

Foodborne Pathogens and Antimicrobial Resistance in Ethiopia: An Urgent Call for Action on “One Health”

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Abstract: Foodborne pathogens are the most common cause of foodborne disease worldwide. They contaminate food at all stages of the food chain, at the agricultural production level (soil and irrigation), from animal sources or at the time of preparation by food handlers. Therefore, food security is a critical issue that affects everyone in the world. Current studies indicate that the problem is more severe in low-income countries like Ethiopia. The majority of studies in Ethiopia have been conducted partially on human, animal or environmental samples with conventional culture-based diagnostic methods. Therefore, this review was conducted to gather information on the main foodborne pathogens and identify gaps in their traceability. Clinically important foodborne pathogens in Ethiopia include (*A. lumbricoids*, *Toxocara* spp., *H. nana*, *E. histolytica/dispar*, *G. intestinalis*, *H. diminuta* and *C. belli*), bacteria (*Salmonella* spp., *E. coli* O157:H7, *B. anthracis*, *Yersinia*, *C. perfringens*, *Klebsiella* spp., *M. paratuberculosis*, and *L. monocytogenes*), and viruses like (rotaviruses, enteroviruses and astroviruses, hepatitis E virus). In Ethiopia, all isolated foodborne bacterial pathogens showed high rates of antimicrobial resistance (AMR). In particular, the most studied foodborne pathogens, *Staphylococcus* spp., *Salmonella* spp., and *E. coli* from specific sources, showing high levels of resistance to most of the antibiotics prescribed in Ethiopia. The occurrence and persistence of AMR in food is one of the main factors causing the spread of antimicrobial resistance in different compartments, humans, animals and the environment. Therefore, strategies of coordination and struggle from a One Health perspective is an urgent strategy to control antibiotic resistance in order to achieve better outcomes for human and animal health.

Keywords: foodborne pathogens, antimicrobial resistance, food, Ethiopia, one health

Introduction

Food is essential for survival. To properly maintain human health, it must be pure, nutritious and free of any kind of impurities. Foodborne illness is defined as any infectious or noxious disease caused by eating contaminated food. A foodborne illness outbreak occurs when two or more cases of similar foodborne illness occur after consuming a common food.¹ Physical, chemical, and microbiological agents can all contaminate food. Bacteria, viruses, parasites and fungi are microbial agents that cause foodborne illness.² Food contamination can come from many different sources. Contaminants include contaminated water, flies, animals and pets, dirty cookware and utensils, dust, and additional impurities. Contaminated feces, pus, respiratory secretions, and other infectious waste from unsanitary food handlers can contaminate food.³

Foodborne pathogens are a major public health problem worldwide.⁴ Foodborne pathogens are one of the leading causes of illness in low-income countries, killing approximately 1.9 million people worldwide each year.⁵ Even in developed countries, foodborne illnesses affect about a third of the population each year.⁶ The effects of these pathogens also vary from region to region, as public awareness of food hygiene varies from country to country. Most foodborne pathogens are introduced as exogenous contaminants during handling, processing and preparation rather than as endogenous contaminants.^{7,8} The problem is serious in developing countries such as Ethiopia due to limitations in ensuring optimal hygienic food handling practices.^{9,10}

Pathogenic bacteria contaminate food at all stages of the food chain, from farm to table.¹¹ Humans can acquire pathogens or their infection through the consumption of a variety of contaminated food and water, or through contact with infected livestock and other animal feces. Infected people and the environment are also sources of infection.¹² Some important organisms circulating in different compartments include *Campylobacter* spp., *Salmonella* spp., *Staphylococcus* spp., *Enterococcus* spp. and Enterobacteriaceae produce ESBL. These bacterial pathogens are recognized as a global One Health concern due to the emergence and rapid transmission of antimicrobial resistance (AMR) in humans, animals and the environment. However, there are few studies evaluating foodborne pathogens and their AMR profiles in humans, animals, food and/or the environment in low-resource settings. According to a recent WHO review, high-quality data on the prevalence and abundance of AMR in humans, animals and food is lacking in low-income countries.¹³

The problem of foodborne disease is exacerbated in rural communities where awareness about the causes, transmission, and prevention of foodborne infections is generally lower. Furthermore, well-documented information on the extent of foodborne disease in Ethiopia is lacking as many cases are not properly diagnosed or reported, and foodborne illness patients do not seek medical attention. Therefore, it is difficult to get statistical data or even estimate the extent of the problem, except to say that it is very large. Today, everyday food safety has become a major global concern with the influence of international trade and public health. Therefore, this study aimed to review existing reports on major foodborne pathogens and AMR and point out gaps in the distribution of foodborne pathogens.

Methods

A systematic review was carried out in Ethiopia to assess the level of foodborne pathogen and antimicrobial resistance in food. Literature searches were performed using PubMed, Google Scholar, Scopus, Science Direct, and hard copies of locally published articles were obtained. The literature search was performed between February 1 and March 30, 2021. All available Ethiopian studies and data on foodborne pathogens and antimicrobial resistance from food reported in English have been compiled.

Literature Review, Discussion and Results

Foodborne Parasite

Parasites in food have long been an overlooked group of pathogens. However, they are a significant cause of disease and economic loss worldwide. Thus, the presence of these parasites in these foods warrants improved control efforts. To this end worldwide, significant progress has been made in the prevention and treatment of parasitic diseases in humans.¹⁴ However, parasitic diseases remain one of the most serious public health problems in the world, causing significant morbidity and mortality, especially in tropical and subtropical countries.¹⁵ In addition to causing morbidity and mortality, intestinal parasitic infections (IPs) have been linked to iron-deficiency anemia, childhood stunting, and other mental and physical health problems.^{16,17} It is estimated that 3.5 billion people are affected worldwide and 450 million people become sick from foodborne IPs, with approximately 200,000 deaths each year.¹⁸

Parasite infections can contaminate fruits and vegetables from the moment they are planted to the moment they are consumed.¹⁹ At the pre-harvest stage, the main contributing variables include the use of human and animal manure as natural fertilizers, as well as the irrigation of fruits and vegetables with untreated wastewater. The majority of local farmers in underdeveloped countries use untreated human or animal waste as fertilizer and contaminated water for irrigation, which contributes to the increased transmission of intestinal parasites (IP).²⁰ Storage, transport and marketing conditions, as well as hygienic standards during processing for food service or home consumption, are all factors in the postharvest stage.^{21,22}

Ethiopia is a country with a large number of IPs. There is also a lack of clean water and sanitary practices. As a result, farmlands are predicted to be contaminated by infective IPs, owing to open defecation. Furthermore, natural fertilizer (human and animal excrement) is widely employed by farmers in the country, and irrigation water is frequently contaminated.²³ Raw fruits and vegetables, such as bananas, mangos, tomatoes, salads, and green peppers, are also consumed raw. All of these factors lead to parasite infection of fruits and vegetables, making these foods crucial vehicles for human transmission.²⁴ According to recent studies in Ethiopia, parasites were found in 25.1% to 57.8% of the fruit

and vegetable samples collected during the marketing phase.^{23–31} The incidence of contamination and parasite species, on the other hand, varies depending on weather conditions, sociocultural status, season of sample collection, fruit and vegetable products analyzed, and other factors.

Parasites have also been detected in raw meat, posing a public health risk, especially in communities that consume raw. Zoonotic parasitic infection is a serious public health problem that requires a comprehensive control plan.^{32–36} Ingestion of parasites in contaminated food, drink or soil, as well as direct contact with dogs, livestock, or other domestic animals, leads to transmission of the disease to humans. Backyard slaughter, large numbers of stray dogs, a shortage of slaughterhouses proportional to the population, and improper disposal of diseased organ waste can all contribute to the high level of parasitemia in Ethiopia.³⁵ According to research, dogs, cats and other wild predators have easy access to offal's.³⁵

Zoonotic parasites can lead to significant financial losses¹⁴ due to reduced productivity of infected animals and convict contaminated organs such as liver, kidney and meat. Milk production, animal fertility and skin quality can all be affected.³⁶ According to a 10-year retrospective study in Ethiopia, total economic losses of 1,219,399 Ethiopian Birr (ETB) (\$61,946.9) were incurred, with an average annual cost of \$6194.69 due to waste of food, organs and carcasses of slaughtered cattle. Parasitic diseases in cattle, particularly hydatidosis and fascioliasis, remain a major problem, accounting for more than half (43.7%) of all visceral diseases in calves slaughtered in Ethiopia.³⁴ In addition, statistics show that 38.3% of cattle slaughtered in Ethiopia have one or more abnormalities, resulting in a total loss of \$61,946.9 (average annual loss of \$6194.69) caused by a variety of parasites.³⁴ Farmers and anyone involved in the livestock chain in sub-Saharan African countries like Ethiopia, which are already struggling with food shortages, are particularly concerned about these losses.

In Ethiopia, the most commonly clinically relevant foodborne parasites are *A. lumbricoids*, *H. nana*, *H. diminuta*, *E. histolytica* /*dispar*, *G. intestinalis*, *Toxocara spp.*, and *C. belli*. The levels of parasitic contamination of foods from various sources are summarized in the table below (Table 1).

Foodborne Bacteria

To control infectious diseases, chemotherapy alone will not be enough; a concerted effort to limit and eliminate possible sources of infection is needed. To address this, the periodic detection of infectious pathogens in food and related processes is a priority goal.³⁷ Plants and animals, and things derived from them, make up the majority of human food sources. It is helpful to think of food security as a product or service that can be bought or sold. Food appears to be a common source of microbial contamination, as some foods naturally contain pathogens or may come from diseased animals.³⁸

Bacteria in Fruits and Vegetables

Bacteria are found naturally in all food plants, from pre-harvest to post-consumption. Their presence or absence in food is considered as a sign of quality. The presence of bacteria in the soil can also affect the quality of food. This is most common in fruits and vegetables grown with contaminated irrigation water, as well as human and animal faces, grazing areas, and more.³⁹

Pathogenic bacteria do not exist naturally in vegetables and raw foods. Some sources of pre-harvest bacteria that can cause foodborne illness are irrigation water or wastewater, compost, manure, etc. Salad vegetables, such as lettuce, cabbage, tomatoes, and spinach, are at risk for bacterial contamination from untreated irrigation water.⁴⁰ A report by Adjrah et al, 2013⁴¹ found that manure used to support the growth of vegetables and crops contained significant amounts of harmful bacteria such as *Salmonella*, *E. coli* O157:H7, *B. anthracis*, *Yersinia*, *C. perfringens*, *Klebsiella spp.* *M. paratuberculosis*, and *L. monocytogenes*. Similarly, fields where wild animals or livestock have grazed are more likely to be contaminated with enteric infections like as *Salmonella* and *L. monocytogenes*, which can survive in agricultural soils for months.⁴² Therefore, the place of cultivation is most likely the first factor affecting the safety of fruits and vegetables.⁴³ Pre-harvest parameters such as planting material selection and crop management should all aim to provide a high quality product.⁴⁴ Microbial contamination of fruits and vegetables can occur during transportation, washing, peeling, slicing, pruning, packaging, and post-harvest handling.⁴⁵ Similarly, food preparation areas are frequently contaminated with *Micrococcus*

Table I Researches Done in Ethiopia That Showed Level of Parasite Contamination in Different Food Items

Author	Food Items	Source of Sample	Isolated Parasite	Reference
Bekele F. et al, 2017	Tomato, lettuce, cabbage, carrots, green pepper and avocado	Local open-market	<i>A. lumbricoids</i> , <i>Toxocara</i> spp., <i>H. nana</i> , <i>E. histolytica/dispar</i> , <i>G. intestinalis</i> , <i>H. diminuta</i> and <i>C. belli</i>	[23]
Tefera T et al ¹⁸	Tomato, lettuce, cabbage, carrots, green pepper, tomato, banana and mango	Local open markets	<i>Toxocara</i> spp., <i>H. nana</i> , <i>E. histolytica/dispar</i> , <i>G. intestinalis</i> , <i>H. diminuta</i> and Hookworms, <i>A. lumbricoids</i> , <i>Strongyloides</i> , <i>Cyclospora</i> spp and <i>Cryptosporidium</i> spp	[24]
Endale A et al, 2018	Lettuce, cabbage, spinach, carrot, green pepper, mango, tomato. orange banana	Local open markets	<i>E. histolytica/dispar</i> , <i>G. lambila</i> , <i>Strongyloides</i> , <i>A. lumbricoids</i> <i>Cyclospora</i> spp., <i>Cryptosporidium</i> spp., <i>I. Belli</i> , <i>Hymenolepis</i> spp., and <i>T. trichiura</i>	[25]
Alemu G. et al, 2019	Lettuce, cabbage, spinach, carrot, green pepper, mango, tomato	Local open markets	<i>E. histolytica/dispar</i> , <i>E. coli</i> , <i>G. intestinalis</i> , Hookworms, <i>A. lumbricoids</i> , <i>Strongyloides</i> , <i>Cyclospora</i> spp., <i>Cryptosporidium</i> spp., <i>F. hepatica</i> , <i>E. vermicularis</i> and <i>T. trichiura</i>	[26]
Gebremariam Gk, Girmay TG. 2020	Lettuce, cabbage, spinach, green pepper, potato tomato	Open market	<i>E. histolytica/dispar</i> , <i>G. lamblia</i> , <i>Cryptosporidium</i> spp., <i>A. lumbricoids</i> , <i>Taenia</i> spp., Hook worm spp., <i>E. Vermicularis</i> , <i>Fasciola</i> spp.	[27]
Delesa DA. 2017 ²²	Lettuce, cabbage, carrots	Farm and market	<i>Giardia</i> spp., <i>Entamoeba</i> spp., <i>Ascaris</i> spp., and <i>Cryptosporidium</i> spp.	[28]
Kifleyohannes T et al, 2021	Cabbage, carrots, tomato, green pepper, lettuce, guava	Local open-market, Backyards of farmers, irrigated farm lands	<i>Giardia</i> spp., <i>Cryptosporidium</i> spp. and <i>Entamoeba</i> spp.	[29]
Alemu G. et al, 2018	Tomato, cabbage, carrots, green pepper and salad	Local open markets	<i>G. lamblia</i> , <i>Cryptosporidium</i> spp., Hookworms, <i>B. coli</i> , <i>E. histolytica</i> , <i>Cyclospora</i> spp., <i>Stronglode</i> , <i>A. lumbricoids</i>	[30]
Bekele F et al, 2020	Tomato, lettuce, cabbage, carrots, green pepper and avocado	Local open markets	<i>Toxocara</i> spp., <i>H. nana</i> , <i>E. histolytica/dispar</i> , <i>G. intestinalis</i> , <i>H. diminuta</i> and Hookworms, <i>A. lumbricoids</i>	[31]
Getahun et al, 2020	Meat	Abattoirs	<i>Echinococcus</i> , <i>Fasciola</i> and <i>Cysticercus</i>	[32]
Muhammed S & Birhanu T. 2020	Organ (lung, liver, heart and kidneys)	Municipal Abattoirs	<i>Bovine hydatidosis</i> , <i>Bovine Fasciolosis</i> and <i>Cysticercus bovis</i>	[33]
Molla D. et al, 2018	Organ (lung, liver, heart, kidneys, head and tongue)	Abattoir condemnations	<i>Hydatidosis</i> , <i>Fasciolosis</i> and <i>Cysticercus</i>	[34]

spp. and *Staphylococcus* spp.⁴⁶ from the hands of the handlers. This means that dishwashing liquid, food preparation surfaces, serving equipment and main meals are all contaminated.⁴⁶ This suggests that the degree of microbiological contamination of fruits and vegetables is influenced by the individual handling of the suppliers. The microbiological safety of fruits and vegetables is also affected by the movement of consumers from one region to another.⁴⁷

Eating fruits and vegetables is now generally considered a risk factor for infection with enteric pathogens.^{48,49} Eating fruits and vegetables has been linked to foodborne illness outbreaks in many countries. During foodborne outbreaks, enteric pathogens such as *E. coli* and *Salmonella* spp. are among the most serious concerns.⁴² *Salmonella* spp., *E. coli* O157:H7, *B. anthracis*, *Yersinia*, *C. perfringens*, *Klebsiella* spp., *M. paratuberculosis*, and *L. monocytogenes* are the most common clinically important foodborne bacteria in Ethiopia. The table below summarizes the level of bacterial contamination in various food items from various sources in Ethiopia (Table 2).

Table 2 Researches Done in Ethiopia That Showed Level of Bacterial Contamination in Fruits and Vegetables

Author	Food Items	Source of Sample	Result and Isolated Bacteria	Reference
Mengistu DA. et al, 2022	Locally prepared fresh fruit juice	Juice houses	<i>E. coli</i> and other indicator organisms (Total coliform, fecal coliform and total viable bacterial count)	[50]
Berhanu M. et al, 2020	Packed and fresh fruit juice (non-refrigerated)	Supermarkets and cafes	<i>E. coli</i> , <i>S. aureus</i> , <i>Shigella</i> spp., <i>Salmonella</i> spp., <i>Klebsiella</i> spp., <i>Pseudomonas</i> spp.	[51]
Derra FA. et al, 2020	Mushroom, carrot, potato, Lentil, moringa, onion, salads and others	Retrospective data food microbiology laboratory of EPHI	<i>E. coli</i> , <i>S. aureus</i> , and indicator organism (total coliform, fecal coliform, Mesophilic aerobic bacteria and yeast and mold cont.)	[52]
Dobo B. 2019	Orange, banana, tomato, cabbage	Local markets	<i>S. aureus</i> , <i>Salmonella</i> spp., and indicator organism (total coliform, fecal coliform, Mesophilic aerobic bacteria and yeast and mold count)	[53]
Alemu et al, 2018	Tomato, cabbage, green pepper, carrot, lettuce	Local vendor	<i>E. coli</i> , <i>S. aureus</i> , <i>Shigella</i> spp., <i>Salmonella</i> spp.	[30]
Shiferaw M. and Kibret M. 2018	Avocado, guava	Juice houses	<i>Shigella</i> spp., <i>Salmonella</i> spp., and Indicator organisms (total coliform count, fecal coliform count, <i>S. aureus</i> count, Mesophilic aerobic bacteria and total yeast and mold counts)	[54]
Delesa DA. 2017	Lettuce, cabbage, carrots	Farm and market	Indicator organisms (total aerobic mesospheric count, total coliform count, fecal coliform count and total yeast and mold counts)	[28]
Weldeznaga D. and Muleta D. 2016	Lettuce, cabbage, tomato, carrots, potato, green pepper, onion	From irrigation farm land with river	<i>S. aureus</i> , <i>Salmonella</i> spp., and indicator organism (total coliform and fecal coliform)	[55]
Leul A. and Kibret M. 2012	Mango and pineapple juice	Cafes and juice houses	<i>Citrobacter</i> spp., <i>Salmonella</i> spp., <i>E. coli</i> , <i>Enterobacter</i> spp., <i>Klebsiella</i> spp., <i>Pseudomonas</i> spp. and indicator organism	[56]
Ayele W. 2009	Some vegetable like lettuce	Locally produced and marketed vegetable	<i>E. coli</i> (indicator organism)	[57]
Solomon ET. et al, 2002	Lettuce	Irrigation (study purpose)	<i>E. coli</i> O157:H7	[58]

Zoonotic Foodborne Bacteria

Due to the limited scope of the study and the lack of a coordinated epidemiological surveillance system, it is difficult to assess the burden of zoonotic bacterial infections in Ethiopia. In addition, underreporting of cases and the prevalence of other diseases considered high priority may mask the problem of foodborne infections.⁵⁹ A study in Jimma, South West Ethiopia,⁶⁰ Gondar, Ethiopia⁶¹ and Wolita Sodo, South Ethiopia⁶² showed high carriage rates of *Salmonella* spp., isolated from raw beef. The same study, conducted in the same area, showed high rates of *E. coli* and multidrug resistant (MDR) *E. coli* O157:H7 in meat from abattoirs and butchers.⁶³ In another study conducted at abattoirs and butchers in Addis Ababa, Ethiopia, a significant frequency of *S. aureus* was found.⁶⁴ A similar study conducted in Addis Ababa, Ethiopia, in a selected number of butcher shops also found inadequate meat quality and microbiological safety.⁶⁵ Additionally, another study found that cooked meat and fish posed a public health risk in Ethiopia.⁶⁶

According to a review report, pathogenic bacteria such as *E. coli*, *Salmonella* spp., *Campylobacter* spp., *L. monocytogenes*, *Brucella* spp., and *C. burnetii* were the most common isolates from milk and dairy products in Ethiopia.¹⁰ Another research report⁶⁷ revealed that dairy products in Ethiopia have poor microbiological characteristics. Research in the East Wollega Zone of Sibu Sire districts⁶⁸ and Lume and Siraro districts of Oromia state⁶⁹ showed poor hygienic practices and microbiological quality of milk obtained from farmers and markets. Collectively, a number of review articles^{70–73} and primary research⁷⁴ have identified zoonotic bacterial pathogens such as Anthrax, *E. coli*, *Brucella* spp., *C. burnetii*, *Leptospirosis*, *Salmonella* spp., *M. bovis*, *F. tularensis*, *L. monocytogenes*, *Campylobacter* spp., *Streptococcus* spp., *Bartonella* spp., *S. aureus*, *Ehrlichia*, *O. tsutsugamushi*, *Y. pestis*, and *R. rickettsii* are still a public health problem in Ethiopia.

Antimicrobial Resistance

The discovery of bacteria as the source of disease and the discovery of antibiotics as effective therapeutic agents has contributed to the protection of humans and animals over the past century.⁷⁵ Despite the advancement of many antibiotic discoveries, the development of antimicrobial resistant strains of bacteria has risen to the top of the list of global public health priorities. The World Health Organization can make the fight against the spread and emergence of antimicrobial resistance a top priority (WHO). Antimicrobial resistance may represent a global threat to the effectiveness of antibiotics, raising questions about their usefulness for both preventing and treating infections.⁷⁶ Antimicrobial resistance is driven by complex interacting factors that can be described as a resistance network or interface of a multifaceted One Health System,⁷⁷ which links clinical factors (human and animal health) and environmental factors (persistence of antimicrobial agents and AMR microorganisms in soil and food).^{78,79} Besides the indiscriminate use of antibiotics in human health, animal husbandry and agriculture, unsanitary food conditions can upset the microbial balance in favor of resistant bacteria through selective pressure. According to several studies conducted around the world, food is one of the potential sources of antibiotic resistance.^{79,80} The rise of antibiotic-resistant pathogens in food highlights the common requirement that these foods be cooked before consumption.

In the case of AMR, these diverse causes can be divided into two components: “selection”, mainly through antibiotic use, and “transmission”, the human-animal-environment axis. In livestock, the majority of global antibiotic use is in animals raised for food,^{81,82} and it is widely accepted that antibiotic use in animals is responsible for AMR,⁷⁰ although the magnitude of these effects is not well characterized and potentially specific to the AMR mechanism and organism. However, antibiotic use in humans is also a major cause of AMR.⁸³ In terms of transmission, resistant bacteria can be shared between humans, animals, and the environment through human-to-human contact (open community, patient contact, family transmission, workplace contact, traveler’s), human-sanitary contact (occupational contact with animals, consumption or preparation of food), animal-animal contact (involving agriculture or the movement of wild or domestic animals) and human-environment or animal-environment interactions.⁷⁹ It is important to note that we do not know the aforementioned selection and transmission factors that contribute to the increasing prevalence of AMR and the increasing incidence of AMR diseases. AMR selection and transfer is not a completely independent process. Although the use of antibiotics in humans and animals is believed to be present, environmental pollution from these sources and the use of antibiotics in non-animal agriculture increase the overall prevalence of antibiotic resistance. The relationship between antibiotic use and resistance is complex and dependent on pre-existing microbial population structures.⁸² Previous research has found that there is no single “quick fix” method for solving this problem. Instead, the prevention and reduction of the burden of AMR in the One Health system should take a coordinated, multifaceted approach that focuses on the specifics of antibiotic use, as well as type and prevalence of AMR in each system, while taking into account potential interactions within and between compartments.⁸⁴ It must also be multidisciplinary, multi-sector and collaborative.

In Ethiopia, several studies have shown an increase in antibiotic resistance in most bacterial species of human, animal, food and environmental origin.⁸⁵ Various studies in patients, animals, food and the environment have shown the risk of loss of valuable antimicrobials. For example, several studies in Ethiopia have found high rates of AMR isolates in foods (both plant and animal sources).^{70–73} However, the studies are not comprehensive and are difficult to compare. Evidence is often based on fragmented data from individual studies. Almost all studies have focused on specific pathogens, mainly *Staphylococci* spp., *Salmonella* spp., and *E. coli* from certain specific sources. Nevertheless, according to a review study,⁸⁵ the pooled prevalence

of AMR bacteria from live food-producing animals was 20%. High estimates of pooled AMR prevalence were found to be 29% for bacteria identified in milk and environmental samples and 28% for meat.⁸⁵ In foods of non-animal origin, the prevalence was lower with 13%.⁸⁵ Microorganisms reported to have higher MDR patterns are: *Staphylococcus* spp., (96%), *Salmonella* spp., (81%) and *E. coli* (77%).⁷⁸ Another review studies have also identified foodborne pathogens (*Salmonella* spp., pathogenic *E. coli*, *Shigella* spp., and *Campylobacter* spp) (11%) were extracted from samples (meat and meat products, dairy and dairy products, poultry products, and food contact surfaces).⁸⁶ The consumption of animal products such as meat, dairy and eggs, as well as vegetable products (fruits and vegetables), is increasing due to rapid population expansion, urbanization, increasing income per capita income, globalization and changes in consumption habits.^{70–73} Thus, the risk of the spread of emerging and re-emerging antibiotic resistance is increased by human-animal-ecosystem interactions and causes deaths and economic losses each year.⁸⁷

Foodborne Viral Infection

In recent years, viruses have been widely recognized as the main source of foodborne illness. Many known outbreaks of foodborne viruses have been traced to an infected food handler who contaminated prepared and ready-to-eat foods. Although viruses are now considered an extremely common cause of foodborne illnesses in many countries, they are rarely recognized because tools for the analysis and diagnosis of these viruses are generally unavailable. However, in recent years, there have been significant methodological advances available for the detection and identification of viruses in food.⁴⁷ These advances can help improve the assessment of the true burden of viral foodborne illness, as well as measures to prevent and control viral contamination of food. Viruses can infect humans in a variety of ways, but the most common foodborne viruses are those that infect the digestive system and are excreted in the feces and, in some cases, vomit. Norovirus is the most common cause of viral gastroenteritis worldwide, and hepatitis A virus, which can also be spread through the gastrointestinal tract, remains a global health concern. Rotavirus, Enterovirus and Astrovirus, Hepatitis E virus, emerging viruses (eg, Nipah virus, Highly Pathogenic Avian Influenza virus (HPAI), and Coronavirus that causes SARS) are also significantly associated with infection via food route.⁴⁷ In Ethiopia, data on foodborne viral infections are inadequate. However, one study of young people participating in a drinking water intervention trial found elevated levels of virome in the gut.⁸⁸

Conclusion and Recommendation

In Ethiopia, the presence of foodborne pathogens (parasites and bacteria) is significant and is considered as One Health concern. As a result, different sectors (public health, veterinary medicine, food safety, and the Water, Sanitation, and Hygiene (WASH) program) work together to deliver valuable interventions. It is necessary to develop food safety risk communication as well as a country-specific action plan to accelerate the response. It is essential to implement and identify practical, feasible and cost-effective interventions, as well as political commitment through high-level advocacy. It is essential to strengthen surveillance and diagnostic capacity at all levels, both in the veterinary and human sectors, to detect cases early and identify active reservoirs to protect benefits of humans and animals.

There is also a high prevalence of antibiotic resistance (AMR) in most bacterial species from food sources (animals and plants). Sources of contamination must be identified to control the spread of AMR through food. The prudent strategy of using antibiotics under expert supervision must be promoted and reinforced. Instead of acting as a “AMR amplifier”, we propose that the food chain could act as a “resistance modulator” to reduce the prevalence of resistant microorganisms by controlling careful parameters of the food process. However, further research is needed to establish both the processes involved in the spread of AMR and the food processing and storage parameters that are important for their reduction. Then, on this basis, robust control methods to prevent the spread of AMR among bacteria along the food chain can be created and implemented.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically

reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

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References

1. Guteta D. *Sanitary Survey of Food and Drinking Establishments in Ambo Town West Showa Zone Oromia Region* [Doctoral dissertation]; Addis Ababa University; 2007.
2. World Health Organization. Foodborne disease outbreaks: guidelines for investigation and control. World Health Organization; 2008.
3. Stewart GF, Amerine MA. *Introduction to Food Science and Technology*. Elsevier; 2012.
4. Amore G, Beloeil PA, Bocca V, et al; European Food Safety Authority (EFSA). Zoonoses, antimicrobial resistance and food-borne outbreaks guidance for reporting 2020 data; 2021.
5. Heredia N, Garcia S. Animals as sources of food-borne pathogens: a review. *Anim Nutr*. 2018;4(3):250–255. doi:10.1016/j.aninu.2018.04.006
6. Foundation AN, Safety F; American Medical Association, Centers for Disease Control and Prevention. Diagnosis and management of foodborne illnesses: a primer for physicians and other health care professionals. MMWR. Recommendations and reports: morbidity and mortality weekly report. *Recomm Rep*. 2004;53(RR-4):1–33.
7. Rane S. Street vended food in developing world: hazard analyses. *Indian J Microbiol*. 2011;51(1):100–106. doi:10.1007/s12088-011-0154-x
8. Paudyal N, Anihouvi V, Hounhouigan J, et al. Prevalence of foodborne pathogens in food from selected African countries—A meta-analysis. *Int J Food Microbiol*. 2017;249:35–43. doi:10.1016/j.ijfoodmicro.2017.03.002
9. Girma G. Prevalence, Antibiogram and growth potential of Salmonella and Shigella in Ethiopia: implications for public health. A Review. *Prevalence*. 2015;33:45.
10. Keba A, Rolon ML, Tamene A, et al. Review of the prevalence of foodborne pathogens in milk and dairy products in Ethiopia. *Int Dairy j*. 2020;109:104762.
11. Chen L, Alali W. recent discoveries in human serious foodborne pathogenic Bacteria: resurgence, pathogenesis, and control strategies. *Front Microbiol*. 2018;9:2412. doi:10.3389/fmicb.2018.02412
12. Linscott AJ. Food-borne illnesses. *Clin Microbiol Newsletter*. 2011;33(6):41–45.
13. Shrivastava SR, Shrivastava PS, Ramasamy J. World health organization releases global priority list of antibiotic-resistant bacteria to guide research, discovery, and development of new antibiotics. *J Med Soc*. 2018;32(1):76. doi:10.4103/jms.jms_25_17
14. Liu X, Wu M, Liu Y, Li J, Yang D, Jiang L. Foodborne parasites dominate current parasitic infections in Hunan Province, China. *Front Cell Infect Microbiol*. 2021;4:974.
15. Nyarango RM, Aloo PA, Kabiru EW, Nyanchongi BO. The risk of pathogenic intestinal parasite infections in Kisii Municipality, Kenya. *BMC Public Health*. 2008;8(1):1–6. doi:10.1186/1471-2458-8-237
16. Evans AC, Stephenson LS. Not by drugs alone: the fight against parasitic helminths. *World Health Forum*. 1995;16(3):258–261.
17. World Health Organization. Deworming for health and development: report of the Third Global Meeting of the Partners for Parasite Control. World Health Organization; 2005.
18. World Health Organization. WHO estimates of the global burden of foodborne diseases: foodborne disease burden epidemiology reference group 2007–2015. World Health Organization; 2015.
19. Adenusi AA, Abimbola WA, Adewoga TO. Human intestinal helminth contamination in pre-washed, fresh vegetables for sale in major markets in Ogun State, southwest Nigeria. *Food Control*. 2015;50:843–849. doi:10.1016/j.foodcont.2014.10.033
20. Idahosa OT. Parasitic contamination of fresh vegetables sold in Jos markets. *Global J Med Res*. 2011;11(1):21–25.
21. Alhabbal AT. The prevalence of parasitic contamination on common cold vegetables in Alqalamoun Region. *Int J Pharm Sci Rev Res*. 2015;30(1):94–97.
22. Amoah P, Drechsel P, Abaidoo RC, Klutse A. Effectiveness of common and improved sanitary washing methods in selected cities of West Africa for the reduction of coliform bacteria and helminth eggs on vegetables. *Trop med int health*. 2007;12:40–50. doi:10.1111/j.1365-3156.2007.01940.x
23. Bekele F, Tefera T, Biresaw G, Yohannes T. Parasitic contamination of raw vegetables and fruits collected from selected local markets in Arba Minch town, Southern Ethiopia. *Infect Dis Poverty*. 2017;6(1):1–7. doi:10.1186/s40249-016-0226-6
24. Tefera T, Biruksew A, Mekonnen Z, Eshetu T. Parasitic contamination of fruits and vegetables collected from selected local markets of Jimma Town, Southwest Ethiopia. *Int Scholar Res Notices*. 2014;2014:1–7. doi:10.1155/2014/382715
25. Endale A, Tafa B, Bekele D, Tesfaye F. Detection of medically important parasites in fruits and vegetables collected from local markets in Dire Dawa, Eastern Ethiopia. *Glob J Med Res*. 2018;18:29–36.
26. Alemu G, Mama M, Misker D, Haftu D. Parasitic contamination of vegetables marketed in Arba Minch town, southern Ethiopia. *BMC Infect Dis*. 2019;19(1):1–7. doi:10.1186/s12879-019-4020-5
27. Kiros Gebremariam G, Girmay TG. Parasitic contamination of fresh vegetables in open air markets of Aksum, Ethiopia; 2020.
28. Delesa DA. Intestinal parasitic and bacteriological contamination of raw vegetables from selected farms and markets in Nekemte, Ethiopia. *IJARBS*. 2017;4(12):191–200. doi:10.22192/ijarbs.2017.04.12.019
29. Kiflejohannes T, Debenham JJ, Robertson LJ. Is fresh produce in Tigray, Ethiopia a potential transmission vehicle for *Cryptosporidium* and *Giardia*? *Foods*. 2021;10(9):1979. doi:10.3390/foods10091979
30. Alemu G, Mama M, Siraj M. Bacterial contamination of vegetables sold in Arba Minch town, Southern Ethiopia. *BMC Res Notes*. 2018;11(1):1–5.

31. Bekele F, Shumbej T, Dendir A, et al. Contamination rate of commonly consumed fresh vegetables and fruits with parasites of medically importance in Wolkite and Butajira Towns of Gurage Zone, Southern Ethiopia. *Int J Public Health*. 2020;9(3):211–215.
32. Getahun D, Van Henten S, Abera A, et al. Cysts and parasites in an abattoir in Northwest Ethiopia; an urgent call for action on “one health”. *J Infect Develop Countries*. 2020;14(06.1):53S–57S. doi:10.3855/jidc.11713
33. Muhammed S, Birhanu T. Major bovine parasitic zoonosis at selected municipal abattoirs, North Shewa Zone, Oromia, Central Ethiopia. *Int J Veter Sci Res*. 2020;6(2):148–153.
34. Molla D, Nazir S, Mohammed A, Tintagu T. Parasitic infections as major cause of abattoir condemnations in cattle slaughtered at an Ethiopian abattoir: 10-year retrospective study. *J Helminthol*. 2020;94. doi:10.1017/S0022149X1900004X
35. Regassa B. Review on hydatidosis in small ruminant and its economic and public health significance. *J Dairy Vet Sci*. 2019;11(2):1–8.
36. Mitiku M. Epidemiology of cystic Bovine hydatidosis: emphasis on abattoir findings in Ethiopia. *J Veter Med Res*. 2017;4:56.
37. Jaykus LA. Epidemiology and detection as options for control of viral and parasitic foodborne disease. *Emerg Infect Dis*. 1997;3(4):529. doi:10.3201/eid0304.970418
38. Wirtanen G, Salo S. Microbial contaminants & contamination routes in food industry. In: *Microbial Contaminants & Contamination Routes in Food Industry: 1st SAFOODNET Open Seminar 2007*. VTT Technical Research Centre of Finland; 2022.
39. Burnett SL, Beuchat LR. Human pathogens associated with raw produce and unpasteurized juices, and difficulties in decontamination. *J Ind Microbiol Biotechnol*. 2000;25(6):281–287. doi:10.1038/sj.jim.7000106
40. Taban BM, Halkman AK. Do leafy green vegetables and their ready-to-eat [RTE] salads carry a risk of foodborne pathogens? *Anaerobe*. 2011;17(6):286–287. doi:10.1016/j.anaerobe.2011.04.004
41. Adjrah Y, Soncy K, Anani K, et al. Socio-economic profile of street food vendors and microbiological quality of ready-to-eat salads in Lomé. *Int Food Res J*. 2013;20(1):e43.
42. Alegbeleye OO, Singleton I, Sant’Ana AS. Sources and contamination routes of microbial pathogens to fresh produce during field cultivation: a review. *Food Microbiol*. 2018;73:177–208. doi:10.1016/j.fm.2018.01.003
43. Brackett R. Incidence, contributing factors, and control of bacterial pathogens in produce. *Postharvest Biol Technol*. 1999;15(3):305–311. doi:10.1016/S0925-5214(98)00096-9
44. Mritunjay SK, Kumar V. Potential hazards of microbial contamination associated with raw eaten salad vegetables and fresh produces. *Middle East J Sci Res*. 2015;23(4):741–749.
45. Oranusi S, Olorunfemi OJ. Microbiological safety evaluation of street vended ready-to-eat fruits sold in Ota, Ogun state, Nigeria. *Int J Res Biol Sci*. 2011;1(3):27–32.
46. Mensah P, Yeboah-Manu D, Owusu-Darko K, Ablordey A. Street foods in Accra, Ghana: how safe are they? *Bull World Health Organ*. 2002;80(7):546–554.
47. World Health Organization. Microbiological hazards in fresh leafy vegetables and herbs: meeting report. World Health Organization; 2008.
48. Heaton JC, Jones K. Microbial contamination of fruit and vegetables and the behaviour of enteropathogens in the phyllosphere: a review. *J Appl Microbiol*. 2008;104(3):613–626. doi:10.1111/j.1365-2672.2007.03587.x
49. Buck JW, Walcott RR, Beuchat LR. Recent trends in microbiological safety of fruits and vegetables. *Plant Health Progress*. 2003;4(1):25. doi:10.1094/PHP-2003-0121-01-RV
50. Mengistu DA, Mulugeta Y, Mekbib D, Baraki N, Gobena T. Bacteriological quality of locally prepared fresh fruit juice sold in juice houses of Eastern Ethiopia. *Environ Health Insights*. 2022;16:11786302211072949. doi:10.1177/11786302211072949
51. Berhanu M, Adal M, Sahile S. Microbial quality spectrum of packed and fresh fruit juices in Gondar town supermarkets and Cafes, Northwestern Ethiopia. *J Microbiol Res*. 2020;10:45–54.
52. Derra FA, Bedada T, Edichio R, et al. Evaluation of the consumption and contamination level of Vegetables and Fruits in Ethiopia; 2020.
53. Dobo B. Fungal and bacterial contamination of fresh fruits and vegetables sold in Hawassa town of southern Ethiopia. *Global Sci J*. 2019;7(3):1038–1049.
54. Shiferaw M, Kibret M. Microbial quality of avocado and guava fruits used for preparation of freshly squeezed juices from juice houses of Bahir Dar town, northwest Ethiopia. *Int J Sci Res Publ*. 2018;8:100–113.
55. Weldezigina D, Muleta D. Bacteriological contaminants of some fresh vegetables irrigated with Awetu River in Jimma Town, Southwestern Ethiopia. *Adv Biol*. 2016;2016:1–11. doi:10.1155/2016/1526764
56. Leul A, Kibret M. Bacteriological safety of freshly squeezed Mango and pineapple juices served in juice houses of Bahir Dar town, northwest Ethiopia. *Int J Sci Basic Appl Res*. 2012;6:24–35.
57. Ayele W. A preliminary survey of the microbiological and parasitological quality of some locally produced and marketed vegetables in Arba Minch, Ethiopia. Arba Minch Town ROSA project booklet; 2009: 43–44.
58. Solomon EB, Potenski CJ, Matthews KR. Effect of irrigation method on transmission to and persistence of *Escherichia coli* O157: H7 on lettuce. *J Food Prot*. 2002;65(4):673–676. doi:10.4315/0362-028X-65.4.673
59. Edget A, Dagmar N, Biruhtesfa A. Review on common foodborne pathogens in Ethiopia. *Afr J Microbiol Res*. 2014;8(53):4027–4040.
60. Geresu MA, Desta WZ. Carriage, risk factors, and antimicrobial resistance patterns of *Salmonella* isolates from raw beef in Jimma, Southwestern Ethiopia. *Infect Drug Resist*. 2021;14:2349. doi:10.2147/IDR.S313485
61. Ejo M, Garedew L, Alebachew Z, Worku W. Prevalence and antimicrobial resistance of salmonella isolated from animal-origin food items in Gondar, Ethiopia. *Biomed Res Int*. 2016;2016:1–8. doi:10.1155/2016/4290506
62. Wabeto W, Abraham Y, Anjulo AA. Detection and identification of antimicrobial-resistant *Salmonella* in raw beef at Wolaita Sodo municipal abattoir, Southern Ethiopia. *J Health Popul Nutr*. 2017;36(1):1–7. doi:10.1186/s41043-017-0131-z
63. Sebsibe MA, Asfaw ET. Occurrence of multi-drug resistant *Escherichia coli* and *Escherichia coli* O157: H7 in meat and swab samples of various contact surfaces at abattoir and butcher shops in Jimma town, Southwest district of Ethiopia. *Infect Drug Resist*. 2020;13:3853. doi:10.2147/IDR.S277890
64. Adugna F, Pal M, Girmay G. Prevalence and Antibigram Assessment of *Staphylococcus aureus* in Beef at Municipal Abattoir and Butcher Shops in Addis Ababa, Ethiopia. *Biomed Res Int*. 2018;2018:1–7. doi:10.1155/2018/5017685
65. Zerabruk K, Retta N, Muleta D, Tefera AT. Assessment of microbiological safety and quality of minced meat and meat contact surfaces in selected butcher shops of Addis Ababa, Ethiopia. *J Food Qual*. 2019;2019:1–9. doi:10.1155/2019/3902690

66. Bedada TL, Feto TK, Awoke KS, et al. Microbiological and public health status of cooked meat and fish in Ethiopia. *Open Microbiol J.* 2020;14(1):123–129. doi:10.2174/1874285802014010123
67. Zahara L, Zinewi H. The Microbial properties of dairy products in Ethiopia: a Review. *Int J Dairy Sci Technol.* 2015;2(1):088–094.
68. Adugna C, Eshetu M. Hygienic practice, microbial quality and physico-chemical properties of milk collected from farmers and market chains in Eastern Wollega zone of Sibu Sire districts, Ethiopia. *Int J Agricult Sci Food Technol.* 2021;7(1):125–132.
69. Amenu K, Spengler M, Markemann A, Zárate AV. Microbial quality of water in rural households of Ethiopia: implications for milk safety and public health. *J Health Popul Nutr.* 2014;32(2):190.
70. Abebe E, Gugsu G, Ahmed M. Review on major food-borne zoonotic bacterial pathogens. *J Trop Med.* 2020;2020:1–19. doi:10.1155/2020/4674235
71. Pieracci EG, Hall AJ, Gharpure R, et al. Prioritizing zoonotic diseases in Ethiopia using a one health approach. *One Health.* 2016;2:131–135. doi:10.1016/j.onehlt.2016.09.001
72. Asante J, Noreddin A, El Zowalaty ME. Systematic review of important bacterial zoonoses in Africa in the last decade in light of the “One Health” concept. *Pathogens.* 2019;8(2):50. doi:10.3390/pathogens8020050
73. Cavalerie L, Wardeh M, Lebrasseur O, et al. One hundred years of zoonoses research in the Horn of Africa: a scoping review. *PLoS neglected tropical diseases.* *PLoS Neglected Trop Dis.* 2021;15(7):e0009607. doi:10.1371/journal.pntd.0009607
74. Mekonnen SA, Gezehagn A, Berju A, et al. Health and economic burden of foodborne zoonotic diseases in Amhara region, Ethiopia. *PLoS One.* 2021;16(12):e0262032. doi:10.1371/journal.pone.0262032
75. World Bank. *Pulling Together to Beat Superbugs: Knowledge and Implementation Gaps in Addressing Antimicrobial Resistance.* Washington, DC; 2019.
76. World Health Organization. WHO report on surveillance of antibiotic consumption: 2016–2018 early implementations; 2019.
77. Holmes AH, Moore LS, Sundsfjord A, et al. Understanding the mechanisms and drivers of antimicrobial resistance. *Lancet.* 2016;387(10014):176–187. doi:10.1016/S0140-6736(15)00473-0
78. O'Neill J. Review on antimicrobial resistance: tackling drug-resistant infections globally: final report and recommendations; 2016.
79. Abbassi MS, Badi S, Lengiz S, Mansouri R, Salah H, Hynds P. Hiding in plain sight-wildlife as a neglected reservoir and pathway for the spread of antimicrobial resistance: a narrative review. *FEMS Microbiol Ecol.* 2022;98(6):fiac045. doi:10.1093/femsec/fiac045
80. Woolhouse M, Ward M, Van Bunnik B, Farrar J. Antimicrobial resistance in humans, livestock and the wider environment. *Philos Trans R Soc B Biol Sci.* 2015;370(1670):20140083. doi:10.1098/rstb.2014.0083
81. Van Boeckel TP, Glennon EE, Chen D, et al. Reducing antimicrobial use in food animals. *Science.* 2017;357(6358):1350–1352. doi:10.1126/science.aao1495
82. Van Boeckel TP, Pires J, Silvester R, et al. Global trends in antimicrobial resistance in animals in low-and middle-income countries. *Science.* 2019;365(6459):eaaw1944. doi:10.1126/science.aaw1944
83. Laxminarayan R, Matsoso P, Pant S, et al. Access to effective antimicrobials: a worldwide challenge. *Lancet.* 2016;387(10014):168–175. doi:10.1016/S0140-6736(15)00474-2
84. World Health Organization. The evolving threat of antimicrobial resistance: options for action. World Health Organization; 2012.
85. Gameda BA, Assefa A, Jaleta MB, Amenu K, Wieland B. Antimicrobial resistance in Ethiopia: a systematic review and meta-analysis of prevalence in foods, food handlers, animals, and the environment. *One Health.* 2021;13:100286. doi:10.1016/j.onehlt.2021.100286
86. Belina D, Hailu Y, Gobena T, Hald T, Njage PM. Prevalence and epidemiological distribution of selected foodborne pathogens in human and different environmental samples in Ethiopia: a systematic review and meta-analysis. *One Health Outlook.* 2021;3(1):1–30. doi:10.1186/s42522-021-00048-5
87. Erkyihun GA, Gari FR, Edao BM, Kassa GM. A review on one health approach in Ethiopia. *One Health Outlook.* 2022;4(1):1–3. doi:10.1186/s42522-022-00064-z
88. Altan E, Aiemojy K, Phan TG, et al. Enteric virome of Ethiopian children participating in a clean water intervention trial. *PLoS One.* 2018;13(8):e0202054. doi:10.1371/journal.pone.0202054

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