


# Different Induction of PD-L1 (CD274) and PD-1 (CD279) Expression in THP-1-Differentiated Types 1 and 2 Macrophages

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**Background:** Phorbol 12-myristate 13-acetate (PMA)-induced differentiation of human monocytic THP-1 cells is an experimental model for preparing resting macrophages (M<sub>0</sub>) for cell polarization toward the different functional specializations of macrophages.

**Methods:** In this study, we examined the expression of immune checkpoints by using flow cytometry following multicolor staining. The blockade of immune checkpoint by using neutralizing antibodies was performed to assess their role in PMA-induced THP-1-differentiated macrophages.

**Results:** Upon the inducible macrophage differentiation caused by PMA, increased expression levels of CD11b and CD68 were measured and characterized according to their adherent phenotype accompanied by the generation of cellular complexity. While the cell growth rate was abolished post-differentiation, some cells underwent cell death. Notably, we found increases in the expression of programmed cell death protein 1, also known as PD-1 (CD279), and its ligand PD-L1 (CD274), mainly in differentiated M<sub>0</sub> (CD68<sup>+</sup>CD11b<sup>+</sup>) macrophages. However, neutralizing PD-L1/PD-1 neither blocked THP-1 cell differentiation toward macrophages nor inhibited macrophage polarization in M<sub>1</sub> and M<sub>2</sub>. In specializing macrophages, a decrease both in CD274 and CD279 was found in M<sub>2</sub>.

**Conclusion:** These results revealed the inducible expression of PD-L1/PD-1 in PMA-induced THP-1-differentiated M<sub>0</sub> macrophages followed by a decrease in M<sub>2</sub> macrophages.

**Keywords:** PMA, THP-1, macrophage, differentiation, PD-L1, PD-1

## Introduction

Macrophages, a type of phagocyte of innate immunity, are responsible for recognizing, engulfing, and destroying targets such as pathogens and apoptotic cells.<sup>1,2</sup> Compared to circulating neutrophils and monocytes, macrophages are professional phagocytes that participate in immune defense. In the human body, macrophages are mainly and notably recruited to the lungs, liver, brain, spleen, and lymph nodes. A relatively low macrophages can be detected in the peripheral blood.<sup>3</sup> It is noted that the different types of activated macrophages produce diverse proinflammatory cytokines/chemokines for triggering intercellular inflammation in response to infections, allergens, and cancers.<sup>1,2,4</sup> For the regulation in immunity, macrophages can be differentiated toward antigen-presenting cells to modulate cellular T and humoral B cell immunity.<sup>5</sup> Therefore, in macrophages, their biological roles and cellular regulation are heterogeneous.

Macrophages are generally and functionally divided into two specialized classes: classically (type I or M<sub>1</sub>) and alternatively (type II or M<sub>2</sub>) activated

macrophages.<sup>2</sup> M<sub>1</sub> macrophages are responsible for proinflammation and most antimicrobe and anticancer activities, and M<sub>2</sub> macrophages promote anti-inflammation and tumorigenesis, including tumor growth and metastasis.<sup>4</sup> To differentiate specialized tissue macrophages, the possible modulators derived from infectious pathogens and cancer cells are essential for macrophage maturation, activation, and polarization.<sup>6,7</sup> Notably, blood monocytes are activated and recruited into inflamed tissues and mostly constitute an effective process for further differentiation toward macrophages.<sup>1,3,4,8</sup> The processing protocols of monocyte/macrophage differentiation and polarization are currently used for immune cell therapy in treating inflammatory disorders, infectious diseases, and cancers.<sup>9,10</sup>

For monocyte-derived macrophage differentiation, artificial strategies are processed by exogenously treating monocytes with bioactive growth factors, such as granulocyte-macrophage colony-stimulating factor (GM-CSF) and M-CSF, as well as chemical agents, such as 1,25-dihydroxy vitamin D<sub>3</sub> and phorbol-12-myristate-13-acetate (PMA).<sup>11</sup> In addition to isolated primary monocytes, human monocytic cell lines such as THP-1 and HL-60 are the most frequently used cell model systems.<sup>11,12</sup> Following PMA treatment, THP-1 cells are differentiated into macrophage-like cells, resulting in cells with increased adherence and decreased growth activity.<sup>13</sup> At the resting stage of macrophages (M<sub>0</sub>), the cells can be polarized into M<sub>1</sub> and M<sub>2</sub> by stimulation with lipopolysaccharide (LPS)/interferon (IFN)- $\gamma$  and interleukin (IL)-4/IL-10/IL-13/tumor growth factor (TGF)- $\beta$ , respectively.<sup>8</sup> Recent studies have discovered the aberrant expression of immune checkpoints such as programmed cell death protein 1, also known as PD-1 (CD279), and its ligand PD-L1 (CD274) in M<sub>2</sub> macrophages, which are involved in tumorigenesis.<sup>14–16</sup> However, the regulation of PD-L1/PD-1 expression in monocyte/macrophage differentiation is unknown. In this study, by using PMA-induced THP-1 differentiation toward M<sub>0</sub>, M<sub>1</sub>, and M<sub>2</sub> macrophages, we examined the regulation of PD-L1/PD-1 expression and its possible role in macrophage differentiation before polarization.

## Methods

### Cells, Culture Condition, and Reagents

Human THP-1 monocytic cells (ATCC TIB-202) were cultured in RPMI-1640 medium (Invitrogen Life Technologies, Carlsbad, CA, USA) supplemented with 10% heat-inactivated fetal bovine serum (FBS, Invitrogen Life Technologies) and maintained at 37°C with 5% CO<sub>2</sub>.

Phorbol 12-myristate 13-acetate (PMA) and LPS were purchased from Sigma-Aldrich (St. Louis, MO, USA) and dissolved in dimethyl sulfoxide (DMSO, Sigma-Aldrich) and double-distilled H<sub>2</sub>O, respectively. Cytokines, including IFN- $\gamma$ , IL-4, and IL-13, were purchased from PeproTech (Rocky Hill, NJ). Neutralizing antibodies against CD274 (Cat#10084-R639) and CD279 (Cat# 10377-HN94) was purchased from Sino Biological (Wayne, PA, USA).

### Differentiation Protocol

The protocol for THP-1 macrophage differentiation was performed according to previous works with modifications.<sup>13,17</sup> THP-1 cells were divided into two groups and seeded in a 6-well culture plate ( $2 \times 10^5$  cells/mL). Following seeding for 24 h, cells were treated with one microliter per mL of DMSO as a control group and 150 ng/mL of PMA as an M<sub>0</sub> group for 48 h. After that, PMA-containing medium was washed away from all cells and replaced with PMA-free culture medium for 24 h then the cells were treated without (for M<sub>0</sub>) or with LPS (1  $\mu$ g/mL)/IFN- $\gamma$  (10 ng/mL) (for M<sub>1</sub>) and IL-4 (25 ng/mL)/IL-13 (25 ng/mL) (for M<sub>2</sub>) for an additional 72 h.

### Morphological Observation

Cell images were taken for the determination of control and PMA-induced THP-1 macrophage differentiation using an inverted microscope (AxioVert. A1 Inverted Microscope, Carl Zeiss, Germany) under the same magnification of light intensity. Representative images were observed under 10 $\times$  magnification, and the area of observation was chosen randomly from the whole well.

### Immunofluorescence Staining

For flow cytometric analysis, the cells were detached with 0.01% trypsin and then re-suspended in 0.1 mL of Flow Cytometry Staining Buffer (Thermo Fisher Scientific, Carlsbad, CA, USA) in the final cell concentration with  $1 \times 10^7$  cells/mL. Cells were stained with fluorescence-conjugated antibodies, including antibodies against human CD11b (Cat# 47-0118-42), CD68 (Cat# 12-0689-42), CD80 (Cat# 67-0809-42), CD209 (Cat# 45-2099-42), CD274 (Cat# 47-5983-42), and CD279 (Cat# 67-2799-42) (Thermo Fisher Scientific). The cells were washed twice in phosphate buffered saline and then analyzed using flow cytometry (Attune Nxt, Invitrogen Life Technologies) with excitation set at 405, 488, and 633 nm, and the data were

analyzed using FCS Express 7 Cytometry software (De Novo Software, CA, USA).

## MTT Assay and Lactate Dehydrogenase (LDH) Assay

For the detection of cell growth and cytotoxicity, MTT Cell Proliferation Kit (Sigma-Aldrich) and Cytotoxicity Detection kit assays (Roche Diagnostics, Lewes, UK) were performed, respectively, according to the manufacturer's instructions. A microplate reader (SpectraMax 340PC; Molecular Devices Corporation, Sunnyvale, CA, USA) was used to measure the absorbance at 570 and 490 nm for MTT and LDH, respectively. The data were analyzed using Softmax Pro software (Molecular Devices Corporation).

## Statistical Analysis

The values are expressed as the means  $\pm$  standard deviation (SD). The groups were compared using Student's two-tailed unpaired *t*-test or a one-way analysis of variance analysis. A *P* value of 0.05 was considered significant.

## Results

### Stimulation of Phorbol 12-Myristate 13-Acetate (PMA) Promotes Macrophage Differentiation in Human Monocytic THP-1 Cells

To create an inducible model of macrophage differentiation, human monocytic THP-1 cells were used in this study according to previous works with partial modification.<sup>12,13</sup> With PMA stimulation for 2 days, THP-1 cells were cultured only in PMA-free medium for an additional 4 days as a differentiated phenotype of THP-1 M<sub>0</sub> (Figure 1A). Image observation showed an adherent phenotype with the enlarged cell size in PMA-stimulated THP-1 cells (Figure 1B). Using flow cytometric analysis, PMA-stimulated THP-1 cells displayed an increase in cell size as well as cellular complexity characterized by high granularity (Figure 1C). Several cell markers could be identified in differentiated macrophages,<sup>17</sup> and immunostaining of specific markers followed by flow cytometric analysis showed increased expression of CD11b and CD68 (Figure 1D). As supported by the above experiments, the stimulation of PMA effectively promoted

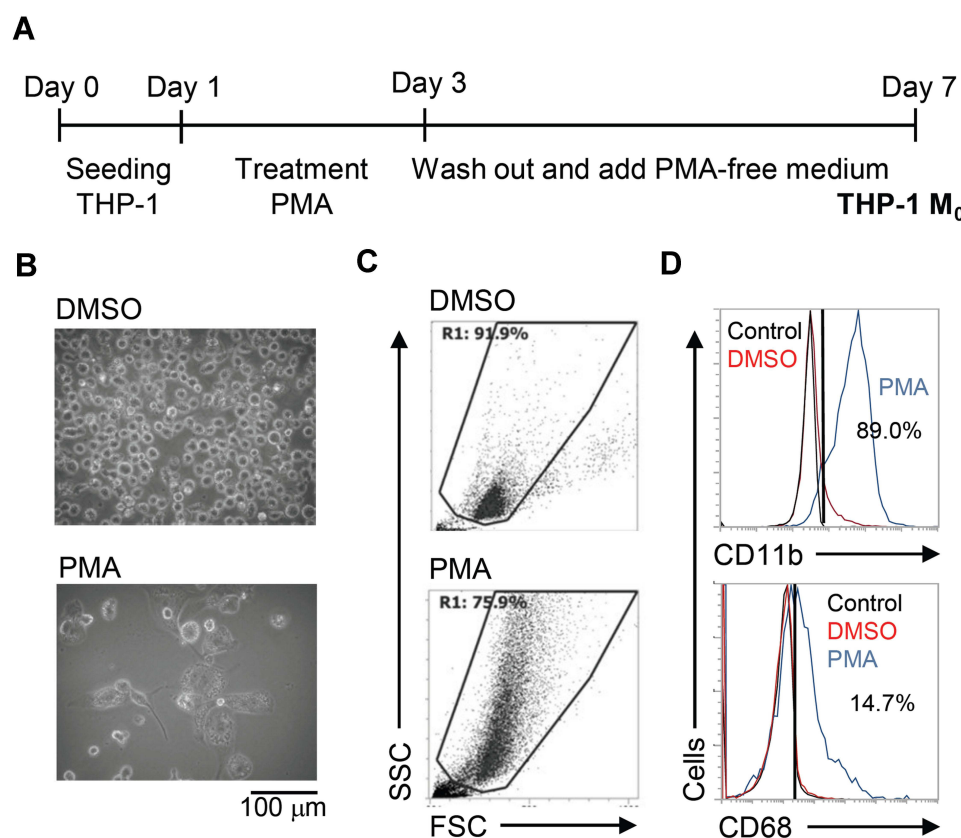
the differentiation of THP-1 cells toward a macrophage-like phenotype.

### Phorbol 12-Myristate 13-Acetate (PMA) Treatment Suppresses Cell Growth Accompanied by Cytotoxicity in THP-1-Differentiated Macrophages

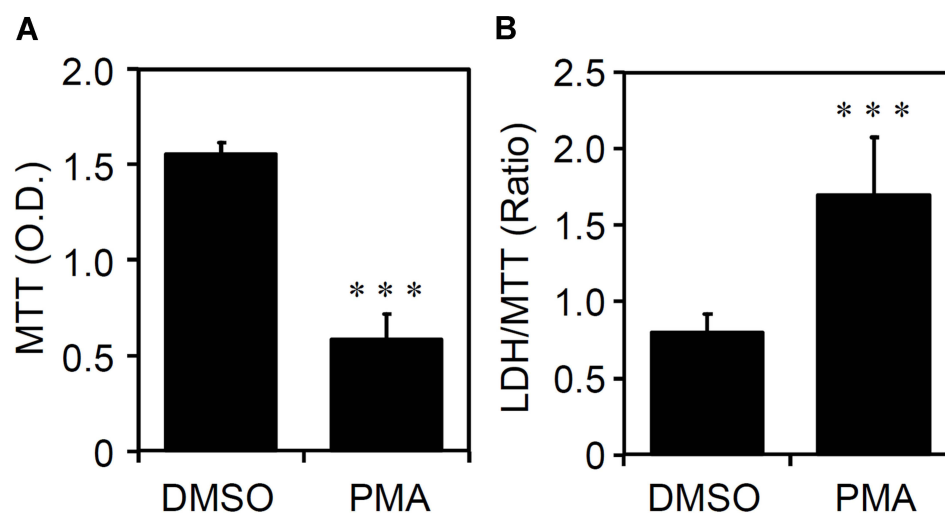
The cell fate of THP-1-differentiated macrophages shows heterogeneity of cell growth and apoptosis.<sup>18</sup> Through an MTT cell proliferation assay, which can measure the reduction of tetrazolium into an insoluble formazan product in viable cells,<sup>19</sup> PMA-stimulated THP-1 cells significantly (*p* < 0.05) exhibited a decrease in the cell growth rate (Figure 2A). Compared to the morphological observation, as shown in Figure 1A, the cell density was dramatically reduced in PMA-stimulated THP-1 cells. Among these cells, the induction of cytotoxicity is demonstrated by the formation of detached cells. By using an LDH cytotoxicity assay, which assesses the level of plasma membrane damage in a cell population,<sup>20</sup> PMA treatment also caused cellular injury in THP-1-differentiated macrophages (Figure 2B). The results show that PMA induces inducible macrophage differentiation accompanied by cell growth inhibition as well as cytotoxicity.

### Expression of PD-L1 (CD274) and PD-1 (CD279) in Phorbol 12-Myristate 13-Acetate (PMA)-Stimulated THP-1-Differentiated Macrophages

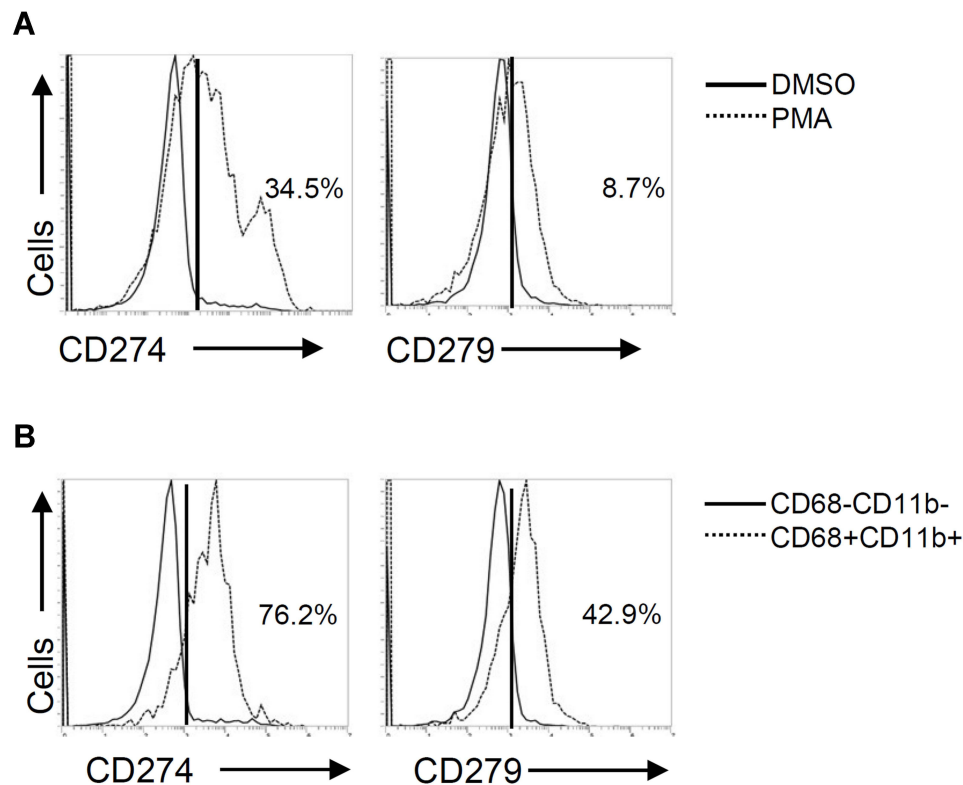
PD-L1 (CD274) and PD-1 (CD279) are expressed not only in cancer cells but also in immune cells, such as tumor-infiltrating lymphocytes and tumor-associated macrophages.<sup>21</sup> In PMA-stimulated THP-1 cells, flow cytometric analysis demonstrated considerably increased expression of both PD-L1 (CD274) and PD-1 (CD279) (Figure 3A). Compared to PD-1 (CD279), the expression of PD-L1 (CD274) was more inducible in THP-1-differentiated macrophages. As shown by the gating of CD68<sup>+</sup>CD11b<sup>+</sup> macrophages, the expression levels of PD-L1 (CD274) and PD-1 (CD279) were higher than those in the gating of CD68<sup>+</sup>CD11b<sup>+</sup> cells (Figure 3B). These results revealed the inducible expression of PD-L1 (CD274) and PD-1 (CD279) in THP-1-differentiated macrophages.



**Figure 1** Phorbol 12-myristate 13-acetate (PMA) treatment triggers macrophage differentiation in human monocytic THP-1 cells. **(A)** Experimental flowchart of the PMA stimulation performed in this study. **(B)** Cell morphology evaluation showed cell growth in PMA-treated THP-1 cells. **(C)** Flow cytometric dot-plot, plotting forward-scattered (FSC) versus side-scattered (SSC) from a population of THP-1 cells, showing the cell size and complexity. **(D)** Immunostaining followed by flow cytometric histogram analysis showed the expression of CD11b and CD68. Treatment of DMSO was used as control. For all images and flow cytometric analysis, representative staining data of isotype control, DMSO, and PMA were selectively obtained from three individual experiments. For the flow cytometric analysis, the percentage of positive cells in PMA treatment is shown.



**Figure 2** Phorbol 12-myristate 13-acetate (PMA) treatment causes cell growth inhibition and cytotoxicity in THP-1-differentiated macrophages. According to the experimental design shown in Figure 1A, **(A)** MTT and **(B)** lactate dehydrogenase (LDH)/MTT assays showed cell growth and cytotoxicity, respectively, in PMA-treated THP-1 cells. The original optical density (O.D.) of MTT and the ratio of LDH/MTT were shown. All quantitative data are shown as the mean  $\pm$  SD of three independent experiments. \*\*\* $p < 0.001$ .



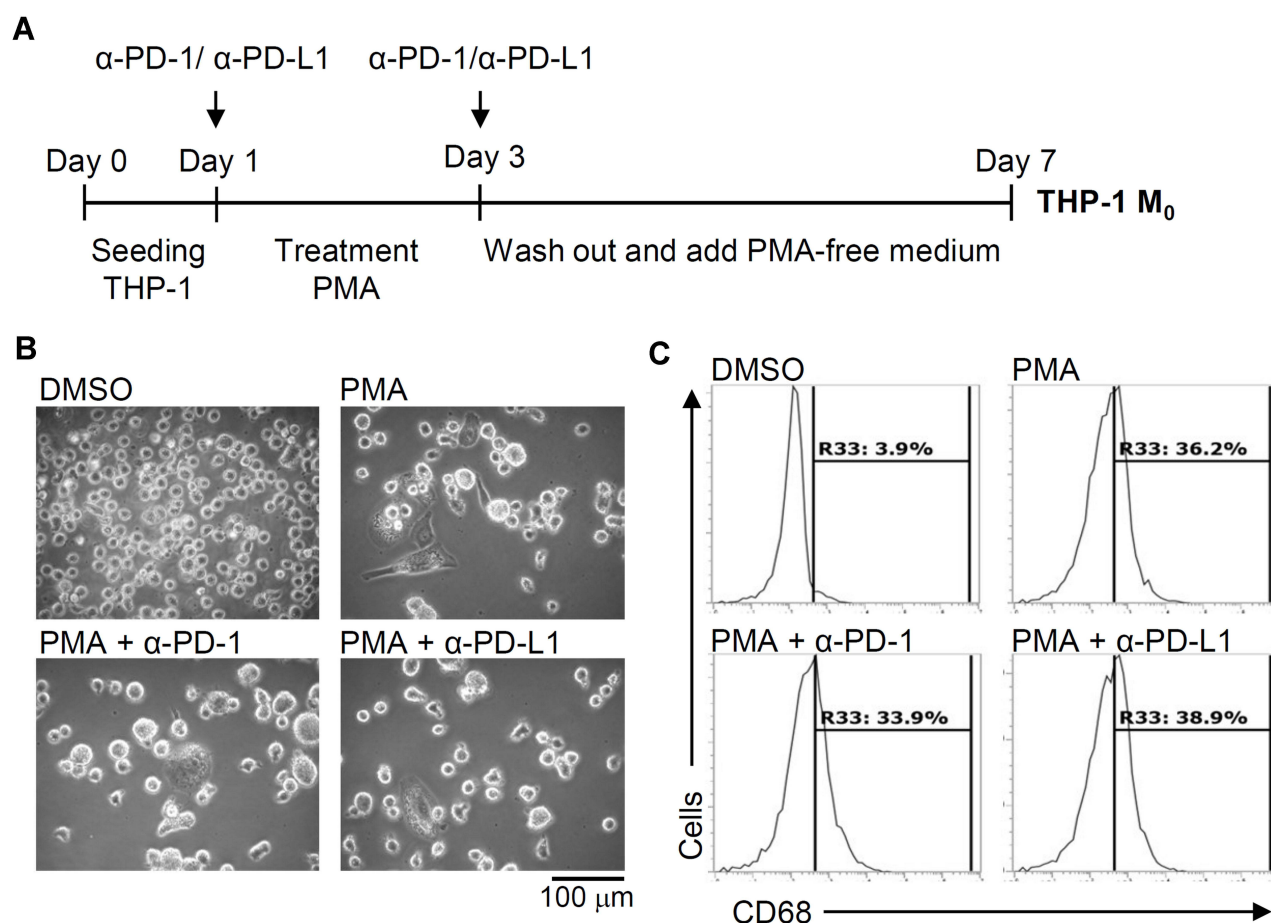
**Figure 3** Phorbol 12-myristate 13-acetate (PMA) treatment induces PD-L1 (CD274) and PD-1 (CD279) expression in THP-1-differentiated macrophages. According to the experimental design shown in Figure 1A, immunostaining followed by flow cytometric histogram and dot-plot analysis showed the expression of PD-L1 (CD274) and PD-1 (CD279) in PMA-treated THP-1 cells (A) without or (B) with the gating of CD68<sup>+</sup>CD11b<sup>+</sup> and CD68<sup>+</sup>CD11b<sup>+</sup> cells. Representative data were selectively obtained from three individual experiments, and the percentage of positive cells is shown.

### Blockade of PD-L1 (CD274) and PD-1 (CD279) Does Not Disturb Phorbol 12-Myristate 13-Acetate (PMA)-Induced Macrophage Differentiation

To investigate the role of inducible expression of PD-L1 (CD274) and PD-1 (CD279) in THP-1-differentiated macrophages, we next examined the blockade effects on PD-L1 (CD274) and PD-1 (CD279) using neutralizing antibodies as schematically represented in Figure 4A. Following PMA treatment concurrently incubated with anti-PD-L1 (CD274) and anti-PD-1 (CD279) neutralizing antibodies, no other blockade effects were identified in PMA-induced increased cell adherence and decreased cell density (Figure 4B). As shown by the flow cytometric analysis, the PMA-induced increased expression of CD68 was not affected by the blockade of PD-L1 (CD274) and PD-1 (CD279) (Figure 4C). The results illustrate inducible macrophage differentiation independent of the inducible expression of PD-L1 (CD274) and PD-1 (CD279) in THP-1-differentiated macrophages.

### Expression of PD-L1 (CD274) and PD-1 (CD279) and Its Roles in THP-1-Differentiated Macrophages M<sub>1</sub> and M<sub>2</sub>

We next investigated the different expression of inducible PD-L1 (CD274) and PD-1 (CD279) in THP-1-differentiated macrophages. Following the inducible THP-1-differentiated macrophages (M<sub>0</sub>), we used the combination treatment of LPS/IFN- $\gamma$  and IL-4/IL-13, respectively, to induce the polarization of M<sub>0</sub> into M<sub>1</sub> and M<sub>2</sub> as schematically represented in Figure 5A.<sup>8</sup> As shown by the flow cytometric analysis, the LPS/IFN- $\gamma$ - and IL-4/IL-13-induced increased expression of CD80 (M<sub>1</sub> marker) and CD209 (M<sub>2</sub> marker) were measured to show the different polarization. In THP-1 M<sub>1</sub> and THP-1 M<sub>2</sub> macrophages, the different expression of PD-L1 (CD274) and PD-1 (CD279) were shown (Figure 5B). The results elucidate the different expression of inducible PD-L1 (CD274) and PD-1 (CD279) in the different specializing macrophages while a decrease both in these checkpoints was found in THP-1-differentiated M<sub>2</sub> macrophages. To further



**Figure 4** Pharmacologically inhibiting PD-L1 (CD274) and PD-1 (CD279) does not affect phorbol 12-myristate 13-acetate (PMA)-induced macrophage differentiation. **(A)** In the absence and presence of neutralizing antibodies (5 μg/mL) against CD274 (α-PD-L1) and CD279 (α-PD-1), THP-1 cells were treated with PMA according to the experimental design. **(B)** Cell morphology showed cell growth. **(C)** Immunostaining followed by flow cytometric histogram analysis showed the expression of CD68. For all images and flow cytometric analysis, representative data were selectively obtained from three individual experiments, and the percentage of positive cells is shown.

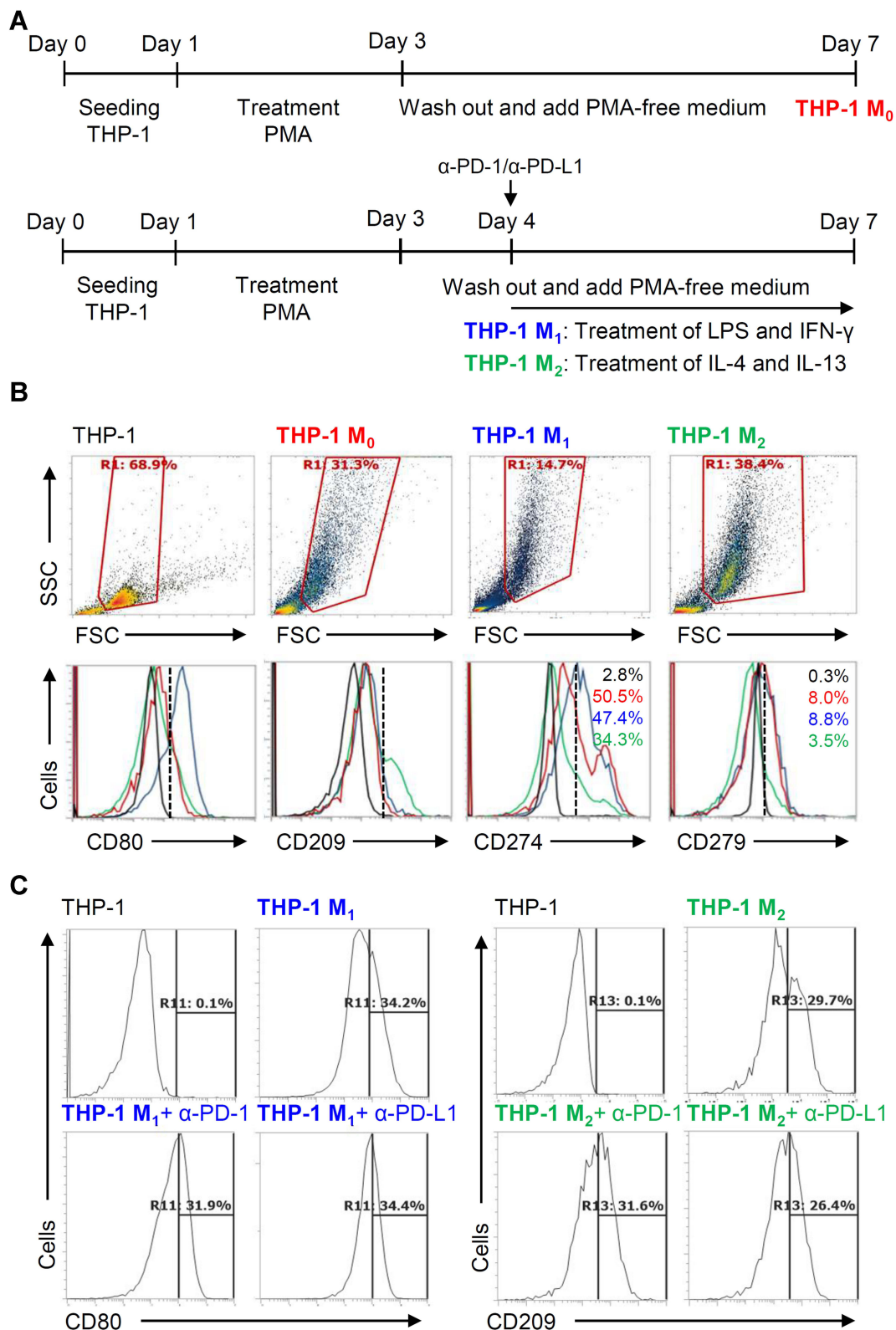
investigate the inhibition of PD-L1/PD-1 on the polarization of M<sub>1</sub> and M<sub>2</sub> macrophages, anti-PD-L1 (CD274) and anti-PD-1 (CD279) neutralizing antibodies were treated in the processing of THP-1-differentiated M<sub>1</sub> and M<sub>2</sub> macrophages (Figure 5C). The results showed that PD-1/PD-L1 was also not involved in macrophage polarization.

## Discussion

For anticancer immunotherapy, the blockade of immune checkpoints such as PD-L1 (CD274) and PD-1 (CD279) is currently and widely executed by targeting cancer cells and tumor-infiltrating lymphocytes in principle. However, tumor-associated macrophages also express PD-L1 (CD274) and PD-1 (CD279),<sup>14,15,21</sup> increasing alternative therapeutic efficacy by targeting these immune checkpoints for anticancer activity. In this study, by using an inducible model of macrophage

differentiation, we demonstrated increased expression of PD-L1 (CD274) and PD-1 (CD279) in PMA-activated human monocytic THP-1 cells. However, pharmacologically neutralizing PD-L1 (CD274) and PD-1 (CD279) did not affect THP-1 macrophage differentiation as well as polarization. Interestingly, a decrease both in these immune checkpoints in M<sub>2</sub> macrophages although the previous studies showed the blockade of PD-L1 (CD274) and PD-1 (CD279) increases macrophage phagocytosis as anticancer phenomena.<sup>15</sup> However, in this study, the biological effect of down-regulated PD-L1 (CD274) and PD-1 (CD279) in cytokine-driven THP-1 M<sub>2</sub> macrophages remains unclear. Future studies are urgently needed to verify the biological roles of PD-L1 (CD274) and PD-1 (CD279) in monocyte/macrophage differentiation.

No further pioneering study has investigated the regulation of PD-L1 (CD274) and PD-1 (CD279) expression



**Figure 5** Different expression of PD-L1 (CD274) and PD-1 (CD279) in THP-1-differentiated macrophages. **(A)** In PMA-stimulated THP-1-differentiated M<sub>0</sub> macrophages, cells were then treated with LPS (1 μg/mL)/IFN-γ (10 ng/mL) and IL-4 (25 ng/mL)/IL-13 (25 ng/mL) for polarization of M<sub>1</sub> and M<sub>2</sub>, respectively, in the absence and presence of neutralizing antibodies (5 μg/mL) against CD274 (α-PD-L1) and CD279 (α-PD-1) according to the experimental design. **(B)** For immunostaining, cells were stained with CD80 and CD209 for dissecting M<sub>1</sub> and M<sub>2</sub>, respectively. Immunostaining followed by flow cytometric histogram analysis showed the expression of CD274 and CD279 in these cells. **(C)** Furthermore, the expression of CD80 and CD209 in M<sub>1</sub> and M<sub>2</sub> without or with the blockade of CD274 and CD279 were shown. For all flow cytometric analysis, representative data were selectively obtained from three individual experiments, and the percentage of positive cells is shown. THP-1 (black); THP-1 M<sub>0</sub> (red); THP-1 M<sub>1</sub> (blue); THP-1 M<sub>2</sub> (green).

in monocyte/macrophage differentiation. Notably, recent studies illustrate that PD-L1 (CD274) is continually expressed on nonclassical monocytes at steady state to promote immunoregulatory function, probably by causing T cell death,<sup>22</sup> and the presence of PD-1 (CD279)-expressing intermediate monocytes is upregulated in pre-term cases of septic shock or fatal outcomes, showing an immunosuppressive role.<sup>23</sup> Classically activated M<sub>1</sub> macrophages express more PD-L1 (CD274) due to the presence of type 1 helper T cells and a combination of LPS/IFN- $\gamma$ .<sup>24</sup> In contrast, alternatively activated macrophages, which are polarized by IL-4 stimulation, slightly express PD-L1 (CD274) but effectively express PD-1 (CD279).<sup>14,15</sup> In THP-1-differentiated M<sub>0</sub> macrophages, both PD-L1 (CD274) and PD-1 (CD279) were evaluated. For macrophage polarization, the different expression in PD-L1 (CD274) and PD-1 (CD279) expression could be critical in polarized M<sub>1</sub> and M<sub>2</sub>, respectively.

Because PD-L1/PD-1 are the main immune checkpoint proteins on the surface of T cells for cancer immune escape,<sup>25</sup> the significance of macrophage-associated PD-L1/PD-1 in the relevance to macrophage differentiation and polarization is therefore speculated. However, the role of the evaluated expression of PD-L1 (CD274) and PD-1 (CD279) in M<sub>0</sub> macrophages remains unclear. Importantly, patient tumors with a high number of PD-L1 (CD274)-positive macrophages show favorable survival.<sup>14,26,27</sup> In contrast, PD-1 (CD279) expressed on macrophages is speculated to be immunosuppressive.<sup>14,15,26</sup> According to the current studies, signaling of PD-L1 (CD274) and PD-1 (CD279) may determine the different regulations on macrophage polarization, while PD-L1 (CD274) triggers CD86 and MHC II expression and PD-1 (CD279) promotes phagocytic inhibition.<sup>14</sup> The anti- and pro-tumor functions of PD-L1 (CD274) and PD-1 (CD279) have been demonstrated by their aberrant expression and functional regulation in macrophages; however, the mechanisms regulating their expression in macrophages remain unclear.

To control PD-L1 (CD274) and PD-1 (CD279) expression, many immune regulators and signaling pathways are responsible for their transcriptional induction.<sup>25,28</sup> For PD-L1 (CD274) expression, proinflammatory cytokines, including type I and type II IFNs, tumor necrosis factor  $\alpha$ , vascular endothelial growth factor, protein kinases, and transcription factors, have been reported.<sup>25,28</sup> PD-1 (CD279) is induced on activated T cells through mechanisms involving signaling of T-cell antigen receptors and cytokine receptors. In general, the nuclear factor of

activated T cells and IFN-regulating factors are required for the transcription of PD-1 in T cells.<sup>29</sup> Under PMA stimulation, the activation of mitogen-activated protein kinase signaling networks,<sup>30</sup> RhoA/ROCK signaling,<sup>31</sup> and several transcription factors<sup>32,33</sup> are essential for macrophage differentiation. Their potential regulation of the expression of PD-L1 (CD274) and PD-1 (CD279) is therefore proposed not only for macrophage differentiation but also for immune modulation.

Limitations of this work need further validation, including the selected differentiation models, the primary responses, and the biological roles of PD-L1/PD-1 in M<sub>0</sub> macrophages. Although PMA-differentiated THP-1 is commonly used in macrophage study, other physiological inducible models of macrophage differentiation, such as GM-CSF, M-CSF, and vitamin D<sub>3</sub>,<sup>11</sup> are needed to validate the expression of PD-L1 (CD274) and PD-1 (CD279). In conclusion, these works identified increased expression of PD-L1 (CD274) and PD-1 (CD279) in PMA-activated human monocytic THP-1 cells toward M<sub>0</sub> macrophages and a decreased PD-L1/PD-1 in M<sub>2</sub> macrophages. Although the blockade of PD-L1/PD-1 did not interfere with macrophage differentiation and polarization, as an inducible model of differentiated macrophages, more investigations on the expression and regulation of PD-L1/PD-1 may advance its physiopathological roles in macrophage intracellular regulation and activation toward M<sub>1</sub> and M<sub>2</sub>.

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## Disclosure

The authors report no conflicts of interest in this work.

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