

Pathological Mechanisms and Molecular Imaging Advances in Alzheimer's Disease

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Abstract: Alzheimer's disease (AD) is one of the most prevalent neurodegenerative disorders globally, where early diagnosis plays a pivotal role in delaying disease progression and improving patient outcomes. In recent years, the rapid, multidisciplinary advances in molecular imaging and emerging technologies have significantly advanced our understanding of AD pathogenesis, early diagnosis, and intervention strategies. Imaging tools such as positron emission tomography (PET) and magnetic resonance imaging (MRI), alongside emerging technologies like retinal imaging, nanosensors, and quantum dots (QDs), are continuously enhancing AD diagnostic pathways. Studies on the gut microbiome and extracellular vesicles (EVs) offer novel insights into AD pathogenesis. Furthermore, AI-driven multimodal data fusion techniques hold great promise for improving diagnostic accuracy. Future research will increasingly focus on multi-target synergistic intervention strategies, standardization of multimodal imaging, and the integration of AI with molecular diagnostics and treatment to enable early detection and personalized precision therapy for AD.

Keywords: Alzheimer's disease, molecular imaging, gut microbiome, magnetic resonance imaging, positron emission tomography

Introduction

According to WHO statistics, life expectancy is increasing globally, with most individuals now expected to live beyond 60 years. The number and percentage of elderly individuals are increasing in all countries, shifting the demographic center of gravity toward older populations. The aging population also brings about a rapid increase in aging-related diseases, bringing a heavy burden to society and families. And one of the more common ones is dementia. Alzheimer's disease (AD) is the most common cause of dementia, accounting for 50%-75% of cases. The prevalence of AD increased with age and increased significantly after age 80.¹ AD is a neurodegenerative disorder characterized by progressive cognitive decline. In 2018, AD International estimated that approximately 50 million people worldwide were affected by dementia, highlighting the growing importance of AD research.

Clinically, AD is categorized into three stages based on cognitive impairment: the asymptomatic stage with biomarkers (preclinical AD), the prodromal stage with mild cognitive impairment (MCI) and/or mild behavioral changes, and the dementia stage characterized by dysfunction. This progression is continuous and progressive. Currently, there is no clinical method to reverse AD fully. Moreover, patients typically do not exhibit clinical symptoms in the early stages. Once symptoms appear, it indicates the disease has reached a stage difficult to reverse.²

Given the progressive and irreversible nature of AD, early diagnosis plays a vital role in optimizing patient outcomes. For patients, timely diagnosis increases the likelihood of delaying disease progression through pharmacological intervention. It also enables physicians to make comprehensive assessments and formulate individualized treatment plans. For family members, early intervention can reduce long-term caregiving burdens and medical expenses. By 2050, 71% of

people with dementia are expected to be living in low- and middle-income countries,³ so early diagnosis will reduce the financial burden.

The diagnosis of AD involves multiple branches of medicine, and the mainstream direction is to detect and diagnose biomarkers, including imaging biomarkers and fluid biomarkers. This review begins with the pathophysiological mechanisms of AD and proceeds to discuss diagnostic techniques that are grounded in the disease's pathology and physiological changes. Molecular imaging techniques, such as positron emission tomography (PET), can detect neurodegeneration through characteristic changes in biomarkers like β -amyloid ($A\beta$), Tau proteins, and energy metabolism. Additionally, magnetic resonance imaging (MRI) is used to assess structural and functional brain changes in AD. Genetic testing related to the apolipoprotein E (*APOE*) locus, retinal imaging, and electroencephalography are increasingly applied in clinical practice. The integration of these advanced technologies is expected to enable earlier and more accurate identification and intervention of AD.

Pathophysiology of AD

The pathogenesis of AD is not a linear sequence of events but rather a complex interplay of interdependent mechanisms. Multiple interrelated mechanisms contribute to a cycle that accelerates disease progression (Figure 1). Microscopic examination reveals that the hallmark pathological features of AD include neuritic plaques composed of $A\beta_{42}$ and neurofibrillary tangles (NFTs) consisting of highly phosphorylated Tau. These proteins play a critical role in disrupting neural connectivity, ultimately leading to neuronal death and brain tissue damage. Additionally, neurodegeneration, along with synaptic and neuronal loss, is observed.

Amyloid Accumulation

The amyloid hypothesis is the most widely accepted theory of AD pathogenesis, proposing that $A\beta$ deposition is the initiating event in the disease cascade. $A\beta$ is produced from amyloid precursor protein (APP) through sequential cleavage by β and γ secretases.⁴ $A\beta$ plaques accumulate in various brain regions and are primarily composed of $A\beta_{40}$ and $A\beta_{42}$, two metabolic by-products of APP.⁵ These plaques are recognized by the brain as foreign substances. They activate microglia and induce cytokine release, initiating inflammatory and immune responses that ultimately lead to neuronal death and neurodegeneration.⁶ $A\beta$ aggregates are assembled from $A\beta$ monomers into a variety of unstable oligomeric species, collectively termed (o $A\beta$). This amyloid plaque is not only neurotoxic in its own right but also alters kinase/phosphatase activity. In the early stages of disease progression, $A\beta$ enhances the activity of several kinases, including glycogen synthase kinase-3 β (GSK-3 β) and cyclin-dependent kinase 5 (CDK-5).^{7,8} GSK-3 β plays a central role in Tau phosphorylation by targeting multiple serine/threonine residues. This exacerbates Tau-induced neurotoxicity.⁷ Hyperphosphorylated Tau, dissociated from microtubules, has a higher propensity to form NFTs.⁹ Additionally, N-methyl-D-aspartate receptor (NMDAR) subunits which are critical regulators of synaptic function in AD, can co-immunoprecipitate with o $A\beta$. $A\beta$ evokes immediate cellular Ca^{2+} influx through the activation of NMDARs in primary neurons.¹⁰

$A\beta$ promotes Tau pathology and, in conjunction with Tau, accelerates the progression of AD.¹¹ Tau is considered a key factor triggering downstream effects in the pathogenesis of AD.¹² Tau and $A\beta$ disrupt mitochondrial calcium homeostasis, leading to mitochondrial dysfunction and reduced neuronal viability.¹³ In turn, mitochondrial dysfunction promotes $A\beta$ accumulation, NFT formation, and neurodegeneration.¹⁴ The synergistic effects of $A\beta$ and Tau also affect microglia and astrocytes.¹⁵ Activated microglia and reactive astrocytes initiate neuroinflammatory cascades, thereby accelerating neurodegeneration. Ultimately, this leads to blood-brain barrier (BBB) disruption and cognitive decline.¹⁶

Neurofibrillary Degeneration

In 1988, scientists isolated Tau from plaques in the brains of AD patients, suggesting for the first time that Tau proteins may be responsible for AD.¹⁷ Tau is a soluble microtubule-associated protein predominantly expressed in neurons. Under normal physiological conditions, Tau localizes to axons and binds microtubules to maintain cytoskeletal stability. Under pathological conditions, Tau becomes hyperphosphorylated due to aberrant kinase activity, resulting in its detachment from microtubules and the formation of insoluble aggregates. These aggregates further form NFTs. Both 3R and 4R Tau

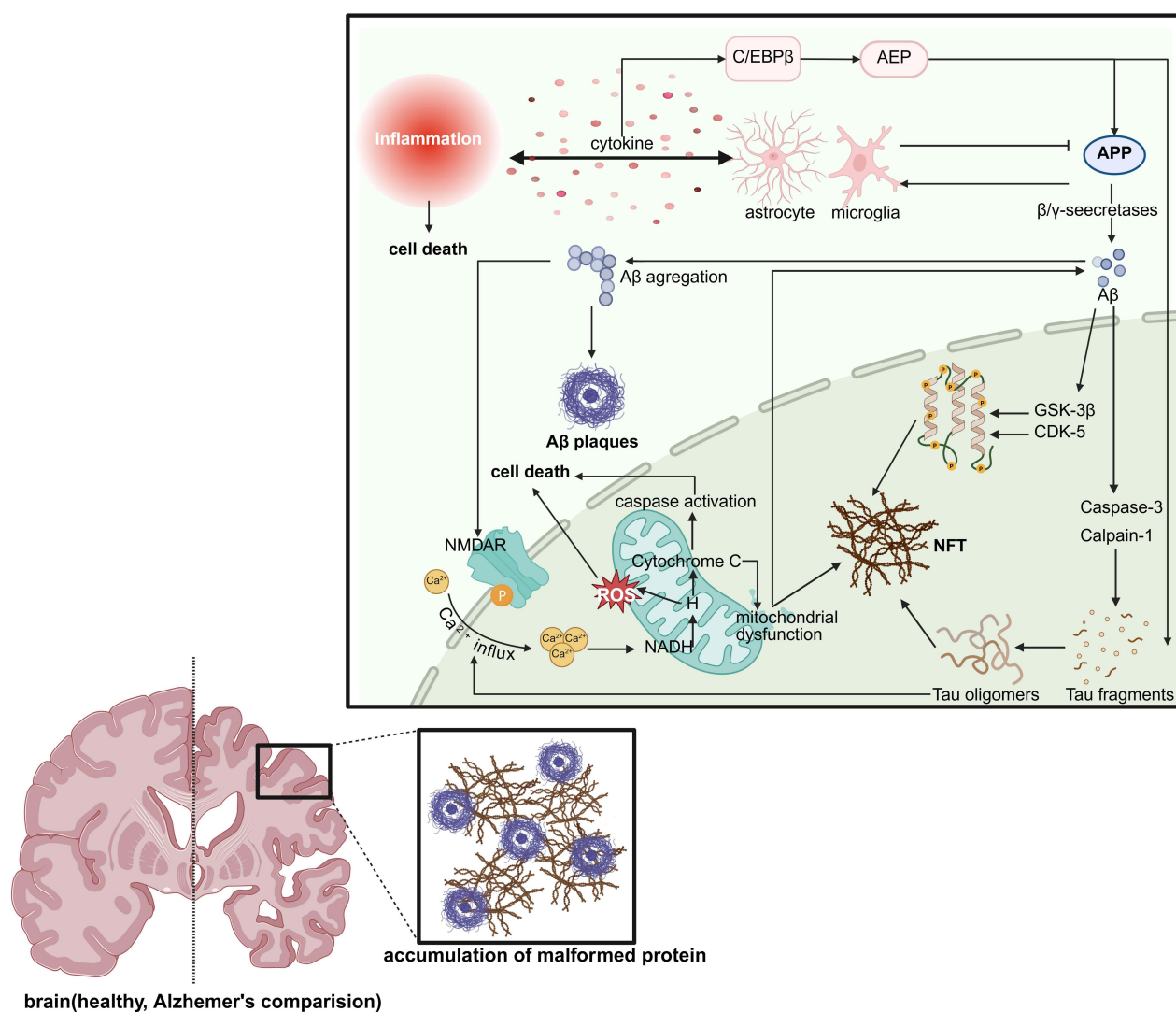


Figure 1 Pathophysiology of Alzheimer's disease.

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isoforms are present in AD, and their dysregulated expression is closely linked to disease progression.¹⁸ Moreover, pathological Tau can be internalized by healthy neurons through mechanisms such as endocytosis and receptor-mediated uptake, thereby perpetuating a vicious cycle of AD progression.¹⁹ Additionally, hyperphosphorylated Tau is intrinsically neurotoxic and mediates Aβ-induced toxicity, with Aβ neurotoxicity being largely dependent on the presence of Tau.²⁰ Tau protein is currently a key biomarker and therapeutic target in AD diagnosis and treatment. Total tau (t-Tau) and phosphorylated tau (p-Tau) levels in colony-stimulating factor (CSF) are accurate biomarkers for tracking progression from MCI to advanced dementia.²¹ Before the appearance of pathological Aβ plaques, plasma p-Tau231 and p-Tau217 showed significant differences in capturing brain Aβ better altered.²²

Synaptic Dysfunction and Neurotransmitter Imbalance

The cholinergic hypothesis posits that cholinergic nerve inactivation contributes to cognitive decline in AD patients. The cholinergic system plays a fundamental role in regulating memory and attention. These cognitive functions rely on intact synaptic structures and functions. Damage to basal forebrain cholinergic neurons that innervate the cortex can result in attention deficits, contributing to the characteristic decline in learning and memory observed in AD patients.²³

Significantly reduced levels of acetylcholine have been found in the cortical cerebrospinal fluid of AD patients,²⁴ and loss of synapses associated with cognition can be found in postmortem brain tissue of patients.²⁵ In the basal nucleus of Meynert, nicotinic acetylcholine receptors density is significantly reduced,²⁶ accompanied by a marked depletion of choline acetyltransferase (ChAT). The reduction in ChAT has been reported to correlate with the severity of dementia.²⁷ These findings indicate that cholinergic inactivation is a key contributor to memory deficits in AD.

AD is also increasingly recognized as a synaptic disorder.²⁸ Experimental data indicate that synapses in AD patients exhibit a reduced number, altered shapes, and changes in the expression of proteins.^{29,30} And varying degrees of loss and alteration of markers on the synaptic surface are observed.³¹ Disruption of synapses leads to disruption of neuronal network activity. This disruption affects both excitatory and inhibitory synapses.³² Disruption of the excitation-inhibition (E/I) balance may induce seizures with epileptiform symptoms, which aligns with the significantly higher incidence of epilepsy in AD patients compared to the general population.^{33,34}

Neuroinflammation

Neuroinflammation is also recognized to be one of the key components in the pathogenesis of AD, and it has been proved that neuroinflammation can be an indicator of early detection of AD.³⁵ Activated microglia, cytokines, and pathological astrocytes have been observed in the brains of AD patients.^{36,37}

Microglia are brain-resident phagocytes that contribute to central nervous system (CNS) development and play essential roles in neuronal regulation and connectivity. Due to the presence of the BBB, microglia are the main cells in the brain that accomplish peripheral immune activity. In response to harmful stimuli, microglia mediate an acute immune response. For example, if A β protein is deposited in the brain, microglia can reduce the accumulation of A β protein by increasing clearance or phagocytosis. However, activated microglia exist in both anti-inflammatory and pro-inflammatory states. Therefore, if the adverse stimulation exists for a long time and cannot be solved, the immune function of microglia will lead to increased damage. Chronic microglial activation dysregulates neuroinflammatory responses, leading to neuronal atrophy, synaptic loss, A β production, and neurotoxic effects.^{38,39} Several immune cytokines, including tumor necrosis factor- α (TNF- α), interleukins (IL), and type I interferon, are closely linked to various AD processes.^{40,41} Interferon-induced transmembrane protein 3 (IFITM3) has been shown to enhance γ -secretase activity, increasing A β deposition, which leads to further A β secretion in response to chronic inflammation.⁴² At the same time, the accumulated A β will further activate microglia,⁴³ stimulate the release of pro-inflammatory factors, and interfere with the synthesis of anti-inflammatory factors to induce neuroinflammation and neurodegeneration.^{44,45} Researchers also suggest that AD is primarily caused by reduced clearance of A β , rather than by its overproduction.⁴⁶

Gut Microbiome Disruption

The gut microbiome refers to the microbial community residing in the human gut, comprising bacteria, fungi, and phages. It is now hypothesized that dysbiosis of the gut microbiome may influence pathophysiologic changes in AD. This effect is not unidirectional but bidirectional. The bidirectional “microbiota-gut-brain” axis is widely recognized. The CNS connects to the gut via sympathetic and parasympathetic nerves. The gut can be connected to the brain through enteroendocrine, short-chain fatty acids, and neurotransmitters. First, dysregulation of the gut microbiome can lead to the development of an inflammatory response. These pro-inflammatory substances not only increase intestinal epithelial permeability, facilitating their translocation into the circulatory system, but also cross the BBB to activate microglia and astrocytes in the brain.⁴⁷ Among the inflammatory cytokines, C/EBP β acts as a transcription factor for A β activation. It promotes the expression of asparagine endopeptidase (AEP), which cleaves APP and Tau, facilitating the formation of A β and NFTs.⁴⁸ Bacterial DNA, particularly from *Escherichia coli* and *Porphyromonas gingivalis*, which are significantly correlated with AD, also promoted Tau aggregation to some extent.⁴⁹ Notably, recent studies have shown that transplanting fecal microbiota from healthy wild-type mice into transgenic AD mice leads to significant improvements in A β plaque deposition and NFT formation.⁵⁰ Collectively, these findings suggest that disruption of the gut microbiome may contribute to the pathogenesis of AD.

Oxidative Stress

Mitochondrial dysfunction and oxidative stress (OS) have long been implicated in the early pathogenesis of AD. A key contributor is the reduction in cytochrome c oxidase levels, which impairs mitochondrial function. This mitochondrial dysfunction is exacerbated by OS-induced hyperactivation of GSK-3, leading to disruption of mitochondrial membrane permeability and excessive production of reactive oxygen species (ROS). Metal ions, especially copper, may bind to A β plaque and produce ROS. The resulting ROS not only oxidize modified A β peptides and hinder their clearance, but also damage lipids and proteins in neuronal membranes, thereby increasing membrane permeability and vulnerability.⁵¹ In parallel, A β plaques impair calcium ion storage in the endoplasmic reticulum, resulting in elevated cytosolic calcium levels. This rise in calcium depletes cellular glutathione stores and further accelerates intracellular ROS accumulation.⁵² Additionally, overactivation of NMDARs enhances calcium influx, promoting the formation of both ROS and reactive nitrogen species (RNS).⁵³ A β peptides can directly activate nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, triggering the synthesis of free radicals and intensifying oxidative damage.⁵⁴

Molecular Imaging Techniques for AD Detection

Molecular imaging plays a key role in the early diagnosis and monitoring of AD by visualizing pathological biomarkers (Figure 2).

Application of PET Imaging in AD Detection

PET imaging belongs to the category of molecular imaging and functional imaging. It is founded on advancements in basic research and the development of specific imaging agents. It has been used to study various aspects such as neuronal synaptic function, A β deposition, Tau proteins, as well as various neurotransmitter and receptor changes in a visual imaging manner. And patients with AD exhibit pathological changes 2–15 years before the onset of overt clinical symptoms.⁵⁵ Therefore, PET imaging is currently employed for the early clinical diagnosis of AD. PET imaging is very helpful in atypical/unspecified cases (81.1% of the cases) and in patients with MCI (88.2% of the cases).⁵⁶ Additionally, PET can assist in excluding other disease types. Studies indicate that FDG-PET demonstrates high sensitivity (96.7%) at a Clinical Dementia Rating (CDR) of 1. However, MRI and CT findings in AD are not specific.⁵⁷ PET imaging can be classified into various types based on the clinical applications of different PET imaging agents.

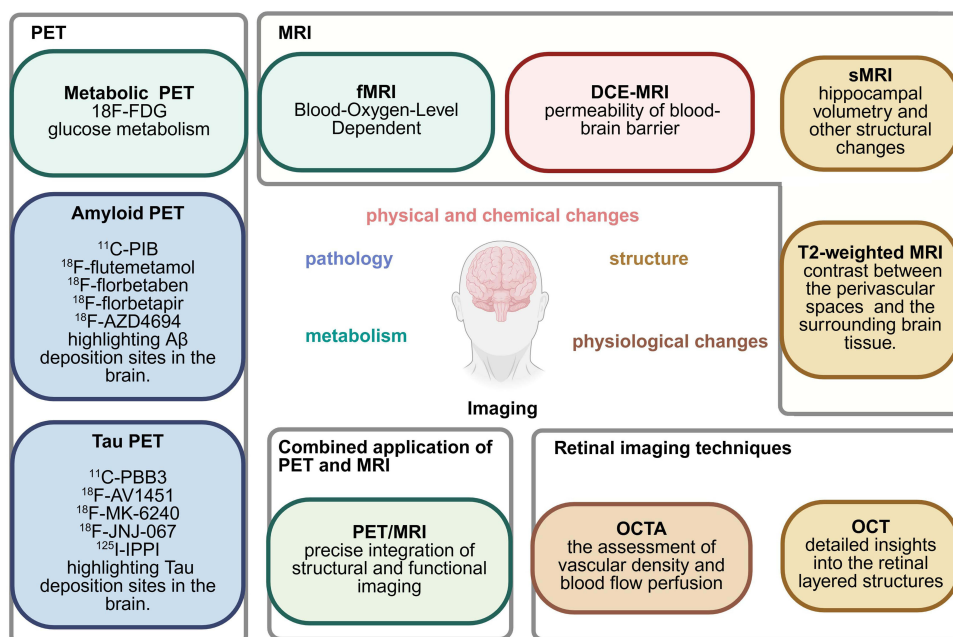


Figure 2 Molecular imaging techniques for AD detection.

Notes: Created in BioRender. Liu, C. (2025) <https://BioRender.com/f41q369>.

A β Class

According to the pathophysiology of AD described above, A β plaques are formed and deposited in different regions of the brain. Among them, A β ₄₂ shows a greater tendency to aggregate and can be detected in the brains of AD patients.⁵⁸ The A β PET imaging agent (amyloid imaging agent) is based on this pathogenesis in AD patients. This technique enables non-invasive in vivo detection of amyloid plaques. It demonstrates high sensitivity (69%–95%) and specificity (83%–89%) in detecting amyloid pathology in patients with neuropathologically confirmed AD.⁵⁹ Amyloid PET can also detect amyloid pathology in clinically atypical variants of AD, such as posterior cortical atrophy, frontal variant AD, or logopenic variant primary progressive aphasia.⁶⁰ Currently, ¹¹C-Pittsburgh Compound B (¹¹C-PiB) and ¹⁸F-flutemetamol are commonly used in clinical diagnosis. In addition, ¹⁸F-florbetaben (NCT02681172), ¹⁸F-florbetapir, and ¹⁸F-AZD4694 are also being actively tested and gradually promoted in the clinics.^{61–64}

¹¹C-PiB is a derivative of the A β stain thioflavin T, exhibiting high affinity and specificity for A β .⁶⁵ ¹¹C-PiB PET can diagnose early amyloid deposition and early AD patients earlier and more specifically.⁶⁶ However, the short half-life of ¹¹C, approximately 20 minutes. This leads to a serious obstacle to the widespread dissemination of ¹¹C-PiB PET in routine clinics.

To overcome the limitations of short-lived radionuclides, ¹⁸F isotope was developed and subsequently approved by the US Food and Drug Administration (FDA). With a half-life of approximately 110 minutes, ¹⁸F offers a practical alternative to ¹¹C, extending the imaging window and improving clinical usability. ¹⁸F-flutemetamol, a fluorinated analog of thioflavin T and a derivative of the widely studied ¹¹C-PiB, demonstrates high specificity and sensitivity in detecting A β plaque distribution in AD patients. These findings have been confirmed through postmortem pathological verification.⁶⁷ However, both ¹¹C-PiB and ¹⁸F-flutemetamol exhibit off-target signals in white matter during PET imaging. Notably, the white matter uptake of both tracers increases with age, but ¹⁸F-flutemetamol shows a higher degree of white matter retention compared to ¹¹C-PiB.⁶⁸ Recent studies suggest that in younger individuals without cognitive impairment, ¹⁸F-flutemetamol may provide better imaging performance due to its stronger contrast between gray and white matter, thereby reducing the likelihood of false positives. Conversely, in cognitively unimpaired older adults, ¹¹C-PiB may outperform ¹⁸F-flutemetamol because of its superior sensitivity in detecting subtle or near-threshold cortical A β deposits, particularly in higher-order cortical regions.⁶⁹

Amyloid imaging agents share a common limitation: different amyloid-positive diseases may exhibit similar deposition patterns, making it difficult for PET imaging to differentiate among disease types.⁷⁰ Additionally, A β ₄₀ is primarily confined to neuritic plaques.⁷¹ But amyloid PET has weak binding affinity for A β ₄₀. And it cannot quantify the reduction in A β levels over time, further diminishing its diagnostic value.⁷² Notably, A β burden does not clearly correlate with the clinical manifestations of AD.⁷³ In summary, these limitations reduce the utility of amyloid imaging for monitoring disease progression.

Tau Protein Class

In addition to A β , NFTs are a pathological hallmark that exists in the brains of AD patients. Tau imaging can distinguish between early and late NFT pathology by assessing the varying rates of Tau deposition at different stages. Older adults without AD may sometimes yield positive results on Amyloid PET scans, potentially leading to false-positive interpretations. The use of Tau imaging—either independently or in conjunction with amyloid imaging—can significantly improve diagnostic precision by confirming the presence of neurodegenerative lesions, thereby reducing false-positive rates.^{74,75} In contrast, Tau PET imaging has demonstrated slightly higher accuracy than amyloid PET, both in detecting disease presence and in predicting the progression of AD.^{76,77} In addition, data suggest that Tau-associated radioligand is specific for PHF-Tau deposition.⁷⁸ Therefore, it can be utilized to differentiate AD from other Tauopathies.⁷⁴ Owing to its diagnostic value, Tau PET imaging has become a focal point of AD research in recent years and has made significant advancements.

First-generation radioactive ligands, such as ¹¹C-PBB3 and ¹⁸F-AV1451, have been developed, with ¹⁸F-AV1451 being the most widely used. Studies have demonstrated that ¹⁸F-AV1451 exhibits specific binding in AD-related brain regions (eg, amygdala, entorhinal cortex, parahippocampus, fusiform, etc). However, its early diagnostic capability is limited, as ¹⁸F-AV1451 does not effectively distinguish MCI from cognitively normal individuals.⁷⁹ To enhance the

selectivity for various Tau protofibril subtypes and reduce off-target binding, second-generation PET radioligands have been developed. Among the most advanced are ^{18}F -MK-6240.^{80,81} In a cross-sectional study of ^{18}F -MK-6240, both ^{18}F -MK-6240 and ^{18}F -AV1451 showed statistically significant differences when comparing cognitively unimpaired amyloid negative ($\text{A}\beta^-$ CU) and $\text{A}\beta^+$ CU. The group separation was more pronounced with ^{18}F -MK-6240. This suggests that ^{18}F -MK-6240 may be more effective in detecting early Tau accumulation.⁸² In addition to fluorine-based ligands, radioactive iodine derivatives such as ^{125}I are increasingly being utilized in in vitro studies of NFTs. Notably, the novel radioligand ^{125}I -IPPI has demonstrated high binding affinity to Tau, with no reported obvious off-target binding.^{83,84}

In summary, Tau PET shows strong potential for diagnosing, staging, and predicting AD. However, its broader clinical application is limited by off-target binding and low sensitivity. Ongoing efforts aim to improve its clinical utility by developing second-generation Tau imaging agents. Table 1 presents PET imaging agents used for detecting AD, including those targeting $\text{A}\beta$ and Tau proteins.

Energy Metabolism Class (^{18}F -FDG)

Cerebral glucose metabolism is a key indicator of synaptic activity and neuronal density. One hallmark of AD is a reduction in glucose metabolism within the brain, which reflects the functional loss of neurons.⁹⁴ To evaluate the cerebral metabolic rate of glucose (CMRgl), the radiotracer ^{18}F -fluorodeoxyglucose (^{18}F -FDG) is widely used in PET imaging. ^{18}F -FDG is a radiolabeled glucose analog that substitutes for glucose in metabolic pathways, allowing clinicians to visualize and quantify regional brain metabolism. As such, ^{18}F -FDG PET provides a reliable biomarker for assessing intracellular glucose utilization. The prevailing hypothesis suggests that metabolic disturbances in the AD brain occur prior to the onset of clinical symptoms and overt pathological changes. Therefore, ^{18}F -FDG PET is particularly valuable for early diagnosis. Compared to structural imaging methods like MRI, ^{18}F -FDG PET offers superior sensitivity for detecting early functional changes. PET imaging features in AD patients include hypometabolism in the frontal cortex, posterior cingulate cortex, parietal cortex, and temporal cortex regions.⁹⁵ Notably, patterns of hypometabolism vary among different AD subtypes.⁹⁶ Therefore, ^{18}F -FDG PET can be used for the identification of AD subtypes. In distinguishing AD from non-AD dementias, ^{18}F -FDG PET has demonstrated a sensitivity of 0.86 and a specificity of 0.88, also highlighting its diagnostic reliability.⁹⁷ ^{18}F -FDG PET remains the primary modality for functional brain imaging in the detection and diagnosis of AD.

^{18}F -FDG PET also has shortcomings. It has high sensitivity, but its specificity is significantly lower.⁹⁸ In addition, there remains controversy over whether elevated blood glucose levels alter the distribution of ^{18}F -FDG in the brains of cognitively normal individuals, potentially influencing the diagnostic interpretation of AD.^{99,100} Currently, research is underway on emerging imaging agents for mitochondrial dysfunction in the parahippocampus in early-stage AD.¹⁰¹

Application of MRI in AD Detection

MRI, characterized by its high spatial resolution, non-invasiveness, and lack of radiation exposure, can detect patterns of brain damage and abnormalities.¹⁰² It plays a crucial role in elucidating the neuropathological mechanisms of AD and MCI, aiding in the differentiation of AD from other brain disorders, and predicting the progression from MCI to AD.¹⁰³ Currently, MRI encompasses various modalities; this paper focuses on structural MRI (sMRI), functional MRI (fMRI), vascular imaging using MRI, and the combined application of PET/MRI.

Structural MRI (sMRI)

sMRI provides detailed images of brain anatomy, allowing precise observation of morphological and volumetric changes across various regions. This capability is crucial for identifying structural alterations associated with AD.

In AD, significant structural changes occur, notably hippocampal atrophy, which manifests in the disease's early stages. Reduction in hippocampal volume correlates closely with cognitive decline, with atrophy rates differing between AD patients and healthy individuals. Neocortical atrophy in AD follows a specific pattern, spreading from the medial temporal lobe to other regions, distinct from atrophy patterns observed in normal aging. Atrophy of the entorhinal cortex is evident in both AD and MCI patients and is closely linked to cognitive decline. Subcortical structures

Table I Examples of PET Imaging Agents

Tracer	Study Population	Main Findings	Researcher
¹¹ C-PiB (A β)	40 probable AD dementia (ProAD) patients 20 possible AD dementia (PosAD) patients 25 MCI patients	¹¹ C-PiB PET improved diagnostic accuracy in cases with high diagnostic uncertainty after MRI and SPECT.	Omachi Y (2015) ⁸⁵
¹⁸ F-florbetapir (A β)	10 AD patients 8 FTD patients 10 healthy controls	¹⁸ F-florbetapir effectively differentiates AD patients from frontotemporal dementia (FTD) patients and healthy controls.	Kobylecki C (2015) ⁸⁶
¹⁸ F-florbetaben (A β)	45 MCI patients	¹⁸ F-florbetaben-positive individuals demonstrate a higher progression rate to AD over 2- and 4-year follow-ups, supporting the identification of prodromal AD in MCI patients and enabling timely intervention.	Ong KT (2015) ⁸⁷
¹⁸ F-flutemetamol (A β)	131 MCI patients 41 AD patients 20 dementia NOS patients 5 SCD patients 10 non-AD patients	¹⁸ F-flutemetamol PET positivity rates varied among clinical groups, facilitating the differentiation of cognitive impairment subtypes and informing diagnostic decisions in patients with MCI.	Leuzy A (2019) ⁸⁸
¹¹ C-PBB3 (Tau)	7 AD patients 7 healthy controls	Significant differences were detected between AD patients and healthy controls in the cerebral cortex (including the hippocampal formation).	Kimura Y (2015) ⁸⁹
AV-1451 (Tau)	13 AD patients 84 healthy controls	AV-1451 PET imaging demonstrated significant elevations in Tau protein in the entorhinal cortex, lateral occipital cortex, inferior temporal cortex, and amygdala in preclinical AD, highlighting the importance of these regions in early AD diagnosis.	Mishra S (2017) ⁹⁰
AV-1451 (Tau)	364 participants 203 AD patients 161 MCI patients	An advanced AD flortaucipir pattern was associated with increased risk of clinical progression over 18 months. This indicates the value of PET in managing and assessing clinical progression in patients with cognitive impairment.	Lu M (2021) ⁹¹
¹⁸ F-MK-6240 (Tau)	6 AD patients 4 healthy controls	All but one AD/MCI participant exhibited increased MK-6240 retention in brain regions associated with AD-related NFT deposition, particularly in the medial temporal lobe. Late-stage AD patients also demonstrated tracer retention in neocortical regions.	Lohith TG (2019) ⁹²
[¹⁸ F]-JNJ-067 (Tau)	17 participants 5 MCI patients 5 AD patients 3 PSP patients 4 healthy controls	Significant [¹⁸ F]-JNJ-067 binding was detected in AD participants. However, the lack of significant differences between amyloid-negative HCs and amyloid-positive MCI participants suggest this tracer has limited sensitivity in early clinical and preclinical cohorts.	Baker SL (2021) ⁹³

—including the amygdala, thalamus, basal ganglia, and basal forebrain—also exhibit varying degrees of atrophy, each correlating with cognitive decline and disease progression.¹⁰⁴

sMRI effectively demonstrates brain atrophy characteristics in AD patients, plays a role in the early diagnosis of AD by detecting atrophy in specific brain regions such as the hippocampus and medial temporal lobe in people with MCI.¹⁰⁵ When combined with appropriate analytical methods and classification algorithms, sMRI can accurately distinguish patients with AD, MCI, and healthy controls. One study demonstrated that sMRI exhibits exceptional performance in differentiating AD from healthy controls and MCI patients, achieving precise classification. In both AD vs healthy control and AD vs MCI models, the area under the curve (AUC) reached 1.00, indicating that the sMRI model can accurately distinguish AD patients from both groups.¹⁰⁶

Since sMRI can track structural brain changes over time, it is valuable for monitoring disease progression and developing individualized disease progression models. Additionally, analyzing sMRI data from AD patients at various stages helps elucidate patterns of brain structural changes during disease progression, providing insights into AD's pathological mechanisms. For instance, integrating deep learning algorithms with sMRI data can uncover neurodegenerative patterns in AD, including spatial and spatiotemporal connectivity. Studies indicate that neurodegenerative brain regions exhibit stable spatial connectivity over longitudinal studies. sMRI serves as a critical imaging tool for investigating these patterns, enhancing our understanding of AD's pathological mechanisms.¹⁰⁷

However, sMRI has notable limitations: its diagnostic accuracy is limited, with low sensitivity and specificity for key regions like the hippocampus and medial temporal lobe.¹⁰⁵ It lacks molecular specificity, unable to directly detect AD's histological hallmarks like A β proteins and NFTs. Additionally, it requires specialized expertise, leading to high measurement variability.¹⁰⁸

In summary, sMRI effectively captures AD-related structural brain changes, aiding early detection, differential diagnosis, progression monitoring, and pathological research. However, its limitations restrict its standalone utility. Still, when combined with other biomarkers or advanced analytical tools, it remains valuable for advancing AD understanding and clinical management.

Functional MRI

Functional MRI (fMRI) indirectly reflects neuronal activity by detecting changes in hemodynamic dynamics when the brain is active, it can non-invasively acquire brain function information of humans in cognitive, behavioral and other states with a certain spatial and temporal resolution across the whole brain,¹⁰⁹ aiding in the identification of functional abnormalities in AD patients.

Cognitive tasks, such as memory encoding, elicit changes in brain activity in AD patients. For example, during the process of encoding new information, hippocampal activity is reduced in AD patients. Conversely, increased activity may be detected in other regions, such as the prefrontal cortex, which is considered an attempted compensatory mechanism employed by other neural networks when the hippocampus is impaired.¹¹⁰

During resting-state fMRI, AD patients show reduced connectivity within the default mode network (DMN), encompassing the posterior cingulate cortex, precuneus, and medial prefrontal cortex. In patients with MCI, such connectivity abnormalities are mainly manifested as reduced functional connectivity between the posterior cingulate cortex and the anterior cingulate cortex, with a lesser degree than in AD patients.¹¹¹ A study leveraged machine learning to capitalize on these resting-state fMRI patterns, training models to distinguish AD, MCI, and other groups. Among these patterns, reduced DMN connectivity in AD patients is a feature that differentiates AD from healthy controls. Machine learning algorithms effectively classified these conditions, highlighting the potential of resting-state fMRI to assist in the differential diagnosis of AD.¹¹²

However, fMRI has several limitations in its application. It does not directly capture neural activity but reflects it indirectly through changes in blood flow, which increases the difficulty of interpreting signals. Its temporal resolution is relatively low, and as a single imaging modality, it struggles to explicitly link specific abnormalities in functional connectivity to a particular disease, similar changes may also occur in other disorders, resulting in a deficiency in establishing disease-specific associations.¹¹³ These factors collectively restrict its application in disease research.

Nonetheless, its ability to detect early functional changes in AD and MCI remains valuable. Combining it with other modalities may further enhance its clinical utility.

Vascular Detection Using MRI

AD is a complex neurodegenerative disorder, with its pathogenesis closely linked to vascular factors. MRI plays a fundamental role in vascular assessment in AD, primarily by detecting perivascular spaces (PVS) and evaluating the integrity of BBB.

MRI Detection of PVS

PVS are fluid-filled spaces surrounding cerebral blood vessels, and their enlargement can be detected on T2-weighted MRI images. Enlarged PVS serves as a marker for cerebral small vessel disease and amyloid pathology. In cerebral amyloid angiopathy (CAA), PVS severity in the centrum semiovale (CSO) exceeds non-CAA cases, with distinct patterns in the basal ganglia observed both in vivo and postmortem.¹¹⁴ Research suggests that an increased PVS burden is linked to a progressive rise in dementia risk. Higher PVS grades in the CSO are associated with an elevated risk of dementia and AD.¹¹⁵ Emerging diffusion techniques like diffusion tensor imaging along the perivascular spaces (DTI-ALPS), enable assessment of fluid movement within PVS, with a lower DTI-ALPS index linked to AD dementia and reduced cognitive function.¹¹⁶ Automated segmentation of PVS yields continuous measures, capturing spatial distribution and shape complexity of PVS, which offers more detailed and objective information beyond the classification provided by visual scoring.¹¹⁷ These MRI-based methods for detecting PVS, from basic observation to advanced techniques, aid in understanding AD-related mechanisms and improve detection.

MRI Detection of the BBB

BBB dysfunction is a key pathological feature of AD, which occurs in the early stage of the disease, even before the accumulation of pathological proteins, and precedes neurodegeneration. Therefore, monitoring BBB abnormalities provides an important basis for the early detection of AD.¹¹⁸ A recent study has utilized the non-invasive diffusion-prepared pseudo-continuous arterial spin labeling (DP-pCASL) MRI technique to measure the water exchange rate (Kw) across the BBB. This technique, which differentiates magnetically tagged water signals from different compartments, has shown potential in sensitively detecting BBB functionality. Results revealed decreased Kw in multiple brain regions across the AD continuum, suggest that MRI-detected Kw alterations could serve as an indicator for tracking BBB dysfunction in AD, contributing to early identification and disease monitoring.¹¹⁹

Combined Application of PET and MRI

PET/MRI is an emerging imaging technology that combines the anatomical and quantitative advantages of MRI with the physiological information obtained from PET.¹²⁰ This hybrid approach reduces radiation exposure and enables one-stop examinations, streamlining clinical workflows. In AD research, PET/MRI captures subtle regional brain alterations, enhances early detection (including preclinical and MCI stages), and improves differential diagnosis by quantifying hippocampal/thalamic metabolic activity and predicting MCI-to-AD progression.¹²¹ Despite its advantages, PET/MRI has limitations, including insufficient accuracy in early attenuation correction, image artifacts, inadequate sensitivity to prodromal diseases, and low equipment popularity. Looking ahead, deep learning will facilitate correction and low-dose imaging, wider anti-amyloid therapies will expand its primary care use, it is expected to become the standard imaging technique for neurodegenerative diseases by 2030.¹²²

Application of Retinal Imaging Technology in AD Detection

The retina and brain originate from the same tissue during embryonic development. As a component of the CNS, the retina exhibits structural and functional similarities to the brain, encompassing neurons, glial cells, and blood barriers. This similarity provides an anatomical and physiological basis for the association between the two, allowing the retina to serve as a window that contributes to a better understanding of processes such as healthy aging and neurodegeneration in

the CNS.¹²³ Therefore, retinal imaging has great potential as a non-invasive, convenient, and relatively low-cost test for early diagnosis and disease monitoring of AD.

Pathological Features of AD in the Retina

AD manifests in the retina through structural, functional, and vascular alterations. Structurally, retinal nerve fiber layer (RNFL) thinning is prominent in advanced AD, particularly in the superior and inferior quadrants, likely reflecting the vulnerability of extramacular large ganglion cell axons.¹²⁴ MCI patients also exhibit RNFL and ganglion cell-inner plexiform layer (GC-IPL) thinning, with macular parameters showing stronger correlations with cognitive decline than peripapillary metrics.^{125,126} Functionally, AD patients display visual deficits (reduced contrast sensitivity, impaired motion perception) and circadian rhythm disturbances linked to melanopsin-containing retinal ganglion cell (mRGC) loss, alongside abnormal pupillary light responses.¹²⁷ Vascularly, AD-associated retinal changes include venular narrowing, increased tortuosity, reduced fractal dimensions, and choroidal thinning, indicative of microvascular dysfunction.^{128,129}

Common Retinal Imaging Techniques

The most widely used retinal imaging techniques today include optical coherence tomography (OCT) and optical coherence tomography angiography (OCTA).

OCT is a high-resolution imaging technique that employs low-coherence light interference to generate cross-sectional retinal scans. It offers detailed insights into the retinal layered structures, including the RNFL, ganglion cell layer (GCL), inner plexiform layer, inner nuclear layer, outer plexiform layer, outer nuclear layer, and retinal pigment epithelium. In AD research, OCT is primarily employed to assess retinal thinning, particularly reductions in RNFL and GCL thickness, which may be linked to neuronal damage and neurodegeneration in AD patients.¹³⁰

OCTA, an extension of OCT technology, is a vascular imaging modality that enables three-dimensional visualization of the retinal and choroidal microvasculature. It facilitates the assessment of vascular density and blood flow perfusion. OCTA enables the detection of retinal vascular abnormalities, including reduced vascular density and morphological alterations.¹³¹ OCTA quantifies radial peripapillary capillary network perfusion by evaluating parameters such as capillary perfusion density (CPD) and capillary flow index (CFI), providing novel insights for AD research. Studies indicate that CPD and CFI measurements obtained via OCTA in the peripapillary region of AD patients demonstrate high repeatability, supporting the reliability of OCTA in assessing peripapillary vascular perfusion.¹³²

OCT and OCTA provide valuable structural and vascular insights in AD, revealing retinal thinning and microvascular changes. Their non-invasive nature and repeatable measurements support their potential as biomarkers, though further validation is needed.

Advantages and Limitations of Retinal Imaging in AD Diagnosis

Retinal imaging provides multiple benefits for AD diagnosis. It is non-invasive, as techniques like OCT, OCTA, and fundus photography impose a minimal physical burden on patients. These examinations are rapid, relatively simple to perform, and well-tolerated, making retinal imaging suitable for large-scale population screening and regular monitoring of high-risk AD groups. This facilitates the early detection of individuals with AD or those at risk. Additionally, retinal imaging holds the potential for early detection. Structural and functional retinal changes may precede the onset of clinical AD symptoms,¹³³ enhancing the potential of retinal imaging techniques for early detection. OCT can detect retinal thinning in patients with early-stage memory impairment, reflecting neurodegenerative changes in the AD brain and aiding early diagnosis.¹³⁴ However, diagnostic accuracy is confounded by age-related retinal thinning, genetic variability, and comorbidities (such as glaucoma, diabetes), which obscure AD-specific interpretations.^{135,136} Technical heterogeneity across OCT/OCTA devices further limits cross-study comparability.¹²⁹ Moreover, retinal degeneration is not exclusive to AD but also occurs in other neurodegenerative and aging-related conditions, including age-related macular degeneration, diabetes, hypertension, and glaucoma. This overlap in clinical features reduces the diagnostic specificity of retinal imaging techniques for AD.^{137,138} While retinal imaging offers a promising non-invasive method for AD detection, its diagnostic accuracy is constrained by overlapping pathologies, technical variability, and other

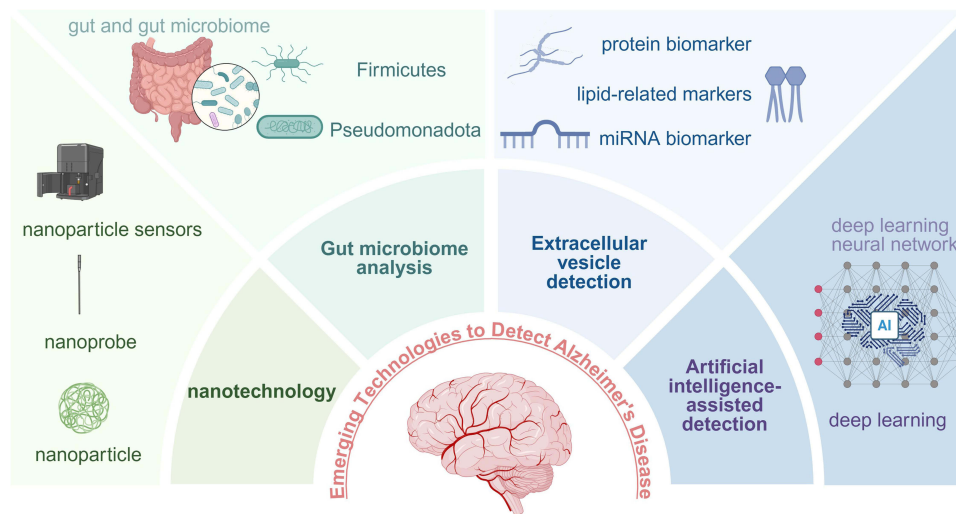


Figure 3 Emerging technologies to detect Alzheimer's disease.

Notes: Created in BioRender. Liu, C. (2025) <https://BioRender.com/q14r466>.

factors. Further optimization is required to improve its clinical utility. While retinal imaging offers a promising non-invasive method for AD detection, its diagnostic accuracy is constrained by overlapping pathologies, technical variability, and other factors. Further optimization is required to improve its clinical utility.

Emerging Detection Technologies

Advancements in biomarker-based diagnostic tools are revolutionizing the detection of AD. Emerging technologies prioritize early detection, enhanced accuracy, and greater accessibility (Figure 3).

Nanotechnology

Traditional AD detection methods primarily utilize PET and MRI to detect A β deposition, pathological Tau proteins, and neurodegeneration. However, these conventional approaches have limitations. For instance, PET imaging is associated with high costs, the need for IV access for radiotracer administration, radiation exposure, and limited accessibility.¹³⁹ MRI have limited sensitivity to the earliest stages of AD.¹⁴⁰ In this context, nanotechnology has emerged as a promising approach for AD detection, offering potential solutions to overcome the limitations of existing methods and enhance early diagnosis.

Nanoparticle Sensors

Nanoparticle-based sensors utilize unique electrical and optical properties for biomarker detection. Electrochemical sensors employ nanomaterials to improve electron transfer between biomolecules and electrodes, enhancing sensitivity. For example, nanomaterials such as gold nanoparticles (AuNPs) and graphene oxide, with high surface area-to-volume ratio and excellent electrical, thermal, and catalytic properties, significantly amplify signals, provide a larger surface area for bioreceptor immobilization, accelerate electron transfer, thereby enhancing detection sensitivity to enable accurate detection of low-abundance exosomal miRNAs and support early diagnosis of neurodegenerative diseases.¹⁴¹ Optical sensors exploit light-biomarker interactions for label-free, real-time detection, and combining them with nanoparticles—their main advantage is the ability to modify optical properties—can enhance sensitivity, specificity, reduce screening time, aiding early AD biomarker detection.¹⁴²

Nanoprobes

Nanoprobes exhibit unique physicochemical properties such as high surface area, tunable surface chemistry, and biocompatibility, enabling highly sensitive detection of AD biomarkers. Furthermore, nanoprobe-based detection methods are non-invasive, reducing patient discomfort and lowering detection costs. Magnetic Nanoprobes play a crucial role

in AD detection. Research indicates that iron can selectively deposit in the core of A β plaques in the form of crystalline magnetic nanoparticles, exhibiting superparamagnetic properties that can be used for AD diagnosis.¹⁴³ For example, curcumin-conjugated superparamagnetic iron oxide nanoparticles coated with polyethylene glycol-poly(lactic acid) can cross BBB to detect A β plaques. Anti-A β functionalized SPIONs can specifically attach to A β plaques, and A β antibody-functionalized magnetic nitrogen-doped graphene exhibits strong selectivity for A β , enabling electrochemical detection.¹⁴⁴

Nanoparticles

QDs have unique properties to overcome conventional dyes and imaging limitations. They enable early AD detection by tracking *in vivo* A β aggregation states, and detect biomarkers like APOE accurately.¹⁴⁵ Moreover, their tunable optics and versatile surface chemistry facilitate precise bioimaging, aiding in identifying pathological hallmarks. However, limitations persist: heavy metal-based QDs raise biocompatibility concerns with proinflammatory effects; BBB penetration remains a critical barrier; and clinical translation is hindered by the need for rigorous trials to validate safety and efficacy in humans.¹⁴⁶ Future efforts need to enhance QDs' biocompatibility, BBB penetration, and clinical translation to unlock their AD diagnostic potential.

AuNPs are extensively employed in biosensing due to their exceptional optical and electronic properties, tunable morphology and size, facile synthesis, high chemical stability, excellent conductivity, catalytic activity, and ease of functionalization.¹⁴⁷ Applications include electrochemical immunosensors for A β 42 detection and plasmonic biosensors for simultaneous detection of multiple AD biomarkers.¹⁴⁸

Gut Microbiome

Based on the pathophysiologic changes in AD patients, it is known that alterations in the gut microbiome may lead to abnormalities in intestinal function or microbial content, which ultimately affects the onset and progression of AD through the “microbiota-gut-brain” axis. Gut microbiome-derived biomarkers have been considered for neurodegenerative diseases, such as multiple sclerosis, with proven differential diagnostic capabilities.¹⁴⁹ Therefore, it is thought that analyzing the alterations in gut microbes in AD patients may assist in determining whether a patient has AD.

Gut Microbiome Analysis

Several studies have suggested that the composition of the gut microbiome can be an indicator of preclinical AD. Ferreira and colleagues analyzed no significant differences in overall Bacteroidota to Firmicutes ratios between healthy and preclinical AD individuals compared to healthy and symptomatic AD individuals.¹⁵⁰ In contrast, Liu et al conducted a study involving 97 participants (33 AD, 32 amnesic mild cognitive impairment [aMCI], and 32 cognitively normal controls), sequencing to characterize the gut bacterial community. They found that microbial diversity was significantly reduced in AD patients, with a notably lower relative abundance of Firmicutes compared to controls. Additionally, Pseudomonadota (formerly known as Proteobacteria) was highly enriched in the AD group relative to healthy controls.¹⁵¹ Similarly, Pan et al found that the relative abundance of Bacteroidota was lower in individuals with MCI compared to healthy controls, while Fusobacteria was significantly more abundant in the MCI group.¹⁵² A related meta-analysis also identified trends in gut microbiome composition among AD patients, showing that the abundance of Bacteroides species varies across regions (higher in the US cohort, lower in the Chinese cohort).¹⁵³ Taken together, these findings indicate that gut microbiome composition is altered in individuals with AD.¹⁵⁴ As such, analyzing the structure of the gut microbiota may aid in the diagnosis or early detection of AD.

Beyond compositional shifts, the secretory products of gut bacteria are also altered in AD. Various metabolites, such as Arachidonic, show a progressive increase as the disease progresses.¹⁵⁵ Similar alterations in microbial metabolites are considered promising biomarker candidates for AD diagnosis.

Shortcomings and Prospects of Detecting Gut Microbiome

Although gut microbiome testing meets the criteria for AD screening, being both sensitive and noninvasive. It also faces several limitations. Analyzing and interpreting the composition of the gut microbiome is time-consuming. And the

microbiome is highly dynamic and influenced by various confounding factors such as health status, lifestyle, and dietary habits, which can affect the reliability of test results. Additionally, there is currently no unified standard for evaluating gut microbiome profiles in relation to AD. Different diagnostic frameworks may yield varying assessments of disease severity based on microbiome composition. Therefore, a standardized research protocol for this screening method is still needed. Research in the field of gut microbiology has primarily focused on bacterial communities, while the roles of mycobiome and virome remain relatively underexplored. Moreover, current advancements in gut microbiome research have emphasized therapeutic applications—such as probiotics, prebiotics, and fecal microbiota transplantation—rather than the development of diagnostic tools.¹⁵⁴

Extracellular Vesicle Detection

Extracellular vesicles (EVs), including apoptotic bodies, microvesicles and exosomes, carry biomolecules like proteins and RNAs to modulate cellular activity.¹⁵⁶ In recent years, the potential of EVs in AD diagnosis has attracted much attention, especially as biomarkers.

A β Metabolism and Tau Phosphorylation-Related Proteins

EVs derived from neurons and glial cells transport proteins implicated in A β metabolism, including APP, β -secretase (BACE1), and γ -secretase complex components such as presenilin-1 and presenilin-2.¹⁵⁷ Dysregulation of these proteins contributes to neurotoxic A β accumulation, and alterations in their levels within EVs may serve as potential biomarkers for AD diagnosis and disease progression monitoring. For instance, studies on CSF-derived EVs in AD patients have reported significant alterations in the levels of proteins such as Heat shock protein A1A (HSPA1A), and Prostaglandin F2 receptor negative regulator (PTGFRN) throughout disease progression.¹⁵⁸ EVs also contain proteins involved in Tau phosphorylation, including Tau kinases such as GSK-3 β and CDK-5, and phosphatases such as protein phosphatase 2A. Aberrant Tau phosphorylation is a hallmark of AD and serves as a predictive marker for cognitive decline and a metric for assessing treatment efficacy.¹⁵⁹

miRNA Biomarkers and Lipid-Related Biomarkers

EVs protect miRNAs from degradation, enhancing their stability and clinical relevance.¹⁶⁰ A study reported that the number of AD dementia-related miRNAs in EVs was nearly twice that in serum. Moreover, EV-derived miRNAs exhibited a stronger correlation with medial temporal lobe atrophy, a neuroimaging biomarker of AD pathology, underscoring their diagnostic superiority.¹⁶¹ Lipidomic analysis of brain-derived EVs (BDEVs) reveals AD-associated dysregulation, including altered glycerophospholipids, sphingolipids, and reduced docosahexaenoic acid. Compared to bulk lipid analysis of the frontal cortex, BDEVs offer superior sensitivity in detecting lipid dysregulation associated with AD, suggesting their utility for early AD diagnosis through peripheral blood studies.¹⁶²

Other Protein Biomarkers

EVs contain neuroinflammatory proteins (IL-1 β , TNF- α , CCL2, CXCL10), synaptic dysfunction-associated proteins (synaptic vesicle proteins, neurotransmitter receptors, synaptic scaffold proteins), and neuronal injury markers (neuron-specific enolase, neurofilament light chain, and ubiquitin carboxy-terminal hydrolase L1). Increased levels of these EV-associated biomarkers correlate with disease severity and cognitive decline in AD patients, highlighting their potential utility in monitoring disease progression and therapeutic response.¹⁶³

Advantages and Challenges of Extracellular Vesicle Detection in AD Diagnosis

Extracellular vesicle detection offers significant advantages in AD diagnosis: their lipid bilayer ensures cargo stability against enzymatic degradation, improving reliability. EVs carry molecules reflecting AD-specific neuropathology, providing mechanistic insights. Early biomarker changes in EVs enable potential early-stage detection for timely intervention. However, standardization issues in isolation methods and variability in biomarker specificity hinder clinical translation, necessitating further research.¹⁶⁴

Artificial Intelligence-Assisted Detection

Artificial intelligence (AI) has greatly enhanced the efficiency and accuracy of detecting AD. AI technologies, such as machine learning and deep learning, enable the analysis of neuroimaging data from CT, MRI, and PET to detect early pathological features of AD, offering robust support for early diagnosis and treatment.¹⁶⁵

AI Technologies for AD Detection

Machine learning and deep learning play pivotal roles in AD detection and diagnosis. Researchers employ deep learning models for diagnosis, prognosis prediction, and forecasting patient health outcomes following pharmacological interventions. For instance, deep learning can automatically extract features from neuroimaging data, assisting radiologists in accurate diagnosis. Various models such as convolutional neural network, recurrent neural network, and transfer learning are applied to process multimodal data like PET and MRI, enabling AD detection, segmentation, and severity grading. Compared with traditional methods, it reduces the subjectivity and time consumption of manual feature extraction, improving diagnostic efficiency and accuracy, providing new directions and technical support for AD diagnosis.¹⁶⁶

Deep learning techniques offer notable advantages in processing complex three-dimensional data and are extensively utilized in AD research. For example, integrating 3D convolutional neural network with PET imaging achieves 96% accuracy in AD versus normal control classification and 84.2% accuracy in MCI converter versus non-converter classification.¹⁶⁷ Other deep learning architectures, including artificial neural networks and recurrent neural networks, have also been employed to predict AD progression.

Advantages and Challenges of AI in AD Detection

AI overcomes limitations of manual detection by autonomously extracting features from neuroimaging data, improving early diagnosis. Moreover, AI models, particularly deep learning-based algorithms, efficiently process large-scale neuroimaging data to detect complex pathological patterns.¹⁶⁸ It enhances accuracy by detecting complex patterns and quantifying biomarkers like amyloid plaques. AI also enables precise image segmentation, aiding quantitative analysis of disease-related changes thereby providing objective and accurate data for AD diagnosis, monitoring, and treatment evaluation.^{169,170}

Despite its potential, AI encounters challenges in AD detection. These include disease heterogeneity, subtle early symptoms overlapping with other conditions, scarce longitudinal data, ethical issues like informed consent, data privacy concerns, difficulties in clinical integration, and limited diverse training datasets, necessitating further clinical validation studies.¹⁷¹

Conclusion and Future Perspectives

In recent years, advances in biomedical research have deepened our understanding of AD across multiple critical domains, including pathophysiological mechanisms, imaging modalities, and emerging diagnostic technologies. Pathologically, AD is driven by a complex interplay of mechanisms, including A β aggregation, hyperphosphorylated Tau-mediated NFT formation, synaptic dysfunction, neuroinflammation, gut-brain axis dysregulation, and OS, each contributing to a self-perpetuating cycle of neurodegeneration. In terms of imaging, molecular and structural techniques remain foundational for early diagnosis. PET enables visualization of core biomarkers like A β and Tau, while MRI captures structural and functional brain changes. Complementing these, retinal imaging offers non-invasive insights into neurodegeneration via retinal structural and vascular alterations, emerging as a promising screening tool. Emerging technologies further expand diagnostic capabilities: nanotechnology enhances biomarker detection sensitivity; gut microbiome analysis reveals bidirectional microbiota-brain interactions; EVs carry AD-specific molecules for early detection; and AI integrates multimodal data to boost diagnostic accuracy.

Early diagnosis, enabled by these advancements, is clinically pivotal. Detecting AD in its preclinical or prodromal stages is paramount for implementing timely interventions, potentially delaying cognitive decline, improving patient outcomes, and enhancing patients' quality of life. Multimodal strategies, combining biomarkers, diverse imaging techniques, and novel technologies, are key to overcoming single-modality limitations and enhancing diagnostic precision. Such multimodal strategies lay the foundation for timely interventions and hold crucial clinical significance for improving AD prognosis.

Looking forward, AD research will focus on standardizing multimodal imaging protocols, developing multi-target therapeutic interventions, and deepening AI integration with molecular diagnostics. Interdisciplinary collaboration across clinical medicine, neuroscience, AI, and bioengineering will be critical to translating these innovations into personalized precision therapies. Such progress holds the potential to transform AD into a condition amenable to early, targeted intervention—offering transformative hope to millions worldwide.

Data Sharing Statement

All data generated or analyzed during this study are included in this published article.

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Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

References

1. Chambers-Richards T, Chireh B, D'Arcy C. Trends in prevalence of self-reports of Alzheimer's disease/dementia among non-institutionalized individuals 45+ in Canada, 1994–2014. *Journal of Public Health Research*. 2022;11(4):22799036221135221. doi:10.1177/22799036221135221
2. Rafii MS, Aisen PS. Detection and treatment of Alzheimer's disease in its preclinical stage. *Nat Aging*. 2023;3(5):520–531. doi:10.1038/s43587-023-00410-4
3. (ADI) AsDI. Dementia statistics. Available from: <https://www.alzint.org/about/dementia-facts-figures/dementia-statistics/>. Accessed September 06, 2025.
4. Zhang Y, Chen H, Li R, Sterling K, Song W. Amyloid β -based therapy for Alzheimer's disease: challenges, successes and future. *Signal Transduct Target Ther*. 2023;8(1):248. doi:10.1038/s41392-023-01484-7
5. Gu L, Guo Z. Alzheimer's A β 42 and A β 40 form mixed oligomers with direct molecular interactions. *Biochem Biophys Res Commun*. 2021;534:292–296. doi:10.1016/j.bbrc.2020.11.092
6. Miao JF, Ma HX, Yang Y, et al. Microglia in Alzheimer's disease: pathogenesis, mechanisms, and therapeutic potentials. *Front Aging Neurosci*. 2023;15:1201982. doi:10.3389/fnagi.2023.1201982
7. Sayas CL, Ávila J. GSK-3 and Tau: a Key Duet in Alzheimer's Disease. *Cells*. 2021;10(4):721. doi:10.3390/cells10040721
8. Kuruva CS, Reddy PH. Amyloid beta modulators and neuroprotection in Alzheimer's disease: a critical appraisal. *Drug Discov Today*. 2017;22(2):223–233. doi:10.1016/j.drudis.2016.10.010
9. Jackson GR, Wiedau-Pazos M, Sang TK, et al. Human wild-type tau interacts with wingless pathway components and produces neurofibrillary pathology in *Drosophila*. *Neuron*. 2002;34(4):509–519. doi:10.1016/S0896-6273(02)00706-7
10. Guo T, Zhang D, Zeng Y, Huang TY, Xu H, Zhao Y. Molecular and cellular mechanisms underlying the pathogenesis of Alzheimer's disease. *Mol Neurodegener*. 2020;15(1):40. doi:10.1186/s13024-020-00391-7
11. Jack CR, Holtzman DM. Biomarker modeling of Alzheimer's disease. *Neuron*. 2013;80(6):1347–1358. doi:10.1016/j.neuron.2013.12.003
12. Li X, Lei P, Tuo Q, et al. Enduring Elevations of Hippocampal Amyloid Precursor Protein and Iron Are Features of β -Amyloid Toxicity and Are Mediated by Tau. *Neurotherapeutics*. 2015;12(4):862–873. doi:10.1007/s13311-015-0378-2
13. Esteras N, Abramov AY. Mitochondrial Calcium Deregulation in the Mechanism of Beta-Amyloid and Tau Pathology. *Cells*. 2020;9(9):2135. doi:10.3390/cells9092135
14. Atlante A, Valenti D, Latina V, Amadoro G. Dysfunction of Mitochondria in Alzheimer's Disease: ANT and VDAC Interact with Toxic Proteins and Aid to Determine the Fate of Brain Cells. *Int J Mol Sci*. 2022;23(14):7722. doi:10.3390/ijms23147722
15. Heneka MT, Carson MJ, El Khoury J, et al. Neuroinflammation in Alzheimer's disease. *Lancet Neurol*. 2015;14(4):388–405. doi:10.1016/S1474-4422(15)70016-5
16. Lawrence JM, Schardin K, Wigdahl B, Nonnemacher MR. Roles of neuropathology-associated reactive astrocytes: a systematic review. *Acta Neuropathol Commun*. 2023;11(1):42. doi:10.1186/s40478-023-01526-9
17. Wischik CM, Novak M, Edwards PC, Klug A, Tichelaar W, Crowther RA. Structural characterization of the core of the paired helical filament of Alzheimer disease. *Proc Natl Acad Sci U S A*. 1988;85(13):4884–4888. doi:10.1073/pnas.85.13.4884
18. Jung HJ, Park SS, Mok JO, Lee TK, Park CS, Park SA. Increased expression of three-repeat isoforms of tau contributes to tau pathology in a rat model of chronic type 2 diabetes. *Exp Neurol*. 2011;228(2):232–241. doi:10.1016/j.expneurol.2011.01.012

19. Evans LD, Wassmer T, Fraser G, et al. Extracellular Monomeric and Aggregated Tau Efficiently Enter Human Neurons through Overlapping but Distinct Pathways. *Cell Rep.* 2018;22(13):3612–3624. doi:10.1016/j.celrep.2018.03.021
20. Zhang HQ, Wei W, Zhao M, et al. Interaction between A β and Tau in the Pathogenesis of Alzheimer's Disease. *Int J Bio Sci.* 2021;17(9):2181–2192. doi:10.7150/ijbs.57078
21. Mattsson N, Lonneborg A, Boccardi M, Blennow K, Hansson O, Geneva Task Force Roadmap A. Clinical validity of cerebrospinal fluid A β 42, tau, and phospho-tau as biomarkers for Alzheimer's disease in the context of a structured 5-phase development framework. *Neurobiol Aging.* 2017;52:196–213. doi:10.1016/j.neurobiolaging.2016.02.034
22. Milà-Alomà M, Ashton NJ, Shekari M, et al. Plasma p-tau231 and p-tau217 as state markers of amyloid- β pathology in preclinical Alzheimer's disease. *Nat Med.* 2022;28(9):1797–1801. doi:10.1038/s41591-022-01925-w
23. Ferreira-Vieira TH, Guimaraes IM, Silva FR, Ribeiro FM. Alzheimer's disease: targeting the Cholinergic System. *Curr Neuropharmacol.* 2016;14(1):101–115. doi:10.2174/1570159X13666150716165726
24. Jia JP, Jia JM, Zhou WD, et al. Differential regional acetylcholine and choline concentrations in the cerebrospinal fluid of patients with Alzheimer's disease and vascular dementia. *Chin Med J.* 2004;117(8):1161–1164.
25. Scheff SW, Price DA, Schmitt FA, Scheff MA, Mufson EJ. Synaptic loss in the inferior temporal gyrus in mild cognitive impairment and Alzheimer's disease. *J Alzheimers Dis.* 2011;24(3):547–557. doi:10.3233/JAD-2011-101782
26. Ogawa M, Iida Y, Nakagawa M, et al. Change of central cholinergic receptors following lesions of nucleus basalis magnocellularis in rats: search for an imaging index suitable for the early detection of Alzheimer's disease. *Nucl Med Biol.* 2006;33(2):249–254. doi:10.1016/j.nucmedbio.2005.06.013
27. Ikonomic MD, Mufson EJ, Wu J, Bennett DA, DeKosky ST. Reduction of choline acetyltransferase activity in primary visual cortex in mild to moderate Alzheimer's disease. *Arch Neurol.* 2005;62(3):425–430. doi:10.1001/archneur.62.3.425
28. Meftah S, Gan J. Alzheimer's disease as a synaptopathy: evidence for dysfunction of synapses during disease progression. *Front Synaptic Neurosci.* 2023;15:1129036. doi:10.3389/fnsyn.2023.1129036
29. Montero-Crespo M, Domínguez-álvaro M, Alonso-Nanclares L, DeFelipe J, Blázquez-Llorca L. Three-dimensional analysis of synaptic organization in the hippocampal CA1 field in Alzheimer's disease. *Brain.* 2020;144(2):553–573. doi:10.1093/brain/awaa046
30. Kong Y, Huang L, Li W, et al. The Synaptic Vesicle Protein 2A Interacts With Key Pathogenic Factors in Alzheimer's Disease: implications for Treatment. *Front Cell Develop Biol.* 2020;9:609908. doi:10.3389/fcell.2021.609908
31. Reddy PH, Mani G, Park BS, et al. Differential loss of synaptic proteins in Alzheimer's disease: implications for synaptic dysfunction. *J Alzheimers Dis.* 2005;7(2):103–117. discussion 73–80. doi:10.3233/JAD-2005-7203
32. Kurucu H, Colom-Cadena M, Davies C, et al. Inhibitory synapse loss and accumulation of amyloid beta in inhibitory presynaptic terminals in Alzheimer's disease. *Eur J Neurol.* 2022;29(5):1311–1323. doi:10.1111/ene.15043
33. Barbour AJ, Gourmaud S, Lancaster E, et al. Seizures exacerbate excitatory: inhibitory imbalance in Alzheimer's disease and 5XFAD mice. *Brain.* 2024;147(6):2169–2184. doi:10.1093/brain/awae126
34. van van Hugte EJH, Schubert D, Nadif Kasri N. Excitatory/inhibitory balance in epilepsies and neurodevelopmental disorders: depolarizing γ -aminobutyric acid as a common mechanism. *Epilepsia.* 2023;64(8):1975–1990. doi:10.1111/epi.17651
35. Hampel H, Caraci F, Cuello AC, et al. A Path Toward Precision Medicine for Neuroinflammatory Mechanisms in Alzheimer's Disease. *Front Immunol.* 2020;11:456. doi:10.3389/fimmu.2020.00456
36. Si ZZ, Zou CJ, Mei X, et al. Targeting neuroinflammation in Alzheimer's disease: from mechanisms to clinical applications. *Neural Regen Res.* 2023;18(4):708–715. doi:10.4103/1673-5374.353484
37. Miklossy J, Martins RN, Schwab C, McGeer PL. Inflammatory Aspects of Alzheimer Disease and Other Neurodegenerative Disorders. *J Alzheimers Dis.* 2008;13(4):359–369. doi:10.3233/JAD-2008-13402
38. Hong S, Beja-Glasser VF, Nfonoyim BM, et al. Complement and microglia mediate early synapse loss in Alzheimer mouse models. *Science.* 2016;352(6286):712–716. doi:10.1126/science.aad8373
39. Schnöder L, Hao W, Qin Y-D, et al. Deficiency of Neuronal p38 α MAPK Attenuates Amyloid Pathology in Alzheimer Disease Mouse and Cell Models through Facilitating Lysosomal Degradation of BACE1*. *J Biol Chem.* 2015;291(5):2067–2079. doi:10.1074/jbc.M115.695916
40. Chen Z, Balachandran YL, Chong WP, Chan K WY. Roles of Cytokines in Alzheimer's Disease. *Int J Mol Sci.* 2024;25(11):5803. doi:10.3390/ijms25115803
41. Roy ER, Chiu G, Li S, et al. Concerted type I interferon signaling in microglia and neural cells promotes memory impairment associated with amyloid β plaques. *Immunity.* 2022;55(5):879–94.e6. doi:10.1016/j.immuni.2022.03.018
42. Hur JY, Frost GR, Wu X, et al. The innate immunity protein IFITM3 modulates γ -secretase in Alzheimer's disease. *Nature.* 2020;586(7831):735–740. doi:10.1038/s41586-020-2681-2
43. Zhang G, Peng Q, Guo X, et al. Microglia-derived Galectin-9 drives amyloid- β pathology in Alzheimer's disease. *Aging Cell.* 2025;24(2):e14396. doi:10.1111/accel.14396
44. Jian M, Kwan JS, Bunting M, Ng RC, Chan KH. Adiponectin suppresses amyloid- β oligomer (A β O)-induced inflammatory response of microglia via AdipoR1-AMPK-NF- κ B signaling pathway. *J Neuroinflammation.* 2019;16(1):110. doi:10.1186/s12974-019-1492-6
45. Torrisi SA, Geraci F, Tropea MR, et al. Fluoxetine and Vortioxetine Reverse Depressive-Like Phenotype and Memory Deficits Induced by A β (1–42) Oligomers in Mice: a Key Role of Transforming Growth Factor- β 1. *Front Pharmacol.* 2019;10:693. doi:10.3389/fphar.2019.00693
46. Mawuenyega KG, Sigurdson W, Ovod V, et al. Decreased clearance of CNS beta-amyloid in Alzheimer's disease. *Science.* 2010;330(6012):1774. doi:10.1126/science.1197623
47. Das TK, Ganesh BP. Interlink between the gut microbiota and inflammation in the context of oxidative stress in Alzheimer's disease progression. *Gut Microbes.* 2023;15(1):2206504. doi:10.1080/19490976.2023.2206504
48. Manen KV, Guariglia S, Del C. Alonso A, McCoy EC. Expression of neuronal protein Tau in *Candida albicans*. *Journal of Yeast and Fungal Research.* 2014;5(5):67–73. doi:10.5897/JYFR2014.0136
49. Tetz G, Pinho M, Pritzkow S, Mendez N, Soto C, Tetz V. Bacterial DNA promotes Tau aggregation. *Sci Rep.* 2020;10(1):2369. doi:10.1038/s41598-020-59364-x
50. Kim MS, Kim Y, Choi H, et al. Transfer of a healthy microbiota reduces amyloid and tau pathology in an Alzheimer's disease animal model. *Gut.* 2020;69(2):283–294. doi:10.1136/gutjnl-2018-317431

51. Radak Z, Zhao Z, Goto S, Koltai E. Age-associated neurodegeneration and oxidative damage to lipids, proteins and DNA. *Mol Aspects Med.* 2011;32(4–6):305–315. doi:10.1016/j.mam.2011.10.010
52. Fonseca ACRG, Moreira PI, Oliveira CR, Cardoso SM, Pinton P, Pereira CF. Amyloid-Beta Disrupts Calcium and Redox Homeostasis in Brain Endothelial Cells. *Molecular Neurobiology.* 2015;51(2):610–622. doi:10.1007/s12035-014-8740-7
53. Constantino LC, Vandresen-Filho S, Tasca CI. Neuroprotection induced by NMDA preconditioning as a strategy to understand brain tolerance mechanism. *Neural Regen Res.* 2015;10(4):542–543. doi:10.4103/1673-5374.155415
54. Park L, Anrather J, Zhou P, et al. NADPH-oxidase-derived reactive oxygen species mediate the cerebrovascular dysfunction induced by the amyloid beta peptide. *J Neurosci.* 2005;25(7):1769–1777. doi:10.1523/JNEUROSCI.5207-04.2005
55. Vermunt L, Sikkes SAM, van den Hout A, et al. Duration of preclinical, prodromal, and dementia stages of Alzheimer's disease in relation to age, sex, and APOE genotype. *Alzheimers Dement.* 2019;15(7):888–898. doi:10.1016/j.jalz.2019.04.001
56. Laforce R, Buteau JP, Paquet N, Verret L, Houde M, Bouchard RW. The value of PET in mild cognitive impairment, typical and atypical/unclear dementias: a retrospective memory clinic study. *Am J Alzheimers Dis Other Demen.* 2010;25(4):324–332. doi:10.1177/1533317510363468
57. Morinaga A, Ono K, Ikeda T, et al. A comparison of the diagnostic sensitivity of MRI, CBF-SPECT, FDG-PET and cerebrospinal fluid biomarkers for detecting Alzheimer's disease in a memory clinic. *Dement Geriatr Cognit Disord.* 2010;30(4):285–292. doi:10.1159/000320265
58. Almeida ZL, Vaz DC, Brito RMM. Morphological and Molecular Profiling of Amyloid- β Species in Alzheimer's Pathogenesis. *Molecular Neurobiology.* 2025;62(4):4391–4419. doi:10.1007/s12035-024-04543-4
59. Beach TG, Schneider JA, Sue LI, et al. Theoretical impact of Florbetapir (18F) amyloid imaging on diagnosis of Alzheimer dementia and detection of preclinical cortical amyloid. *J Neuropathol Exp Neurol.* 2014;73(10):948–953. doi:10.1097/NEN.0000000000000114
60. Bergeron D, Gorno-Tempini ML, Rabinovici GD, et al. Prevalence of amyloid- β pathology in distinct variants of primary progressive aphasia. *Ann Neurol.* 2018;84(5):729–740. doi:10.1002/ana.25333
61. Librizzi D, Cabanel N, Zavorotnyy M, et al. Clinical Relevance of [(18F)]Florbetaben and [(18F)]FDG PET/CT Imaging on the Management of Patients with Dementia. *Molecules.* 2021;26(5):1282. doi:10.3390/molecules26051282
62. Martínez G, Veranooj RW, Fuentes Padilla P, Zamora J, Bonfill Cosp X, Flicker L. 18F PET with florbetapir for the early diagnosis of Alzheimer's disease dementia and other dementias in people with mild cognitive impairment (MCI). *Cochrane Database Syst Rev.* 2017;11(11):Cd012216. doi:10.1002/14651858.CD012216.pub2
63. Therriault J, Benedet AL, Pascoal TA, et al. Determining Amyloid- β Positivity Using (18F)-AZD4694 PET Imaging. *J Nucl Med.* 2021;62(2):247–252. doi:10.2967/jnumed.120.245209
64. Aliaga A, Therriault J, Quispialaya K, et al. Autoradiographic comparison between [(11C)]PiB and [(18F)]AZD4694 in human brain tissue. *EJNMMI Res.* 2025;15(1):30. doi:10.1186/s13550-025-01216-8
65. Klunk WE, Engler H, Nordberg A, et al. Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B. *Ann Neurol.* 2004;55(3):306–319. doi:10.1002/ana.20009
66. Myburgh PJ, Sai KKS. Two decades of 11C PiB synthesis, 2003-2023: a review. *Am. J. Nucl. Med. Mol. Imaging.* 2024;14(1):48–62. doi:10.62347/ADSK6584
67. Curtis C, Gamez JE, Singh U, et al. Phase 3 Trial of Flutemetamol Labeled With Radioactive Fluorine 18 Imaging and Neuritic Plaque Density. *JAMA Neurol.* 2015;72(3):287–294. doi:10.1001/jamaneurol.2014.4144
68. Zeydan B, Schwarz CG, Przybelski SA, et al. Comparison of (11C)-Pittsburgh Compound B and (18F)-Flutemetamol White Matter Binding in PET. *J Nucl Med.* 2022;63(8):1239–1244. doi:10.2967/jnumed.121.263281
69. Zeydan B, Johnson DR, Schwarz CG, et al. Visual assessments of 11 C-Pittsburgh compound-B PET vs. 18 F-flutemetamol PET across the age spectrum. *Nucl Med Commun.* 2024;45(12):1047–1054. doi:10.1097/MNM.0000000000001902
70. Alongi P, Chiaravalloti A, Berti V, et al. Amyloid PET in the diagnostic workup of neurodegenerative disease. *Clin Transl Imaging.* 2021;9(4):383–397. doi:10.1007/s40336-021-00428-x
71. Beach TG, Maarouf CL, Intorcía A, et al. Antemortem-Postmortem Correlation of Florbetapir (18F) PET Amyloid Imaging with Quantitative Biochemical Measures of A β 42 but not A β 40. *J Alzheimers Dis.* 2018;61(4):1509–1516. doi:10.3233/JAD-170762
72. Meier SR, Sehlin D, Roshanbin S, et al. 11C-PiB and 124I-antibody PET Provide Differing Estimates of Brain Amyloid- β After Therapeutic Intervention. *J Nucl Med.* 2022;63(2):302–309. doi:10.2967/jnumed.121.262083
73. Pike KE, Savage G, Villemagne VL, et al. β -amyloid imaging and memory in non-demented individuals: evidence for preclinical Alzheimer's disease. *Brain.* 2007;130(11):2837–2844. doi:10.1093/brain/awm238
74. Fleisher AS, Pontecorvo MJ, Devous MD, et al. Positron Emission Tomography Imaging With [18F]flortaucipir and Postmortem Assessment of Alzheimer Disease Neuropathologic Changes. *JAMA Neurol.* 2020;77(7):829–839. doi:10.1001/jamaneurol.2020.0528
75. Altomare D, Caprioglio C, Assal F, et al. Diagnostic value of amyloid-PET and tau-PET: a head-to-head comparison. *Eur J Nucl Med Mol Imaging.* 2021;48(7):2200–2211. doi:10.1007/s00259-021-05246-x
76. Groot C, Smith R, Collij LE, et al. Tau Positron Emission Tomography for Predicting Dementia in Individuals With Mild Cognitive Impairment. *JAMA Neurol.* 2024;81(8):845–856. doi:10.1001/jamaneurol.2024.1612
77. Biel D, Brendel M, Rubinski A, et al. Tau-PET and in vivo Braak-staging as prognostic markers of future cognitive decline in cognitively normal to demented individuals. *Alzheimers Res Ther.* 2021;13(1):137. doi:10.1186/s13195-021-00880-x
78. Kunach P, Vaquer-Alicea J, Smith MS, et al. Cryo-EM structure of Alzheimer's disease tau filaments with PET ligand MK-6240. *Nat Commun.* 2024;15(1):8497. doi:10.1038/s41467-024-52265-x
79. Zhao Q, Liu M, Ha L, Zhou Y. Quantitative (18F)-AV1451 Brain Tau PET Imaging in Cognitively Normal Older Adults, Mild Cognitive Impairment, and Alzheimer's Disease Patients. *Front Neurol.* 2019;10:486. doi:10.3389/fneur.2019.00486
80. Forsberg Morén A, Varrone A. Timing is everything: tau imaging across stages of Alzheimer's disease. *Brain.* 2020;143(9):2634–2636. doi:10.1093/brain/awaa220
81. Gérard T, Colmant L, Malotau V, et al. The spatial extent of tauopathy on [(18F)]MK-6240 tau PET shows stronger association with cognitive performances than the standard uptake value ratio in Alzheimer's disease. *Eur J Nucl Med Mol Imaging.* 2024;51(6):1662–1674. doi:10.1007/s00259-024-06603-2

82. Bourgeat P, Krishnadas N, Doré V, et al. Cross-Sectional and Longitudinal Comparison of Tau Imaging with 18F-MK6240 and 18F-Flortaucipir in Populations Matched for Age, MMSE and Brain Beta-Amyloid Burden. *J Prev Alzheimers Dis.* 2023;10(2):251–258. doi:10.14283/jpad.2023.17
83. Sison SA, Paclibar CG, Liang C, Mukherjee J. Radioiodinated Tau Imaging Agent III Molecular Modeling, Synthesis, and Evaluation of a New Tau Imaging Agent, [(125)I]ISAS in Post-Mortem Human Alzheimer's Disease Brain. *Molecules.* 2024;29(14):3308. doi:10.3390/molecules29143308
84. Mukherjee J, Liang C, Patel KK, Lam PQ, Mondal R. Development and evaluation of [(125) I]JIPPI for Tau imaging in postmortem human Alzheimer's disease brain. *Synapse.* 2021;75(1):e22183. doi:10.1002/syn.22183
85. Omachi Y, Ito K, Arima K, et al. Clinical impact of (11)C-Pittsburgh compound-B positron emission tomography carried out in addition to magnetic resonance imaging and single-photon emission computed tomography on the diagnosis of Alzheimer's disease in patients with dementia and mild cognitive impairment. *Psychiatry Clin Neurosci.* 2015;69(12):741–751. doi:10.1111/pcn.12326
86. Kobylecki C, Langheinrich T, Hinz R, et al. 18F-florbetapir PET in patients with frontotemporal dementia and Alzheimer disease. *J Nucl Med.* 2015;56(3):386–391. doi:10.2967/jnumed.114.147454
87. Ong KT, Villemagne VL, Bahar-Fuchs A, et al. Aβ imaging with 18F-florbetaben in prodromal Alzheimer's disease: a prospective outcome study. *J Neurol Neurosurg Psychiatry.* 2015;86(4):431–436. doi:10.1136/jnnp-2014-308094
88. Leuzy A, Savitcheva I, Chiotis K, et al. Clinical impact of [(18)F]flutemetamol PET among memory clinic patients with an unclear diagnosis. *Eur J Nucl Med Mol Imaging.* 2019;46(6):1276–1286. doi:10.1007/s00259-019-04297-5
89. Kimura Y, Ichise M, Ito H, et al. PET Quantification of Tau Pathology in Human Brain with 11C-PBB3. *J Nucl Med.* 2015;56(9):1359–1365. doi:10.2967/jnumed.115.160127
90. Mishra S, Gordon BA, Su Y, et al. AV-1451 PET imaging of tau pathology in preclinical Alzheimer disease: defining a summary measure. *Neuroimage.* 2017;161:171–178. doi:10.1016/j.neuroimage.2017.07.050
91. Lu M, Pontecorvo MJ, Devous MD, et al. Aggregated Tau Measured by Visual Interpretation of Flortaucipir Positron Emission Tomography and the Associated Risk of Clinical Progression of Mild Cognitive Impairment and Alzheimer Disease: results From 2 Phase III Clinical Trials. *JAMA Neurol.* 2021;78(4):445–453. doi:10.1001/jamaneurol.2020.5505
92. Lohith TG, Bennacef I, Vandenberghe R, et al. Brain Imaging of Alzheimer Dementia Patients and Elderly Controls with (18)F-MK-6240, a PET Tracer Targeting Neurofibrillary Tangles. *J Nucl Med.* 2019;60(1):107–114. doi:10.2967/jnumed.118.208215
93. Baker SL, Provost K, Thomas W, et al. Evaluation of [(18)F]-JNJ-64326067-AAA tau PET tracer in humans. *J Cereb Blood Flow Metab.* 2021;41(12):3302–3313. doi:10.1177/0271678X211031035
94. Chen YF, Wang JK, Cui CL, et al. Evaluating the association between brain atrophy, hypometabolism, and cognitive decline in Alzheimer's disease: a PET/MRI study. *Ageing-Us.* 2021;13(5):7228–7246. doi:10.18632/aging.202580
95. Bouter Y, Glasnek RM, Wenzel JM, Bouter C. 18F-FDG-PET and Multimodal Biomarker Integration: a Powerful Tool for Alzheimer's Disease Diagnosis. *Nuclear Medicine and Molecular Imaging.* 2025. doi:10.1007/s13139-025-00932-2
96. Ryoo HG, Choi H, Shi K, et al. Distinct subtypes of spatial brain metabolism patterns in Alzheimer's disease identified by deep learning-based FDG PET clusters. *Eur. J. Nucl. Med. Mol. Imaging.* 2024;51(2):443–454. doi:10.1007/s00259-023-06440-9
97. Na S, Kang DW, Kim GH, et al. The Usefulness of (18)F-FDG PET to Differentiate Subtypes of Dementia: the Systematic Review and Meta-Analysis. *Dement Neurocogn Disord.* 2024;23(1):54–66. doi:10.12779/dnd.2024.23.1.54
98. Quispialaya KM, Therriault J, Aliaga A, et al. Comparison of Plasma p-tau217 and [(18)F]FDG-PET for Identifying Alzheimer Disease in People With Early-Onset or Atypical Dementia. *Neurology.* 2025;104(2):e210211. doi:10.1212/WNL.000000000000210211
99. Burns CM, Chen K, Kaszniak AW, et al. Higher serum glucose levels are associated with cerebral hypometabolism in Alzheimer regions. *Neurology.* 2013;80(17):1557–1564. doi:10.1212/WNL.0b013e31828f17de
100. Apostolova I, Lange C, Suppa P, et al. Impact of plasma glucose level on the pattern of brain FDG uptake and the predictive power of FDG PET in mild cognitive impairment. *Eur J Nucl Med Mol Imaging.* 2018;45(8):1417–1422. doi:10.1007/s00259-018-3985-4
101. Terada T, Obi T, Bunai T, et al. In vivo mitochondrial and glycolytic impairments in patients with Alzheimer disease. *Neurology.* 2020;94(15):e1592–e604. doi:10.1212/WNL.00000000000009249
102. Yu B, Shan Y, Ding J. A literature review of MRI techniques used to detect amyloid-beta plaques in Alzheimer's disease patients. *Ann Palliat Med.* 2021;10(9):10062–10074. doi:10.21037/apm-21-825
103. Feng Q, Ding Z. MRI Radiomics Classification and Prediction in Alzheimer's Disease and Mild Cognitive Impairment: a Review. *Curr Alzheimer Res.* 2020;17(3):297–309. doi:10.2174/1567205017666200303105016
104. Pini L, Pievani M, Bocchetta M, et al. Brain atrophy in Alzheimer's Disease and aging. *Ageing Res Rev.* 2016;30:25–48. doi:10.1016/j.arr.2016.01.002
105. Lombardi G, Crescioli G, Cavado E, et al. Structural magnetic resonance imaging for the early diagnosis of dementia due to Alzheimer's disease in people with mild cognitive impairment. *Cochrane Database Syst Rev.* 2020;3(3):Cd009628. doi:10.1002/14651858.CD009628.pub2
106. Farina FR, Emek-Savaş DD, Rueda-Delgado L, et al. A comparison of resting state EEG and structural MRI for classifying Alzheimer's disease and mild cognitive impairment. *Neuroimage.* 2020;215:116795. doi:10.1016/j.neuroimage.2020.116795
107. Pan D, Zeng A, Yang B, et al. Deep Learning for Brain MRI Confirms Patterned Pathological Progression in Alzheimer's Disease. *Adv Sci.* 2023;10(6):e2204717. doi:10.1002/adv.202204717
108. Aramadaka S, Mannam R, Sankara Narayanan R, et al. Neuroimaging in Alzheimer's Disease for Early Diagnosis: a Comprehensive Review. *Cureus.* 2023;15(5):e38544. doi:10.7759/cureus.38544
109. Finn ES, Poldrack RA, Shine JM. Functional neuroimaging as a catalyst for integrated neuroscience. *Nature.* 2023;623(7986):263–273. doi:10.1038/s41586-023-06670-9
110. Johnson KA, Fox NC, Sperling RA, Klunk WE. Brain imaging in Alzheimer disease. *Cold Spring Harb Perspect Med.* 2012;2(4):a006213. doi:10.1101/cshperspect.a006213
111. Ibrahim B, Suppiah S, Ibrahim N, et al. Diagnostic power of resting-state fMRI for detection of network connectivity in Alzheimer's disease and mild cognitive impairment: a systematic review. *Hum Brain Mapp.* 2021;42(9):2941–2968. doi:10.1002/hbm.25369
112. Sadeghi MA, Stevens D, Kundu S, et al. Detecting Alzheimer's Disease Stages and Frontotemporal Dementia in Time Courses of Resting-State fMRI Data Using a Machine Learning Approach. *J Imaging Inform Med.* 2024;37(6):2768–2783. doi:10.1007/s10278-024-01101-1

113. Forouzannezhad P, Abbaspour A, Fang C, et al. A survey on applications and analysis methods of functional magnetic resonance imaging for Alzheimer's disease. *J Neurosci Methods*. 2019;317:121–140. doi:10.1016/j.jneumeth.2018.12.012
114. Perosa V, Oltmer J, Munting LP, et al. Perivascular space dilation is associated with vascular amyloid- β accumulation in the overlying cortex. *Acta Neuropathol*. 2022;143(3):331–348. doi:10.1007/s00401-021-02393-1
115. Romero JR, Pinheiro A, Aparicio HJ, DeCarli CS, Demissie S, Seshadri S. MRI-Visible Perivascular Spaces and Risk of Incident Dementia: the Framingham Heart Study. *Neurology*. 2022;99(23):e2561–e71. doi:10.1212/WNL.00000000000021293
116. Voorter PHM, van Dinther M, Jansen WJ, et al. Blood-Brain Barrier Disruption and Perivascular Spaces in Small Vessel Disease and Neurodegenerative Diseases: a Review on MRI Methods and Insights. *J Magn Reson Imaging*. 2024;59(2):397–411. doi:10.1002/jmri.28989
117. Barisano G, Lynch KM, Sibilia F, et al. Imaging perivascular space structure and function using brain MRI. *Neuroimage*. 2022;257:119329. doi:10.1016/j.neuroimage.2022.119329
118. Matsuo K, Nshihara H. Rebuilding insight into the pathophysiology of Alzheimer's disease through new blood-brain barrier models. *Neural Regen Res*. 2024;19(9):1954–1960. doi:10.4103/1673-5374.390978
119. Chen G, Li H, Shao X, et al. Decreased water exchange rate across the blood-brain barrier throughout the Alzheimer's disease continuum: evidence from Chinese data. *Alzheimers Dement*. 2025;21(3):e70089. doi:10.1002/alz.70089
120. Ehman EC, Johnson GB, Villanueva-Meyer JE, et al. PET/MRI: where might it replace PET/CT? *J Magn Reson Imaging*. 2017;46(5):1247–1262. doi:10.1002/jmri.25711
121. Gao F. Integrated Positron Emission Tomography/Magnetic Resonance Imaging in clinical diagnosis of Alzheimer's disease. *Eur J Radiol*. 2021;145:110017. doi:10.1016/j.ejrad.2021.110017
122. Shepherd TM, Dogra S. Clinical Translation of Integrated PET-MRI for Neurodegenerative Disease. *J Magn Reson Imaging*. 2025. doi:10.1002/jmri.70046
123. London A, Benhar I, Schwartz M. The retina as a window to the brain—from eye research to CNS disorders. *Nat Rev Neurol*. 2013;9(1):44–53. doi:10.1038/nrneurol.2012.227
124. Mammadova N, Nepl TK, Denburg NL, West Greenlee MH. Reduced Retinal Thickness Predicts Age-Related Changes in Cognitive Function. *Front Aging Neurosci*. 2020;12:81. doi:10.3389/fnagi.2020.00081
125. Zhang Y, Wang Y, Shi C, Shen M, Lu F. Advances in retina imaging as potential biomarkers for early diagnosis of Alzheimer's disease. *Transl Neurodegener*. 2021;10(1):6. doi:10.1186/s40035-021-00230-9
126. Mejia-Vergara AJ, Karanjia R, Sadun AA. OCT parameters of the optic nerve head and the retina as surrogate markers of brain volume in a normal population, a pilot study. *J Neurol Sci*. 2021;420:117213. doi:10.1016/j.jns.2020.117213
127. Gaire BP, Koronyo Y, Fuchs DT, et al. Alzheimer's disease pathophysiology in the Retina. *Prog Retin Eye Res*. 2024;101:101273. doi:10.1016/j.preteyeres.2024.101273
128. Smith EE, Beaudin AE. New insights into cerebral small vessel disease and vascular cognitive impairment from MRI. *Curr Opin Neurol*. 2018;31(1):36–43. doi:10.1097/WCO.0000000000000513
129. Klyucherev TO, Olszewski P, Shalimova AA, et al. Advances in the development of new biomarkers for Alzheimer's disease. *Transl Neurodegener*. 2022;11(1):25. doi:10.1186/s40035-022-00296-z
130. López-Cuenca I, Salobrar-García E, Elvira-Hurtado L, et al. The Value of OCT and OCTA as Potential Biomarkers for Preclinical Alzheimer's Disease: a Review Study. *Life (Basel)*. 2021;11(7):712. doi:10.3390/life11070712
131. Ibrahim Y, Xie J, Macerollo A, et al. A Systematic Review on Retinal Biomarkers to Diagnose Dementia from OCT/OCTA Images. *J Alzheimers Dis Rep*. 2023;7(1):1201–1235. doi:10.3233/ADR-230042
132. Ma JP, Robbins CB, Stinnett SS, et al. Repeatability of Peripapillary OCT Angiography in Neurodegenerative Disease. *Ophthalmol Sci*. 2021;1(4):100075. doi:10.1016/j.xops.2021.100075
133. Cipollini V, Abdolrahimzadeh S, Troili F, et al. Neurocognitive Assessment and Retinal Thickness Alterations in Alzheimer Disease: is There a Correlation? *J Neuroophthalmol*. 2020;40(3):370–377. doi:10.1097/WNO.0000000000000831
134. van de Kreeke JA, Legdeur N, Badissi M, et al. Ocular biomarkers for cognitive impairment in nonagenarians; a prospective cross-sectional study. *BMC Geriatr*. 2020;20(1):155. doi:10.1186/s12877-020-01556-1
135. van de Kreeke JA, Nguyen HT, Konijnenberg E, et al. Longitudinal retinal layer changes in preclinical Alzheimer's disease. *Acta Ophthalmol*. 2021;99(5):538–544. doi:10.1111/aos.14640
136. Ashok A, Singh N, Chaudhary S, et al. Retinal Degeneration and Alzheimer's Disease: an Evolving Link. *Int J Mol Sci*. 2020;21(19):7290. doi:10.3390/ijms21197290
137. Cheung CY, Mok V, Foster PJ, Trucco E, Chen C, Wong TY. Retinal imaging in Alzheimer's disease. *J Neurol Neurosurg Psychiatry*. 2021;92(9):983–994. doi:10.1136/jnnp-2020-325347
138. Casciano F, Zauli E, Celeghini C, et al. Retinal Alterations Predict Early Prodromal Signs of Neurodegenerative Disease. *Int J Mol Sci*. 2024;25(3):1689. doi:10.3390/ijms25031689
139. Lee K, Mahmud M, Marx D, et al. Clinical Arterial Spin-Labeling MR Imaging to Screen for Typical and Atypical Neurodegenerative Disease in the New Era of Alzheimer Treatment. *AJNR Am J Neuroradiol*. 2024;45(5):632–636. doi:10.3174/ajnr.A8164
140. Xie L, Wisse LEM, Das SR, et al. Longitudinal atrophy in early Braak regions in preclinical Alzheimer's disease. *Hum Brain Mapp*. 2020;41(16):4704–4717. doi:10.1002/hbm.25151
141. Yu J, Zhou R, Liu S, et al. Electrochemical Biosensors for the Detection of Exosomal microRNA Biomarkers for Early Diagnosis of Neurodegenerative Diseases. *Anal Chem*. 2025;97(10):5355–5371. doi:10.1021/acs.analchem.4c02619
142. Song N, Sun S, Chen K, et al. Emerging nanotechnology for Alzheimer's disease: from detection to treatment. *J Control Release*. 2023;360:392–417. doi:10.1016/j.jconrel.2023.07.004
143. Plascencia-Villa G, Ponce A, Collingwood JF, et al. High-resolution analytical imaging and electron holography of magnetite particles in amyloid cores of Alzheimer's disease. *Sci Rep*. 2016;6(1):24873. doi:10.1038/srep24873
144. Panghal A, Flora SJS. Nanotechnology in the diagnostic and therapy for Alzheimer's disease. *Biochim Biophys Acta Gen Subj*. 2024;1868(3):130559. doi:10.1016/j.bbagen.2024.130559
145. Chopra H, Bibi S, Singh I, et al. Nanomedicines in the Management of Alzheimer's Disease: current View and Future Prospects. *Front Aging Neurosci*. 2022;14:879114. doi:10.3389/fnagi.2022.879114

146. Sinha T, Bokhari SFH, Khan MU, et al. Gazing Beyond the Horizon: a Systematic Review Unveiling the Theranostic Potential of Quantum Dots in Alzheimer's Disease. *Cureus*. 2024;16(4):e58677. doi:10.7759/cureus.58677
147. Carneiro P, Morais S, Pereira MC. Nanomaterials towards Biosensing of Alzheimer's Disease Biomarkers. *Nanomaterials*. 2019;9(12):1663. doi:10.3390/nano9121663
148. Gopalan D, Pandey A, Alex AT, et al. Nanoconstructs as a versatile tool for detection and diagnosis of Alzheimer biomarkers. *Nanotechnology*. 2021;32(14):142002. doi:10.1088/1361-6528/abcdcb
149. Farrokhi V, Nemati R, Nichols FC, et al. Bacterial lipopeptide, Lipid 654, is a microbiome-associated biomarker for multiple sclerosis. *Clin Transl Immunology*. 2013;2(11):e8. doi:10.1038/cti.2013.11
150. Ferreira AL, Choi J, Ryou J, et al. Gut microbiome composition may be an indicator of preclinical Alzheimer's disease. *Sci Transl Med*. 2023;15(700):eabo2984. doi:10.1126/scitranslmed.abo2984
151. Liu P, Wu L, Peng G, et al. Altered microbiomes distinguish Alzheimer's disease from amnesic mild cognitive impairment and health in a Chinese cohort. *Brain Behav Immun*. 2019;80:633–643. doi:10.1016/j.bbi.2019.05.008
152. Pan Q, Li YQ, Guo K, et al. Elderly Patients with Mild Cognitive Impairment Exhibit Altered Gut Microbiota Profiles. *J Immunol Res*. 2021;2021:5578958. doi:10.1155/2021/5578958
153. Jemimah S, Chabib CMM, Hadjileontiadis L, AlShehhi A. Gut microbiome dysbiosis in Alzheimer's disease and mild cognitive impairment: a systematic review and meta-analysis. *PLoS One*. 2023;18(5):e0285346. doi:10.1371/journal.pone.0285346
154. Chandra S, Sisodia SS, Vassar RJ. The gut microbiome in Alzheimer's disease: what we know and what remains to be explored. *Molecular Neurodegeneration*. 2023;18(1):9. doi:10.1186/s13024-023-00595-7
155. Zhao H, Zhou X, Song Y, et al. Multi-omics analyses identify gut microbiota-fecal metabolites-brain-cognition pathways in the Alzheimer's disease continuum. *Alzheimers Res Ther*. 2025;17(1):36. doi:10.1186/s13195-025-01683-0
156. van Niel G, D'Angelo G, Raposo G. Shedding light on the cell biology of extracellular vesicles. *Nat Rev Mol Cell Biol*. 2018;19(4):213–228. doi:10.1038/nrm.2017.125
157. Soares Martins T, Pelech S, Ferreira M, et al. Phosphoproteome Microarray Analysis of Extracellular Particles as a Tool to Explore Novel Biomarker Candidates for Alzheimer's Disease. *Int J Mol Sci*. 2024;25(3):1584. doi:10.3390/ijms25031584
158. Muraoka S, DeLeo AM, Sethi MK, et al. Proteomic and biological profiling of extracellular vesicles from Alzheimer's disease human brain tissues. *Alzheimers Dement*. 2020;16(6):896–907. doi:10.1002/alz.12089
159. Singh G, Mehra A, Arora S, et al. Exosome-mediated delivery and regulation in neurological disease progression. *Int J Biol Macromol*. 2024;264(Pt 2):130728. doi:10.1016/j.ijbiomac.2024.130728
160. Wang Y, Yuan P, Ding L, et al. Circulating extracellular vesicle-containing microRNAs reveal potential pathogenesis of Alzheimer's disease. *Front Cell Neurosci*. 2022;16:955511. doi:10.3389/fncel.2022.955511
161. Chai YL, Strohm L, Zhu Y, et al. Extracellular Vesicle-Enriched miRNA-Biomarkers Show Improved Utility for Detecting Alzheimer's Disease Dementia and Medial Temporal Atrophy. *J Alzheimers Dis*. 2024;99(4):1317–1331. doi:10.3233/JAD-230572
162. Su H, Rustam YH, Masters CL, et al. Characterization of brain-derived extracellular vesicle lipids in Alzheimer's disease. *J Extracell Vesicles*. 2021;10(7):e12089. doi:10.1002/jev2.12089
163. Pei J, Palanisamy CP, Jayaraman S, et al. Proteomics profiling of extracellular vesicle for identification of potential biomarkers in Alzheimer's disease: a comprehensive review. *Ageing Res Rev*. 2024;99:102359. doi:10.1016/j.arr.2024.102359
164. Lee S, Mankhong S, Kang JH. Extracellular Vesicle as a Source of Alzheimer's Biomarkers: opportunities and Challenges. *Int J Mol Sci*. 2019;20(7):1728. doi:10.3390/ijms20071728
165. Illakiya T, Karthik R. Automatic Detection of Alzheimer's Disease using Deep Learning Models and Neuro-Imaging: current Trends and Future Perspectives. *Neuroinformatics*. 2023;21(2):339–364. doi:10.1007/s12021-023-09625-7
166. Zhao Z, Chuah JH, Lai KW, et al. Conventional machine learning and deep learning in Alzheimer's disease diagnosis using neuroimaging: a review. *Front Comput Neurosci*. 2023;17:1038636. doi:10.3389/fncom.2023.1038636
167. Arya AD, Verma SS, Chakarabarti P, et al. A systematic review on machine learning and deep learning techniques in the effective diagnosis of Alzheimer's disease. *Brain Inform*. 2023;10(1):17. doi:10.1186/s40708-023-00195-7
168. Borchert RJ, Azevedo T, Badhwar A, et al. Artificial intelligence for diagnostic and prognostic neuroimaging in dementia: a systematic review. *Alzheimers Dement*. 2023;19(12):5885–5904. doi:10.1002/alz.13412
169. Fabrizio C, Termine A, Caltagirone C, Sancesario G. Artificial Intelligence for Alzheimer's Disease: promise or Challenge? *Diagnostics*. 2021;11(8):1473. doi:10.3390/diagnostics11081473
170. Monsour R, Dutta M, Mohamed AZ, Borkowski A, Viswanadhan NA. Neuroimaging in the Era of Artificial Intelligence: current Applications. *Fed Pract*. 2022;39(Suppl 1):S14–s20. doi:10.12788/fp.0231
171. Kale M, Wankhede N, Pawar R, et al. AI-driven innovations in Alzheimer's disease: integrating early diagnosis, personalized treatment, and prognostic modelling. *Ageing Res Rev*. 2024;101:102497. doi:10.1016/j.arr.2024.102497

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