

Role of Cytokines on the Progression of Liver Fibrosis in Mice Infected with *Echinococcus multilocularis*

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Background: Liver fibrosis is a significant pathological change of *Echinococcus multilocularis* (*E. multilocularis*) infection. This study aimed to explore the role of cytokines on the progression of liver fibrosis in mice infected with *E. multilocularis*.

Methods: Liver histopathological features at 2, 8, 30, 90 and 180 d were quantified by inflammatory severity score. The expression levels of inflammatory cytokines, fibrosis-related cytokines and hepatic cell apoptosis were measured using qRT-PCR and immunohistochemistry.

Results: At the early stage of infection, parasite stimulation triggers the rapid recruitment of immune cells, such as macrophages and neutrophils. These infiltrated immune cells then produce a large number of cytokines, such as iNOS (inducible nitric oxide synthase), a pro-inflammatory cytokine; TGF- β (transforming growth factor) activated HSCs (hepatic stellate cells) to promote the proliferation of fibroblasts and secretion of ECM (extracellular matrix); MMP9 (matrix metalloproteinase 9) degraded basal ECM and facilitated its replacement by a highly dense interstitial matrix. At the middle and late stages of infection, the expression of IL-10 (interleukin-10) with general inhibitory effect was increased. The imbalance of fiber formation and degradation aggravated liver fibrosis. Meanwhile, the whole process of *E. multilocularis* infection was accompanied by the necrosis and apoptosis of hepatic cells.

Conclusion: Along with the expansion of parasitic infection, dynamic changes in cytokine expression were observed on the liver fibrosis progression, which is helpful to provide some new ideas for the prevention and treatment of liver fibrosis in mice infected with *E. multilocularis*.

Keywords: *Echinococcus multilocularis*, liver fibrosis, infection immunity, cytokines, parasite

Introduction

Alveolar echinococcosis (AE) is one of the major public hazard diseases, characterized by continuous and infiltrative tumor-like growth of the *Echinococcus multilocularis* (*E. multilocularis*) metacestode stage over years or even decades in the organs of the intermediate host with estimated 17,400 infections per year.^{1,2} Studies showed *E. multilocularis* infections in China, mainly distributed in Xinjiang, Qinghai, Sichuan province and so on.³ *E. multilocularis* infection mainly affects the liver, and if the treatment is not timely, the fatality rate is often very high.

Liver fibrosis, as one of the histopathological phenomena in mice infected with *E. multilocularis*, is associated with aberrant apoptosis of hepatocytes, collagen

formation and liver immune cells inflammation.⁴ In chronic and persistent inflammatory injuries, such as alcoholic hepatitis, viral hepatitis, parasitic diseases, and so on, this abnormal regulation of liver tissue repair mechanisms leads to irreversible fibrosis, more seriously, liver cirrhosis or liver cancer. Parasite stimulation triggers the rapid recruitment of immune cells, such as neutrophils and macrophages. These infiltrated immune cells then produce a large number of cytokines, such as iNOS (inducible Nitric Oxide Synthase) and IL-10 (interleukin-10).⁵ TGF- β (transforming growth factor) is a key mediator in immune response, matrix synthesis, cell proliferation, apoptosis, etc.,⁶ more importantly, TGF- β is also the main pro-fibrosis cytokine involved in the process of liver fibrosis.⁷ TGF- β activates HSCs (hepatic stellate cells) to promote the proliferation of fibroblasts and secretion of ECM (extracellular matrix), which finally results in fibrosis occurrence.⁸ Liver microenvironment contains the interactions between HSC, liver resident immune cells, hepatocytes and/or sinusoidal endothelial cells, which are crucial for host defense and tissue remodeling.⁹ During liver pathophysiology, the interactions between HSCs and immune cells are critical to the deposition of ECM and the development of fibrosis, which are also mediated by cytokine signaling.^{10,11} Within hepatic damage, HSCs produce MMPs (matrix metalloproteinases) to degrade the basal ECM, facilitating its replacement by a highly dense interstitial matrix, while at the same time, promoting TGF- β to release from pre-existing ECM, which is a reservoir of cytokines.¹² Therefore, it seems likely that the downregulation of MMPs may lead to a decrease in circulating TGF- β by inhibiting the secretion of the profibrogenic factor and blocking the deposition of high-density interstitial ECM, which may provide another possible antifibrotic mechanism. Caspases are a family of intracellular cysteine proteases that mediate apoptosis and inflammation through the processing and activation of pro-inflammatory cytokines, such as IL-1 β (interleukin-1 β), IL-18, and IL-33.¹³ Studies have shown that hepatocyte apoptosis induces liver fibrosis,¹⁴ which is an excessive wound healing response to chronic liver injury, and hepatocytes death and collagen accumulation could reversely guide released chemokines by recruiting immune cells to further aggravate the inflammatory response.^{15,16}

Although many CCL4-induced models have allowed us to gain a deeper understanding of the complexity of liver inflammation and repair, the composition of the ECM in *E. multilocularis* infection is less well studied. In our

study, a mice model of *E. multilocularis* infection was successfully established, dynamic changes of cytokine expression were observed on the liver fibrosis progression, which is helpful to provide some ideas for the prevention and treatment of liver fibrosis in mice infected with *E. multilocularis*.

Materials and Methods

Animals and Infected-Model

Establishment

BALB/c mice (female, 8-week-old) purchased from the Experimental Animal Science Research Department (Urumqi, China) were maintained in an air-conditioned animal room with a 12-h light/dark cycle and provided with rodent chow and water. *E. multilocularis* protocercariae were obtained from intraperitoneal lesions maintained in BALB/c. All animals were treated according to the guidelines of the Institutional Animal Care, with protocols approved by the Institutional Animal Use and Care Committee of Xinjiang Medical University (No. 20170214-106). All animals received humane care according to the criteria outlined in the Guide for the Care and Use of Laboratory Animals prepared by the National Academy of Sciences and published by the National Institutes of Health (NIH publication 86-23, revised 1985). Specifically, mice were intraperitoneally (left lower quadrant) injected with 5% chloral hydrate 0.15–0.20 mL/mouse through medical syringe needle, and peripheral blood was collected by retro-orbital bleeding after successfully anesthetized; then, whole liver samples were obtained surgically; at last, mice were euthanized using cervical dislocation; the dead body further packed with special medical waste bags to freeze under -20°C and delivered for biosafety handling. A 50 mL syringe was used to aseptically remove the swelling solution, and the remaining tissue was removed and placed into a sterile mortar for grinding. The grinded tissue was diluted by the prepared saline (0.9% saline mixed with 500 U/mL penicillin and 100 U/mL streptomycin) and screened by a 200-mesh nylon. The filtrate was collected in a 50 mL centrifuge tube and left to settle in the protocercaria. After most of the protocercariae was precipitated, the supernatant was discarded, and then rinsed with the prepared saline until it was clear. The final precipitation was reconstituted with PBS and the protocercariae counted (DMI 4000B microscope, Leica, Germany) and adjusted to the appropriate parasite concentration before injection. The parasite vitality was determined by eosin

exclusion. Only parasite batches exhibiting over 95% vitality were used.¹⁷ After suspension, it was prepared into a cercaria suspension of 2000 cercaria/mL for later use. Mice were anesthetized by 2.5% chloral Hydrate, each mouse was intraperitoneally injected with 1 mL of cercaria suspension (in the control group, mice injected with the same amount of normal saline).

Serum and Liver Tissue Samples

Mice peripheral blood was collected by retro-orbital bleeding after being successfully anesthetized at the time points (2, 8, 30, 90 and 180 d). Some liver tissue was fixed in 10% formalin for 24 h and embedded in paraffin, while others were quickly frozen in liquid nitrogen and stored at -80°C .

Measuring ALT and AST Expression in Serum

Mice peripheral blood was collected by retro-orbital bleeding after being successfully anesthetized (anticoagulant, Edathamil dipotassium salt, EDTA-K2). Liver injury was estimated by the increased activity of serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST), which were evaluated by an automatic biochemistry analyzer (Hitachi, Ltd, Kokubunji, Tokyo, Japan).

Liver Histopathological Evaluation

The liver tissues were fixed in 4% paraformaldehyde solution, embedded in paraffin for histological examinations and stained with haematoxylin-eosin (H&E) and Masson's trichrome, assessing liver pathological changes and fibrosis degrees. For H&E staining, the sections were dehydrated into water, stained with hematoxylin (Baoman Biotechnology, Shanghai, China) for 10 min and rapidly washed with running water around 30 s. Then, the sections were differentiated by 1% hydrochloric acid and alcohol, washed by running water again around 1 min, stained with eosin at room temperature around 5 min, dehydrated by graded ethanol and cleared by xylene once around 1 min. The sections were sealed with neutral gum, and the pathological changes were observed and photographed under a microscope. Two experienced pathologists blindly gave a score of severity in mice infected with *E. multilocularis* (Table 1).

For Masson staining, sections were dewaxed into water, stained with hematoxylin for 10 minutes, rinsed with water, followed by hydrochloric acid for several seconds, rinsed

with water again, and stained with ammonia for several minutes. Then, garnet magenta, 12-molybdenum-phosphate solution and green staining solution were successively dyed for about 10 s, about 3 min, about 4 min. The slices were washed with water and air-dried, sealed with neutral adhesive, observed and photographed under a microscope. $\times 200$ Histopathological changes were observed under a microscope. Image J was used to analyze 5 discontinuous random organization fields and obtain the mean value.

Immunohistochemistry (IHC) for Detection of Cytokines Secretion in Liver Tissue

Liver tissue sections (3 μm) were hydrated and inactivated with 3% H_2O_2 . Incubate with goat serum at room temperature for 30 min. iNOS antibody (Abcam, Cambridge, UK, ab115819, dilution 1:200), IL-10 antibody (Bioss, Beijing, China, bs-0698R, 1:200), TGF- β antibody (Affinity, Cincinnati, US, AF5347, dilution 1:200), α -SMA antibody (Affinity, Cincinnati, US, AF1032, dilution 1:200), COL1 α 1 antibody (Affinity, Cincinnati, US, AF7001, dilution 1:200), MMP9 antibody (Affinity, Cincinnati, US, AF5228, dilution 1:200) or Caspase 3 antibody (Affinity, Cincinnati, US, AF6311, dilution 1:200) were incubated overnight at 4°C . The tablets were washed the next day, and the second antibody labeled with horseradish peroxidase was incubated at 37°C for 60 min. Sections were developed with DIAMinobenzidine (DAB) and stained with hematoxylin. The slides were rinsed with distilled water and air-dried, sealed and placed under a microscope for observation. Image J software was used to evaluate the mean optical density of immunohistochemical images.

Quantitative Real-Time PCR for Detecting mRNA Expression

Total RNA was isolated from liver tissue employing a TrizolTM isolation kit (Takara Bio, Dalian, China) as per manufacturer's instructions, subsequently converted to cDNA, which was carried out using PrimeScript reagent kit with gDNA Eraser (Takara Bio, Dalian, China), and subjected to real-time PCR using SYBR Premix Ex TaqII (Takara Bio). The primers for GAPDH, iNOS, IL-10, TGF- β , α -SMA, coll1 α 1, MMP9 and caspase 3 were synthesized by Sangon Biotech (Shanghai, China), details showed in Table 2. Real-Time PCR was operated on ABI Prism 7500 Sequence Detection System (BioRad, Life

Table 1 Hepatic Infection of *Echinococcus multilocularis* Severity Score (Mouse)

Degree of Inflammation		Vesicles* (Number)	Abnormal Structures and Substances	Score
Hepatic Lobule	Portal Area and Surrounding			
No inflammation	No inflammation	0	No obvious change	0
Degeneration, small amount spotted and focal necrosis	Mild inflammation (Inflammatory cells occupy less than 1/3 of the portal area)	1–3	Granuloma	1
Degeneration, large amount spotted and focal necrosis		4–6		2
Confluent necrosis or bridging necrosis	Moderate inflammation (Inflammatory cells occupy 1/3~2/3 of the portal area)	7–9	Laminated layer and germinal layer	3
Extensive bridging of necrosis and involved many hepatic lobules	Severe inflammation (Inflammatory cells occupy more than 2/3 of the portal area)	≥10	Brood capsule and protoscolex	4

Note: *The average number of vesicle counts was observed in at least 3 different fields under low power microscope (magnification, ×100).

Science Research, Hercules, CA, USA). PCR conditions were as follows: one cycle at 95°C for 30 s, 40 cycles at 95°C for 5 s, at 61°C for 30 min. All samples were run in triplicates. Relative mRNA abundances were determined with $2^{-\Delta\Delta C_t}$ method using GAPDH gene to normalize.

Statistical Analysis

The quantitative analysis of morphology results was carried out using Image J. Data were analyzed by SPSS 21.0 (IBM, Chicago, IL, USA) or GraphPad Prism 8.0 software (GraphPad Software, San Diego, CA, USA). Results were expressed as means ± standard error of mean (SEM). Differences between groups were analyzed, according to variable distribution using *t*-test/Mann–Whitney for two groups or ANOVA/Kruskal–Wallis (with Bonferroni or Dunn posttests, respectively) for three or more groups. A *P*-value <0.05 was considered to be statistically significant.

Results

Severity of Inflammation and Fibrosis Was Aggravated in Mice Infected with *E. multilocularis*

Liver specimens stained by H&E from the control group showed normal features with hepatocytes, liver architecture, central veins and portal areas. The mice infected with *E. multilocularis* resulted in distortions of the normal hepatic architecture with swollen hepatocytes around the central veins and interruption of hepatic tissue by numerous fibrous septa together with diffuse cellular infiltration around the fibrous bands and dilated portal areas. Furthermore, the vesicle structure could be observed in 90d and 180d infection (Figure 1). Based on the characteristics of *E. multilocularis*

infection in mice, we invited two experienced pathologists to develop inflammatory infection scores (Table 1). After summing up the scores, the higher the score, the more severe the infection (Figure 2A). Masson staining results analyzed by image J showed that in the model group, the area of collagenous fibrosis in the liver tissues significantly increased (Figures 1 and 2B).

Liver Function of Mice Infected with *E. multilocularis* at Different Stages

The increased ALT and AST indicated the damage to hepatocytes. In the model group of mice infected with

Table 2 Primer Sequence

Gene (Mouse)	Primer (5'→3')
iNOS	Forward:GCAGAGATTGGAGGCCTTGTC Reverse:GGGTTGTTGCTGAACTTCCAGTC
IL-10	Forward:ATAAGAGCAAGGCAGTGGAGC Reverse:GGCCTTGTAGACACCTTGGTC
TGF-β	Forward:AGCTGCGCTTGCAGAGATTA Reverse:GACAGCCACTCAGGCGTATC
α-SMA	Forward:GTCCCAGACATCAGGGAGTAA Reverse:TCGGATACTTCAGCGTCAGGA
Col1α1	Forward:TAAGGGTCCCCAATGGTGAGA Reverse:GGGTCCCTCGACTCCTACAT
MMP9	Forward:GGACCCGAAGCGGACATTG Reverse:CGTCGTCGAAATGGGCATCT
caspase 3	Forward:CTCGCTCTGGTACGGATGTG Reverse:TCCCATAAATGACCCCTTCATCA
GAPDH	Forward:TGGTGAAGCAGGCATCTGAG Reverse:TGAAGTCGCAGGAGACAACC

Abbreviations: iNOS, inducible nitric oxide synthase; IL-10, interleukin-10; TGF-β, transforming growth factor-β; α-SMA, α-smooth muscle actin; Col1α1, collagen type I alpha 1; MMP9, matrix metalloproteinase 9; GAPDH, glyceraldehyde-3-phosphate dehydrogenase.

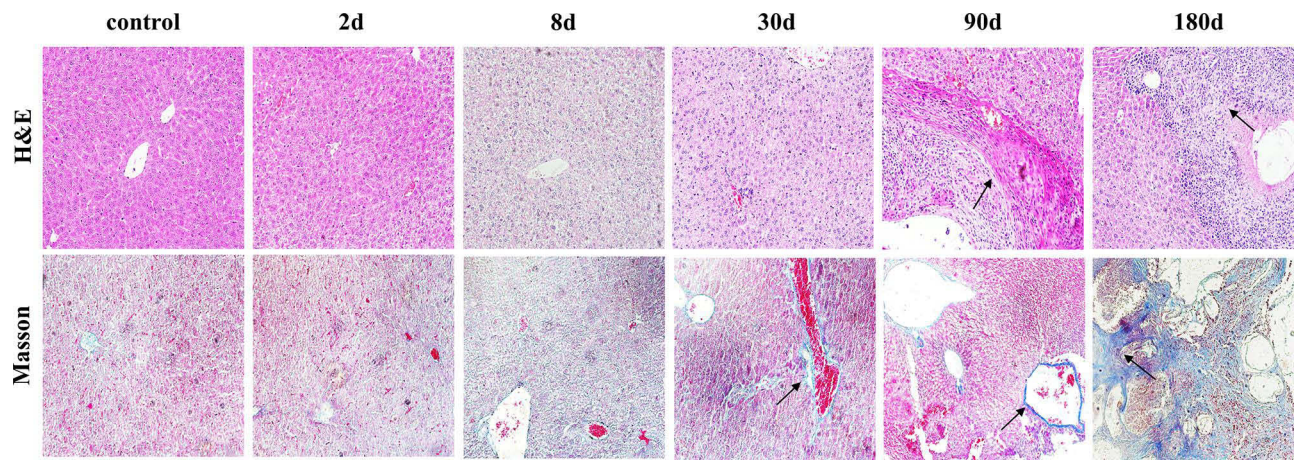


Figure 1 Pathological changes in the liver of mice infected with *Echinococcus multilocularis* (x200).

Note: Immune cell infiltration and fibrosis change are showed with arrows.

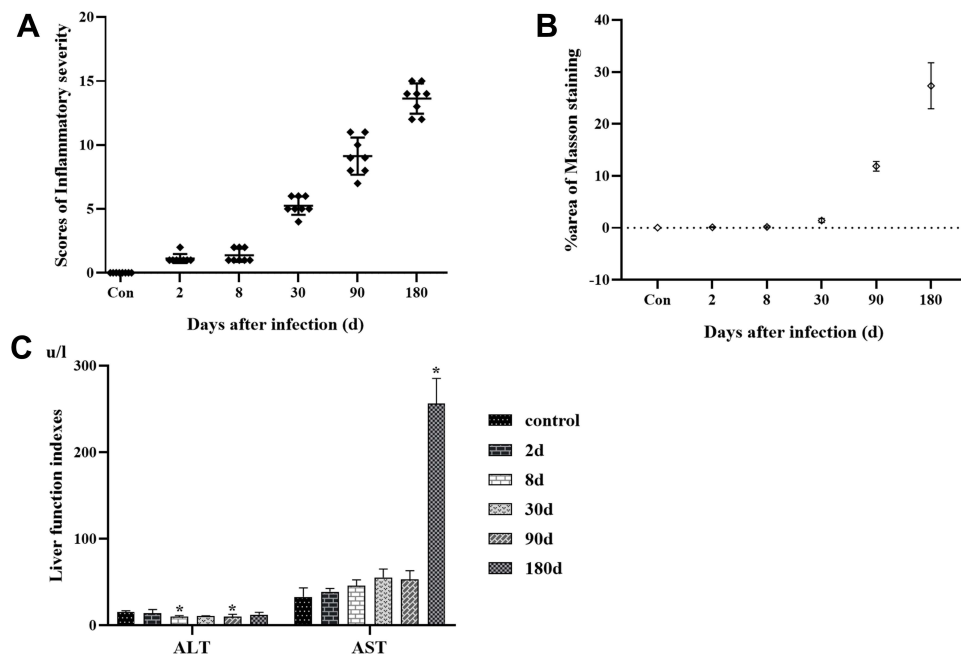


Figure 2 (A) Liver inflammatory severity scores Note, n=8 in each group, ♦Indicates the score of inflammatory severity in each mouse; (B) dynamic quantification of fibrosis area Note, ◇Indicates the average area percent in Masson staining of different group; (C) liver function in mice infected with *Echinococcus multilocularis*.

Note: *Compared with the control group, $P < 0.05$.

Abbreviations: ALT, alanine aminotransferase; AST, aspartate aminotransferase.

E. multilocularis, serum ALT increased at a low level in each period of infection, and the increase was not obvious, and even decreased at 8d and 90d points after infection compared with the control group ($P < 0.05$). The reason might be related to the disorder of the liver immune micro-environment and hepatocytes necrosis (Figure 2C). When hepatocytes necrosis is severe, AST in mitochondria is released. A high AST can reflect the severity of liver cell injury and more importantly the severity of liver cell injury.

Compared with the control group, it showed a rapid increase at 180d after infection ($P < 0.05$) (Figure 2C).

Dynamic Changes of Inflammatory Cytokines in the Liver of Mice Infected with *E. multilocularis*

iNOS, as one pro-inflammatory cytokine, responds positively when stimulated by parasites and expressed in almost all hepatocytes, mainly in hepatocyte plasma and

infiltrated inflammatory cells. As shown in Figures 3 and 4A, at the 8d infection, the expression of iNOS was strongly positive ($P < 0.05$, compared with the control or

2d model group), which suggested that inflammatory cells quickly recruited a large number of inflammatory cells to the injured liver tissue at the early stage of parasitic

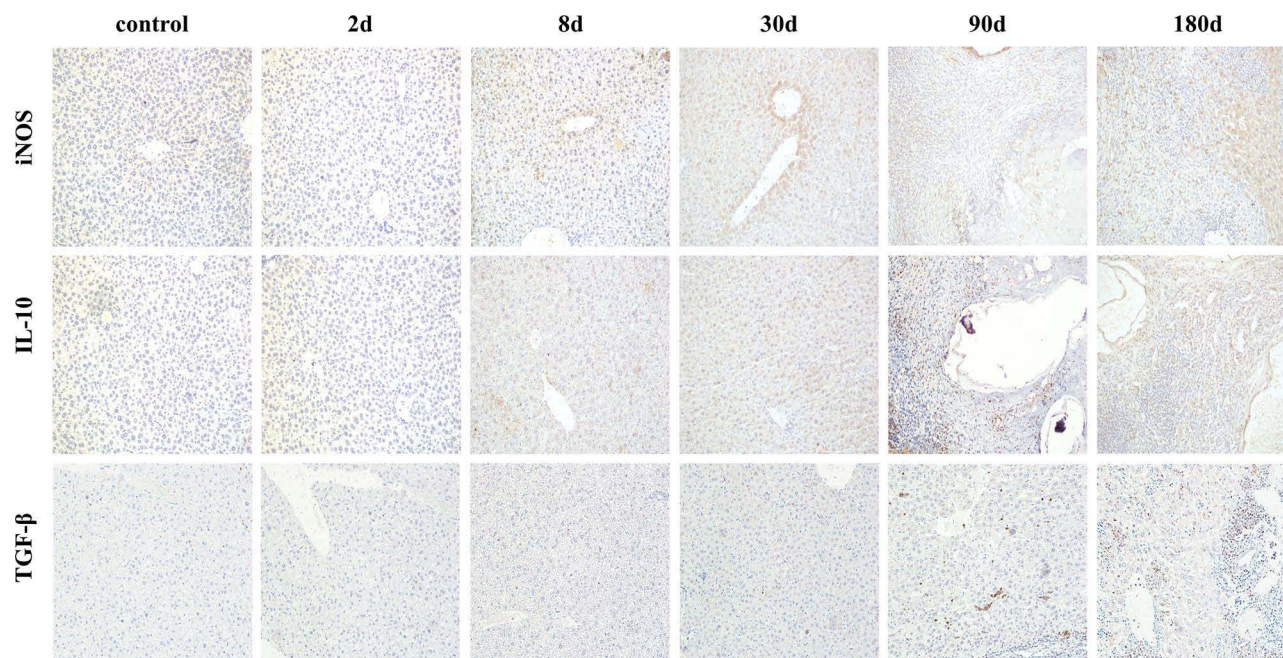


Figure 3 Dynamic changes of inflammatory cytokines in immunohistochemistry (x200).

Abbreviations: iNOS, inducible nitric oxide synthase; IL-10, interleukin-10; TGF-β, transforming growth factor-β.

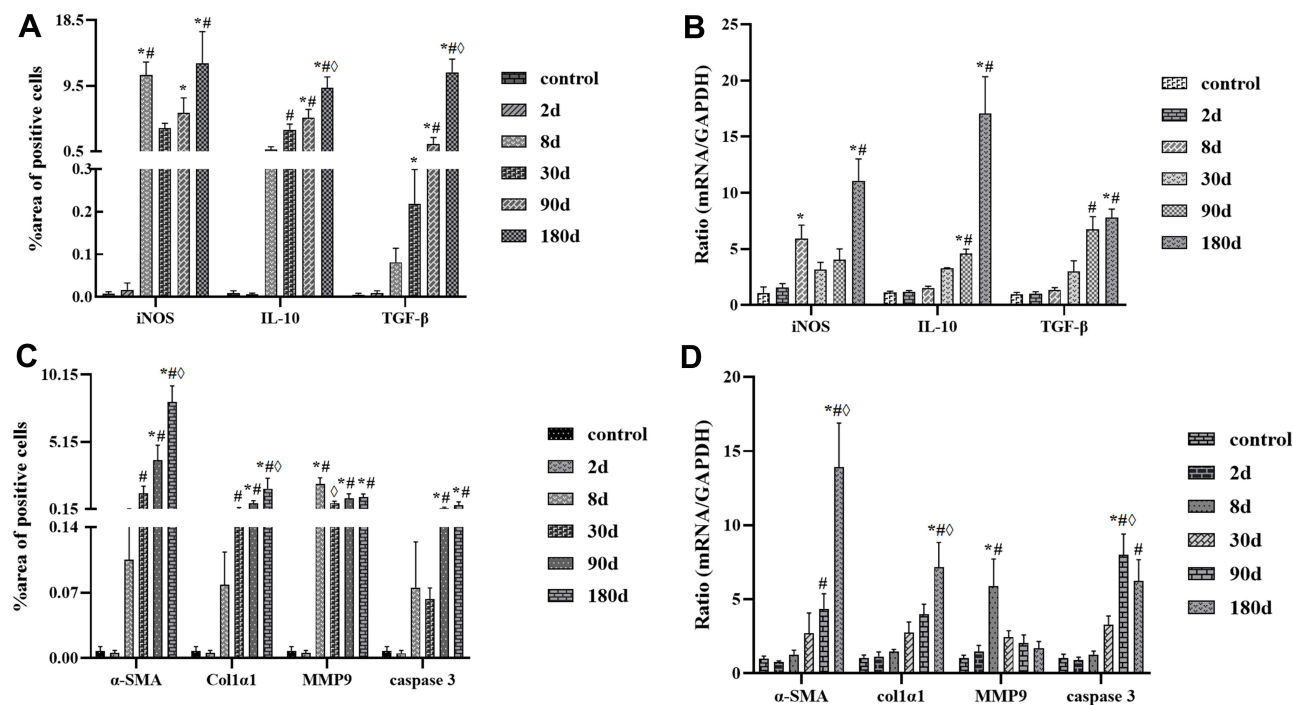


Figure 4 (A and C) Dynamic quantification of cytokines' expression in protein level; (B and D) Dynamic quantification of cytokines' expression in mRNA level.

Notes: *Compared with the control group, $P < 0.05$; #Compared with the infection model of 2d group; ◇Compared with the infection model of 8d group.

Abbreviations: iNOS, inducible nitric oxide synthase; IL-10, interleukin-10; TGF-β, transforming growth factor-β; α-SMA, α-smooth muscle actin; Col1α1, collagen type I alpha 1; MMP9, matrix metalloproteinase 9.

invasion to promote the immune response. IL-10 can underregulate the pro-inflammatory response and liver fibrosis,¹⁸ it showed a weak positive expression at the early stage of infection but a strong positive expression mainly at the middle and late stages of infection. TGF- β , as an important mediator of inflammatory response and tissue repair, was strongly positive mainly at the middle and late stages of infection (Figures 3 and 4A). At the gene expression level, the results were essentially similar to those at the protein level (Figure 4B). With continuous stimulation of parasite, the lesion continues to expand, and the liver microenvironment is further disturbed for a long time. Mainly at the middle and late stages of infection, iNOS, IL-10 and TGF- β all showed high expression levels ($P < 0.05$, compared with the control group), especially at 30d, 90d and 180d model group after infection. With the expansion of parasitic lesions, the liver of mice infected with *E. multilocularis* formed immune tolerance at the middle and late stages (Figures 3, 4A and B).

Dynamic Changes of Fibrosis Related Cytokines and Cell Apoptosis in the Liver of Mice Infected with *E. multilocularis*

With the continuous infection of *E. multilocularis*, chronic damage leads to a disorder in the liver microenvironment, in which the inflammatory response is gradually aggravated, a large number of hepatocytes are necrotic and apoptotic, and the protective self-repair of fibrosis also widely exists in liver tissue. As a major member of liver fibrosis, HSC is mainly activated by TGF- β , which mediates inflammatory response and fibrosis changes (active marker, α -SMA), and rapidly transforms into myofibroblast-like cell, which further promotes the accumulation of ECM and produces collagen. MMP9 as a double-edged sword in chronic infection, can promote the deposition and degradation of collagen and regulate the development of fibrosis level, liver fibrosis promotes the self-repair of liver tissue at the early stage of infection, and it is reversible. As the infection continues to progress, the degree of fibrosis increases and the liver microenvironment becomes disorganized and irreparable, which may eventually lead to cirrhosis and cancer. In our study, α -SMA, as a marker of HSC activation, was highly expressed at both protein and gene levels at the middle and late stages of infection, such as, at 90d and 180d points ($P < 0.05$) (Figures 4C, D and 5). COL1 α 1, as a major component of collagen deposition, was expressed similarly to α -SMA. MMP9

increased rapidly at 8d point (compared with the control group, $P < 0.05$), and then decreased rapidly at the middle and late stages of reinfection. Although the expression was higher than that in the control group, it was not statistically significant ($P > 0.05$). As for liver cell apoptosis, Caspase 3 was highly expressed on 90d and 180d after infection (compared with the control group, $P < 0.05$), and the abnormal expression of these indicators suggested the continuous development and aggravation of liver fibrosis in mice infected with *E. multilocularis* (Figures 4C, D and 5).

Discussion

Human alveolar echinococcosis (AE), caused by the metacystode larval stage of the fox-tapeworm *E. multilocularis*, is a chronic zoonosis associated with significant modulation of the host immune response. Liver fibrosis, as one of the histopathological phenomena in mice infected with *E. multilocularis*, is associated with aberrant apoptosis of hepatocytes, collagen formation and liver immune cells inflammation.⁴ Interactions between HSCs, liver resident immune cells, hepatocytes and/or sinusoidal endothelial cells, which constitute the liver microenvironment, are crucial not only for host defense but also for tissue remodeling.⁹ The cross-talk between HSCs and immune cells in the hepatic microenvironment during liver pathophysiology is crucial for ECM deposition and fibrosis development.^{10,11}

The disease is characterized by an increased infiltration of immune cells around the lesion, creating an “immunosuppressive” microenvironment that is in favor of persistent parasite infection. At the early stage of infection, parasite stimulation triggers the rapid recruitment of immune cells, such as macrophages and neutrophils. These infiltrated immune cells then produce a large number of cytokines, such as iNOS, small amounts of NO generated by eNOS are thought to be important for liver homeostasis and protective against pathological conditions. In contrast, large amounts of NO generated by iNOS are implicated in the etiologies of many liver diseases, including liver fibrosis.¹⁹ In the liver, IL-10 can be produced by a variety of cell types including hepatocytes, Kupffer cells, sinusoidal endothelial cells, HSCs, and lymphocytes; its receptor is expressed by progenitor and HSC, the predominant cell types involved in liver fibrogenesis. IL-10 regulates the production of collagen and collagen enzyme, and affects the ECM remodeling by promoting HSC apoptosis and regulating function.²⁰ That day number

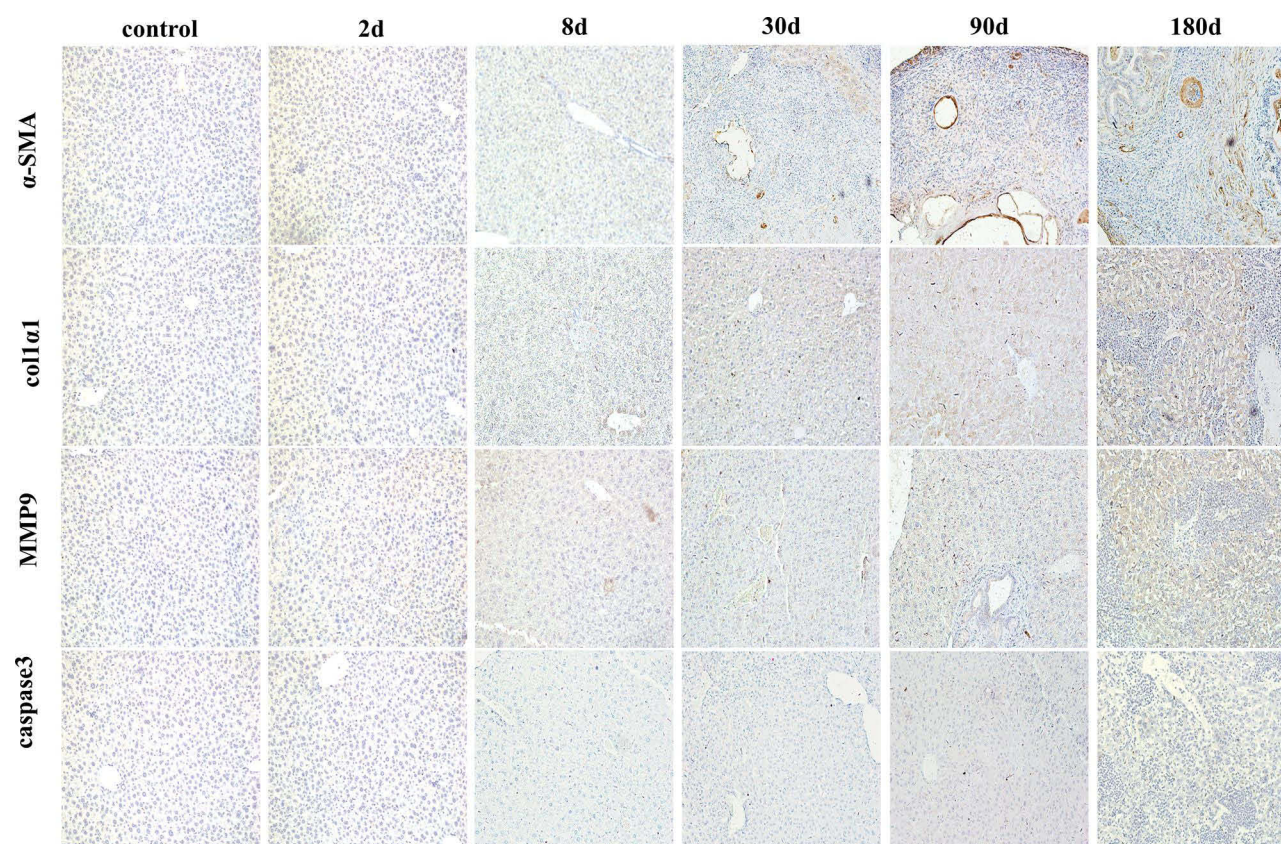


Figure 5 Dynamic changes of fibrosis related cytokines and cell apoptosis in immunohistochemistry (x200).

Abbreviations: α -SMA, α -smooth muscle actin; Coll α 1, collagen type I alpha 1; MMP9, matrix metalloproteinase 9.

2–60, 60–180 and 180–360 were, respectively, divided as early, middle, and late stages.²¹ In the most susceptible mice, impairment of vital functions due to *E. multilocularis* progression and metastases is fast and occurs. In addition, susceptible mice mean that the liver tissue is almost completely fibrotic and even calcified without normal cells after more than 180 days of infection. We designated these stages: 2 d, 8 d and 30 d after infection as early stages; 90d and 180d after infection as middle and late stages. In our study, the dynamic changes of liver inflammatory cytokines in mice infected with *E. multilocularis*, showed that iNOS increased rapidly at the early stage (8d), promoting the acute immune response, and maintaining a high level mainly at the middle and late stages of infection. The anti-inflammatory cytokine, IL-10 was mainly elevated mainly at the middle and late stages of infection.

TGF- β is a necessary mediator in many fields, such as the immune response, inflammation, matrix synthesis, cell growth, apoptosis, and differentiation.⁶ Human cystic echinococcosis (CE) is another type of echinococcosis, echinococcosis caused by inoculation of the eggs of

E. granulosus. Our previous research has shown that the expression of TGF- β 1 was upregulated in patients with hepatic CE, which was closely associated with liver fibrosis.²² Our results showed that TGF- β did not change significantly at the early stage of *E. multilocularis* infection, but the level of TGF- β increased at 30d point, and maintained a high expression level at 90d and 180d points. The expression of α -SMA and COL1 α 1 at different stages was similar to TGF- β .

It has also been suggested that fibronectin peptides are generated by limited digestion of MMP9 during liver injury, which acts as local regulatory signals that induce the apoptosis of HSCs and prevent chronic fibrogenesis and fibrosis.²³ Increased evidence has demonstrated that MMP9 promotes fibrogenesis by activating latent TGF- β , which stimulates the activation of HSCs.²⁴ MMP9 has been demonstrated to induce the apoptosis of activated HSCs.²⁵ In our study, the expression of MMP9 in liver tissue of mice infected with *E. multilocularis* was highly expressed at the early stage (8d), and decreased at the middle and late stages, indicating that early inflammatory cells secreted proteases including MMP9, which decreased

the deposition of ECM, which might be related to the innate immune response.

Pharmacologic inhibition may attenuate liver injury, inflammation, and fibrosis.²⁶ Typically, necroptosis is regarded as a pro-inflammatory mode of cell death that releases intracellular components. However, many studies have shown that necroptosis may have anti-inflammatory effects by blocking the excessive production of chemokines,^{27,28} and our study also supports this conclusion as caspase 3 is highly expressed at 90d and 180d infection both in protein and gene level. Apoptosis seems to play a positive role in the procession of fibrosis in mice infected with *E. multilocularis*.

In the experimental model of secondary infection in mice in our study, initially, inflammatory cells are recruited to the liver, which release cytokines mainly to fight against parasitic infections to exert its pro-inflammatory role, later a large number of anti-inflammatory cytokines are released to promote liver repair. Gradually, it develops immune tolerance to *E. multilocularis* infection; liver fibrosis is constantly progressing; hepatocyte apoptosis or necroptosis; eventually, liver immune microenvironment disorder and fails to repair itself. Many studies on the cytokine profile in AE, in humans and in the experimental models, have stressed that it was never a pure profile, but always a rather mixed profile,^{17,22} such selective and opposite dynamics of inflammatory cytokines at different infected stages release may prevent pathogenic inflammation, and constitute an appropriate response for attraction of effector cells into the periparasitic tissues with the capacity to limit *E. multilocularis* metacystode proliferation and dissemination. Results of observations in humans and experimental studies in animals suggest that, in the absence of fully effective antiparasitic chemotherapy for AE, modulation of the host's immune response could be envisaged to fight against the parasite and to prevent the disease and/or its complications. The balance of liver microenvironment is regulated tightly by cytokines through an intricate interplay of positive and negative regulatory signals. Our study provides evidence that cytokines have played key roles in the host-parasite interaction in *E. multilocularis* infection.

In conclusion, the cellular and molecular mechanisms of liver inflammation and fibrosis in mice infected with *E. multilocularis* are being investigated limitedly. Liver inflammation and fibrogenesis are controlled by complex immunologic pathways that implicate many possible therapeutic targets. Our research may provide some new ideas for *E. multilocularis* infection.

Data Sharing Statement

The data used to support the findings of this study are included within the article.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

Funding

This work was supported by the National Natural Science Foundation (82060372, 81760372). Scientific Research Project of Science Department of Xinjiang Autonomous Region (2020E0277) and State Key Laboratory of Pathogenesis, Prevention and Treatment of High Incidence Diseases in Central Asia Fund (SKL-HIDCA-2020-YG3, SKL-HIDCA-2021-5).

Disclosure

Fengming Tian and Tao Jiang are co-first authors of this study. The authors declare that they have no competing interests.

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