

Esculetin Alleviates Inflammation, Oxidative Stress and Apoptosis in Intestinal Ischemia/Reperfusion Injury via Targeting SIRT3/AMPK/mTOR Signaling and Regulating Autophagy

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Aim: Intestinal ischemia/reperfusion (I/R) injury is a challenging pathological phenomenon accountable for significant mortality in clinical scenarios. Substantial evidence has supported the protective role of esculetin in myocardial I/R injury. This study is designed to reveal the specific impacts of esculetin on intestinal I/R injury and disclose the underlying mechanism.

Methods: First, intestinal I/R injury model and intestinal epithelial cell line hypoxia/reoxygenation (H/R) model were established. Pathologic damages to intestinal tissues were observed through H&E staining. Serum diamine oxidase (DAO) levels were examined. RT-qPCR and Western blot examined the expression of inflammatory mediators. Commercial kits were used for detecting the levels of oxidative stress markers. TUNEL assay and caspase 3 activity assay measured cell apoptosis. Immunofluorescence (IF) staining measured autophagy levels. Western blot analyzed the expression of apoptosis-, Sirtuin 3 (SIRT3)/AMP activated protein kinase (AMPK)/mammalian target of rapamycin (mTOR) signaling- and autophagy-related proteins. Molecular docking verified the interaction of esculetin with SIRT3. Cell viability was explored via CCK-8 assay.

Results: The experimental results revealed that esculetin treatment mitigated pathological damage of intestinal tissues, reduced serum DAO level, ameliorated inflammation, oxidative stress and apoptosis and promoted autophagy in intestinal I/R rats. Moreover, esculetin bond to SIRT3 and activated SIRT3/AMPK/mTOR signaling both in vitro and in vivo. Furthermore, esculetin treatment enhanced cell viability and SIRT3 silencing reversed the impacts of esculetin on autophagy, inflammation, oxidative stress and apoptosis in H/R cell model.

Conclusion: In a word, esculetin activated SIRT3/AMPK/mTOR signaling and autophagy to protect against inflammation, oxidative stress and apoptosis in intestinal I/R injury.

Keywords: intestinal I/R injury, autophagy, esculetin, SIRT3/AMPK/mTOR signaling

Introduction

Intestine is one of the most susceptible organs to ischemia/reperfusion (I/R) injury and intestinal I/R injury is manifested as intestinal injury caused by restoration of blood supply in the intestine following interruption of intestinal blood flow.¹ Intestinal I/R injury is a frequent tissue organ damage that may occur secondary to intestinal and mesenteric vascular diseases including intestinal obstruction and acute mesenteric ischemia.² Further, intestinal I/R injury is viewed as an essential pathological process of multiple human diseases, including severe infection, trauma and hemorrhagic shock.³ Moreover, intestinal I/R injury is intensively acknowledged to be associated with increased morbidity and mortality as it may progress to multiple organ dysfunction and systemic inflammatory response syndrome.⁴ Thereafter, an in-depth

study of discovering satisfactory treatments and effective drugs to improve the therapeutic effect of intestinal I/R injury is of great necessity.

Esculetin, also named 6,7-dihydroxy-2-chromeone (Figure 1A), is a natural derivative of coumarins available from various plants including *Artemisia capillaris*, *Ceratostigma willmottianum* and *Euphorbia lathyris*.⁵ As reported, esculetin is capable of protecting against coagulation, bacteria, inflammatory response, oxidative stress, tumors and viruses, implying the pleiotropic pharmacological effects of esculetin.⁶ Importantly, previous studies have underlined that esculetin may alleviate I/R injury.^{7,8} Meanwhile, esculetin has also been revealed to suppress intestinal injury.⁹ Hence, whether esculetin also plays the protective role in intestinal I/R injury remains to be elaborated.

Sirtuin 3 (SIRT3), a member of Sirtuins family located in the mitochondrial matrix,¹⁰ has been mentioned to participate in mitochondrial function, energy metabolism, signal regulation and apoptosis.¹¹ Also, there are numerous evidence highlighting the role of SIRT3 in I/R injury.^{12–14} Furthermore, SIRT3 can be activated by esculetin in atherosclerosis and acute coronary artery syndrome.^{15,16} Noticeably, it is reported that SIRT3 can stimulate AMPK and then inactivate mTOR.¹⁷

The goal of this study is to explore the impacts of esculetin on intestinal I/R injury and to discover its potential regulatory mechanism related to SIRT3/AMPK/mTOR signaling.

Materials and Methods

Establishment of Intestinal I/R Injury Model

A total of 30 healthy male Sprague-Dawley rats (weighed 180–220 g) supplied by Beijing Vital River Laboratory Animal Technology Co., Ltd (China) were allowed free access to food and water for 1 week and housed in a specific pathogen-free room with controlled temperature of $24 \pm 2^\circ\text{C}$ with a 12 h light/dark cycle. The rats were randomly assigned to 5 groups (n=6 per group): Control group, Sham group, I/R group, I/R+Esculetin 10 mg/kg group and I/R+Esculetin 25 mg/kg group. Rats in I/R group received 1 h of intestinal ischemia and 2 h of reperfusion described as follows: Rats were intraperitoneally injected with pentobarbital sodium (30 mg/kg of body weight) for anesthesia. A mid-incision was made

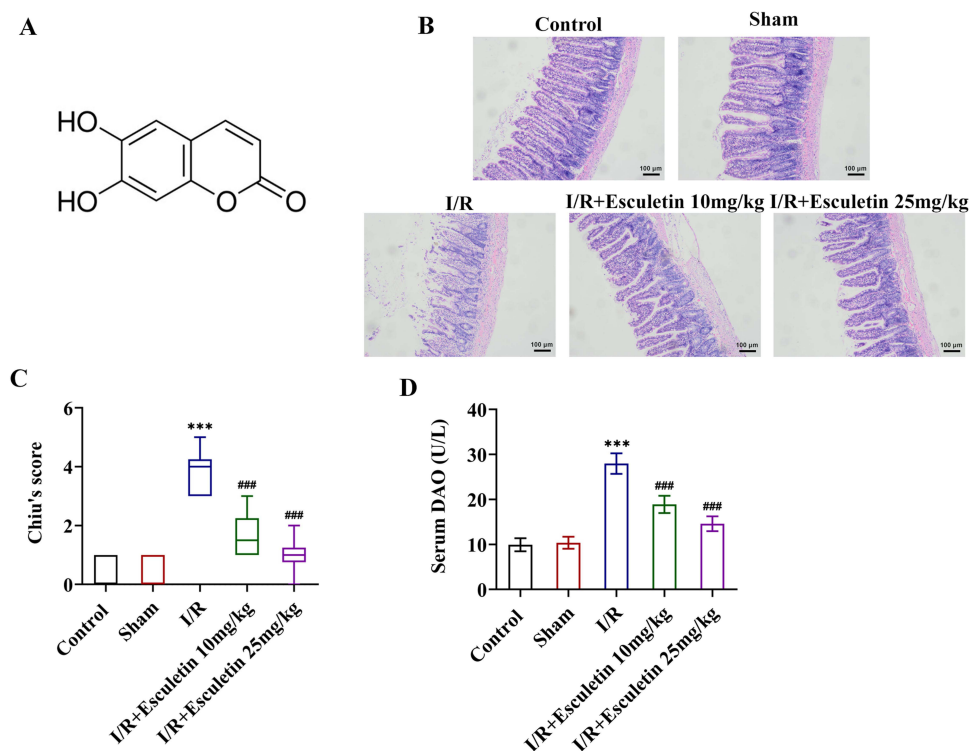


Figure 1 Esculetin alleviates tissue pathological damage and reduces serum DAO levels in intestinal I/R rats. **(A)** The structural formula of esculetin. **(B)** H&E staining and **(C)** Chiu's score assessed the pathological injury of rat intestinal tissues. **(D)** DAO assay kit tested serum DAO levels. *** $p < 0.001$ vs Sham; #### $p < 0.001$ vs I/R.

to expose the superior mesenteric artery, and a noninvasive microvascular clamp was used to occlude the superior mesenteric artery for 1 h for ischemia. Next, the clamp was removed gently for reperfusion. Rats in Sham group underwent the same protocol without vascular occlusion. Rats in I/R+Esculetin 10 mg/kg group were orally administered with esculetin (10 mg/kg; dissolved in the 0.5% sodium carboxymethyl cellulose (CMC-Na) buffer; Sigma-Aldrich, St. Louis, MO, USA) 1 h before surgery. Rats in I/R+Esculetin 25 mg/kg group were orally administered with 25 mg/kg of esculetin 1 h before surgery. Rats in the Control, Sham and I/R groups received the same amount of 0.5% CMC-Na buffer. At the end of the procedures, the rats were sacrificed, and intestinal tissues and blood samples from the abdominal aorta were harvested. All animal experiments were conducted in conformity with the National Institute of Health Guide for the Care and Use of Laboratory Animals,¹⁸ and were approved by the Ethics Committee of Xijing Hospital (Approval number: 20220969).

Hematoxylin and Eosin (H&E) Staining

The rat intestinal tissues fixed by 10% formalin were embedded in paraffin and cut into sections (4 μ m), followed by being stained with hematoxylin solution (Shanghai Enzyme-linked Biotechnology Co., Ltd.) and eosin (Shanghai Enzyme-linked Biotechnology Co., Ltd.). With the aid of a light microscope (Keyence), the images were obtained. Histopathological lesion of intestinal tissues was assessed by the criteria of Chiu's score method.¹⁹

Detection of Serum Diamine Oxidase (DAO) Levels

The serum was harvested following centrifugation of blood samples at 3000 g for 15 min, and DAO levels in the serum were examined with DAO assay kit (cat. no. ml092920; Mlbio, China).

Reverse Transcription-Quantitative PCR (RT-qPCR)

The extraction of total RNA from intestinal tissues and IEC-6 cells was conducted using the Trizol reagent (Beijing Leagene Biotech Co., Ltd.), followed by reverse-transcription from RNA to cDNA according to the guidelines of TRUEscript 1st Strand cDNA Synthesis Kit (CODONX, Beijing, China). Amplification of cDNA was performed via PCR using the SYBR Green qPCR Mix (CODONX, Beijing, China). PCR primer sequences used in this study were listed as follows: TNF- α , forward, 5'-ACTGAACTTCGGGGTGATTG-3' and reverse, 5'-GCT TGGTGGTTTGCTACGAC-3'; IL-1 β , forward, 5'-CAGCTTTCGACAGTGAGGAGAA-3' and reverse, 5'-TC TTGTCGAGATGCTGCTGT-3'; IL-6, forward, 5'-TGATGGATGCTTCCAAACTG-3' and reverse, 5'-GA GCATTGGAAGTTGGGGTA-3'; SIRT3, forward, 5'-TGCACGGTCTGTCTGAAGGTC-3' and reverse, 5'-TG TCAGGTTTCACAACGCCAG-3'; GAPDH, forward, 5'-CTCTACCCACGGCAAGTTC-3' and reverse, 5'-GC CAGTAGACTCCACGACATA-3'. The mRNA levels of above genes were calculated with the $2^{-\Delta\Delta C_t}$ method,²⁰ and GAPDH served as the internal control.

Western Blot

The total protein was extracted from rat intestinal tissues or IEC-6 cells adopting RIPA buffer (Mlbio, China) and then subjected to 10% SDS-PAGE for separation, followed by transferring to PVDF membranes. Thereafter, the membranes were blocked with 5% non-fat milk for 2 h and then immunoblotted with primary antibodies overnight at 4°C and goat anti-rabbit HRP antibody (cat. no. ab205718; 1/2000; Abcam) for 1 h at room temperature. The blots were visualized by the ECL reagent (Mlbio, China) and the gray analysis was implemented with ImageLab4.0 software. Inducible nitric oxide synthase (iNOS; cat. no. ab178945; 1/1000; Abcam), cyclooxygenase-2 (COX2; cat. no. ab179800; 1/1000; Abcam), cleaved caspase 3 (cat. no. #9661; 1/1000; Cell Signaling Technology), caspase 3 (cat. no. ab184787; 1/2000; Abcam), cleaved PARP1 (cat. no. ab32064; 1/1000; Abcam), PARP1 (cat. no. ab191217; 1/1000; Abcam), LC3B (cat. no. ab192890; 1/2000; Abcam), Beclin1 (cat. no. ab207612; 1/2000; Abcam), SIRT3 (cat. no. ab189860; 1/1000; Abcam), p-AMPK (cat. no. ab133448; 1/1000; Abcam), AMPK (cat. no. ab207442; 1/1000; Abcam), p-mTOR (cat. no. #5536; 1/1000; Cell Signaling Technology), mTOR (cat. no. ab134903; 1/10,000; Abcam) and GAPDH (cat. no. ab181602; 1/10,000; Abcam) primary antibodies were utilized in this study.

Evaluation of Superoxide Dismutase (SOD) and Malondialdehyde (MDA)

The SOD activity (cat. no. ml092620) and MDA content (cat. no. ml094962) were examined with corresponding kits (Mlbio, China) in the light of the manufacturer's guidance. Absorbance value at 450 nm was determined using a microplate reader (BioTek Instruments, Inc.).

Terminal-Deoxynucleotidyl Transferase Mediated Nick End Labeling (TUNEL)

The apoptosis of intestinal tissues was examined by TUNEL assay. The sections (5 μ m) were prepared from paraformaldehyde-fixed and paraffin-embedded rat intestinal tissues. After deparaffinization and hydration, the sections were digested with Proteinase K for 15 min, followed by incubation with 50 μ L TUNEL reaction mixture (Beyotime, Shanghai, China) for 1 h. Finally, the images were photographed under a fluorescence microscope (Olympus, Tokyo, Japan).

Caspase 3 Activity Assay

In the light of the manufacturer's guidance, caspase 3 activity was determined using a commercial Caspase 3 Activity kit (cat. no. C1115; Beyotime). Absorbance value at 450 nm was determined using a microplate reader (BioTek Instruments, Inc.).

Immunofluorescence (IF) Staining

After deparaffinization and hydration, the sections of rat intestinal tissues were treated with 3% hydrogen peroxide and then blocked with 10% normal goat serum in PBS for 1 h. After washing, the sections were incubated with the primary antibody against LC3B (cat. no. ab192890; Abcam) overnight at 4°C. On the second day, Alexa Fluor[®] 488 goat anti-rabbit IgG antibody (cat. no. ab150081; Abcam) was added and incubated for another 1 h at room temperature. Nuclear DNA was labeled with DAPI. Images were observed by a fluorescence microscope (Olympus).

Molecular Docking

The crystal structure of SIRT3 protein was retrieved from the Protein Data Bank (<https://www.rcsb.org/>). The three-dimensional structure of esculetin was obtained in SDF format from the PubChem database (<https://pubchem.ncbi.nlm.nih.gov>). The molecular docking between esculetin and SIRT3 was performed using Autodock software (The Scripps Research Institute, La Jolla, CA, USA) and the interaction models were obtained by evaluating the best binding energies. The conformation was visualized utilizing PyMoL software.

Cell Culture and Hypoxia/Reoxygenation (H/R) Model

Rat intestinal epithelial IEC-6 cells from American Type Culture Collection (ATCC, Manassas, VA) were kept in Dulbecco's modified Eagle's medium (DMEM; Biochrom AG, Berlin, Germany) supplemented with 10% fetal bovine serum (FBS; Biochrom AG) and incubated at 37°C with 5% CO₂. For H/R induction, IEC-6 cells were cultured under hypoxic conditions (5% CO₂ and 95% N₂) at 37°C for 6 h, after which the cells were transferred to normal oxygen conditions (95% air and 5% CO₂) in a humidified incubator at 37°C for 6 h.²¹ To measure the impacts of esculetin on H/R cell model, cells were pre-treated by 5, 10 and 20 μ M of esculetin prior to H/R induction.⁷

Cell Counting Kit-8 (CCK-8)

Cell activity was assessed by CCK-8 assay. IEC-6 cells were placed into 96-well plates (5 $\times$ 10³ cells/well). After the indicated treatment, 10 μ L CCK-8 solution (Mlbio, China) was added to each well and the plates were incubated at 37°C with 5% CO₂ for extra 2 h. Cell activity was assessed through detecting absorbance value at 450 nm with the application of a microplate reader (BioTek Instruments, Inc.).

Plasmid Transfection

The specific shRNAs targeting SIRT3 (ShRNA-SIRT3-1/2) and the negative control (ShRNA-NC) were synthesized by GenePharma (Shanghai, China). IEC-6 cells were subjected to plasmid transfection adopting Lipofectamine 3000 (Invitrogen) as per the manufacturer's recommendation. Subsequent experiments were carried out 48 h post-transfection.

Statistical Analyses

GraphPad Prism 8 software (GraphPad Software, Inc.) was utilized to carry out statistical analyses. All data were presented as the mean $\pm$ standard deviation. Statistical significances were measured by using of one-way ANOVA along with Tukey's post hoc test. The significance level was $P < 0.05$.

Results

Esculetin Alleviates Tissue Pathological Damage and Reduces Serum DAO Levels in Intestinal I/R Rats

To uncover the effects of esculetin on intestinal I/R injury in vivo, the pathological changes of rat intestinal tissues were observed by H&E staining. As displayed in [Figure 1B](#) and [C](#), no apparent alternations were observed in the morphology of rat intestinal tissues in the Control group and Sham group. Swelling, shortening, and loss of intestinal villi and the increased Chiu's score were noticed in the I/R group, suggesting the severe pathological damage of rat intestinal tissues following I/R. After administration of 10 or 25 mg/kg of esculetin, the rat intestinal I/R injury was mitigated and Chiu's score was prominently declined. Also, relative to the Sham group, serum DAO levels were significantly elevated in I/R group and esculetin treatment markedly decreased serum DAO levels ([Figure 1D](#)). Taken together, esculetin might play the protective role in I/R-induced intestinal injury.

Esculetin Ameliorates Inflammatory Response, Oxidative Stress and Apoptosis in Intestinal I/R Rats

Further, through RT-qPCR and Western blot, it was noted that I/R treatment remarkably raised the expression of inflammatory factors TNF- α , IL-1 β , IL-6, iNOS and COX2 by contrast with Sham group and esculetin dose-dependently cut down TNF- α , IL-1 β , IL-6, iNOS and COX2 expression ([Figure 2A](#) and [B](#)). Additionally, relative to the Sham group, MDA level was elevated, whereas SOD level was lessened in I/R group. Treatment with esculetin conversely declined MDA level and augmented SOD level in I/R group in a concentration-dependent manner ([Figure 2C](#) and [D](#)). Besides, the results from TUNEL assay elucidated that the apoptotic rate was strengthened in the I/R group, accompanied by enhanced caspase 3 activity and upregulated cleaved caspase 3/caspase 3 and cleaved PARP1/PARP1 expression in contrast with Sham group. Following esculetin administration, the apoptotic rate was attenuated, and Caspase 3 activity, caspase 3/caspase 3 and cleaved PARP1/PARP1 expression were declined ([Figure 3A–C](#)). To conclude, esculetin suppressed inflammatory response, oxidative stress and apoptosis in intestinal I/R rat model.

Esculetin Activates Autophagy and SIRT3/AMPK/mTOR Signaling in Intestinal I/R Rats

Importantly, Western blot analysis revealed that SIRT3 and p-AMPK/AMPK were downregulated while p-mTOR/mTOR protein level was upregulated following I/R injury, which was partly hindered by esculetin administration ([Figure 4A](#)). Through IF assay, it was discovered that the autophagy level decreased in I/R group was elevated by esculetin administration ([Figure 4B](#)). This finding was also further validated by the results that the LC3II/I and Beclin1 expression was lowered following I/R injury while was conversely elevated under esculetin administration ([Figure 4C](#)). Overall, esculetin stimulated SIRT3/AMPK/mTOR signaling and autophagy in intestinal I/R injury in vivo.

Esculetin Regulates Autophagy in H/R Cell Model via Activating SIRT3/AMPK/mTOR Signaling

Via molecular docking, the interaction between esculetin and SIRT3 was confirmed ([Figure 5A](#)). As CCK-8 assay depicted, various concentrations of esculetin elicited no obvious effects on the viability of IEC-6 cells ([Figure 5B](#)).

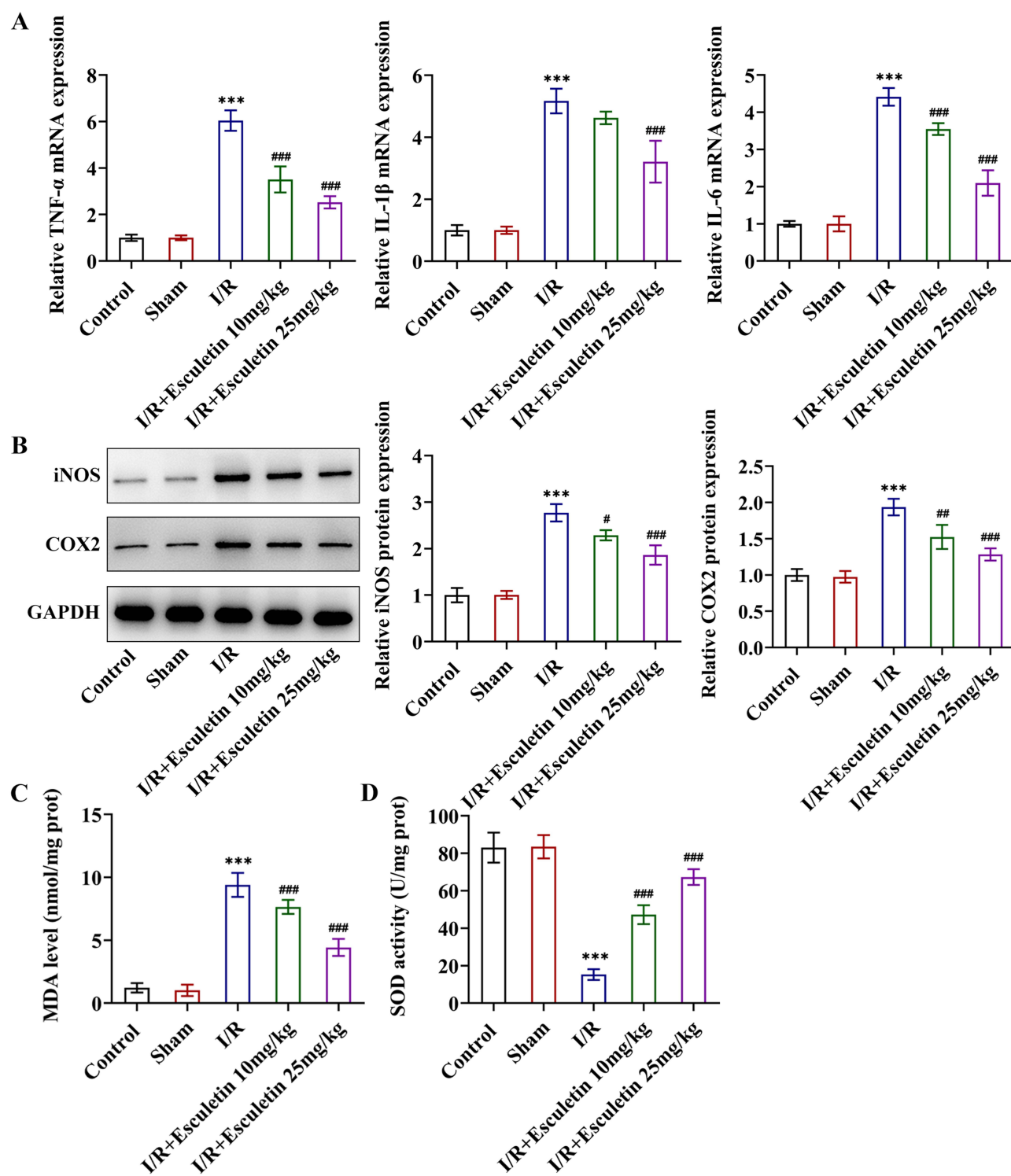


Figure 2 Esculetin ameliorates inflammatory response and oxidative stress in intestinal I/R rats. **(A)** RT-qPCR examined the mRNA levels of TNF- α , IL-1 β and IL-6. **(B)** Western blot analysis examined the protein expression of iNOS and COX2. Related kits examined **(C)** MDA content and **(D)** SOD activity. *** $p < 0.001$ vs Sham; # $p < 0.05$, ### $p < 0.001$ vs I/R.

Moreover, the suppressed IEC-6 cell viability in H/R cell model was restored by esculetin in a concentration-dependent manner (Figure 5C). In agreement with the in vivo results, SIRT3 and p-AMPK/AMPK expressions were downregulated, while p-mTOR/mTOR expression was up-regulated in H/R-exposed IEC-6 cells, which were then partly reversed by esculetin treatment dose-dependently (Figure 5D).

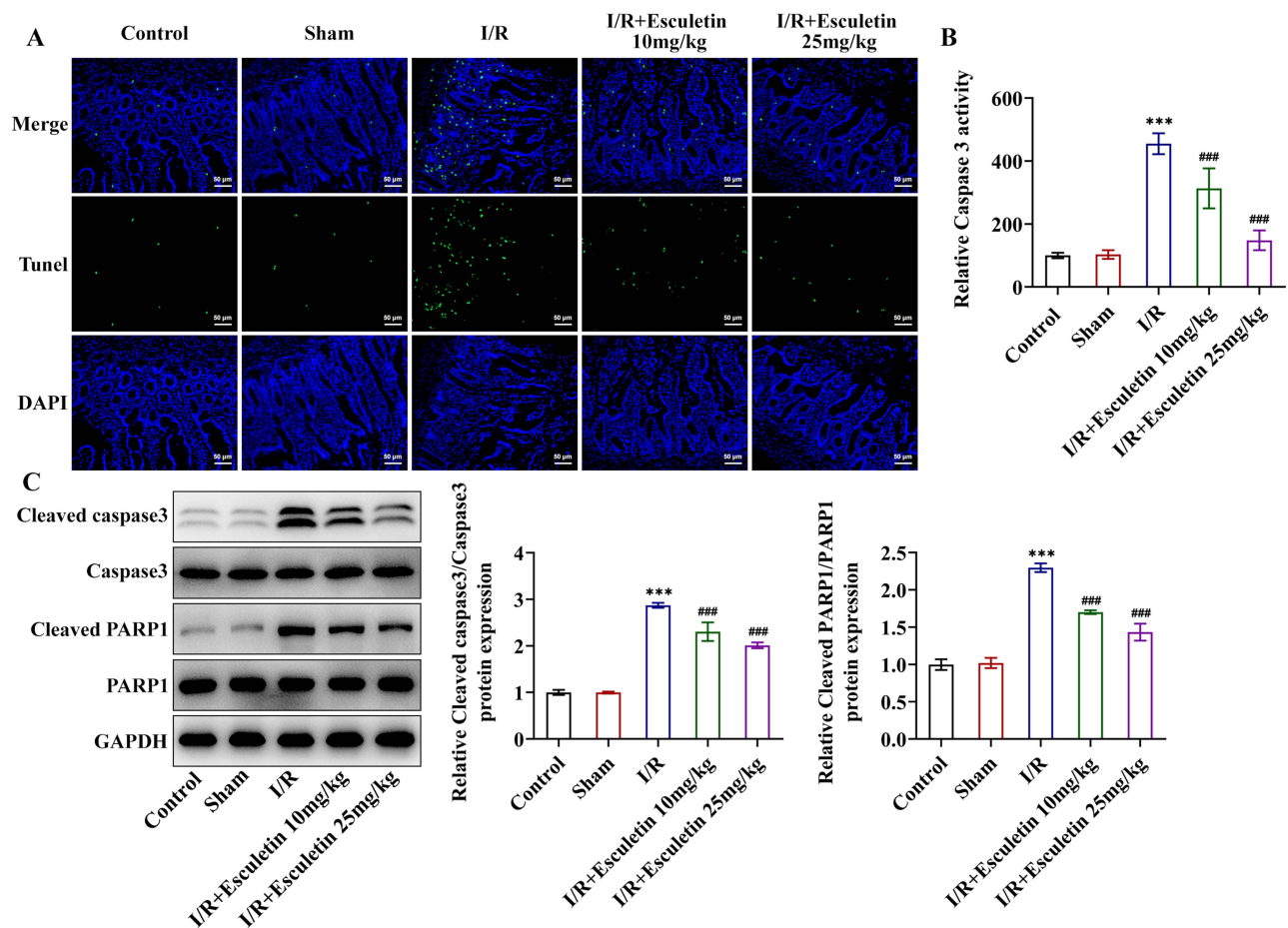


Figure 3 Esculetin reduces apoptosis in intestinal I/R rats. (A) TUNEL assay measured apoptosis. (B) Caspase 3 activity assay examined caspase 3 activity. (C) Western blot analysis of the expression of apoptosis-associated proteins. *** $p < 0.001$ vs Sham, #### $p < 0.001$ vs I/R.

To validate the involvement of SIRT3 in esculetin-mediated H/R cell injury, SIRT3 was knocked down by transfection of ShRNA-SIRT3-1/2 plasmids. Moreover, ShRNA-SIRT3-1 was chosen for the ensuing experiments as it exhibited a more remarkable interference efficacy than ShRNA-SIRT3-2 (Figure 5E). It was observed from IF assay that esculetin enhanced the autophagy level in H/R cell model, which was partly hindered by further depletion of SIRT3 (Figure 5F). As expected, LC3II/I and Beclin1 expression were diminished in IEC-6 cells exposed to H/R induction, and this result was reversed by esculetin. Further, the effect of esculetin was abolished by SIRT3 knockdown (Figure 5G). Collectively, esculetin activated SIRT3/AMPK/mTOR signaling to modulate autophagy in H/R cell model.

Esculetin Stimulates SIRT3/AMPK/mTOR Signaling to Relieve Inflammation, Oxidative Stress and Apoptosis in H/R Cell Model

Furthermore, esculetin administration noticeably lowered H/R exposure-induced TNF- α , IL-1 β , IL-6, iNOS and COX2 expression and SIRT3 deficiency improved TNF- α , IL-1 β , IL-6, iNOS and COX2 expression again (Figure 6A–C). Besides, esculetin decreased MDA level, whereas raised SOD level in H/R cell model and SIRT3 conversely elevated MDA level and declined SOD level (Figure 6D and E). At the same time, esculetin obstructed the apoptosis of IEC-6 cells exposed to H/R and this effect was abrogated by SIRT3 knockdown (Figure 6F). The inhibitor effects of esculetin treatment on Caspase 3 activity, cleaved caspase 3/Caspase 3 and cleaved PARP1/PARP1 expression in H/R-induced IEC-6 cells were also weakened after SIRT3 knockdown (Figure 6G and H). All these results pointed out that esculetin protected against H/R-induced inflammation, oxidative stress and apoptosis in IEC-6 cells via activating SIRT3.

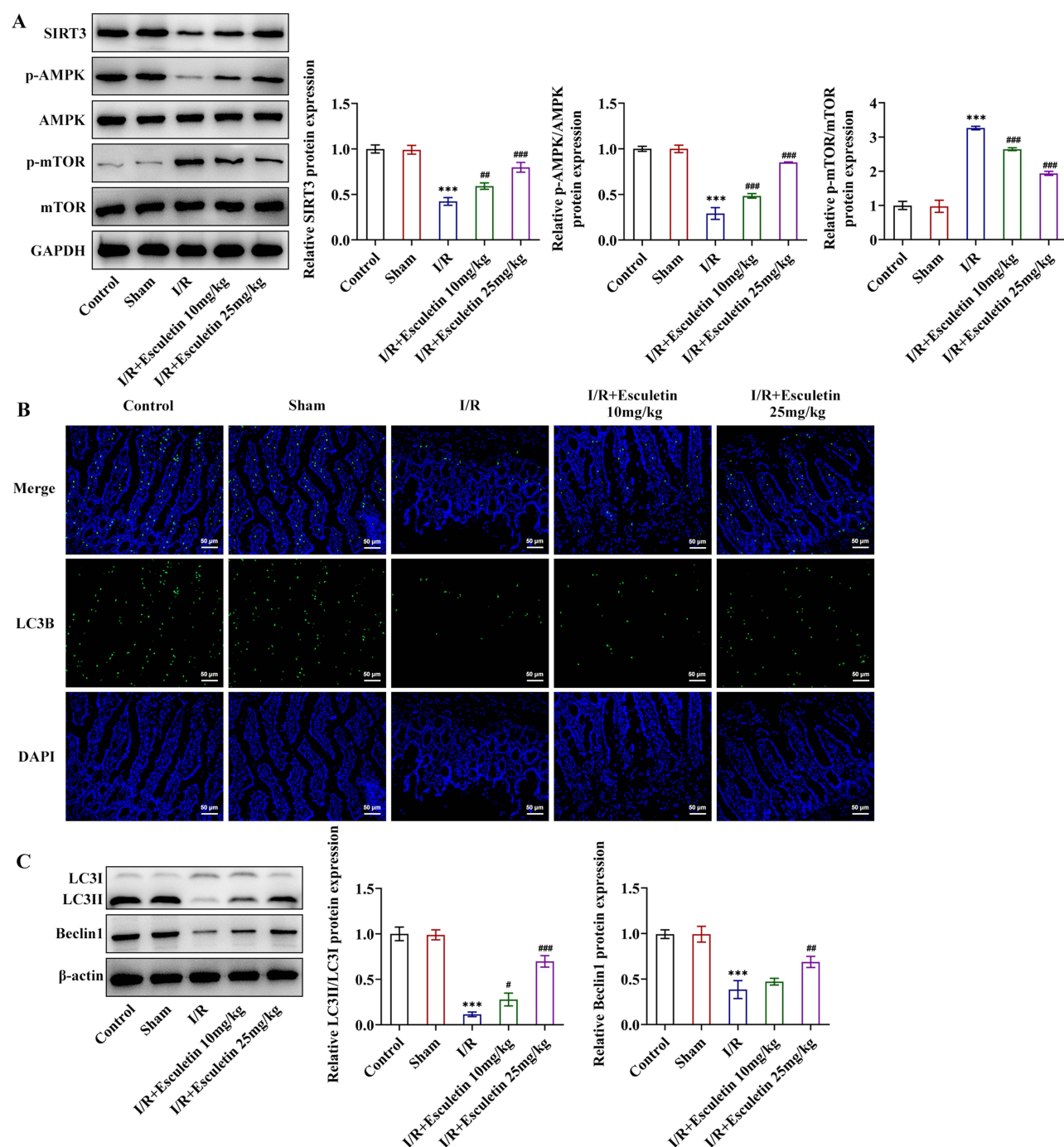


Figure 4 Esculetin activates autophagy and SIRT3/AMPK/mTOR signaling in intestinal I/R rats. **(A)** Western blot analysis of the expression of SIRT3/AMPK/mTOR signaling-associated proteins. **(B)** IF assay evaluated autophagy marker LC3B. **(C)** Western blot analysis of the expression of autophagy-associated proteins. *** $p < 0.001$ vs Sham; # $p < 0.05$, ## $p < 0.01$, ### $p < 0.001$ vs I/R.

Discussion

Intestinal I/R injury is a complex and grave disorder, the pathological process of which may involve extensive fields such as inflammatory response, apoptosis, autophagy, oxidative stress and so on.^{22–24} DAO is an intracellular enzyme existing in the cytoplasm of upper villus cells of the intestinal mucosa.²⁵ Serum DAO level may indirectly reflect the permeability and barrier function of intestinal mucosa and the integrity of intestinal epithelial cells lining mucosal surfaces as DAO may be released into blood when intestinal villus is damaged.²⁵ Consistently, in the present study, severe pathological

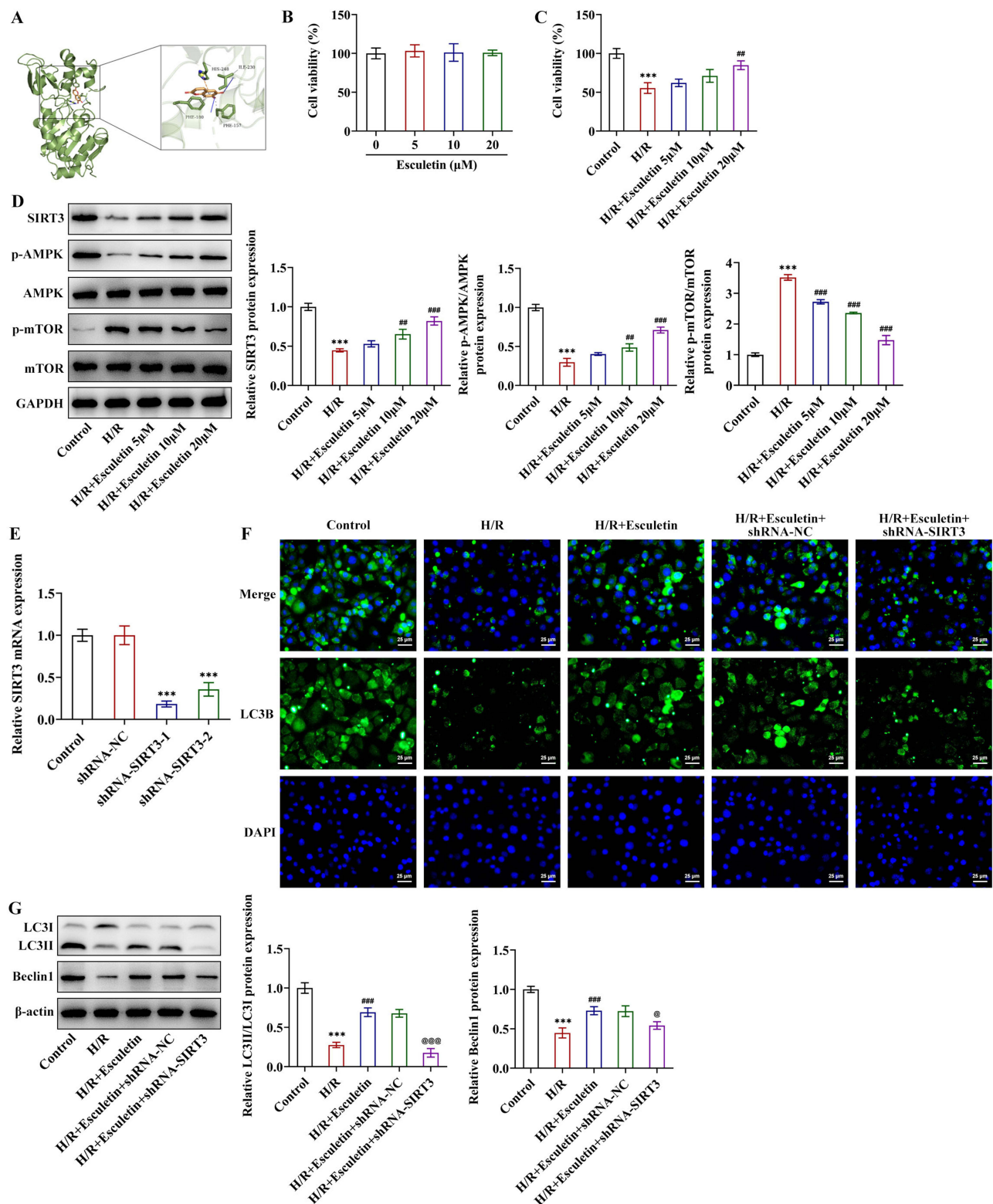


Figure 5 Esculetin regulates autophagy in H/R cell model via activating SIRT3/AMPK/mTOR signaling. **(A)** Molecular docking confirmed the binding of esculetin with SIRT3. **(B)** IEC-6 cells were treated with different concentrations of esculetin, and CCK-8 assay assessed cell viability. **(C)** IEC-6 cells were pre-treated with 5 μM, 10 μM and 20 μM of esculetin prior to H/R induction, and CCK-8 assay assessed cell viability. **(D)** Western blot analysis of the expression of SIRT3/AMPK/mTOR signaling-associated proteins. *** $p < 0.001$ vs Control; ## $p < 0.01$, ### $p < 0.001$ vs H/R. **(E)** RT-qPCR analysis of the mRNA level of SIRT3 to assess the interference efficacy of ShRNA-SIRT3-1/2 plasmids. *** $p < 0.001$ vs shRNA-NC. **(F)** IF assay evaluated the autophagy marker LC3B. **(G)** Western blot analysis of the expression of autophagy-associated proteins. *** $p < 0.001$ vs Control; ### $p < 0.001$ vs H/R; @ $p < 0.05$, @@@ $p < 0.001$ vs H/R+Esculetin+shRNA-NC.

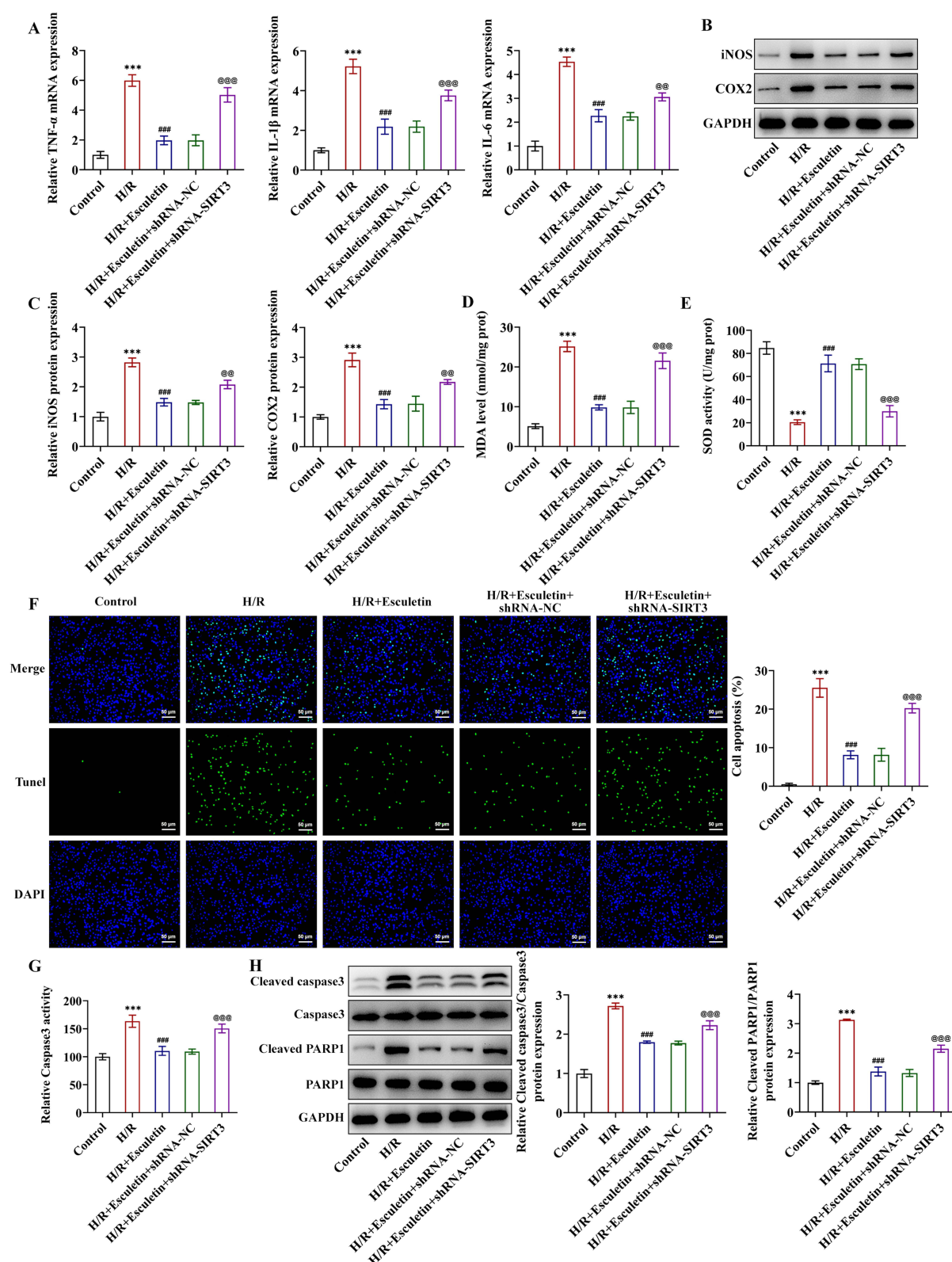


Figure 6 Escluletin activates SIRT3/AMPK/mTOR signaling to relieve inflammation, oxidative stress and apoptosis in H/R cell model. **(A)** RT-qPCR examined the mRNA levels of TNF- α , IL-1 β and IL-6. **(B)** and **(C)** Western blot analysis examined the protein expression of iNOS and COX2. Related kits examined **(D)** MDA content and **(E)** SOD activity. **(F)** TUNEL assay measured apoptosis. **(G)** Caspase 3 activity assay examined caspase 3 activity. **(H)** Western blot analysis of the expression of apoptosis-associated proteins. *** $p < 0.001$ vs Control; ### $p < 0.001$ vs H/R; @@ $p < 0.01$, @@@ $p < 0.001$ vs H/R+Escluletin+shRNA-NC.

damage of rat intestinal tissues, enhanced Chiu's score and serum DAO levels were observed following I/R injury. Previous study has introduced a variety of pharmacological properties of esculetin, a natural dihydroxy coumarin.²⁶ In this study, esculetin was discovered to dose-dependently reduce pathological injury of intestinal tissues and decrease Chiu's score and serum DAO levels in intestinal I/R rat model. Hence, esculetin might play the protective role in I/R-induced intestinal injury.

Intestine can release a great number of pro-inflammatory mediators and induce inflammatory response following I/R injury.²⁷ TNF- α is an initiating factor of inflammatory response and a leading cause of inflammatory response-induced intestinal injury.²⁸ IL-1 β and IL-6 are also inflammatory factors which are produced secondary to endothelial injury and may damage intestinal mucosa and intestinal functions.²⁹ As reported, up-regulated iNOS and COX2 expression are closely associated with inflammatory response and even intestinal I/R injury.³⁰ As expected, we found that the production of TNF- α , IL-1 β , IL-6 and the expression of iNOS and COX2 were greatly increased following I/R treatment, demonstrating that I/R triggered inflammatory response in intestinal tissues. Anyway, this inflammatory response was remarkably restricted by esculetin in a concentration-dependent manner. Additionally, it is commonly acknowledged that the overproduction of oxidative stress marker MDA or absence of antioxidant SOD may contribute to acute lung injury and experimental acute liver lesion elicited by intestinal I/R injury.^{31,32} Intestinal epithelial cell apoptosis is also deemed as a pivotal mechanism leading to loss of intestinal barrier function stimulated by intestinal I/R injury.¹³ Here, the diminished SOD activity and the elevated MDA level and apoptotic rate confirmed the occurrence of oxidative stress and cell apoptosis in intestinal tissues following I/R injury. These changes were partly abolished by esculetin treatment, demonstrating that esculetin ameliorated oxidative stress and apoptosis in intestinal I/R rats, in line with the previous report that esculetin was able to protect against oxidative stress and apoptosis to alleviate myocardial I/R injury.⁷

Autophagy is a degradation process of cellular components mediated by lysosomes.³³ Recent research has exposed that autophagy has been widely recognized to be implicated in intestinal I/R injury and activation of autophagy can ameliorate intestinal I/R injury.^{34,35} In this study, the autophagy level was decreased in the intestinal tissues of intestinal I/R rat model, while esculetin treatment could promote the autophagy level, partly accounting for the protective role of esculetin against intestinal I/R injury. SIRT3 is a pivotal deacetylase mainly modulating mitochondrial action and ROS generation.³⁶ Further research has proposed that SIRT3 can stimulate AMPK signaling to inhibit mTOR and regulate autophagy.¹⁷ Accumulating evidence has revealed that SIRT3 may act as a suppressor in I/R injury, including intestinal I/R injury.^{13,14,37} Further, SIRT3 can be activated by esculetin in atherosclerosis.¹⁵ Consistent with these findings, the interaction between esculetin and SIRT3 was also verified by molecular docking in this study. Meanwhile, esculetin promoted the activation of SIRT3/AMPK/mTOR signaling in both I/R-caused intestinal injury and H/R-induced IEC-6 cells, and the inhibitory effects of esculetin on inflammatory response, oxidative stress, apoptosis and the promotive effect of esculetin on autophagy in H/R-induced IEC-6 cells were weakened by SIRT3 knockdown, indicating that the protective role of esculetin against H/R-induced cell injury was partly achieved by up-regulating SIRT3.

Taken together, esculetin activated SIRT3/AMPK/mTOR signaling and autophagy to relieve inflammatory response, oxidative stress and apoptosis in intestinal I/R injury both in vitro and in vivo. This study expounded the protective role of esculetin against intestinal I/R injury and uncovered its potential molecular mechanism, suggesting the potential of esculetin as an effective drug for the treatment of intestinal I/R injury.

Data Sharing Statement

All data in this study have been included in this article.

Funding

This study was supported by National key research and development program (2018YFF01012100).

Disclosure

The authors declare that there are no conflicts of interest in this work.

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