

# RETRACTED ARTICLE: Research on Correlations of miR-196a Expression with Progression and Prognosis of Cutaneous Squamous Cell Carcinoma

Wei Zhang<sup>1</sup>, Xiaowen Yang<sup>2</sup>, Jianping Lv<sup>1</sup>, Shichang An<sup>3</sup>

<sup>1</sup>Department of Cosmetic Plastic Surgery, Weifang People's Hospital, Weifang, Shandong, 261000, People's Republic of China; <sup>2</sup>Department of Stomatology, The People's Hospital of Zhaoyuan City, Yantai, Shandong, 265400, People's Republic of China; <sup>3</sup>Department of Oral and Maxillofacial Surgery, Weifang People's Hospital, Weifang, Shandong, 261000, People's Republic of China

Correspondence: Shichang An, Department of Oral and Maxillofacial Surgery, Weifang People's Hospital, 151 Changwen Street, Weifang, Shandong, 261000, People's Republic of China, Tel +86-0536-8192557, Email anshichang2021@163.com

**Purpose:** This study aimed to investigate the correlation between miR-196a expression and progression and prognosis of cutaneous squamous cell carcinoma (CSCC).

**Patients and Methods:** Tissue samples and corresponding paracarcinoma tissue samples from 117 patients with CSCC were collected. The qRT-PCR analysis was used to detect the expression levels of miR-196a. Kaplan–Meier curve and Cox regression analyses were used to analyze the relationship between miR-196a expression and patients' prognosis. The CCK-8 and transwell assays were used to explore the effects of miR-196a on the abilities of cell proliferation, migration, or invasion.

**Results:** miR-196a expression was significantly up-regulated in CSCC tissue cell lines, compared with adjacent normal tissues or cell lines, respectively. High expression of miR-196a was associated with positive lymph node metastasis, high TNM stages, and a lower five-year survival rate. The expression level of miR-196a was up-regulated and the proliferation, migration or invasion ability of cells were significantly increased according to the CCK-8 and transwell assays.

**Conclusion:** miR-196a is highly expressed in CSCC, thus affecting the occurrence and development of CSCC. More importantly, miR-196a was shown to have potential as a prognostic marker for CSCC.

**Keywords:** miR-196a, progression, prognosis, cutaneous squamous cell carcinoma

## Introduction

Cutaneous squamous cell carcinoma (CSCC), also known as epidermoid carcinoma, is the second most common skin malignancy incidence rate below basal cell carcinoma.<sup>1,2</sup> The malignancy and metastasis rate of CSCC are higher than basal cell carcinoma. The majority of CSCC can be induced by long-term sunlight, such as solar keratosis, burn, posttraumatic scar and chronic inflammatory ulcer.<sup>3,4</sup> In recent years, the incidence rate of CSCC is increasing. It has been reported that patients with a history of CSCC can develop skin tumors of the same tissue type at the same time or again.<sup>5</sup> The risk of recurrence is high and seriously affects human health.<sup>6</sup> The risk of skin cancer is increased in the population with fair skin, old age, more than 30 years of sun exposure, genetic susceptibility factors, and long-term adverse stimulation.<sup>7,8</sup> Surgery is the cornerstone of the treatment of CSCC, and radiotherapy, as well as systemic treatments, are sometimes also implemented.<sup>9</sup> Now, more and more scientists are devoted to finding CSCC-specific indicators to effectively evaluate the prognosis of cancer patients and to provide a new approach for the treatment of CSCC.

MicroRNAs (miRNAs) are a class of small non-coding single-stranded RNA molecules, about 22 nucleotides in length, which are characterized by not being converted into proteins, but by performing their functions in different ways.<sup>10,11</sup> miRNAs are involved in a range of physiological processes, including growth, development, differentiation, and apoptosis. Studies have indicated that about 50% of miRNA are located in tumor gene-related regions on

chromosomes, and are often accompanied by abnormal miRNA expression in tumors.<sup>12,13</sup> The research about miRNA and CSCC showed that many miRNAs were abnormally expressed in CSCC, such as miR-135b, miR-424, and miR-196a with up-regulated expression, while miR-378, miR-145, and miR-140-3p showed a down-regulated trend.<sup>14</sup> But, there has been no clear report on correlations of miR-196a expression with progression and prognosis of CSCC.

In this paper, we studied the expression of miR-196a in CSCC and analyzed the association between miR-196a expression and CSCC prognosis. Meanwhile, the significance of miR-196a in CSCC cell function was further explored.

## Patients and Methods

### Patients and Tissue Samples

The CSCC tumor tissues and corresponding adjacent normal tissues from 117 patients with CSCC admitted to Weifang People's Hospital were collected during March 2011 and March 2015. All collected samples were immediately placed in liquid nitrogen and stored at  $-80^{\circ}\text{C}$  for use. None of the patients in the study received other treatments before Surgery, including chemotherapy, radiation, and immunotherapy. The clinical characteristics of the patients were collected for further study. All patients were required to sign written informed consent. This study was approved by the Medical Ethics Committee of Weifang People's Hospital (20110146) and conducted in accordance with the Declaration of Helsinki. All patients were followed up for 5 years to investigate the significance of miR-196a on the prognosis of patients.

### Cell Culture and Transfection

Four CSCC cell lines (A431, SCC13, HSC-1, and SCL-1) and one human normal skin cell line (HaCaT) were obtained from the Cell Repository, Chinese Academy of Sciences (Shanghai, China). All cell lines were cultured in DMEM (Gibco, Grand Island, NY) containing 10% fetal bovine serum (FBS; Thermo Fisher Scientific, Waltham, USA) and the cultural atmosphere was at  $37^{\circ}\text{C}$  with  $\text{CO}_2$ .

The CSCC cells were seeded in six-well plates. Then miR-196a mimic, miR-196a inhibitor, and the negative controls (NC) were respectively transfected into cells, and refer to the manufacturers' introduction of Lipofectamine 2000 (Thermo Fisher Scientific, Waltham, MA, USA) for detailed procedures of cell transfection.

### RNA Extraction and Quantitative Real-Time PCR

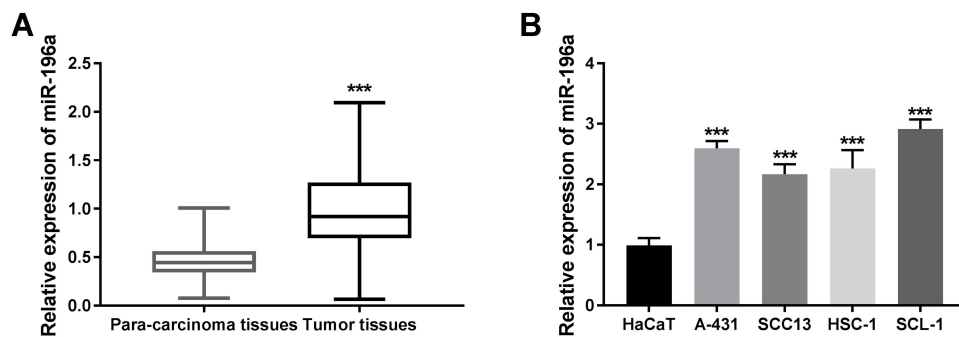
Total RNA was extracted by RNA extraction kit, and the concentration of RNA was determined by NanoDrop 2000 (Thermo Fisher Scientific, Waltham, USA). RNA extraction kit was TRIzol reagent (Thermo Fisher Scientific, Waltham, MA, USA). TaqMan miRNA reverse transcription kit (Thermo Fisher Scientific) was used to reverse-transcribe of total RNA into cDNA. Real-time quantitative PCR (qRT-PCR) was performed using the SYBR-Green I Master mix kit (Invitrogen; Thermo Fisher Scientific, Waltham, USA) on an ABI 7500 Fast Real-Time PCR (Applied Biosystems, Inc., Foster, CA, USA). The relative expression level of miR-196a was calculated by the  $2^{-\Delta\Delta\text{Ct}}$  method and normalized to U6.

### Cell Proliferation Assay

Cell viability was assessed by a cell proliferation assay using Cell Counting Kit-8 (CCK-8, Dojindo Laboratories, Kumamoto, Japan). The cells were inoculated in 96-well plates, mixed gently, placed in  $37^{\circ}\text{C}$ , 5%  $\text{CO}_2$  incubator for conventional culture, and CCK-8 reagent was added at 0, 24 h, 48 h, and 72 h, respectively. Finally, the 96-well plate was placed on an ultraviolet spectrophotometer to detect the OD value at 450 nm.

### Cell Migration and Invasion Assays

The ability of cell invasion and migration was mainly evaluated by Transwell assays using chambers with 8  $\mu\text{m}$  pore polycarbonate filters (Corning, NY, USA). For cell invasion assay, the upper chamber should be precoated with Matrigel (BD, USA), whereas cell migration assay is not required. The other steps in both migration and invasion assays were the same. The density of A431 cells and SCL-1 cells was  $4 \times 10^4$  cells/well. The CSCC cells were scattered in a serum-free medium and placed in the upper chamber. Then 500  $\mu\text{L}$  medium containing 10% FBS was added to the lower chamber to ensure no air bubbles between the lower complete medium and the Transwell chamber. It was incubated in an incubator



**Figure 1** The expression of miR-196a was increased in CSCC tissues and cell lines. **(A)** The expression levels of miR-196a in 117 pairs of CSCC tissue samples and para-carcinoma tissues were detected using qRT-PCR. \*\*\* $P < 0.001$  **(B)** The expression levels of miR-196a in 4 CSCC cell lines were measured using qRT-PCR. \*\*\* $P < 0.001$ .

for 12 h. After being rinsed, the Transwell compartment was wiped with a cotton swab to eliminate un-migrated or un-invaded cells in the upper chamber. The cells migrated or invaded to the lower chamber were fixed with 4% paraformaldehyde, stained with 0.1% crystal violet dye at room temperature, and left overnight for 15 min, then dried upside down and counted under an inverted fluorescence microscope.

## Statistical Analysis

SPSS 22.0 software (IBM, Armonk, NY, USA) was used for data statistics, and GraphPad 5.0 (GraphPad Software, Inc., La Jolla, CA, USA) was used as the mapping tool. The Student's *t*-test was used for comparison between two groups, and a one-way analysis of variance was used for comparison between multiple groups. Mean  $\pm$  SD was used to represent all results. Kaplan–Meier curve and Cox regression analysis were used to investigate the value of miR-196a on the prognosis of CSCC.  $P < 0.05$  was considered statistically significant.

## Results

### Expression of miR-196a in CSCC Tissues and Cell Lines

To reveal the expression level of miR-196a in CSCC, qRT-PCR was used to detect the expression of miR-196a in CSCC and adjacent normal tissues. Figure 1A showed that the expression level of miR-196a in CSCC tissues was higher than that in normal tissues ( $P < 0.001$ ). In the detection of cell lines, it was found that the expression levels of miR-196a in CSCC cell lines (A-431, SCC13, HSC-1, SCL-1) were also significantly higher than that in normal cell lines HaCaT (Figure 1B,  $P < 0.001$ ).

### Relationship Between Clinicopathological Characteristics and miR-196a Expression in CSCC Patients

To analyze the relationship between miR-196a expression and the clinicopathological characteristics of patients, CSCC patients were divided into two groups according to the median expression of miR-196a, the high expression group ( $n = 59$ ) and the low expression group ( $n = 58$ ). As shown in Table 1, the expression level of miR-196a was closely related to lymph node metastasis ( $P = 0.048$ ) and TNM staging ( $P = 0.006$ ), but other clinicopathological characteristics showed no significant correlation with miR-196a ( $P > 0.05$ ).

### Upregulation of miR-196a is Associated with Poor Prognosis in CSCC Patients

The prognostic significance of miR-196a in CSCC was further analyzed through the Kaplan–Meier survival curve and Cox regression analysis. Kaplan–Meier survival curve was used to study the relationship between miR-196a expression level and 5-year survival of patients, the results showed that the 5-year survival rate in the high miR-196a expression group was significantly lower than that in the low miR-196a expression group ( $P = 0.031$ , Figure 2). Cox regression analysis was used to investigate prognostic independent factors, and Table 2 showed that miR-196a (HR = 4.367,  $P =$

**Table I** Association Between miR-196a Expression and Different Clinical Characteristics of Patients with CSCC

Clinical Characteristics	Cases n = 117	miR-196a Expression		P values
		Low (n = 58)	High (n = 59)	
Age				0.521
≤ 65	47	25	22	
> 65	70	33	37	
Sex				0.916
Male	66	33	33	
Female	51	25	26	
Location				0.522
Head, face	66	31	35	
Arm, others	51	27	24	
Tumor size (cm)				0.31
≤ 5	53	29	24	
> 5	64	29	35	
Differentiation				0.039
Well-moderate	50	24	26	
Poor	67	34	33	
Lymph node metastasis				0.048
Negative	83	46	37	
Positive	34	12	22	
TNM stage				0.006
I-II	81	47	34	
III-IV	36	11	25	

0.018), lymph node metastasis (HR = 3.038,  $P = 0.025$ ) and TNM staging (HR = 3.595,  $P = 0.020$ ) were the independent prognostic indicators of CSCC patients.

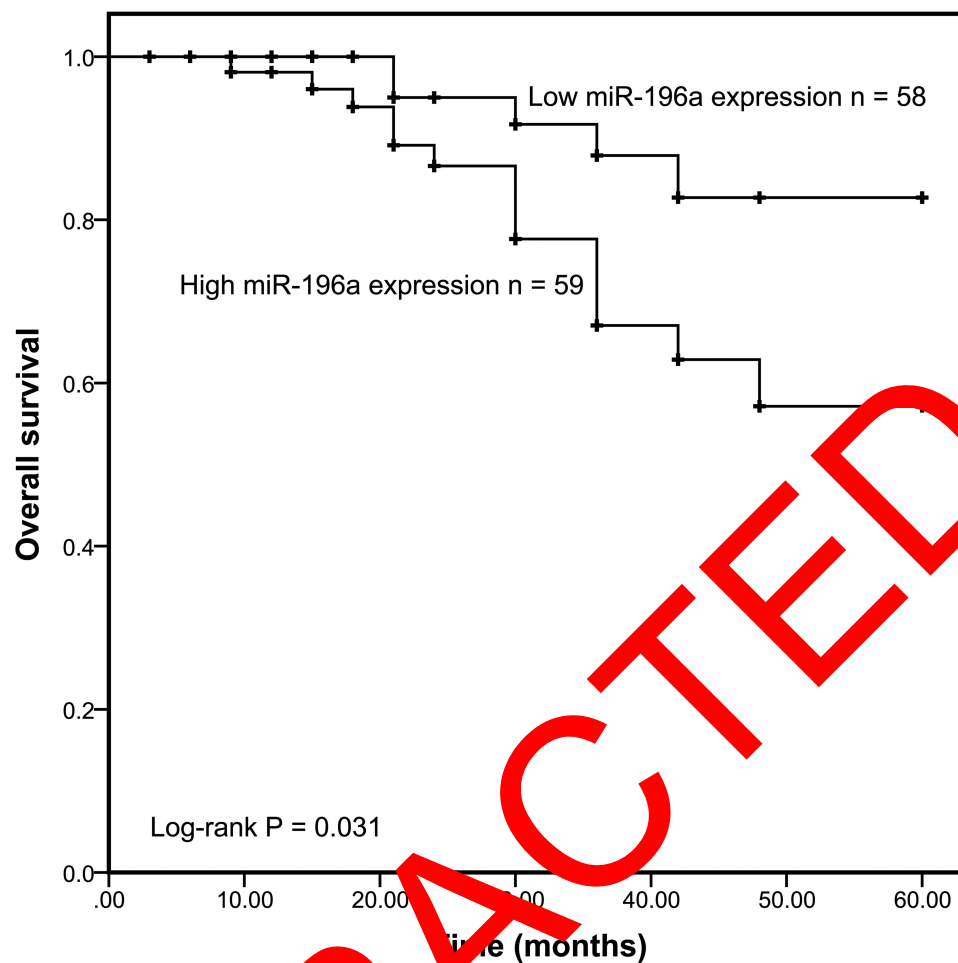
### Upregulation of miR-196a Promotes CSCC Cell Proliferation, Migration, and Invasion

A431 and SCL-1 were transfected with miR-196a mimics or inhibitors to test the tumor cellular functions. For cell proliferation, the results of qRT-PCR showed that miR-196a mimic significantly increased the expression level of miR-196a, but miR-196a inhibitor exerted the opposite effect ( $P < 0.01$ , Figure 3A). Figure 3B shows the experimental results of CCK-8, the up-regulated expression of miR-196a promoted the proliferation of CSCC cells, while down-regulated expression inhibited the proliferation of CSCC cells ( $P < 0.05$ ).

Transwell assays were conducted to study the effects of miR-196a on cell migration and invasion. The results in Figure 4A and B are shown that miR-196a mimic group showed enhanced cell migration and invasion abilities, while the miR-196a inhibitor group showed weakened migration and invasion abilities, compared with control and NC ( $P < 0.001$ ).

### Discussion

Cancer research has always been a hot topic among scientists. At present, the treatment of cancer is mainly surgery, chemotherapy, and radiotherapy, but its prognosis is not satisfactory. miRNA has been proved to play a regulatory role in the growth cycle, apoptosis, tumor invasion and metastasis, and new angiogenesis of tumor cells.<sup>10,15,16</sup> About 50% of miRNA genes are located in tumor gene-related regions on chromosomes and are often accompanied by abnormal miRNA expression in tumors.<sup>17,18</sup> For example, a previous study indicated that miR-139-5p plays a role as a tumor suppressor gene in NSCLC, which is of great significance for the prognosis of NSCLC patients.<sup>19</sup> A study by Tian et al confirmed that microRNA-621 was abnormally low in bladder cancer and inhibited the proliferation and metastasis of cancer cells by inhibiting Wnt/β-catenin signal transduction.<sup>20</sup> For CSCC, it was found that miRNAs play important roles in the pathophysiological process of CSCC development.<sup>21</sup> The abnormal expression of miRNAs in CSCC provides

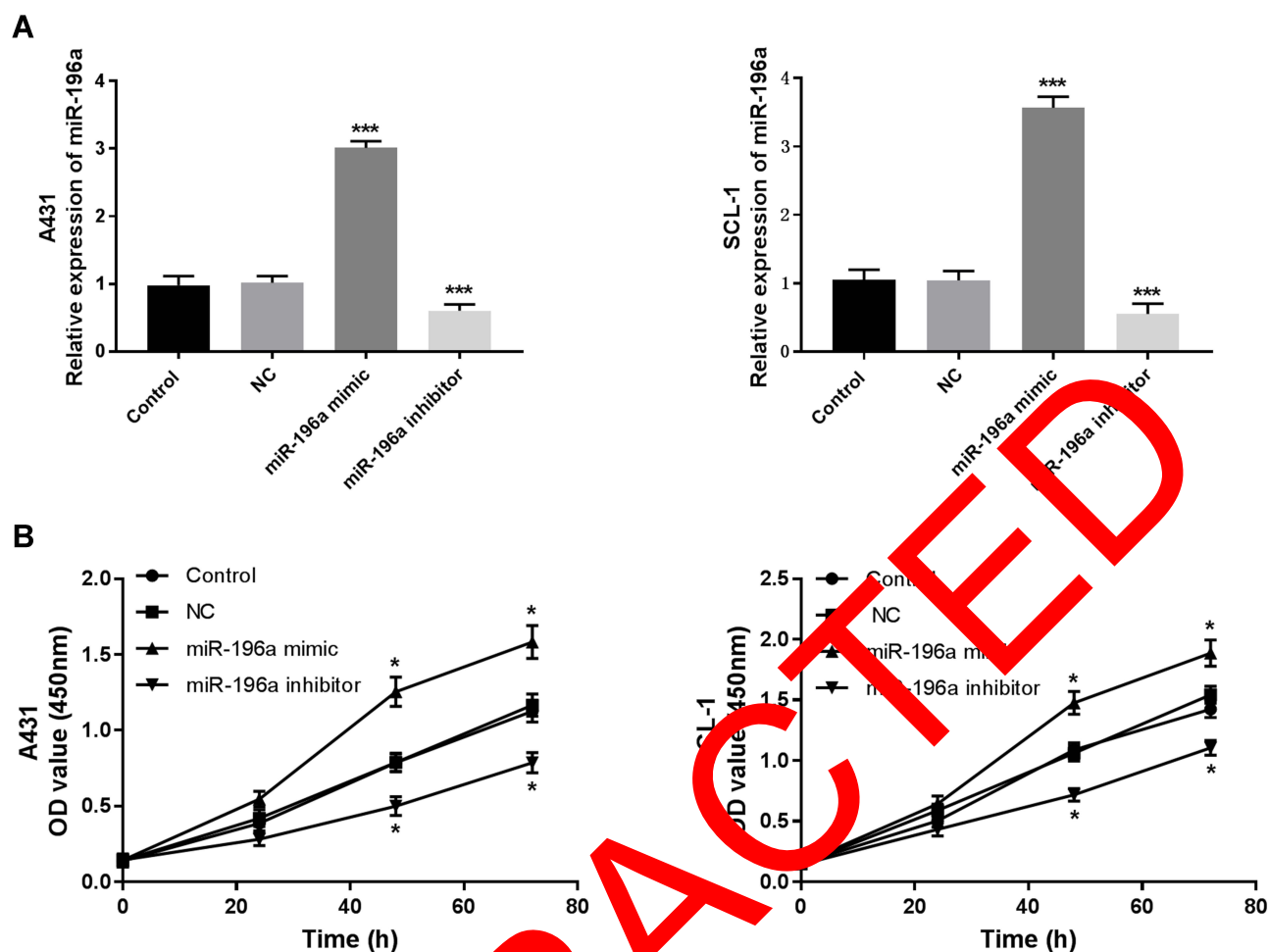


**Figure 2** Kaplan–Meier survival curve in relation to miR-196a expression level in patients with CSCC. Patients with high miR-196a expression exhibited a shorter overall survival time than patients with low miR-196a expression (Log-rank P = 0.031).

a reliable theoretical basis for prognosis.<sup>23</sup> Wang et al found that low-expressed miR-27a promoted tumor growth and metastasis by targeting EGFR, and miR-27a may play an important role in the process of CSCC.<sup>24</sup> miR-196a is an abnormally expressed miRNA that is common in cancer research. He Xin reported abnormal high expression of miR-196a in colorectal cancer.<sup>25</sup> Wang et al found that miR-196a plays a key role in glioma susceptibility.<sup>26</sup> Therefore, it is very necessary to study the role of miR-196a in CSCC.

**Table 2** Multivariate Cox Analysis of Factors for Survival of CSCC Patients

Variables	Multivariate Cox Analysis		
	HR	95% CI	P-value
miR-196a	4.367	1.282–14.874	0.018
Age	2.355	0.841–6.595	0.103
Sex	0.410	0.139–1.210	0.106
Location	0.451	0.162–1.256	0.128
Tumor size	0.672	0.230–1.964	0.468
Differentiation	1.519	0.585–4.327	0.363
Lymph node metastasis	3.038	1.150–8.027	0.025
TNM stage	3.595	1.222–10.582	0.020

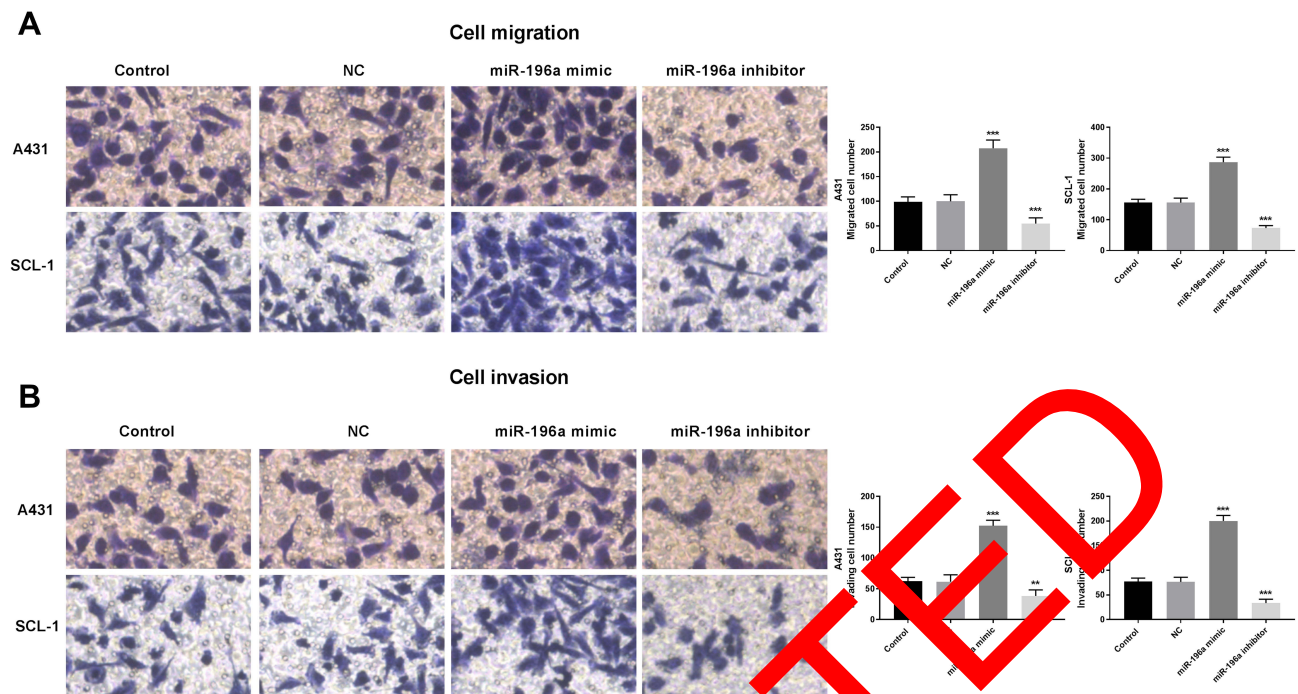


**Figure 3** Effects of miR-196a expression levels on proliferation in A431 and SCL-1 cells. **(A)** the expression level of miR-196a was analyzed by qRT-PCR after transient transfection with miR-196a mimic/inhibitor/ NC. **(B)** The CCK-8 assay was used to study cell proliferation. Increased expression of miR-196a accelerated cell proliferative abilities, while knockdown of miR-196a \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ .

In the current study, it was found that miR-196a was highly expressed in CSCC tissues and CSCC cell lines. The high expression of miR-196a in CSCC tissues was related to positive lymph node metastasis and high TNM stage. The above results suggest that miR-196a expression may be involved in the development of CSCC. Moreover, patients with high miR-196a expression had a worse prognosis than patients with low miR-196a expression. Cox regression analysis results revealed that miR-196a expression, lymph node metastasis, and TNM stage may be independent prognostic risk predictors in CSCC, which implied that miR-196a expression may be a prognostic marker in CSCC. miR-196a is up-regulated in many other cancers. In thyroid carcinoma, miR-196a was also up-regulated, and the prognosis of patients with up-regulated miR-196a was tended poor, miR-196a is an independent prognostic risk factor for thyroid cancer.<sup>27</sup> In gastric cancer, miR-196a is overexpressed and associated with a shorter overall survival time, and miR-196a can be used as a promising new prognostic marker for gastric cancer.<sup>28</sup> Interestingly, miR-196a expression was reduced in melanoma cells compared to healthy melanocytes and mediated the melanoma progression through regulating homeobox B7 (HOXB7) and bone morphogenetic proteins 4 (BMP4) expression.<sup>29</sup> Thus, given the differences in pathogenesis between CSCC and melanoma, miR-196a may play a different role in the tumor progression, which needs to be further investigated in future studies.

The effects of miR-196a on the proliferation, migration, and invasion of CSCC cells were also explored. In this paper, the upregulated cell lines of miR-196a were successfully constructed to further confirm the effect of miR-196a on CSCC cells. The results showed that the proliferation, migration, and invasion ability of A431 and SCL-1





**Figure 4** Effects of miR-196a on cell migration and invasion abilities in A431 and SCL-1 cells. Cell migration and invasion abilities were assessed with Transwell assay. Elevated miR-196a facilitated cell migration and invasion capacities. (magnification 200 ×). \* $P < 0.01$ , \*\*\* $P < 0.001$ .

were significantly increased after the upregulation of miR-196a expression. It suggested that up-regulation of miR-196a could significantly promote the progression of CSCC. The cellular functions of miR-196a on tumors have been widely reported. For example, in epithelial ovarian cancer, miR-196a may play a role as an oncogenic gene, the high expression of miR-196a is closely related to the poor differentiation of patients, and miR-196a can promote the migration and invasion of epithelial ovarian cancer through its downstream target gene HOXA10.<sup>30</sup> In esophageal squamous carcinoma, miR-196a acts as an oncogene and regulates the proliferation, invasion, and migration of esophageal squamous cell carcinoma cells by targeting ANXA1, which is of great significance for the treatment and prognosis of esophageal squamous cell carcinoma.<sup>31</sup> Similar results have been found in osteosarcomas. miR-196a acts as an oncogene in osteosarcoma to promote tumor cellular functions by targeting HOXA5 and is associated with poor prognosis in osteosarcoma patients.<sup>32</sup>

There are still some shortcomings in this study, for example, the sample size is small, which may make the results biased. On the other hand, the expression of miR-196a was measured in tumor tissue samples. In view of the retrospective study, future studies will also take into account blood samples to further support the clinical value of miR-196a in CSCC in a large scale of samples. Also, the downstream target genes of miR-196a have not been discussed yet, which will be completed in future mechanism studies. For these two problems, CSCC specimens meeting the experimental requirements are also being collected continuously, and further studies will be conducted when the sample size is large enough.

## Conclusion

In conclusion, miR-196a was highly expressed in CSCC. The increased expression of miR-196a was closely related to positive lymph node metastasis and high TNM staging in patients with CSCC. The prognosis of CSCC patients with high expression levels of miR-196a was poor. Moreover, miR-196a promoted tumor cell proliferation, migration, and invasion. Therefore, miR-196a may serve as a new therapeutic target for CSCC and provide a new direction for the prognosis of patients.

## Disclosure

The authors report no conflicts of interest in this work.

## References

1. Que SKT, Zwald FO, Schmults CD. Cutaneous squamous cell carcinoma: incidence, risk factors, diagnosis, and staging. *J Am Acad Dermatol*. 2018;78(2):237–247. doi:10.1016/j.jaad.2017.08.059
2. Waldman A, Schmults C. Cutaneous squamous cell carcinoma. *Hematol Oncol Clin North Am*. 2019;33(1):1–12. doi:10.1016/j.hoc.2018.08.001
3. Firnhaber JM. Basal cell and cutaneous squamous cell carcinomas: diagnosis and treatment. *Am Fam Physician*. 2020;102(6):339–346.
4. Prieto-Granada C, Rodriguez-Waitkus P. Cutaneous squamous cell carcinoma and related entities: epidemiology, clinical and histological features, and basic science overview. *Curr Probl Cancer*. 2015;39(4):206–215. doi:10.1016/j.cuprob.2015.07.005
5. Motaparthy K, Kapil JP, Velazquez EF. Cutaneous Squamous Cell Carcinoma: review of the Eighth Edition of the American Joint Committee on Cancer Staging Guidelines, Prognostic Factors, and Histopathologic Variants. *Adv Anat Pathol*. 2017;24(4):171–194. doi:10.1097/PAP.0000000000000157
6. Palazzo E, Morasso MI, Pincelli C. Molecular approach to cutaneous squamous cell carcinoma: from pathways to therapy. *Int J Mol Sci*. 2020;21(4). doi:10.3390/ijms21041211
7. Fu T, Aasi SZ, Hollmig ST. Management of high-risk squamous cell carcinoma of the skin. *Curr Treat Options Oncol*. 2016;17(7):34. doi:10.1007/s11864-016-0408-2
8. Sherwood V, Leigh IM. WNT signaling in cutaneous squamous cell carcinoma: a future treatment strategy? *J Invest Dermatol*. 2016;136(9):1760–1767. doi:10.1016/j.jid.2016.05.108
9. Corchado-Cobos R, García-Sancha N, González-Sarmiento R. Cutaneous squamous cell carcinoma: from biology to therapy. *International Journal of Molecular Sciences*. 2020;21(8). doi:10.3390/ijms21082956
10. Lee YS, Dutta A. MicroRNAs in cancer. *Annu Rev Pathol*. 2009;4:199–227. doi:10.1146/annurev.pathol.4.1107.07.0222
11. Rupaimoole R, Slack FJ. MicroRNA therapeutics: towards a new era for the management of cancer and other diseases. *Nat Rev Drug Discov*. 2017;16(3):203–222. doi:10.1038/nrd.2016.246
12. Lv Y, Huang S. Role of non-coding RNA in pancreatic cancer. *Oncol Lett*. 2019;18(3):963–973. doi:10.3892/ol.2019.10758
13. Sun B, Liu C, Li H, et al. Research progress on the interactions between long non-coding RNAs and microRNAs in human cancer. *Oncol Lett*. 2020;19(1):595–605. doi:10.3892/ol.2019.11182
14. Sand M, Skrygan M, Georgas D, et al. Microarray analysis of microRNA expression in cutaneous squamous cell carcinoma. *J Dermatol Sci*. 2012;68(3):119–126. doi:10.1016/j.jdermsci.2012.09.004
15. Acunzo M, Romano G, Wernicke D, Croce CM. MicroRNA and cancer—a new view. *Adv Biol Regul*. 2015;57:1–9. doi:10.1016/j.jbior.2014.09.013
16. García-Sancha N, Corchado-Cobos R, Pérez-Losada J, Cañueto J. MicroRNA regulation in cutaneous squamous cell carcinoma. *Int J Mol Sci*. 2019;20:9. doi:10.3390/ijms20092181
17. Marmé D. Advances in cancer therapy: liquid biopsy. *Cancer Treat*. 2017;40(7–8):402–403. doi:10.1159/000479186
18. Vishnoi A, Rani S. MiRNA biogenesis and regulation of diseases: an overview. *Methods Mol Biol*. 2017;1509:1–10.
19. Yong-Hao Y, Xian-Guo W, Ming X, Jin-Ping Z. Expression and clinical significance of miR-139-5p in non-small cell lung cancer. *J Int Med Res*. 2019;47(2):867–874. doi:10.1177/030006051881555
20. Tian H, Wang X, Lu J, Tian W, Chen P. MicroRNA-606 inhibits cell proliferation and metastasis in bladder cancer by suppressing Wnt/β-catenin signaling. *Chem Biol Interact*. 2019;300:244–251. doi:10.1016/j.cbi.2019.05.042
21. Li S, Luo C, Zhou J, Zhang Y. MicroRNA-141 directly targets high-mobility group box 1 and inhibits the cancer cell proliferation, migration and invasion in cutaneous squamous cell carcinoma. *Exp Ther Med*. 2017;14(6):5611–5618. doi:10.3892/etm.2017.5245
22. Ma X, Wu D, Zhang X, Shao X, Hu G. microRNA-214 prevents traits of cutaneous squamous cell carcinoma via VEGFA and Bcl-2. *Technol Cancer Res Treat*. 2020;19(5):33033820980098. doi:10.1177/1533033820980098
23. Wang S, Cao KE, He Q, Yin Z, Zou J. miR-199a-5p induces cell invasion by suppressing E-cadherin expression in cutaneous squamous cell carcinoma. *Oncol Lett*. 2016;12(2):97–101. doi:10.3892/ol.2016.4602
24. Wang Y, Deng X, Fan Y, Niu M, Zhou M. miR-27a downregulation promotes cutaneous squamous cell carcinoma progression via targeting EGFR. *Front Oncol*. 2019;9:155. doi:10.3389/fonc.2019.01565
25. Xin H, Wang C, Liu M. miR-196a-2 promotes metastasis of colorectal cancer via targeting IκBα. *BMC Cancer*. 2019;19(1):30. doi:10.1186/s12885-019-5245-1
26. Yang S, Zheng Y, Zhou L, et al. miR-499 rs3746444 and miR-196a-2 rs11614913 are associated with the risk of glioma, but not the prognosis. *Mol Ther Nucleic Acids*. 2020;22:340–351. doi:10.1016/j.omtn.2020.08.038
27. Fu YT, Zhang DQ, Zhou L, et al. Has-MiR-196a-2 is up-regulated and acts as an independent unfavorable prognostic factor in thyroid carcinoma. *Eur Rev Med Pharmacol Sci*. 2018;22(9):2707–2714. doi:10.26355/eurrev\_201805\_14967
28. Mu YP, Tang S, Sun WJ, Gao WM, Wang M, Su XL. Association of miR-193b down-regulation and miR-196a up-regulation with clinicopathological features and prognosis in gastric cancer. *Asian Pac J Cancer Prev*. 2014;15(20):8893–8900. doi:10.7314/APJCP.2014.15.20.8893
29. Braig S, Mueller DW, Rothhammer T, Bosserhoff AK. MicroRNA miR-196a is a central regulator of HOX-B7 and BMP4 expression in malignant melanoma. *Cell Mol Life Sci*. 2010;67(20):3535–3548. doi:10.1007/s00018-010-0394-7
30. Yang B, Li SZ, Ma L, Liu HL, Liu J, Shao JJ. Expression and mechanism of action of miR-196a in epithelial ovarian cancer. *Asian Pac J Trop Med*. 2016;9(11):1105–1110. doi:10.1016/j.apjtm.2016.09.002
31. Hu C, Peng J, Lv L, et al. miR-196a regulates the proliferation, invasion and migration of esophageal squamous carcinoma cells by targeting ANXA1. *Oncol Lett*. 2019;17(6):5201–5209. doi:10.3892/ol.2019.10186
32. Wang X, Zhang L, Zhang X, Xing C, Liu R, Zhang F. MiR-196a promoted cell migration, invasion and the epithelial-mesenchymal transition by targeting HOXA5 in osteosarcoma. *Cancer Biomark*. 2020;29(2):291–298. doi:10.3233/CBM-201674



RETRACTED

Clinical, Cosmetic and Investigational Dermatology

Dovepress

### **Publish your work in this journal**

Clinical, Cosmetic and Investigational Dermatology is an international, peer-reviewed, open access, online journal that focuses on the latest clinical and experimental research in all aspects of skin disease and cosmetic interventions. This journal is indexed on CAS. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/clinical-cosmetic-and-investigational-dermatology-journal>