

Mechanisms of Resistance to ALK Inhibitors and Corresponding Treatment Strategies in Lung Cancer

Jiajun Xie^{1,*}, Yinghao Gao^{2,*}, Weiguo Xu¹, Jing Zhu¹

¹Department of Respiratory and Critical Care Medicine, Mian yang Central Hospital, School of Medicine, University of Electronic Science and Technology of China, Mianyang, People's Republic of China; ²Department of pulmonology, Mianyang hospital of T.C.M, Mianyang, People's Republic of China

*These authors have contributed equally to this work

Correspondence: Jing Zhu; Weiguo Xu, Department of Respiratory and Critical Care Medicine, Mian yang Central Hospital, School of Medicine, University of Electronic Science and Technology of China, Mian yang, People's Republic of China, Tel +86-0816-2243351, Email zhujing_1126@126.com; xwg522630@126.com

Abstract: Lung cancer continues to be a leading cause of cancer-related mortality and morbidity worldwide. The echinoderm microtubule-associated protein-like 4-anaplastic lymphoma kinase (EML4-ALK) fusion gene accounts for approximately 3%-5% of gene mutation types. Targeted therapies for ALK mutations have made significant advancements in recent decades, enabling a considerable number of patients to achieve the goal of five-year survival benefits. However, overcoming the drug resistance that arises with current ALK tyrosine kinase inhibitors (TKIs) remain a major challenge in ALK-targeted therapies. In this review, we briefly discuss the primary and secondary mechanisms of resistance to ALK-TKIs, and explore treatment strategies based on progressive resistance models. Meanwhile, novel drugs and combination therapies are being actively researched and developed to address these challenges. The aim is to offer new insights into the mechanisms of resistance and the corresponding treatment strategies to ALK inhibitors.

Keywords: EML4-ALK, ALK-TKIs, resistance mechanism, treatment strategies, lung cancer

Introduction

Lung cancer is highly malignant¹ and poses a significant threat to human health and life.² Non-small cell lung cancer (NSCLC), one of the most prevalent types, is characterized by high invasiveness, propensity for metastasis, poor prognosis, and high mortality rate.³ In recent years, advancements in genotyping, translational research and drug development have ushered lung cancer treatment into the era of the molecular stratification.⁴ Genetic targets for NSCLC are constantly being identified, matched targeted therapies in development. Among these, epidermal growth factor receptor (EGFR) and anaplastic lymphoma kinase (ALK) are the most commonly driver genes, with the ALK fusion gene present in approximately 5% of NSCLC cases, making it the second most frequent after EGFR mutations.¹ For ALK-positive NSCLC patients, the efficacy of chemotherapy is notably limited.⁵ These patients are also at high risk for intracranial metastasis and are often affected by significant drug-related side effects, such as myelosuppression and gastrointestinal reactions.^{6,7}

The advent of targeted drugs marks the beginning of precision medicine era in the lung cancer treatment. The application of ALK-tyrosine kinase inhibitors (ALK-TKIs) has significantly improved OS and progression-free survival (PFS) in patients with ALK mutations.⁶ Additionally, ALK-TKIs offer patients a better quality of life by minimizing adverse reactions and providing convenient administration options. At present, the first-line ALK-TKIs available for clinicians to choose include crizotinib, ceritinib, alectinib, brigatinib and lorlatinib. This article focuses on ALK fusion

genes, the indications and clinical research data of currently approved ALK inhibitors, mechanisms of drug resistance, and treatment strategies following the development of resistance.

ALK Fusion Gene

ALK was first reported as a component of a subtype of anaplastic large cell lymphoma (ALCL).⁸ Abnormal activation of ALK, due to point mutation or staining of ALK, triggers downstream signals that can drive tumorigenesis.⁹ ALK chromosomal rearrangement is a carcinogenic driving factor of NSCLC, typically resulting from the fusion of the 3'-terminal ALK kinase domain with the 5'-terminal gene partner. To date, more than 90 distinct fusion gene partners have been identified in NSCLC,¹⁰ with the echinoderm microtubule associated protein like 4- anaplastic lymphoma kinase (EML4-ALK) fusion accounting for approximately 85% of cases.¹¹ ALK fusion, mediated by various fusion partners, can form dimers without relying on their ligands, thereby activating the protein kinase domains and triggering multiple downstream signaling pathways that contribute to tumorigenesis (Figure 1).¹² Although ALK fusion partners can influence the intrinsic characteristics of the fusion proteins by altering their stability and kinase activity,¹³ ALK-TKIs are currently not selected according to the fusion type.

The EML4-ALK fusion gene results from a small inversion on the short arm of chromosome 2, where EML4 is fused with the ALK sequence fragment separated by 12 bases.¹¹ (Figure 2) EML4-ALK is more prevalent in young patients, nonsmokers or light smokers, Caucasian adenocarcinoma patients with a high proportion of signet ring cells, and Asian adenocarcinoma patients with a predominance of solid components.^{14,15} ALK mutations are mutually exclusive with other genetic changes, so EGFR and Kirsten rats arcomaviral oncogene homolog (KRAS) mutations are rare in ALK-positive patients.¹⁶ Due to multiple splicing sites at the 5' end of the EML4 gene, at least 17 variant subtypes of EML4-ALK have been identified, including V1, V2, V3a/b, V5a/b, V5', V7 and non-EML4-ALK. Among these, the V1 (E13; A20) and V3 variants (E6; A20) are the most common.¹⁷ These variant subtypes have been linked to differences in drug sensitivity,¹⁸ protein stability,¹⁹ and the emergence of drug resistance mutations.²⁰ Specifically, patients with V1 and V2 variants exhibit significantly higher sensitivity to ALK-TKIs than those with V3 variants, and patients with V5a variants have significantly shorter PFS during ALK-TKIs treatment compare to those with V1 and V2 variants.¹⁸

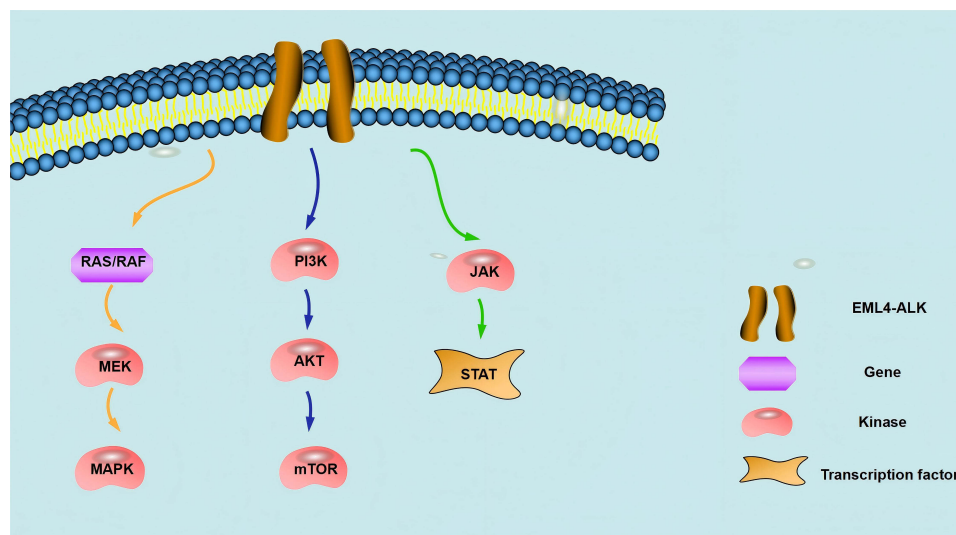


Figure 1 EML4-ALK signaling pathway.

Note: EML4-ALK is localized in the cytoplasm and primarily promotes cell proliferation, invasion, metastasis, and inhibits apoptosis through three signaling pathways: the MAPK/MEK/ERK, PI3K/AKT, and JAK/STAT3 pathways. In the figure, purple represents genes, pink denotes kinases, and light yellow indicates transcription factors.

Abbreviations: EML4-ALK, Echinoderm microtubule associated protein like 4- anaplastic lymphoma kinase; RAS/RAF, rat sarcom/Rapidly Accelerated Fibrosarcoma; MEK, mitogen-activated extracellular signal-regulated kinase; MAPK, mitogen-activated protein kinase; PI3K, Phosphatidylinositol-3-kinase; JAK, just another kinase; STAT, Signal Transducer and Activator of Transcription; mTOR, Mammalian target of rapamycin.

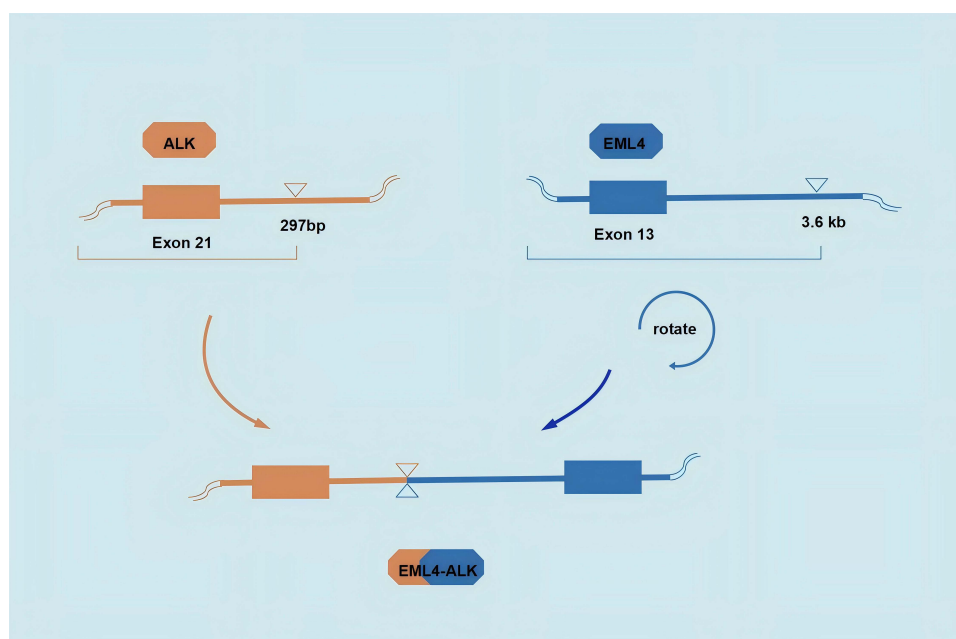


Figure 2 EML4-ALK fusion gene structure.

Note: ALK gene and EML4 gene are directed to chromosome 2p from opposite directions. 297bp upstream of exon ALK 21 is connected to 3.6kb downstream of exon EML4 13, forming EML4-ALK fusion gene (V1 variant subtype).

Abbreviation: EML4-ALK, Echinoderm microtubule associated protein like 4- anaplastic lymphoma kinase.

ALK-TKI Drug Resistance Mechanism

Despite their efficacy, resistance to ALK-TKIs remains a major challenge. This article explores both pharmacological and physiological mechanisms of drug resistance. Physiological drug resistance can be further categorized into primary and secondary drug resistance, with secondary drug resistance mainly including ALK-dependent and ALK-independent resistance mechanisms.

Pharmacological Resistance Mechanism

One important reason for crizotinib resistance is its limited ability to cross the blood–brain barrier. The accumulation of crizotinib in the intracranial space is prevented by p-glycoprotein (ABCBI/ABCG2),²¹ resulting in a concentration of the drug in the blood 384 times higher than in the cerebrospinal fluid.²² Intracranial progression after crizotinib treatment was as high as 41% in the PROFILE 1005 and PROFILE 1007 studies.²³ No pharmacokinetic-related resistance has been observed with second- and third-generation ALK-TKIs to date.

Physiological Resistance Mechanisms

Primary Drug Resistance

The mainstream view is that ALK-positive NSCLC patients with disease progression within 3–6 months of ALK-TKI treatment are considered to have primary drug resistant to ALK-TKIs.²⁴ Most evidence of primary drug resistance comes from case reports. Factors such as KRAS gene co-mutation,^{25,26} EGFR gene co-mutation,²⁷ Bcl-2 interacting mediator of cell death (Bim) gene deletion polymorphism,²⁸ MYC copy number amplification,²⁹ EML4-ALK rearrangement mutation with low mutant allele fraction (MAF),³⁰ and etc., have all been considered to be associated with primary drug resistance. ALK fusion protein and EGFR mutation can co-exist and be expressed in the same tumor cells. Variations in protein phosphorylation levels may affect TKI response in patients with ALK/EGFR co-mutation.²⁸ Additionally, poor clinical outcomes were reported in seven patients with co-mutations of ALK and K-RAS after initial crizotinib treatment.³¹ The traditional “rejection theory” has been challenged by the gradual detection of co-existing mutations

in NSCLC. K-RAS co-mutations, in particular, may serve as a negative predictor of TKI response in patients with ALK-positive NSCLC.³²

High somatic coding mutation loads and DNA repair gene mutations (including TP53) were also found to be involved in primary drug resistance after crizotinib treatment, as demonstrated by Whole Exome Sequencing(WES).³² A study of 18 primary drug-resistant patients, who had a median progression-free survival (mPFS) of only 2.2 months, was published at American Society of Clinical Oncology (ASCO).³³ Primary resistance has been believed to be associated with rare ALK fusion partners (such as ZC3H8-ALK, ALK-LOC1027238, and ALK-DTNB-ASXL2), PTEN/mTOR mutations, the ALK G3709A mutation, and KIT mutations. Paola found that mutations affecting TP53 gene, especially non-disruptive mutations, are able to affect prognosis of ALK-positive patients in NSCLC. This phenomenon is linked to the acquisition of carcinogenic gain-of-function (GOF) mutations, despite non-destructive mutations maintaining some activity of the wild-type TP53 protein.³⁴ Using WES and RNA sequencing data, Professor Jason demonstrated that TP53 mutations were associated with a significantly poorer PFS, though TP53 deletions did not adversely affect outcomes, indicating potential functional distinctions between TP53 deletion and mutation in tumor biology.³⁵ Furthermore, the loss of genes within the ALK region was linked to a shorter PFS. Kevin pointed out that the poor prognostic ALK phenotype likely represents a heterogenous population of primary resistance, early development of resistance and an aggressive disease course.³⁶ The addition of each poor prognostic factor is suspected to be additive or synergistic in conferring a poor prognostic ALK phenotype. The prognostic association of variant 3 is likely related to the stability of the protein and acquired ALK G1202R resistance mutations, which most commonly affect the first- and second-generation inhibitors, but not lorlatinib. Baseline mutations in TP53, cell cycle regulation, and DNA repair likely relate to higher genomic instability and predispose the cancer cell to acquire novel signaling bypass pathways. Baseline alterations in the signaling pathways also likely promote the escape. Cell cycle disruptions and other alterations in DNA repair are other co-mutations that may strongly predict early resistance and progression. Given the limited research on the mechanisms of primary drug resistance, the mechanisms of primary drug resistance are expected to be further explored through large sample studies in the future.

Secondary Drug Resistance

Based on distinct drug resistance mechanisms and corresponding clinical treatment strategies, the secondary drug resistance to ALK-TKIs can be categorized into ALK-dependent and non-ALK-dependent drug resistance. ALK-dependent drug resistance mainly includes secondary gene mutations in the kinase domain and amplification of ALK fusion gene. On the other hand, non-ALK-dependent drug resistance mechanisms primarily involves abnormal activation of bypass signal path, histological transformation, abnormal protein expression, and gene epigenetic regulation.

Secondary Gene Mutations in the Kinase Region

Secondary gene mutations in ALK kinase domain are a common mechanism of drug resistance to ALK-TKIs. These mutations lead to changes in protein conformation and the spatial conformation of the drug-kinase binding region, thus interfering with TKI binding and resulting in acquired drug resistance.³⁷ ALK mutations mainly occur in several areas,³⁸ including the ATP-binding pocket, solvent front, ribose binding pocket, N/C-terminal to the α C-helix, and Asp-Phe-Gly. Approximately 20% of patients develop drug-resistant mutations after receiving first-generation ALK-TKI therapy. The most common mutation associated with crizotinib resistance is L1196M, where a leucine residue is replaced by methionine, which has a longer thioether side chain, preventing crizotinib from binding to the ALK kinase due to steric hindrance. Subsequently, other crizotinib mutations were discovered, including the co-mutation G1202R+G1269A.

The probability of gene mutation after second-generation ALK-TKI treatment exceeds 50%. The G1202R mutation in solvent-region, where glycinate residue is substituted by the larger-volume arginine, is one of the most common gene mutations in second-generation ALK inhibitors. This mutation can disrupt the interaction between the piperidine ring in the structure of ceritinib and alectinib and the kinase domain.³⁹ Some mutations cause ceritinib resistance by producing P-ring fluctuations.^{40,41} Alectinib, Brigatinib and Ensartinib were all detected with different drug resistance mutations, of which G1269A could be inhibited by other second-generation ALK-TKIs in addition to ensartinib.⁴² Lorlatinib is primarily associated with compound resistance mutations.⁴³ Among them, the G1202R+L1196M mutation is currently

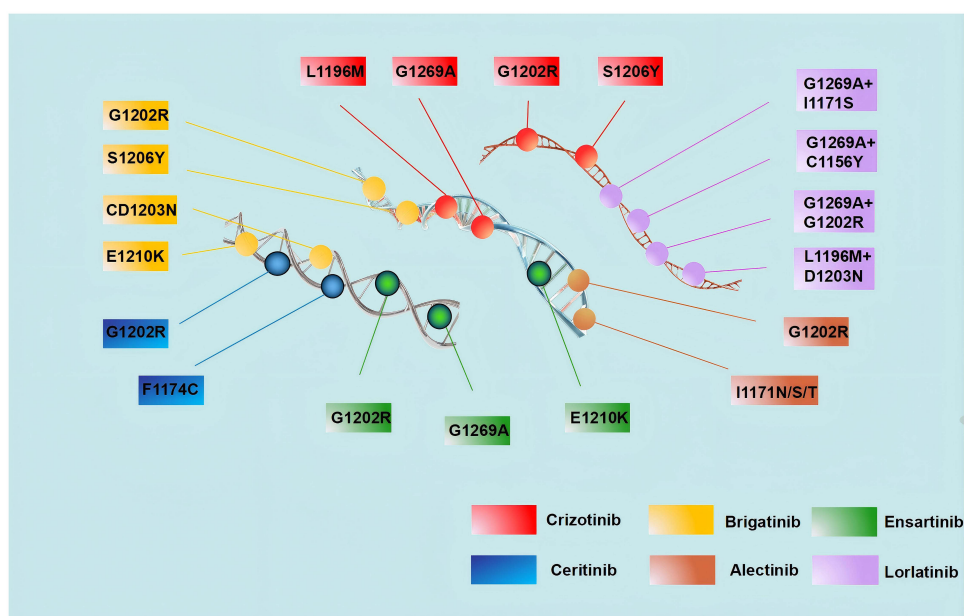


Figure 3 Secondary gene mutations to ALK-TKIs.

Note: The figure shows common gene mutations for the first/second/third-generation ALK-TKIs. Red indicates mutations frequently observed with crizotinib, yellow represents brigatinib-associated mutations, green denotes mutations common to ensartinib, blue corresponds to ceritinib-specific mutations, orange highlights alectinib-related mutations, and purple identifies compound mutations typically associated with lorlatinib.

the most resistant to lorlatinib ($IC_{50} = 1000$ nmol/L) and confers resistant to all ALK-TKIs.⁴⁴ Variant subtypes of EML4-ALK may have different drug resistance mutations. The V3 variant has a higher probability of secondary drug resistance mutations than V1 and V2 variants, and its mutation type tend to be more unique and complex^{17,45} (Figure 3).

ALK Fusion Gene Amplification

Crizotinib-induced increased EML4-ALK copy number was first found in a drug-resistant H3122 cell line (H3122CR).⁴⁶ Subsequently, an increased copy number of the ALK gene was found in two crizotinib-resistant patients, suggesting that gene amplification may also play a role in acquired resistance.⁴⁷ A fusion between Ral GTPase activating protein catalytic subunit alpha 1 Gene (RALGAPA1) and Neuregulin-1 (NRG1) was found in patients treated with alectinib, and gene sequencing confirmed the presence of NRG1 fusion in the cell line (H3122-NRG1) binding RALGAPA1-NRG1 fusion. Moreover, H3122-NRG1 cells showed significant resistance to crizotinib.⁴⁸ Currently, only a small fraction of ALK-TKI-resistant patients have been found to have ALK amplification, and the existing data cannot identify the conditions and critical trigger values of gene amplification leading to drug resistance. Therefore, fusion gene amplification is not considered to be the dominant resistance mechanism of ALK-TKIs.

Abnormal Activation of the Bypass Signal Pathway (Table 1)

When the ALK signaling pathway is inhibited, bypass signaling pathways, such as EGFR, KIT and IGF-1R, are activated.³⁹ These bypass pathways circumvent the original target and activate downstream pathways, leading to ALK inhibitor resistance.⁴⁹ The main ligands of the EGF receptor are epidermal growth factor (EGF) and transforming growth factor- α (TGF- α). As a ligand of c-Met, hepatocyte growth factor (HGF) activates the transmembrane signaling pathway after binding to corresponding receptors, which can promote tumor development, metastasis, invasion and may contribute to drug resistance.⁵⁰ EGFR upregulates its ligand expression and is the most common bypass activation pathway,⁵¹ which can cause resistance to crizotinib, alectinib, and ceritinib. Abnormal activation of EGFR with increased autophosphorylation, is also thought to contribute to crizotinib resistance.⁵² In in vitro experiments, heparin-binding epidermal growth factor-like growth factor (HB-EGF) was found to activate EGFR and trigger resistance to crizotinib through Erk1/2 and Akt transduction bypass signals.⁵³ The abnormal P13K-AKT pathway is associated with patients with crizotinib

Table 1 Activation Pathways of Abnormal Bypass Signaling Pathways to ALK-TKIs

ALK-TKI	Bypass Signal Path	Mode	Ref
Crizotinib	EGFR	Up-regulation	[51]
Crizotinib	EGFR	Autophosphorylation increase	[52]
Crizotinib	HB-EGF	Abnormal activation	[53]
Crizotinib	PI3K-AKT	Abnormal path	[54]
Crizotinib	HER2/3	Amplification	[55]
Crizotinib	PKC	Activation	[55]
Ceritinib	MAP2K1	Mutation	[56]
Ceritinib	SRC signaling	Activation	[58]
Ceritinib	ERK1/2	Activation	[59]
Alectinib	TGF- α	Unknow	[60]
Alectinib	MET	Excitation	[61]
Lorlatinib	NF2	Afunction	[62]
Lorlatinib	MiR-100-5p	Up-regulation	[63]

Abbreviations: EGFR, epidermal growth factor receptor; HB-EGF, heparin-binding epidermal growth factor-like growth factor; PI3K-AKT: Phosphatidylinositol-3-kinase-AK; HER2/3: human epidermal growth factor receptor 2/3; PKC, Protein Kinase C; MAP2K1, mitogen-activated protein kinase kinase 1; SRC, sarcoma gene; ERK1/2, extracellular regulated protein kinases1/2; TGF- α , transforming growth factor- α ; MET, mesenchymal to epithelial transition factor; NF2, neurofibromatosis type 2.

secondary EGFR L858R mutation resistance.⁵⁴ Ligand-mediated human epidermal growth factor receptor 2/3 (HER-2/3) activation and Protein Kinase C(PKC) activation⁵⁵ are also implicated in crizotinib resistance. In a cell model study, the MEK inhibitor smetinib (AZD6244) combined with ceritinib was active in ALK-positive drug-resistant tumors with mitogen-activated protein kinase kinase 1(MAP2K1)-activating mutations,⁵⁶ indicating that acquired resistance can also result from reactivation of downstream effector proteins. The RAS-MEK pathway may be a key downstream effect of EML4-ALK.⁵⁷ Activation of the sarcoma gene (SRC) signaling pathway may also contribute to ceritinib resistance.⁵⁸ SHP099, a newly discovered SHP2 small molecule inhibitor, when combined with ceritinib, prevents the growth of drug-resistant PDCs by blocking the reactivation of RAS, ERK1, and ERK2 (ERK1/2).⁵⁹ TGF- α -induced EGFR pathway activation signal transduction pathway has been observed in alectinib-resistant models. Cell sensitivity to alectinib was restored after the pathway was knocked out.⁶⁰ HGF-induced MET activation stimulates the phosphorylation of the adaptor protein Grb-2-associated binder 1 (Gab1) and activates the downstream signaling pathway, contributing to drug resistance in the alectinib bypass signaling pathway.⁶¹ Loss of neurofibromatosis type 2 (NF2) function has been identified as a bypass resistance mechanism of Lorlatinib.⁶² Additionally, the upregulation of miR-100-5p may inhibit the expression of mTOR pathway-related mRNA, thus leading to ALK-TKI drug resistance.⁶³

Histological Transformation

Histological transformation is another key mechanism of ALK-TKI resistance in NSCLC. Epithelial to mesenchymal transition(EMT) is considered to be the most common morphological change, characterized by loss of epithelial tissue markers and increased expression of interstitial tissue markers, which may be related to upregulation of vimentin expression and downregulation of e-cadherin.⁶⁴ Morphological phenotypic changes were observed in artificially induced crizotinib-resistant spindle cells (H2228/CR), suggesting that EMT may be associated with crizotinib resistance.⁴⁹ Enrichment of mutated genes in crizotinib-resistant tissues revealed four pathways related to EMT, further confirming that EMT plays a role in crizotinib resistance.⁶⁵ Several cases have been reported in which patients with ALK-positive lung adenocarcinoma transformed into small cell lung cancer (SCLC) or squamous cell carcinoma (SCC) after targeted therapy, resulting in drug resistance.^{66,67} Deletion of the retinoblastoma (RB) and p53 genes,⁶⁸ as well as mutations in the PTEN gene,⁶⁹ may have important status in SCLC transformation. For epithelial to EMT-mediated resistance, SRC inhibitors combined with lorlatinib are highly sensitive to patient-derived cell lines.⁶²

Abnormal Protein Expression

The overexpression of P-glycoprotein (P-gp) is found in ceritinib-resistant NSCLC. After downregulation of P-gp, drug-resistant cells become sensitive to ceritinib again.⁷⁰ P-gp may lead to increased drug effects and decreased drug exposure in the central nervous system, which may provide a reference for the poor intracranial activity of ceritinib.⁷⁰ Other studies have shown that exosomes of drug-resistant cells may induce drug resistance in sensitive strains and enhance their cell migration ability.⁷¹

In addition to gene mutations and signaling pathway abnormalities, gene epigenetic regulation plays an important role in the resistance to ALK-TKIs. DNA methylation, one of the most important mechanisms of epigenetic regulation, has been shown that DNA methylation plays a key role in the resistance to ALK-TKIs. Kim built an in vitro model of ceritinib-resistant NSCLC and employed genome-wide DNA methylation analysis combination with single-cell RNA-seq to identify cytidine deaminase (CDA). CDA was hypomethylated and upregulated in ceritinib-resistant cells. Treatment with epigenome-related nucleosides, such as 5-formyl-2'-deoxycytidine, selectively decreased CDA-overexpressing resistant cells via accumulation of DNA damage.⁷² Non-coding RNAs, such as microRNAs (miRNAs) and long non-coding RNAs (lncRNAs), play an important role in regulating gene expression and cell signaling pathways. Byoung investigated the dynamic changes in the transcriptome and enhancer landscape during development of acquired resistance to ALK-TKIs. Histone H3 lysine 27 acetylation (H3K27ac) was profoundly altered during acquisition of resistance, and enhancer remodeling induced expression changes in both miRNAs and mRNAs. The research suggests that enhancer remodeling and altered expression of miRNAs play key roles in cancer drug resistance.⁷³ Future research could focus on exploring the epigenetic mechanisms and provide new therapeutic targets for overcoming resistance.

Treatment Strategies to Drug Resistance

Resistance to ALK-TKIs is inevitable. The subsequent treatment strategies are different according to the progressive models. Meanwhile, novel drug molecules and combination therapies are being researched and developed (Figure 4).

Treatment of “Oligoprogression”

Patients with progressive disease characterized by ≤ 5 metastases or ≤ 2 affected organs after systemic treatment with ALK-TKI were defined as “oligoprogression”.⁷⁴ Patients with oligoprogression can continue with the original TKI therapy combined with local ablation therapy (LAT), such as radiotherapy, radiofrequency ablation, cryoablation, and surgery. Stereotactic radiotherapy (SRT) is a common method used for tumor control, offering low toxicity and effective treatment for oligoradiotherapy in oncogene-driven NSCLC.⁷⁵ Crizotinib combined with radiotherapy has been shown to reduce tumor proliferation, microvascular density and perfusion, while enhancing the inhibitory effect on ALK-positive NSCLC tumor cells.^{76,77} Clinical studies have demonstrated that patients with extracranial oligoprogression who were treated with crizotinib and LAT had actuarial local lesion control rates of 100% and 86% at 6 and 12 months, respectively. The median duration of crizotinib use was prolonged by local treatment (28 months vs 10 months).⁷⁸ However, a preclinical study presented an opposing view, showing that radiotherapy combined with ALK inhibitors did not significantly improve outcomes compared to radiotherapy alone, and EML4-ALK cells treated with ALK inhibitors showed no significant sensitization to radiation.⁷⁹ At present, questions about the safety, mode, and timing of local treatment for minimal progression remain to be answered by more precise prospective studies.

Management of Intracranial Progression

The incidence of brain metastases at the time of first diagnosis in patients with ALK-positive NSCLC is as high as 20%,⁸⁰ and this rate gradually increases with prolonged targeted drug therapy.⁸¹ For patients with 1–4 localized brain metastases after ALK-TKI treatment, stereotactic radiosurgery (SRS) is widely used for its low toxicity and protection of normal brain tissue.⁸² In addition, localized intracranial progression after crizotinib treatment can be managed by increasing the drug dosage.⁸³ Compared with crizotinib, second- and third-generation ALK-TKIs have superior blood–brain barrier penetration and intracranial activity,⁸⁴ which can improve drug resistance caused by low concentration of crizotinib in the brain. Ceritinib, alectinib, brigatinib, and lorlatinib have all been proven effective for treating localized CNS metastases after crizotinib resistance.^{85–88} In patients progressed with second-generation ALK-TKI, lorlatinib readministration achieved a 63.0%

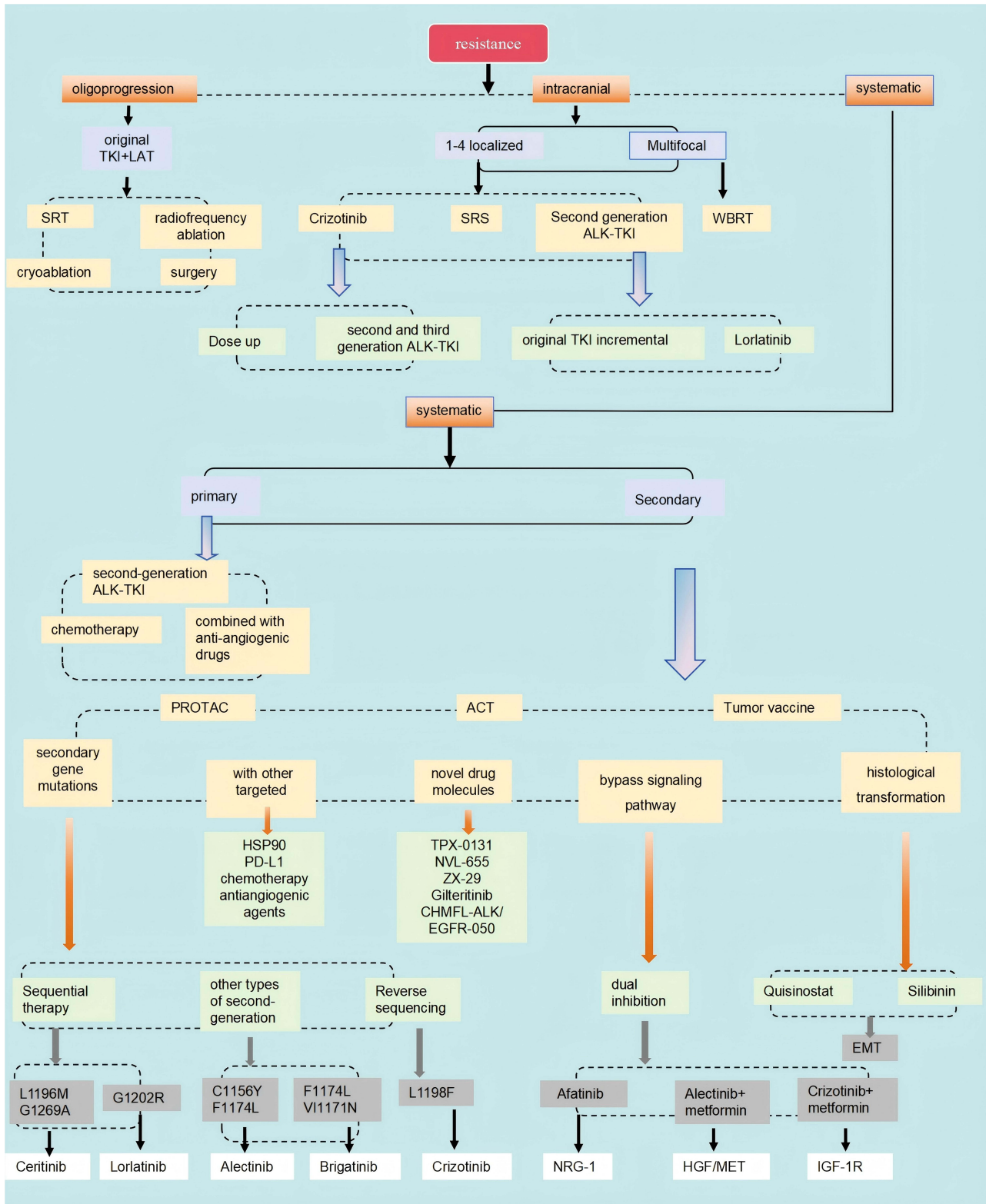


Figure 4 Summary of treatment strategies for drug resistance.
Note: This figure represents the contents of the chapter "Treatment strategies to drug resistance".

intracranial ORR.⁸⁹ Alectinib and brigatinib have also been reported to reinduce CNS disease control by dose escalation.^{90,91} Therefore, for patients with intracranial progression after second-generation ALK-TKI treatment, lorlatinib or increased doses of the original TKI may be considered. For multifocal CNS progression (more than 4 intracranial lesions), whole brain radiation therapy (WBRT) can relieve symptoms in more than 50% of patients as standard treatment.⁹² However, WBRT has a limited effect on survival, with the median survival extension ranging from only 4 to 6 months.⁹² For patients with metastatic adenocarcinoma, combination chemotherapy has certain CNS efficacy.⁹³

Treatment of Systemic Progress

Treatment of Primary Drug Resistance

Patients with EML4-ALK rearrangement who progressed after 1 month of crizotinib treatment were reported to be sensitive to chemotherapy.³⁰ Ceritinib considered beneficial for patients with primary crizotinib resistance.⁹⁴ ALK-positive patients with primary resistance to crizotinib can derive survival benefits from either second-generation ALK-TKIs or chemotherapy, but there is no significant difference between the two.⁹⁵ Nick has reported that dual inhibition of ALK and vascular endothelial growth factor (VEGF) may be a reasonable combined approach.⁹⁶ In addition, previous cases have shown that TKIs combined with anti-VEGF drugs can allow the drugs to reach the tumor site and overcome primary drug resistance.⁹⁷ However, more evidence is needed to support the use of anti-angiogenic therapy in ALK rearranged NSCLC, including pre-clinical, translational, and clinical data.

Treatment of Secondary Gene Mutations in the Kinase Region (Sequential Therapy and ALK Sequencing Analysis)

Sequential therapy is a common strategy to overcome resistance to ALK-TKIs secondary gene mutations, and more than 70% of patients who were resistant to crizotinib chose to continue treatment with progeny ALK-TKIs.⁹⁸ Ceritinib overcomes the common drug resistance mutations of crizotinib by binding specifically to the anterior cleavage site of ALK. The inhibitory effect of alectinib in patients who progressed after treatment with crizotinib was demonstrated in the ALTA-3 trial. Lorlatinib has a good therapeutic effect on almost all the discovered drug-resistant mutations, including high-resistance mutations such as G1202R. In clinical studies, lorlatinib demonstrated an effective rate of 39% in patients who progressed after second-generation TKI treatment, with an ORR reaching 48%.⁸⁹ In addition, after developing resistance to a second-generation ALK-TKI, it is possible to switch to another second-generation ALK-TKI. Alectinib showed high activity against F1174L (ceritinib secondary mutation) and D1203N (brigatinib secondary mutation). After alectinib resistance, I1171/N/S and V1180L mutation sites could be inhibited by Ceritinib. Brigatinib can inhibit ensartinib and alectinib secondary mutations G1269A and I1171N, and even inhibit G1202R. Brigatinib also achieved mPFS at 7 months in patients who progressed after multiline therapy (at least 2 ALK-TKIs).^{99–101} The G1269A mutation developed after enshatinib treatment can be inhibited by other ALK second-generation targeting drugs (excluding enshatinib).¹⁰²

The reverse selection of ALK inhibitors is also recognized in clinical practice. In patients who switch to alectinib after ceritinib resistance, secondary mutations in I1171T and V1180L can be overcome with ceritinib.¹⁰³ Multiple sequential treatments with ALK-TKIs can increase the probability of complex mutations, such as E1210K+D1203N and F1174C+D1203N, but both can be overcome by lorlatinib.^{104,105} Some of the complex mutations after lorlatinib treatment may still be sensitive to second generation ALK-TKIs. These findings suggest that selecting different ALK-TKI combinations for sequential therapy is crucial. Even combining two different ALK inhibitors could potentially extend the survival of patients with ALK mutations.

Patients with ALK gene mutations detected by tissue testing demonstrated significantly higher ORR and PFS when treated with lorlatinib compared to those with negative mutation status (69% vs 27% and 11.0 vs 5.4 months, respectively).¹⁰⁶ Therefore, repeated tissue biopsy and NGS resequencing can provide a basis for clinical decision-making. For highly resistant mutations where no ALK mutation is detected, alternative therapeutic targets should be considered. Circulating tumor DNA obtained from liquid biopsy provides insight into the extent of intra-tumor heterogeneity, potentially supplementing results from re-biopsy in a non-invasive manner. The drug-resistant mutation sites and drug-resistant mutation sites that can be covered by ALK-TKIs in [Table 2](#). Clinical studies on sequential treatment of ALK-TKIs are enrolled in [Table 3](#).

Table 2 Table of Resistant Mutation Sites and Drug Resistance Mutation Sites to ALK-TKIs

ALK-TKI	Probability (%)	Drug	Resistant mutation site	Ref	Resistant mutation sites can be covered	Ref
First generation	20%	Crizotinib	L1196M, G1269A G1202R, S1206Y I1151Tins, L1152R C1156Y, F1174L G1202R+G1269A	[47,107–109]	L1198F	[110]
Second generation	>50%	Ceritinib	G1202R, F1174C T1151K, G1123S	[40,41,52,104]	L1196M, G1269A I1171T, S1206Y I1171N/S, V1180L I1171N+L1196M I1171N +G1269A	[111–115]
		Alectinib	G1202R, I1171N/S/T	[105]	C1156Y, F1174L D1203N, G1269A	[102,105]
		Brigatinib	G1202R, S1206Y CD1203N, E1210K	[116]	G1202R, L1196M F1174L/V, G1269A I1171N, L1198F V1180L I1171N+L1198F I1171N+L1196M I1171N +L1256F	[86,115]
		Ensartinib	G1269A, G1202R, E1210K	[117]	F1174, C1156Y L1196M, S1206R T1151	[118]
Third generation	Unknown	Lorlatinib	G1269A+I1171S/C1156Y/ G1202 L1196M+D1203N G1202R+L1196M/F1174L	[43]	G1202R, I1151Tins I1171N, F1174L E1210K+D1203N F1174C+D1203N	[104–106]

Notes: Resistant mutation site: Secondary resistance mutations that occur after the use of the corresponding drug. Overlays of resistant mutation sites:Secondary drug resistance mutations that occur after ALK-TKIS treatment can be inhibited by corresponding drugs.

Table 3 Clinical Studies on Sequential Treatment of ALK-TKIs

Clinical trial Identifier	First-line	Sequential drug	Phase	Design	Primary endpoint	Secondary Endpoint	Status
NCT03596866	Crizotinib	Brigatinib /Alectinib	3	RCT	PFS	PFS, ORR, DOR	Active not recruiting
NCT04362072	Alectinib/ ceritinib	Lorlatinib	4	Single-Arm	OR	IC-OR, TTR DOR, PFS	Active Not recruiting
NCT04111705	Alectinib/ brigatinib	Lorlatinib	2	Single-Arm	ORR	PFS, DCR DOR, OS	Active Not recruiting
NCT02706626	Alectinib/ ceritinib	Brigatinib	2	Single-Arm	ORR	None	Terminated
NCT04074993	unknown	Brigatinib	2	Single-arm	ORR	DOR, IC-DOR	Active Not recruiting

Abbreviations: RCT, randomized Controlled Trial; PFS, progression-free survival; ORR, objective remission rate; DOR, Duration of Response; MTD, maximum tolerated dose; OR, Objective Response; IC-OR, Intracranial Objective Response; TTR, Time to Response; AEs, Incidence of Adverse events; IC-DOR, Intracranial duration of response.

Treatment of Abnormal Activation of the Bypass Signaling Pathway

To successfully overcome resistance caused by bypass signaling activation, a key therapeutic feature is the dual inhibition of both the oncogene driver and the bypass pathway. Combining ALK-TKIs with EGFR-TKIs can effectively improve drug resistance caused by activation of the downstream EGFR signaling pathway.¹¹⁹ Alectinib combined with metformin may overcome alectinib resistance caused by activation of the HGF/MET signaling pathway.¹²⁰ Similarly, crizotinib combined with metformin also improves resistance mediated by the IGF-1R signaling pathway.¹²¹ In an ALK-rearranged NSCLC cell line containing the origin of the MEK mutation, the combination of the MEK inhibitors selumetinib and

crizotinib reversed H3122CR resistance by inhibiting downstream Ras/MAPK signaling pathways.¹²² Cerivastatin can induce Yes-associated protein(YAP) targeting oncogenes such as AXL, Recombinant Human Cysteine-Rich Angiogenic Inducer 61 (CYR61) and TGF β R2 in cells, providing a theoretical basis for the use of YAP in the treatment of ALK-TKI-resistant patients.¹²³ The combination of alectinib and cerivastatin can successfully reverse the resistance to both alectinib and lorlatinib.¹²⁴ In addition, mTOR inhibitors can overcome lorlatinib resistance caused by mutations leading to the loss of NF2 function.⁶²

Treatment of Histological Transformation

Histone deacetylase (HDAC) inhibitors, such as Quisinostat, can overcome drug resistance by inhibiting EMT through the upregulation of miR-200c.¹²⁵ When ceritinib is combined with Panobinostat,⁷³ it downregulates the expression of neuro-cadherin and vimentin, key markers associated with EMT. This enhances the sensitivity and inhibitory activity of ceritinib to drug-resistant cells, and exerts a strong inhibitory effect on the H3122 cell line and mouse transplanted tumors. Finally, the purpose of reversing drug resistance is achieved. Silibinin overcomes the resistance of lorlatinib and brigatinib due to EMT, restoring function through the transforming growth factor- β /drosophila mothers against decapentaplegic protein (TGF- β /SMAD) signaling pathway.¹²⁶

Combination With Antitumor Drugs of Other Targets (Table 4)

Heat shock protein 90 (HSP90) is highly sensitive to the EML4-ALK fusion protein, and the use of HSP90 inhibitors can block the binding of HSP90 to ATP and promote proteasome-mediated degradation of HSP90 target proteins.¹²⁷ At present, the dual-target inhibitor of HSP90 and ALK-TKI (NCT712217) has entered phase II clinical trials and is expected to combat resistance to ALK-TKI.¹²⁸ After 5 months of nivolumab treatment, the tumor tissue disappeared completely, with a complete response lasting up to 18 months.¹²⁹ The proposed mechanism involves overexpression of the EML4-ALK fusion protein, which upregulates STAT3 protein via hypoxia-inducing factors and signal transcription. This, in turn, increases the expression of Programmed Cell Death-Ligand 1(PD-L1), leading to T-cell suppression and immune escape of tumor cells.¹²⁹ The clinical study evaluating the combination of ALK-TKIs and nivolumab for progressive NSCLC (NCT02393625) has been closed due to grade 3 or greater hepatotoxicity observed.¹³⁰ Meanwhile, the clinical study NCT02584634, which explores the combination of ALK-TKIs and Avelumab, has been transferred to a sub-study for further evaluate efficacy. Retrospective studies have shown that ALK-TKI combined with chemotherapy

Table 4 Clinical Studies to ALK-TKIs Combined With Antitumor Drugs of Other Targets

Clinical trial Identifier	ALK-TKI	Combination drug	Intervention Model	Phase	Primary endpoint	Secondary endpoint	Status
NCT02393625	Ceritinib	Nivolumab	Parallel Assignment	I	MTD, ORR	DOR, DCR TTR, PFS	Active, not recruiting
NCT03087448	Ceritinib	Trametinib	Single Group Assignment	I/II	MTD	ORR, DCR PFS, OS	Terminated
NCT03202940	Alectinib	Cobimetinib	Single Group Assignment	IB/II	MTD	ORR, PFS, OS, DOR	Recruiting
NCT02521051	Alectinib	Bevacizumab	Single Group Assignment	I/II	Recommended phase II dose Safety and tolerability	CNS-ORR CNS-DCR CNS-PFS	Unknown
NCT04005144	Brigatinib	Binimetinib	Single Group Assignment	I	Safety, tolerability	ORR, PFS, OS	Terminated
NCT04292119	Lorlatinib	Crizotinib, Binimetinib, TNO155	Parallel Assignment	I/2	MTD, ORR	PFS, DOR	Unknown
NCT04227028	Brigatinib	Bevacizumab	Single Group Assignment	IB	Incidence of AE MTD	ORR, DOR PFS, OS	Recruiting
NCT02321501	Ceritinib	Everolimus	Single Group Assignment	I/Ib	MTD	RR, PFS,	Active, not recruiting

Abbreviations: MTD, maximum tolerated dose; ORR, objective response rate; DOR, Duration of Response; DCR, disease control rate; TTR, Time to Response; PFS, progression-free survival; OS, overall survival; CNS, Central nervous system; AE, adverse events; RR, Response rate.

is a potential treatment option.¹³¹ Additionally, a combination of ALK-TKIs with antiangiogenic agents (NLCTG1501) showed a DCR of 67% and an ORR of 8%.¹³²

Novel Drugs to Overcome Drug Resistance

Fourth-Generation ALK-TKIs

TPX-0131, a fourth generation of ALK-TKI, was developed to overcome both mutations that arised from continuous ALK-TKIs use and acquired complex double mutation based on G1202R. TPX-0131 is a highly permeable macrocyclic molecule in the CNS that tightly binds to adenine sites in ATP pockets. TPX-0131 can inhibit a variety of ALK-resistant mutations, especially SFM G1202R and the complex L1196M/G1202R. The efficacy of TPX-0131 against G1202R is over 100 times greater than that of lorlatinib, but its efficacy against I1171N and G1269S is less potent.¹³³ TPX-0131 can effectively inhibit both wild-type ALK (IC50: 10 nmol/L) and 26 ALK mutants. In animal experiments, TPX-0131 can completely shrink tumors compared with lorlatinib, and the level of TPX-0131 in rat cerebrospinal fluid is approximately 66% of that in plasma.¹³³ The clinical trial for TPX-0131 (NCT04849273) is currently underway, targeting Alk-positive advanced NSCLC patients who have received less than 3 ALK-TKI treatments, and the primary outcome was ORR.

NVL-655 was designed to increase ALK selectivity and reduce CNS adverse events caused by off-target inhibition of the tropomyosin receptor kinase (TRK) family. NVL-655 effectively inhibit single and complex mutations, including solvent frontier G1202R, G1202R/L1196M, G1202R/G1269A, and G1202R/L1198F.¹³⁴ NVL-655 showed significant efficacy in Ba/F3 transplantation models of EML4-ALK V1 variant subtypes, with much higher in vivo tumor activity than lorlatinib. NVL-655 is highly selective for ALK and does not affect TrkB, so it can minimize TRK-related CNS adverse events.¹³⁵ A phase I/II clinical study of ALKOVE-1 (NCT05384626) is currently underway. The Phase I study is focused on dose escalation and safety to recommend the Phase II dose, while the Phase II study evaluates ORR and efficacy in ALK-positive lung cancer using blinded independent review committees (BICR).

Other New ALK-TKIs

ZX-29, a novel ALK-TKI, demonstrates impressive potency with IC50 values of 2.1 nM for wild-type ALK, 1.3 nM for ALK L1196M, and 3.9 nM for G1202R. ZX-29 can induce protective autophagy and apoptosis by inducing endoplasmic reticulum stress, overcoming cell resistance caused by ALK mutations, and exhibiting more powerful cytotoxicity than ceritinib.^{136,137} Currently, ZX-29 is still in the preclinical development state. Gilteritinib, which acts on R/R acute myeloid leukemia, is an FM-associated receptor tyrosine kinase inhibitor.¹³⁸ Gilteritinib forms hydrogen bonds with residues such as M1199 and E1210, and overcomes single mutations and complex mutations with strong resistance, such as I1171N+L1198H and I1171N+F1174I.^{139,140} Gilteritinib inhibits both wild-type and mutant ALK. Gilteritinib exerts better drug resistance activity than the previous 3 generations of ALK-TKIs by inserting into the ATP-binding site of ALK, providing additional treatment options for drug-resistant patients. Some researchers have also modified ceritinib by replacing the piperidine ring with a group of lower steric hindrance, resulting in compounds effective against both wild-type and crizotinib-resistant mutants, including the highly resistant G1202R.¹⁴¹ Another reported dual-target ALK/EGFR inhibitor, CHMFL-ALK/EGFR-050, has demonstrated strong tumor suppressor activity in vivo and in vitro.¹⁴² CEP-37440, as a dual inhibitor of ALK/FAK, has been shown to be effective against multiple ALK kinase mutations,¹⁴³ with a stronger intracranial drug dose, potentially offering greater therapeutic properties in patients with tumor brain metastases. Other new ALK-TKI candidates include WX-0593,¹⁴⁴ ASP3026,¹⁴⁵ CT-707,¹⁴⁶ TQ-B3139,¹⁴⁷ APG2449 and ZG0418 (Table 5).

PROTACs

Proteolytic targeting chimeras (PROTACs) are an emerging drug development technology that uses the ubiquitin–proteasome system (UPS) to degrade target proteins and has become a hotspot in the field of tumor therapy research.¹⁴⁸ PROTACs can expand the accessibility of targeted proteins, directly degrade without changing enzyme activity and eliminate all functions of the protein, and has low requirements for binding sites. The mechanism leads to the use of PROTACs at lower doses, greatly reducing off-target effects and drug resistance.¹⁴⁸ PROTAC molecules targeting ALK have been gradually developed. MS4077, which uses ceritinib as an ALK ligand and polyethylene glycol (PEG) as

Table 5 Clinical Studies of Novel ALK-TKIs

Drug	Clinical trial Identifier	Phase	Design	Line of therapy	Primary endpoint	Secondary endpoint	Status
TPX-0131	NCT04849273	1	Single-arm	Second line or later	DLTs	AEs	Terminated
NVL-655	NCT05384626	1/2	Single-arm	First-line	DLTs, RP2D, ORR	ORR, DOR, TTR, PFS, OS, CBR	Recruiting
WX-0593	NCT04641754	2	Single-arm	Second line	ORR	PFS, DCR, DOR, TTP	Unknown
ASP3026	NCT01401504	1	Single-arm	First-line	Safety and tolerability	Pharmacokinetics, ORR	Completed
APG-2449	NCT03917043	1	Single-arm	Second line / first-line	MTD, RP2D	Cmax, AUC	Recruiting
ZG0418	NCT03607188	1	Single-Arm	Second line	MTD	None	Recruiting
TQ-B3139	NCT04056572	2	Single-arm	Second line	ORR	PFS, DCR, OS	Unknown

Abbreviations: DLTs, Incidence of first cycle dose-limiting toxicities; AEs, Adverse events; RP2D, Recommended Phase 2 Dose; ORR, objective remission rate; DOR, Duration of Response; TTR, Time to Response; PFS, progression-free survival; OS, overall survival; CBR, clinical benefit rate; DCR, disease control rate; TTP, Time to Progression; MTD, Maximum Tolerated Dose; Cmax, Maximum plasma concentration; AUC, Area under the plasma concentration versus time curve.

a linked chain, can reduce the level of ALK in NCI-H2228 cells.¹⁴⁹ Another novel ALK PROTAC molecule TD-004¹⁵⁰ obtained by replacing the cereblon ligand with the von Hippel-Lindau tumor suppressor(VHL) ligand can effectively promote the degradation of the ALK-EML4 fusion protein in SU-DHL-1 and H3122 cells and inhibit the growth of H3122 xenografts. However, it should be noted that although PROTAC molecules have been partially reported in NSCLC, their poor drug formation and potential off-target toxicity still limit clinical application to a certain extent.

Immunotherapy

Programmed cell death 1 ligand 1 (PD-L1) positive patients occupy a high proportion in ALK-positive NSCLC patients, which¹⁵¹ may be related to the up-regulation of ALK protein in PD-L1 expression in tumor cells.¹⁵² Existing preclinical studies have shown that ALK driver gene positive tumors are resistant to PD-1 and PD-L1 inhibitors.^{153,154} This can be explained by the immune escape of ALK and its aberrations,¹⁵⁵ as well as the poor immunogenicity of the tumor microenvironment in ALK-positive tumors.¹⁵⁶ Clinical trials investigating the combination of immune checkpoint inhibitors with ALK-TKI have yielded no clinical benefits and increased drug toxicity.¹⁵⁷ However, the success of the IMpower150 trial seems to provide new insights into immunotherapy in patients with ALK-positive NSCLC.¹⁵⁸ The combination of VEGF and PD-L1 inhibitors can overcome immunotherapy resistance to ALK-positive NSCLC.¹⁵⁹

Others

Tumor vaccine is a cell fragment or fragment containing a tumor-specific antigen or tumor-associated antigen. Once entering the human body, these cell fragments can activate both fluids and cellular immunity, overcoming the immune suppression state of the body, and suppressing tumor growth. Studies have shown that the novel ALK-TKI vaccine combined with chemotherapy can significantly increase the number of CD8⁺ and CD4⁺ T cells in EML4-ALK mice and improve the survival rate of mice.¹⁶⁰ This provides a prospect for clinical application of the ALK vaccine and its combined use. Adoptive cell therapy (ACT) is a therapeutic method to improve the antitumor effect by diffuses T lymphocytes in vitro and then transfuses them back into the patient.¹⁶¹ A study (NCT03215810) evaluated adoptive therapy of tumor-infiltrating lymphocytes after immunosuppressive resistance in 20 patients, with 3 of 13 patients confirming a response and 11 patients with reduced load, which may suggest a new therapeutic strategy for advanced lung cancer with ALK mutations.¹⁶²

Discussion

Targeted drugs for ALK mutations have been extensively investigated and developed over the past few decades. A considerable number of patients can achieve the goal of five-year survival benefits. First-line use of the second

generation ALK-TKIs showed good mPFS (eg, ALESIA:41.6m,¹⁶³ eXalt3: 25.8m,¹⁶⁴ ALTA-1L:24.0m¹⁶⁵). The third generation ALK-TKIs, when used as first-line therapy, demonstrate favorable 5-year PFS rate (60%) and powerful intracranial control effect.¹⁶⁶ The treatment strategy for ALK-positive patients in the first-line should be selected individually according to the specific conditions of the patient, such as tumor burden, brain metastases, genetic mutation profile, tolerance, and economic conditions.

However, resistance to ALK-TKIs remains unavoidable after first-line use. The mechanism of resistance to lorlatinib, a first-line third-generation ALK-TKI, primarily involves the activation of bypass signaling pathways and complex ALK mutations. ALK-dependent resistance is predominantly driven by secondary ALK mutations, including common complex mutations such as G1202R+L1196M and I1171N+L1196M. Some of these mutations confer high-level resistance to all ALK TKIs, while certain complex mutations may remain sensitive to first- and second-generation TKIs. ALK-independent resistance mechanisms encompass the activation of bypass signaling pathways (35%, eg, MAPK, PI3K/mTOR/PTEN, RTK pathways), MET amplification or rearrangement (22%), and epithelial-mesenchymal transition (EMT). In the Phase III CROWN trial, follow-up treatment options for patients who developed resistance to lorlatinib mainly comprised sequential ALK TKI therapy (61%) and chemotherapy with or without anti-angiogenic agents (34%). Certain complex mutations, such as G1202R+L1196M, can be addressed with dual inhibitors like AG-957/adaphostin. The mechanisms of resistance to second-generation ALK TKIs primarily encompass ALK-dependent mutations (such as G1202R, L1196M) and ALK-independent mechanisms (including bypass activation, MET amplification, and EMT). Approximately 50–60% of these mutations occur within the ALK kinase domain. Lorlatinib effectively inhibits a wide range of ALK-resistant mutations, including G1202R and complex mutations, and addresses central nervous system metastasis. Sequential treatment strategies can prolong the utility of lorlatinib, preserving it as a critical option for later-line therapy. However, the adverse effects of lorlatinib, such as hyperlipidemia and CNS reactions, may limit its long-term use. Sequential approaches can help to balance efficacy and safety. First-line treatment should be individualized based on tumor burden, presence of brain metastasis, mutation profile, drug tolerance, and economic considerations. Upon developing resistance, biopsies should be conducted to determine the underlying resistance mechanism.

Mechanism of ALK-TKIs resistance is gradually becoming clear, and primary drug resistance typically occurs within 3–6 months of treatment. Previous reports revealed that mutations affecting TP53 gene and the poor prognostic ALK phenotype are able to affect prognosis of ALK-positive patients in NSCLC. Given the limited research on the mechanisms of primary drug resistance, there is currently a lack of effective treatment strategies and large-scale clinical study data. The mechanisms of primary drug resistance are expected to be further explored through large sample studies in the future.

Acquired drug resistance mechanisms, such as secondary gene mutation, bypass activation, and EMT, have been gradually recognized, and multiple therapeutic strategies targeting these resistance mechanisms have been developed. For example, the new generation of ALK-TKIs and bypass signaling pathway inhibitors have been targeted and applied in clinical practice, effectively improving the prognosis of patients. Sequential therapy remains a key strategy to overcome the resistance of secondary ALK-TKI gene mutations, but it may change the resistance mechanism of patients. For example, the ALK G1269A and L1196M mutations are the most common mutations in crizotinib-resistant patients, while the second-generation ALK-TKI resistance is mainly driven by ALK G1202R mutations. Multiple resistance mutations often occur after lorlatinib treatment. Repeated biopsies for drug-resistant patients to obtain a drug resistance profile are particularly important in clinical decision-making, and liquid biopsies have the advantage of being noninvasive. Patients treated multiple times with different ALK-TKIs are more easily have coactivation of ALK and bypass signaling pathways. Therefore, developing more long-lasting ALK-TKIs and exploring other targets to provide effective second hits, alongside the use of bypass pathway inhibitors, is critical in delaying drug resistance. While promising, all strategies of addressing drug resistance have shortcomings. The rapid development of new ALK inhibitors is encouraging, such as TPX-0131 and NVL-655, which have obtained amazing data in clinical studies. ZX-29, gilteritinib and other novel TKI preclinical data are also promising. However, the lengthy development cycle, high costs, and the inability to meet urgent clinical needs remain challenges. Combination therapy may offer a solution to enhance the efficacy of ALK-TKIs. For example, other targeted therapies, such as Hsp90, mTOR, and PD-L1, can help counteract the downstream pathway, enhance tumor growth inhibition, and reduce tumor survival and the ability to

adapt to TKI monotherapy by bypassing secondary mutations and signaling. However, the combination of drugs may lead to potential interactions, pharmacokinetic properties of different drugs, and even enhance toxicity and reduce efficacy. The potential of targeted protein-degrading drugs developed by PROTAC technology has also attracted much attention. However, the PROTAC molecule has some problems, such as an excessively large structure, poor water solubility and serious off-target toxicity.

In the future, a deeper understanding of ALK's roles in tumor biology, as well as the mechanisms underlying tumorigenesis and drug resistance, will be essential for overcoming current limitations. Continued exploration of new therapeutic approaches will be key to advancing chronic disease management for ALK-positive NSCLC patients.

Conclusion

The development of ALK-TKIs has significantly prolonged the survival of ALK-positive patients. Although resistance to ALK-TKIs is inevitable, appropriate subsequent therapies can still provide significant clinical benefits. Repeated biopsies are crucial for elucidating the mechanisms of drug resistance in these patients, enabling informed clinical decision-making. The treatment to resistance should be considered on the basis of the resistance mechanisms, the progressive models, drug availability, and therapeutic efficacy. In summary, with ongoing discoveries in resistance mechanisms and advancements in drug development, lung cancer is anticipated to become a manageable chronic condition.

Data Availability Statement

Data Availability Statements are available in section "MDPI Research Data Policies" at <https://www.mdpi.com/ethics>.

Institutional Review Board Statement

The study was conducted in accordance with the Declaration of Helsinki, No applicable for studies not involving humans or animals.

Informed Consent Statement

"Not applicable." for studies not involving humans.

Acknowledgments

Thanks to Zongchang Zhang for his contribution to data analysis, data sources and table completion. Thanks to Gan Yu for revising the language of this article.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

Funding

This study was supported by NHC Key Laboratory of Nuclear Technology Medical Transformation (Mian yang Central Hospital) (Grant No. 2021HYX002), Incubation Project of Mian yang Central Hospital (Grant No.2019FH14) and Mianyang Traditional Chinese Medicine Association Traditional Chinese Medicine Inheritance and Innovation Technology Project (Grant No.MYSZYX-202429). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Disclosure

The authors declare no conflicts of interest in this work.

References

1. Miller KD, Nogueira L, Devasia T, et al. Cancer treatment and survivorship statistics, 2022. *Ca a Cancer J Clinicians*. 2022;72(5):409–436. doi:10.3322/caac.21731
2. Herbst RS, Morgensztern D, Boshoff C. The biology and management of non-small cell lung cancer. *Nature*. 2018;553:7689):446–454. doi:10.1038/nature25183
3. Mao Q, Jiang F, Yin R, et al. Interplay between the lung microbiome and lung cancer. *Cancer Lett*. 2018;415:40–48. doi:10.1016/j.canlet.2017.11.036
4. Thai AA, Solomon BJ, Sequist LV, Gainor JF, Heist RS. Lung cancer. *Lancet (London, England)*. 2021;398(10299):535–554. doi:10.1016/s0140-6736(21)00312-3
5. Wang M, Wang G, Ma H, Shan B. Crizotinib Versus Chemotherapy on ALK-positive NSCLC: a Systematic Review of Efficacy and Safety. *Curr Cancer Drug Targets*. 2019;19(1):41–49. doi:10.2174/1568009617666170623115846
6. Solomon BJ, Mok T, Kim DW, et al. First-line crizotinib versus chemotherapy in ALK-positive lung cancer. *New Engl J Med*. 2014;371(23):2167–2177. doi:10.1056/NEJMoa1408440
7. Soria JC, Tan DSW, Chiari R, et al. First-line ceritinib versus platinum-based chemotherapy in advanced ALK-rearranged non-small-cell lung cancer (ASCEND-4): a randomised, open-label, phase 3 study. *Lancet (London, England)*. 2017;389(10072):917–929. doi:10.1016/s0140-6736(17)30123-x
8. Morris SW, Kirstein MN, Valentine MB, et al. Fusion of a kinase gene, ALK, to a nucleolar protein gene, NPM, in non-Hodgkin's lymphoma. *Science (New York, NY)*. 1995;267:5196):316–7. doi:10.1126/science.267.5196.316-b
9. Hallberg B, Palmer RH. Mechanistic insight into ALK receptor tyrosine kinase in human cancer biology. *Nat Rev Cancer*. 2013;13(10):685–700. doi:10.1038/nrc3580
10. Ou SI, Zhu VW, Nagasaka M. Catalog of 5' Fusion Partners in ALK-positive NSCLC Circa 2020. *JTO Clin Res Rep*. 2020;1(1):100015. doi:10.1016/j.jtocrr.2020.100015
11. Soda M, Choi YL, Enomoto M, et al. Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature*. 2007;448(7153):561–566. doi:10.1038/nature05945
12. Cognigni V, Pecci F, Lupi A, et al. The Landscape of ALK-Rearranged Non-Small Cell Lung Cancer: a Comprehensive Review of Clinicopathologic, Genomic Characteristics, and Therapeutic Perspectives. *Cancers*. 2022;14(19):4765. doi:10.3390/cancers14194765
13. Childress MA, Himmelberg SM, Chen H, Deng W, Davies MA, Lovly CM. ALK Fusion Partners Impact Response to ALK Inhibition: differential Effects on Sensitivity, Cellular Phenotypes, and Biochemical Properties. *mol Cancer Res*. 2018;16(11):1724–1736. doi:10.1158/1541-7786.Mcr-18-0171
14. Sacher AG, Dahlberg SE, Heng J, Mach S, Jänne PA, Oxnard GR. Association Between Younger Age and Targetable Genomic Alterations and Prognosis in Non-Small-Cell Lung Cancer. *JAMA Oncol*. 2016;2(3):313–320. doi:10.1001/jamaoncol.2015.4482
15. Gou LY, Wu YL. Prevalence of driver mutations in non-small-cell lung cancers in the People's Republic of China. *Lung Cancer (Auckland, NZ)*. 2014;5:1–9. doi:10.2147/lett.S40817
16. Sabir SR, Yeoh S, Jackson G, Bayliss R. EML4-ALK Variants: biological and Molecular Properties, and the Implications for Patients. *Cancers*. 2017;9(9):118. doi:10.3390/cancers9090118
17. Lin JJ, Zhu VW, Yoda S, et al. Impact of EML4-ALK Variant on Resistance Mechanisms and Clinical Outcomes in ALK-Positive Lung Cancer. *J Clin Oncol*. 2018;36(12):1199–1206. doi:10.1200/jco.2017.76.2294
18. Christopoulos P, Endris V, Bozorgmehr F, et al. EML4-ALK fusion variant V3 is a high-risk feature conferring accelerated metastatic spread, early treatment failure and worse overall survival in ALK(+) non-small cell lung cancer. *Int J Cancer*. 2018;142(12):2589–2598. doi:10.1002/ijc.31275
19. Heuckmann JM, Balke-Want H, Malchers F, et al. Differential protein stability and ALK inhibitor sensitivity of EML4-ALK fusion variants. *Clin Cancer Res*. 2012;18(17):4682–4690. doi:10.1158/1078-0432.Ccr-11-3260
20. Früh M, Peters S. EML4-ALK Variant Affects ALK Resistance Mutations. *J Clin Oncol*. 2018;36(12):1257–1259. doi:10.1200/jco.2017.77.2764
21. Tang SC, Nguyen LN, Sparidans RW, Wagenaar E, Beijnen JH, Schinkel AH. Increased oral availability and brain accumulation of the ALK inhibitor crizotinib by coadministration of the P-glycoprotein (ABCB1) and breast cancer resistance protein (ABCG2) inhibitor elacridar. *Int J Cancer*. 2014;134(6):1484–1494. doi:10.1002/ijc.28475
22. Costa DB, Kobayashi S, Pandya SS, et al. CSF concentration of the anaplastic lymphoma kinase inhibitor crizotinib. *J Clin Oncol*. 2011;29(15):e443–5. doi:10.1200/jco.2010.34.1313
23. Ou SH, Jänne PA, Bartlett CH, et al. Clinical benefit of continuing ALK inhibition with crizotinib beyond initial disease progression in patients with advanced ALK-positive NSCLC. *Ann Oncol*. 2014;25(2):415–422. doi:10.1093/annonc/mdt572
24. Wang Y, Zhang Y, Chen R, Tian X. Autocrine EGF and TGF- α promote primary and acquired resistance to ALK/c-Met kinase inhibitors in non-small-cell lung cancer. *Pharmacol Res Perspectives*. 2023;11(1):e01047. doi:10.1002/prp2.1047
25. Schmid S, Gautschi O, Rothschild S, et al. Clinical Outcome of ALK-Positive Non-Small Cell Lung Cancer (NSCLC) Patients with De Novo EGFR or KRAS Co-Mutations Receiving Tyrosine Kinase Inhibitors (TKIs). *J Thorac Oncol*. 2017;12(4):681–688. doi:10.1016/j.jtho.2016.12.003
26. Mengoli MC, Barbieri F, Bertolini F, Tiseo M, Rossi G. K-RAS mutations indicating primary resistance to crizotinib in ALK-rearranged adenocarcinomas of the lung: report of two cases and review of the literature. *Lung Cancer*. 2016;93:55–58. doi:10.1016/j.lungcan.2016.01.002
27. Long Y, Zhang K, Li Y, Yu M, Zhu J, Huang M. Durable complete response after Afatinib and crizotinib in an advanced non-small cell lung cancer patient with EGFR L861Q mutation and acquired MET amplification: a case report. *Ann Palliative Med*. 2020;9(5):3609–3613. doi:10.21037/apm-19-482
28. Zhang L, Jiang T, Li X, et al. Clinical features of Bim deletion polymorphism and its relation with crizotinib primary resistance in Chinese patients with ALK/ROS1 fusion-positive non-small cell lung cancer. *Cancer*. 2017;123(15):2927–2935. doi:10.1002/cncr.30677
29. Rihawi K, Alfieri R, Fiorentino M, et al. MYC Amplification as a Potential Mechanism of Primary Resistance to Crizotinib in ALK-Rearranged Non-Small Cell Lung Cancer: a Brief Report. *Transl Oncol*. 2019;12(1):116–121. doi:10.1016/j.tranon.2018.09.013

30. Zhang L, Li Y, Zhang S, Gao C, Nie K, Ji Y. Primary resistance to crizotinib treatment in a non-small cell lung cancer patient with an EML4-ALK rearrangement: a case report. *Cancer Biol Med.* 2018;15(2):178–181. doi:10.20892/j.issn.2095-3941.2018.0003
31. Yang JJ, Zhang XC, Su J, et al. Lung cancers with concomitant EGFR mutations and ALK rearrangements: diverse responses to EGFR-TKI and crizotinib in relation to diverse receptors phosphorylation. *Clin Cancer Res.* 2014;20(5):1383–1392. doi:10.1158/1078-0432.Ccr-13-0699
32. Xiao D, Deng Q, He D, et al. High Tumor Mutation Burden and DNA Repair Gene Mutations are Associated with Primary Resistance to Crizotinib in ALK-Rearranged Lung Cancer. *Onco Targets Ther.* 2021;14:4809–4817. doi:10.2147/ott.S325443
33. Kang. Primary resistance to ALK inhibitor in ALK-positive nonsmall-cell lung cancer. abstract presented at: American Society of Clinical Oncology (ASCO) Annual Meeting; 2017;
34. Canale M, Petracci E, Cravero P, et al. Prognosis of ALK-rearranged non-small-cell lung cancer patients carrying TP53 mutations. *Transl Oncol.* 2022;23:101471. doi:10.1016/j.tranon.2022.101471
35. Couëtoux du Tertre M, Marques M, Tremblay L, et al. Analysis of the Genomic Landscape in ALK+ NSCLC Patients Identifies Novel Aberrations Associated with Clinical Outcomes. *mol Cancer Ther.* 2019;18(9):1628–1636. doi:10.1158/1535-7163.Mct-19-0105
36. Chan SWS, Zeng J, Young J, et al. A Poor Prognostic ALK Phenotype: a Review of Molecular Markers of Poor Prognosis in ALK Rearranged Nonsmall Cell Lung Cancer. *Clin Lung Cancer.* 2025;26(1):e22–e32.e2. doi:10.1016/j.clc.2024.10.009
37. Lin JJ, Shaw AT. Resisting Resistance: targeted Therapies in Lung Cancer. *Trends Cancer.* 2016;2(7):350–364. doi:10.1016/j.trecan.2016.05.010
38. Zhang S, Wang F, Keats J, et al. Crizotinib-resistant mutants of EML4-ALK identified through an accelerated mutagenesis screen. *Chem. Biol. Drug Des.* 2011;78(6):999–1005. doi:10.1111/j.1747-0285.2011.01239.x
39. Kong X, Pan P, Sun H, et al. Drug Discovery Targeting Anaplastic Lymphoma Kinase (ALK). *J Med Chem.* 2019;62(24):10927–10954. doi:10.1021/acs.jmedchem.9b00446
40. He MY, Li WK, Meiler J, Zheng QC, Zhang HX. Insight on mutation-induced resistance to anaplastic lymphoma kinase inhibitor ceritinib from molecular dynamics simulations. *Biopolymers.* 2019;110(2):e23257. doi:10.1002/bip.23257
41. Zhu VW, Cui JJ, Fernandez-Rocha M, Schrock AB, Ali SM, Ou SI. Identification of a novel T1151K ALK mutation in a patient with ALK-rearranged NSCLC with prior exposure to crizotinib and ceritinib. *Lung Cancer.* 2017;110:32–34. doi:10.1016/j.lungcan.2017.05.018
42. Horn L, Whisenant JG, Wakelee H, et al. Monitoring Therapeutic Response and Resistance: analysis of Circulating Tumor DNA in Patients With ALK+ Lung Cancer. *J Thorac Oncol.* 2019;14(11):1901–1911. doi:10.1016/j.jtho.2019.08.003
43. Zhu VW, Nagasaka M, Madison R, Schrock AB, Cui J, Ou SI. A Novel Sequentially Evolved EML4-ALK Variant 3 G1202R/S1206Y Double Mutation In Cis Confers Resistance to Lorlatinib: a Brief Report and Literature Review. *JTO Clin Res Rep.* 2021;2(1):100116. doi:10.1016/j.jtocrr.2020.100116
44. Yoda S, Lin JJ, Lawrence MS, et al. Sequential ALK Inhibitors Can Select for Lorlatinib-Resistant Compound ALK Mutations in ALK-Positive Lung Cancer. *Cancer Discovery.* 2018;8(6):714–729. doi:10.1158/2159-8290.Cd-17-1256
45. Hou D, Zheng X, Song W, et al. Association of anaplastic lymphoma kinase variants and alterations with ensartinib response duration in non-small cell lung cancer. *Thoracic Cancer.* 2021;12(17):2388–2399. doi:10.1111/1759-7714.14083
46. Katayama R, Khan TM, Benes C, et al. Therapeutic strategies to overcome crizotinib resistance in non-small cell lung cancers harboring the fusion oncogene EML4-ALK. *Proc Natl Acad Sci USA.* 2011;108(18):7535–7540. doi:10.1073/pnas.1019559108
47. Doebele RC, Pilling AB, Aisner DL, et al. Mechanisms of resistance to crizotinib in patients with ALK gene rearranged non-small cell lung cancer. *Clin Cancer Res.* 2012;18(5):1472–1482. doi:10.1158/1078-0432.Ccr-11-2906
48. McCoach CE, Le AT, Gowan K, et al. Resistance Mechanisms to Targeted Therapies in ROS1(+) and ALK(+) Non-small Cell Lung Cancer. *Clin Cancer Res.* 2018;24(14):3334–3347. doi:10.1158/1078-0432.Ccr-17-2452
49. Kim HR, Kim WS, Choi YJ, Choi CM, Rho JK, Lee JC. Epithelial-mesenchymal transition leads to crizotinib resistance in H2228 lung cancer cells with EML4-ALK translocation. *mol Oncol.* 2013;7(6):1093–1102. doi:10.1016/j.molonc.2013.08.001
50. Corso S, Giordano S. Cell-autonomous and non-cell-autonomous mechanisms of HGF/MET-driven resistance to targeted therapies: from basic research to a clinical perspective. *Cancer Discovery.* 2013;3(9):978–992. doi:10.1158/2159-8290.Cd-13-0040
51. Isozaki H, Takigawa N, Kiura K. Mechanisms of Acquired Resistance to ALK Inhibitors and the Rationale for Treating ALK-positive Lung Cancer. *Cancers.* 2015;7(2):763–783. doi:10.3390/cancers7020763
52. Katayama R, Shaw AT, Khan TM, et al. Mechanisms of acquired crizotinib resistance in ALK-rearranged lung Cancers. *Sci, Trans Med.* 2012;4(120):120ra17. doi:10.1126/scitranslmed.3003316
53. Yamada T, Takeuchi S, Nakade J, et al. Paracrine receptor activation by microenvironment triggers bypass survival signals and ALK inhibitor resistance in EML4-ALK lung cancer cells. *Clin Cancer Res.* 2012;18(13):3592–3602. doi:10.1158/1078-0432.Ccr-11-2972
54. Kim S, Kim TM, Kim DW, et al. Heterogeneity of genetic changes associated with acquired crizotinib resistance in ALK-rearranged lung cancer. *J Thorac Oncol.* 2013;8(4):415–422. doi:10.1097/JTO.0b013e318283dccc
55. Wilson FH, Johannessen CM, Piccioni F, et al. A functional landscape of resistance to ALK inhibition in lung cancer. *Cancer Cell.* 2015;27(3):397–408. doi:10.1016/j.ccell.2015.02.005
56. Crystal AS, Shaw AT, Sequist LV, et al. Patient-derived models of acquired resistance can identify effective drug combinations for cancer. *Science (New York, NY).* 2014;346(6216):1480–1486. doi:10.1126/science.1254721
57. Hrustanovic G, Olivás V, Pazarentzos E, et al. RAS-MAPK dependence underlies a rational polytherapy strategy in EML4-ALK-positive lung cancer. *Nature Med.* 2015;21(9):1038–1047. doi:10.1038/nm.3930
58. Cooper MR, Chim H, Chan H, Durand C. Ceritinib: a new tyrosine kinase inhibitor for non-small-cell lung cancer. *Ann Pharmacotherapy.* 2015;49(1):107–112. doi:10.1177/1060028014553619
59. Dardaai L, Wang HQ, Singh M, et al. SHP2 inhibition restores sensitivity in ALK-rearranged non-small-cell lung cancer resistant to ALK inhibitors. *Nature Med.* 2018;24(4):512–517. doi:10.1038/nm.4497
60. Miyawaki M, Yasuda H, Tani T, et al. Overcoming EGFR Bypass Signal-Induced Acquired Resistance to ALK Tyrosine Kinase Inhibitors in ALK-Translocated Lung Cancer. *mol Cancer Res.* 2017;15(1):106–114. doi:10.1158/1541-7786.Mcr-16-0211
61. Isozaki H, Hotta K, Ichihara E, et al. Protocol Design for the Bench to Bed Trial in Alectinib-Refractory Non-Small-Cell Lung Cancer Patients Harboring the EML4-ALK Fusion Gene (ALRIGHT/OLCSG1405). *Clin Lung Cancer.* 2016;17(6):602–605. doi:10.1016/j.clc.2016.05.005

62. Recondo G, Mezquita L, Facchinetti F, et al. Diverse Resistance Mechanisms to the Third-Generation ALK Inhibitor Lorlatinib in ALK-Rearranged Lung Cancer. *Clin Cancer Res.* 2020;26(1):242–255. doi:10.1158/1078-0432.Ccr-19-1104
63. Lai Y, Kacal M, Kanony M, et al. miR-100-5p confers resistance to ALK tyrosine kinase inhibitors Crizotinib and Lorlatinib in EML4-ALK positive NSCLC. *Biochem. Biophys. Res. Commun.* 2019;511(2):260–265. doi:10.1016/j.bbrc.2019.02.016
64. Usman S, Waseem NH, Nguyen TKN, et al. Vimentin Is at the Heart of Epithelial Mesenchymal Transition (EMT) Mediated Metastasis. *Cancers.* 2021;13(19):4985. doi:10.3390/cancers13194985
65. Wei J, van der Wekken AJ, Saber A, et al. Mutations in EMT-Related Genes in ALK Positive Crizotinib Resistant Non-Small Cell Lung Cancers. *Cancers.* 2018;10(1):10. doi:10.3390/cancers10010010
66. Oya Y, Yoshida T, Uemura T, Murakami Y, Inaba Y, Hida T. Serum ProGRP and NSE levels predicting small cell lung cancer transformation in a patient with ALK rearrangement-positive non-small cell lung cancer: a case report. *Oncol Lett.* 2018;16(4):4219–4222. doi:10.3892/ol.2018.9158
67. Park S, Han J, Sun JM. Histologic transformation of ALK-rearranged adenocarcinoma to squamous cell carcinoma after treatment with ALK inhibitor. *Lung Cancer.* 2019;127:66–68. doi:10.1016/j.lungcan.2018.11.027
68. Ou SI, Lee TK, Young L, et al. Dual occurrence of ALK G1202R solvent front mutation and small cell lung cancer transformation as resistance mechanisms to second generation ALK inhibitors without prior exposure to crizotinib. Pitfall of solely relying on liquid re-biopsy?. *Lung Cancer.* 2017;106:110–114. doi:10.1016/j.lungcan.2017.02.005
69. Levaq D, D'Haene N, de Wind R, Rimmeling M, Berghmans T. Histological transformation of ALK rearranged adenocarcinoma into small cell lung cancer: a new mechanism of resistance to ALK inhibitors. *Lung Cancer.* 2016;102:38–41. doi:10.1016/j.lungcan.2016.10.012
70. Katayama R, Sakashita T, Yanagitani N, et al. P-glycoprotein Mediates Ceritinib Resistance in Anaplastic Lymphoma Kinase-rearranged Non-small Cell Lung Cancer. *EBioMedicine.* 2016;3:54–66. doi:10.1016/j.ebiom.2015.12.009
71. Kwok HH, Ning Z, Chong PW, et al. Transfer of Extracellular Vesicle-Associated-RNAs Induces Drug Resistance in ALK-Translocated Lung Adenocarcinoma. *Cancers.* 2019;11(1):104. doi:10.3390/cancers11010104
72. Heo H, Kim JH, Lim HJ, et al. DNA methylome and single-cell transcriptome analyses reveal CDA as a potential druggable target for ALK inhibitor-resistant lung cancer therapy. *Exp mol Med.* 2022;54(8):1236–1249. doi:10.1038/s12276-022-00836-7
73. Yun MR, Lim SM, Kim SK, et al. Enhancer Remodeling and MicroRNA Alterations Are Associated with Acquired Resistance to ALK Inhibitors. *Cancer Res.* 2018;78(12):3350–3362. doi:10.1158/0008-5472.Can-17-3146
74. Dingemans AC, Hendriks LEL, Berghmans T, et al. Definition of Synchronous Oligometastatic Non-Small Cell Lung Cancer-A Consensus Report. *J Thorac Oncol.* 2019;14(12):2109–2119. doi:10.1016/j.jtho.2019.07.025
75. Campo M, Al-Halabi H, Khandekar M, Shaw AT, Sequist LV, Willers H. Integration of Stereotactic Body Radiation Therapy With Tyrosine Kinase Inhibitors in Stage IV Oncogene-Driven Lung Cancer. *oncologist.* 2016;21(8):964–973. doi:10.1634/theoncologist.2015-0508
76. Dai Y, Wei Q, Schwager C, et al. Synergistic effects of crizotinib and radiotherapy in experimental EML4-ALK fusion positive lung cancer. *Radiotherapy Oncol.* 2015;114(2):173–181. doi:10.1016/j.radonc.2014.12.009
77. Sun Y, Nowak KA, Zaorsky NG, et al. ALK inhibitor PF02341066 (crizotinib) increases sensitivity to radiation in non-small cell lung cancer expressing EML4-ALK. *mol Cancer Ther.* 2013;12(5):696–704. doi:10.1158/1535-7163.Mct-12-0868
78. Gan GN, Weickhardt AJ, Scheier B, et al. Stereotactic radiation therapy can safely and durably control sites of extra-central nervous system oligoprogressive disease in anaplastic lymphoma kinase-positive lung cancer patients receiving crizotinib. *Int J Radiat Oncol Biol Phys.* 2014;88(4):892–898. doi:10.1016/j.ijrobp.2013.11.010
79. Fleschutz K, Walter L, Leistner R, Heinzlerling L. ALK Inhibitors Do Not Increase Sensitivity to Radiation in EML4-ALK Non-small Cell Lung Cancer. *Anticancer Res.* 2020;40(9):4937–4946. doi:10.21873/anticancer.14497
80. Tan AC, Itchins M, Khasraw M. Brain Metastases in Lung Cancers with Emerging Targetable Fusion Drivers. *Int J mol Sci.* 2020;21(4):1416. doi:10.3390/ijms21041416
81. Rangachari D, Yamaguchi N, VanderLaan PA, et al. Brain metastases in patients with EGFR-mutated or ALK-rearranged non-small-cell lung cancers. *Lung Cancer.* 2015;88(1):108–111. doi:10.1016/j.lungcan.2015.01.020
82. Habets EJ, Dirven L, Wiggeraad RG, et al. Neurocognitive functioning and health-related quality of life in patients treated with stereotactic radiotherapy for brain metastases: a prospective study. *Neuro-Oncology.* 2016;18(3):435–444. doi:10.1093/neuonc/nov186
83. Wang S, Chen J, Xie Z, et al. Pulsatile crizotinib treatment for brain metastasis in a patient with non-small-cell lung cancer. *J Clin Pharm Therapeutics.* 2017;42(5):627–630. doi:10.1111/jcpt.12550
84. Venur VA, Ahluwalia MS Targeted Therapy in Brain Metastases: ready for Primetime? *American Society of Clinical Oncology Educational Book American Society of Clinical Oncology Annual Meeting.* 2016;35:e123–30. doi:10.1200/edbk_100006
85. Chow LQM, Barlesi F, Bertino EM, et al. ASCEND-7: efficacy and Safety of Ceritinib Treatment in Patients with ALK-Positive Non-Small Cell Lung Cancer Metastatic to the Brain and/or Leptomeninges. *Clin Cancer Res.* 2022;28(12):2506–2516. doi:10.1158/1078-0432.Ccr-21-1838
86. Zhang S, Anjum R, Squillace R, et al. The Potent ALK Inhibitor Brigatinib (AP26113) Overcomes Mechanisms of Resistance to First- and Second-Generation ALK Inhibitors in Preclinical Models. *Clin Cancer Res.* 2016;22(22):5527–5538. doi:10.1158/1078-0432.Ccr-16-0569
87. He M, Li W, Zheng Q, Zhang H. A molecular dynamics investigation into the mechanisms of alectinib resistance of three ALK mutants. *J Cell Biochem.* 2018;119(7):5332–5342. doi:10.1002/jcb.26666
88. Solomon BJ, Bauer TM, Mok TSK, et al. Efficacy and safety of first-line lorlatinib versus crizotinib in patients with advanced, ALK-positive non-small-cell lung cancer: updated analysis of data from the phase 3, randomised, open-label CROWN study. *Lancet Respir Med.* 2023;11(4):354–366. doi:10.1016/s2213-2600(22)00437-4
89. Solomon BJ, Besse B, Bauer TM, et al. Lorlatinib in patients with ALK-positive non-small-cell lung cancer: results from a global phase 2 study. *Lancet Oncol.* 2018;19(12):1654–1667. doi:10.1016/s1470-2045(18)30649-1
90. Gainer JF, Chi AS, Logan J, et al. Alectinib Dose Escalation Reinduces Central Nervous System Responses in Patients with Anaplastic Lymphoma Kinase-Positive Non-Small Cell Lung Cancer Relapsing on Standard Dose Alectinib. *J Thorac Oncol.* 2016;11(2):256–260. doi:10.1016/j.jtho.2015.10.010

91. Urbanska EM, Santoni-Rugiu E, Melchior LC, Carlsen JF, Sørensen JB. Intracranial Response of ALK(+) Non-Small-cell Lung Cancer to Second-line Dose-escalated Brigatinib After Alectinib Discontinuation Due to Drug-induced Hepatitis and Relapse After Whole Brain Radiotherapy Followed by Stereotactic Radiosurgery. *Clin Lung Cancer*. 2021;22(4):e528–e532. doi:10.1016/j.clcc.2020.04.012
92. Chen H, Wu A, Tao H, et al. Concurrent versus sequential whole brain radiotherapy and TKI in EGFR-mutated NSCLC patients with brain metastasis: a single institution retrospective analysis. *Medicine*. 2018;97(44):e13014. doi:10.1097/md.00000000000013014
93. Bearz A, Garassino I, Tiseo M, et al. Activity of Pemetrexed on brain metastases from Non-Small Cell Lung Cancer. *Lung Cancer*. 2010;68(2):264–268. doi:10.1016/j.lungcan.2009.06.018
94. Facchinetti F, Caramella C, Auger N, et al. Crizotinib primary resistance overcome by ceritinib in a patient with ALK-rearranged non-small cell lung cancer. *Tumori*. 2016;102(Suppl. 2):S46–S49. doi:10.5301/tj.5000520
95. Ma D, Zhang Y, Xing P, et al. Clinical features and outcomes of ALK rearranged non-small cell lung cancer with primary resistance to crizotinib. *Thoracic Cancer*. 2019;10(5):1213–1219. doi:10.1111/1759-7714.13071
96. Tan AC, Pavlakis N. Anti-Angiogenic Therapy in ALK Rearranged Non-Small Cell Lung Cancer (NSCLC). *Int J mol Sci*. 2022;23(16):8863. doi:10.3390/ijms23168863
97. Nakasuka T, Ichihara E, Makimoto G, Maeda Y, Kiura K. Primary Resistance to Alectinib Was Lost after Bevacizumab Combined Chemotherapy in ALK-Rearranged Lung Adenocarcinoma. *J Thorac Oncol*. 2019;14(8):e168–e169. doi:10.1016/j.jtho.2019.03.009
98. Elsayed M, Bozorgmehr F, Kazdal D, et al. Feasibility and Challenges for Sequential Treatments in ALK-Rearranged Non-Small-Cell Lung Cancer. *Front Oncol*. 2021;11:670483. doi:10.3389/fonc.2021.670483
99. Popat S, Brustugun OT, Cadranel J, et al. Real-world treatment outcomes with brigatinib in patients with pretreated ALK+ metastatic non-small cell lung cancer. *Lung Cancer*. 2021;157:9–16. doi:10.1016/j.lungcan.2021.05.017
100. Descourt R, Perol M, Rousseau-Bussac G, et al. Brigatinib in patients with ALK-positive advanced non-small-cell lung cancer pretreated with sequential ALK inhibitors: a multicentric real-world study (BRIGALK study). *Lung Cancer*. 2019;136:109–114. doi:10.1016/j.lungcan.2019.08.010
101. Nishio M, Yoshida T, Kumagai T, et al. Brigatinib in Japanese Patients With ALK-Positive NSCLC Previously Treated With Alectinib and Other Tyrosine Kinase Inhibitors: outcomes of the Phase 2 J-ALTA Trial. *J Thorac Oncol*. 2021;16(3):452–463. doi:10.1016/j.jtho.2020.11.004
102. Yang Y, Zhou J, Zhou J, et al. Efficacy, safety, and biomarker analysis of ensartinib in crizotinib-resistant, ALK-positive non-small-cell lung cancer: a multicentre, phase 2 trial. *Lancet Respir Med*. 2020;8(1):45–53. doi:10.1016/s2213-2600(19)30252-8
103. Ou SH, Greenbowe J, Khan ZU, et al. I1171 missense mutation (particularly I1171N) is a common resistance mutation in ALK-positive NSCLC patients who have progressive disease while on alectinib and is sensitive to ceritinib. *Lung Cancer*. 2015;88(2):231–234. doi:10.1016/j.lungcan.2015.02.005
104. Gainor JF, Dardaie I, Yoda S, et al. Molecular Mechanisms of Resistance to First- and Second-Generation ALK Inhibitors in ALK-Rearranged Lung Cancer. *Cancer Discovery*. 2016;6(10):1118–1133. doi:10.1158/2159-8290.Cd-16-0596
105. Okada K, Araki M, Sakashita T, et al. Prediction of ALK mutations mediating ALK-TKIs resistance and drug re-purposing to overcome the resistance. *EBioMedicine*. 2019;41:105–119. doi:10.1016/j.ebiom.2019.01.019
106. Shaw AT, Solomon BJ, Besse B, et al. ALK Resistance Mutations and Efficacy of Lorlatinib in Advanced Anaplastic Lymphoma Kinase-Positive Non-Small-Cell Lung Cancer. *J Clin Oncol*. 2019;37(16):1370–1379. doi:10.1200/jco.18.02236
107. Choi YL, Soda M, Yamashita Y, et al. EML4-ALK mutations in lung cancer that confer resistance to ALK inhibitors. *New Engl J Med*. 2010;363(18):1734–1739. doi:10.1056/NEJMoa1007478
108. Lovly CM, Pao W. Escaping ALK inhibition: mechanisms of and strategies to overcome resistance. *Sci, Trans Med*. 2012;4(120):120ps2. doi:10.1126/scitranslmed.3003728
109. Yu Y, Ou Q, Wu X, et al. Concomitant resistance mechanisms to multiple tyrosine kinase inhibitors in ALK-positive non-small cell lung cancer. *Lung Cancer*. 2019;127:19–24. doi:10.1016/j.lungcan.2018.11.024
110. Shaw AT, Friboulet L, Leshchiner I, et al. Resensitization to Crizotinib by the Lorlatinib ALK Resistance Mutation L1198F. *New Engl J Med*. 2016;374(1):54–61. doi:10.1056/NEJMoa1508887
111. Friboulet L, Li N, Katayama R, et al. The ALK inhibitor ceritinib overcomes crizotinib resistance in non-small cell lung cancer. *Cancer Discovery*. 2014;4(6):662–673. doi:10.1158/2159-8290.Cd-13-0846
112. Roskoski R Jr. Anaplastic lymphoma kinase (ALK) inhibitors in the treatment of ALK-driven lung cancers. *Pharmacol Res*. 2017;117:343–356. doi:10.1016/j.phrs.2017.01.007
113. Ni Z, Zhang TC. Computationally unraveling how ceritinib overcomes drug-resistance mutations in ALK-rearranged lung cancer. *Journal of Molecular Modeling*. 2015;21(7):175. doi:10.1007/s00894-015-2716-z
114. Hida T, Seto T, Horinouchi H, et al. Phase II study of ceritinib in alectinib-pretreated patients with anaplastic lymphoma kinase-rearranged metastatic non-small-cell lung cancer in Japan: ASCEND-9. *Cancer Sci*. 2018;109(9):2863–2872. doi:10.1111/cas.13721
115. Camidge DR, Dziadziuszko R, Peters S, et al. Updated Efficacy and Safety Data and Impact of the EML4-ALK Fusion Variant on the Efficacy of Alectinib in Untreated ALK-Positive Advanced Non-Small Cell Lung Cancer in the Global Phase III ALEX Study. *J Thorac Oncol*. 2019;14(7):1233–1243. doi:10.1016/j.jtho.2019.03.007
116. Zhang SS, Nagasaka M, Zhu VW, Ou SI. Going beneath the tip of the iceberg. Identifying and understanding EML4-ALK variants and TP53 mutations to optimize treatment of ALK fusion positive (ALK+) NSCLC. *Lung Cancer*. 2021;158:126–136. doi:10.1016/j.lungcan.2021.06.012
117. Ma Y, Pan H, Liu Y, et al. Ensartinib in advanced ALK-positive non-small cell lung cancer: a multicenter, open-label, two-staged, phase 1 trial. *J Thoracic Dis*. 2022;14(12):4751–4762. doi:10.21037/jtd-22-1606
118. Horn L, Infante JR, Reckamp KL, et al. Ensartinib (X-396) in ALK-Positive Non-Small Cell Lung Cancer: results from a First-in-Human Phase I/II, Multicenter Study. *Clin Cancer Res*. 2018;24(12):2771–2779. doi:10.1158/1078-0432.Ccr-17-2398
119. Yamaguchi N, Lucena-Araujo AR, Nakayama S, et al. Dual ALK and EGFR inhibition targets a mechanism of acquired resistance to the tyrosine kinase inhibitor crizotinib in ALK rearranged lung cancer. *Lung Cancer*. 2014;83(1):37–43. doi:10.1016/j.lungcan.2013.09.019
120. Chen H, Lin C, Peng T, et al. Metformin reduces HGF-induced resistance to alectinib via the inhibition of Gab1. *Cell Death Dis*. 2020;11(2):111. doi:10.1038/s41419-020-2307-5
121. Li L, Wang Y, Peng T, et al. Metformin restores crizotinib sensitivity in crizotinib-resistant human lung cancer cells through inhibition of IGF1-R signaling pathway. *Oncotarget*. 2016;7(23):34442–34452. doi:10.18632/oncotarget.9120

122. Yoshida R, Sasaki T, Minami Y, et al. Activation of Src signaling mediates acquired resistance to ALK inhibition in lung cancer. *Int J Oncol.* 2017;51(5):1533–1540. doi:10.3892/ijo.2017.4140
123. Yun MR, Choi HM, Lee YW, et al. Targeting YAP to overcome acquired resistance to ALK inhibitors in ALK-rearranged lung cancer. *EMBO Mol. Med.* 2019;11(12):e10581. doi:10.15252/emmm.201910581
124. Tsuji T, Ozasa H, Aoki W, et al. Alectinib Resistance in ALK-Rearranged Lung Cancer by Dual Salvage Signaling in a Clinically Paired Resistance Model. *mol Cancer Res.* 2019;17(1):212–224. doi:10.1158/1541-7786.Mcr-18-0325
125. Fukuda K, Takeuchi S, Arai S, et al. Epithelial-to-Mesenchymal Transition Is a Mechanism of ALK Inhibitor Resistance in Lung Cancer Independent of ALK Mutation Status. *Cancer Res.* 2019;79(7):1658–1670. doi:10.1158/0008-5472.Can-18-2052
126. Verdura S, Encinar JA, Teixidor E, et al. Silibinin Overcomes EMT-Driven Lung Cancer Resistance to New-Generation ALK Inhibitors. *Cancers.* 2022;14(24). doi:10.3390/cancers14246101
127. Butler LM, Ferraldeschi R, Armstrong HK, Centenera MM, Workman P. Maximizing the Therapeutic Potential of HSP90 Inhibitors. *mol Cancer Res.* 2015;13(11):1445–1451. doi:10.1158/1541-7786.Mcr-15-0234
128. Jhaveri K, Ochiaia SO, Dunphy MP, et al. Heat shock protein 90 inhibitors in the treatment of cancer: current status and future directions. *Expert Opin Invest Drugs.* 2014;23(5):611–628. doi:10.1517/13543784.2014.902442
129. Rigaud C, Abbou S, Minard-Colin V, et al. Efficacy of nivolumab in a patient with systemic refractory ALK+ anaplastic large cell lymphoma. *Pediatr Blood Cancer.* 2018;65(4). doi:10.1002/pbc.26902
130. Spigel DR, Reynolds C, Waterhouse D, et al. Phase 1/2 Study of the Safety and Tolerability of Nivolumab Plus Crizotinib for the First-Line Treatment of Anaplastic Lymphoma Kinase Translocation - Positive Advanced Non-Small Cell Lung Cancer (CheckMate 370). *J Thorac Oncol.* 2018;13(5):682–688. doi:10.1016/j.jtho.2018.02.022
131. Lin JJ, Schoenfeld AJ, Zhu VW, et al. Efficacy of Platinum/Pemetrexed Combination Chemotherapy in ALK-Positive NSCLC Refractory to Second-Generation ALK Inhibitors. *J Thorac Oncol.* 2020;15(2):258–265. doi:10.1016/j.jtho.2019.10.014
132. Watanabe S, Sakai K, Matsumoto N, et al. Phase II Trial of the Combination of Alectinib with Bevacizumab in Alectinib Refractory ALK-Positive Nonsquamous Non-Small-Cell Lung Cancer (NLCTG1501). *Cancers.* 2022;15(1):204. doi:10.3390/cancers15010204
133. Murray BW, Zhai D, Deng W, et al. TPX-0131, a Potent CNS-penetrant, Next-generation Inhibitor of Wild-type ALK and ALK-resistant Mutations. *mol Cancer Ther.* 2021;20(9):1499–1507. doi:10.1158/1535-7163.Mct-21-0221
134. Ou SI, Nagasaka M, Brazel D, Hou Y, Zhu VW. Will the clinical development of 4th-generation “double mutant active” ALK TKIs (TPX-0131 and NVL-655) change the future treatment paradigm of ALK+ NSCLC?. *Transl Oncol.* 2021;14(11):101191. doi:10.1016/j.tranon.2021.101191
135. Pelish HE, Tangpeerachaikul A, Kohl NE, Porter JR, Shair MD, Horan JC. NUV-655 (NVL-655) is a selective, brain-penetrant ALK inhibitor with antitumor activity against the lorlatinib-resistant G1202R/L1196M compound mutation. *Cancer Res.* 2021;81(13).
136. Gou W, Li Z, Xu X, et al. ZX-29, a novel ALK inhibitor, induces apoptosis via ER stress in ALK rearrangement NSCLC cells and overcomes cell resistance caused by an ALK mutation. *Biochim. Biophys. Acta, Mol. Cell. Res.* 2020;1867(7):118712. doi:10.1016/j.bbamcr.2020.118712
137. Zhou X, Zhang X, Wu Z, et al. The novel ALK inhibitor ZX-29 induces apoptosis through inhibiting ALK and inducing ROS-mediated endoplasmic reticulum stress in Karpas299 cells. *J Biochem Molecular Toxicology.* 2021;35(3):e22666. doi:10.1002/jbt.22666
138. Pulte ED, Norsworthy KJ, Wang Y, et al. FDA Approval Summary: gilteritinib for Relapsed or Refractory Acute Myeloid Leukemia with a FLT3 Mutation. *Clin Cancer Res.* 2021;27(13):3515–3521. doi:10.1158/1078-0432.Ccr-20-4271
139. Mizuta H, Okada K, Araki M, et al. Gilteritinib overcomes lorlatinib resistance in ALK-rearranged cancer. *Nat Commun.* 2021;12(1):1261. doi:10.1038/s41467-021-21396-w
140. Dhillon S. Gilteritinib: first Global Approval. *Drugs.* 2019;79(3):331–339. doi:10.1007/s40265-019-1062-3
141. Mathi GR, Kang CH, Lee HK, et al. Replacing the terminal piperidine in ceritinib with aliphatic amines confers activities against crizotinib-resistant mutants including G1202R. *Eur. J. Med. Chem.* 2017;126:536–549. doi:10.1016/j.ejmech.2016.11.046
142. Chen Y, Wu J, Wang A, et al. Discovery of N-(5-((5-chloro-4-((2-(isopropylsulfonyl)phenyl)amino)pyrimidin-2-yl)amino)-4-methoxy-2-(4-methyl-1,4-diazepan-1-yl)phenyl)acrylamide (CHMFL-ALK/EGFR-050) as a potent ALK/EGFR dual kinase inhibitor capable of overcoming a variety of ALK/EGFR associated drug resistant mutants in NSCLC. *Eur. J. Med. Chem.* 2017;139:674–697. doi:10.1016/j.ejmech.2017.08.035
143. Ott GR, Cheng M, Learn KS, et al. Discovery of Clinical Candidate CEP-37440, a Selective Inhibitor of Focal Adhesion Kinase (FAK) and Anaplastic Lymphoma Kinase (ALK). *J Med Chem.* 2016;59(16):7478–7496. doi:10.1021/acs.jmedchem.6b00487
144. Shi Y, Fang J, Hao X, et al. Safety and activity of WX-0593 (Iruplinkib) in patients with ALK- or ROS1-rearranged advanced non-small cell lung cancer: a phase 1 dose-escalation and dose-expansion trial. *Signal Transduct Targeted Ther.* 2022;7(1):25. doi:10.1038/s41392-021-00841-8
145. Li T, LoRusso P, Maitland ML, et al. First-in-human, open-label dose-escalation and dose-expansion study of the safety, pharmacokinetics, and antitumor effects of an oral ALK inhibitor ASP3026 in patients with advanced solid tumors. *J Hematol Oncol.* 2016;9(1):23. doi:10.1186/s13045-016-0254-5
146. Song P, Zhang X, Yang D, Wang H, Si X, Zhang L. Single-center study to determine the safety and efficacy of CT-707 in Chinese patients with advanced anaplastic lymphoma kinase-rearranged non-small-cell lung cancer. *Thoracic Cancer.* 2020;11(5):1216–1223. doi:10.1111/1759-7714.13376
147. Ma Y, Zhao H, Xue J, et al. First-in-human phase I study of TQ-B3139 (CT-711) in advanced non-small cell lung cancer patients with ALK and ROS1 rearrangements. *Eur J Cancer.* 2022;173:238–249. doi:10.1016/j.ejca.2022.06.037
148. Sun X, Rao Y. PROTACs as Potential Therapeutic Agents for Cancer Drug Resistance. *Biochemistry.* 2020;59(3):240–249. doi:10.1021/acs.biochem.9b00848
149. Zhang C, Han XR, Yang X, et al. Proteolysis Targeting Chimeras (PROTACs) of Anaplastic Lymphoma Kinase (ALK). *Eur. J. Med. Chem.* 2018;151:304–314. doi:10.1016/j.ejmech.2018.03.071
150. Kang CH, Lee DH, Lee CO, Du Ha J, Park CH, Hwang JY. Induced protein degradation of anaplastic lymphoma kinase (ALK) by proteolysis targeting chimera (PROTAC). *Biochem. Biophys. Res. Commun.* 2018;505(2):542–547. doi:10.1016/j.bbrc.2018.09.169
151. Chang GC, Yang TY, Chen KC, et al. ALK variants, PD-L1 expression, and their association with outcomes in ALK-positive NSCLC patients. *Sci Rep.* 2020;10(1):21063. doi:10.1038/s41598-020-78152-1
152. Ota K, Azuma K, Kawahara A, et al. Induction of PD-L1 Expression by the EML4-ALK Oncoprotein and Downstream Signaling Pathways in Non-Small Cell Lung Cancer. *Clin Cancer Res.* 2015;21(17):4014–4021. doi:10.1158/1078-0432.Ccr-15-0016

153. Gettinger S, Politi K. PD-1 Axis Inhibitors in EGFR- and ALK-Driven Lung Cancer: lost Cause?. *Clin Cancer Res.* 2016;22(18):4539–4541. doi:10.1158/1078-0432.Ccr-16-1401
154. Mazieres J, Drilon A, Lusque A, et al. Immune checkpoint inhibitors for patients with advanced lung cancer and oncogenic driver alterations: results from the IMMUNOTARGET registry. *Ann Oncol.* 2019;30(8):1321–1328. doi:10.1093/annonc/mdz167
155. Wang L, Lui VWY. Emerging Roles of ALK in Immunity and Insights for Immunotherapy. *Cancers.* 2020;12(2). doi:10.3390/cancers12020426
156. Budezies J, Kirchner M, Kluck K, et al. Deciphering the immunosuppressive tumor microenvironment in ALK- and EGFR-positive lung adenocarcinoma. *Cancer Immunol Immunother.* 2022;71(2):251–265. doi:10.1007/s00262-021-02981-w
157. Patel M, Jabbour SK, Malhotra J. ALK inhibitors and checkpoint blockade: a cautionary tale of mixing oil with water?. *J Thoracic Dis.* 2018;10 (Suppl 18):S2198–s2201. doi:10.21037/jtd.2018.06.118
158. Reck M, Mok TSK, Nishio M, et al. Atezolizumab plus bevacizumab and chemotherapy in non-small-cell lung cancer (IMpower150): key subgroup analyses of patients with EGFR mutations or baseline liver metastases in a randomised, open-label phase 3 trial. *Lancet Respir Med.* 2019;7(5):387–401. doi:10.1016/s2213-2600(19)30084-0
159. Gaissmaier L, Christopoulos P. Immune Modulation in Lung Cancer: current Concepts and Future Strategies. *Respiration.* 2020;1–27. doi:10.1159/000510385
160. Voena C, Menotti M, Mastini C, et al. Efficacy of a Cancer Vaccine against ALK-Rearranged Lung Tumors. *Cancer Immunol Res.* 2015;3 (12):1333–1343. doi:10.1158/2326-6066.Cir-15-0089
161. Wang Z, Cao YJ. Adoptive Cell Therapy Targeting Neoantigens: a Frontier for Cancer Research. *Front Immunol.* 2020;11:176. doi:10.3389/fimmu.2020.00176
162. Creelan BC, Wang C, Teer JK, et al. Tumor-infiltrating lymphocyte treatment for anti-PD-1-resistant metastatic lung cancer: a phase 1 trial. *Nature Med.* 2021;27(8):1410–1418. doi:10.1038/s41591-021-01462-y
163. Zhou C, Kim SW, Reungwetwattana T, et al. Alectinib versus crizotinib in untreated Asian patients with anaplastic lymphoma kinase-positive non-small-cell lung cancer (ALESIA): a randomised phase 3 study. *Lancet Respir Med.* 2019;7(5):437–446. doi:10.1016/s2213-2600(19)30053-0
164. Horn L, Wang Z, Wu G, et al. Ensartinib vs Crizotinib for Patients With Anaplastic Lymphoma Kinase-Positive Non-Small Cell Lung Cancer: a Randomized Clinical Trial. *JAMA Oncol.* 2021;7(11):1617–1625. doi:10.1001/jamaoncol.2021.3523
165. Garcia Campelo MR, Lin HM, Zhu Y, et al. Health-related quality of life in the randomized phase III trial of brigatinib vs crizotinib in advanced ALK inhibitor-naïve ALK + non-small cell lung cancer (ALTA-1L). *Lung Cancer.* 2021;155:68–77. doi:10.1016/j.lungcan.2021.03.005
166. Solomon BJ, Liu G, Felip E, et al. Lorlatinib Versus Crizotinib in Patients With Advanced ALK-Positive Non-Small Cell Lung Cancer: 5-Year Outcomes From the Phase III CROWN Study. *J Clin Oncol.* 2024;42(29):3400–3409. doi:10.1200/jco.24.00581

International Journal of General Medicine

Publish your work in this journal

The International Journal of General Medicine is an international, peer-reviewed open-access journal that focuses on general and internal medicine, pathogenesis, epidemiology, diagnosis, monitoring and treatment protocols. The journal is characterized by the rapid reporting of reviews, original research and clinical studies across all disease areas. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/international-journal-of-general-medicine-journal>

Dovepress
Taylor & Francis Group