

Garlic and Its Bioactive Derivatives as Host-Directed Therapies in Sepsis

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Abstract: Sepsis, a life-threatening dysregulated host response to infection, urgently requires novel adjunctive therapies due to the limitations of antibiotics and rising multidrug resistance. Garlic and its bioactive organosulfur compounds, such as allicin and diallyl sulfides, demonstrate significant therapeutic potential for sepsis management through diverse mechanisms. Their efficacy primarily stems from simultaneous immunomodulation and antioxidant activity. Garlic derivatives suppress the NF- κ B pathway to curtail excessive pro-inflammatory cytokine release and activate the Nrf2/HO-1 pathway to mitigate oxidative stress. Emerging research highlights that they help mitigate mitochondrial dysfunction by enhancing the PINK1/Parkin-mediated mitophagy pathway, thereby preserving cellular integrity. Beyond these host-directed effects, they exert direct, broad-spectrum antimicrobial activity against critical pathogens, including *Pseudomonas aeruginosa*, *Escherichia coli*, and *Plasmodium* species. Notably, they act synergistically with antibiotics and are effective against multidrug-resistant (MDR) strains. The pleiotropic actions of garlic compounds also confer protection against sepsis-induced multi-organ injury in major organs such as the lung, kidney, and liver. This combination of direct antimicrobial effects, immunomodulation, and organ protection positions garlic derivatives as a promising integrative therapeutic strategy. Given this broad therapeutic potential, further efforts will be required to translate these pleiotropic benefits into clinical practice and establish their efficacy through rigorous clinical trials for sepsis adjunctive therapy ultimately. Notably, current evidence predominantly relies on preclinical data, while critical clinical insights, such as the pharmacokinetics of allicin in humans, remain lacking, posing potential translational challenges.

Keywords: sepsis, garlic, nuclear factor kappa B, NF- κ B, diallyl disulfide, DADS, S-allyl cysteine, SAC

Introduction

Sepsis, a life-threatening organ dysfunction induced by a dysregulated host response to infection, remains a leading cause of mortality in intensive care units (ICU) worldwide.¹⁻³ Its pathophysiology is complex and multidimensional, involving concurrent hyperinflammation, immune suppression, and metabolic reprogramming, which collectively contribute to high mortality and long-term morbidity.^{2,4} While antibiotics are cornerstone therapies, the rising threat of antimicrobial resistance and the considerable economic burden of sepsis underscore the urgent need for novel therapeutic strategies.⁵⁻⁷ Notably, sepsis etiology is not confined to bacteria but also include other pathogens encompasses fungi and viruses. Viral pathogens, including dengue, Ebola, and COVID-19 (SARS-CoV-2), are responsible for a significant proportion, estimated at around 30%, of all sepsis cases.⁸ Given that sepsis is fundamentally characterized by a dysregulated host response,^{9,10} therapeutic strategies have consequently evolved beyond pathogen-targeted approaches to include modulation of the host's aberrant immune activity. This shift has established host-directed therapies as a central and expanding frontier in sepsis management.¹¹⁻¹⁴

Phytochemicals, with their pleiotropic mechanisms, have emerged as promising host-directed candidates for modulating the dysregulated immune response in sepsis. Beyond their primary pharmacological roles, compounds such as glycyrrhizin, silymarin, and curcumin have shown efficacy in attenuating inflammation and organ injury in preclinical models, largely through pathways involving HMGB1, nuclear factor kappa B (NF- κ B), and Nrf2.¹⁵⁻¹⁸ Among these,

garlic (*Allium sativum* L.) and its bioactive derivatives present a particularly compelling case, given their documented anti-inflammatory, antimicrobial, and antioxidant properties relevant to sepsis pathology.^{19–22} Garlic's therapeutic potential is exemplified by bioactive compounds including allicin, S-allyl cysteine (SAC), and sucrose methyl 3-formyl-4-methylpentanoate (SMFM). Experimental studies indicate these agents modulate key signaling cascades—such as NF- κ B, JAK-STAT, PI3K-Akt, and p38-MAPK—thereby reducing pro-inflammatory cytokines (eg, TNF- α , IL-6, IL-1 β) and mitigating oxidative stress and organ damage in models of sepsis.^{15,23–27}

Despite the promising preclinical evidence supporting the therapeutic potential of garlic and its bioactive derivatives, in mitigating sepsis-induced organ injuries through anti-inflammatory, antioxidant, and autophagy-modulating mechanisms, a significant translational gap persists. Given that the current evidence is predominantly derived from animal models, key translational barriers remain. These include a lack of human pharmacokinetic and safety data, undefined optimal dosing, and insufficient clinical efficacy validation. Furthermore, challenges related to the compound's bioavailability, stability, and the absence of specific regulatory frameworks for its therapeutic application must be resolved to assess its true translational relevance. The preclinical and clinical data was extracted into [Table 1](#).

Given the broad-spectrum biological activities of garlic and its derivatives, including anti-infection, antioxidant, and immunomodulatory effects, they represent a promising therapeutic avenue for sepsis. This review compiles and evaluates evidence from preclinical and clinical, in vitro, and in vivo studies to delineate their efficacy in sepsis management.

The Mechanism of Garlic and Its Derivatives on Sepsis

NF- κ B Pathway

The NF- κ B signaling pathway is mediated through two distinct mechanisms: the canonical and non-canonical pathways.^{48–50} TRAF6 serving as a crucial regulatory node in the canonical pathway activation. The canonical NF- κ B activation pathway is regulated by the I κ B kinase (IKK) complex, comprising catalytic subunits IKK α /IKK β and regulatory subunit NEMO. IKK activation triggers I κ B α phosphorylation, leading to polyubiquitination and proteasomal degradation. This releases NF- κ B dimers for nuclear translocation and target gene regulation. IKK β is the principal catalytic subunit phosphorylating I κ B α . TNF receptor-associated factor 6 (TRAF6), an E3 ubiquitin ligase, undergoes K63-linked autopolyubiquitination, hence activating its signaling activity. K63-polyubiquitinated TRAF6, as well as presumably TRAF6-modified NEMO, serves as scaffolds for recruiting the TAK1-TAB2 kinase complex. Activated TAK1 then phosphorylates and activates IKK β .

Evidence indicates that allicin protects against sepsis-induced lung injury by inhibiting the TLR4/MyD88/NF- κ B pathway. Allicin dose-dependently suppressed TRAF6, thereby inhibiting NF- κ B activation and the subsequent production of inflammatory and apoptotic mediators.⁴¹ In psoriatic models, allicin has been shown to directly inhibit the TRAF6/MAPK/NF- κ B and STAT3/NF- κ B cascades in IL-17-stimulated keratinocytes, thereby disrupting the inflammatory positive feedback loop. Consequently, allicin reduces the release of multiple inflammatory factors (eg, IL-17A/F, IL-22, IL-12, IL-20), chemokines (CXCL2, CXCL5, CCL20), and antimicrobial peptides (S100A1/9).³³ Similarly, in studies on bovine mammary epithelial cells, allicin mitigates lipopolysaccharide (LPS)-induced inflammation by suppressing the TLR4/NF- κ B pathway and NLRP3 inflammasome activation, leading to decreased levels of TNF- α , IL-1 β , IL-6, and IL-8. Consistent with these findings, allicin also alleviated LPS-induced mastitis in a mouse model.³²

In vitro research⁴⁰ demonstrated that aged black garlic extract (BG10) potently inhibited the production of nitric oxide (NO) and IL-6. This anti-inflammatory effect was achieved by suppressing the nuclear translocation of NF- κ B, thereby downregulating the expression of its downstream targets, inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 (COX-2), while concurrently increasing the intracellular levels of I κ B α . Moreover, SAC, as an active component of garlic, showed multiple therapeutic benefits, including anti-inflammatory and antioxidant properties.^{51–53} SAC has been demonstrated to ameliorate acute kidney injury by preserving mitochondrial integrity and attenuating oxidative stress, inflammation, and apoptosis. In 2019, MR et al found that pretreatment with SAC (100 mg/kg) in an LPS-induced AKI model led to a significant reduction in renal function markers (BUN and creatinine) and a concurrent increase in inflammatory mediators (NF- κ B, TLR4, IL-1 β , and IL-6), suggesting dose-dependent efficacy.³⁹

Table 1 Summary of Preclinical and Clinical Data

Author/Year/[Reference]	Types of Animal/Participants	Types of Model Inducer	Intervention Group	Laboratory Data	Clinical Data	Effect (Conclusion)
Li XJ et al /2024/ ²⁸	c57BL/6 mice/ HK2 cells	CLP/LPS	Allicin	Y		Positive
Zhou S et al /2024/ ²⁹	BALB/c female mice	<i>S. aureu</i>	GELNs	Y		Positive
Chen Y et al /2024/ ³⁰	Male C57BL/6j mice	LPS	SPRC	Y		Positive
Avendaño-Ortiz J et al /2023/ ³¹	Healthy volunteers and patients with sepsis	LPS	TASE		Y	Positive
Che HY et al /2023/ ³²	Pregnant ICR mice/ MAC-T cell line	LPS	Allicin	Y		Positive
Zhang L et al /2023/ ³³	Male and female BALB/c mice	IMQ	Allicin	Y		Positive
Tong YC et al /2023/ ²²	Cats	-	Garlic oil	Y		Positive
Chang Z et al /2022/ ³⁴	Sprague–Dawley female rats	<i>Escherichia coli</i>	Allicin	Y		Positive
Liu M et al /2022/ ³⁵	C57BL/6j male mice/ THP-1 cells	LPS	Alliin (CAS: 556–27-4)	Y		Positive
Redondo-Calvo FJ et al /2022/ ³⁶	Male Sprague–Dawley® rats	<i>Escherichia coli</i> ATCC 25922	TASE/ BGE	Y		Positive
Zhu L et al /2022/ ³⁷	Female C57BL/6 strain mice	<i>C. rodentium</i>	PTS, PTSO	Y		Positive
Torres KAM et al /2021/ ³⁸	<i>Streptococcus agalactiae</i>	-	CGE	Y		Positive
Zhang Y et al /2021/Uncited	Male SPF DBA/1 mice	Chicken collagen type II	Allicin	Y		Positive
Khajevand-Khazaei MR et al /2019/ ³⁹	Male C57BL/6 mice	LPS	SAC	Y		Positive
You BR et al /2019/ ⁴⁰	RAW264.7 cells	LPS	BG10	Y		Positive
Shen N et al /2019/ ⁴¹	Male SD rats	LPS	Allicin	Y		Positive
Shi L et al /2017/ Uncited	RAW264.7 cell/Male ICR mice	LPS/DSS	Alliin	Y		Positive
Anandasadagopan SK et al /2017/ ⁴²	Male Wistar albino rats	Cr (VI)	SAC	Y		Positive
Percival SS et al / 2016/ ⁴³	Healthy human participants / RCT	-	AGE		Y	
Lee SK et al /2015/ ²⁶	Male wild type ICR mice	CLP	SMFM	Y		Positive
Bayraktar O et al /2015/ ²⁷	Male Wistar rats	LPS	SAC	Y		Positive
Salama AA et al /2014/ ⁴⁴	<i>Babesia</i> parasites, <i>B. bovis</i> and <i>B. caballi</i>	BALB/c mice /In vitro	Allicin	Y		Positive
Kim MJ et al /2014/ Uncited	Male C57BL/6 mice /RAW 264.7	LPS	AGE/RGE	Y		Positive

(Continued)

Table 1 (Continued).

Author/Year/[Reference]	Types of Animal/Participants	Types of Model Inducer	Intervention Group	Laboratory Data	Clinical Data	Effect (Conclusion)
Watson CJ et al /2014/Uncited	Asymptomatic women who were culture-positive for <i>Candida</i> species (n=63)/RCT	-	Garlic		Y	Positive
Feng Y et al /2012/ ⁴⁵	Female BALB/c mice	<i>Plasmodium yoelii</i> 17XL	Allicin	Y		Positive
Park HJ et al /2012/Uncited	RAW 264.7 macrophages	LPS	ARGE	Y		Positive
Kuo CH et al /2011/Uncited	Male Wistar rats	Endotoxin	Garlic oil	Y		Positive
Yalindag-Ozturk N et al /2011/ ⁴⁶	A critically ill infant	-	Garlic extract		Y	Positive
Smyth AR et al /2010/Uncited	Patients (n=26)/RCT	-	Garlic		Y	Positive
Cutler RR et al /2009/ ²¹	Clinical isolates of Lancefield GBS	In vitro	Allicin extract and topical gel formulation	Y		Positive
Yu J et al /2007/ Uncited	Patients (n=30)	SGD	Garlicin		Y	Negative
Coppi A et al /2006/ ⁴⁷	Female Swiss Webster mice	<i>Plasmodium berghei</i> and <i>yoelii</i>	Allicin	Y		Positive
Stjernberg L et al /2000/Uncited	Military personnel /PRDBIS	-	Garlic		Y	Positive

Abbreviations: AGE, aged black garlic; BG10, black garlic extract BG10; BGE, ceftriaxone plus black garlic extract; CGE, crude garlic extract; CLP, cecal ligation and puncture; LPS, lipopolysaccharide; Cr (VI), hexavalent chromium; DAT, Diallyl trisulfide; DSS, dextran sulfate sodium; GBS, group B streptococci; GELNs, garlic- derived exosome-like nanovesicles; IMQ, imiquimod; MAC-T cell line, immortalized dairy cow mammary epithelial cell line; PTS, propyl propane thiosulfinate; PTSO, propyl propane thiosulfonate; RGE, raw garlic extract; SMFM, sucrose methyl 3-formyl-4-methylpentanoate; SAC, S-allyl cysteine; ARGE, aged red garlic extract; SGD, selected gut decontamination; TPA, 12- O-tetradecanoylphorbol- 13- acetate; TASE, thiosulfinate-enriched *Allium sativum* extract; *S. aureus*, *Staphylococcus aureus*.

Nrf2/HO-1 Pathway

Nrf2 is a key transcription factor that regulates cellular redox homeostasis by controlling antioxidant and cytoprotective gene expression. By binding to antioxidant response elements (AREs), Nrf2 activates a suite of Phase II detoxifying enzymes, thereby maintaining redox equilibrium and enhancing cellular defense mechanisms in pathological circumstances.⁵⁴ Studies have demonstrated that Nrf2 activation mitigates sepsis-induced organ dysfunction by attenuating excessive inflammatory responses and oxidative damage.⁵⁵ Notably, these protective effects are particularly evident in the amelioration of complications such as acute lung injury, acute kidney injury, and sepsis-associated myocardial depression.^{28,56,57}

Furthermore, evidence indicates that Nrf2 activation stimulates mitochondrial biogenesis, reduces apoptosis, and enhances tissue regeneration, underscoring its vital role in preserving organ integrity during sepsis.^{58–60} Activation of the Nrf2 signaling pathway has been shown to mitigate sepsis-induced inflammatory responses, oxidative stress, and organ dysfunction, thereby contributing to improved clinical outcomes. These findings highlight the therapeutic potential of targeting Nrf2 for developing novel strategies to reduce complications and enhance prognosis in sepsis. In a mouse model of sepsis-induced acute kidney injury (S-AKI), allicin was found to downregulate levels of serum creatinine, blood urea nitrogen, UALB, KIM-1, and NGAL, indicating improved renal function. Allicin also inhibited inflammatory and apoptotic processes, as reflected by reduced levels of inflammatory cytokines and apoptosis-related proteins.²⁸

PINK1/Parkin Pathway

Sepsis can lead to irreversible tissue damage and organ dysfunction, significantly contributing to the pathogenesis of type 5 cardiorenal syndrome (CRS-5).⁶¹ Studies have confirmed that activation of PINK1/Parkin-mediated mitophagy protects against sepsis-associated acute kidney injury by reducing inflammatory cytokine levels and restoring renal function.⁶² S-propargyl-cysteine (SPRC), a garlic-derived chemical, is a hydrogen sulfide (H₂S) donor that reduces macrophage inflammation and reactive oxygen species (ROS) production during sepsis and promotes an M1-to-M2 transition. This protective effect is mediated through the PINK1/Parkin pathway, which triggers adaptive autophagy in response to LPS challenge, thereby attenuating immune dysregulation.³⁰ Furthermore, allicin has been shown to mitigate LPS-induced macrophage pyroptosis by enhancing PINK1/Parkin-dependent mitophagy, leading to reduced mitochondrial ROS and suppression of NLRP3 inflammasome activation.³⁵

Other

The above mechanism is shown as [Figure 1](#). Garlic and its derivatives have been shown can ameliorate clinical symptoms in diverse pathogen infections.^{29,63,64} However, their precise roles and underlying mechanisms in sepsis remain incompletely understood and necessitate further investigation.

The Effect of Garlic and Its Derivatives on Pathogens

Anti-Protozoal Potential

Malaria

As early as 2006, allicin, a cysteine protease inhibitor present in garlic extracts, has been proven to possess ability to inhibit malaria infection. Malaria parasite proteases are essential for completing the parasite's life cycle and represent promising targets for novel antimalarial drugs. Allicin has been shown to inhibit the processing of the circumsporozoite protein (CSP) and block sporozoite invasion of host cells in both in vitro and in vivo models.⁴⁷ In 2012, a study further demonstrated that allicin confers protection against *Plasmodium yoelii* 17XL infection by enhancing both innate and adaptive immune responses.⁴⁵ The protective mechanism is primarily mediated through the upregulation of pro-inflammatory mediators-including IFN- γ , TNF, IL-12p70, and NO, as well as the activation of CD4⁺ T cells, dendritic cells (DCs), and macrophages. Additionally, allicin promotes the maturation of CD11c⁺ DCs, thereby strengthening the host's overall anti-malarial immunity.

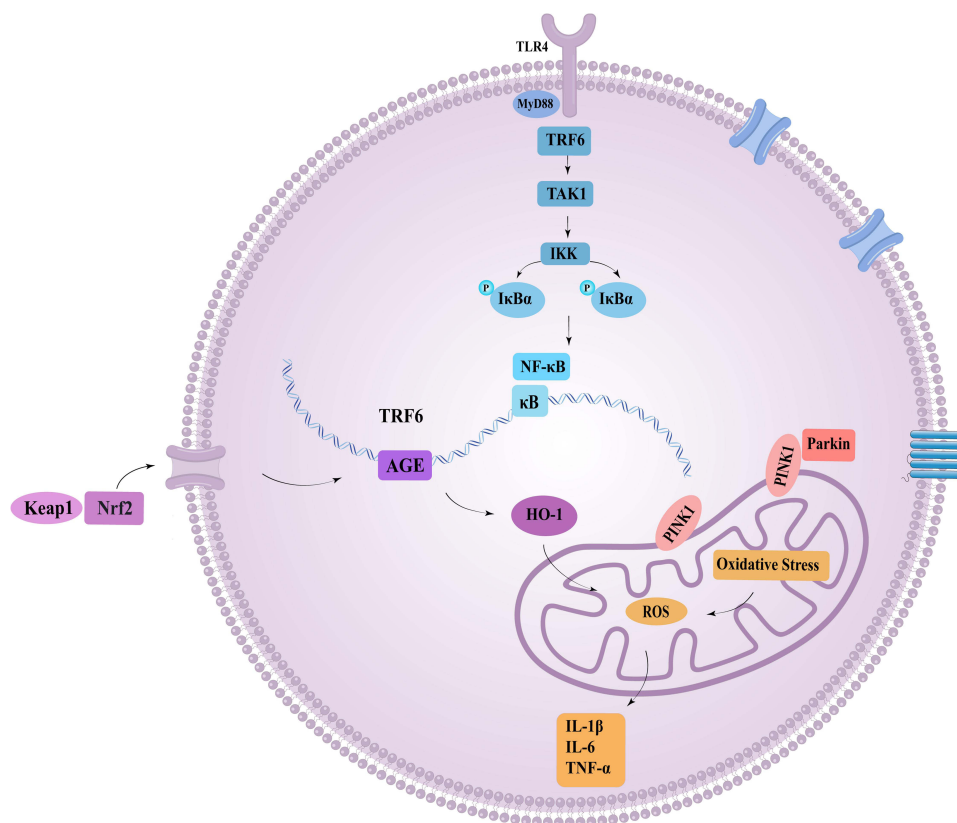


Figure 1 The role of garlic and its derivatives in the signaling pathway during sepsis development.

Babesia and Theileria equi Parasites

Further evidence suggests allicin demonstrated dose-dependent (30 mg/kg for 5 days) *in vitro* growth inhibition against several *Babesia* and *Theileria* parasite species.⁴⁴ Importantly, it significantly inhibited merozoite invasion into erythrocytes. In a mouse model of *Babesia microti* infection, allicin treatment significantly reduced parasitemia. Notably, combining allicin with the common antibabesial drug diminazene aceturate produced a synergistic inhibitory effect both *in vitro* and *in vivo*, suggesting its potential as a beneficial combination therapy for babesiosis.

Anti-Bacterial Potential

Pseudomonas aeruginosa

Pseudomonas aeruginosa (PA) is a prevalent opportunistic pathogen in healthcare-associated settings, particularly in ICU, where it is a leading cause of bloodstream and respiratory infections. This pathogen exhibits intrinsic resistance to multiple antibiotic classes and a demonstrated capacity to develop further resistance to novel agents.⁶⁵ The potential of garlic-based compounds as adjuvant therapy was initially reported in 2011, when a critically ill infant with severe multidrug-resistant PA pneumonia and bacteremia showed gradual clinical improvement following treatment with garlic and antibiotics.⁴⁶ A growing body of evidence now points to the underlying mechanisms. Emerging evidence indicates that diallyl disulfide (DADS), a key component of garlic oil, interferes with the quorum-sensing (QS) networks of *P. aeruginosa* PAO1. It suppresses the major genes (*las*, *rhl*, *pqs*) across all three QS systems, consequently reducing the production of associated virulence factors such as LasA/LasB proteases, LecA/LecB lectins, pyocyanin, and biofilm.⁶⁶ Similarly, diallyl sulfide (DAS), another garlic-derived compound, inhibits PA QS systems. This inhibition attenuates the production of host-damaging virulence factors and may enhance the synthesis of certain host-beneficial nutritional factors.⁶⁷

Group B Streptococcus

Group B *streptococcus* (GBS) remains a leading cause of invasive neonatal disease and associated morbidity. Maternal GBS colonization represents the primary risk factor for neonatal sepsis.⁶⁸ To mitigate the incidence of early-onset GBS disease, current clinical guidelines recommend universal culture-based screening during late gestation and intrapartum antibiotic prophylaxis (IAP).⁶⁹ Nevertheless, the widespread use of antibiotics contributes to the selection and dissemination of antimicrobial-resistant strains, highlighting the urgent need for alternative therapeutic strategies. In this context, recent investigations have identified specific garlic-derived compounds that exhibit inhibitory effects against clinical isolates of GBS, including γ -glutamyl-S-allyl-cysteine (fraction 18), γ -glutamyl-phenylalanine (fraction 20), and E- and Z-stereoisomers of ajoene (fraction 42).³⁸

Escherichia coli

Allicin has been demonstrated to significantly suppress the upregulation of the MALT1 and AKT/NF- κ B pathways and the expression of cytokines such as IL-6 and IL-1 β in *Escherichia coli*-induced urinary tract infections (UTI).³⁴ Separately, an evaluation of thirty clinical *E. coli* isolates revealed that the antibacterial activity of garlic extract (AGE) and manuka honey against extended-spectrum beta-lactamase (ESBL)-producing strains, as well as their interactive effects, are concentration-dependent.⁷⁰

Furthermore, extensive studies have demonstrated that garlic and its extracts exhibit broad-spectrum antimicrobial, antibiofilm, alongside selective cytotoxic properties.^{37,71–75} These bioactive compounds demonstrate efficacy against various pathogens including *Klebsiella pneumoniae*, *Clostridioides difficile* strains, *Bacillus anthracis*, *Citrobacter spp.*, and diverse fungi, particularly drug-resistant strains.

Anti-Viral Potential

Clinical and preclinical evidence supports the antiviral properties of garlic and its bioactive compounds.⁷⁶ A randomized controlled trial (RCT) demonstrated that supplementation with AGE significantly enhanced the proliferation of $\gamma\delta$ -T cells and natural killer (NK) cells in healthy subjects.⁷⁷ Although the incidence of colds and flu was unchanged, AGE consumption led to a marked reduction in illness severity, including fewer symptoms and days of impairment. A subsequent RCT confirmed these immunomodulatory findings, further positing that the improved cellular function with AGE may be associated with more efficient immune activity and potentially reduced inflammation.⁴³ Similarly, a long-acting garlic tablet formulation markedly decreased the incidence of acute respiratory infections in children compared to placebo.⁷⁸ Beyond these common viral illnesses, research indicates a broader antiviral potential. Organosulfur compounds from garlic show inhibitory activity against human immunodeficiency virus type 1 (HIV-1).⁷⁹ Furthermore, a selenium-enriched garlic powder demonstrated the ability to inhibit SARS-CoV-2 replication in vitro, potentially through mechanisms involving antioxidant activity and downregulation of host proteins crucial for viral entry.⁸⁰ These effects are attributed to multiple mechanisms, including direct inhibition of viral replication, blocking viral entry into host cells, and modulation of immune responses, positioning garlic as a promising candidate for non-specific prophylaxis against viral infections.

In summary, emerging evidence collectively supports the potential of garlic-derived bioactive compounds and their formulated extracts as promising plant-based candidates for adjunctive sepsis therapy. Their utility is attributed to multifaceted immunomodulatory, antioxidant, and antimicrobial effects against resistant pathogens.

Pleiotropic Effects of Garlic and Its Derivatives as Sepsis Adjunctive Therapy

Synergistic Effects in Enhancing Antibiotic Efficacy

Emerging evidence indicates that garlic and its bioactive constituents can enhance the efficacy of conventional antibiotics—such as gentamicin, ciprofloxacin, and ceftriaxone—through synergistic interactions.^{36,81,82} This synergy not only enhances bactericidal outcomes but may also allow for dose reduction, thereby potentially mitigating side effects and delaying the development of resistance. The underlying mechanisms involve multiple pathways that compromise

bacterial defenses, including inhibition of efflux pumps, impairment of biofilm formation, and increased membrane permeability, collectively promoting antibiotic penetration and intracellular accumulation. Notably, ajoene, a sulfur-rich compound derived from garlic, exhibits promising antitubercular properties.⁸² When combined with first-line anti-*Mycobacterium tuberculosis* (TB) drugs such as isoniazid and rifampicin, it significantly enhances anti-biofilm activity. The direct bactericidal activity of ajoene is mediated through the efflux pumps inhibition and subsequent augmentation of ROS.

Potential as a Novel Anti-Resistance Strategy

The global escalation of antimicrobial resistance (AMR) underscores the critical need for alternative therapeutic agents. *Allium sativum* (garlic) and its organosulfur compounds have re-emerged as promising candidates due to their broad-spectrum antimicrobial activity against multidrug-resistant (MDR) pathogens. Evidence indicates that garlic and its derivatives can effectively inhibit key virulence traits, including biofilm formation and QS systems, in diverse bacteria.^{83–86} As evidenced by studies on AGE exhibits strong efficacy against drug-resistant *C. albicans* and its associated polymicrobial biofilms, highlighting its potential as a safe and effective option for developing synergistic adjuvant therapies.⁸³

The mechanism of garlic and its derivatives demonstrates potential as a novel strategy to combat antimicrobial resistance. Research has identified garlic as a source of natural quorum-sensing inhibitors (QSIs), which can suppress fungal growth, prevent biofilm formation, and potentially augment host immune responses. These properties suggest its utility as a complementary therapeutic approach, particularly for infections in immunocompromised patients and those caused by drug-resistant fungal pathogens.⁸⁶

Immunomodulatory Role in Sepsis

The pathophysiology of sepsis is defined by a simultaneous and dysregulated host response, featuring concurrent hyperinflammation and profound immunosuppression.⁸⁷ This immunopathology typically evolves in a biphasic manner: an initial phase of systemic inflammatory response syndrome (SIRS), often manifesting as a “cytokine storm”, is frequently followed by a compensatory anti-inflammatory response syndrome (CARS). The CARS phase leads to protracted immunosuppression, characterized by features such as neutrophilia, an increase in regulatory T cell numbers, a monocytic endotoxin tolerance phenotype, elevated levels of IL-10 and TGF- β , and significant lymphocyte apoptosis. Consequently, an effective therapeutic regimen requires a dual-pronged strategy: mitigating the early cytokine storm and reversing the subsequent state of immune paralysis.^{88–90}

Immunomodulation has emerged as a pivotal therapeutic strategy for sepsis, shifting the paradigm from a sole focus on pathogen eradication to the active restoration of the host's dysregulated immune response. The dual immunomodulatory effect of thiosulfinate-enriched garlic extract (TASE) on LPS-stimulated monocytes, dependent on inflammatory status, has been demonstrated. TASE was shown to enhance cytokine production in low-inflammatory states while suppressing it in hyperinflammatory contexts, primarily through inhibition of the HIF-1 α pathway and downregulation of IRAK-M, VEGFA and PD-L1.³¹ Additionally, NaSH and SPRC significantly attenuated the inflammatory response by suppressing the secretion of pro-inflammatory cytokines, reducing ROS generation.³⁰ This potential was linked to a phenotypic shift in macrophages from pro-inflammatory M1 to anti-inflammatory M2, coupled with the induction of PINK1/Parkin-mediated mitophagy, which improved mitochondrial function and dampened the inflammatory cascade. Earlier-cited studies also have substantially deepened our understanding of the immunomodulatory mechanisms of garlic and its extracts in sepsis.^{48,91} Together, this evolving body of evidence underscores the potential of garlic and its bioactive derivatives as a multi-target agent for re-establishing immune homeostasis in sepsis.

The Role in Sepsis-Induced Multi-Organ Dysfunction

Sepsis, a life-threatening condition arising from a dysregulated host response to infection, can progress to multi-organ dysfunction syndrome (MODS), a process driven by uncontrolled systemic inflammation, oxidative stress, and immune paralysis.^{92–95} Given the limitations of conventional antibiotics and the lack of specific therapies targeting the host response, natural products with pleiotropic properties have garnered increasing attention. Garlic and its bioactive

constituents, particularly organosulfur compounds like allicin and SAC, have emerged as promising candidates for mitigating sepsis-associated organ injury via multiple interconnected mechanisms.

A prominent mechanism is the activation of the Nrf2 pathways. A 2024 study demonstrated that the combination of Selenomethionine (SeMet) and allicin synergistically attenuates oxidative stress and intestinal barrier injury in both cellular and murine models by activating the Nrf2- NQO1 axis and suppressing endoplasmic reticulum stress. This protective effect was abolished by an Nrf2 inhibitor, confirming the pathway's centrality.⁹⁶ Similarly, SAC has shown efficacy in reducing liver and lung injury in LPS- induced septic rats by lowering markers of oxidative stress and inhibiting DNA fragmentation.²⁷ The protective role of SAC against oxidative damage was also demonstrated in a model of chromium (VI)- induced hepatotoxicity, where it reduced NF- κ B and TNF- α expression.⁴² A 2024 study found that SPRC, protected against LPS-induced cardiorenal injury in mice.³⁰ These findings are consistent with reports on specialized pro-resolving mediators such as Resolvin D1, which protects against septic acute kidney injury by modulating inflammation and apoptosis,⁹⁷ underscoring a key therapeutic axis in sepsis management.

Limitations and Future Directions

Allicin is investigated as a potential adjunctive agent for sepsis management, primarily based on its multimodal mechanisms of action. Preclinical evidence suggests that its value stems not only from its direct anti-pathogen effect, but also from its ability to regulate host response dysregulation. A recent study (2023) found that garlic treatment in LPS-induced septic rats activates transient receptor potential vanilloid 1 (TRPV1) channels, upregulates calcitonin gene-related peptide (CGRP), and alleviates systemic oxidative stress and inflammatory damage.²⁴ Furthermore, garlic-derived exosome-like nanoparticles (GELNs) are shown to alleviate ulcerative colitis in mice. Orally administered GELNs ameliorate colitis symptoms, reduce proinflammatory cytokines, and protect the intestinal barrier. The mechanism involves the remodeling of gut microbiota, specifically through GELNs-enriched microRNAs that promote the growth of the beneficial bacterium *Bacteroides thetaiotaomicron*.⁹⁸

However, the current evidence landscape possesses significant gaps that limit translational confidence. The promising data are almost exclusively derived from in vitro studies and animal experiments. Critical questions regarding optimal dosing, pharmacokinetics in critically ill patients, formulation stability, and safety profiles in human sepsis remain largely unaddressed. There is a notable absence of large-scale, rigorous randomized controlled trials to substantiate its clinical efficacy and safety. Consequently, while the consolidated mechanistic narrative supports allicin's biological plausibility, its definitive role in sepsis therapy await validation through targeted clinical research designed to bridge these translational gaps.

Conclusion

In conclusion, contemporary research provides compelling preclinical evidence that garlic and its extracts can attenuate the cascade of events leading to MODS in sepsis. Their ability to simultaneously target inflammation, oxidative stress, and apoptosis positions them as potential multi-target adjunctive therapies. However, translating these findings into clinical practice requires rigorous human trials to validate efficacy, determine optimal dosing, and standardize bioactive compound formulations for sepsis management.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

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Disclosure

We declare that we have no financial and personal relationships with other people or organizations that can inappropriately influence our work. There is no professional or other personal interest of any nature or kind in any product, service and/or company that could be construed as influencing the position presented in, or the review of the manuscript.

References

- Schlapbach LJ, Watson RS, Sorce LR, et al; Society of critical care medicine pediatric sepsis definition task force. International consensus criteria for pediatric sepsis and septic shock. *JAMA*. 2024;331(8):665–674. doi:10.1001/jama.2024.0179
- Martin-Loeches I, Singer M, Leone M. Sepsis: key insights, future directions, and immediate goals. A review and expert opinion. *Intensive Care Med*. 2024;50(12):2043–2049. doi:10.1007/s00134-024-07694-z
- Borges A, Bento L. Organ crosstalk and dysfunction in sepsis. *Ann Intensive Care*. 2024;14(1):147. doi:10.1186/s13613-024-01377-0
- Wiersinga WJ, van der Poll T. Immunopathophysiology of human sepsis. *EBioMedicine*. 2022;86:104363. doi:10.1016/j.ebiom.2022.104363
- Gebremeskel F, Gebremedhin H, Mehari M. Magnitude of neonatal sepsis and factors associated with it among neonates admitted to the intensive care units of neonate in the primary hospital of Hawzen, Tigray, Ethiopia, 2020. *Glob Health Epidemiol Genom*. 2024;2024:7393056. doi:10.1155/2024/7393056
- Kingren MS, Starr ME, Saito H. Divergent sepsis pathophysiology in older adults. *Antioxid Redox Signal*. 2021;35(16):1358–1375. doi:10.1089/ars.2021.0056
- Liu Z, Ting Y, Li M, Tan Y, Long Y, Li Y. From immune dysregulation to organ dysfunction: understanding the enigma of Sepsis. *Front Microbiol*. 2024;15:1415274. doi:10.3389/fmicb.2024.1415274
- Gürtler GL, Schramm W, Seitz R. Viral sepsis - pathophysiology and disease manifestation. *Infection*. 2025;53(3):775–784. doi:10.1007/s15010-025-02486-z
- Bode C, Weis S, Sauer A, Wendel-Garcia P, David S. Targeting the host response in sepsis: current approaches and future evidence. *Crit Care*. 2023;27(1):478. doi:10.1186/s13054-023-04762-6
- Miller WD, Keskey R, Alverdy JC. Sepsis and the Microbiome: a Vicious Cycle. *J Infect Dis*. 2021;223(12 Suppl 2):S264–S269. doi:10.1093/infdis/jiaa682
- de la Fuente-Nunez C, Cesaro A, Hancock REW. Antibiotic failure: beyond antimicrobial resistance. *Drug Resist Updat*. 2023;71:101012. doi:10.1016/j.drug.2023.101012
- Moosazadeh Moghaddam M, Fazel P, Fallah A, et al. Host and pathogen-directed therapies against microbial infections using exosome- and antimicrobial peptide-derived stem cells with a special look at pulmonary infections and sepsis. *Stem Cell Rev Rep*. 2023;19(7):2166–2191. doi:10.1007/s12015-023-10594-2
- Kaufmann SHE, Dorhoi A, Hotchkiss RS, Bartenschlager R. Host-directed therapies for bacterial and viral infections. *Nat Rev Drug Discov*. 2018;17(1):35–56. doi:10.1038/nrd.2017.162
- Netea MG, van de Veerdonk FL, Giamarellos-Bourboulis EJ. Host-directed therapy in pandemic preparedness. *JAMA*. 2025;333(8):661–662. doi:10.1001/jama.2024.26152
- Alikiaii B, Bagherniya M, Askari G, Johnston TP, Sahebkar A. The role of phytochemicals in sepsis: a mechanistic and therapeutic perspective. *Biofactors*. 2021;47(1):19–40. doi:10.1002/biof.1694
- Qiang X, Peng Y, Wang Z, et al. Synthesis of glycyrrhizin analogues as HMGB1 inhibitors and their activity against sepsis in acute kidney injury. *Eur J Med Chem*. 2023;259:115696. doi:10.1016/j.ejmech.2023.115696
- Lim HR, Yi YS. Silymarin in *Silybum marianum* ameliorated acute lethal sepsis in mice by targeting caspase-11 noncanonical inflammasome in macrophages. *J Ethnopharmacol*. 2026;355(Pt B):120742. doi:10.1016/j.jep.2025.120742
- Qiu F, Zeng C, Liu Y, Pan H, Ke C. J147 ameliorates sepsis-induced depressive-like behaviors in mice by attenuating neuroinflammation through regulating the TLR4/NF- κ B signaling pathway. *J Mol Histol*. 2023;54(6):725–738. doi:10.1007/s10735-023-10147-4
- Orekhov AN, Tertov VV, Sobenin IA, Pivovarova EM. Direct anti-atherosclerosis-related effects of garlic. *Ann Med*. 1995;27(1):63–65. doi:10.3109/07853899509031938
- Stepień AE, Trojaniak J, Tabarkiewicz J. Anti-cancer and anti-inflammatory properties of black garlic. *Int J Mol Sci*. 2024;25(3):1801. doi:10.3390/ijms25031801
- Cutler RR, Odent M, Hajj-Ahmad H, et al. In vitro activity of an aqueous allicin extract and a novel allicin topical gel formulation against Lancefield group B streptococci. *J Antimicrob Chemother*. 2009;63(1):151–154. doi:10.1093/jac/dkn457
- Tong YC, Li PC, Yang Y, et al. Detection of antibiotic resistance in feline-origin ESBL *Escherichia coli* from different areas of China and the resistance elimination of garlic oil to cefquinome on ESBL *E. coli*. *Int J Mol Sci*. 2023;24(11):9627. doi:10.3390/ijms24119627
- Usmani J, Kausar H, Akbar S, et al. Molecular docking of bacterial protein modulators and pharmacotherapeutics of *Carica papaya* leaves as a promising therapy for sepsis: synchronising in silico and in vitro studies. *Molecules*. 2023;28(2):574. doi:10.3390/molecules28020574
- Torres-Narváez JC, Pérez-Torres I, Del Valle-Mondragón L, et al. Garlic prevents the oxidizing and inflammatory effects of sepsis induced by bacterial lipopolysaccharide at the systemic and aortic level in the rat. Role of trpv1. *Heliyon*. 2023;9(11):e21230. doi:10.1016/j.heliyon.2023.e21230
- Rauf A, Abu-Izneid T, Thiruvengadam M, et al. Garlic (*Allium sativum* L.): its chemistry, nutritional composition, toxicity, and anticancer properties. *Curr Top Med Chem*. 2022;22(11):957–972. doi:10.2174/1568026621666211105094939
- Lee SK, Park YJ, Ko MJ, et al. A novel natural compound from garlic (*Allium sativum* L.) with therapeutic effects against experimental polymicrobial sepsis. *Biochem Biophys Res Commun*. 2015;464(3):774–779. doi:10.1016/j.bbrc.2015.07.031
- Bayraktar O, Tekin N, Aydın O, Akyuz F, Musmul A, Burukoglu D. Effects of S-allyl cysteine on lung and liver tissue in a rat model of lipopolysaccharide-induced sepsis. *Naunyn-Schmiedeberg's Arch Pharmacol*. 2015;388(3):327–335. doi:10.1007/s00210-014-1076-z
- Li XJ, Liu T, Wang Y. Allicin ameliorates sepsis-induced acute kidney injury through Nrf2/HO-1 signaling pathway. *J Nat Med*. 2024;78(1):53–67. doi:10.1007/s11418-023-01745-3

29. Zhou S, Huang P, Cao Y, Hua X, Yang Y, Liu S. Garlic-derived exosome-like nanovesicles-based wound dressing for staphylococcus aureus infection visualization and treatment. *ACS Appl Bio Mater.* 2024;7(3):1888–1898. doi:10.1021/acsabm.3c01256
30. Chen Y, Cao W, Li B, et al. The potential role of hydrogen sulfide in regulating macrophage phenotypic changes via PINK1/parkin-mediated mitophagy in sepsis-related cardiorenal syndrome. *Immunopharmacol Immunotoxicol.* 2024;46(2):139–151. doi:10.1080/08923973.2023.2281901
31. Avendaño-Ortiz J, Redondo-Calvo FJ, Lozano-Rodríguez R, et al. Thiosulfinate-enriched allium sativum extract exhibits differential effects between healthy and sepsis patients: the implication of HIF-1 α . *Int J Mol Sci.* 2023;24(7):6234. doi:10.3390/ijms24076234
32. Che HY, Zhou CH, Lyu CC, et al. Allicin alleviated LPS-Induced mastitis via the TLR4/NF- κ B signaling pathway in bovine mammary epithelial cells. *Int J Mol Sci.* 2023;24(4):3805. doi:10.3390/ijms24043805
33. Zhang L, Ma X, Shi R, et al. Allicin ameliorates imiquimod-induced psoriasis-like skin inflammation via disturbing the interaction of keratinocytes with IL-17A. *Br J Pharmacol.* 2023;180(5):628–646. doi:10.1111/bph.15983
34. Chang Z, An L, He Z, et al. Allicin suppressed Escherichia coli-induced urinary tract infections by a novel MALT1/NF- κ B pathway. *Food Funct.* 2022;13(6):3495–3511. doi:10.1039/D1FO03853B
35. Liu M, Lu J, Yang S, Chen Y, Yu J, Guan S. Alliin alleviates LPS-induced pyroptosis via promoting mitophagy in THP-1 macrophages and mice. *Food Chem Toxicol.* 2022;160:112811. doi:10.1016/j.fct.2022.112811
36. Redondo-Calvo FJ, Bejarano-Ramírez N, Baladrón V, et al. Black garlic and thiosulfinate-enriched extracts as adjuvants to ceftriaxone treatment in a rat peritonitis model of sepsis. *Biomedicines.* 2022;10(12):3095. doi:10.3390/biomedicines10123095
37. Zhu L, Andersen-Civil AIS, Castro-Meija JL, et al. Garlic-derived metabolites exert antioxidant activity, modulate gut microbiota composition and limit citrobacter rodentium infection in mice. *Antioxidants.* 2022;11(10):2033. doi:10.3390/antiox11102033
38. Torres KAM, Lima SMRR, Torres LMB, Gamberini MT, Silva Junior PID. Garlic: an alternative treatment for group B streptococcus. *Microbiol Spectr.* 2021;9(3):e0017021. doi:10.1128/Spectrum.00170-21
39. Khajevand-Khazaei MR, Azimi S, Sedighnejad L, et al. S-allyl cysteine protects against lipopolysaccharide-induced acute kidney injury in the C57BL/6 mouse strain: involvement of oxidative stress and inflammation. *Int Immunopharmacol.* 2019;69:19–26. doi:10.1016/j.intimp.2019.01.026
40. You BR, Yoo JM, Baek SY, Kim MR. Anti-inflammatory effect of aged black garlic on 12-O-tetradecanoylphorbol-13-acetate-induced dermatitis in mice. *Nutr Res Pract.* 2019;13(3):189–195. doi:10.4162/nrp.2019.13.3.189
41. Shen N, Cheng A, Qiu M, Zang G. Allicin improves lung injury induced by sepsis via regulation of the toll-like receptor 4 (TLR4)/Myeloid differentiation primary response 88 (MYD88)/nuclear factor kappa B (NF- κ B) pathway. *Med Sci Monit.* 2019;25:2567–2576. doi:10.12659/MSM.914114
42. Anandasadagopan SK, Sundaramoorthy C, Pandurangan AK, Nagarajan V, Srinivasan K, Ganapasam S. S-Allyl cysteine alleviates inflammation by modulating the expression of NF- κ B during chromium (VI)-induced hepatotoxicity in rats. *Hum Exp Toxicol.* 2017;36(11):1186–1200. doi:10.1177/0960327116680275
43. Percival SS. Aged garlic extract modifies human immunity. *J Nutr.* 2016;146(2):433S–436S. doi:10.3945/jn.115.210427
44. Salama AA, AbouLaila M, Terkawi MA, et al. Inhibitory effect of allicin on the growth of *Babesia* and *Theileria equi* parasites. *Parasitol Res.* 2014;113(1):275–283. doi:10.1007/s00436-013-3654-2
45. Feng Y, Zhu X, Wang Q, et al. Allicin enhances host pro-inflammatory immune responses and protects against acute murine malaria infection. *Malar J.* 2012;11:268. doi:10.1186/1475-2875-11-268
46. Yalindag-Ozturk N, Ozdamar M, Cengiz P. Trial of garlic as an adjunct therapy for multidrug resistant *Pseudomonas aeruginosa* pneumonia in a critically ill infant. *J Altern Complement Med.* 2011;17(4):379–380. doi:10.1089/acm.2010.0445
47. Coppi A, Cabinian M, Mirelman D, Sinnis P. Antimalarial activity of allicin, a biologically active compound from garlic cloves. *Antimicrob Agents Chemother.* 2006;50(5):1731–1737. doi:10.1128/AAC.50.5.1731-1737.2006
48. Mulero MC, Huxford T, Ghosh G. NF- κ B, I κ B, and IKK: integral components of immune system signaling. *Adv Exp Med Biol.* 2019;1172:207–226.
49. Yamamoto M, Gohda J, Akiyama T, Inoue JI. TNF receptor-associated factor 6 (TRAF6) plays crucial roles in multiple biological systems through polyubiquitination-mediated NF- κ B activation. *Proc Jpn Acad Ser B Phys Biol Sci.* 2021;97(4):145–160. doi:10.2183/pjab.97.009
50. Sun SC. The non-canonical NF- κ B pathway in immunity and inflammation. *Nat Rev Immunol.* 2017;17(9):545–558. doi:10.1038/nri.2017.52
51. Johnson P, Loganathan C, Iruthayaraj A, Poomani K, Thayumanavan P. S-allyl cysteine as potent anti-gout drug: insight into the xanthine oxidase inhibition and anti-inflammatory activity. *Biochimie.* 2018;154:1–9. doi:10.1016/j.biochi.2018.07.015
52. Ruiz-Sánchez E, Pedraza-Chaverri J, Medina-Campos ON, Maldonado PD, Rojas P. S-allyl cysteine, a garlic compound, produces an antidepressant-like effect and exhibits antioxidant properties in mice. *Brain Sci.* 2020;10(9):592. doi:10.3390/brainsci10090592
53. Colín-González AL, Ali SF, Túnez I, Santamaría A. On the antioxidant, neuroprotective and anti-inflammatory properties of S-allyl cysteine: An update. *Neurochem Int.* 2015;89:83–91. doi:10.1016/j.neuint.2015.06.011
54. Liu H, Wang L, Zhou J. Nrf2 and its signaling pathways in sepsis and its complications: a comprehensive review of research progress. *Medicine.* 2025;104(16):e42132. doi:10.1097/MD.00000000000042132
55. Gunne S, Heinicke U, Parnham MJ, Laux V, Zacharowski K, von Knethen A. Nrf2-A molecular target for sepsis patients in critical care. *Biomolecules.* 2020;10(12):1688. doi:10.3390/biom10121688
56. Shen K, Wang X, Jia Y, et al. miR-125b-5p in adipose derived stem cells exosome alleviates pulmonary microvascular endothelial cells ferroptosis via Keap1/Nrf2/GPX4 in sepsis lung injury. *Redox Biol.* 2023;62:102655. doi:10.1016/j.redox.2023.102655
57. Lu SM, Yang B, Tan ZB, et al. TaoHe ChengQi decoction ameliorates sepsis-induced cardiac dysfunction through anti-ferroptosis via the Nrf2 pathway. *Phytomedicine.* 2024;129:155597. doi:10.1016/j.phymed.2024.155597
58. Li Y, Feng YF, Liu XT, et al. Songorine promotes cardiac mitochondrial biogenesis via Nrf2 induction during sepsis. *Redox Biol.* 2021;38:101771. doi:10.1016/j.redox.2020.101771
59. Xie K, Wang F, Yang Y, et al. Monotropein alleviates septic acute liver injury by restricting oxidative stress, inflammation, and apoptosis via the AKT (Ser473)/GSK3 β (Ser9)/Fyn/NRF2 pathway. *Int Immunopharmacol.* 2024;142(Pt B):113178. doi:10.1016/j.intimp.2024.113178
60. Duan H, Yang X, Cai S, et al. Nrf2 mitigates sepsis-associated encephalopathy-induced hippocampus ferroptosis via modulating mitochondrial dynamic homeostasis. *Int Immunopharmacol.* 2024;143(Pt 1):113331. doi:10.1016/j.intimp.2024.113331

61. Liu Y, Zheng C, Liu X, Zhang XQ, Zhang Y, Yu C. Early prediction of sepsis-induced cardiorenal syndrome: superiority of myoglobin over troponin I. *Ren Fail.* 2025;47(1):2542523. doi:10.1080/0886022X.2025.2542523
62. Hu C, Wu Z, Li T, et al. Dendrobine attenuates sepsis-associated acute kidney injury by promoting PINK1/PARKIN-mediated mitophagy. *Int Immunopharmacol.* 2025;157:114741. doi:10.1016/j.intimp.2025.114741
63. Giang TV, Hoa LNM, Hien TT, et al. Traditional vietnamese medicine containing garlic extract for patients with non-severe COVID-19: a Phase-II, double-blind, randomized controlled trial. *Cureus.* 2023;15(7):e42484. doi:10.7759/cureus.42484
64. Agostinelli E, Marzaro G, Gambari R, Finotti A. Potential applications of components of aged garlic extract in mitigating pro-inflammatory gene expression linked to human diseases (Review). *Exp Ther Med.* 2025;30(1):134. doi:10.3892/etm.2025.12884
65. Zakhour J, Sharara SL, Hindy JR, Haddad SF, Kanj SS. Antimicrobial treatment of pseudomonas aeruginosa severe sepsis. *Antibiotics.* 2022;11(10):1432. doi:10.3390/antibiotics11101432
66. Li WR, Ma YK, Xie XB, et al. Diallyl disulfide from garlic oil inhibits pseudomonas aeruginosa quorum sensing systems and corresponding virulence factors. *Front Microbiol.* 2019;9:3222. doi:10.3389/fmicb.2018.03222
67. Li WR, Zeng TH, Yao JW, et al. Diallyl sulfide from garlic suppresses quorum-sensing systems of Pseudomonas aeruginosa and enhances biosynthesis of three B vitamins through its thioether group. *Microb Biotechnol.* 2021;14(2):677–691. doi:10.1111/1751-7915.13729
68. Kaki S, Natemeier M, Clark M, Jurecki M. Beyond colonization: an atypical presentation of maternal group B streptococcus infection. *Cureus.* 2025;17(6):e85225. doi:10.7759/cureus.85225
69. Chan YTV, Lau SYF, Hui SYA, et al. HA-DH Liaison Committee, COC (O&G). Incidence of neonatal sepsis after universal antenatal culture-based screening of group B streptococcus and intrapartum antibiotics: a multicentre retrospective cohort study. *BJOG.* 2023;130(1):24–31. doi:10.1111/1471-0528.17279
70. Idris AR, Afegbua SL. Single and joint antibacterial activity of aqueous garlic extract and Manuka honey on extended-spectrum beta-lactamase-producing *Escherichia coli*. *Trans R Soc Trop Med Hyg.* 2017;111(10):472–478. doi:10.1093/trstmh/trx084
71. Sharma S, Raj K, Riyaz M, Singh DD. Antimicrobial studies on garlic lectin. *Probiotics Antimicrob Proteins.* 2023;15(6):1501–1512. doi:10.1007/s12602-022-10001-1
72. Kaur B, Kumar N, Chawla S, et al. A comparative study of in-vitro and in-silico anti-candidal activity and GC-MS profiles of snow mountain garlic vs. normal garlic. *J Appl Microbiol.* 2022;133(3):1308–1321. doi:10.1111/jam.15537
73. Sunil M, Kurangi B, Dodamani S, Khalil M, Chopra A. Antimicrobial, antibiofilm, cytotoxicity, and substantivity of aged garlic extract against oral bacteria: an in-vitro study. *BMC Complement Med Ther.* 2025;25(1):266. doi:10.1186/s12906-025-05012-8
74. Kaur R, Tiwari A, Manish M, Maurya IK, Bhatnagar R, Singh S. Common garlic (*Allium sativum* L.) has potent Anti-Bacillus anthracis activity. *J Ethnopharmacol.* 2021;264:113230. doi:10.1016/j.jep.2020.113230
75. Aleksić A, Stojanović-Radić Z, Harmanus C, Kuijper EJ, Stojanović P. In vitro anti-clostridial action and potential of the spice herbs essential oils to prevent biofilm formation of hypervirulent Clostridioides difficile strains isolated from hospitalized patients with CDI. *Anaerobe.* 2022;76:102604. doi:10.1016/j.anaerobe.2022.102604
76. Rouf R, Uddin SJ, Sarker DK, et al. Antiviral potential of garlic (*Allium sativum*) and its organosulfur compounds: a systematic update of pre-clinical and clinical data. *Trends Food Sci Technol.* 2020;104:219–234. doi:10.1016/j.tifs.2020.08.006
77. Nantz MP, Rowe CA, Muller CE, Creasy RA, Stanilka JM, Percival SS. Supplementation with aged garlic extract improves both NK and $\gamma\delta$ -T cell function and reduces the severity of cold and flu symptoms: a randomized, double-blind, placebo-controlled nutrition intervention. *Clin Nutr.* 2012;31(3):337–344. doi:10.1016/j.clnu.2011.11.019
78. Andrianova IV, Sobenin IA, Sereda EV, Borodina LI, Studenikin MI. Effect of long-acting garlic tablets “allicor” on the incidence of acute respiratory viral infections in children. *Ter Arkh.* 2003;75(3):53–56.
79. Gökalp F. The inhibition effect of garlic-derived compounds on human immunodeficiency virus type 1 and saquinavir. *J Biochem Mol Toxicol.* 2018;32(11):e22215. doi:10.1002/jbt.22215
80. Majeed M, Nagabhushanam K, Lawrence L, Prakasan P, Mundkur L. The mechanism of anti-viral activity of a novel, hydroponically selenium-enriched garlic powder (SelenoForce®) against SARS-CoV-2 virus. *Glob Adv Integr Med Health.* 2024;13:27536130241268100. doi:10.1177/27536130241268100
81. Magryś A, Olender A, Tchrzewska D. Antibacterial properties of *Allium sativum* L. against the most emerging multidrug-resistant bacteria and its synergy with antibiotics. *Arch Microbiol.* 2021;203(5):2257–2268. doi:10.1007/s00203-021-02248-z
82. Sarangi A, Das BS, Pahuja I, et al. Ajoene: a natural compound with enhanced antimycobacterial and antibiofilm properties mediated by efflux pump modulation and ROS generation against *M. Smegmatis*. *Arch Microbiol.* 2024;206(12):453. doi:10.1007/s00203-024-04189-9
83. Ashrit P, Sadanandan B, Shetty K, Vaniyampambath V. Polymicrobial biofilm dynamics of multidrug-resistant candida albicans and ampicillin-resistant *Escherichia coli* and antimicrobial inhibition by aqueous garlic extract. *Antibiotics.* 2022;11(5):573. doi:10.3390/antibiotics11050573
84. Alabdullatif M, Ramirez-Arcos S. Biofilm-associated accumulation-associated protein (Aap): a contributing factor to the predominant growth of *Staphylococcus epidermidis* in platelet concentrates. *Vox Sang.* 2019;114(1):28–37. doi:10.1111/vox.12729
85. Shahid M, Naureen I, Riaz M, Anjum F, Fatima H, Rafiq MA. Biofilm inhibition and antibacterial potential of different varieties of garlic (*Allium sativum*) against sinusitis isolates. *Dose Response.* 2021;19(4):15593258211050491. doi:10.1177/15593258211050491
86. Li N, Zhang J, Yu F, et al. Garlic-derived quorum sensing inhibitors: a novel strategy against fungal resistance. *Drug Des Devel Ther.* 2024;18:6413–6426. doi:10.2147/DDDT.S503302
87. van der Poll T, Shankar-Hari M, Wiersinga WJ. The immunology of sepsis. *Immunity.* 2021;54(11):2450–2464. doi:10.1016/j.immuni.2021.10.012
88. Hotchkiss RS, Karl IE. The pathophysiology and treatment of sepsis. *N Engl J Med.* 2003;348(2):138–150. doi:10.1056/NEJMr021333
89. Hotchkiss RS, Monneret G, Payen D. Sepsis-induced immunosuppression: from cellular dysfunctions to immunotherapy. *Nat Rev Immunol.* 2013;13(12):862–874. doi:10.1038/nri3552
90. van der Poll T, van de Veerdonk FL, Scicluna BP, Netea MG. The immunopathology of sepsis and potential therapeutic targets. *Nat Rev Immunol.* 2017;17(7):407–420. doi:10.1038/nri.2017.36
91. Samynathan R, Subramanian U, Venkidasamy B, Shariati MA, Chung IM, Thiruvengadam M. S-allylcysteine (SAC) exerts renoprotective effects via regulation of TGF- β 1/Smad3 pathway mediated matrix remodeling in chronic renal failure. *Curr Pharm Des.* 2022;28(8):661–670. doi:10.2174/1381612828666220401114301

92. Tang F, Zhao XL, Xu LY, Zhang JN, Ao H, Peng C. Endothelial dysfunction: pathophysiology and therapeutic targets for sepsis-induced multiple organ dysfunction syndrome. *Biomed Pharmacother.* 2024;178:117180. doi:10.1016/j.biopha.2024.117180
93. Malavika M, Sanju S, Poorna MR, et al. Role of myeloid derived suppressor cells in sepsis. *Int Immunopharmacol.* 2022;104:108452. doi:10.1016/j.intimp.2021.108452
94. Srdić T, Đurašević S, Lakić I, et al. From molecular mechanisms to clinical therapy: understanding sepsis-induced multiple organ dysfunction. *Int J Mol Sci.* 2024;25(14):7770. doi:10.3390/ijms25147770
95. Fan JB, Li QY, Feng XF, et al. The “cytokine storm” in infection and sepsis: win the battle but lose the war. *Mil Med Res.* 2026;12(1):95. doi:10.1186/s40779-025-00678-0
96. Liu Y, Lv X, Yuan H, Wang X, Huang J, Wang L. Selenomethionine and allicin synergistically mitigate intestinal oxidative injury by activating the Nrf2 pathway. *Toxics.* 2024;12(10):719. doi:10.3390/toxics12100719
97. Zhao YL, Zhang L, Yang YY, et al. Resolvin D1 protects lipopolysaccharide-induced acute kidney injury by down-regulating nuclear factor-kappa B signal and inhibiting apoptosis. *Chin Med J.* 2016;129(9):1100–1107. doi:10.4103/0366-6999.180517
98. Wang X, Liu Y, Dong X, et al. peu-MIR2916-p3-enriched garlic exosomes ameliorate murine colitis by reshaping gut microbiota, especially by boosting the anti-colitic *Bacteroides thetaiotaomicron*. *Pharmacol Res.* 2024;200:107071. doi:10.1016/j.phrs.2024.107071

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