ Studies have reported that Con A can directly induce autophagic cell death of hepatocytes.^{26,27} Con A, after endocytosis, binds to mitochondria to cause mitochondrial dysfunction and autophagy.^{28,29} The role of autophagy in hepatitis is worthy of further exploration, particularly whether H₂S-preconditioning affects autophagy during Con A-induced hepatitis and whether the PI3K-AKT-mTOR pathway mediates it. In our study, the results showed that LC3-II (an important autophagy marker) was inhibited by H₂S preconditioning in hepatocytes, but Beclin-1 was not affected (Figure 4). Con A could upregulate the levels of p-AKT1, and NaHS preconditioning could further enhance AKT1 phosphorylation compared to the Con A group in hepatocytes, while p-AKT1 was markedly decreased after using PAG (Figure 5). This indicates that PtdIns3K-AKT1 may play an important role in the autophagy processes of Con A-induced hepatitis. Chang et al³⁰ reported that either 3-MA or small interfering RNA for BNIP3 and LC3, but neither Beclin-1 nor ATG5, partially inhibited Con A-induced cell death. Petiot et al³¹ also used RNA interference technology and found that Con A triggers autophagic cell death through increasing of BNIP3 protein and the generation of LC3-II, but other autophagy proteins such as Beclin-1 and ATG5 are not involved. This may explain why Beclin-1 was not affected in our experiment.

The mechanism underlying how H₂S inhibits apoptosis and autophagy through activation of the PtdIns3K-AKT1 signaling during Con A-induced acute hepatitis is not clear. We intend to probe this unknown mechanism in future studies.

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Disclosure

The authors report no conflicts of interest in this work.

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