



# Exploring the Interaction Between Sjögren's Syndrome and Osteoporosis: Pathophysiological Mechanisms and Management Strategies

Zhenghao Shi <sup>1,2,\*</sup>, Jiaxu Chen <sup>1,2,\*</sup>, Luan Xue <sup>1,2</sup>

<sup>1</sup>Department of Rheumatology and Immunology, Yueyang Hospital of Integrated Traditional Chinese and Western Medicine, Shanghai, People's Republic of China; <sup>2</sup>Shanghai University of Traditional Chinese Medicine, Shanghai, People's Republic of China

\*These authors contributed equally to this work

Correspondence: Luan Xue, Email xelco@163.com

**Abstract:** Sjögren's syndrome (SS) is a systemic autoimmune disorder predominantly affecting middle-aged and elderly women, characterized by lymphocytic infiltration of exocrine glands—particularly the salivary and lacrimal glands—resulting in xerophthalmia and xerostomia. Osteoporosis (OP), sharing similar age of onset and gender predilection, is a skeletal disorder defined by reduced bone mineral density and heightened fracture risk. The comorbidity of SS and OP represents a prevalent clinical phenomenon, with studies reporting an incidence rate of 33.1% to 51.6% among SS patients, significantly higher than that observed in healthy elderly populations. Vitamin D metabolism, widespread abnormal immune responses, hormonal imbalances, metabolic acidosis, and the RANKL/RANK/OPG axis exert significant contributory roles in the pathogenesis of both conditions. Treatment regimens for SS and OP may present certain overlaps yet potential contradictions. In this narrative review, we summarize the bidirectional relationship between these two diseases, thoroughly discuss the existing challenges in their management, and emphasize that recommending a comprehensive management strategy for SS patients with concurrent OP is crucial for enhancing patients' quality of life.

**Keywords:** Sjögren's syndrome, osteoporosis, immune system, pathophysiological mechanisms

## Introduction

Sjögren's Syndrome (SS) is a complex systemic autoimmune disorder that most often affects middle-aged and elderly women.<sup>1,2</sup> The clinical manifestations of SS range from localized symptoms of xerophthalmia and xerostomia to systemic involvement of multiple organ systems, including the respiratory, musculoskeletal, hematological, renal, and neurological systems.<sup>3,4</sup> Furthermore, patients experience significant subjective burden, which may lead to psychiatric conditions such as anxiety, depression, and insomnia, severely impacting quality of life.<sup>5</sup> The complex clinical presentation results in high disease heterogeneity, posing substantial challenges for disease management.<sup>3</sup>

Osteoporosis (OP) is characterized by reduced bone mineral density (BMD) and structural deterioration, leading to fragile bones susceptible to fractures.<sup>6–8</sup> Similar to SS, OP shows a predilection for middle-aged and postmenopausal women.<sup>9–11</sup> The incidence of OP is significantly higher in patients with SS, with multiple studies confirming that individuals with SS are particularly susceptible to developing OP. A study of 128 Chinese patients with primary SS found that 32.8% had osteopenia and 51.6% met the diagnostic criteria for OP.<sup>12</sup> In another study involving 118 SS patients, the prevalence of OP was 33.1%.<sup>13</sup> Contributing factors include genetic determinants, hormonal influences, inadequate dietary intake of calcium and vitamin D (VD), physical inactivity, and prolonged use of certain medications, all of which are closely associated with the comorbidity of these two conditions.<sup>14</sup>

Moreover, the comorbidity of SS and OP undoubtedly increases disease burden, enhances fracture risk, and creates medication contradictions that further complicate disease management.<sup>15</sup> This review elucidates the key comorbidity mechanisms between SS and OP, examines current pharmacotherapeutic approaches, and provides a basis for clinical

medication selection and new drug development. It also calls for more targeted future research to establish better disease prevention, risk stratification, and management strategies for comorbid patients.

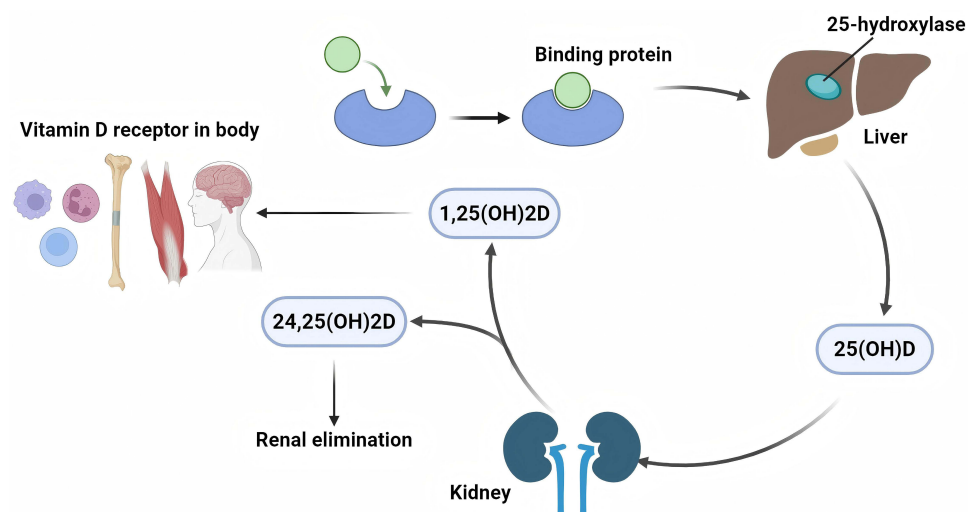
## SS and OP Share Similar Pathophysiological Mechanisms

### VD and VD Metabolites

VD, best known for its role in skeletal health, has emerged as a key regulator of immune.<sup>16</sup> The metabolic process of VD in the endocrine system is complex and extensive, involving numerous enzymes, metabolites, the vitamin D receptor (VDR), VD binding protein (DBP), and other regulatory hormones.<sup>17</sup> Upon absorption, VD binds with DBP and is transported to the liver, where it undergoes conversion by 25-hydroxylase to form 25-hydroxyvitamin D (25(OH)D). This compound then binds to DBP and is transported to various tissues: primarily the kidneys, where it is filtered through the glomerulus; subsequently, it is captured by transmembrane proteins and absorbed into tubular epithelial cells, during which hydroxylation occurs to produce 1,25-dihydroxyvitamin D (1,25(OH)2D), the active form of VD. The final hydroxylation step in the kidney, acting on both 25(OH)D and 1,25(OH)2D, results in the formation of 24,25-dihydroxyvitamin D (24,25(OH)2D), an inactive metabolite that is readily eliminated from the body<sup>18–20</sup> (Figure 1).

Fundamentally, VD facilitates increased intestinal absorption of calcium and phosphorus, playing a vital role in maintaining normal serum calcium and phosphate levels, which is essential for satisfactory bone mineralization.<sup>21,22</sup> Meanwhile, multiple tissues, including bone and immune cells, express VDR that respond to 1,25(OH)2D, which is crucial for normal bone metabolism and immune system function.<sup>23</sup>

Bone remodeling is a dynamic equilibrium process jointly constituted by osteoclast-mediated bone resorption and osteoblast-mediated bone formation.<sup>24</sup> Osteoblasts—derived from mesenchymal stem cells—accomplish new bone formation by synthesizing matrix proteins (including type I collagen) and regulating hydroxyapatite deposition.<sup>25</sup> In contrast, as specialized multinucleated giant cells, osteoclasts degrade mineralized bone matrix through the secretion of acidic substances and proteolytic enzymes.<sup>26</sup> VD plays an essential role in bone formation and remodeling processes by regulating the functions of osteoblasts and osteoclasts.<sup>27</sup> Studies utilizing VDR knockout mice have demonstrated that osteoblasts from these animals exhibit enhanced expression of alkaline phosphatase, bone sialoprotein, and osteocalcin compared to wild-type counterparts, accompanied by accelerated mineralization capacity both in vitro and in vivo.<sup>28</sup> Furthermore, additional studies highlight the precise phase-specific regulatory mechanisms of VD in osteogenesis, which exhibit distinct temporal-dependent characteristics. Specifically, early addition of 1,25(OH)2D to osteoblast cultures



**Figure 1** The overview of VD Metabolites. VD binds to vitamin D binding protein and is transported to the liver, where it is converted into 25(OH)D. After binding to DBP, 25(OH)D is absorbed into the renal tubular epithelial cells, where it undergoes hydroxylation to form 1,25(OH)2D. 1,25(OH)2D can bind to VD receptors in the body and exert various physiological effects.

suppresses the expression of collagen type 1 and alkaline phosphatase, whereas its administration during later differentiation stages stimulates their expression, thereby exerting biphasic regulatory effects on osteoblast function.<sup>29</sup>

Furthermore, under VD regulation, osteoblasts also participate in bone resorption processes. Numerous studies have demonstrated that under induction by 1,25(OH)<sub>2</sub>D, osteoblasts produce membrane-associated RANKL, which activates RANK on osteoclasts and their precursors to stimulate both differentiation and activation of osteoclast precursors.<sup>30</sup> In contrast, osteoblasts from VDR-knockout mice fail to support 1,25(OH)<sub>2</sub>D-induced osteoclastogenesis, whereas when co-cultured with wild-type osteoblasts in the presence of 1,25(OH)<sub>2</sub>D, osteoclast precursors regain the ability to differentiate into osteoclasts.<sup>31</sup>

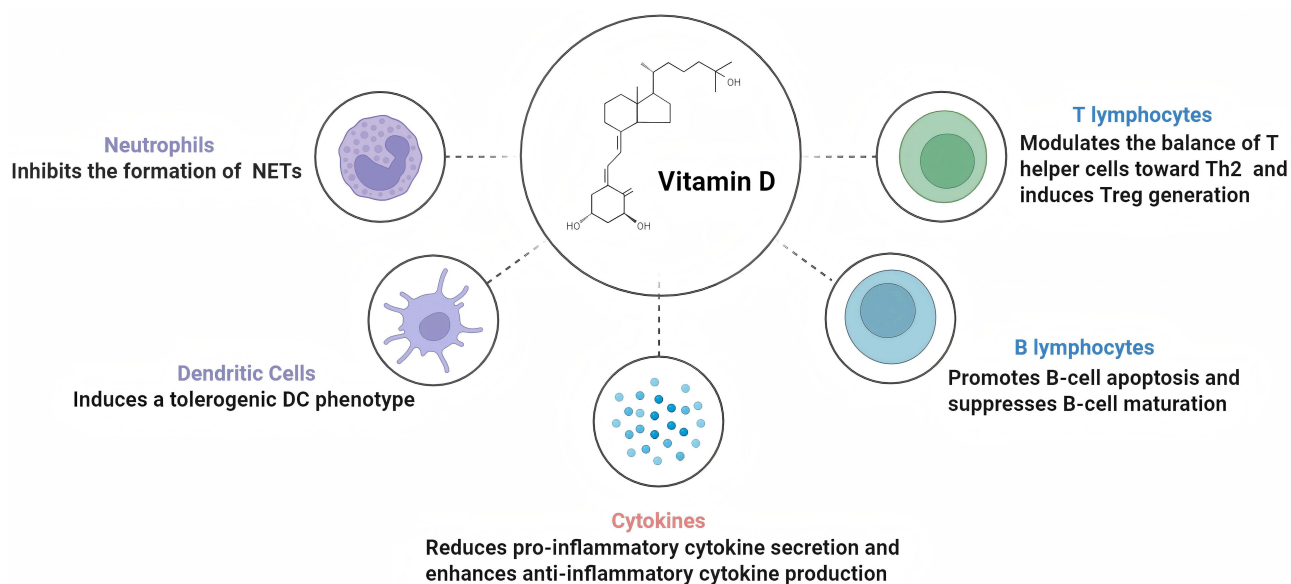
Although the exact pathogenesis of SS remains unclear, it has been widely confirmed to be associated with various immune cells that express VDR and are regulated by 1,25(OH)<sub>2</sub>D. Reduced levels of 1,25(OH)<sub>2</sub>D have been observed in the peripheral blood of SS patients, showing a certain correlation with clinical and immuno-inflammatory status.<sup>32,33</sup> Additionally, in murine models, 1,25(OH)<sub>2</sub>D or VDR agonists have been shown to reduce lymphocytic infiltration in salivary glands<sup>34</sup> (Figure 2).

For neutrophils, 1,25(OH)<sub>2</sub>D decreases neutrophil extracellular traps (NETs) formation and exhibits an antagonistic effect against the type I interferon signaling pathway in neutrophils of SS patients.<sup>35</sup>

Regarding dendritic cells (DCs), 1,25(OH)<sub>2</sub>D induces a shift toward a tolerogenic phenotype capable of producing anti-inflammatory factors, promoting regulatory T (Treg) cell differentiation, reducing antigen presentation, and thereby decreasing T cell activation.<sup>36,37</sup>

In SS patients, CD8<sup>+</sup> T cells, Treg cells, and plasma cells accumulate around acini within exocrine glands, whereas their proportions are reduced in peripheral blood. The degree of this infiltration positively correlates with disease activity.<sup>38–40</sup> Through modulating cytokine secretion by DCs, 1,25(OH)<sub>2</sub>D can shift the T helper (Th) cell balance from a Th1/Th17-dominant state toward a Th2 phenotype.<sup>41,42</sup> Conversely, observations indicate that decreased peripheral Th2 cell frequencies and an elevated Th1/Th2 ratio may constitute the immunological mechanism underlying interstitial lung disease development in SS patients, reflecting higher disease activity.<sup>43</sup>

Notably, 1,25(OH)<sub>2</sub>D induces the generation of Tregs by binding to the FoxP3 promoter region.<sup>44</sup> Animal models have demonstrated that VD analogs increase the number of FoxP3<sup>+</sup> Tregs in salivary glands and alleviate glandular inflammation.<sup>34</sup> Moreover, 1,25(OH)<sub>2</sub>D induces apoptosis of activated B lymphocytes and modulates the NF- $\kappa$ B pathway through regulation of CD40 expression, thereby suppressing the generation of plasma cells and class-switched memory B cells.<sup>45,46</sup>



**Figure 2** Overview of the effects of vitamin D on cytokines and immune cells, including neutrophils, dendritic cells, B lymphocytes, and T lymphocytes.

Regarding cytokine modulation, 1,25(OH)<sub>2</sub>D significantly reduces secretion of the pro-inflammatory cytokines IL-6 and IL-12 while enhancing production of the anti-inflammatory cytokine IL-10.<sup>47</sup>

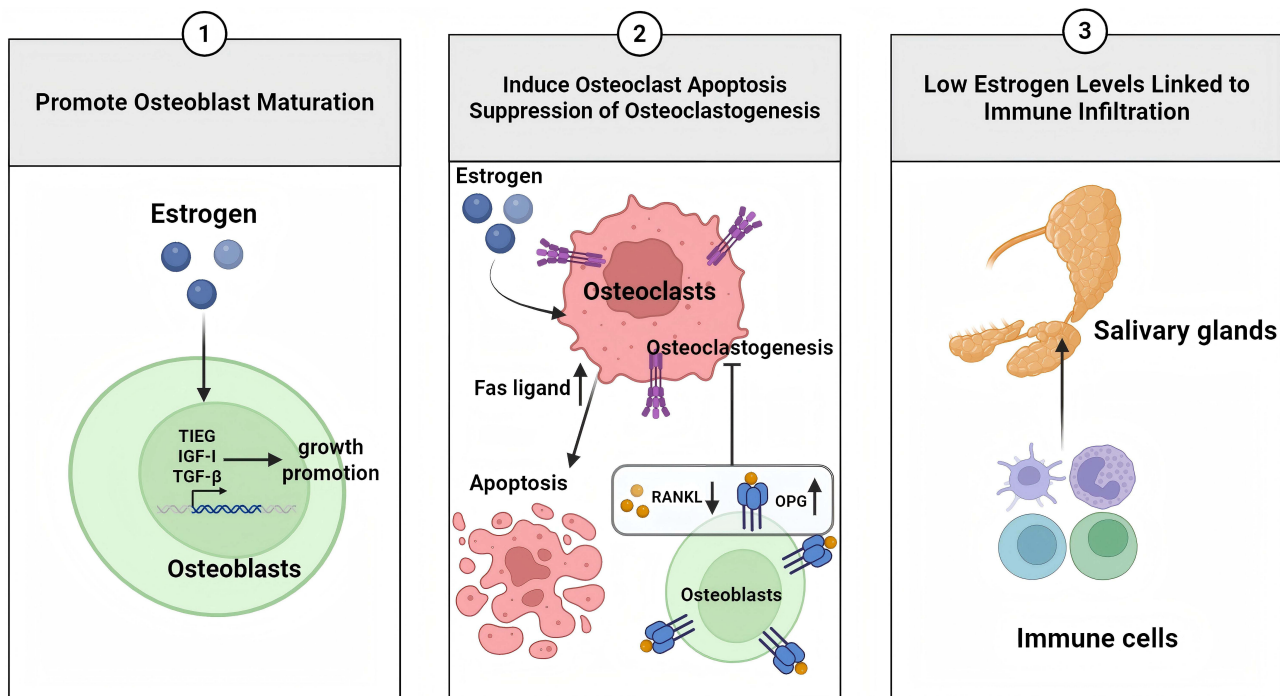
## Sex Hormones

Sex hormones are a class of hormones that regulate sexual characters, sexual development, and reproductive functions in organisms. Sex hormones mainly include estrogens, progesterones, and androgen.<sup>48</sup> In addition to some fundamental functions related to sexual differentiation and the development and maintenance of sexual traits, sex hormones also participate significantly in the regulation of various physiological processes including but not limited to immune function, bone health, and cardiovascular system.<sup>23,49,50</sup> It is intuitive, therefore, that sex hormones may influence both conditions since both SS and OP mainly affect middle-aged and elderly women.

### Estrogen

Estrogen influences both osteoblasts and osteoclasts and is important for multiple processes maintaining bone metabolic homeostasis (Figure 3). It induces osteoblasts to upregulate the expression of genes such as TIEG, IGF-I, and TGF- $\beta$ , confirmed to directly promote osteoblast growth.<sup>51–57</sup> Estrogen stimulates Fas ligand expression in osteoclasts, which in turn exerts pro-apoptotic effects on osteoclast activity. Furthermore, estrogen decreases RANKL expression and increases OPG expression, thereby inhibiting osteoclast differentiation.<sup>58–60</sup> Additionally, estrogen participates in complex bone metabolic balance by promoting the expression of 1,25(OH)<sub>2</sub>D receptors, modulating osteoblast responses to parathyroid hormone (PTH), and promoting IGFBP-4 expression while reducing its proteolytic degradation.<sup>61</sup> PTH stimulates bone resorption, renal tubular calcium reabsorption, and the production of 1,25(OH)<sub>2</sub>D.<sup>62</sup> IGFBP-4 binds to IGF, thereby reducing its availability to IGF receptors and partially inhibiting IGF's biological effects on osteoblasts, including cell proliferation, differentiation, and bone formation and remodeling.<sup>63</sup>

Clinically, multiple studies have found decreased estrogen levels and an abnormal estrogen-progesterone ratio in patients with SS.<sup>64–66</sup> Elevated levels of Bcl-2, Fas, and FasL in the salivary gland tissues of SS patients correlate with



**Figure 3** The Role of Estrogen in Bone Metabolism and Sjögren's Syndrome. This figure depicts three vertically arranged sections illustrating the multifaceted effects of estrogen: 1. Estrogen exerts growth-promoting effects on osteoblasts by upregulating the expression of TIEG, IGF-I, and TGF- $\beta$ . 2. Estrogen induces osteoclast apoptosis through increased FAS ligand expression. Concurrently, it reduces RANKL secretion and enhances osteoblastic OPG production, which competitively binds RANKL to further inhibit osteoclast maturation. 3. Decreased estrogen levels are associated with enhanced immune cell infiltration in the exocrine glands of SS patients.

inflammatory severity and are significantly higher than in healthy controls.<sup>67</sup> Animal studies demonstrate that ovariectomy induces selective apoptosis of submandibular salivary gland acinar cells in mice, which subsequently develop focal lymphadenitis and SS-like disease with positive autoantibodies.<sup>68</sup> In ovariectomized rabbit and mouse models, estrogen supplementation prevents lacrimal cell death and lymphocytic infiltration.<sup>69,70</sup> These findings suggest acinar cell atrophy and apoptosis in SS may relate to estrogen deficiency, though specific mechanisms require further investigation.

### Progesterone

Progesterone, also known as P4, is a 21-carbon steroid hormone. Progesterone provides multiple protections for the developing fetus and plays a key role in neuroglial cells, synapses, dendritic activity, neural development, and tissue repair. Additionally, progesterone also acts on a wide range of tissues, including the bones, heart, and brain.<sup>71–73</sup> Studies have found that the bone formation marker BAP increases during the luteal phase of the ovulatory cycle when progesterone levels are high, but does not increase during an anovulatory cycle. The bone resorption marker pyridinoline decreases during the luteal phase of the ovulatory cycle when progesterone levels are high, with a smaller decrease during an anovulatory cycle.<sup>74</sup> This suggests that elevated progesterone could be linked to an increase in bone formation while resulting in a slight reduction of bone resorption. Another research has indicated that elevated levels of progesterone can effectively suppress osteoblasts differentiation by as much as 80%.<sup>75</sup> Considering these factors, an appropriate amount of progesterone could be beneficial in managing OP.<sup>76,77</sup>

There is a scarcity of research exploring the influence of progesterone on SS, indicating a need for additional studies in this area.

### Androgen

Testosterone is the primary androgen circulating in men, binding to both albumin and sex hormone-binding globulin in the bloodstream. In peripheral tissues, testosterone is converted into 5 $\alpha$ -dihydrotestosterone by the enzyme 5 $\alpha$ -reductase. Both testosterone and 5 $\alpha$ -dihydrotestosterone are capable of activating the androgen receptor (AR).<sup>78</sup> Additionally, testosterone can be converted into estradiol (E2) by the enzyme aromatase, which is also expressed locally in bone tissue. The AR is expressed in both osteoblasts and osteocytes.<sup>79</sup> Studies have shown that osteoblasts from cortical bone have a stronger AR binding capacity compared to those from trabecular bone, although there is no gender difference in AR expression.<sup>80,81</sup> This suggests that both testosterone and E2 may play important roles in maintaining skeletal health in both sexes. Testosterone directly inhibits osteoclast differentiation and bone resorption, while the inhibitory effect of E2 on osteoclasts is mainly mediated indirectly through osteoblasts.<sup>82</sup> The effects of androgens on bone cells are regulated by three main factors: TGF- $\beta$ , IGFs, and IL-6. Androgens may promote bone formation by activating TGF- $\beta$  and IGFs or reduce osteoclastogenesis by inhibiting IL-6.<sup>83–85</sup> Additionally, androgens may directly affect osteoclasts or indirectly influence osteoclast formation by affecting bone marrow stromal cells, thereby regulating osteoclastogenesis induced by RANKL.<sup>86</sup>

There is currently no clear conclusion regarding the relationship between androgens and SS. Some studies have found that patients with SS have lower serum levels of dehydroepiandrosterone, 5 $\alpha$ -dihydrotestosterone, and dehydroepiandrosterone sulfate. As age increases and the aging process begins, androgen levels naturally decline. By the age of 40, women's testosterone levels gradually decrease to half of what they were at 20 years old. Given the age of onset in SS patients, the decline in androgen levels may be closely related to the onset of SS.<sup>66,87,88</sup> Androgens can regulate the function of the lacrimal glands and meibomian glands, modulate lipid production in these tissues, and promote the formation of the lipid layer of the tear film. Androgen deficiency may lead to meibomian gland dysfunction and evaporative dry eye.<sup>89,90</sup> Some studies have also found defects in androgen deficiency and the processing of dehydroepiandrosterone secretion in the salivary glands of SS patients.<sup>91</sup> Therefore, the dry mouth and dry eyes in SS patients may be related to androgens, but further research is needed to verify this.

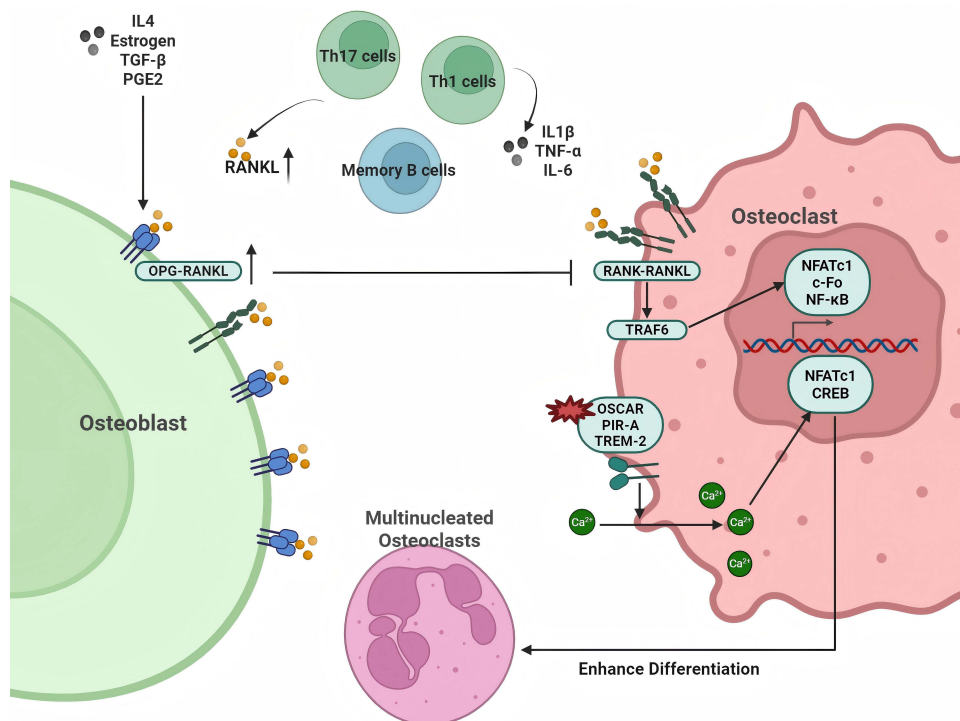
In summary, sex hormones, especially estrogens and androgens, play an essential role in regulating bone health, glandular function, and various physiological processes. Estrogen influences bone metabolism by regulating osteoblasts and osteoclasts, while also affecting the health of glands such as the lacrimal and salivary glands. Androgens, meanwhile, help maintain BMD and strength by balancing bone resorption and formation. They also play a critical role in the

function of the lacrimal and meibomian glands, which affect tear production and eye health. As a result, any abnormalities in sex hormone levels, particularly a deficiency or imbalance of estrogens and androgens, could significantly contribute to the co-occurrence of SS and OP. These hormonal changes not only directly impact bone and glandular function but may also indirectly trigger the development and progression of these conditions through immune system mechanisms. Gaining a deeper understanding of how sex hormones affect the comorbidity of SS and OP may offer new avenues for the treatment and management of these diseases.

## RANKL/RANK/OPG Axis

The RANKL-RANK-OPG axis plays a pivotal role in mediating communication between osteoblasts and osteoclasts.<sup>92,93</sup> Upon RANK activation, signaling adaptor proteins such as TRAF6 are recruited and activated, triggering a kinase cascade that leads to nuclear translocation and activation of transcription factors including NFATc1, c-Fos, and NF- $\kappa$ B—key regulators of osteoclast-specific gene expression. Co-stimulatory signals mediated by ITAM-containing proteins (DAP12 and FcR $\gamma$ ), which bind to receptors such as OSCAR, PIR-A, or TREM-2, activate the Syk-PLC $\gamma$  pathway and induce calcium influx. This calcium influx subsequently activates calcium-dependent phosphatase signaling, enhancing NFATc1 and CREB activity to promote osteoclast-specific gene expression. This process initiates osteoclast differentiation, ultimately leading to the formation of mature multinucleated osteoclasts through cell fusion and incomplete cytokinesis.<sup>94–97</sup>

Osteoprotegerin (OPG), primarily expressed by osteoblasts, is regulated by estrogen, IL-4, TGF- $\beta$ , and PGE2.<sup>98</sup> By competitively binding to RANK with RANKL, OPG inhibits osteoclast formation and suppresses bone resorption activity<sup>96</sup> (Figure 4).



**Figure 4** The communication between osteoblasts and osteoclasts through RANKL/RANK/OPG axis. Upon RANK activation, adaptor proteins such as TRAF6 are recruited and activated, leading to the activation of kinases and subsequent nuclear translocation and activation of transcription factors including NFATc1, c-Fos, and NF- $\kappa$ B, which are essential for regulating osteoclast-specific gene expression. Co-stimulatory signals mediated by ITAM-containing proteins DAP12 and FcR $\gamma$ , through binding to receptors such as OSCAR, PIR-A, or TREM-2, further trigger the Syk-PLC $\gamma$  pathway and calcium influx. The resulting calcium influx activates calcium-dependent phosphatase signaling, enhancing NFATc1 and CREB activity to promote osteoclast-specific gene expression and initiate osteoclast differentiation. Subsequently, cells undergo fusion and incomplete division to form mature multinucleated osteoclasts. OPG competes with RANKL for binding to RANK, thereby inhibiting osteoclastogenesis and reducing bone resorption.

In SS, activated Th1 and Th17 cells lead to elevated levels of IL-1 $\beta$ , TNF- $\alpha$ , and IL-6, which promote RANKL expression.<sup>99–103</sup> Furthermore, activated Th17 cells and memory B cells further produce RANKL. These cytokines collectively suppress OPG expression, ultimately disrupting bone homeostasis.<sup>104,105</sup> However, direct evidence confirming this mechanism in SS remains limited.

Relevant studies have been conducted in systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA). Among SLE patients, the frequency of the OPG 245 G allele is significantly higher in those with low BMD than in those with normal BMD (31.4% vs 18.1%,  $p = 0.04$ ),<sup>106</sup> indicating that OPG 245 G may be a genetic factor underlying OP in autoimmune diseases. In another study that collected blood and synovial fluid from 167 RA patients and 25 healthy volunteers, elevated levels of inflammatory cytokines were observed in RA. Exposure to IL-21 or IL-23 down-regulated the production of cytokines, TNF- $\alpha$ , IFN- $\gamma$ , IL-17, and RANKL in these cells. This further suggests that increased TNF- $\alpha$  and IL-6 levels may underlie the elevated RANKL observed in SS, and that the IL-21/IL-23 axis plays a pivotal role in modulating both inflammation and RANKL expression.<sup>107</sup>

## Renal Tubular Acidosis (RTA)

RTA refers to a collection of transport abnormalities marked by a diminished reabsorption of bicarbonate (HCO $_3^-$ ) in the proximal tubule, decreased secretion of distal hydrogen ions (H $^+$ ), or a combination of both. This leads to impaired net acid excretion and ongoing hyperchloremic metabolic acidosis, all while the glomerular filtration rate stays within the normal range.<sup>108</sup> From a pathophysiological perspective, RTA is mainly categorized into three distinct types. Type 2, or proximal RTA, results from the impaired reabsorption of filtered bicarbonate in the proximal tubule. In contrast, type 1 (distal) RTA (dRTA) is caused by a deficiency in distal H $^+$  secretion, while type 4 RTA is characterized by decreased secretion of both H $^+$  and K $^+$  in the collecting ducts, often due to lower aldosterone secretion or response. Most cases observed in children are primary and stem from inherited factors. Types 1 and 4 RTA might also arise secondary to other underlying issues, especially when symptoms emerge at a later stage. Type III or combined RTA is a type of renal tubular dysfunction that exhibits features of both proximal RTA and dRTA.<sup>109</sup>

The prevalence of dRTA in patients with SS varies significantly across different studies, ranging from 6.8% to 70%.<sup>110–113</sup> SS may be caused by tubulointerstitial infiltration of T cells, B cells, and plasma cells, or, less commonly, by autoantibodies. The majority of patients exhibit clinical symptoms as a direct consequence of lymphocyte infiltration in the interstitium, which promotes interstitial fibrosis, leading to chronic kidney disease. dRTA is the most common electrolyte disturbance in SS, involving the connecting tubules and collecting ducts.<sup>114</sup> dRTA is associated with growth retardation, skeletal pain, abnormal bone metabolism, and pathological fractures, which mainly originates from the physicochemical dissolution of minerals caused by acidosis, leading to changes in bone cell function. Studies have found significant suppression of both bone formation and bone resorption in dRTA patients, along with a mild increase in osteoid volume.<sup>115</sup> Additionally, most dRTA patients have a significant reduction in BMD in the spine and hips.<sup>116</sup>

## The Medications Used to Treat SS May Lead to OP GCs

GCs are widely used to suppress inflammation and immune system responses due to their potent anti-inflammatory, immunosuppressive, and metabolic regulatory functions. They exert therapeutic effects by binding to GCs receptors in the body, thereby regulating a series of biological processes. GCs can significantly inhibit the overactivity of the immune system, which is why they are extensively used in diseases caused by immune system dysfunction, such as SS.<sup>117</sup>

Glucocorticoid-induced osteoporosis (GIO) is the most common secondary cause of OP and the most common iatrogenic cause. Bone loss caused by GIO can be roughly divided into two stages. The initial stage, within the first year of treatment, sees a rapid loss of BMD at a rate of about 6–12%. In the long-term stage, the rate of BMD loss slows down, with an annual loss of about 3% thereafter.<sup>118,119</sup> In a study, the risk of vertebral fractures in individuals taking oral GCs was nearly three times higher than that of the general population, while the risk of hip fractures doubled.<sup>120</sup> Additionally, the incidence of fractures was influenced by the duration of use (cumulative dose) and the current dose.<sup>121</sup>

The pathophysiology of GIO primarily involves a long-term reduction in bone formation, with an increase in bone resorption during the early phase (the first year after starting treatment). GCs inhibit the formation of osteoblasts, induce apoptosis in both osteoblasts and osteocytes, and prolong the lifespan of osteoclasts.<sup>119</sup>

GCs exert multiple direct effects on osteoblast signaling pathways. They enhance the expression of PPAR $\gamma$ 2, KLF15, C/EBP $\alpha$ , and aP2, which leads to the preferential differentiation of multipotent progenitor cells into adipocytes rather than osteoblasts, thus reducing the number of osteoblasts.<sup>122</sup> In the WNT- $\beta$ -catenin signaling pathway, GCs increase the expression of sclerostin and other inhibitory factors, while suppressing the expression of WNT16 in a dose- and time-dependent manner. This results in a decrease in osteoblastogenesis and bone loss.<sup>123</sup> At low or physiological doses, GCs appear to induce autophagy and support the activity and function of osteoblasts. However, at doses higher than physiological levels, this process is inhibited, leading to a loss of osteoblast-related gene expression and increased apoptosis.<sup>124</sup> Additionally, GCs affect the RANK-RANKL-OPG axis by increasing RANKL production and decreasing the transcription of OPG mRNA in osteoblasts and osteocytes. GCs influence osteoblasts to elevate the ratio of RANKL to OPG, which enhances bone resorption and promotes the differentiation and maturation of osteoclasts.<sup>125</sup> However, the long-term impact of GCs on osteoclast function is still not fully understood.

GCs affect bone metabolism through various indirect pathways. They suppress the transcription of the IGF1 gene, thereby reducing bone formation, whereas IGF1 typically promotes bone formation by stimulating type I collagen synthesis, inhibiting collagen degradation, and preventing osteoblast apoptosis.<sup>126</sup> Additionally, GCs disrupt calcium homeostasis by decreasing calcium absorption in the intestine and inhibiting calcium reabsorption in the renal tubules, which can lead to hypocalcemia and secondary hyperparathyroidism.<sup>127</sup> GCs may also enhance the sensitivity of bone cells to PTH by increasing the number and affinity of PTH receptors. They inhibit growth hormone secretion, reducing its anabolic effects on bone, and suppress gonadotropin secretion, leading to decreased estrogen and testosterone levels, which further promotes bone resorption.<sup>125</sup> These combined effects contribute to bone loss caused by GCs.

In summary, although GCs are effective in treating inflammatory and immune-related conditions, prolonged use can result in considerable bone loss and a higher risk of fractures. This is due to the intricate combination of direct and indirect effects on bone metabolism. The main factors contributing to GIO include the inhibition of osteoblast function, increased bone resorption, and disturbances in calcium balance. A thorough understanding of these mechanisms is essential for creating strategies to reduce bone loss and safeguard bone health in individuals undergoing GCs treatment.

## Disease-Modifying Antirheumatic Drugs (DMARDs)

### Methotrexate (MTX)

MTX is one of an often-prescribed DMARDs that is extensively used for the management of a variety of conditions such as cancers and disorders of the immune system. It operates by targeting dihydrofolate reductase, which leads to a decrease in folate production. Folate plays an essential role in the process of DNA and RNA synthesis in cells, and the reduction of its production significantly hinders the rate of cell division.<sup>128</sup>

Currently, the impact of MTX on BMD among patients with SS is still not well-established, as there exists a scarcity of research and ongoing discussions surrounding the issue. The administration of MTX may result in a reduction of osteoblasts, osteocytes, and chondrocytes within the growth plate, concurrently elevating the quantity and activity of osteoclasts.<sup>129</sup> Between 2011 and 2019, there were reports of five instances in which individuals exhibited reduced BMD or sustained fractures following the administration of MTX. Nonetheless, the sites of the fractures were uncommon for those typically seen with OP, and all individuals showed signs of recovery within a few months following the cessation of MTX.<sup>130</sup> Moreover, research indicates that there was an increase in BMD among patients with RA following treatment with MTX. Notably, findings in this area appear to be inconsistent, possibly due to the diverse mechanisms underlying autoimmune disorders and the differing intensities of disease activity.<sup>131-133</sup> The enhancement of BMD observed in individuals with RA might be attributed to the attenuation of the inflammatory response through the use of MTX.

### Cyclophosphamide (CTX)

CTX is categorized as an alkylating compound. Its mechanism involves the introduction of alkyl groups into the DNA structure, which leads to breaks or cross-links within the DNA strands; this interference disrupts the processes of DNA

replication and repair, ultimately resulting in cellular apoptosis.<sup>134</sup> Due to the rapid division rate of cancerous cells, the agent is notably effective in suppressing their growth.<sup>135</sup> Simultaneously, CTX exerts a degree of suppression on immune system cells, which is why it is frequently utilized in managing diseases related to the immune system.<sup>136,137</sup> The occurrence of OP due to CTX is significantly connected to the decline in mesenchymal stem cells within the bone marrow and the inhibition of bone development. Alterations in the microenvironment resulting from CTX might enhance the persistence of osteoclasts, thus accelerating the processes of bone metastasis and bone degradation. Simultaneously, CTX has the capacity to impede the formation of osteoclasts through the inhibition of the RANKL signaling pathway, which contributes to the deceleration of bone resorption. The influence of CTX on bone metabolism encompasses both the suppression of osteoblast formation and the reduction of osteoclast development. These influences can result from the decrease in osteoclast function, the downregulation of bone degradation indicators, and the modulation of osteoblast formation processes, thereby contributing to the maintenance of bone integrity.<sup>138</sup> Research indicated that CTX may lead to OP in male C57BL/6 mice, likely due to its impact on hindering the formation of osteoblasts by diminishing both the quantity and differentiation of bone mesenchymal stem cells (MSCs), as well as reducing the activity and generation of osteoblasts.<sup>139</sup>

### Calcineurin Inhibitors (CNI)

CNI are a class of drugs that regulate immune responses by inhibiting the activity of calcineurin. Calcineurin is a calcium-dependent enzyme that typically plays a role when T cells are stimulated. Calcineurin dephosphorylates the transcription factor NFAT (Nuclear Factor of Activated T-cells), allowing it to enter the nucleus and activate immune responses. Therefore, inhibiting the activity of calcineurin can prevent T cell activation and reduce immune responses.<sup>140</sup> The main calcineurin inhibitors used in clinical practice are Cyclosporine (CsA) and Tacrolimus (FK506). CsA binds to cyclophilin A, inhibiting the function of calcineurin. FK506, similar to CsA, has a more specific mechanism. It binds to the FKBP12 protein, and the resulting complex inhibits the activity of calcineurin.<sup>141</sup>

Studies have found that both CsA and FK506 significantly reduce the bone strength of the femoral shaft and the BMD of the tibia and femur in rats. Histological analysis of bone tissue shows that administration of both drugs leads to a decrease in bone volume, trabecular number and thickness, as well as an increase in trabecular separation. CsA increases both bone formation and bone resorption, leading to high-turnover bone loss, while FK506 increases bone resorption without affecting bone formation, resulting in bone loss.<sup>142</sup> Another study found that both CsA and FK506 reduced BMD in liver transplant patients, with FK506 treatment having more favorable long-term effects on bone metabolism compared to CsA treatment.<sup>142</sup>

### Azathioprine (AZA)

AZA is converted into 6-mercaptopurine (6-MP) in the body. 6-MP inhibits the synthesis of purines, which in turn affects the synthesis of DNA and RNA, suppressing the proliferation of T cells and B cells, and slowing down the immune response.<sup>143</sup> A study found that MicroCT imaging of mice showed a significant adverse effect of azathioprine treatment on trabecular bone microstructure, with a significant reduction in bone volume/tissue volume and trabecular number.<sup>144</sup> Another study found that monotherapy with azathioprine did not alter the levels of ionized calcium, 1,25(OH)<sub>2</sub>D, or PTH, but a decrease in osteocalcin was observed, indicating impaired osteoblast activity.<sup>145</sup>

## Management of Patients with SS-OP Comorbidity

There exists a significant therapeutic paradox between immunosuppressive therapy for SS and OP management, which manifests in three key aspects: Firstly, while immunosuppressants like glucocorticoids can effectively control inflammatory activity in SS, they simultaneously accelerate bone loss by inhibiting osteoblast function and promoting osteoclast activation. Furthermore, DMARDs such as MTX and CNI exhibit differential effects on bone metabolism, with some agents potentially exacerbating bone deterioration. Secondly, OP treatments including bisphosphonates and anti-RANKL monoclonal antibodies, while inhibiting bone resorption, may interfere with immune cell function and potentially compromise immunoregulation. Lastly, a clinical dilemma exists in treatment timing - the acute phase of SS necessitates intensive immunosuppression, yet this period often coincides with when drug-induced skeletal adverse

effects are most difficult to counteract, while early initiation of bone-protective therapy may be challenging due to high disease activity.

Currently, there is a significant lack of clinical management guidelines for autoimmune diseases complicated with OP. This knowledge gap is particularly prominent in research on individual specific autoimmune diseases complicated with OP. To date, only OP associated with RA has been partially studied. Other autoimmune disease areas still lack targeted consensus on bone health management. This current situation urgently needs to be addressed through multicenter clinical trials and systematic reviews. Furthermore, the current treatment for SS remains largely empirical, with a notable absence of standardized clinical guidelines. Based on these current circumstances, we propose a distinctive management strategy for SS-OP comorbidity (Figure 5).

In SS, multiple factors including inflammatory activation, VD deficiency, hypoestrogenism, and acid-base metabolic abnormalities may exacerbate bone destruction. First and foremost, it is crucial to monitor BMD and assess bone metabolism regularly. This includes periodic measurements of serum calcium, phosphorus, 25(OH)D, and PTH to evaluate metabolic risk. And higher doses of calcium and VD supplementation than peers may be necessary, but there are few direct studies in the field of SS. For RA patients, women aged 51 and above and men aged 71 and above require an increased intake of 1200mg per day. For individuals aged 50 and older, a daily VD supplementation of 800–1000 IU is routinely recommended, and is beneficial for reducing RA disease activity.<sup>146,147</sup>

Additionally, monitoring bone turnover markers such as  $\beta$ -C-terminal telopeptide and procollagen type I N-terminal propeptide provides valuable insights into ongoing bone remodeling processes.<sup>148,149</sup> Given that immune system activation in SS may enhance bone resorption, comprehensive bone metabolism tests are vital for these patients as they facilitate the detection of early bone loss and enable timely intervention. Furthermore, for patients receiving long-term corticosteroid treatment or those with additional risk factors (such as advanced age, low body weight, previous fracture history, smoking, excessive alcohol consumption, or physical inactivity),<sup>8</sup> regular dual-energy X-ray absorptiometry examination can monitor OP progression, and adjusting treatment regimens accordingly.<sup>150</sup> More supplementation with calcium and VD is recommended to maintain adequate serum 25(OH)D levels in this population to slow down bone damage.

Comprehensive assessment of BMD and bone metabolism not only facilitates early and timely intervention but also helps gain a better understanding of the skeletal health status of patients with SS and OP, allowing for effective therapeutic selection. Selection of specific agents should be tailored to patient characteristics: bisphosphonates may be considered for patients with mild renal impairment, with caution regarding esophageal dysmotility; denosumab represents a viable option for those with significant renal dysfunction,<sup>151,152</sup> while monitoring for infection risk; and teriparatide can be utilized in patients with high fracture risk, with attention to treatment duration limitations. In all

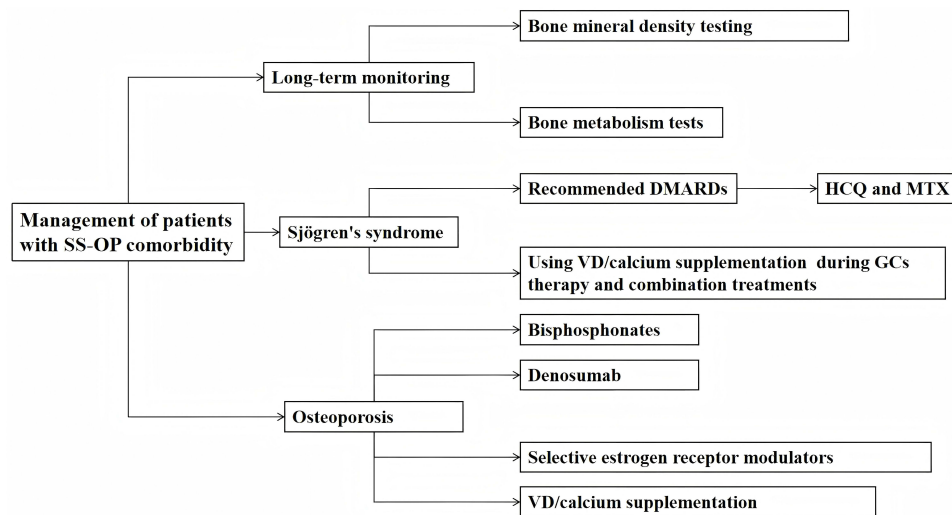


Figure 5 Overview of the management strategies for the comorbidity of Sjögren's Syndrome and Osteoporosis.

cases, careful consideration of potential SS-related comorbidities such as renal damage, esophageal motility disorders, and infection risk is essential for rational therapeutic selection and optimal patient outcomes.

In patients with comorbid SS and OP, the apply of DMARDs needs to be considered with caution, as some DMARDs may exacerbate OP or increase the risk of fractures. Recommended DMARDs include hydroxychloroquine (HCQ), and MTX. HCQ has been widely used in the treatment of SS to alleviate symptoms and control disease progression. Multiple studies have found that the impact of HCQ on bone metabolism may be relatively minor, and its anti-inflammatory effects may provide some protection for bone health.<sup>153</sup> MTX is a commonly used medication for treating immune system-related diseases, especially in conditions such as RA, psoriasis, and SLE, where it has shown significant efficacy. It works by suppressing the overactivity of the immune system to alleviate symptoms and control disease progression. In the treatment of SS, MTX also has certain applications, particularly in managing patients with notable arthritis, inflammation, or other systemic manifestations.<sup>154,155</sup> Although it has been mentioned earlier that MTX may have some effect on bone metabolism, the research is not conclusive, and the impact of MTX is relatively minor. In addition, some DMARDs are not recommended for SS patients with OP, such as CsA, AZA, FK506, etc., as these medications can have an adverse effect on bone metabolism, as mentioned earlier.

## Challenges and Perspectives

The current understanding of the SS-OP association remains limited due to several fundamental challenges: (1) The pathogenesis of SS, a complex systemic autoimmune disorder, remains incompletely understood; (2) Significant clinical heterogeneity among SS patients creates population standardization difficulties; (3) Osteoporosis's strong dependence on age and gender introduces unavoidable confounding effects in current studies.

Building upon current evidence, future research should prioritize two key translational directions for SS-OP comorbidity: first, the development of specific biomarkers focusing on shared inflammatory mediators that concurrently associate with glandular dysfunction and accelerated bone loss, along with predictive markers for rapid BMD decline; second, the design of targeted clinical trials investigating therapeutic strategies against common inflammatory pathways, with dual emphasis on both immunomodulation and osteoprotection to comprehensively address disease progression. Current bone health management guidelines for autoimmune diseases primarily focus on GIO,<sup>156</sup> while lacking comprehensive evaluation of other medications and individualized analysis across different autoimmune conditions. Future research should address these gaps to optimize patient management.

## Summary and Conclusion

OP poses a significant health challenge for individuals with SS, significantly increasing the risk of fractures, pain, and mobility limitations. These symptoms not only negatively impact physical health but also greatly interfere with daily life and overall quality of life. The two conditions are interrelated, with their interaction forming a vicious cycle, where each exacerbates the other's pathological state. For instance, individuals with SS often experience chronic inflammation and immune system abnormalities, leading to an imbalance in bone metabolism, which makes the bones more susceptible to damage and, in turn, triggers OP. Therefore, increased clinical focus on bone health in SS patients is warranted.

This review elucidates the interaction mechanisms between SS and OP. VD deficiency may worsen bone health problems and affect the progression of SS through immune system involvement. Hormones such as estrogen and androgens are vital for both bone and glandular health, and any imbalance in these hormones may serve as a link between SS and OP. Additionally, The RANKL/RANK/OPG signaling axis constitutes a pivotal interface coordinating bone metabolism and immune function. dRTA, the most prevalent electrolyte disorder in SS patients, may serve as a key pathogenic contributor to OP development. Furthermore, medications commonly used to treat SS may promote the development and progression of OP by inhibiting bone formation and accelerating bone resorption.

For patients with SS-OP comorbidity, we propose a rational management approach that includes: regular BMD monitoring and bone metabolism evaluation, prioritizing the use of DMARDs with minimal impact on bone metabolism, and combining bone-modulating agents with calcium/VD supplementation to synergistically control both autoimmune activity and bone loss progression.

These findings provide critical evidence for establishing integrated management protocols for bone health in SS patients, and offer fundamental insights for developing multidisciplinary approaches to autoimmune diseases with comorbid bone disorders.

## Acknowledgments

Zhenghao Shi and Jiaxu Chen are co-first authors for this study. The authors have no specific acknowledgements.

## Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

## Funding

This review was funded by National Natural Science Foundation of China, grant number 82374318.

## Disclosure

ZH.S., JX.C. and L.X. declare that they have no conflicts of interest in this work.

## References

1. Brito-Zerón P, Baldini C, Bootsma H, et al. Sjögren syndrome. *Nat Rev Dis Primers*. 2016;2:16047. doi:10.1038/nrdp.2016.47
2. Mavragani CP, Moutsopoulos HM. The geoepidemiology of Sjögren's syndrome. *Autoimmun Rev*. 2010;9(5):A305–10. doi:10.1016/j.autrev
3. Baldini C, Fulvio G, La Rocca G, Ferro F. Update on the pathophysiology and treatment of primary Sjögren syndrome. *Nat Rev Rheumatol*. 2024;20(8):473–491. doi:10.1038/s41584-024-01135-3
4. Ramos-Casals M, Brito-Zeron P, Seror R, et al. Characterization of systemic disease in primary Sjogren's syndrome: EULAR-SS task force recommendations for articular, cutaneous, pulmonary and renal involvements. *Rheumatology*. 2015;54(12):223–228. doi:10.1093/rheumatology/kev200
5. James-Goulbourne T, Murugesan V, Kissin EY. Sonographic features of salivary glands in Sjögren's syndrome and its mimics. *Curr Rheumatol Rep*. 2020;22(8):36. PMID: 32562049. doi:10.1007/s11926-020-00914-7
6. Lane JM, Russell L, Khan SN. Osteoporosis. *Clin Orthop Relat Res*. 2000;372:139–150. doi:10.1097/00003086-200003000-00016
7. Black DM, Rosen CJ. Clinical practice. Postmenopausal osteoporosis. *N Engl J Med*. 2016;374(3):254–262. doi:10.1056/NEJMcp1513724
8. Lane NE. Epidemiology, etiology, and diagnosis of osteoporosis. *Am J Obstet Gynecol*. 2006;194(2 Suppl):S3–11. doi:10.1016/j.ajog.2005.08.047
9. Kelsey JL. Risk factors for osteoporosis and associated fractures. *Public Health Rep*. 1989;104:14–20.
10. Gambert SR, Schultz BM, Hamdy RC. Osteoporosis: clinical features, prevention, and treatment. *Endocrinol Metab Clin North Am*. 1995;24(2):317–371. doi:10.1016/S0889-8529(18)30044-6
11. Arceo-Mendoza RM, Camacho PM. Postmenopausal osteoporosis: latest guidelines. *Endocrinol Metab Clin North Am*. 2021;50(2):167–178. doi:10.1016/j.ecl.2021.03.009
12. Hu Z, Zhu S, Zhang B. Impaired bone health in patients with primary Sjogren's syndrome. *Arthritis Rheumatol*. 2016;2016:1.
13. Lai EL, Huang WN, Chen HH, et al. Ten-year fracture risk by FRAX and osteoporotic fractures in patients with systemic autoimmune diseases. *Lupus*. 2019;28:945–953. doi:10.1177/0961203319855122
14. Salman-Monte TC, Sanchez-Piedra C, Fernandez Castro M, et al. Prevalence and factors associated with osteoporosis and fragility fractures in patients with primary Sjögren syndrome. *Rheumatol Int*. 2020;40(8):1259–1265. doi:10.1007/s00296-020-04615-3
15. Gravani F, Papadaki I, Antypa E, et al. Subclinical atherosclerosis and impaired bone health in patients with primary Sjogren's syndrome: prevalence, clinical and laboratory associations. *Arthritis Res Ther*. 2015;17(1):99. doi:10.1186/s13075-015-0613-6
16. Sassi F, Tamone C, D'Amelio P. Vitamin D: nutrient, hormone, and immunomodulator. *Nutrients*. 2018;10(11):1656. doi:10.3390/nu10111656
17. Liu ES, LeBoff MS. Editorial: the role of vitamin D metabolites in the evaluation of bone health: are they physiologically relevant? *J Bone Miner Res*. 2024;39(1):1–2. doi:10.1093/jbmr/zjad012
18. Kumar R. The metabolism and mechanism of action of 1,25-dihydroxyvitamin D3. *Kidney Int*. 1986;30(6):793–803. doi:10.1038/ki.1986.258
19. Haussler MR, Cordy PE. Metabolites and analogues of vitamin D: which for what? *JAMA*. 1982;247(6):841–844. doi:10.1001/jama.1982.03320310085047
20. Henry HL, Norman AW. Vitamin D: metabolism and biological actions. *Annu Rev Nutr*. 1984;4:493–520. doi:10.1146/annurev.nu.04.070184.002425
21. Delrue C, Speeckaert MM. Vitamin D and Vitamin D-Binding Protein in Health and Disease. *Int J Mol Sci*. 2023;24(5):4642. doi:10.3390/ijms24054642
22. Kim JH, Kim K, Kim I, Seong S, Koh JT, Kim N. Stanniocalcin 1 and 1,25-dihydroxyvitamin D3 cooperatively regulate bone mineralization by osteoblasts. *Exp Mol Med*. 2024;56(9):1991–2001. doi:10.1038/s12276-024-01302-2
23. Ismailova A, White JH. Vitamin D, infections and immunity. *Rev Endocr Metab Disord*. 2022;23(2):265–277. doi:10.1007/s11154-021-09679-5

24. Raggatt LJ, Partridge NC. Cellular and molecular mechanisms of bone remodeling. *J Biol Chem.* 2020;285(33):25103–25108. doi:10.1074/jbc.R109.041087
25. Hu K, Olsen BR. Osteoblast-derived VEGF regulates osteoblast differentiation and bone formation during bone repair. *J Clin Invest.* 2016;126(2):509–526. doi:10.1172/JCI82585
26. Kylmaja E, Nakamura M, Tuukkanen J. Osteoclasts and remodeling based bone formation. *Curr Stem Cell Res Ther.* 2016;11(8):626–633. doi:10.2174/1574888x10666151019115724
27. van Driel M, van Leeuwen JPTM. Vitamin D and bone: a story of endocrine and auto/paracrine action in osteoblasts. *Nutrients.* 2023;15(3):480. doi:10.3390/nu15030480
28. Yamamoto Y, Yoshizawa T, Fukuda T, et al. Vitamin D receptor in osteoblasts is a negative regulator of bone mass control. *Endocrinology.* 2013;154(3):1008–1020. doi:10.1210/en.2012-1542
29. Chen C, Adhikari R, White DL, Kim WK. Role of 1,25-Dihydroxyvitamin D3 on osteogenic differentiation and mineralization of chicken mesenchymal stem cells. *Front Physiol.* 2021;12:479596. doi:10.3389/fphys.2021.479596
30. Lee SK, Kalinowski J, Jastrzebski S, Lorenzo JA. 1,25(OH)<sub>2</sub> vitamin D<sub>3</sub>-stimulated osteoclast formation in spleen-osteoblast cocultures is mediated in part by enhanced IL-1 alpha and receptor activator of NF-kappa B ligand production in osteoblasts. *J Immunol.* 2002;169(5):2374–2380. PMID: 12193704. doi:10.4049/jimmunol.169.5.2374
31. Nakamichi Y, Udagawa N, Suda T, Takahashi N. Mechanisms involved in bone resorption regulated by vitamin D. *J Steroid Biochem Mol Biol.* 2018;177:70–76. doi:10.1016/j.jsbmb.2017.11.005
32. Lee SJ, Oh HJ, Choi BY, et al. Serum 25-hydroxyvitamin D3 and BAFF levels are associated with disease activity in primary Sjögren's syndrome. *J Immunol Res.* 2016;2016:5781070. doi:10.1155/2016/5781070
33. Erten Ş, Şahin A, Altunoğlu A, Gemcioğlu E, Koca C. Comparison of plasma vitamin D levels in patients with Sjögren's syndrome and healthy subjects. *Int J Rheum Dis.* 2015;18(1):70–75. doi:10.1111/1756-185X.12298
34. Trotta MC, Herman H, Balta C, et al. Oral administration of vitamin D3 prevents corneal damage in a knock-out mouse model of Sjögren's syndrome. *Biomedicine.* 2023;11(2):616. doi:10.3390/biomedicine11020616
35. Peng Y, Wu X, Zhang S, et al. The potential roles of type I interferon activated neutrophils and neutrophil extracellular traps (NETs) in the pathogenesis of primary Sjögren's syndrome. *Arthritis Res Ther.* 2022;24(1):170. doi:10.1186/s13075-022-02860-4
36. Griffin MD, Lutz W, Phan VA, Bachman LA, McKean DJ, Kumar R. Dendritic cell modulation by 1alpha,25 dihydroxyvitamin D3 and its analogs: a vitamin D receptor-dependent pathway that promotes a persistent state of immaturity in vitro and in vivo. *Proc Natl Acad Sci U S A.* 2001;98:6800–6805. doi:10.1073/pnas.121172198
37. Ferreira GB, Gysemans CA, Demengeot J, et al. 1,25-Dihydroxyvitamin D3 promotes tolerogenic dendritic cells with functional migratory properties in NOD mice. *J Immunol.* 2014;192:4210–4220. doi:10.4049/jimmunol.1302350
38. An Q, Zhao J, Zhu X, et al. Exploiting the role of T cells in the pathogenesis of Sjögren's syndrome for therapeutic treatment. *Front Immunol.* 2022;13:995895. doi:10.3389/fimmu.2022.995895
39. Furuzawa-Carballeda J, Hernández-Molina G, Lima G, Rivera-Vicencio Y, Férrez-Blando K, Llorente L. Peripheral regulatory cells immunophenotyping in primary Sjögren's syndrome: a cross-sectional study. *Arthritis Res Ther.* 2013;15(3):R68. doi:10.1186/ar4245
40. Gao CY, Yao Y, Li L, et al. Tissue-resident memory CD8+CD8+ T cells acting as mediators of salivary gland damage in a murine model of Sjögren's syndrome. *Arthritis Rheumatol.* 2019;71:121–132. doi:10.1002/art.40676
41. Boonstra A, Barrat FJ, Crain C, Heath VL, Savelkoul HF, O'Garra A. 1alpha,25-Dihydroxyvitamin d3 has a direct effect on naive CD4(+) T cells to enhance the development of Th2 cells. *J Immunol.* 2001;167(9):4974–4980. doi:10.4049/jimmunol.167.9.4974
42. Daniel C, Sartory NA, Zahn N, Radeke HH, Stein JM. Immune modulatory treatment of trinitrobenzene sulfonic acid colitis with calcitriol is associated with a change of a T helper (Th) 1/Th17 to a Th2 and regulatory T cell profile. *J Pharmacol Exp Ther.* 2008;324(1):23–33. doi:10.1124/jpet.107.127209
43. Shi L, Wang J, Guo HX, Han XL, Tang YP, Liu GY. Circulating Th2 cell reduction and Th1/Th2 imbalance are correlated with primary Sjögren's syndrome-associated interstitial lung disease. *Arthritis Res Ther.* 2022;24(1):121. doi:10.1186/s13075-022-02811-z
44. Jeffery LE, Burke F, Mura M, et al. 1,25-Dihydroxyvitamin D3 and IL-2 combine to inhibit T cell production of inflammatory cytokines and promote development of regulatory T cells expressing CTLA-4 and FoxP3. *J Immunol.* 2009;183(9):5458–5467. doi:10.4049/jimmunol.0803217
45. Bhatt K, Rafi W, Shah N, Christakos S, Salgame P. 1,25 (OH)<sub>2</sub>D treatment alters the granulomatous response in M. tuberculosis infected mice. *Sci Rep.* 2016;6:34469. doi:10.1038/srep34469
46. Medrano M, Carrillo-Cruz E, Montero I, Perez-Simon JA. Vitamin D: effect on haematopoiesis and immune system and clinical applications. *Int J Mol Sci.* 2018;19(9):2663. doi:10.3390/ijms19092663
47. Zhang Y, Leung DY, Richers BN, et al. Vitamin D inhibits monocyte/macrophage proinflammatory cytokine production by targeting MAPK phosphatase-1. *J Immunol.* 2012;188(5):2127–2135. doi:10.4049/jimmunol.1102412
48. Gevers Leuven JA. Sex steroids and lipoprotein metabolism. *Pharmacol Ther.* 1994;64(1):99–126. doi:10.1016/0163-7258(94)90035-3
49. Khazai N, Judd SE, Tangpricha V. Calcium and vitamin D: skeletal and extraskeletal health. *Curr Rheumatol Rep.* 2008;10(2):110–117. doi:10.1007/s11926-008-0020-y
50. Fannon S, Marts S. Sex, hormones and the cardiovascular system. *Trends Endocrinol Metab.* 2002;13(8):320–321. doi:10.1016/s1043-2760(02)00678-1
51. Oursler MJ. Estrogen regulation of gene expression in osteoblasts and osteoclasts. *Crit Rev Eukaryotic Gene Exp.* 1998;8:125–140. doi:10.1615/CritRevEukarGeneExpr.v8.i2.20
52. Tau KR, Hefferran TE, Waters KM, et al. Estrogen regulation of a transforming growth factor-beta inducible early gene that inhibits deoxyribonucleic acid synthesis in human osteoblasts. *Endocrinology.* 1998;139:1346–1353. doi:10.1210/endo.139.3.5830
53. Ernst M, Hrath JK, Rodan GA. Estradiol effects on proliferation, messenger ribonucleic acid for collagen and insulin-like growth factor-I and parathyroid hormone-stimulated adenylate cyclase activity in osteoblastic cells from calvariae and long bones. *Endocrinology.* 1989;125:825–833. doi:10.1210/endo-125-2-825
54. Ernst M, Rodan GA. Estradiol regulation of insulin-like growth factor-I expression in osteoblastic cells: evidence for transcriptional control. *Mol Endocrinol.* 1991;5:1081–1089. doi:10.1210/mend-5-8-1081

55. Oursler MJ, Osdoby P, Pyfferoen J, Riggs BL, Spelsberg TC. Avian osteoclasts as estrogen target cells. *Proc Natl Acad Sci USA*. 1991;88:6613–6617. doi:10.1073/pnas.88.15.6613
56. Noël M, Chevenne D, Porquet D. Utility of insulin-like growth factor-I and its binding protein assays. *Curr Opin Clin Nutr Metab Care*. 2001;4(5):399–405. doi:10.1097/00075197-200109000-00010
57. Deng Z, Fan T, Xiao C, et al. TGF- $\beta$  signaling in health, disease, and therapeutics. *Signal Transduct Target Ther*. 2024;9(1):61. doi:10.1038/s41392-024-01764-w
58. Kim HN, Ponte F, Nookaew I, et al. Estrogens decrease osteoclast number by attenuating mitochondria oxidative phosphorylation and ATP production in early osteoclast precursors. *Sci Rep*. 2020;10(1):11933. doi:10.1038/s41598-020-68890-7
59. Imai Y, Youn MY, Kondoh S, et al. Estrogens maintain bone mass by regulating expression of genes controlling function and life span in mature osteoclasts. *Ann N Y Acad Sci*. 2009;1173(Suppl 1):E31–9. doi:10.1111/j.1749-6632.2009.04954.x
60. Shevde NK, Bendixen AC, Dienger KM, Pike JW. Estrogens suppress RANK ligand-induced osteoclast differentiation via a stromal cell independent mechanism involving c-Jun repression. *Proc Natl Acad Sci U S A*. 2000;97(14):7829–7834. doi:10.1073/pnas.130200197
61. Christakos S, Prince R. Estrogen, vitamin D, and calcium transport. *J Bone Miner Res*. 2003;18(10):1737–1739. doi:10.1359/jbmr.2003.18.10.1737
62. Goltzman D. Physiology of Parathyroid Hormone. *Endocrinol Metab Clin North Am*. 2018;47(4):743–758. doi:10.1016/j.ecl.2018.07.003
63. Zhang M, Faugere MC, Malluche H, Rosen CJ, Chernausk SD, Clemens TL. Paracrine overexpression of IGFBP-4 in osteoblasts of transgenic mice decreases bone turnover and causes global growth retardation. *J Bone Miner Res*. 2003;18(5):836–843. doi:10.1359/jbmr.2003.18.5.836
64. Hayashi Y, Arakaki R, Ishimaru N. Apoptosis and estrogen deficiency in primary Sjögren syndrome. *Curr Opin Rheumatol*. 2004;16(5):522–526. doi:10.1097/01.bor.0000135450.78047.78
65. Suzuki T, Schaumberg DA, Sullivan BD, et al. Do estrogen and progesterone play a role in the dry eye of Sjögren's syndrome? *Ann N Y Acad Sci*. 2002;966:223–225. doi:10.1111/j.1749-6632.2002.tb04218.x
66. Kontinen YT, Fuellen G, Bing Y, et al. Sex steroids in Sjögren's syndrome. *J Autoimmun*. 2012;39(1–2):49–56. doi:10.1016/j.jaut.2012.01.004
67. Benchabane S, Slimani-Kaddouri A, Acheli D, Bendimerad-Iratene T, Mesbah R, Touil-Boukoffa C. Association between increased Bcl-2, Fas and FasL levels and inflammation extent in labial salivary glands during primary Sjogren's syndrome. *Endocr Metab Immune Disord Drug Targets*. 2022;22(3):328–338. doi:10.2174/1871530321666210809155147
68. Da Y, Niu K, Wang K, et al. A comparison of the effects of estrogen and Cimicifuga racemosa on the lacrimal gland and submandibular gland in ovariectomized rats. *PLoS One*. 2015;10(3):e0121470. doi:10.1371/journal.pone.0121470
69. Azzarolo AM, Eihausen H, Schechter J. Estrogen prevention of lacrimal gland cell death and lymphocytic infiltration. *Exp Eye Res*. 2003;77(3):347–354. doi:10.1016/S0014-4835(03)00120-9
70. Mostafa S, Seamon V, Azzarolo AM. Influence of sex hormones and genetic predisposition in Sjogren's syndrome: a new clue to the immunopathogenesis of dry eye disease. *Exp Eye Res*. 2012;96(1):88–97. doi:10.1016/j.exer.2011.12.016
71. Compston JE. Sex steroids and bone. *Physiol Rev*. 2015;81(1):419–447. doi:10.1152/physrev.2001.81.1.419
72. Bernstein P, Pohost G. Progesterone, progestins, and the heart. *Rev Cardiovasc Med*. 2010;11(4):228–236. doi:10.3909/ricm0557
73. Bencker C, Gschwandtner L, Nayman S, et al. Progestagens and progesterone receptor modulation: effects on the brain, mood, stress, and cognition in females. *Front Neuroendocrinol*. 2025;76:101160. doi:10.1016/j.yfrne.2024.101160
74. Schmidmayr M, Magdolen U, Tübel J, Kiechle M, Burgkart R, Seifert-Klauss V. Progesterone enhances differentiation of primary human osteoblasts in long-term cultures. The influence of concentration and cyclicity of progesterone on proliferation and differentiation of human osteoblasts in vitro. *Geburtsh Frauenheilk*. 2008;68:722–728. doi:10.1055/s-2008-1038711
75. Rickard DJ, Waters KM, Ruesink TJ, et al. Estrogen receptor isoform-specific induction of progesterone receptors in human osteoblasts. *J Bone Miner Res*. 2002;17:580–592. doi:10.1359/jbmr.2002.17.4.580
76. Prior JC. Progesterone for the prevention and treatment of osteoporosis in women. *Climacteric*. 2018;21(4):366–374. doi:10.1080/13697137.2018.1467400
77. Prior JC, Seifert-Klauss VR, Giustini D, Adachi JD, Kalyan S, Goshtasebi A. Estrogen-progestin therapy causes a greater increase in spinal BMD than estrogen therapy—a systematic review and meta-analysis of controlled trials with direct randomization. *J Musculoskelet Neuronal Interact*. 2017;17:146–154.
78. Goldman AL, Bhasin S, Wu FCW, Krishna M, Matsumoto AM, Jasuja R. A reappraisal of Testosterone's binding in circulation: physiological and clinical implications. *Endocr Rev*. 2017;38(4):302–324. doi:10.1210/er.2017-00025
79. Vanderschueren D, Vandenput L, Boonen S, et al. Androgens and bone. *Endocr Rev*. 2004;25:389–425. doi:10.1210/er.2003-0003
80. Sakamoto A, Chen M, Nakamura T, Xie T, Karsenty G, Weinstein LS. Deficiency of the G-protein alpha-subunit G(s)alpha in osteoblasts leads to differential effects on trabecular and cortical bone. *J Biol Chem*. 2005;280(22):21369–21375. doi:10.1074/jbc.M500346200
81. Sims NA, Vrahnas C. Regulation of cortical and trabecular bone mass by communication between osteoblasts, osteocytes and osteoclasts. *Arch Biochem Biophys*. 2014;561:22–28. doi:10.1016/j.abb.2014.05.015
82. Michael H, Härkönen PL, Väänänen HK, Hentunen TA. Estrogen and testosterone use different cellular pathways to inhibit osteoclastogenesis and bone resorption. *J Bone Miner Res*. 2005;20(12):2224–2232. doi:10.1359/JBMR.050803
83. Bodine PV, Riggs BL, Spelsberg TC. Regulation of c-fos expression and TGF-beta production by gonadal and adrenal androgens in normal human osteoblastic cells. *J Steroid Biochem Mol Biol*. 1995;52(2):149–158. doi:10.1016/0960-0760(94)00165-i
84. Pfeilschifter J, Scheidt-Nave C, Leidig-Bruckner G, et al. Relationship between circulating insulin-like growth factor components and sex hormones in a population-based sample of 50- to 80-year-old men and women. *J Clin Endocrinol Metab*. 1996;81(7):2534–2540. doi:10.1210/jcem.81.7.8675573
85. Bellido T, Jilka RL, Boyce BF, et al. Regulation of interleukin-6, osteoclastogenesis, and bone mass by androgens. The role of the androgen receptor. *J Clin Invest*. 1995;95(6):2886–2895. doi:10.1172/JCI117995
86. Martin A, Yu J, Xiong J, et al. Estrogens and androgens inhibit association of RANKL with the pre-osteoblast membrane through post-translational mechanisms. *J Cell Physiol*. 2017;232(12):3798–3807. doi:10.1002/jcp.25862
87. Xuan Y, Zhang X, Wu H. Impact of sex differences on the clinical presentation, pathogenesis, treatment and prognosis of Sjögren's syndrome. *Immunology*. 2024;171(4):513–524. doi:10.1111/imm.13740

88. Porola P, Laine M, Virkki L, Poduval P, Kontinen YT. The influence of sex steroids on Sjögren's syndrome. *Ann N Y Acad Sci.* 2007;1108:426–432. doi:10.1196/annals.1422.045
89. Sullivan DA, Wickham LA, Rocha EM, et al. Androgens and dry eye in Sjögren's syndrome. *Ann N Y Acad Sci.* 1999;876:312–324. doi:10.1111/j.1749-6632.1999.tb07656.x
90. Baudouin C. The pathology of dry eye. *Surv Ophthalmol.* 2001;45(Suppl 2):S211–20. doi:10.1016/s0039-6257(00)00200-9
91. Porola P, Virkki L, Przybyla BD, et al. Androgen deficiency and defective intracrine processing of dehydroepiandrosterone in salivary glands in Sjögren's syndrome. *J Rheumatol.* 2008;35(11):2229–2235. doi:10.3899/jrheum.080220
92. Boyce BF, Xing L. Functions of RANKL/RANK/OPG in bone modeling and remodeling. *Arch Biochem Biophys.* 2008;473(2):139–146. doi:10.1016/j.abb.2008.03.018
93. Kim JM, Lin C, Stavre Z, Greenblatt MB, Shim JH. Osteoblast-osteoclast communication and bone homeostasis. *Cells.* 2020;9(9):2073. doi:10.3390/cells9092073
94. Kobayashi Y, Udagawa N, Takahashi N. Action of RANKL and OPG for osteoclastogenesis. *Crit Rev Eukaryot Gene Expr.* 2009;19(1):61–72. doi:10.1615/critrevukargeneexpr.v19.i1.30
95. Hofbauer LC, Heufelder AE. Role of receptor activator of nuclear factor-kappaB ligand and osteoprotegerin in bone cell biology. *J Mol Med.* 2001;79(5–6):243–253. doi:10.1007/s001090100226
96. Udagawa N, Koide M, Nakamura M, et al. Osteoclast differentiation by RANKL and OPG signaling pathways. *J Bone Miner Metab.* 2021;39(1):19–26. doi:10.1007/s00774-020-01162-6
97. Ono T, Hayashi M, Sasaki F, Nakashima T. RANKL biology: bone metabolism, the immune system, and beyond. *Inflamm Regen.* 2020;40:2. doi:10.1186/s41232-019-0111-3
98. Simonet WS, Lacey DL, Dunstan CR, et al. Osteoprotegerin: a novel secreted protein involved in the regulation of bone density. *Cell.* 1997;89(2):309–319. doi:10.1016/s0092-8674(00)80209-3
99. Zhou P, Zheng T, Zhao B. Cytokine-mediated immunomodulation of osteoclastogenesis. *Bone.* 2022;164:116540. doi:10.1016/j.bone.2022.116540
100. Zhang YH, Heulsmann A, Tondravi MM, Mukherjee A, Abu-Amer Y. Tumor necrosis factor-alpha (TNF) stimulates RANKL-induced osteoclastogenesis via coupling of TNF type 1 receptor and RANK signaling pathways. *J Biol Chem.* 2001;276(1):563–568. doi:10.1074/jbc.M00819820
101. Verstappen GM, Corneth OBJ, Bootsma H, Kroese FGM. Th17 cells in primary Sjögren's syndrome: pathogenicity and plasticity. *J Autoimmun.* 2018;87:16–25. doi:10.1016/j.jaut.2017.11.003
102. Alunno A, Carubbi F, Bistoni O, et al. T regulatory and T helper 17 cells in primary Sjögren's syndrome: facts and perspectives. *Mediators Inflamm.* 2015;2015:243723. doi:10.1155/2015/243723
103. Matsui K, Sano H. T helper 17 cells in primary Sjögren's syndrome. *J Clin Med.* 2017;6(7):65. doi:10.3390/jcm6070065
104. Sato K, Suematsu A, Okamoto K, et al. Th17 functions as an osteoclastogenic helper T cell subset that links T cell activation and bone destruction. *J Exp Med.* 2006;203(12):2673–2682. doi:10.1084/jem.20061775
105. Yeo L, Toellner KM, Salmon M, et al. Cytokine mRNA profiling identifies B cells as a major source of RANKL in rheumatoid arthritis. *Ann Rheum Dis.* 2011;70(11):2022–2028. doi:10.1136/ard.2011.153312
106. Bonfá AC, Seguro LP, Carparo V, Bonfá E, Pereira RM. RANKL and OPG gene polymorphisms: associations with vertebral fractures and bone mineral density in premenopausal systemic lupus erythematosus. *Osteoporos Int.* 2015;26:1563–1571. doi:10.1007/s00198-015-3029-x
107. Bhattacharya G, Sengupta S, Jha R, et al. IL-21/23 axis modulates inflammatory cytokines and RANKL expression in RA CD4+ T cells via p-Akt1 signaling. *Front Immunol.* 2023;14:1235514. doi:10.3389/fimmu.2023.1235514
108. Palmer BF, Kelepouris E, Clegg DJ. Renal tubular acidosis and management strategies: a narrative review. *Adv Ther.* 2021;38(2):949–968. doi:10.1007/s12325-020-01587-5
109. Laing CM, Unwin RJ. Renal tubular acidosis. *J Nephrol.* 2006;19(Suppl 9):S46–52.
110. Maripuri S, Grande JP, Osborn TG, et al. Renal involvement in primary Sjögren's syndrome: a clinicopathologic study. *Clin J Am Soc Nephrol.* 2009;4:1423–1431. doi:10.2215/CJN.00980209
111. Kidder D, Rutherford E, Kipgen D, Fleming S, Geddes C, Stewart GA. Kidney biopsy findings in primary Sjögren syndrome. *Nephrol Dial Transpl.* 2015;30:1363–1369. doi:10.1093/ndt/gfv042
112. Jain A, Srinivas BH, Emmanuel D, Jain VK, Parameshwaran S, Negi VS. Renal involvement in primary Sjögren's syndrome: a prospective cohort study. *Rheumatol Int.* 2018;38:2251–2262. doi:10.1007/s00296-018-4118-x
113. Ren H, Wang WM, Chen XN, et al. Renal involvement and followup of 130 patients with primary Sjögren's syndrome. *J Rheumatol.* 2008;35:278–284.
114. François H, Mariette X. Renal involvement in primary Sjögren syndrome. *Nat Rev Nephrol.* 2016;12(2):82–93. doi:10.1038/nrneph.2015.174
115. Domrongkitchaiporn S, Pongsakul C, Stitchantrakul W, et al. BMD and histology in distal renal tubular acidosis. *Kidney Int.* 2001;59(3):1086–1093. doi:10.1046/j.1523-1755.2001.0590031086.x
116. Sánchez A, Libman J. Trastornos de los mecanismos renales de acidificación en pacientes osteoporóticos [Renal acidification mechanism disorders in patients with osteoporosis]. *Medicina.* 1995;55(3):197–202.
117. Rhen T, Cidlowski JA. Antiinflammatory action of glucocorticoids--new mechanisms for old drugs. *N Engl J Med.* 2005;353(16):1711–1723. doi:10.1056/NEJMra050541
118. Chen M, Fu W, Xu H, Liu CJ. Pathogenic mechanisms of glucocorticoid-induced osteoporosis. *Cytokine Growth Factor Rev.* 2023;70:54–66. doi:10.1016/j.cytogfr.2023.03.002
119. Chotiyarnwong P, McCloskey EV. Pathogenesis of glucocorticoid-induced osteoporosis and options for treatment. *Nat Rev Endocrinol.* 2020;16(8):437–447. doi:10.1038/s41574-020-0341-0
120. van Staa TP, Leufkens HG, Cooper C. The epidemiology of corticosteroid-induced osteoporosis: a meta-analysis. *Osteoporos Int.* 2002;13(10):777–787. doi:10.1007/s001980200108
121. Adami G, Saag KG. Glucocorticoid-induced osteoporosis update. *Curr Opin Rheumatol.* 2019;31(4):388–393. doi:10.1097/BOR.0000000000000608

122. Ahmed M, Gaffen SL. IL-17 inhibits adipogenesis in part via C/EBP $\alpha$ , PPAR $\gamma$  and Krüppel-like factors. *Cytokine*. 2013;61(3):898–905. doi:10.1016/j.cyto.2012.12.007
123. Cheng CH, Chen LR, Chen KH. Osteoporosis due to hormone imbalance: an overview of the effects of estrogen deficiency and glucocorticoid overuse on bone turnover. *Int J Mol Sci*. 2022;23(3):1376. doi:10.3390/ijms23031376
124. Wang J, Zhang Y, Cao J, et al. The role of autophagy in bone metabolism and clinical significance. *Autophagy*. 2023;19(9):2409–2427. doi:10.1080/15548627.2023.2186112
125. Manelli F, Giustina A. Glucocorticoid-induced osteoporosis. *Trends Endocrinol Metab*. 2000;11(3):79–85. doi:10.1016/s1043-2760(00)00234-4
126. Lovšin N, Marc J. Glucocorticoid receptor regulates TNFSF11 transcription by binding to glucocorticoid responsive element in TNFSF11 proximal promoter region. *Int J Mol Sci*. 2021;22(3):1054. doi:10.3390/ijms22031054
127. Carmeliet G, Van Cromphaut S, Daci E, Maes C, Bouillon R. Disorders of calcium homeostasis. *Best Pract Res Clin Endocrinol Metab*. 2003;17(4):529–546. doi:10.1016/j.beem.2003.08.001
128. Rajagopalan PT, Zhang Z, McCourt L, Dwyer M, Benkovic SJ, Hammes GG. Interaction of dihydrofolate reductase with methotrexate: ensemble and single-molecule kinetics. *Proc Natl Acad Sci U S A*. 2002;99(21):13481–13486. doi:10.1073/pnas.172501499
129. Georgiou KR, King TJ, Scherer MA, Zhou H, Foster BK, Xian CJ. Attenuated Wnt/ $\beta$ -catenin signalling mediates methotrexate chemotherapy-induced bone loss and marrow adiposity in rats. *Bone*. 2012;50(6):1223–1233. doi:10.1016/j.bone.2012.03.027
130. Robin F, Cadiou S, Albert JD, Bart G, Coiffier G, Guggenbuhl P. Methotrexate osteopathy: five cases and systematic literature review. *Osteoporos Int*. 2021;32(2):225–232. doi:10.1007/s00198-020-05664-x
131. Yu JJ, Wu F, Ma RR, Wu TJ, Zhang Y, Ying ZH. The effects of iguratimod combined with methotrexate and hydroxychloroquine on BMD in patients with rheumatoid arthritis. *Pharmazie*. 2021;76(10):507–510. doi:10.1691/ph.2021.1630
132. Poutoglidou F, Pourzitaki C, Manthou ME, et al. Effects of long-term methotrexate, infliximab, and tocilizumab administration on bone microarchitecture and tendon morphology in healthy wistar rats. *Cureus*. 2021;13(4):e14696. doi:10.7759/cureus.14696
133. Morgan SL, Chen DT, Carlee J, Baggott JE. Effect of methotrexate therapy on BMD and body composition in rat adjuvant arthritis. *J Rheumatol*. 2004;31(9):1693–1697.
134. Gershwin ME, Goetzl EJ, Steinberg AD. Cyclophosphamide: use in practice. *Ann Intern Med*. 1974;80(4):531–540. doi:10.7326/0003-4819-80-4-531
135. Emadi A, Jones RJ, Brodsky RA. Cyclophosphamide and cancer: golden anniversary. *Nat Rev Clin Oncol*. 2009;6(11):638–647. doi:10.1038/nrclinonc.2009.146
136. Puéchal X, Iudici M, Perrodeau E, et al; French Vasculitis Study Group. Rituximab vs cyclophosphamide induction therapy for patients with granulomatosis with polyangiitis. *JAMA Network Open*. 2022;5(11):e2243799. doi:10.1001/jamanetworkopen.2022.43799
137. Campochiaro C, Allanore Y, Braun-Moscovici Y, Matucci-Cerinic M, Balbir-Gurman A. Is cyclophosphamide still the gold standard in early severe rapidly progressive systemic sclerosis? *Autoimmun Rev*. 2024;23(1):103439. doi:10.1016/j.autrev.2023.103439
138. Al-Rawithi S, El-Yazigi A, Ernst P, Al-Fiar F, Nicholls PJ. Urinary excretion and pharmacokinetics of acrolein and its parent drug cyclophosphamide in bone marrow transplant patients. *Bone Marrow Transplant*. 1998;22(5):485–490. doi:10.1038/sj.bmt.1701355
139. Zhao D, Wang C, Zhao Y, et al. Cyclophosphamide causes osteoporosis in C57BL/6 male mice: suppressive effects of cyclophosphamide on osteoblastogenesis and osteoclastogenesis. *Oncotarget*. 2017;8(58):98163–98183. doi:10.18632/oncotarget.21000
140. Barbarino JM, Staatz CE, Venkataramanan R, Klein TE, Altman RB. PharmGKB summary: cyclosporine and tacrolimus pathways. *Pharmacogenet Genomics*. 2013;23(10):563–585. doi:10.1097/FPC.0b013e328364db84
141. Dai A, Kim SJ. Systemic calcineurin inhibitors tacrolimus and voclosporin: a review of off-label dermatologic uses. *J Am Acad Dermatol*. 2024;90(2):358–367. doi:10.1016/j.jaad.2023.05.074
142. Kanda J, Izumo N, Furukawa M, et al. Effects of the calcineurin inhibitors cyclosporine and tacrolimus on bone metabolism in rats. *Biomed Res*. 2018;39(3):131–139. doi:10.2220/biomedres.39.131
143. Van Scoik KG, Johnson CA, Porter WR. The pharmacology and metabolism of the thiopurine drugs 6-mercaptopurine and azathioprine. *Drug Metab Rev*. 1985;16(1–2):157–174. doi:10.3109/03602538508991433
144. Morgan S, Hooper KM, Milne EM, Farquharson C, Stevens C, Staines KA. Azathioprine has a deleterious effect on the bone health of mice with DSS-induced inflammatory bowel disease. *Int J Mol Sci*. 2019;20(23):6085. doi:10.3390/ijms20236085
145. Bryer HP, Isserow JA, Armstrong EC, et al. Azathioprine alone is bone sparing and does not alter cyclosporin A-induced osteopenia in the rat. *J Bone Miner Res*. 1995;10(1):132–138. doi:10.1002/jbmr.5650100119
146. Ranjbar M, Rahimlou M, Fallah M, Djafarian K, Mohammadi H. Effects of vitamin D supplementation in patients with rheumatoid arthritis: a systematic review and meta-analysis. *Heliyon*. 2025;11:e42463. doi:10.1016/j.heliyon.2025.e42463
147. LeBoff MS, Greenspan SL, Insogna KL, et al. The clinician's guide to prevention and treatment of osteoporosis. *Osteoporos Int*. 2022. doi:10.1007/s00198-021-05900-y
148. Greenblatt MB, Tsai JN, Wein MN. Bone Turnover Markers in the Diagnosis and Monitoring of Metabolic Bone Disease. *Clin Chem*. 2017;63(2):464–474. doi:10.1373/clinchem.2016.259085
149. Voulgaridou G, Papadopoulou SK, Detopoulou P, et al. Vitamin D and calcium in osteoporosis, and the role of bone turnover markers: a narrative review of recent data from RCTs. *Diseases*. 2023;11(1):29. doi:10.3390/diseases11010029
150. Ensrud KE, Crandall CJ. Osteoporosis. *Ann Intern Med*. 2017;167(3):ITC17–ITC32. doi:10.7326/AITC201708010
151. Billington E, Aghajafari F, Skulsky E, Kline GA. Bisphosphonates. *BMJ*. 2024;386:e076898. doi:10.1136/bmj-2023-076898
152. Diab DL, Watts NB. Denosumab in osteoporosis. *Expert Opin Drug Saf*. 2014;13(2):247–253. doi:10.1517/14740338.2014.860133
153. Yao Z, Ayoub A, Srinivasan V, et al. Hydroxychloroquine and a low antiresorptive activity bisphosphonate conjugate prevent and reverse ovariectomy-induced bone loss in mice through dual antiresorptive and anabolic effects. *Bone Res*. 2024;12(1):52. doi:10.1038/s41413-024-00352-6
154. Dubertret L. Methotrexate in dermatology and rheumatology. *Joint Bone Spine*. 2005;72(2):104–105. doi:10.1016/j.jbspin.2004.03.005
155. Gupta K, Ravindran V. Low-dose methotrexate in Rheumatology: a reinvented drug. *J R Coll Physicians Edinb*. 2025;10:14782715241312256. doi:10.1177/14782715241312256
156. Tanaka Y, Soen S, Hirata S, et al. The 2023 Guidelines for the management and treatment of glucocorticoid-induced osteoporosis. *J Bone Miner Metab*. 2024;42:143–154. doi:10.1007/s00774-024-01502-w

**International Journal of General Medicine**

**Publish your work in this journal**

The International Journal of General Medicine is an international, peer-reviewed open-access journal that focuses on general and internal medicine, pathogenesis, epidemiology, diagnosis, monitoring and treatment protocols. The journal is characterized by the rapid reporting of reviews, original research and clinical studies across all disease areas. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/international-journal-of-general-medicine-journal>

**Dovepress**  
Taylor & Francis Group