


The Landscape of Potential Targeted Treatments and Chemotherapy Efficacy Biomarkers for TNBC

Yujuan Kang ^{*}, Yanqing Liu^{*}, Zhi Liang^{*}, Song Zhang^{*}, Guangdong Qiao

Department of Breast Surgery, The Affiliated Yantai Yuhuangding Hospital of Qingdao University, Yantai, People's Republic of China

^{*}These authors contributed equally to this work

Correspondence: Guangdong Qiao, Department of Breast Surgery, The Affiliated Yantai Yuhuangding Hospital of Qingdao University, Yantai, People's Republic of China, Email qiaoguangdong@ytyhdy2.wecom.work

Abstract: Triple negative breast cancer (TNBC) is characterized as an estrogen receptor (ER) negative, progesterone receptor (PR) negative, and human epidermal growth factor receptor (HER-2) negative disease with an enhanced systemic metastatic incidence, chemotherapeutic insensitivity, and drug resistance. The internal characteristics of TNBC are extraordinary complex and are accompanied by numerous gene mutations and constitutively activated signaling networks, which, in turn, augment TNBC malignancy. There is an urgency to uncover new and potent therapeutic targets for TNBC. Herein, we discussed the historical as well as newly discovered subtypes and properties of TNBC. We also presented a systematic outline of the aberrant gene expression and abnormal activated signaling networks in TNBC. Subsequently, we summarized the corresponding targeted therapeutic drugs, and reviewed the current targeted TNBC therapies launched in clinical trials, thereby providing a direction for future TNBC therapy. We also provided a detailed list of coding genes involved in chemo-sensitive and chemo-resistance of TNBC, which can potentially serve as indicators of chemotherapeutic efficacy. Finally, we discussed the novel and future potential treatments of TNBC. This review highlighted the new era of TNBC treatment in the coming years and provided the gene expression profile of TNBC patients who may benefit from chemotherapy.

Keywords: TNBC, immunotherapy, PI3K-AKT-mTOR, BRCA, EGFR, FGFR, PD-L1, CDK, chemo-sensitive, chemo-resistance

Introduction

About 2.1 million women suffer from breast cancer (BC) in a given year, and BC is the largest contributor of cancer-linked deaths among women.¹ BC is immunohistochemically (IHC) stratified into five sub-cohorts, namely, Luminal A, Luminal B, HER2, TNBC, and Normal-Like BC.² TNBC, marked with no ER, PR, and HER2 contents, make up approximately 15–20% of all BC patients.³ TNBC including six TNBC categories,^{4,5} namely, two Basal-Like (BL1 and BL2), a mesenchymal (M), a mesenchymal stem-like (MSL), an immunomodulatory (IM), and a luminal androgen receptor (LAR).⁵ Due to the poor response of TNBC patients to anti-HER2 monoclonal antibodies (mAb)-based and endocrine interventions, classic anticancer treatments, such as, surgery, radiotherapy, and chemotherapy remain the primary strategies for treating TNBC.⁶ Among TNBC patients, disease progression and recurrence are common within the initial 3–5 years following diagnosis, and distant metastases frequently occur in the brain and lung.⁷ The primary TNBC intervention is chemotherapy. Anthracyclines and Taxanes are the most effective drugs.⁸ Even though 30% of TNBC patients respond adequately to chemotherapy and have good prognosis, presently, there are no available interventions for chemo-resistant patients.⁹ Among an unselected TNBC population, BRCA mutation incidence is about 20%.¹⁰ Emerging evidences revealed that platinum agent introduction in metastatic and neoadjuvant settings achieves satisfactory outcomes in patients with BRCA mutations.^{11,12} Furthermore, others have successfully employed poly ADP-ribose polymerase (PARP) inhibitors to cure TNBCs with BRCA mutations.¹³ With growing advancements in medicinal technology, immunotherapeutic agents have emerged as drugs with robust responses and enhancements in patient overall survival (OS).¹⁴ TNBC exhibits excess tumor infiltrating lymphocytes (TILs),¹⁵ relative to other subtypes, along with markedly enhanced PD-L1 expression¹⁶ and OS strongly linked to the severity of immune cell

invasion.¹⁷ Hence, immune checkpoint blockade (ICB) therapies, such as, programmed death-1 (PD-1)- or programmed death-ligand 1 (PD-L1)-specific antibodies were shown to be highly efficacious in suppressing TNBC development.^{18,19}

In addition, dysregulated gene expression and signaling networks are the most prominent feature of TNBC.²⁰ Presently, several molecular targeted drugs are recognized as crucial for the treatment of TNBC. Notably, even though some TNBC patients are responsive to chemotherapy, there is still a considerable need for novel targeted therapeutic strategy discovery.²¹ In this review, we summarized the abnormal TNBC gene expression landscape, existing molecular targeted treatments, and chemotherapeutic efficacy biomarkers from the following four aspects: (1) The Molecular Subtypes of TNBC; (2) The Essential Characteristics and Potential Molecular Targeted Therapy of TNBC; (3) Coding Genes Involved in Chemo-Sensitive and Chemo-Resistance of TNBC; (4) Potential Novel Therapeutic Targets for TNBC.

Results

The Molecular Subtypes of TNBC

TNBC is a diverse disease that displays considerable differences in pathological characteristics, physiological activities, and genetic profiles.^{5,22} The established TNBC subtypes are BL1, BL2, M, MSL, IM, and LAR,⁵ and they are all unique in their features. The BL1 subtype is characterized by enhanced MYC, PI3K catalytic subunit α (PIK3CA), CDK6, AKT2, KRAS, FGFR1, IGF1R, CCNE1, and CDKN2A/B amplification, and augmented prevalence of heterozygous or homozygous deletion of DNA repair-associated genes (BRCA2, PTEN [phosphatase and tensin homolog], MDM2, RB1, and TP53). It is often combated with PARP inhibitors (PARPi) and genotoxic agents.²³ The BL2 subtype features aberrant activation of EGFR, MET, NGF, Wnt/ β -catenin, and IGF-1R pathway activations, and it is treated with mTOR inhibitors and growth factor inhibitors.²³ The M subtype features constitutive stimulation of the cell migration-associated axis, extracellular matrix-receptor association network, and differentiation axes [Wnt, anaplastic lymphoma kinase, transforming growth factor (TGF)- β],²³ and it is targeted with mTOR inhibitors and drugs against the epithelial-mesenchymal-transition (EMT).²⁴ The MSL subtype exhibited markedly reduced cell proliferation-associated genes, with simultaneous elevations in the stemness-associated genes (ABCA8, PROCR, ENG, ALDH1A1, PER1, ABCB1, TERT2IP, BCL2, BMP2, and THY), HOX genes (HOXA5, HOXA10, MEIS1, MEIS2, MEOX1, MEOX2, and MSX1), and mesenchymal stem cell (MSC) markers (BMP2, ENG, ITGAV, KDR, NGFR, NT5E, PDGFR, THY1, and VCAM1), and this is managed using PI3K inhibitors, v-src sarcoma viral oncogene homolog (Src) antagonists, or antiangiogenic drugs. A Src inhibitor (Dasatinib) may also be used to treat M and MSL TNBC.²³ Of note, the M and MSL categories are linked to an enhanced angiogenesis signature score.²⁵ The IM subtype is featured with abundant immune signaling networks, including, Th1/Th2, NK cell, B cell receptor, dendritic cell (DC), T cell receptor, interleukin (IL)-12, and IL-7 networks, and this category is treated with immune checkpoint inhibitor (ICI).²³ The LAR subtype have enhanced AR signaling, as is evidenced by IHC, which depicts upregulated AR, downstream genes, and auxiliary activators (DHCR24, ALCAM, FASN, FKBP5, APOD, PIP, SPDEF, and CLDN8).^{26,27} LAR tumors also contain PI3KCA (55%), AKT1 (13%) and CDH1 (13%) gene mutations.²⁵ LAR responds to anti-AR therapy.⁵

With advancements in medical technology, there have emerged certain modification to TNBC stratification. Burstein et al proposed a four subtypes classification of TNBC, including, LAR, mesenchymal (MES), basal-like immunosuppressed (BLIS), and basal-like immune activated (BLIA).²⁸ The potential corresponding targets were also identified as follows: (i) LAR: AR and MUC1, (ii) MES: growth factor receptors, (iii) BLIS: an immunosuppressing molecule (VTCN1), and (iv) BLIA: STAT-related molecules and cytokines.²⁸ Alternatively, Zhao et al identified AR, CD8, FOXC1, and DCLK1 as TNBC bioindicators, and stratified TNBCs into five categories, based on IHC: (a) IHC-based LAR (IHC-LAR; AR-positive [+]), (b) IHC-based immunomodulatory (IHC-IM; AR-negative [-], CD8+), (c) IHC-based basal-like immune-suppressed (IHC-BLIS; AR-, CD8-, FOXC1+), (d) IHC-based mesenchymal (IHC-MES; AR-, CD8-, FOXC1-, DCLK1+), and (e) IHC-based unclassifiable (AR-, CD8-, FOXC1-, DCLK1-). The IHC-LAR category is marked with a strongly activated HER2 network. The IHC-IM category has CD8⁺ T cell invasion into the tumor parenchyma. The IHC-BLIS category has an enhanced VEGF expression. The IHC-MES category is featured by active JAK/STAT3 axis.²⁹ He et al stratified TNBC according to immune profiles as follows: High (Immunity_H), Medium (Immunity_M), and Low (Immunity_L). Immunity_H displays a vast immune cell invasion and anti-tumor

immunologic activity. Among its immune signatures are excess cellular apoptosis, as well as calcium, MAPK, PI3K–Akt, and RAS networks. Alternatively, Immunity_L presents with a diminished immune profile and enhanced cellular cycle, Hippo axis, DNA replication, mismatch repair, cell adhesion molecule interaction, spliceosome, adherens junction activity, pyrimidine metabolism, glycosylphosphatidylinositol-anchor biosynthesis, and RNA polymerase networks. There are five transcription factor (TF) genes that are specific to the Immunity_H category, namely, CORO1A, STAT4, BCL11B, ZNF831, and EOMES, and two that are specific to the Immunity_L category, namely, IRF8 and SPI1.³⁰ These TNBC classifications facilitate an elevated level of understanding of TNBC (Table 1).

The Essential Characteristics and Potential Molecular Targeted Therapy of TNBC

TNBC prognosis is generally worse than non-TNBC.^{8,31,32} Although chemotherapy is the primary intervention for TNBC, it often produces rather disappointing outcomes. Due to the lack of precision therapeutic targets, TNBC is often aggressive in BC patients.^{33,34} TNBC is a heterogeneous tumor, accompanied by dysregulated gene expression and constitutively activated signaling networks. In this report, we provide a detailed list of gene mutations and aberrantly activated signaling networks in TNBC, as well as corresponding treatment methods for TNBC, based on the following aspects: (a) DNA Damage Response (DDR)-Related BRCA Loss-Function Mutation; (b) PI3K Signaling Network; (c) NOTCH Signaling Network;

Table 1 The Subgroups and Properties of TNBC

TNBC	ER (-)	PR (-)	HER2 (-)	Properties	Therapy	Ref
Lehmann et al	Basal-Like 1 (BL1)			(a) High expression of MYC, PIK3CA, CDK6, AKT2, KRAS, FGFR1, IGF1R, CCNE1, CDKN2A/B; (b) Augmented prevalence of heterozygous or homozygous deletion of DNA repair-associated genes (BRCA2, PTEN, MDM2, RB1, and TP53)	PARPi; Genotoxic Agents	[5, 23]
	Basal-Like 2 (BL2)			(a) Activation of EGFR, MET, NGF, Wnt/ β -catenin, IGF-1R pathways	mTOR Inhibitors; Growth Factor Inhibitors	[5, 23]
	Mesenchymal (M)			a. Activated cell migration-associated axis, extracellular matrix–receptor network, and differentiation axes (Wnt, anaplastic lymphoma kinase, TGF- β); (b) Higher signature score for angiogenesis	mTOR Inhibitors; Drugs against EMT; Src Inhibitor	[5, 23–25]
	Mesenchymal stem-like (MSL)			a. Reduced cell proliferation-related genes; b. Elevation in the stemness-related genes (ABCA8, PROCR, ENG, ALDH1, PER1, ABCB1, TERT2IP, BCL2, BMP2, and THY); c. Elevation in the HOX genes (HOXA5, HOXA10, MEIS1, MEIS2, MEOX1, MEOX2, and MSX1); d. Elevation in the MSC-specific markers (BMP2, ENG, ITGAV, KDR, NGFR, NTSE, PDGFR, THY1, and VCAM1); e. Higher signature score for angiogenesis;	PI3K Inhibitors; Src Inhibitor; Antiangiogenic drugs	[5, 23, 25]
	Immunomodulatory (IM)			a. Enriched immune related signaling networks, including, Th1/Th2, NK cell, B cell receptor, dendritic cell (DC), T cell receptor, interleukin (IL)-12, and IL-7 networks	ICI	[5, 23]
	Luminal Androgen Receptor (LAR)			a. Enhanced AR signaling, downstream genes, and auxiliary activators (DHCR24, ALCAM, FASN, FKBP5, APOD, PIP, SPDEF, and CLDN8) (b) Contain PI3KCA (55%), AKT1 (13%) and CDH1 (13%) gene mutations	AR Antagonist (Bicalutamide)	[5, 25–27]
Burstein et al	Luminal androgen receptor (LAR)			AR and MUC1		[28]
	Mesenchymal (MES)			Growth factor receptors		
	Basal-like immunosuppressed (BLIS)			Immunosuppressing molecule (VTCN1)		
	Basal-like immune activated (BLIA)			STAT-related molecules and cytokines		

(Continued)

Table 1 (Continued).

TNBC	ER (-)	PR (-)	HER2 (-)	Properties	Therapy	Ref
Zhao et al	IHC-based LAR (IHC-LAR; AR-positive [+])			Activation of HER2 network		[29]
	IHC-based immunomodulatory (IHC-IM; AR-negative [-], CD8+)			CD8 ⁺ T cell invasion into the tumor parenchyma		
	IHC-based basal-like immune-suppressed (IHC-BLIS; AR-, CD8-, FOXC1+)			Enhanced VEGF expression		
	IHC-based mesenchymal (IHC-MES; AR-, CD8-, FOXC1-, DCLK1+)			Active JAK/STAT3 axis		
	IHC-based unclassifiable (AR-, CD8-, FOXC1-, DCLK1-)			Unknown		
He et al	Immunity High (Immunity_H)			(a) Excess cellular apoptosis, as well as calcium, MAPK, PI3K-Akt, and RAS networks; (b) Express five TF genes (CORO1A, STAT4, BCL11B, ZNF831, and EOMES)		[30]
	Immunity Medium (Immunity_M)			In the middle		
	Immunity Low (Immunity_L)			(a) Diminished immune profile and enhanced cellular cycle, Hippo axis, DNA replication, mismatch repair, cell adhesion molecule interaction, spliceosome, adherens junction activity, pyrimidine metabolism, glycosylphosphatidylinositol-anchor biosynthesis, and RNA polymerase networks		

(d) CDK-Rb Signaling Network; (e) EGFR Activation; (f) Angiogenesis Signaling Network; (g) HER2-Low TNBC; (h) DDR Signaling Network (i) Immunotherapy, and (j) Androgen Receptor (AR) (Figure 1 and Table 2).

DDR-Related BRCA Loss-Function Mutation

Among the cancer features are DNA damage and impaired repair.⁹⁸ Prior reports demonstrated loss-of-function mutations in two essential BC susceptibility genes (BRCA1 and BRCA2) that modulate DDR, in 5% of BC patients.^{99–101} BRCA1 and BRCA2 proteins modulate DNA replication for stabilization and homologous recombination (HR).¹⁰² Based on one report, 70% of the inherited BRCA1 mutation-harboring BC patients and 16~23% of BRCA2 mutation-harboring BC patients are TNBC.¹⁰³ TNBC PARP enzymes are essential to the base excision repair mechanism, and PARP1 and PARP2 modulate DDR process.¹⁰⁴ BRCA1/2-mutated tumors tend to possess extra sensitivity toward PARP1 inhibition induced synthetic lethality. Thus, PARP1 inhibitors are promising for BRCA1/2 mutated patients.¹⁰⁵ The PARPi Olaparib is highly potent as a monotherapy in BC patients.¹⁰⁶ Presently, there are five BC-combating PARPi are under clinical validation: Olaparib, Rucaparib, Niraparib, Talazoparib, and Veliparib.³⁵ Additionally, emerging preclinical and clinical investigations demonstrated strong efficacy of platinum-based chemotherapy among TNBC patients.¹⁰⁷ Moreover, relevant clinical investigations were launched to accurately assess the therapeutic effect of PARPi in TNBC and BC (<https://clinicaltrials.gov/>) (Supplemental Table 1).

PI3K Signaling Network

Dysregulated phosphoinositide 3-kinase (PI3K)/AKT/mTOR is a hallmark of cancerous development.¹⁰⁵ This axis is essential for the receptor tyrosine kinase (RTK) signaling and tumor growth in BC.¹⁰⁸ In fact, PI3K pathway mutations form a special subgroup among TNBC patients.²⁰ In TNBC, the aberrant PI3K/AKT/mTOR axis activation is regulated by upstream regulator, activating PIK3CA mutations, PTEN loss of function or expression mutation, and proline-rich inositol poly-phosphatase (negative PI3K modulator) downregulation.^{109,110} Herein, we summarized the underlying mechanisms and PI3K signaling-targeted drugs associated with TNBC.

PI3K Inhibitors

In a previous report, it was revealed that pan-PI3K inhibitor (Buparlisib), combined with Buparlisib and Olaparib, diminishes

TNBC CELL

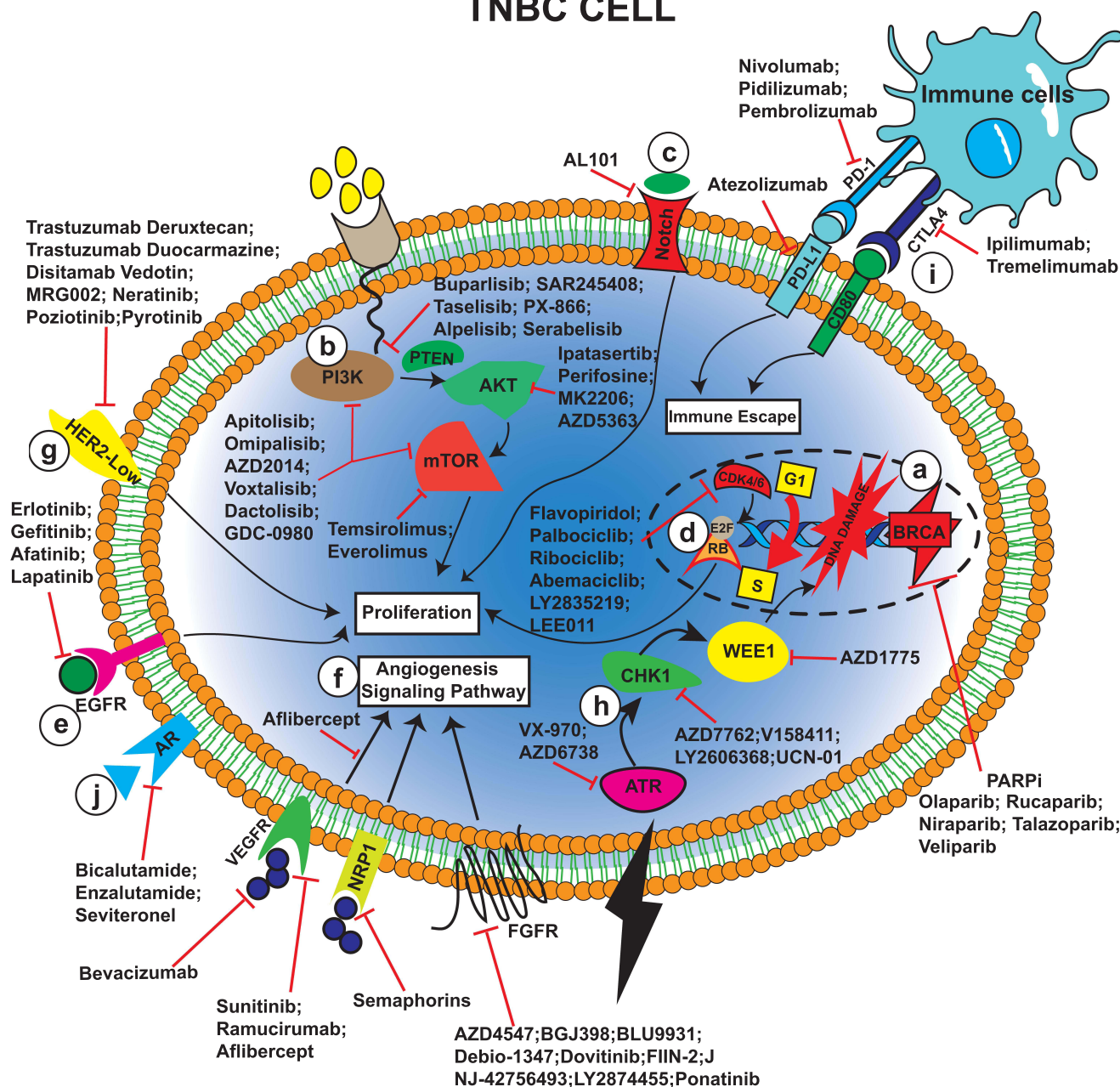


Figure 1 The abnormal gene and cell signal transduction profiles, as well as corresponding targeted TNBC therapy. (a) DNA Damage Response (DDR)-Related BRCA Loss-Function Mutation; (b) PI3K Signaling Network; (c) NOTCH Signaling Network; (d) CDK-Rb Signaling Network; (e) EGFR Activation; (f) Angiogenesis Signaling Network; (g) HER2-Low TNBC; (h) DDR Signaling Network (i) Immunotherapy, and (j) Androgen Receptor (AR).

BRCA1/2 content and encourages HR deficiency in BRCA1 WT TNBC patients.³⁶ In addition, the pan-PI3K inhibitors (SAR245408, Taselisib, and PX-866) are undergoing studied for HER2⁺ and HR⁺ BC.^{37–39} Alpelisib is an isoform-specific PI3K inhibitor under examination in clinical trials.⁴⁰ Moreover, there is ongoing development of Serabelisib, a robust and targeted PIK3CA inhibitor for management of solid tumors.⁴¹ The relevant clinical trials are listed in [Supplemental Table 1](#).

AKT Inhibitors

Ipatasertib inhibits AKT which is downstream of PI3K.⁴² Early AKT inhibitors, namely, Perifosine and MK2206, exhibited no clinical significance in Phase II trials,⁴³ however, novel AKT inhibitors appear to be highly promising in Phase I trials.^{44,45}

Table 2 The Molecular Targeted Medicines of TNBC

Drug Type	Drugs Name	Targeted Gene/Signaling Pathway	Ref
PARP Inhibitors	Olaparib; Rucaparib; Niraparib; Talazoparib; Veliparib	BRCA1/2	[35]
PI3K Inhibitors	Buparlisib; SAR245408; Taselisib; PX-866; Alpelisib; Serabelisib	PI3K	[36–41]
AKT Inhibitors	Ipatasertib; Perifosine; MK2206; AZD5363	AKT	[42–46]
mTOR Inhibitors	Temsirolimus; Everolimus	mTOR	[47]
Dual PI3K/AKT/mTOR Inhibitors	Apitolisib; Omipalisib; AZD2014; Voxtalisib; Dactolisib; GDC-0980	PI3K/AKT/mTOR	[48–53]
Notch Inhibitors	AL101	Notch	[54, 55]
CDK4/6 Inhibitors	Flavopiridol; Palbociclib; Ribociclib, Abemaciclib; LEE011; LY2835219	CDK-Rb Pathway	[56–58]
EGFR TKI	Erlotinib; Gefitinib	EGFR	[59]
Anti-EGFR mAb	Afatinib; Lapatinib	EGFR and ERBB2	[59]
VEGFR mAb	Bevacizumab; Sunitinib; Ramucirumab	VEGF-A and VEGFR	[60–62]
Anti-angiogenic	Semaphorins	NRP1 and NRP2	[63]
VEGF decoy receptor	Aflibercept	VEGFR	[64]
FGFR Inhibitors	AZD4547; BGJ398; BLU9931; Debio-1347; Dovitinib; FIIN-2; JNJ-42756493; LY2874455; Ponatinib	FGFR	[65–74]
Anti-HER2 ADC	Trastuzumab Deruxtecan; Trastuzumab Duocarmazine; Disitamab Vedotin; MRG002	HER2	[75]
Anti-HER2 TKIs	Neratinib; Pozotinib; Pyrotinib	HER2	[76]
ATR Inhibitors	VX-970; AZD6738	ATR-CHK1-WEE1	[35]
CHK1 Inhibitors	AZD7762; VI58411; LY2606368; UCN-01	ATR-CHK1-WEE1	[77–80]
WEE Inhibitors	AZD1775	ATR-CHK1-WEE1	[81]
CTLA-4 Inhibitors	Ipilimumab; Tremelimumab	CTLA-4	[82]
PD-1 Inhibitors	Pembrolizumab; Pidilizumab; Nivolumab	PD-1	[83]
PD-L1 Inhibitors	Atezolizumab	PD-L1	[83]
AR Inhibitors	Bicalutamide; Enzalutamide; Seviteronel	AR	[84]
Ferroptosis related drugs	1-methyl-3-isobutylxanthine; Rosiglitazone	Ferroptosis Pathway	[85]
Targeted ADC bioindicators	Sacituzumab Govitecan (IMMU-132); Ladiratuzumab vedotin (SGN-LIV1A); Glematumumab Vedotin (CDX-011); AVID100; U3-1402; CAB-ROR2-ADC; SAR566658	Trop-2; LIV1; EGFR; gpNMB; EGFR; HER3; ROR2; DS6	[86–89]
Ceritinib	Ceritinib	RTK-ACK1/FAK-AR	[90, 91]
Src Inhibitor	Dasatinib	Src	[92–96]
CDK12/CDK13 Inhibitor	SR-4835	CDK12/CDK13	[97]

Other AKT inhibitors (AZD5363) are under clinical development either as a mono- or chemo-combined therapy or as targeted agents for TNBC therapy.^{43,46} [Supplemental Table 1](#) summarizes the related clinical trials.

mTOR Inhibitors

A phase I investigation explored the synergistic effect of the chemotherapeutic doxorubicin, antiangiogenic bevacizumab, and mTOR inhibitors (Temsolimus, Everolimus) in TNBC patients.⁴⁷ Clinical trials related with them are provided [Supplemental Table 1](#).

Dual PI3K/AKT/mTOR Inhibitors

Apitolisib is a PI3K inhibitor that also targets mTORC.⁴⁸ Omipalisib is reported to have strong pan-PI3K and mTORC suppression abilities in a HER2⁺ model.⁴⁹ Preclinical information confirmed the mTORC1/2 inhibitor AZD2014-based inhibition of p-AKT, relative to everolimus in TNBC patient-based xerograph models.⁵⁰ Voxelalisib is another PI3K and mTOR inhibitor that is presently under investigation in clinical trials as a potential HR⁺ tumor treatment.⁵¹ Dual PI3K/AKT/mTOR inhibitors are often administered alongside other drugs. Dactolisib is a potential PI3K and mTOR inhibitor that will supplement MEK162 (a MEK inhibitor) therapy in treated TNBC.⁵² *De et al* also reported that the synergistic effect of GDC-0980 (pan-PI3K and mTOR inhibitor) and ABT888 (a PARPi) significantly suppressed tumor development in a PTEN-null TNBC model.⁵³ Clinical trials are summarized in [Supplemental Table 1](#).

NOTCH Signaling Network

The NOTCH axis is a robust bioindicator of TNBC. Notch gain of function mutations occurred in 10% of TNBC patients. The NOTCH signaling pathway utilizes four distinct receptor types (Notch-1 through Notch-4) which interact with five specific ligand molecules (Delta-like 1, 3, and 4 along with Jagged proteins 1 and 2).⁵⁴ AL101 is currently being developed as a pan-Notch inhibitor of the NOTCH network.⁵⁵ Relevant clinical trials are summarized in [Supplemental Table 1](#).

CDK-Rb Signaling Network

The cyclin-dependent kinases 4 and 6 (CDK4/6)–RB1 axis modulates the G1 phase restriction point of the cell cycle, and cancers often impair this signal transduction to initiate constitutively active cell proliferation.^{111,112} Mutations and aberrations impacting the Rb axis are another hallmark of TNBC etiology.^{4,113} Pharmaceuticals-based CDK4/6 inhibition (CDK4/6i) generally abrogates RB phosphorylation, thereby affecting the G1/S checkpoint.¹¹⁴ More recently, it was revealed that RB-harboring TNBC are highly responsive to CDK4/6 inhibition, thereby providing ample evidence of the strong potential of CDK4/6i as a successful TNBC therapy.^{21,115–117} Flavopiridol became the first broad-spectrum CDK inhibitor examined in clinical trials.⁵⁶ Palbociclib, a specific CDK4/6 inhibitor, has been approved by the FDA for the treatment of ER-positive advanced BC^{57,58} and is expected to provide new treatment strategies for TNBC patients, especially the LAR subtype of TNBC.²¹ At present, there are three FDA-approved CDK4/6i: Palbociclib, Ribociclib, and Abemaciclib for BC patients. Two other CDK4/6i (LEE011 and LY2835219) are under examination in dose-screening Phase I investigations. Relevant clinical trials are presented in [Supplemental Table 1](#).

EGFR Activation

Unlike non-TNBC,^{118,119} TNBCs exhibited close to 78% EGFR overexpression,^{118,120–123} indicating the possibility of using EGFR as a target for TNBC therapy. Among the reasons for EGFR overexpression in TNBC is EGFR gene amplification, which is highly variable and present in close to 24% TNBC patients,^{124–126} and EGFR gene mutation.^{124,126} Given these evidences, EGFR tyrosine kinase inhibitors (TKI) and anti-EGFR mAb have been examined alone and alongside chemotherapy in the early^{127,128} and metastatic stage of disease.^{129,130} TKIs like Erlotinib and Gefitinib target EGFR; anti-EGFR mAb, which include Afatinib and Lapatinib target EGFR and ERBB2⁵⁹ in TNBC therapy. The available clinical information involving Erlotinib, Afatinib, and Lapatinib usage are presented in [Supplemental Table 1](#).

Angiogenesis Signaling Network

Angiogenesis critically modulates tumor growth.¹³¹ Vascular endothelial growth factor (VEGF) is the primary angiogenic modulator, and it is synthesized during BC.¹³¹ Angiogenic homeostasis is maintained via an intricate

balance of pro- angiogenic agents (VEGF, FGF and PDGF, HIFs) and anti-angiogenic agents (thrombospondin, angiostatin and endostatin).¹³² VEGF are expressed in 30–60% of TNBC,¹³³ thereby raising the possibility of an anti-VEGF based therapy for TNBC. Herein, we summarized relevant and potential therapies targeting angiogenesis in TNBC.

VEGFR

TNBC patients exhibit markedly elevated intra-tumoral VEGF rates, relative to non-TNBC patients.¹³⁴ VEGF family includes five ligands (VEGFA, VEGFB, VEGFC, VEGFD, and placental growth factor), five receptors (VEGFR1, VEGFR2, VEGFR3, NRP1 and NRP2).¹³² Since angiogenesis is critical for tumor cell proliferation, Bevacizumab, a VEGF-A-binding mAb, has gained much attention as a potential TNBC therapeutic target.⁶⁰ Under a neoadjuvant situation, bevacizumab is reported to strongly enhance pathological complete response (pCR).¹³⁵ In addition, the VEGF axis-targeting interaction efficacy can potentially be modified using other ligands, such as, Semaphorins, that associate with both NRP1 and NRP2, suggesting that Semaphorins a potential may be new anti-angiogenic therapeutic target.⁶³ Small molecule TKIs have been developed to prohibit angiogenesis through binding receptors.¹³⁶ Sunitinib represents one such agent, binding multiple moieties including VEGFR1, VEGFR2, PDGFR and KIT.⁶¹ IMC-1121 (Ramucirumab) is a fully humanized mAb of VEGFR, and it is reported to demonstrate strong efficacy against BC.⁶² VEGF-Trap (Aflibercept) is a decoy VEGFR that potentially sequesters VEGF to disrupt its association with its true receptor.⁶⁴ Ongoing clinical investigations evaluating the effectiveness of the aforementioned drugs are summarized in [Supplemental Table 1](#).

Fibroblast Growth Factor Receptors (FGFR)

FGFRs belong to the trans-membranal receptor family that are crucial for angiogenic regulation.⁵⁵ Tumor-based fibroblasts are ubiquitous present in BC, whereby they generate a myriad of growth factors, namely, basic fibroblast growth factor (bFGF).¹³⁷ The bFGF-FGFR axis activation is a hallmark of carcinogenesis.¹³⁸ FGFR1, FGFR2, FGFR3, and FGFR4 make up the FGFR family of RTKs.^{139–141} FGFR1 over-expression is typically found in 18% of TNBC patients, with the FGFR1 gene amplification accounting for 33% of cases,¹⁴² and FGFR2 amplification in <5% of cases.¹⁴³ Clinical trials have reported favorable outcomes for both multi-targeted TKIs¹⁴⁴ and FGFR-specific TKIs.¹⁴⁵ Unlike TKIs, anti-FGFR isoform antibodies¹⁴⁶ and FGF ligand inhibitors were also shown to have strong anti-TNBC efficacies.¹⁴⁷ AZD4547,^{65,66} BGJ398 (Infigratinib),⁶⁷ BLU9931,⁶⁸ Debio-1347,⁶⁹ Dovitinib (TKI258),⁷⁰ FIIN-2,⁷¹ JNJ-42756493,⁷² LY2874455⁷³ and Ponatinib⁷⁴ are small but robust molecular compounds that suppress FGFRs at reduced concentrations. Relevant clinical trials were hoped to develop in the future.

HER2-Low TNBC

A new stratification was proposed for BC with reduced HER2 content and undetectable gene amplification (IHC) 1+ or IHC 2+ with negative in situ hybridization (ISH). This is termed as HER2-low.⁵⁵ Anti-HER2 agent technology has also been transformed from the classical binary stratification to the “HER2-low” BC classification.⁷⁵ Antibody-drug conjugates (ADCs) are a relatively new anticancer therapy that facilitates the specific deployment of a robust cytotoxic “payload” to cancer cells via targeted association between antibody and cancer cell surface molecule.¹⁴⁸ At present, multiple ADCs are being examined for drug efficacy and safety.¹⁴⁹ Till now, four anti-HER2 ADCs were reported to produce strong anti-cancer activity among HER2-low BC patients: Trastuzumab Deruxtecan (DXd), Trastuzumab Duocarmazine, Disitamab Vedotin, and MRG002.⁷⁵ The anti-HER2 ADC Dxd (DS-8201a) demonstrated satisfactory outcome in a Phase 1 investigation involving advanced HER2-low solid tumors, which including BC (NCT02564900). Trastuzumab Duocarmazine (SYD985) comprises an anti-HER2 antibody attached to a DNA-alkylating duocarmycin payload via a cleavable linker.⁷⁵ Disitamab Vedotin (RC48-ADC) is an ADC made up of a new anti-HER2 humanized antibody, hertuzumab, which is attached to a microtubular inhibitor monomethyl auristatin E (MMAE) payload via a cleavable linker.⁷⁵ MRG002 is a newly designed HER2-targeted ADC made up of modified trastuzumab linked with a cleavable linker to a MMAE payload.⁷⁵ Several anti-HER2 vaccines are currently being examined among HER2-low BC population, among them, some displayed good outcome among TNBC patients.^{149,150} Anti-HER2 TKIs, Neratinib,⁷⁶

and other anti-HER2 TKIs, namely, Poziotinib and Pyrotinib, are also under investigation as potential anti-cancer agents. Relevant clinical trials are provided in [Supplemental Table 1](#).

DDR Signaling Network

TNBC patients exhibited a high frequency of distinctive DDR alterations.^{4,151–153} The DDR axis, which modulates damaged DNA repair and genomic stability maintenance, is, in turn, regulated by DNA damage sensors, cell cycle checkpoints, diverse DNA repair signals and cellular apoptosis.^{154,155} The ATR-CHK1-WEE1 axis is most targeted among all the DDR networks. VX-970 (VE-822 or Berzosertib) is a robust and specific ATR inhibitor, and the first ATR inhibitor to be examined in clinical trials. In another clinical trial, AZD6738 (Ceralasertib), a targeted ATR inhibitor, was employed as a second ATR inhibitor.³⁵ The CHK1 inhibitor AZD7762 is an ATP-competitive dual CHK1/2 inhibitor, which exerts its primary antitumor property via CHK1 suppression.⁷⁷ TNBC sensitivity to AZD7762 is substantially augmented by RB deficiency *in vitro* and *in xenografts* likely due to the non-existent RB enhanced DNA replication stress.⁷⁸ V158411, another ATP-competitive CHK1/2 inhibitor, demonstrated marked cytotoxicity in TNBC versus HR⁺ BC. LY2606368 (Prexasertib) is a second-generation CHK1/2 dual inhibitor that selectively targets CHK1. LY2606368 monotherapy was proven to be safe and efficacious among high-grade ovarian carcinoma patients, who have numerous commonalities with TNBC patients.⁷⁹ UCN-01 was the first proposed CHK1 inhibitor, and its synergistic effect with gemcitabine inhibits TNBC as UCN-01 drives the gemcitabine-induced S-phase cell arrest through G2/M checkpoints, thereby augmenting DNA damage, and accelerating cellular death.⁸⁰ Despite the recent emergence of several small-molecule WEE1 inhibitors, AZD1775 (Adavosertib) is the most studied, and only molecule to enter clinical trials till date.⁸¹ Relevant clinical trials are provided in [Supplemental Table 1](#).

Immunotherapy

TNBC have strongly upregulated PD-1/PD-L1 levels. Consequently, scientists examined the efficacies of these checkpoint inhibitors in mono- and combined therapies, with modest outcome.¹⁵⁶ CTLA-4, PD-1, and PD-L1 inhibitors are the extensively examined immune checkpoint blockers (ICB). CTLA-4/CD152, found on CD8⁺ and CD4⁺ T cells,¹⁵⁷ critically activates or inhibits T cell-induced immune response. Among the current CTLA-4 inhibitors are Ipilimumab and Tremelimumab.⁸² PD-1/CD279 functions as a single-chain glycoprotein¹⁵⁸ predominantly expressed on the membranes of various immune cells including T lymphocytes, B lymphocytes, NK cells, monocytic cells, DCs and tumor tissues.¹⁵⁹ PD-1 is a major activated T cell immune checkpoint receptor, and it strongly modulates immune suppression. The current PD-1 inhibitors are Pembrolizumab, Pidilizumab, and Nivolumab. Among them, nivolumab is a fully humanized IgG4 mAb⁸³ with remarkable clinical outcome. PD-L1 inhibits and inactivates T-cell via its association with the PD-1 and B7-1 receptors on stimulated T cells.¹⁶⁰ Atezolizumab, a humanized anti-PD-L1 IgG-1 mAb,⁸³ also enhances median OS and ORR among lung cancer patients.¹⁶¹ Relevant clinical trials are summarized in [Supplemental Table 1](#).

Androgen Receptor (AR)

Approximately, 25–75% TNBC patients have enhanced AR expression level, among whom, the strongest expression is seen among patients with the LAR subtype.¹⁶² Using both *in vitro* and *in vivo* investigations, AR⁺ TNBCs were shown to respond well to AR antagonist therapy.^{163,164} Currently, three anti-AR drugs, Bicalutamide, Enzalutamide, and Seviteronel, are under clinical investigation.⁸⁴ All available anti-androgen therapy-based clinical information are provided in [Supplemental Table 1](#).

Coding Genes Involved in Chemo-Sensitive and Chemo-Resistance of TNBC

Unlike non-TNBC, TNBC are generally larger, of enhanced grade, and node positive. Hence, for TNBC, chemotherapy is the best treatment option.⁷ Among the recommended interventions are taxanes and anthracyclines, however, platinum-based regimens could be employed in neo-adjuvant settings.¹⁶⁵ Taxanes are the primary constituents of neoadjuvant chemotherapy. Mechanically, they suppress microtubule depolymerization to repress cancer cell proliferation.¹⁶⁶ TNBC patients are typically more responsive to neoadjuvant therapy than ER⁺ BC. They also have considerably worse OS, relative to non-TNBC tumor patients despite having better pCR rates; thus, this is known as the “triple negative paradox”.¹⁶⁷ Given that chemotherapy is the main form of TNBC treatment, chemoresistance is the primary contributor of TNBC recurrence and metastasis. Currently,

there is a lack of clinically employed bioindicators for sensitive or resistant tumor classification. Moreover, there are no treatment approaches geared toward the chemo-refractory cases.¹⁶⁸ In this report, we first discussed the TNBC-associated chemotherapy-sensitivity and chemotherapy-resistance coding genes. The chemo-sensitive genes are strongly diminished in chemo-resistant TNBCs. It is speculated that future TNBC treatment may utilize overexpression of the chemo-sensitive genes (Figure 2 and Table 3). Alternatively, the chemo-resistant genes are ubiquitously expressed in chemo-resistant TNBC. Thus, future research may entail suppression of the aforementioned genes in TNBC (Figure 3 and Table 4). Due to the strongly altered states of these genes, they are served as strong candidates for estimating the chemotherapeutic response of TNBC patients, which, in turn, can assist in enhancing patient care and prognosis. However, whether these genes can become indicators of drug resistance in patients still has a long way to go.

Chemotherapy-Sensitivity Genes of TNBC

CLDN1

Approximately 77% TNBC display little to no claudin-1 (CLDN1) expression, and this deficiency is strongly associated with worse outcome. One study reported that CLDN1 enhances TNBC cell line sensitivity to chemotherapeutic agents (paclitaxel, abbreviated as PTX) for BC therapy. These evidences confirm CLDN1 as a chemotherapeutic response indicator for TNBC patients.¹⁶⁹

DAB2IP

DAB2IP abrogates TNBC chemoresistance (Docetaxel) by suppressing the RAC1-triggered nuclear β -catenin transfer. Decitabine treatment re-expresses DAB2IP by preventing DNA methylation, thereby enhancing its potential in the management of TNBC.¹⁷⁰

Thioredoxin-Interacting Protein (TXNIP)

TXNIP originates from the α -arrestin family of proteins,¹⁷¹ which tightly modulate various physiological activities via interaction with other proteins.¹⁷² TXNIP accelerates cellular apoptosis, and inhibits chemo-resistant TNBC cellular growth both in vitro and in vivo by enhancing ROS-based DNA damage. The small molecular c-Myc inhibitor 10058-F4 enhances TXNIP content and augments intracellular ROS production to downregulate DOX-stimulated chemoresistance in TNBC. Combination with DOX further enhances the chemotherapeutic cytotoxic properties of 10058-F4.¹⁷³

C/EBP- β LIP

Doxorubicin effectiveness depends on the number of drug efflux transporters like P-glycoprotein (Pgp) present within tumor cells.¹⁷⁴ One report revealed that blocking C/EBP- β LIP degradation strongly enhances nitric oxide content, while reducing Pgp expression and activity, which, in turn, restores ER stress-dependent apoptosis and Doxorubicin-induced immunogenic cell apoptosis. This, in turn, rescues the anthracycline therapeutic activity in Pgp-positive TNBC.¹⁷⁵

cAMP Response Element Binding Protein 3-Like 1 (CREB3L1)

Researchers reported high CREB3L1 levels in chemotherapeutic sensitive cancers than chemotherapeutic resistant cancers. Similarly, high CREB3L1-harboring TNBC responds better to doxorubicin-based chemotherapy than low CREB3L1-harboring TNBC.¹⁷⁶

Atractylenolide-I (ATL-I)

ATL-I exerts strong anti-tumor influences against BC.¹⁷⁷ In particular, ATL-I reduces the CTGF transcript levels in TNBC, which reduces TNBC cell migration, diminishes the fibroblast to cancer-associated fibroblasts (CAFs) transformation, and enhances TNBC cell sensitivity to PTX.¹⁷⁸

SLFN12

SLFN12 is a newly discovered protein, with strong association with TNBC patient prognosis.¹⁷⁹ SLFN12 was shown to enhance TNBC cell sensitivity to radiation and cytotoxic drugs via reduction of CHK1 and CHK2 phosphorylation, which may be responsible for the OS differences between high and low SLFN12 tumor patients.¹⁸⁰

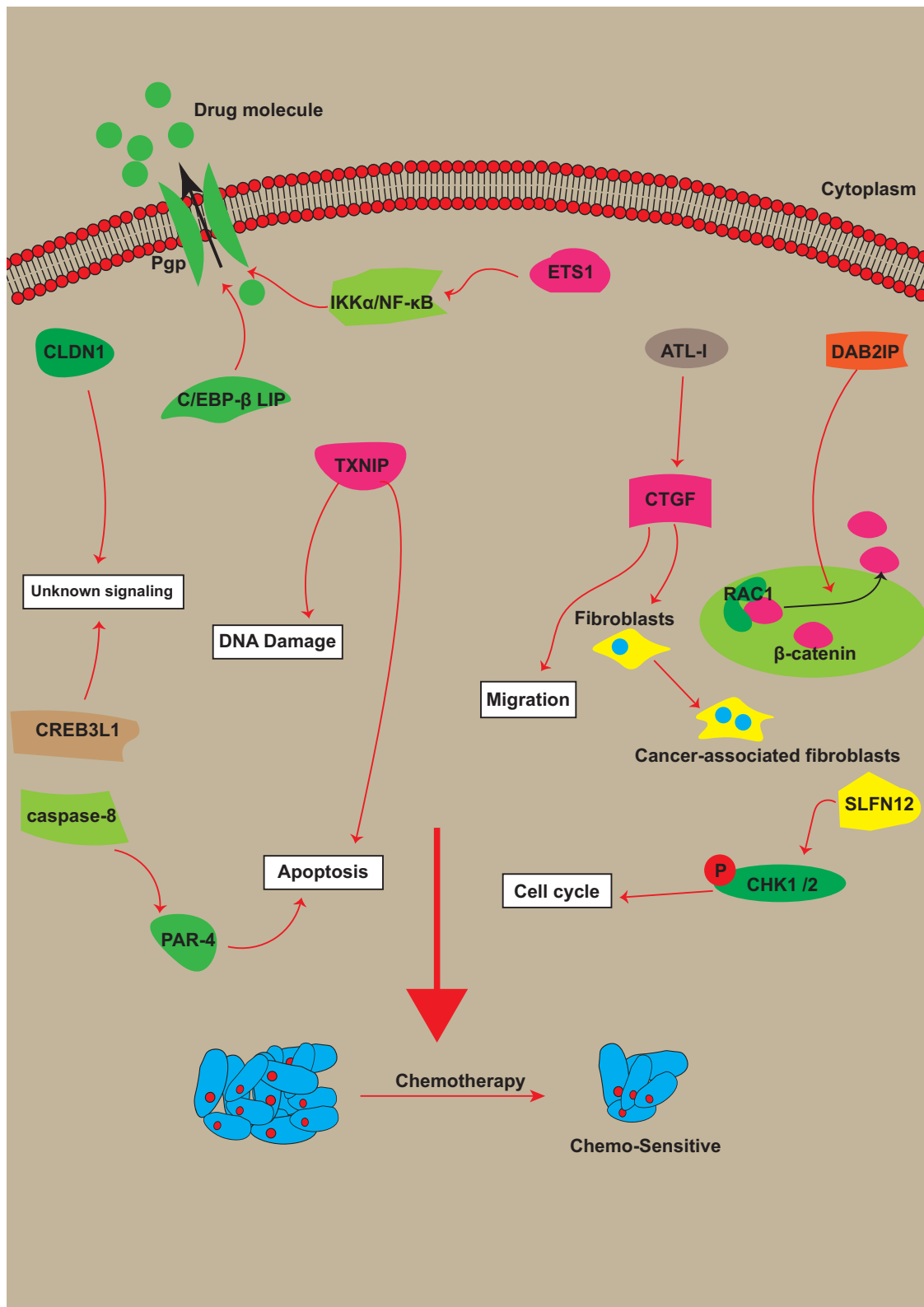


Figure 2 Genes promoting chemotherapeutic sensitivity in TNBC.

Table 3 Chemotherapy-Sensitive Genes of TNBC

Genes	Target Gene	Medication	Refs.
CLDN1	Unknown pathway	PTX	[169]
DAB2IP	RAC1-mediated β -catenin nuclear accumulation	Docetaxel	[170]
TXNIP	ROS-based DNA damage	DOX	[171–173]
C/EBP- β LIP	Pgp	Anthracycline	[174, 175]
CREB3L1	Unknown pathway	Doxorubicin	[176]
ATL-1	CTGF	PTX	[177, 178]
SLFN12	CHK1 and CHK2	PTX	[179, 180]
PAR-4	Caspase-8	Doxorubicin	[181]
ETS-1	IKK α /NF- κ B pathway	Cisplatin	[182]

Prostate Apoptosis Response-4 (PAR-4)

PAR-4 deficiency in TNBC facilitates resistance to DNA damage-driven apoptosis, while simultaneously preventing caspase-8 activation. PAR-4 is downstream of caspase-8, and it functions by cleavage-triggered nuclear transfer of the C-terminal. Herein, researchers demonstrated that the nuclear transfer of the C-terminal PAR-4 fragment strongly depletes cIAP1, then activates caspase-8.¹⁸¹

ETS1

Strong evidence suggests that the ETS1 dysregulation is behind the cisplatin resistance of TNBC. This indicates that ETS1 overexpression can potentially enhance cisplatin chemosensitivity by augmenting ABC transporters via the IKK α /NF- κ B axis.¹⁸²

Chemotherapy-Resistance Genes of TNBC

Autophagy-Related Genes

SERCA2. SERCA2 is intricately linked to TNBC progression, and it enhances TNBC cell chemoresistance. SERCA2 was shown to interact with LC3B via the LIR motif in order to augment WIPI2-independent autophagosome production, which promotes autophagy.¹⁸³

HORMA Domain-containing Protein 1 (HORMAD1). HORMAD1 is a CTA with normal testicular expression, whereby it is engaged in essential physiological activities.¹⁸⁴ In TNBC, however, it is expressed in substantially elevated levels.^{185,186} HORMAD1 modulates autophagy, and autophagic partially augments docetaxel-induced cellular apoptosis.¹⁸⁷

TNFSF13. TNFSF13 accelerates autophagy and desensitizes TNBC cells to chemotherapy by inhibiting the AKT-mTOR axis. Hence, the TNFSF13-autophagy network is an excellent target for reversing TNBC chemoresistance.¹⁸⁸

Cell Cycle-Related Genes

Protein Phosphatase 1 Regulatory Subunit 14B (PPP1R14B). PPP1R14B belongs to the protein phosphatase 1 family of modulatory subunits, and it is strongly elevated in TNBC tissues. PPP1R14B augments STMN1-based α -tubulin acetylation, microtubular stability, and cell-cycle progression, which, in turn, facilitates TNBC cell resistance to PTX.¹⁸⁹

CENPF. CENPF expression is strongly elevated in TNBC, and it is linked to worse outcome in patients undergoing chemotherapy.^{190,191} In vitro CENPF deficiency strongly enhances adriamycin (ADR)-driven cytotoxicity in MDA-MB-231 and ADR-resistant cells (MDA-MB-231/ADR). CENPF also modulates the Chk1-based G2/M phase

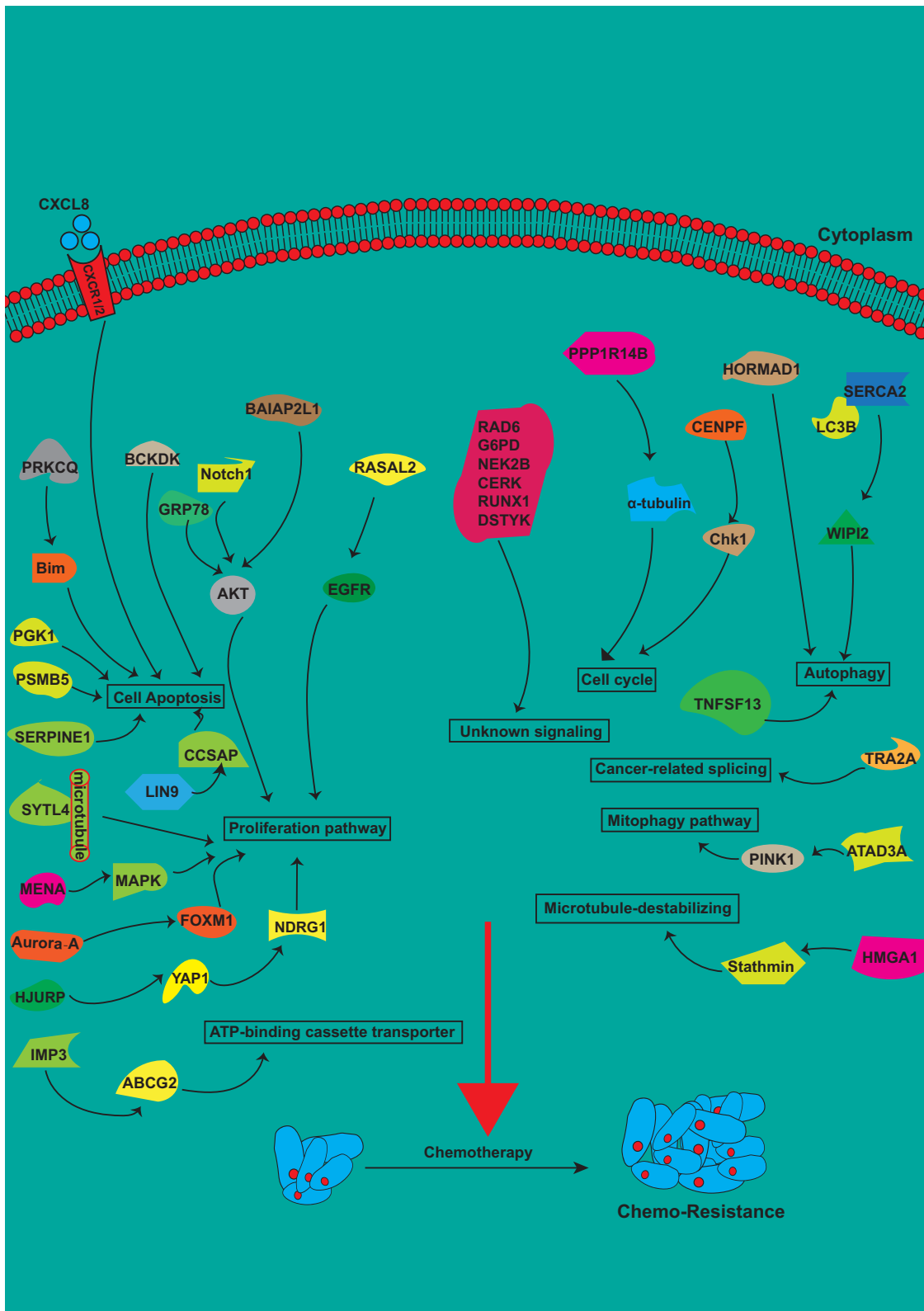


Figure 3 Genes promoting chemotherapeutic resistance in TNBC.

Table 4 Chemotherapy-Resistant Genes of TNBC

Genes	Target Gene/Signaling Pathway	Medicine	Ref.
SERCA2	LC3B	PTX	[183]
HORMAD1	Apoptosis and Autophagy	Docetaxel	[184–187]
TNFSF13	AKT-mTOR	PTX	[188]
PPP1R14B	STMN1	Paclitaxel	[189]
CENPF	Chk1	Adriamycin	[190–192]
Notch1	MVP/AKT	Cisplatin	[193]
HJURP	HJURP/YAPI/NDRG1 axis	Doxorubicin	[194]
BAIAP2L1	RPL3/AKT	Docetaxel	[195]
RASAL2	MEK/EGFR	Cisplatin	[168]
SYTL4	C2A domain	PTX	[196]
MENA	MAPK signaling	PTX	[197]
Aurora-A	FOXMI	PTX	[198]
GRP78	p-AKT/mTOR pathway	Pirarubicin	[199, 200]
CXCL8	Apoptosis signaling	PTX	[201, 202]
BCKDK	BCAA flux	DOX	[203]
PRKCQ	Bim	PTX; doxorubicin	[204]
PSMB5	Apoptosis	PTX	[205]
LIN9	CCSAP	PTX	[206]
PGK1	XAF1	PTX	[207]
SERPINE1	VEGFA	PTX	[208]
ATAD3A	PINK1	PTX	[209]
TRA2A	Splicing factors	PTX	[210]
HMGA1	HMGA1/p27/stathmin	Paclitaxel	[211, 212]
IMP3	ABCG2	Doxorubicin	[213]
RAD6	Microtubule dynamics	PTX	[214]
G6PD	Unknown signaling	DOX	[215]
NEK2B	β -catenin	PTX	[216, 217]
CERK	Unknown signaling	Paclitaxel; Cisplatin	[218]
RUNX1	Unknown signaling	Paclitaxel	[219]
DSTYK	Unknown signaling	DOX	[220]

arrest, and associates with Rb to sequester E2F1 in TNBC. Given the critical E2F1 role in DNA damage response and repair, discovery of a new signal transduction whereby CENPF modulates the Rb-E2F1 network will enable a novel approach to overcoming chemoresistance in TNBC.¹⁹²

Proliferation-Related Genes

Notch1. Notch1 and major vault protein (MVP) are robust regulators of TNBC cell chemoresistance. It does so by modulating MVP expression to stimulate the AKT and EMT axes. Notch1 deficiency increases TNBC cell sensitivity to cisplatin-based chemotherapy.¹⁹³

HJURP. HJURP modulates Yes-Associated Protein (YAP1) protein ubiquitination to control its downstream transcriptional activity. In addition, YAP1 associated with the NDRG1 gene promoter to enhance gene transcription. Subsequently, the HJURP/YAP1/NDRG1 network modulates cell proliferation and doxorubicin resistance in TNBC. Together, these evidences indicate that the HJURP/YAP1/NDRG1 network modulation is essential in TNBC progression and therapeutic response.¹⁹⁴

BAI-Associated Protein 2-Like 1 (BAIAP2L1). Excess BAIAP2L1 expression markedly induces BC proliferation potentially via AKT activation. Conversely, exposure to AKT inhibitor LY294002 diminishes BAIAP2L1 expression in BC cells. BAIAP2L1 interacts with the AA202-288 of ribosomal protein L3 (RPL3) within its SRC homology 3 (SH3) domain. In contrast, its deficiency abolishes AKT signaling to enhance PIK3CA degradation. Additionally, BAIAP2L1 overexpression enhances docetaxel resistance.¹⁹⁵

RASAL2. Emerging evidences revealed that excess RASAL2 forms a chemotherapy-unresponsive TNBC subset. RASAL2 GAP function is critical for kinase inhibitor sensitivity, and RASAL2-high TNBCs maintain basal RAS activity via inhibition of negative feedback modulators SPRY1/2, along with EGFR elevation. As a result, RASAL2 expression reduces feedback compensation following MEK1/2 and EGFR co-suppression thereby inducing synergistic apoptosis in vitro and in vivo. Among TNBC patients, elevated RASAL2 contents indicate clinical chemotherapeutic response (Cisplatin).¹⁶⁸

Synaptotagmin-Like 4 (SYTL4). SYTL4 is a relatively new chemo-resistant gene in TNBC. SYTL4 downregulation stabilizes the microtubular network by decelerating microtubular growth rate. Additionally, SYTL4 expresses alongside microtubules and interacts with the middle region of microtubules via a linker and C2A domain. SYTL4 also induces PTX resistance in TNBC in vitro and in vivo.¹⁹⁶

MENA. The MENA isoform disrupts the balance between dynamic and stable microtubules in PTX-exposed cells. Following PTX exposure, MENA activates MAPK. Conversely, following MEK inhibitor co-treatment, reduced ERK phosphorylation restores PTX sensitivity by enhancing microtubule stabilization in MENA isoform-harboring cells.¹⁹⁷

Aurora Kinase A (Aurora-A). Aurora-A modulates FOXM1 in TNBC. Aurora-A-induced FOXM1 stabilization represents its kinase-independent induction of augmented TNBC cell proliferative capacity. Even though Aurora-A kinase inhibition is a potentially potent TNBC treatment regimen, the report indicated that the antibiotic drug thiostrepton usage, which downregulates Aurora-A and FOXM1 contents, can work together with PTX to revert PTX chemoresistance in TNBC.¹⁹⁸

GRP78. GRP78 is intricately linked to apoptosis, and GRP78 deficiency enhances BC apoptosis.¹⁹⁹ GRP78 silencing strongly reduces MDR1 levels by negatively modulating the Akt/mTOR network in MDA-MB-231R cells.²⁰⁰ Furthermore, it reverts pirarubicin resistance of TNBC using miR-495-3p mimics and the p-AKT/mTOR axis.²⁰⁰

Apoptosis-Related Genes

Cxcl8. CXCL8, otherwise called IL-8, originates from the proangiogenic chemokine family, and it is strongly linked to CXCR1/2.²⁰¹ CXCL8 is ubiquitously present in PTX-resistant TNBC cells. In contrast, CXCL8 deficiency enhances TNBC cell sensitivity to PTX via regulation of the cell apoptosis network.²⁰²

Branched-Chain Ketoacid Dehydrogenase Kinase (BCKDK). Based on transcriptome analysis, BCKDK silencing dysregulates mitochondrial metabolic axes and upregulates the apoptotic axis. BCKDK suppression and simultaneous DOX exposure exacerbate apoptosis, caspase activity, and inhibition of TNBC proliferation. Collectively, BCKDK inactivity in TNBC alters BCAA flux, diminishes protein translation, thereby accelerating cell apoptosis, ATP insufficiency, and susceptibility toward genotoxic stress.²⁰³

PRKCQ. Excess PRKCQ content strongly suppresses PTX- or doxorubicin-induced cell apoptosis. Conversely, PRKCQ deficiency enhances chemotherapy-induced TNBC cell apoptosis. PRKCQ modulates chemotherapy responsiveness by regulating Bim (a pro-apoptotic Bcl2 family member) content. Bim suppression abrogates the simultaneous PRKCQ deficiency- and chemotherapy treatment-induced cell apoptosis.²⁰⁴

Proteasome Subunit Beta 5 (PSMB5). Excess PSMB5 expression is strongly associated with worse TNBC patient outcome. In contrast, PSMB5 deficiency enhances TNBC cell apoptosis and augments chemotherapeutic sensitivity to PTX.²⁰⁵

LIN9. PTX-resistant TNBC cells have strongly elevated LIN9 content, relative to corresponding parental cells. LIN9 knockdown or PTX-resistant TNBC cell exposure to a bromo- and extra-terminal domain inhibitor (BETi) JQ1 substantially diminished LIN9 levels while enhancing PTX-resistant TNBC cell sensitivity to PTX. LIN9 suppression in resistant cell lines visibly reduces tumor cell survivability, increases multinucleated cell production, and enhances tumor cell apoptosis via direct regulation of CCSAP.²⁰⁶

PGK1. PGK1 silencing markedly increased TNBC cell line sensitivity to PTX therapy by enhancing drug-induced apoptosis. Moreover, PGK1-knockdown cells display excess XAF1, as well as apoptotic proteins cleaved caspase-3 and Bax expressions.²⁰⁷

SERPINE1. Enhanced SERPINE1 expression confers BC resistance to PTX treatment via elevation of VEGFA, and inhibition of cell apoptosis.²⁰⁸

Mitophagy-Related Genes

ATAD3A. PINK1 transports PD-L1 to the mitochondria for mitophagy-mediated destruction. PTX also increases ATAD3A content to impair PD-L1 proteostasis via modulation of PINK1-reliant mitophagy. In the clinics, tumor patients who have elevated ATAD3A levels prior to ICI and PTX co-treatment experienced substantially reduced PFS relative to those with ATAD3A-low tumors.²⁰⁹

Cancer-Related Splicing Genes

Transformer2A (TRA2A). TRA2A promotes TNBC cell proliferation, survival, migration, and invasion. Additionally, TRA2A induces PTX resistance of TNBC cells via specific regulation of cancer-associated splicing, independent of other splicing factors.²¹⁰

Microtubule-Destabilizing-Related Splicing Genes

High Mobility Group A1 (HMGA1). HMGA1 is typically ubiquitously expressed during embryogenesis, and it declines to little or no expression in adults. However, it is strongly expressed in multiple tumors.²¹¹ The HMGA1/p27/stathmin network regulates TNBC cell motility. Therefore, HMGA1 silencing in TNBC cells strongly diminishes stathmin content and action on microtubules. This, in turn, impairs cell motility in a p27-reliant fashion. Additionally, in a xenograft mouse model, HMGA1-depleted TNBC cells are more responsive to PTX treatment.²¹²

ATP-Binding Cassette Transporter-Related Genes

IMP3. BC resistance protein (BCRP), otherwise called ABCG2, belongs to the ATP-binding cassette transporter family. The IMP3 protein is reported to enhance chemoresistance (Doxorubicin) of BC cells via modulating ABCG2 levels.²¹³

Unknown Signaling-Related Genes

RAD6. SMI#9 (a RAD6 inhibitor) and PTX share the same mechanism of microtubular modulation. Therefore, the synergistic effect of both PTX and RAD6 inhibitor has potential advantages on enhancing TNBC sensitivities to PTX

while eliminating toxicity. The aforementioned investigations were the first to uncover a role for RAD6 in modulating microtubule dynamics.²¹⁴

Glucose-6-Phosphate Dehydrogenase (G6PD). Metastatic TNBC cells exhibit markedly elevated G6PD levels once DOX resistance form following lung metastasis. Moreover, G6PD suppression abrogates DOX resistance in the same mTNBC cells, while enhancing parental TNBC cell responsiveness to DOX.²¹⁵

NEK2B. Elevated Nek2B expression is intricately linked to worse disease-free survival (DFS) and OS in TNBC.²¹⁶ Nek2B interacts with β -catenin to negatively regulate TNBC patient prognosis. Their synergistic action enhances TNBC chemo-resistance to PTX.²¹⁷

Ceramide Kinase (CERK). Upregulated CERK activates numerous oncogenic networks to promote chemo-resistance in TNBC cells. Additionally, excess CERK expression heavily influences chemo-sensitivity (PTX and cisplatin), and therefore serve as a potential bioindicator of chemo-resistance risk classification in newly diagnosed TNBC patients.²¹⁸

RUNX1. Suppression of RUNX1 transcriptional activity strongly enhances chemotherapy sensitivity in (AR⁺) TNBC cell lines both in culture and forced suspension. Pharmacologic RUNX1 suppression also augments the effects of the aforementioned combinational treatments (PTX).²¹⁹

DSTYK. One investigation knocked out DSTYK using the CRISPR/Cas9 method, and revealed considerable chemo-resistant cell death following drug intervention. In addition, DSTYK knockout enhances the chemotherapeutic drug-driven tumor cell apoptosis in an orthotopic mouse model. Collectively, these results indicate that DSTYK is essential for DOX chemoresistance.²²⁰

Potential Novel Therapeutic Targets for TNBC

Considering the aforementioned TNBC-based evidences, a detailed and extensive list of chemotherapy, as well as existing and ongoing targeted molecular therapy including the following: (1) chemotherapy; (2) DRR axis inhibitors (targeting BRCA, ATR-CHK1-WEE1); (3) proliferation-associated network inhibitors (targeting PI3K-AKT-mTOR; NOTCH; RB; EGFR); (4) ICBs (targeting PD-L1/PD-1); (5) AR inhibitor. With emerging investigations, more researchers have proposed additional approaches to treating TNBC. Here, we have summarized the most current potential TNBC therapies, namely, ferroptosis-associated network; potential targeted ADC markers; Ceritinib; Src Inhibitor; CDK12/CDK13 inhibitor. All TNBC-related therapies and medications are presented in [Figure 4](#) and [Table 2](#).

Ferroptosis-Associated Network

Twelve genes linked to ferroptosis were analyzed for prognostic evaluation in TNBC cases, with seven genetic markers (ASNS, LAMP2, CAV1, DPP4, HELLS, TF, ZFP69B) emerging as viable therapeutic candidates. Additionally, two pharmaceutical compounds (1-methyl-3-isobutylxanthine and rosiglitazone) demonstrated potential efficacy in TNBC treatment scenarios.⁸⁵

Potential Targeted ADC Bioindicators

ADC enhances high cytotoxic drug concentration delivery to ADC-containing cells.⁸⁶ Sacituzumab govitecan (IMMU-132) is an SN-38 conjugate Trop-2-targeting antibody. Trop-2 is present in approximately 90% of all TNBCs.⁸⁷ Ladiratumumab Vedotin (SGN-LIV1A), with monomethyl-auristatin-E (MMAE) as the payload, achieves a 25% ORR and median PFS of 11 months among TNBC patients. Excess glycoprotein-NMB (gpNMB) expression, indicated by $\geq 25\%$ of tumor epithelial cell staining, occurs in about 40% of TNBCs, and in this subgroup, glembatumumab vedotin (CDX-011, an ADC that interacts with gpNMB to deploy MMAE) achieves 40% ORR versus 0% with the author's therapy of choice.⁸⁸ Some new ADC medicines have also been created and engaged in clinical trials. AVID100 is an anti-EGFR ADC that targets EGFR and is conjugated with DM1.⁸⁹ U3-1402 is an anti-HER3 ADC that is conjugated with a topoisomerase I inhibitor exatecan derivative via a peptidyl linker.²²¹ CAB-ROR2-ADC is an ADC composed of a conditionally active biologic antibody directing receptor tyrosine kinase-like orphan receptor 2 (ROR2) conjugated to an undisclosed payload.⁸⁹ Anti-CA6-DM4 immunoconjugate (SAR566658) is a humanized DS6 antibody directed against CA6 conjugated to DM4.⁸⁹ New targeted for ADC drug development is the most potential direction for TNBC therapy.

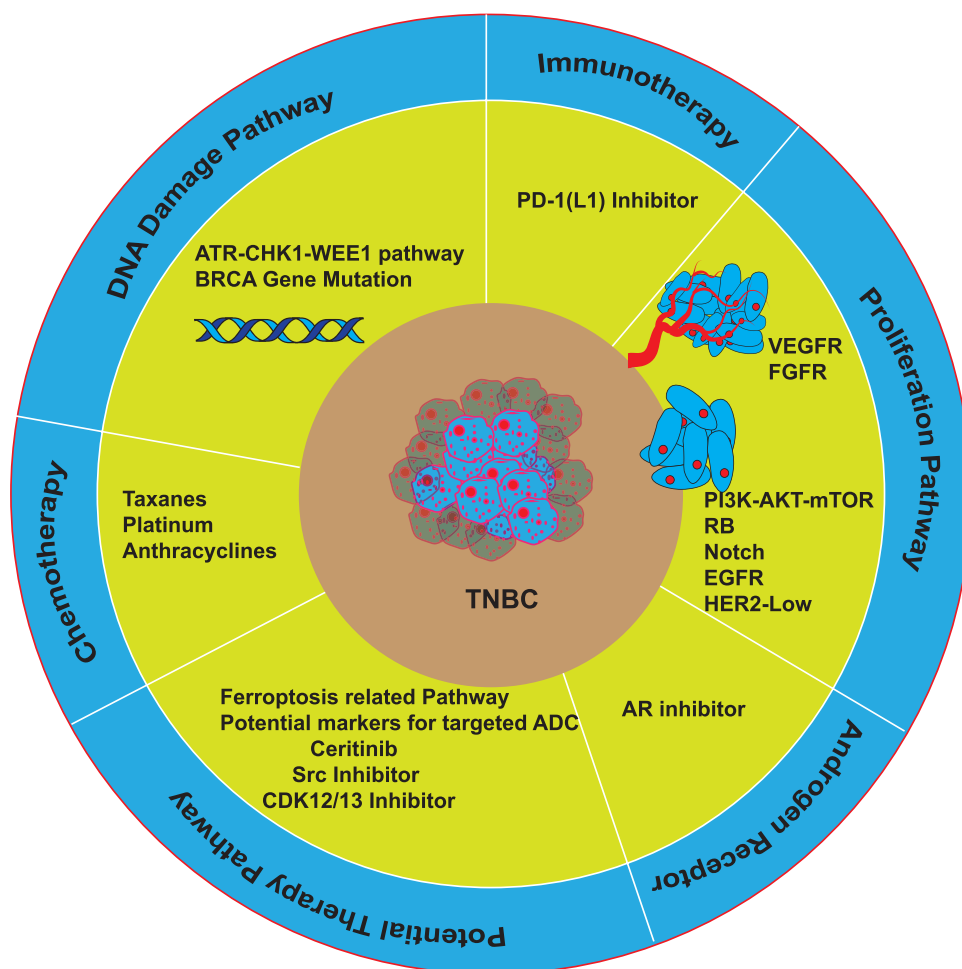


Figure 4 An illustration of all available therapeutic means of TNBC including the DNA damage pathway, immunotherapy, chemotherapy, proliferation pathway, androgen receptor, potential therapeutic pathway.

Ceritinib

Ceritinib, an FDA-approved anti-lung cancer medication,⁹⁰ significantly inhibits LAR TNBC development. Additionally, Activated CDC42 Kinase 1 (ACK1) is a strong ceritinib target in LAR TNBC cells, whereby ceritinib inactivates the RTK-ACK1/FAK-AR network in LAR TNBC cells. Lastly, this investigation established a new treatment approach for AR⁺ TNBC tumors involving a combinational therapy for AR⁻ or AR low TNBCs.⁹¹

Src Inhibitor

Src is a specific tyrosine kinase that has gained recognition as a possible target for multi-cancer treatment.^{92,93} Prior investigations by Finn et al⁹⁴ reported that TNBC cell lines are especially responsive to dasatinib, whereas, Huang et al demonstrated that a BC subgroup harboring the dasatinib-responsive signature includes patients with TNBC.⁹⁵ It was also revealed that dasatinib suppresses different TNBC cellular growth in culture. However, Src content or phosphorylation failed to indicate dasatinib responsiveness in vitro.⁹⁶

CDK12/CDK13 Inhibitor

CDKs are serine/threonine kinase activities are heavily reliant on their association with the cyclin modulatory subunit.²²² The CDK enzyme family modulates the cell cycle and transcription processes.²²³ Quereda et al established a targeted dual CDK12/CDK13 inhibitor (SR-4835) that diminishes core DNA damage response gene expressions by upregulating intronic polyadenylation site cleavage, which results in impaired DNA damage repair, thereby increasing cellular responsiveness to DNA-damaging agents and PARP inhibitors.⁹⁷

Conclusion and Future Perspective

TNBC is a disease marked by enhanced heterogeneity and poor prognosis. Unlike HR-positive BC, TNBC is correlated with a younger patient age, increased visceral metastases risk, enhanced early recurrence risk, and extremely poor outcome.^{9,224,225} Therapy for TNBC patients, who lack ER and PR expressions as well as HER2 amplification, is quite challenging due to the heterogenic characteristic of this disease, as well as the lack of well-established molecular targets.^{167,226,227} Gene expression profiling-related molecular TNBC subtyping is critical for elucidating the underlying mechanisms of this disease, which, in turn, will benefit personalized treatment.²⁹ Hence, an extensive exploration of all subtypes and properties is crucial. Additionally, newly discovered subtype stratification enables clinicians to better manage TNBC. As a result, given the poor outcome of TNBC patients, it is essential to gain an extensive comprehension of the disease, particularly, in relation to the following aspects: (a) DDR-associated BRCA Loss-Function Mutation; (b) PI3K axis; (c) NOTCH network; (d) CDK-Rb axis; (e) EGFR Activation; (f) Angiogenesis network; (g) HER2-Low; (h) DDR signal transduction; (i) Immunotherapy and (j) AR signaling. Based on the aforementioned genes and aberrant signal networks, scientists have developed potential targeted therapies, as is presented in [Table 2](#). The relevant clinical trials are summarized in [Supplemental Table 1](#). It is our hope and speculation that the mentioned medications will be incredibly advantageous to TNBC therapy in near future. Notably, with the emergence of the HER2-low concept, drugs targeting HER2-low are being proposed and placed in clinical trials. Furthermore, given that chemotherapy is the standard intervention for TNBC, herein, we summarized the coding genes associated with TNBC chemoresistance, which may eventually be targets of future TNBC therapy. Additionally, we also listed the known chemo-sensitive and chemo-resistance genes associated with TNBC. Moreover, the CLDN1, DAB2IP, TXNIP, C/EBP- β LIP, CREB3L1, ATL-1, SLFN12, PAR-4, and ETS-1 genes may can be used to predict patient sensitivity to chemotherapy or neoadjuvant-chemotherapy in the clinic. Similarly, the HJURP, CXCL8, SERCA2, PPP1R14B, HORMAD1, DSTYK, Notch1, and CENPF genes serve as indicators of patient resistance to chemotherapy in the clinic. We speculate that the aforementioned chemo-sensitive and chemo-resistant genes will be examined further in coming years for the development of better anti-TNBC therapy. We also discussed the emergence of new treatments for TNBC, namely, the ferroptosis-associated network, ADC, Ceritinib, Src inhibitor, and CDK12/CDK13 inhibitor. Considering that the aforementioned drugs continue to show benefit against TNBC, they may enter clinical trials relatively soon, and be administered to TNBC patients in the clinic.

In summary, this review presents an extensive overview of the gene mutation profile of TNBC, as well as the corresponding targeted immune- and chemotherapies. This will provide a direction and beacon for the development of highly efficacious future TNBC therapy.

Abbreviations

ACK1, Activated CDC42 Kinase 1; ADCs, Antibody-drug conjugates; ADR, Adriamycin; Aurora-A, Aurora kinase A; AR, Androgen Receptor; ATL-I, Atractylenolide-I; BAIAP2L1, BAI-associated protein 2- like 1; BC, Breast Cancer; BCKDK, Branched-chain ketoacid dehydrogenase kinase; BCRP, Breast cancer Resistance Protein; BETi, bromo- and extra-terminal domain inhibitor; bFGF, Basic Fibroblast Growth Factor; BLIA, Basal-like Immune activated; BLIS, Basal-like Immunosuppressed; CAFs, Cancer-Associated Fibroblasts; CDK 4/6, Cyclin-Dependent Kinases 4 and 6; CDK4/6i, CDK4/6 inhibitor; CERK, Ceramide Kinase; CLDN1, Claudin-1; CREB3L1, cAMP response element binding protein 3-like 1; DC, Dendritic Cell; DDR, DNA Damage Response; DFS, Disease-Free Survival; ER, Estrogen Receptor; EMT, Epithelial-Mesenchymal Transition; FGFR, Fibroblast Growth Factor Receptors; G6PD, Glucose-6-phosphate Dehydrogenase; gpNMB, Glycoprotein-NMB; HER-2, Human Epidermal Growth Factor Receptor; HMGA1, High Mobility Group A1; HORMAD1, HORMA domain-containing protein 1; HR, Homologous Recombination; ICB, Immune Checkpoint Blockade; ICI, Immune checkpoint inhibitor; IHC, Immunohistochemical; IHC-BLIS, IHC-based basal-like immune-suppressed; IHC-LAR, IHC-based luminal androgen receptor; IHC-IM, IHC-based immunomodulatory; IHC-MES, IHC-based mesenchymal; IL-8, Interleukin-8; IM, Immunomodulatory; ISH, In Situ Hybridization; LAR, Luminal Androgen Receptor; mAb, Monoclonal Antibodies; MES, Mesenchymal; MSC, Mesenchymal Stem Cell; MSL, Mesenchymal stem-like; MMAE, Monomethyl-Auristatin-E; MVP, Major Vault Protein; OS, Overall Survival; PAR-4, Prostate apoptosis response-4; PARP, Poly ADP-ribose Polymerase; PARPi, Poly ADP-ribose Polymerase Inhibitor; pCR, Pathological

Complete Response; PD-1, programmed death-1; PD-L1, programmed death-ligand 1; PIK3CA, PI3K catalytic subunit α ; Pgp, P-glycoprotein; PI3K, Phosphoinositide 3-kinase; PPP1R14B, Protein phosphatase 1 regulatory subunit; PR, Progesterone Receptor; PSMB5, Proteasome Subunit Beta 5; PTEN, Phosphatase And Tensin Homolog; PTX, Paclitaxel; ROR2, Receptor Tyrosine Kinase-Like Orphan Receptor 2; RPL3, ribosomal protein L3; RTK, Receptor Tyrosine Kinase; SH3, SRC homology 3; Src, v-src Sarcoma Viral Oncogene Homolog; SYTL4, Synaptotagmin-like 4; TF, Transcription Factor; TGF, transforming growth factor; TNBC, Triple Negative Breast Cancer; TILs, Tumor Infiltrating Lymphocytes; TKI, Tyrosine kinase inhibitors; TRA2A, Transformer2A; TXNIP, Thioredoxin-interacting protein; VEGF, Vascular endothelial growth factor; YAP1, Yes-Associated Protein.

Data Sharing Statement

No datasets were generated or analyzed during the current study.

Human and Animal Rights and Informed Consent

This article does not contain any studies with human or animal subjects performed by any of the authors.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

Funding

Beijing Vlove Charity Foundation (ky1322); Yantai Yuhuangding Hospital Youth Launch Fund (kj0521) and Yantai Yuhuangding Hospital Doctoral Research Launch Fund (kj0570).

Disclosure

The authors declare no competing interests.

References

- Junnuthula V, Kolimi P, Nyavanandi D, Sampathi S, Vora LK, Dyawanapelly S. Polymeric Micelles for Breast Cancer Therapy: recent Updates, Clinical Translation and Regulatory Considerations. *Pharmaceutics*. 2022;14(9):1860. doi:10.3390/pharmaceutics14091860
- Feng Y, Spezia M, Huang S, et al. Breast cancer development and progression: risk factors, cancer stem cells, signaling pathways, genomics, and molecular pathogenesis. *Genes Dis*. 2018;5(2):77–106. doi:10.1016/j.gendis.2018.05.001
- Cheang MC, Voduc D, Bajdik C, et al. Basal-like breast cancer defined by five biomarkers has superior prognostic value than triple-negative phenotype. *Clin Cancer Res*. 2008;14(5):1368–1376. doi:10.1158/1078-0432.CCR-07-1658
- Cancer Genome Atlas Network. Comprehensive molecular portraits of human breast tumours. *Nature*. 2012;490(7418):61–70. doi:10.1038/nature11412
- Lehmann BD, Bauer JA, Chen X, et al. Identification of human triple-negative breast cancer subtypes and preclinical models for selection of targeted therapies. *J Clin Invest*. 2011;121(7):2750–2767. doi:10.1172/JCI45014
- Zeichner SB, Terawaki H, Gogineni K. A Review of Systemic Treatment in Metastatic Triple-Negative Breast Cancer. *Breast Cancer*. 2016;10:25–36. doi:10.4137/BCBCR.S32783
- Dent RTrudeau M, Pritchard KI, Hanna WM, et al. Triple-negative breast cancer: clinical features and patterns of recurrence. *Clin Cancer Res*. 2007;13(15):4429–4434. doi:10.1158/1078-0432.CCR-06-3045
- Foulkes WD, Smith IE, Reis-Filho JS. Triple-negative breast cancer. *N Engl J Med*. 2010;363(20):1938–1948. doi:10.1056/NEJMra1001389
- Liedtke C, Mazouni C, Hess KR, et al. Response to neoadjuvant therapy and long-term survival in patients with triple-negative breast cancer. *J Clin Oncol*. 2008;26(8):1275–1281. doi:10.1200/JCO.2007.14.4147
- Gonzalez-Angulo AM, Timms KM, Liu S, et al. Incidence and Outcome of BRCA Mutations in Unselected Patients with Triple Receptor-Negative Breast Cancer. *Clin Cancer Res*. 2011;17(5):1082–1089. doi:10.1158/1078-0432.CCR-10-2560
- Sikov WM, Berry DA, Perou CM, et al. Impact of the addition of carboplatin and/or bevacizumab to neoadjuvant once-per-week paclitaxel followed by dose-dense doxorubicin and cyclophosphamide on pathologic complete response rates in stage II to III triple-negative breast cancer: CALGB 40603 (Alliance). *J Clin Oncol*. 2015;33(1):13–21. doi:10.1200/JCO.2014.57.0572
- Von Minckwitz G, Schneeweiss A, Loibl S, et al. Neoadjuvant carboplatin in patients with triple-negative and HER2-positive early breast cancer (GeparSixto; GBG 66): a randomised Phase 2 trial. *Lancet Oncol*. 2014;15(7):747–756. doi:10.1016/S1470-2045(14)70160-3

13. Rose M, Burgess JT, O'byrne K, Richard DJ, Bolderson E. PARP Inhibitors: clinical Relevance, Mechanisms of Action and Tumor Resistance. *Front Cell Dev Biol.* 2020;8:564601. doi:10.3389/fcell.2020.564601
14. Sharma P, Allison JP. Immune checkpoint targeting in cancer therapy: toward combination strategies with curative potential. *Cell.* 2015;161(2):205–214. doi:10.1016/j.cell.2015.03.030
15. Stanton SE, Adams S, Disis ML. Variation in the Incidence and Magnitude of Tumor-Infiltrating Lymphocytes in Breast Cancer Subtypes: a Systematic Review. *JAMA Oncol.* 2016;2(10):1354–1360. doi:10.1001/jamaoncol.2016.1061
16. Wimberly H, Brown JR, Schalper K, et al. PD-L1 Expression Correlates with Tumor-Infiltrating Lymphocytes and Response to Neoadjuvant Chemotherapy in Breast Cancer. *Cancer Immunol Res.* 2015;3(4):326–332. doi:10.1158/2326-6066.CIR-14-0133
17. Iglesia MD, Vincent BG, Parker JS, et al. Prognostic B-cell signatures using mRNA-seq in patients with subtype-specific breast and ovarian cancer. *Clin Cancer Res.* 2014;20(14):3818–3829. doi:10.1158/1078-0432.CCR-13-3368
18. Yu L, Wang J. T cell-redirecting bispecific antibodies in cancer immunotherapy: recent advances. *J Cancer Res Clin Oncol.* 2019;145(4):941–956. doi:10.1007/s00432-019-02867-6
19. Marra A, Viale G, Curigliano G. Recent advances in triple negative breast cancer: the immunotherapy era. *BMC Med.* 2019;17(1):90. doi:10.1186/s12916-019-1326-5
20. Pascual J, Turner NC. Targeting the PI3-kinase pathway in triple-negative breast cancer. *Ann Oncol.* 2019;30(7):1051–1060. doi:10.1093/annonc/mdz133
21. Asghar US, Barr AR, Cutts R, et al. Single-Cell Dynamics Determines Response to CDK4/6 Inhibition in Triple-Negative Breast Cancer. *Clin Cancer Res.* 2017;23(18):5561–5572. doi:10.1158/1078-0432.CCR-17-0369
22. Metzger-Filho O, Tutt A, De Azambuja E, et al. Dissecting the heterogeneity of triple-negative breast cancer. *J Clin Oncol.* 2012;30(15):1879–1887. doi:10.1200/JCO.2011.38.2010
23. Lehmann BD, Pietenpol JA. Identification and use of biomarkers in treatment strategies for triple-negative breast cancer subtypes. *J Pathol.* 2014;232(2):142–150. doi:10.1002/path.4280
24. Gibson GR, Qian D, Ku JK, Lai LL. Metaplastic breast cancer: clinical features and outcomes. *Am Surg.* 2005;71(9):725–730. doi:10.1177/000313480507100906
25. Bareche Y, Venet D, Ignatiadis M, et al. Unravelling triple-negative breast cancer molecular heterogeneity using an integrative multiomic analysis. *Ann Oncol.* 2018;29(4):895–902. doi:10.1093/annonc/mdy024
26. Hayes MJ, Thomas D, Emmons A, Giordano TJ, Kleer CG. Genetic changes of Wnt pathway genes are common events in metaplastic carcinomas of the breast. *Clin Cancer Res.* 2008;14(13):4038–4044. doi:10.1158/1078-0432.CCR-07-4379
27. Yin L, Duan JJ, Bian XW, Yu SC. Triple-negative breast cancer molecular subtyping and treatment progress. *Breast Cancer Res.* 2020;22(1):61. doi:10.1186/s13058-020-01296-5
28. Burstein MD, Tsimelzon A, Poage GM, et al. Comprehensive genomic analysis identifies novel subtypes and targets of triple-negative breast cancer. *Clin Cancer Res.* 2015;21(7):1688–1698. doi:10.1158/1078-0432.CCR-14-0432
29. Zhao S, Ma D, Xiao Y, et al. Molecular Subtyping of Triple-Negative Breast Cancers by Immunohistochemistry: molecular Basis and Clinical Relevance. *Oncologist.* 2020;25(10):e1481–e1491. doi:10.1634/theoncologist.2019-0982
30. He Y, Jiang Z, Chen C, Wang X. Classification of triple-negative breast cancers based on Immunogenomic profiling. *J Exp Clin Cancer Res.* 2018;37(1):327. doi:10.1186/s13046-018-1002-1
31. Bonotto M, Gerratana L, Poletto E, et al. Measures of outcome in metastatic breast cancer: insights from a real-world scenario. *Oncologist.* 2014;19(6):608–615. doi:10.1634/theoncologist.2014-0002
32. Dent R, Hanna WM, Trudeau M, Rawlinson E, Sun P, Narod SA. Pattern of metastatic spread in triple-negative breast cancer. *Breast Cancer Res Treat.* 2009;115(2):423–428. doi:10.1007/s10549-008-0086-2
33. Bianchini G, De Angelis C, Licata L, Gianni L. Treatment landscape of triple-negative breast cancer - expanded options, evolving needs. *Nat Rev Clin Oncol.* 2022;19(2):91–113. doi:10.1038/s41571-021-00565-2
34. Marra A, Trapani D, Viale G, Criscitiello C, Curigliano G. Practical classification of triple-negative breast cancer: intratumoral heterogeneity, mechanisms of drug resistance, and novel therapies. *NPJ Breast Cancer.* 2020;6(1):54. doi:10.1038/s41523-020-00197-2
35. Jin J, Tao Z, Cao J, Li T, Hu X. DNA damage response inhibitors: an avenue for TNBC treatment. *Biochim Biophys Acta Rev Cancer.* 2021;1875(2):188521. doi:10.1016/j.bbcan.2021.188521
36. Ibrahim YH, Garcia-Garcia C, Serra V, et al. PI3K inhibition impairs BRCA1/2 expression and sensitizes BRCA-proficient triple-negative breast cancer to PARP inhibition. *Cancer Discov.* 2012;2(11):1036–1047. doi:10.1158/2159-8290.CD-11-0348
37. Tolaney S, Burris H, Gartner E, et al. Phase I/II study of pirlalisib (SAR245408) in combination with trastuzumab or trastuzumab plus paclitaxel in trastuzumab-refractory HER2-positive metastatic breast cancer. *Breast Cancer Res Treat.* 2015;149(1):151–161. doi:10.1007/s10549-014-3248-4
38. Peek GW, Tollefsbol TO. Combinatorial PX-866 and Raloxifene Decrease Rb Phosphorylation, Cyclin E2 Transcription, and Proliferation of MCF-7 Breast Cancer Cells. *J Cell Biochem.* 2016;117(7):1688–1696. doi:10.1002/jcb.25462
39. Hoefflich KP, Guan J, Edgar KA, et al. The PI3K inhibitor taselisib overcomes letrozole resistance in a breast cancer model expressing aromatase. *Genes Cancer.* 2016;7(3–4):73–85. doi:10.18632/genesandcancer.100
40. Fritsch C, Huang A, Chatenay-Rivauday C, et al. Characterization of the novel and specific PI3Kalpha inhibitor NVP-BYL719 and development of the patient stratification strategy for clinical trials. *Mol Cancer Ther.* 2014;13(5):1117–1129. doi:10.1158/1535-7163.MCT-13-0865
41. Juric D, De Bono JS, Lorusso PM, et al. A First-in-Human, Phase I, Dose-Escalation Study of TAK-117, a Selective PI3Kalpha Isoform Inhibitor, in Patients with Advanced Solid Malignancies. *Clin Cancer Res.* 2017;23(17):5015–5023. doi:10.1158/1078-0432.CCR-16-2888
42. Lin J, Sampath D, Nannini MA, et al. Targeting activated Akt with GDC-0068, a novel selective Akt inhibitor that is efficacious in multiple tumor models. *Clin Cancer Res.* 2013;19(7):1760–1772. doi:10.1158/1078-0432.CCR-12-3072
43. Leighl NB, Dent S, Clemons M, Hedley D. A Phase 2 study of perifosine in advanced or metastatic breast cancer. *Breast Cancer Res Treat.* 2008;108:87–92. doi:10.1007/s10549-007-9584-x
44. Banerji U, Dean EJ, Perez-Fidalgo JA, Batist G. A Phase I Open-Label Study to Identify a Dosing Regimen of the Pan-AKT Inhibitor AZD5363. *Clin Cancer Res.* 2018;24:2050–2059.

45. Saura C, Roda D, Rosello S, et al. Correction: a First-in-Human Phase I Study of the ATP-Competitive AKT Inhibitor Ipatasertib Demonstrates Robust and Safe Targeting of AKT in Patients with Solid Tumors. *Cancer Discov.* 2018;8:1490.
46. Wisinski KB, Tevaarwerk AJ, Burkard ME, et al. Phase I Study of an AKT Inhibitor (MK-2206) Combined with Lapatinib in Adult Solid Tumors Followed by Dose Expansion in Advanced HER2+ Breast Cancer. *Clin Cancer Res.* 2016;22(11):2659–2667. doi:10.1158/1078-0432.CCR-15-2365
47. Basho RK, Gilcrease M, Murthy RK, et al. Targeting the PI3K/AKT/mTOR Pathway for the Treatment of Mesenchymal Triple-Negative Breast Cancer: evidence From a Phase I Trial of mTOR Inhibition in Combination With Liposomal Doxorubicin and Bevacizumab. *JAMA Oncol.* 2017;3(4):509–515. doi:10.1001/jamaoncol.2016.5281
48. Sutherlin DP, Bao L, Berry M, et al. Discovery of a potent, selective, and orally available class I phosphatidylinositol 3-kinase (PI3K)/mammalian target of rapamycin (mTOR) kinase inhibitor (GDC-0980) for the treatment of cancer. *J Med Chem.* 2011;54(21):7579–7587. doi:10.1021/jm2009327
49. Knight SD, Adams ND, Burgess JL, Dhanak D. Discovery of GSK2126458. *ACS Med Chem Lett.* 2010;1:39–43.
50. Hatem R, El Botty R, Chateau-Joubert S, et al. Targeting mTOR pathway inhibits tumor growth in different molecular subtypes of triple-negative breast cancers. *Oncotarget.* 2016;7(30):48206–48219. doi:10.18632/oncotarget.10195
51. Garcia-Echeverria C, Sellers WR. Drug discovery approaches targeting the PI3K/Akt pathway in cancer. *Oncogene.* 2008;27(41):5511–5526. doi:10.1038/onc.2008.246
52. Maira SM, Stauffer F, Brueggen J, et al. Identification and characterization of NVP-BEZ235, a new orally available dual phosphatidylinositol 3-kinase/mammalian target of rapamycin inhibitor with potent in vivo antitumor activity. *Mol Cancer Ther.* 2008;7(7):1851–1863. doi:10.1158/1535-7163.MCT-08-0017
53. De P, Sun Y, Carlson JH, et al. Doubling down on the PI3K-AKT-mTOR pathway enhances the antitumor efficacy of PARP inhibitor in triple negative breast cancer model beyond BRCA-ness. *Neoplasia.* 2014;16(1):43–72. doi:10.1593/neo.131694
54. Speiser JJ, Ersahin C, Osipo C. The functional role of Notch signaling in triple-negative breast cancer. *Vitam Horm.* 2013;93:277–306.
55. Sukumar J, Gast K, Quiroga D, Lustberg M, Williams N. Triple-negative breast cancer: promising prognostic biomarkers currently in development. *Expert Rev Anticancer Ther.* 2021;21(2):135–148. doi:10.1080/14737140.2021.1840984
56. Hu Y, Gao J, Wang M, Li M. Potential Prospect of CDK4/6 Inhibitors in Triple-Negative Breast Cancer. *Cancer Manag Res.* 2021;13:5223–5237. doi:10.2147/CMAR.S310649
57. Finn RS, Crown JP, Lang I, Boer K. The cyclin-dependent kinase 4/6 inhibitor palbociclib in combination with letrozole versus letrozole alone as first-line treatment of oestrogen receptor-positive, HER2-negative, advanced breast cancer (PALOMA-1/TRIO-18): a randomised phase 2 study. *Lancet Oncol.* 2015;16(1):25–35. doi:10.1016/S1470-2045(14)71159-3
58. Raphael J, Helou J, Pritchard KI, Naimark DM. Palbociclib in hormone receptor positive advanced breast cancer: a cost-utility analysis. *Eur J Cancer.* 2017;85:146–154. doi:10.1016/j.ejca.2017.08.018
59. Katoh M. FGFR inhibitors: effects on cancer cells, tumor microenvironment and whole-body homeostasis (Review). *Int J Mol Med.* 2016;38(1):3–15. doi:10.3892/ijmm.2016.2620
60. Garcia J, Hurwitz HI, Sandler AB, Miles D. Bevacizumab (Avastin(R)) in cancer treatment: a review of 15 years of clinical experience and future outlook. *Cancer Treat Rev.* 2020;86:102017. doi:10.1016/j.ctrv.2020.102017
61. Mendel DB, Laird AD, Xin X, Louie SG, Schreck RE. In vivo antitumor activity of SU11248, a novel tyrosine kinase inhibitor targeting vascular endothelial growth factor and platelet-derived growth factor receptors: determination of a pharmacokinetic/pharmacodynamic relationship. *Clin Cancer Res.* 2003;9(1):327–337.
62. O'sullivan Coyne G, Burotto M. Clinical experience with ramucirumab: outcomes in breast cancer. *Expert Opin Biol Ther.* 2014;14(9):1351–1360. doi:10.1517/14712598.2014.939069
63. Bender RJ, Mac Gabhann F. Expression of VEGF and semaphorin genes define subgroups of triple negative breast cancer. *PLoS One.* 2013;8:e61788. doi:10.1371/journal.pone.0061788
64. Grothey A, Ellis LM. Targeting angiogenesis driven by vascular endothelial growth factors using antibody-based therapies. *Cancer J.* 2008;14(3):170–177. doi:10.1097/PPO.0b013e318178d9de
65. Gavine PR, Mooney L, Kilgour E, et al. AZD4547: an orally bioavailable, potent, and selective inhibitor of the fibroblast growth factor receptor tyrosine kinase family. *Cancer Res.* 2012;72(8):2045–2056. doi:10.1158/0008-5472.CAN-11-3034
66. Kwak Y, Cho H, Hur W, Sim T. Antitumor Effects and Mechanisms of AZD4547 on FGFR2-Deregulated Endometrial Cancer Cells. *Mol Cancer Ther.* 2015;14(10):2292–2302. doi:10.1158/1535-7163.MCT-15-0032
67. Guagnano V, Kauffmann A, Wohrle S, et al. FGFR genetic alterations predict for sensitivity to NVP-BGJ398, a selective pan-FGFR inhibitor. *Cancer Discov.* 2012;2(12):1118–1133. doi:10.1158/2159-8290.CD-12-0210
68. Hagel M, Miduturu C, Sheets M, et al. First Selective Small Molecule Inhibitor of FGFR4 for the Treatment of Hepatocellular Carcinomas with an Activated FGFR4 Signaling Pathway. *Cancer Discov.* 2015;5(4):424–437. doi:10.1158/2159-8290.CD-14-1029
69. Nakanishi Y, Akiyama N, Tsukaguchi T, et al. The fibroblast growth factor receptor genetic status as a potential predictor of the sensitivity to CH5183284/Debio 1347, a novel selective FGFR inhibitor. *Mol Cancer Ther.* 2014;13(11):2547–2558. doi:10.1158/1535-7163.MCT-14-0248
70. Lee SH, Lopes De Menezes D, Vora J, et al. In vivo Target Modulation and Biological Activity of CHIR-258, a Multitargeted Growth Factor Receptor Kinase Inhibitor, in Colon Cancer Models. *Clin Cancer Res.* 2005;11(10):3633–3641. doi:10.1158/1078-0432.CCR-04-2129
71. Tan L, Wang J, Huang Z, et al. Development of covalent inhibitors that can overcome resistance to first-generation FGFR kinase inhibitors. *Proc Natl Acad Sci U S A.* 2014;111(45):E4869–77. doi:10.1073/pnas.1403438111
72. Taberero J, Bahleda R, Dienstmann R, et al. Phase I Dose-Escalation Study of JNJ-42756493, an Oral Pan-Fibroblast Growth Factor Receptor Inhibitor, in Patients With Advanced Solid Tumors. *J Clin Oncol.* 2015;33(30):3401–3408. doi:10.1200/JCO.2014.60.7341
73. Zhao G, Li WY, Chen D, et al. A novel, selective inhibitor of fibroblast growth factor receptors that shows a potent broad spectrum of antitumor activity in several tumor xenograft models. *Mol Cancer Ther.* 2011;10(11):2200–2210. doi:10.1158/1535-7163.MCT-11-0306
74. O'hare T, Shakespeare WC, Zhu X, et al. AP24534, a pan-BCR-ABL inhibitor for chronic myeloid leukemia, potently inhibits the T315I mutant and overcomes mutation-based resistance. *Cancer Cell.* 2009;16(5):401–412. doi:10.1016/j.ccr.2009.09.028
75. Nicolo E, Boscolo Bielo L, Curigliano G, Tarantino P. The HER2-low revolution in breast oncology: steps forward and emerging challenges. *Ther Adv Med Oncol.* 2023;15:17588359231152842. doi:10.1177/17588359231152842

76. Hyman DM, Piha-Paul SA, Won H, et al. Author Correction: HER kinase inhibition in patients with HER2- and HER3-mutant cancers. *Nature*. 2019;566(7745):E11–E12. doi:10.1038/s41586-019-0974-0
77. Qiu Z, Oleinick NL, Zhang J. ATR/CHK1 inhibitors and cancer therapy. *Radiother Oncol*. 2018;126(3):450–464. doi:10.1016/j.radonc.2017.09.043
78. Witkiewicz AK, Chung S, Brough R, et al. Targeting the Vulnerability of RB Tumor Suppressor Loss in Triple-Negative Breast Cancer. *Cell Rep*. 2018;22(5):1185–1199. doi:10.1016/j.celrep.2018.01.022
79. Lee JM, Nair J, Zimmer A, et al. Prexasertib, a cell cycle checkpoint kinase 1 and 2 inhibitor, in BRCA wild-type recurrent high-grade serous ovarian cancer: a first-in-class proof-of-concept phase 2 study. *Lancet Oncol*. 2018;19(2):207–215. doi:10.1016/S1470-2045(18)30009-3
80. Bennett CN, Tomlinson CC, Michalowski AM, et al. Cross-species genomic and functional analyses identify a combination therapy using a CHK1 inhibitor and a ribonucleotide reductase inhibitor to treat triple-negative breast cancer. *Breast Cancer Res*. 2012;14(4):R109. doi:10.1186/bcr3230
81. Guertin AD, Li J, Liu Y, et al. Preclinical evaluation of the WEE1 inhibitor MK-1775 as single-agent anticancer therapy. *Mol Cancer Ther*. 2013;12(8):1442–1452. doi:10.1158/1535-7163.MCT-13-0025
82. Jia H, Truica CI, Wang Y, et al. Immunotherapy for triple-negative breast cancer: existing challenges and exciting prospects. *Drug Resist Updat*. 2017;32:1–15. doi:10.1016/j.drug.2017.07.002
83. Walter S, Weinschenk T, Stenzl A, et al. Multi-peptide immune response to cancer vaccine IMA901 after single-dose cyclophosphamide associates with longer patient survival. *Nat Med*. 2012;18(8):1254–1261. doi:10.1038/nm.2883
84. Arce-Salinas C, Riesco-Martinez MC, Hanna W, Bedard P, Warner E. Complete Response of Metastatic Androgen Receptor-Positive Breast Cancer to Bicalutamide: case Report and Review of the Literature. *J Clin Oncol*. 2016;34:e21–4. doi:10.1200/JCO.2013.49.8899
85. Xu N, Li B, Liu Y, et al. Ferroptosis and triple-negative breast cancer: potential therapeutic targets. *Front Oncol*. 2022;12:1017041. doi:10.3389/fonc.2022.1017041
86. Garrido-Castro AC, Lin NU, Polyak K. Insights into Molecular Classifications of Triple-Negative Breast Cancer: improving Patient Selection for Treatment. *Cancer Discov*. 2019;9(2):176–198. doi:10.1158/2159-8290.CD-18-1177
87. Bardia A, Mayer IA, Diamond JR, et al. Efficacy and Safety of Anti-Trop-2 Antibody Drug Conjugate Sacituzumab Govitecan (IMMU-132) in Heavily Pretreated Patients With Metastatic Triple-Negative Breast Cancer. *J Clin Oncol*. 2017;35(19):2141–2148. doi:10.1200/JCO.2016.70.8297
88. Yardley DA, Weaver R, Melisko ME, et al. EMERGE: a Randomized Phase II Study of the Antibody-Drug Conjugate Glemtatumumab Vedotin in Advanced Glycoprotein NMB-Expressing Breast Cancer. *J Clin Oncol*. 2015;33(14):1609–1619. doi:10.1200/JCO.2014.56.2959
89. Nagayama A, Vidula N, Bardia A. Novel Therapies for Metastatic Triple-Negative Breast Cancer: spotlight on Immunotherapy and Antibody-Drug Conjugates. *Oncology*. 2021;35(3505):249–254. doi:10.46883/ONC.2021.3505.0249
90. Gristina V, La Mantia M, Iacono F, Galvano A, Russo A, Bazan V. The Emerging Therapeutic Landscape of ALK Inhibitors in Non-Small Cell Lung Cancer. *Pharmaceuticals*. 2020;13(12):474. doi:10.3390/ph13120474
91. Dong S, Yousefi H, Iv S, et al. Ceritinib is a novel triple negative breast cancer therapeutic agent. *Mol Cancer*. 2022;21(1):138. doi:10.1186/s12943-022-01601-0
92. Kim LC, Song L, Haura EB. Src kinases as therapeutic targets for cancer. *Nat Rev Clin Oncol*. 2009;6(10):587–595. doi:10.1038/nrclinonc.2009.129
93. Wheeler DL, Iida M, Dunn EF. The role of Src in solid tumors. *Oncologist*. 2009;14(7):667–678. doi:10.1634/theoncologist.2009-0009
94. Finn RS, Dering J, Ginther C, et al. Dasatinib, an orally active small molecule inhibitor of both the src and abl kinases, selectively inhibits growth of basal-type/“triple-negative” breast cancer cell lines growing in vitro. *Breast Cancer Res Treat*. 2007;105(3):319–326. doi:10.1007/s10549-006-9463-x
95. Huang F, Reeves K, Han X, et al. Identification of candidate molecular markers predicting sensitivity in solid tumors to dasatinib: rationale for patient selection. *Cancer Res*. 2007;67(5):2226–2238. doi:10.1158/0008-5472.CAN-06-3633
96. Tryfonopoulos D, Walsh S, Collins DM, et al. Src: a potential target for the treatment of triple-negative breast cancer. *Ann Oncol*. 2011;22(10):2234–2240. doi:10.1093/annonc/mdq757
97. Quereda V, Bayle S, Vena F, et al. Therapeutic Targeting of CDK12/CDK13 in Triple-Negative Breast Cancer. *Cancer Cell*. 2019;36(5):545–558e7. doi:10.1016/j.ccell.2019.09.004
98. Cortesi L, Rugo HS, Jackisch C. An Overview of PARP Inhibitors for the Treatment of Breast Cancer. *Target Oncol*. 2021;16(3):255–282. doi:10.1007/s11523-021-00796-4
99. Rizzolo P, Silvestri V, Falchetti M, Ottini L. Inherited and acquired alterations in development of breast cancer. *Appl Clin Genet*. 2011;4:145–158. doi:10.2147/TACG.S13226
100. Godet I, Gilkes DM. BRCA1 and BRCA2 mutations and treatment strategies for breast cancer. *Integr Cancer Sci Ther*. 2017;4(1). doi:10.15761/ICST.1000228
101. Winter C, Nilsson MP, Olsson E, et al. Targeted sequencing of BRCA1 and BRCA2 across a large unselected breast cancer cohort suggests that one-third of mutations are somatic. *Ann Oncol*. 2016;27(8):1532–1538. doi:10.1093/annonc/mdw209
102. Lord CJ, Ashworth A. The DNA damage response and cancer therapy. *Nature*. 2012;481(7381):287–294. doi:10.1038/nature10760
103. Perou CM, Sorlie T, Eisen MB, et al. Molecular portraits of human breast tumours. *Nature*. 2000;406(6797):747–752. doi:10.1038/35021093
104. Walsh CS. Two decades beyond BRCA1/2: homologous recombination, hereditary cancer risk and a target for ovarian cancer therapy. *Gynecol Oncol*. 2015;137(2):343–350. doi:10.1016/j.ygyno.2015.02.017
105. Bai X, Ni J, Beretov J, Graham P, Li Y. Triple-negative breast cancer therapeutic resistance: where is the Achilles’ heel? *Cancer Lett*. 2021;497:100–111. doi:10.1016/j.canlet.2020.10.016
106. Tutt A, Robson M, Garber JE, et al. Oral poly(ADP-ribose) polymerase inhibitor olaparib in patients with BRCA1 or BRCA2 mutations and advanced breast cancer: a proof-of-concept trial. *Lancet*. 2010;376(9737):235–244. doi:10.1016/S0140-6736(10)60892-6
107. Jin J, Zhang W, Ji W, Yang F, Guan X. Predictive biomarkers for triple negative breast cancer treated with platinum-based chemotherapy. *Cancer Biol Ther*. 2017;18(6):369–378. doi:10.1080/15384047.2017.1323582
108. Cantley LC. The phosphoinositide 3-kinase pathway. *Science*. 2002;296(5573):1655–1657. doi:10.1126/science.296.5573.1655

109. Cossu-Rocca P, Orru S, Muroli MR, et al. Analysis of PIK3CA Mutations and Activation Pathways in Triple Negative Breast Cancer. *PLoS One*. 2015;10(11):e0141763. doi:10.1371/journal.pone.0141763
110. Ooms LM, Binge LC, Davies EM, et al. The Inositol Polyphosphate 5-Phosphatase PIPP Regulates AKT1-Dependent Breast Cancer Growth and Metastasis. *Cancer Cell*. 2015;28(2):155–169. doi:10.1016/j.ccell.2015.07.003
111. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646–674. doi:10.1016/j.cell.2011.02.013
112. Asghar U, Witkiewicz AK, Turner NC, Knudsen ES. The history and future of targeting cyclin-dependent kinases in cancer therapy. *Nat Rev Drug Discov*. 2015;14(2):130–146. doi:10.1038/nrd4504
113. Curtis C, Shah SP, Chin SF, et al. The genomic and transcriptomic architecture of 2,000 breast tumours reveals novel subgroups. *Nature*. 2012;486(7403):346–352. doi:10.1038/nature10983
114. Finn RS, Dering J, Conklin D, et al. PD 0332991, a selective cyclin D kinase 4/6 inhibitor, preferentially inhibits proliferation of luminal estrogen receptor-positive human breast cancer cell lines in vitro. *Breast Cancer Res*. 2009;11(5):R77. doi:10.1186/bcr2419
115. Liu T, Yu J, Deng M, et al. CDK4/6-dependent activation of DUB3 regulates cancer metastasis through SNAIL1. *Nat Commun*. 2017;8(1):13923. doi:10.1038/ncomms13923
116. Yamamoto T, Kanaya N, Somlo G, Chen S. Synergistic anti-cancer activity of CDK4/6 inhibitor palbociclib and dual mTOR kinase inhibitor MLN0128 in pRb-expressing ER-negative breast cancer. *Breast Cancer Res Treat*. 2019;174(3):615–625. doi:10.1007/s10549-018-05104-9
117. Yuan Y, Wen W, Yost SE, Xing Q, Yan J, Mortimer J. Combination therapy with BYL719 and LEE011 is synergistic and causes a greater suppression of p-S6 in triple negative breast cancer. *Sci Rep*. 2019;9(1):7509. doi:10.1038/s41598-019-43429-7
118. Rakha EA, El-Sayed ME, Green AR, Lee AH, Robertson JF, Ellis IO. Prognostic markers in triple-negative breast cancer. *Cancer*. 2007;109(1):25–32. doi:10.1002/ncr.22381
119. Tan DS, Marchio C, Jones RL, Savage K, Dowsett M. Triple negative breast cancer: molecular profiling and prognostic impact in adjuvant anthracycline-treated patients. *Breast Cancer Res Treat*. 2008;111(1):27–44. doi:10.1007/s10549-007-9756-8
120. Choi J, Jung WH, Koo JS. Clinicopathologic features of molecular subtypes of triple negative breast cancer based on immunohistochemical markers. *Histol Histopathol*. 2012;27(11):1481–1493. doi:10.14670/HH-27.1481
121. Gumuskaya B, Alper M, Hucumenoglu S, Altundag K, Uner A, Guler G. EGFR expression and gene copy number in triple-negative breast carcinoma. *Cancer Genet Cytogenet*. 2010;203(2):222–229. doi:10.1016/j.cancergencyto.2010.07.118
122. Martin V, Botta F, Zanellato E, et al. Molecular characterization of EGFR and EGFR-downstream pathways in triple negative breast carcinomas with basal like features. *Histol Histopathol*. 2012;27(6):785–792. doi:10.14670/HH-27.785
123. Reis-Filho JS, Pinheiro C, Lambros MB, Milanezi F, Carvalho S, Savage K. EGFR amplification and lack of activating mutations in metaplastic breast carcinomas. *J Pathol*. 2006;209(4):445–453. doi:10.1002/path.2004
124. Grob TJ, Heilenkotter U, Geist S, et al. Rare oncogenic mutations of predictive markers for targeted therapy in triple-negative breast cancer. *Breast Cancer Res Treat*. 2012;134(2):561–567. doi:10.1007/s10549-012-2092-7
125. Shao MM, Zhang F, Meng G, Xu H. Epidermal growth factor receptor gene amplification and protein overexpression in basal-like carcinoma of the breast. *Histopathology*. 2011;59(2):264–273. doi:10.1111/j.1365-2559.2011.03921.x
126. Nakajima H, Ishikawa Y, Furuya M, et al. Protein expression, gene amplification, and mutational analysis of EGFR in triple-negative breast cancer. *Breast Cancer*. 2014;21(1):66–74. doi:10.1007/s12282-012-0354-1
127. Bernsdorf M, Ingvar C, Jorgensen L, et al. Effect of adding gefitinib to neoadjuvant chemotherapy in estrogen receptor negative early breast cancer in a randomized phase II trial. *Breast Cancer Res Treat*. 2011;126(2):463–470. doi:10.1007/s10549-011-1352-2
128. Matsuda N, Wang X, Lim B, et al. Safety and Efficacy of Panitumumab Plus Neoadjuvant Chemotherapy in Patients With Primary HER2-Negative Inflammatory Breast Cancer. *JAMA Oncol*. 2018;4(9):1207–1213. doi:10.1001/jamaoncol.2018.1436
129. Schuler M, Awada A, Harter P, et al. A phase II trial to assess efficacy and safety of Afatinib in extensively pretreated patients with HER2-negative metastatic breast cancer. *Breast Cancer Res Treat*. 2012;134(3):1149–1159. doi:10.1007/s10549-012-2126-1
130. Yardley DA, Ward PJ, Daniel BR, Eakle JF. Panitumumab, Gemcitabine, and Carboplatin as Treatment for Women With Metastatic Triple-Negative Breast Cancer: a Sarah Cannon Research Institute Phase II Trial. *Clin Breast Cancer*. 2016;16(5):349–355. doi:10.1016/j.clbc.2016.05.006
131. Greenberg S, Rugo HS. Triple-negative breast cancer: role of antiangiogenic agents. *Cancer J*. 2010;16(1):33–38. doi:10.1097/PPO.0b013e3181d38514
132. Papa A, Caruso D, Tomao S, Rossi L, Zaccarelli E, Tomao F. Triple-negative breast cancer: investigating potential molecular therapeutic target. *Expert Opin Ther Targets*. 2015;19(1):55–75. doi:10.1517/14728222.2014.970176
133. Da Silva JL, Cardoso Nunes NC, Izetti P, De Mesquita GG, De Melo AC. Triple negative breast cancer: a thorough review of biomarkers. *Crit Rev Oncol Hematol*. 2020;145:102855. doi:10.1016/j.critrevonc.2019.102855
134. Linderholm BK, Hellborg H, Johansson U, et al. Significantly higher levels of vascular endothelial growth factor (VEGF) and shorter survival times for patients with primary operable triple-negative breast cancer. *Ann Oncol*. 2009;20(10):1639–1646. doi:10.1093/annonc/mdp062
135. Von Minckwitz G, Eidtmann H, Rezai M, et al. German Breast Group, Arbeitsgemeinschaft Gynakologische Onkologie-Breast Study Groups. Neoadjuvant chemotherapy and bevacizumab for HER2-negative breast cancer. *N Engl J Med*. 2012;366(4):299–309. doi:10.1056/NEJMoa1111065
136. Pal SK, Mortimer J. Triple-negative breast cancer: novel therapies and new directions. *Maturitas*. 2009;63(4):269–274. doi:10.1016/j.maturitas.2009.06.010
137. Giulianelli S, Cerliani JP, Lamb CA, et al. Carcinoma-associated fibroblasts activate progesterone receptors and induce hormone independent mammary tumor growth: a role for the FGF-2/FGFR-2 axis. *Int J Cancer*. 2008;123(11):2518–2531. doi:10.1002/ijc.23802
138. Katoh M. Cancer genomics and genetics of FGFR2 (Review). *Int J Oncol*. 2008;33(2):233–237.
139. Eswarakumar VP, Lax I, Schlessinger J. Cellular signaling by fibroblast growth factor receptors. *Cytokine Growth Factor Rev*. 2005;16(2):139–149. doi:10.1016/j.cytogfr.2005.01.001
140. Katoh M, Nakagama H. FGF receptors: cancer biology and therapeutics. *Med Res Rev*. 2014;34(2):280–300. doi:10.1002/med.21288
141. Ornitz DM, Itoh N. The Fibroblast Growth Factor signaling pathway. *Wiley Interdiscip Rev Dev Biol*. 2015;4(3):215–266. doi:10.1002/wdev.176

142. Jafarian AH, Kooshkiforooshani M, Farzad F, Mohamadian Roshan N. The Relationship Between Fibroblastic Growth Factor Receptor-1 (FGFR1) Gene Amplification in Triple Negative Breast Carcinomas and Clinicopathological Prognostic Factors. *Iran J Pathol.* 2019;14(4):299–304. doi:10.30699/IJP.2019.96713.1952
143. Perez-Garcia J, Munoz-Couselo E, Soberino J, Racca F, Cortes J. Targeting FGFR pathway in breast cancer. *Breast.* 2018;37:126–133. doi:10.1016/j.breast.2017.10.014
144. Soria JC, Debraud F, Bahleda R, et al. Corrections to “Phase I/IIa study evaluating the safety, efficacy, pharmacokinetics, and pharmacodynamics of lucitanib in advanced solid tumors”. *Ann Oncol.* 2015;26(2):445. doi:10.1093/annonc/mdl547
145. Nogova L, Sequist LV, Perez Garcia JM, et al. Evaluation of BGJ398, a Fibroblast Growth Factor Receptor 1-3 Kinase Inhibitor, in Patients With Advanced Solid Tumors Harboring Genetic Alterations in Fibroblast Growth Factor Receptors: results of a Global Phase I, Dose-Escalation and Dose-Expansion Study. *J Clin Oncol.* 2017;35(2):157–165. doi:10.1200/JCO.2016.67.2048
146. Bai A, Meetze K, Vo NY, et al. GP369, an FGFR2-IIIb-specific antibody, exhibits potent antitumor activity against human cancers driven by activated FGFR2 signaling. *Cancer Res.* 2010;70(19):7630–7639. doi:10.1158/0008-5472.CAN-10-1489
147. Tolcher AW, Papadopoulos KP, Patnaik A, et al. A phase I, first in human study of FP-1039 (GSK3052230), a novel FGF ligand trap, in patients with advanced solid tumors. *Ann Oncol.* 2016;27(3):526–532. doi:10.1093/annonc/mdv591
148. Lyons TG. Targeted Therapies for Triple-Negative Breast Cancer. *Curr Treat Options Oncol.* 2019;20(11):82. doi:10.1007/s11864-019-0682-x
149. Tarantino P, Hamilton E, Tolaney SM, et al. HER2-Low Breast Cancer: pathological and Clinical Landscape. *J Clin Oncol.* 2020;38(17):1951–1962. doi:10.1200/JCO.19.02488
150. Mittendorf EA, Ardavani A, Litton JK, et al. Primary analysis of a prospective, randomized, single-blinded phase II trial evaluating the HER2 peptide vaccine in breast cancer patients to prevent recurrence. *Oncotarget.* 2016;7(40):66192–66201. doi:10.18632/oncotarget.11751
151. Guler G, Himmetoglu C, Jimenez RE, et al. Aberrant expression of DNA damage response proteins is associated with breast cancer subtype and clinical features. *Breast Cancer Res Treat.* 2011;129(2):421–432. doi:10.1007/s10549-010-1248-6
152. Ribeiro E, Ganzinelli M, Andreis D, et al. Triple negative breast cancers have a reduced expression of DNA repair genes. *PLoS One.* 2013;8(6):e66243. doi:10.1371/journal.pone.0066243
153. Shah SP, Roth A, Goya R, et al. The clonal and mutational evolution spectrum of primary triple-negative breast cancers. *Nature.* 2012;486(7403):395–399. doi:10.1038/nature10933
154. Curtin NJ. DNA repair dysregulation from cancer driver to therapeutic target. *Nat Rev Cancer.* 2012;12(12):801–817. doi:10.1038/nrc3399
155. Albiges L, Goubar A, Scott V, et al. Chk1 as a new therapeutic target in triple-negative breast cancer. *Breast.* 2014;23(3):250–258. doi:10.1016/j.breast.2014.02.004
156. Nanda R, Chow LQ, Dees EC, et al. Pembrolizumab in Patients With Advanced Triple-Negative Breast Cancer: phase Ib KEYNOTE-012 Study. *J Clin Oncol.* 2016;34(21):2460–2467. doi:10.1200/JCO.2015.64.8931
157. Vesely MD, Kershaw MH, Schreiber RD, Smyth MJ. Natural innate and adaptive immunity to cancer. *Annu Rev Immunol.* 2011;29(1):235–271. doi:10.1146/annurev-immunol-031210-101324
158. Riley JL. PD-1 signaling in primary T cells. *Immunol Rev.* 2009;229(1):114–125. doi:10.1111/j.1600-065X.2009.00767.x
159. Shi L, Chen S, Yang L, Li Y. The role of PD-1 and PD-L1 in T-cell immune suppression in patients with hematological malignancies. *J Hematol Oncol.* 2013;6(1):74. doi:10.1186/1756-8722-6-74
160. Shih K, Arkenau HT, Infante JR. Clinical impact of checkpoint inhibitors as novel cancer therapies. *Drugs.* 2014;74(17):1993–2013. doi:10.1007/s40265-014-0305-6
161. Garon EB. Current Perspectives in Immunotherapy for Non-Small Cell Lung Cancer. *Semin Oncol.* 2015;42 Suppl 2:S11–8. doi:10.1053/j.seminoncol.2015.09.019
162. Rampurwala M, Wisinski KB, O’regan R. Role of the androgen receptor in triple-negative breast cancer. *Clin Adv Hematol Oncol.* 2016;14(3):186–193.
163. Barton VN, D’amato NC, Gordon MA, et al. Multiple Molecular Subtypes of Triple-Negative Breast Cancer Critically Rely on Androgen Receptor and Respond to Enzalutamide In Vivo. *Mol Cancer Ther.* 2015;14(3):769–778. doi:10.1158/1535-7163.MCT-14-0926
164. Barton VN, D’amato NC, Gordon MA, L CJ, Elias A, K RJ. Androgen Receptor Biology in Triple Negative Breast Cancer: a Case for Classification as AR+ or Quadruple Negative Disease. *Horm Cancer.* 2015;6(5–6):206–213. doi:10.1007/s12672-015-0232-3
165. Lebert JM, Lester R, Powell E, Seal M, Mccarthy J. Advances in the systemic treatment of triple-negative breast cancer. *Curr Oncol.* 2018;25:S142–S150. doi:10.3747/co.25.3954
166. McGrogan BT, Gilmartin B, Carney DN, Mccann A. Taxanes, microtubules and chemoresistant breast cancer. *Biochim Biophys Acta.* 2008;1785(2):96–132. doi:10.1016/j.bbcan.2007.10.004
167. Carey LA, Dees EC, Sawyer L, et al. The triple negative paradox: primary tumor chemosensitivity of breast cancer subtypes. *Clin Cancer Res.* 2007;13(8):2329–2334. doi:10.1158/1078-0432.CCR-06-1109
168. Koh SB, Ross K, Isakoff SJ, et al. RASAL2 Confers Collateral MEK/EGFR Dependency in Chemoresistant Triple-Negative Breast Cancer. *Clin Cancer Res.* 2021;27(17):4883–4897. doi:10.1158/1078-0432.CCR-21-0714
169. Lemesle M, Geoffroy M, Alpy F, Tomasetto CL, Kuntz S, Grillier-Vuissoz I. CLDN1 Sensitizes Triple-Negative Breast Cancer Cells to Chemotherapy. *Cancers.* 2022;15(1):14. doi:10.3390/cancers15010014
170. Xiong Z, Yang L, Li N, et al. DAB2IP attenuates chemoresistance of triple-negative breast cancer through sequestration of RAC1 to prevent beta-catenin nuclear accumulation. *Clin Transl Med.* 2022;12(12):e1133. doi:10.1002/ctm2.1133
171. Chen KS, Deluca HF. Cloning of the human 1 alpha,25-dihydroxyvitamin D-3 24-hydroxylase gene promoter and identification of two vitamin D-responsive elements. *Biochim Biophys Acta.* 1995;1263(1):1–9. doi:10.1016/0167-4781(95)00060-t
172. Nasoohi S, Ismael S, Ishrat T. Thioredoxin-Interacting Protein (TXNIP) in Cerebrovascular and Neurodegenerative Diseases: regulation and Implication. *Mol Neurobiol.* 2018;55(10):7900–7920. doi:10.1007/s12035-018-0917-z
173. Chen Y, Feng X, Yuan Y, Jiang J, Zhang P, Zhang B. Identification of a novel mechanism for reversal of doxorubicin-induced chemotherapy resistance by TXNIP in triple-negative breast cancer via promoting reactive oxygen-mediated DNA damage. *Cell Death Dis.* 2022;13(4):338. doi:10.1038/s41419-022-04783-z
174. Gottesman MM, Fojo T, Bates SE. Multidrug resistance in cancer: role of ATP-dependent transporters. *Nat Rev Cancer.* 2002;2(1):48–58. doi:10.1038/nrc706

175. Salaroglio IC, Gazzano E, Abdullrahman A, et al. Increasing intratumor C/EBP-beta LIP and nitric oxide levels overcome resistance to doxorubicin in triple negative breast cancer. *J Exp Clin Cancer Res.* 2018;37(1):286. doi:10.1186/s13046-018-0967-0
176. Denard B, Jiang S, Peng Y, Ye J. CREB3L1 as a potential biomarker predicting response of triple negative breast cancer to doxorubicin-based chemotherapy. *BMC Cancer.* 2018;18(1):813. doi:10.1186/s12885-018-4724-8
177. Long F, Lin H, Zhang X, Zhang J, Xiao H, Wang T. Attractylenolide-I Suppresses Tumorigenesis of Breast Cancer by Inhibiting Toll-Like Receptor 4-Mediated Nuclear Factor-kappaB Signaling Pathway. *Front Pharmacol.* 2020;11:598939. doi:10.3389/fphar.2020.598939
178. Wang M, Li XZ, Zhang MX, Ye QY, Chen YX, Chang X. Attractylenolide-I Sensitizes Triple-Negative Breast Cancer Cells to Paclitaxel by Blocking CTGF Expression and Fibroblast Activation. *Front Oncol.* 2021;11:738534. doi:10.3389/fonc.2021.738534
179. Al-Marsoumi S, Vomhof-Dekrey E, Basson MD. Schlafen12 Reduces the Aggressiveness of Triple Negative Breast Cancer through Post-Transcriptional Regulation of ZEB1 That Drives Stem Cell Differentiation. *Cell Physiol Biochem.* 2019;53:999–1014. doi:10.33594/00000191
180. Raafat Elsayed AA, Al-Marsoumi S, Vomhof-Dekrey EE, Basson MD. SLFN12 Over-expression Sensitizes Triple Negative Breast Cancer Cells to Chemotherapy Drugs and Radiotherapy. *Cancer Genomics Proteomics.* 2022;19(3):328–338. doi:10.21873/cgp.20323
181. Guo H, Treude F, Kramer OH, Luscher B, Hartkamp J. PAR-4 overcomes chemo-resistance in breast cancer cells by antagonizing cIAP1. *Sci Rep.* 2019;9(1):8755. doi:10.1038/s41598-019-45209-9
182. Zhang Y, Wu J, Ye M, et al. ETS1 is associated with cisplatin resistance through IKKalpha/NF-kappaB pathway in cell line MDA-MB-231. *Cancer Cell Int.* 2018;18(1):86. doi:10.1186/s12935-018-0581-4
183. Fan M, Gao J, Zhou L, et al. Highly expressed SERCA2 triggers tumor cell autophagy and is a druggable vulnerability in triple-negative breast cancer. *Acta Pharm Sin B.* 2022;12(12):4407–4423. doi:10.1016/j.apsb.2022.05.009
184. Qiao H, Rao H, Yun Y, et al. Impeding DNA Break Repair Enables Oocyte Quality Control. *Mol Cell.* 2018;72:211–221e3. doi:10.1016/j.molcel.2018.08.031
185. Adelaide J, Finetti P, Bekhouche I, et al. Integrated profiling of basal and luminal breast cancers. *Cancer Res.* 2007;67(24):11565–11575. doi:10.1158/0008-5472.CAN-07-2536
186. Chen YT, Venditti CA, Theiler G, et al. Identification of CT46/HORMAD1, an immunogenic cancer/testis antigen encoding a putative meiosis-related protein. *Cancer Immun.* 2005;5:9.
187. Zong B, Sun L, Peng Y, Lei J, Zhang Y, Guo S. HORMAD1 promotes docetaxel resistance in triple negative breast cancer by enhancing DNA damage tolerance. *Oncol Rep.* 2021;45(4):46. doi:10.3892/or.2021.7997
188. Lin HY, Kuei CH, Lee HH, et al. TNFSF13 upregulation confers chemotherapeutic resistance via triggering autophagy initiation in triple-negative breast cancer. *J Mol Med (Berl).* 2020;98(9):1255–1267. doi:10.1007/s00109-020-01952-5
189. Liao L, Zhang YL, Deng L, et al. Protein Phosphatase 1 Subunit PPP1R14B Stabilizes STMN1 to Promote Progression and Paclitaxel Resistance in Triple-Negative Breast Cancer. *Cancer Res.* 2023;83(3):471–484. doi:10.1158/0008-5472.CAN-22-2709
190. Gobel C, Schroeder C, Hube-Magg C, et al. Upregulation of centromere protein F is linked to aggressive prostate cancers. *Cancer Manag Res.* 2018;10:5491–5504. doi:10.2147/CMAR.S165630
191. Chong T, Sarac A, Yao CQ, et al. Deregulation of the spindle assembly checkpoint is associated with paclitaxel resistance in ovarian cancer. *J Ovarian Res.* 2018;11(1):27. doi:10.1186/s13048-018-0399-7
192. Wang D, Xu W, Huang M, et al. CENPF knockdown inhibits Adriamycin chemoresistance in triple-negative breast cancer via the Rb-E2F1 axis. *Sci Rep.* 2023;13(1):1803. doi:10.1038/s41598-023-28355-z
193. Xiao YS, Zeng D, Liang YK, et al. Major vault protein is a direct target of Notch1 signaling and contributes to chemoresistance in triple-negative breast cancer cells. *Cancer Lett.* 2019;440-441:156–167. doi:10.1016/j.canlet.2018.09.031
194. Mao M, Jia Y, Chen Y, et al. HJURP regulates cell proliferation and chemo-resistance via YAP1/NDRG1 transcriptional axis in triple-negative breast cancer. *Cell Death Dis.* 2022;13(4):396. doi:10.1038/s41419-022-04833-6
195. Deng N, Zhang X, Zhang Y. BAIAP2L1 accelerates breast cancer progression and chemoresistance by activating AKT signaling through binding with ribosomal protein L3. *Cancer Sci.* 2023;114(3):764–780. doi:10.1111/cas.15632
196. Liu XY, Jiang W, Ma D, et al. SYTL4 downregulates microtubule stability and confers paclitaxel resistance in triple-negative breast cancer. *Theranostics.* 2020;10(24):10940–10956. doi:10.7150/thno.45207
197. Oudin MJ, Barbier L, Schafer C, et al. MENA Confers Resistance to Paclitaxel in Triple-Negative Breast Cancer. *Mol Cancer Ther.* 2017;16(1):143–155. doi:10.1158/1535-7163.MCT-16-0413
198. Yang N, Wang C, Wang J, et al. Aurora kinase A stabilizes FOXM1 to enhance paclitaxel resistance in triple-negative breast cancer. *J Cell Mol Med.* 2019;23(9):6442–6453. doi:10.1111/jcmm.14538
199. Zhou H, Zhang Y, Fu Y, Chan L, Lee AS. Novel mechanism of anti-apoptotic function of 78-kDa glucose-regulated protein (GRP78): endocrine resistance factor in breast cancer, through release of B-cell lymphoma 2 (BCL-2) from BCL-2-interacting killer (BIK). *J Biol Chem.* 2011;286(29):25687–25696. doi:10.1074/jbc.M110.212944
200. Liu M, Yang J, Lv W, et al. Down-regulating GRP78 reverses pirarubicin resistance of triple negative breast cancer by miR-495-3p mimics and involves the p-AKT/mTOR pathway. *Biosci Rep.* 2022;42:1.
201. Ha H, Debnath B, Neamat N. Role of the CXCL8-CXCR1/2 Axis in Cancer and Inflammatory Diseases. *Theranostics.* 2017;7(6):1543–1588. doi:10.7150/thno.15625
202. Yi M, Peng C, Xia B, Gan L. CXCL8 Facilitates the Survival and Paclitaxel-Resistance of Triple-Negative Breast Cancers. *Clin Breast Cancer.* 2022;22(2):e191–e198. doi:10.1016/j.clbc.2021.06.009
203. Biswas D, Slade L, Duffley L, et al. Inhibiting BCKDK in triple negative breast cancer suppresses protein translation, impairs mitochondrial function, and potentiates doxorubicin cytotoxicity. *Cell Death Discov.* 2021;7(1):241. doi:10.1038/s41420-021-00602-0
204. Byerly JH, Port ER, Irie HY. PRKCQ inhibition enhances chemosensitivity of triple-negative breast cancer by regulating Bim. *Breast Cancer Res.* 2020;22(1):72. doi:10.1186/s13058-020-01302-w
205. Wei W, Zou Y, Jiang Q, et al. PSMB5 is associated with proliferation and drug resistance in triple-negative breast cancer. *Int J Biol Markers.* 2018;33(1):102–108. doi:10.5301/ijbm.5000283
206. Lai H, Wang R, Li S, et al. LIN9 confers paclitaxel resistance in triple negative breast cancer cells by upregulating CCSAP. *Sci China Life Sci.* 2020;63(3):419–428. doi:10.1007/s11427-019-9581-8

207. Sun S, Wu H, Wu X, et al. Silencing of PGK1 Promotes Sensitivity to Paclitaxel Treatment by Upregulating XAF1-Mediated Apoptosis in Triple-Negative Breast Cancer. *Front Oncol.* 2021;11:535230. doi:10.3389/fonc.2021.535230
208. Zhang Q, Lei L, Jing D. Knockdown of SERPINE1 reverses resistance of triple-negative breast cancer to paclitaxel via suppression of VEGFA. *Oncol Rep.* 2020;44(5):1875–1884. doi:10.3892/or.2020.7770
209. Xie XQ, Yang Y, Wang Q, et al. Targeting ATAD3A-PINK1-mitophagy axis overcomes chemoimmunotherapy resistance by redirecting PD-L1 to mitochondria. *Cell Res.* 2023;33(3):215–228. doi:10.1038/s41422-022-00766-z
210. Liu T, Sun H, Zhu D, et al. TRA2A Promoted Paclitaxel Resistance and Tumor Progression in Triple-Negative Breast Cancers via Regulating Alternative Splicing. *Mol Cancer Ther.* 2017;16(7):1377–1388. doi:10.1158/1535-7163.MCT-17-0026
211. Fusco A, Fedele M. Roles of HMGA proteins in cancer. *Nat Rev Cancer.* 2007;7(12):899–910. doi:10.1038/nrc2271
212. Sgubin M, Pegoraro S, Pellarin I, et al. HMGA1 positively regulates the microtubule-destabilizing protein stathmin promoting motility in TNBC cells and decreasing tumour sensitivity to paclitaxel. *Cell Death Dis.* 2022;13(5):429. doi:10.1038/s41419-022-04843-4
213. Samanta S, Pursell B, Mercurio AM. IMP3 protein promotes chemoresistance in breast cancer cells by regulating breast cancer resistance protein (ABCG2) expression. *J Biol Chem.* 2013;288(18):12569–12573. doi:10.1074/jbc.C112.442319
214. Haynes BM, Cunningham K, Shekhar MPV. RAD6 inhibition enhances paclitaxel sensitivity of triple negative breast cancer cells by aggravating mitotic spindle damage. *BMC Cancer.* 2022;22(1):1073. doi:10.1186/s12885-022-10119-z
215. Luo M, Fu A, Wu R, et al. High Expression of G6PD Increases Doxorubicin Resistance in Triple Negative Breast Cancer Cells by Maintaining GSH Level. *Int J Biol Sci.* 2022;18(3):1120–1133. doi:10.7150/ijbs.65555
216. Lee J, Gollahon L. Nek2-targeted ASO or siRNA pretreatment enhances anticancer drug sensitivity in triple-negative breast cancer cells. *Int J Oncol.* 2013;42(3):839–847. doi:10.3892/ijo.2013.1788
217. Shen H, Yan W, Yuan J, Wang Z, Wang C. Nek2B activates the wnt pathway and promotes triple-negative breast cancer chemothezrapy-resistance by stabilizing beta-catenin. *J Exp Clin Cancer Res.* 2019;38(1):243. doi:10.1186/s13046-019-1231-y
218. Zhu S, Xu Y, Wang L, et al. Ceramide kinase mediates intrinsic resistance and inferior response to chemotherapy in triple-negative breast cancer by upregulating Ras/ERK and PI3K/Akt pathways. *Cancer Cell Int.* 2021;21(1):42. doi:10.1186/s12935-020-01735-5
219. Fernandez NB, Sosa SM, Roberts JT, et al. RUNX1 Is Regulated by Androgen Receptor to Promote Cancer Stem Markers and Chemotherapy Resistance in Triple Negative Breast Cancer. *Cells.* 2023;13(1):12. doi:10.3390/cells13010012
220. Ogbu SC, Rojas S, Weaver J, et al. DSTYK Enhances Chemoresistance in Triple-Negative Breast Cancer Cells. *Cells.* 2021;11(1):11. doi:10.3390/cells11010011
221. Hashimoto Y, Koyama K, Kamai Y, et al. A Novel HER3-Targeting Antibody-Drug Conjugate, U3-1402, Exhibits Potent Therapeutic Efficacy through the Delivery of Cytotoxic Payload by Efficient Internalization. *Clin Cancer Res.* 2019;25(23):7151–7161. doi:10.1158/1078-0432.CCR-19-1745
222. Malumbres M. Cyclin-dependent kinases. *Genome Biol.* 2014;15(6):122. doi:10.1186/gb4184
223. Malumbres M, Barbacid M. Cell cycle, CDKs and cancer: a changing paradigm. *Nat Rev Cancer.* 2009;9(3):153–166. doi:10.1038/nrc2602
224. Carey LA, Perou CM, Livasy CA, et al. Race, breast cancer subtypes, and survival in the Carolina Breast Cancer Study. *JAMA.* 2006;295(21):2492–2502. doi:10.1001/jama.295.21.2492
225. Criscitiello C, Azim Jr HA, Schouten PC, Linn SC, Sotiriou C. Understanding the biology of triple-negative breast cancer. *Ann Oncol.* 2012;23(Suppl 6):vi13–8. doi:10.1093/annonc/mds188
226. Pegram MD, Lipton A, Hayes DF, et al. Phase II study of receptor-enhanced chemosensitivity using recombinant humanized anti-p185HER2/neu monoclonal antibody plus cisplatin in patients with HER2/neu-overexpressing metastatic breast cancer refractory to chemotherapy treatment. *J Clin Oncol.* 1998;16(8):2659–2671. doi:10.1200/JCO.1998.16.8.2659
227. Wiggins RG, Woolley PV, Smythe T, et al. Phase-II trial of tamoxifen in advanced breast cancer. *Cancer Chemother Pharmacol.* 1979;3(1):45–48. doi:10.1007/BF00254419

Breast Cancer: Targets and Therapy

Publish your work in this journal

Breast Cancer - Targets and Therapy is an international, peer-reviewed open access journal focusing on breast cancer research, identification of therapeutic targets and the optimal use of preventative and integrated treatment interventions to achieve improved outcomes, enhanced survival and quality of life for the cancer patient. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/breast-cancer—targets-and-therapy-journal>

Dovepress
Taylor & Francis Group