

Progranulin Protects Against Airway Remodeling Through the Modulation of Autophagy via HMGB1 Suppression in House Dust Mite-Induced Chronic Asthma

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Purpose: Airway remodeling is an important feature of chronic asthma, and yet there are few effective therapeutic strategies. Progranulin (PGRN) has been shown to have lung protective functions, but the role of PGRN in asthmatic airway remodeling is unclear. We aim to explore the protective potential of PGRN on house dust mite (HDM)-induced airway remodeling and the underlying mechanisms.

Methods: In this study, a murine model of chronic asthma was established by HDM sensitization and challenge. Recombinant PGRN was intranasally administrated to mice during the phase of HDM challenge. TGF- β 1-treated human airway epithelial BEAS-2B cells were utilized to explore the effect of PGRN on airway epithelia exposed to profibrotic conditions and molecular mechanisms.

Results: We found that PGRN treatment attenuated HDM-induced airway remodeling, as evidenced by the suppression of collagen accumulation, mucus overproduction and airway smooth muscle synthesis in HDM-challenged asthmatic mice lungs. Meanwhile, PGRN also reversed the increased levels of autophagy markers and autophagosomes in airway epithelia under mimic asthmatic conditions, thereby controlling the fibrotic process in vivo and in vitro. Specifically, overexpressed HMGB1 and the subsequent RAGE/MAPKs signaling activation due to HDM exposure were abrogated in PGRN-treated asthmatic mice. Furthermore, knockdown of HMGB1 expression significantly restrained the fibrosis formation in TGF- β 1-induced airway epithelia accompanied by the downregulation of autophagic activity. However, enhancement of extracellular HMGB1 levels blunted the inhibition of autophagic flux by PGRN in airway epithelia, thereby resulting in the augmentation of collagen synthesis and fibrosis.

Conclusion: Taken together, our data revealed that PGRN protected against asthmatic airway remodeling by negatively regulating autophagy via HMGB1 suppression, which might provide new insights into the therapeutic options for HDM-induced chronic asthma.

Keywords: house dust mite-induced asthma, airway remodeling, progranulin, autophagy, HMGB1

Introduction

Asthma is a chronic inflammatory disease of the airways that is mainly caused by inhaled allergens such as house dust mite (HDM), or other external stimulus, leading to symptoms including cough, wheezing, dyspnea, or chest tightness.¹ These clinical manifestations of asthma are closely relevant to the structural changes of the airway

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wall, also known as airway remodeling.² Increased airway smooth muscle (ASM) mass, extracellular matrix (ECM) components and collagen deposition, epithelial-mesenchymal transition (EMT), reticular basement membrane thickening, and goblet cell metaplasia are identified as the typical pathophysiological traits of airway remodeling in asthma.³ Among the complicated mechanisms, TGF- β 1-induced fibrosis of airway walls by promoting matrix accumulation is a key contributor to airway structural remodeling.⁴ Therapeutically, although conventional asthma medications such as inhaled corticosteroid and long-acting β 2-agonists can reduce airway inflammation and hyperresponsiveness, airway fibrosis and remodeling cannot be effectively prevented in chronic asthma.⁵ Therefore, it is necessary to provide a better understanding of the pathogenesis of airway remodeling and explore new beneficial alternatives.

Progranulin (PGRN), a secretory glycoprotein encoded by the GRN gene, is widely expressed in a variety of tissues and cell types, including airway epithelial cells,⁶ macrophages⁷ and microglial cells.⁸ PGRN has pleiotropic properties by participating inflammatory response, regeneration, cell cycle control, and tissue development.^{9–12} Of interest, PGRN is recently regarded as a crucial regulator to prevent fibrotic process in different diseases, such as liver fibrosis,¹³ myocardial infarction¹⁴ and cutaneous wound healing.¹⁵ But the effect of PGRN on house dust mite-induced airway remodeling is still elusive. Whether PGRN modulates asthma-related airway fibrosis and what potential mechanisms remain to be elucidated.

Autophagy is a highly conserved catabolic process in which dysfunctional or excessive proteins and organelles are recycled or delivered to maintain cellular homeostasis through the lysosome-mediated degradation pathway. Emerging studies have suggested that autophagy is linked with several inflammatory diseases due to its potential to modulate genetic and immune mechanism. Of note, autophagy dysregulation plays a pivotal role in the process of fibrotic disorders including asthma airway fibrosis.^{16–18} Autophagy pathway activation in the airways from asthmatics is highly associated with the development of airway remodeling.¹⁹ High mobility group box 1 (HMGB1) is an essential damage-associated molecular pattern (DAMP) protein expressed in various cell types. Extracellular HMGB1 plays critical roles in cell death and injury through acting with the domain receptors, such as receptor for advanced glycation end-products (RAGE) and toll-like receptors (TLRs), leading to mitogen-activated protein

kinase (MAPK) and nuclear factor (NF)- κ B activation.²⁰ HMGB1 release has been reported to crucially involve in the regulation of autophagy in multiple diseases.^{21,22} Our previous study has found that HMGB1 signaling activation is responsible for HDM-induced airway damage. However, the relationship between HMGB1-related autophagy and airway remodeling in HDM-induced chronic asthma under PGRN treatment is poorly reported to date.

In this study, by using a murine model of HDM-induced chronic asthma *in vivo* and TGF- β 1-induced airway epithelial cells *in vitro*, we sought to explore the roles of PGRN in asthmatic airway remodeling and autophagy, including its mechanism. Our findings demonstrated that PGRN was a protective regimen for asthmatic airway remodeling and fibrosis by modulating autophagy via HMGB1 suppression, indicating PGRN might be a potential therapeutic strategy for HDM-induced asthma.

Materials and Methods

Animal Model of Chronic Asthma

Healthy female Balb/c mice at age of 6–8 weeks old (purchased from Changzhou Cavens Laboratory Animal Co. Ltd.) were maintained under specific pathogen-free conditions. All animal protocols were performed in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals. The animal study was reviewed and approved by the Animal Care and Use Committee of Tongji University. To investigate the protective effects of PGRN against HDM-induced airway remodeling, mice were randomly divided into the control group, the HDM group and the HDM+PGRN group ($n=8/\text{group}$). The mice in HDM group were sensitized with intraperitoneal (i.p.) injection of 50 μ g HDM (cat. no. XPB70D3A2.5, Greer Labs, Lenoir, NC, USA) with aluminum hydroxide (Sigma-Aldrich, St. Louis, MO, USA) on days 0, 7 and 14, and then challenged by intranasal (i.n.) administration of 10 μ g HDM from day 21 three times a week for 4 consecutive weeks. The mice in HDM+PGRN group were intranasally administrated with recombinant PGRN (ProSpec, Rehovot, Israel) (2 μ g dissolved in 20 μ l PBS) 30min prior to each inhalation of HDM during challenge phase. The mice in the control group were just treated with PBS instead of HDM or PGRN. All mice were sacrificed 24h after the last challenge. The bronchoalveolar lavage fluid (BALF) and lung tissues were collected to analyze inflammatory cells counts and histopathological features, respectively.

Cell Culture and Treatments

Human airway epithelial cell line BEAS-2B was purchased from Cell Bank of the Chinese Academy of Sciences (Shanghai, China). Cells were cultured in DMEM (Hyclone, Camarillo, CA, USA) containing 10% fetal bovine serum (Gibco, Grand Island, NY, USA), 1% penicillin-streptomycin (Gibco) at 37°C in 5% CO₂. To identify the effect of PGRN on airway fibrosis, the cells were pretreated with recombinant PGRN, and then (2 hours later) incubated with 10ng/mL TGF- β 1 (Peprotech, Rocky Hill, NJ, USA) for another 24h or 48h respectively. Phosphate-buffered saline (PBS) was used as negative control.

Cell Transfection

The siRNAs targeting HMGB1 gene (siRNA-HMGB1) and siRNA-NC were purchased from GenePharma Company (Shanghai, China). BEAS-2B cells were planted in 6-well (2×10^5) and cultured overnight before transfection. Then, the cells were subjected to the transient transfection by the Lipofectamine 2000 (Invitrogen, Camarillo CA, USA) according to the manufacturer's instructions. All treated BEAS-2B cells were collected for further different experiments.

Analysis of Lung Morphology

Lung tissue samples were in 4% formaldehyde and then embedded in paraffin. The tissue sections (5 μ m) were stained with haematoxylin and eosin (H&E), periodic acid Schiff (PAS) and Masson's trichrome. The images were obtained by a light microscope (Olympus X73, Tokyo, Japan). The lung inflammation score was calculated as previously described.²³

Western Blot Analysis

Lung tissues or cells were washed by PBS and harvested with RIPA. The protein lysates were electrophoresed in SDS-PAGE gel and then transferred onto PVDF membranes. After blocked with 5% BSA, the membranes were incubated overnight at 4°C with primary antibodies against α -smooth muscle actin (α -SMA) (1:1000; ab21027), Collagen-I (1:1000; ab260043) collagen-III (1:1000; ab184993), RAGE (1:1000; ab216329), p62 (1:1000; ab56416) (Abcam, Cambridge, MA, USA), LC3 (1:1000; 4108S), HMGB1 (1:1000; 6893), phospho(p)-JNK (1:1000; 4668), JNK (1:1000; 9252), p-p38 (1:1000; 9211), p38 (1:1000; 9212), p-ERK1/2 (1:1000; 9101),

ERK1/2 (1:1000; 8544) (Cell Signaling Technology, Danvers, MA), Beclin-1 (1:1000; 11306) (Proteintech, Rosemont, IL, USA) and GAPDH (1:1000; 5174) (Cell Signaling Technology). Then the membranes were rinsed in TBST and incubated with HRP-conjugated anti-rabbit secondary antibodies for 2h at room temperature. ECL system was used to detect the protein blots.

Immunohistochemical Staining

Lung tissues were fixed with 4% paraformaldehyde and incubated with primary antibodies against α -SMA (1:500; ab32575), Collagen-I (1:500; ab34710), Beclin-1 (1:500; ab62557) (Abcam) or LC3 (1:500; 3868) (Cell Signaling Technology) overnight at 4°C, and then incubated with according secondary antibodies for 1h at room temperature. Finally, the slides were counterstained with hematoxylin. The captured images were analyzed using an Image Pro Plus 4.5.

Immunofluorescence Staining

The immunofluorescence staining was performed to detect the expression levels of indicated proteins in cells and lung tissues. Briefly, BEAS-2B cells were incubated with the primary anti- α -SMA antibody (1:300; ab32575) (Abcam) or anti-LC3 antibody (1:500; 18725-1-AP) (Proteintech) overnight (4°C). Followed by washing with PBS, the samples were stained with goat anti-rabbit fluorophore-conjugated secondary antibodies for 1h at room temperature. The nuclei were counterstained with DAPI. The images were obtained by Leica fluorescence microscope.

Transmission Electron Microscopy (TEM)

Cells from different groups were prefixed with 2.5% ice-cold glutaraldehyde for 1 h. After rinsed by PBS, the samples were pro-fixed with 1%OsO₄ and embedded in pure acetone and resin embedding fluid (2:1). Then ultra-thin sections (50–60nm) were double stained with 4% uranyl acetate and lead citrate. Transmission electron microscopy was applied to detect the formation of autophagosomes in airway epithelial cells.

mRFP-GEP-LC3 Adenovirus Transduction

An mRFP-GEP-LC3 dual-fluorescence assay was performed to determine the autophagic flux. Briefly, when the cells reached nearly 60% confluence, they were transfected with mRFP-GEP-LC3 adenovirus (Genomeditech, Shanghai, China) prior to the indicated

experiments. The cells images were captured to demonstrate the autophagic flow by a confocal microscope (Olympus FV 1000, Tokyo, Japan).

Procedures for the measurements of quantitative reverse transcription PCR (qRT-PCR), flow cytometric analysis and enzyme-linked immunosorbent assay (ELISA) are presented as online [Supplemental Materials](#).

Statistical Analysis

Data were presented as mean±standard error of the mean (SEM). Multiple groups' comparisons were performed using one-way ANOVA with the Bonferroni's post hoc test. Statistical analysis was conducted using SPSS19.0. $P < 0.05$ was considered as statistically significant.

Results

PGRN Administration Mitigates HDM-Induced Airway Inflammation and Remodeling in Mice

To determine the function of PGRN in allergic airway inflammation and remodeling, a HDM-induced murine asthma model was utilized (Figure 1A). Compared to the control group, HDM-challenged asthmatic mice had a remarkable enhancement in inflammatory cells infiltration and airway wall thickened, while PGRN administration effectively alleviated these histopathological changes in asthmatic mice (Figure 1B and C). Accordingly, lung inflammation score was also lower in HDM+PGRN group than those in HDM alone group (Figure 1D).

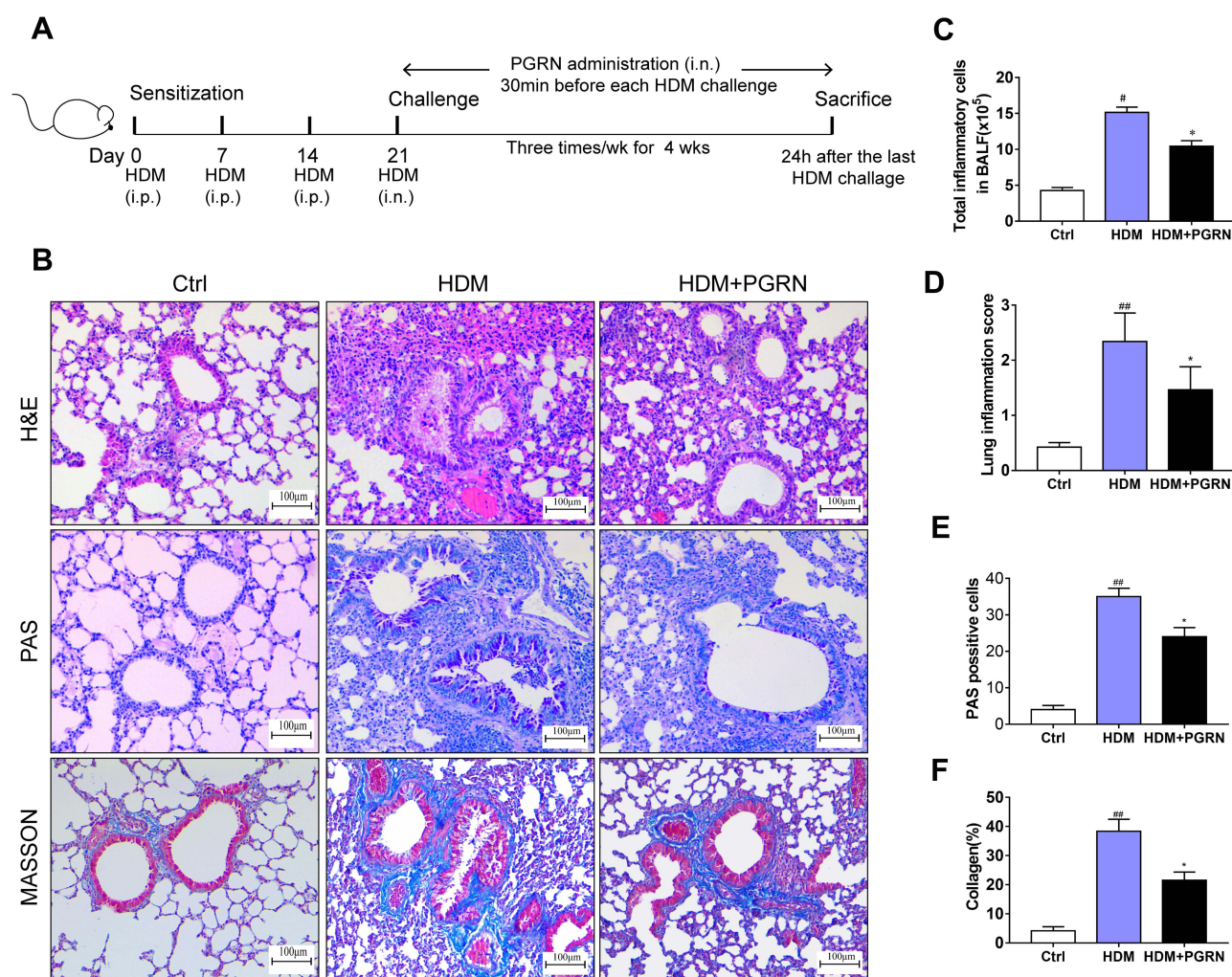


Figure 1 Effects of PGRN on airway remodeling in HDM-challenged asthmatic mice. **(A)** Schematic diagram of the experimental plan for HDM-induced murine asthma. **(B)** H&E, PAS and Masson's trichrome staining of lung sections from different groups. **(C)** The counts of total inflammatory cells in BALF from mice analyzed by Wright staining. **(D)** Levels of inflamed lung areas in mice. **(E)** Quantification of PAS-positive mucous cells in airways of each group. **(F)** Analysis of collagen deposition in airway tissues. Quantitation of lung morphology analysis was based on 5 random areas from each sample. Data are expressed as means ± SEMs of three independent experiments with four mice per group. [#] $p < 0.05$ vs control group, ^{##} $p < 0.01$ vs control group, ^{*} $p < 0.05$ vs HDM group.

Compared with HDM+PGRN group, HDM-challenged mice had a stronger PAS-positive staining in airway tissues which represented mucus overproduction and goblet cell hyperplasia (Figure 1B and E). Additionally, in contrast to the control group, Masson's trichrome staining showed that collagen deposition and airway wall thickness were obviously increased in HDM-induced asthmatic mice, which were significantly inhibited by PGRN treatment (Figure 1B and F). For another, PGRN effectively reduced HDM-mediated Th2-type allergic airway inflammation (Supplementary Figure S1A–C). Since airway remodeling is characterized by myofibroblast differentiation, subepithelial fibrosis, and increased ECM deposition, we further assessed the expression of fibrosis-related markers α -SMA and Collagen-I in lung tissues by immunohistochemistry analysis. Our findings showed that α -SMA and Collagen-I were markedly expressed in peribronchial tissues of asthmatic mice in contrast to those in the control group. However, compared with the HDM group, the expression of α -SMA and Collagen-I was significantly decreased in the HDM+PGRN group (Figure 2A–C). Meanwhile, the elevated mRNA levels of fibrosis-related

genes α -SMA, Collagen-I, TGF- β 1 and MMP9 (Figure 2D–G) in HDM-challenged mice were markedly repressed by PGRN treatment. Collectively, the results indicated that PGRN had an ability to attenuate airway inflammation and remodeling in HDM-induced asthma model.

PGRN Suppresses Fibrotic Process in TGF- β 1-Treated Human Airway Epithelial Cells

To identify the effect of PGRN on airway epithelia under the mimic fibrotic conditions in vitro, human airway epithelial BEAS-2B cells were treated with variable concentrations PGRN for 24h under the induction of 10ng/mL TGF- β 1, a known profibrotic factor to induce myofibroblast differentiation. Our results showed that PGRN (20, 50, or 100ng/mL) dose-dependently inhibited the expression of TGF- β 1-induced fibrosis markers α -SMA, Collagen-I and Collagen-III. (Supplementary Figure S2A–D). BEAS-2B cells exposed to PGRN also revealed the mRNA upregulation of EMT biomarker E-cadherin accompanied by the reduction of N-cadherin (Supplementary Figure S2E). Then, BEAS-2B cells were incubated with TGF- β 1 for different time duration under 100ng/mL

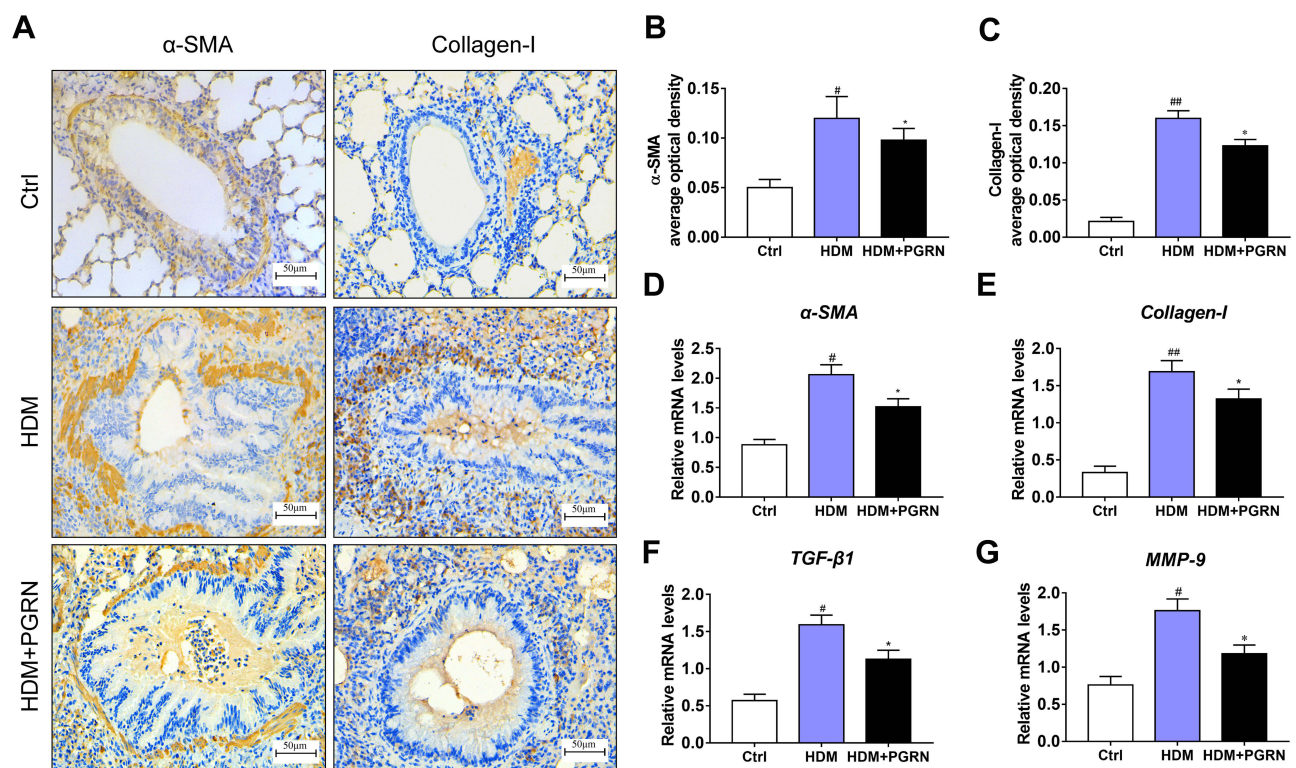


Figure 2 PGRN restrains HDM-induced collagen synthesis and fibrosis in mice. (A) Protein expression of α -SMA and collagen-I in mice was detected by immunohistochemical staining. (B and C) α -SMA and collagen-I expression levels according to average optical density. The relative mRNA expression of fibrosis-related genes including α -SMA (D), collagen-I (E), TGF- β 1 (F), MMP-9 (G) were measured by qRT-PCR. For immunochemistry assay, results are obtained from 5 random areas of each sample. Data are expressed as means $\pm$ SEMs of three independent experiments with four mice per group. # p <0.05 vs control group, ## p <0.01 vs control group, * p <0.05 vs HDM group.

PGRN treatment or not. As illustrated in Figure 3A and B, compared with the controls, the expression of fibrosis-related markers α -SMA, Collagen-I and Collagen-III was dramatically upregulated in TGF- β 1-treated cells in a time-dependent manner. Meanwhile, PGRN treatment decreased the protein levels of these markers evoked by TGF- β 1 stimulation. Furthermore, immunofluorescence staining was applied to assess α -SMA expression in BEAS-2B cells with different treatments. Compared to the control, α -SMA positive cells were apparently increased in response to TGF- β 1 exposure, especially in the 48h TGF- β 1-treated group. However, TGF- β 1-induced α -SMA overexpression was partly reduced in the cells with PGRN treatment (Figure 3C and D). These data suggested that PGRN could restrain TGF- β 1-mediated fibrosis by decreasing the expression of ECM components in airway epithelia.

PGRN Downregulates Autophagic Activity in Asthmatic Mice and Human BEAS-2B Cells

In order to evaluate whether PGRN could regulate autophagy associated with asthmatic airway remodeling and fibrosis, we further examined the expression of acknowledged autophagy markers LC3 and Beclin-1 in different mice lungs. Immunohistochemistry analysis revealed that LC3 and Beclin-1 were highly expressed in the airways of HDM-challenged asthmatic mice. Few staining with anti-LC3 and anti-Beclin-1 from control mice were observed. Compared with HDM group, the asthmatic mice receiving PGRN administration had moderate expression of the two autophagy markers in airway tissues (Figure 4A–C). In addition, the degree of Beclin-1 and LC3 expression was concomitant with the pathological changes of airway tissues in the mice. In vitro, for autophagy

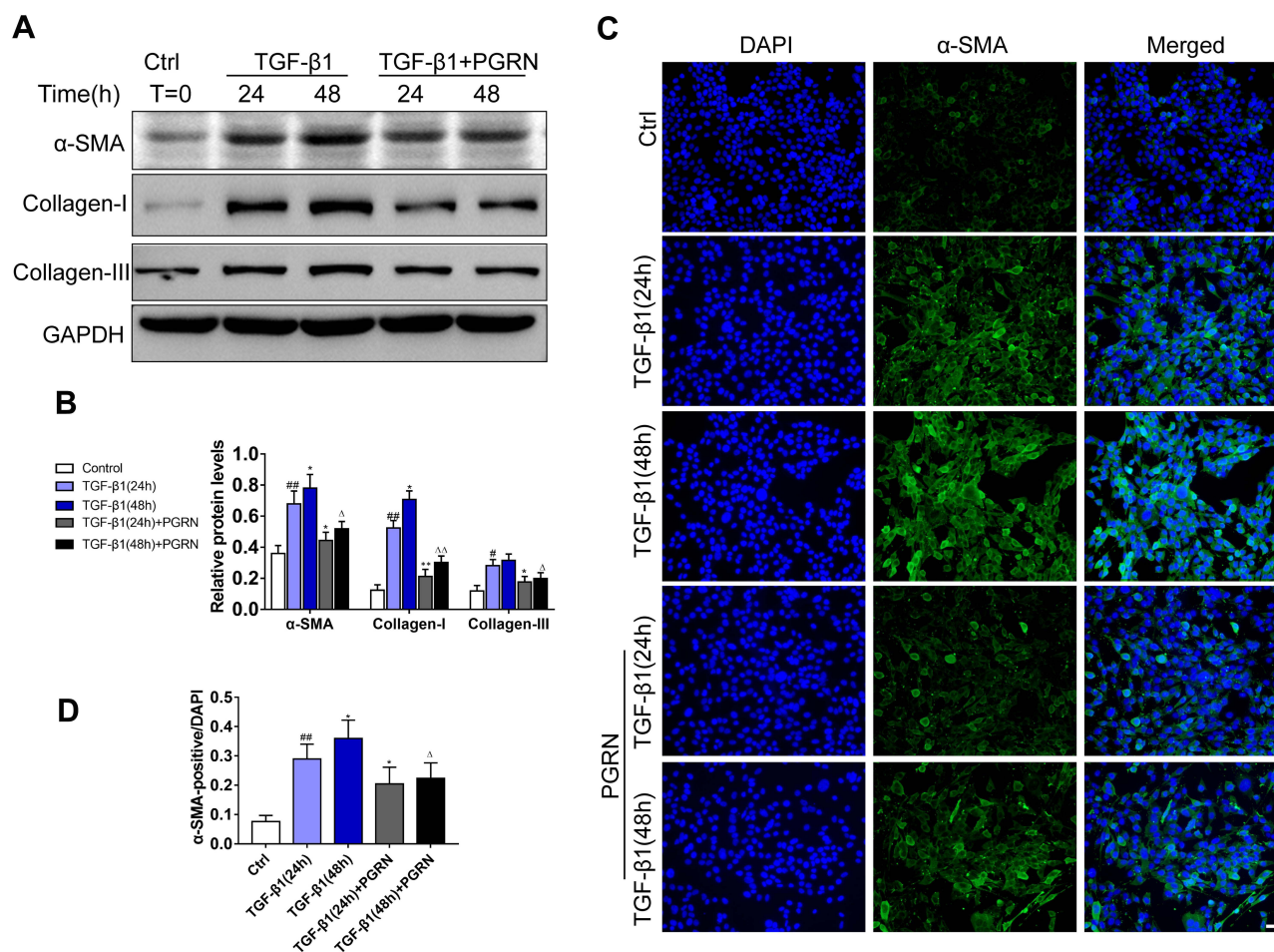


Figure 3 PGRN inhibits TGF- β 1-mediated fibrotic process in BEAS-2B cells. **(A)** After the induction by 48h or 72h TGF- β 1 (10ng/mL) with or without PGRN, Western blotting was performed to detect the expression of α -SMA, collagen-I and collagen-III. **(B)** Grey values of the indicated proteins were shown as bar graphs. **(C)** α -SMA expression (green) was detected by immunofluorescence assay in BEAS-2B cells under different treatments. **(D)** The ratio of α -SMA positive cells from different groups. Data are expressed as means $\pm$ SEM of three independent experiments. [#] p <0.05 vs control group, ^{*} p <0.05 vs control group, ^{##} p <0.01 vs control group, ^{*} p <0.05 vs TGF- β 1 (24h) group, ^{**} p <0.01 vs TGF- β 1 (48h) group, ^Δ p <0.05 vs TGF- β 1 (48h) group, ^{ΔΔ} p <0.01 vs TGF- β 1 (48h) group.

assay, the ultrastructural assessment of epithelial cells was analyzed by TEM. Figure 4D and E displayed that PGRN treatment reversed the increased numbers of autophagosomes in BEAS-2B cells under 48h TGF- β 1 induction. In parallel, we found that increased apoptotic cell death following TGF- β 1 treatment was also restrained by PGRN (Figure 4F and G). These findings confirm the negative regulation of PGRN on autophagy which takes part in asthmatic fibrosis and airway remodeling.

PGRN Blocks HDM-Induced HMGB1 Expression and the Downstream RAGE-MAPKs Pathways in Mice

Previous studies have verified that HMGB1 plays a role in the modulation of autophagy as an essential DAMP molecular.^{24,25} To explore the key role of HMGB1 in

asthma-related autophagy and whether inhibition of autophagy by PGRN is linked with HMGB1-mediated signaling, we evaluated HMGB1 expression and its potential signaling pathway in mice lungs under different groups. As expected, the protein levels of HMGB1 in HDM-challenged asthmatic mice lungs were significantly higher than those in the control group, whereas markedly diminished in PGRN-treated mice. Of note, RAGE is believed as a usual kind of pattern recognition receptors (PRRs) binding HMGB1. HMGB1/RAGE axis is critically implicated in the modulation of inflammation, autophagy and fibrosis via the downstream MAPKs pathway in asthma pathogenesis.²⁶ To further test the possible signaling transduction of HMGB1 in PGRN-treated asthmatic mice, we then examined the changes of RAGE, JNK, ERK1/2 and p38. The immunoblotting analysis showed that upregulation of RAGE expression triggered by HDM was inhibited

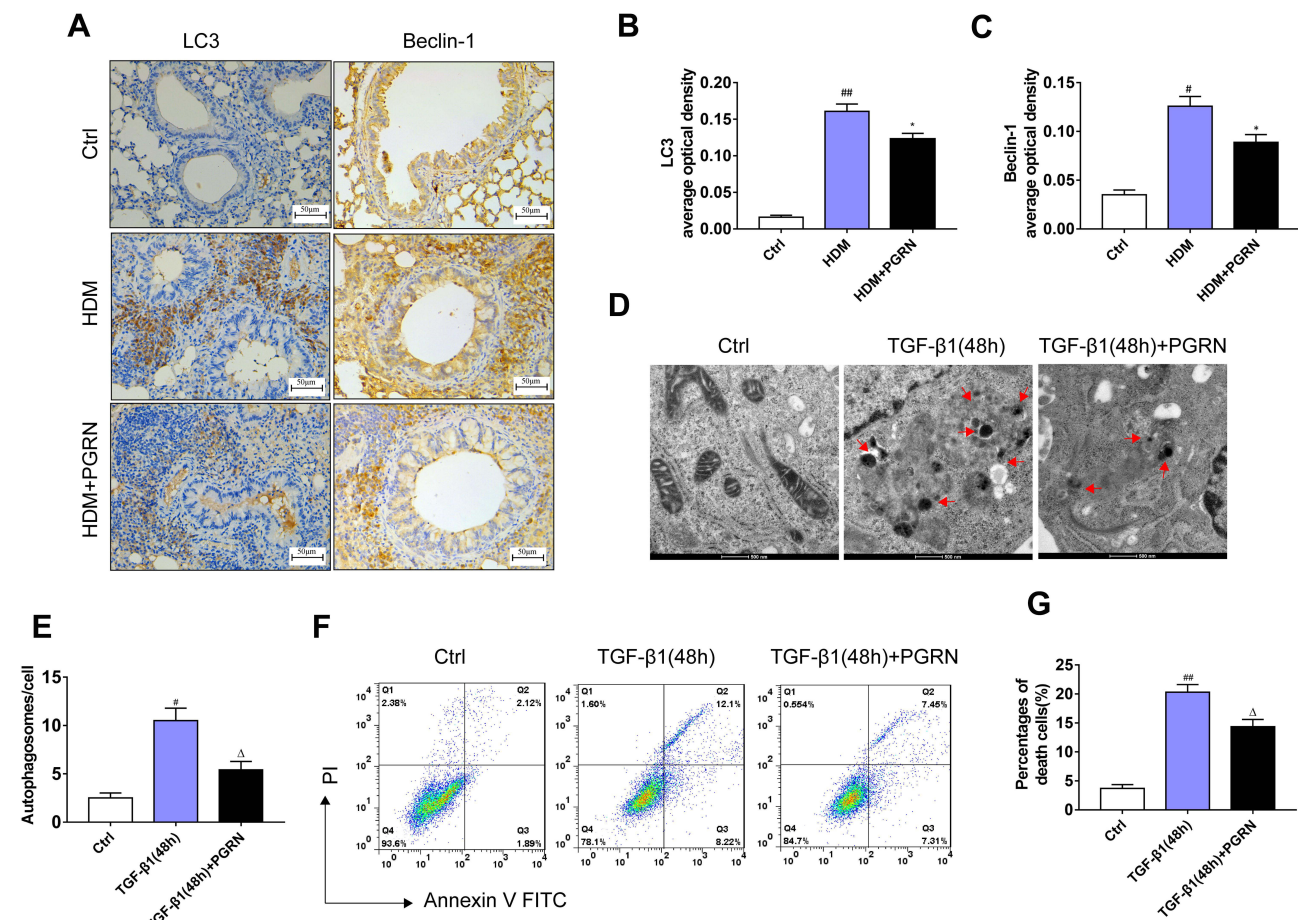


Figure 4 PGRN reduces airway autophagy in asthmatic mice and TGF- β 1-treated BEAS-2B cells. (A) LC3 and Beclin-I expression in airway tissues of mice was assessed by immunohistochemistry. (B and C) Semi-quantitative analysis showed that PGRN significantly decreased the expression levels of LC3 and Beclin-I in mice exposed to HDM. (D) Representative TEM images of autophagosomes (red arrows) in TGF- β 1-induced BEAS-2B cells with PGRN treatment or not. Scale bar=500nm. (E) Quantification of the numbers of autophagosomes in cells. (F) Apoptosis of cells was measured by the FITC-annexin V/PI assay. (G) Flow cytometry indicated that PGRN reduced apoptotic cell death in TGF- β 1-treated cells. For analysis of immunohistochemistry assay, results are pooled from 5 random areas with four mice per group. Data are expressed as means $\pm$ SEM of three independent experiments. ^{##} $p < 0.05$ vs control group, ^{##} $p < 0.01$ vs control group, ^{*} $p < 0.05$ vs HDM group, ^Δ $p < 0.05$ vs TGF- β 1 (48h) group.

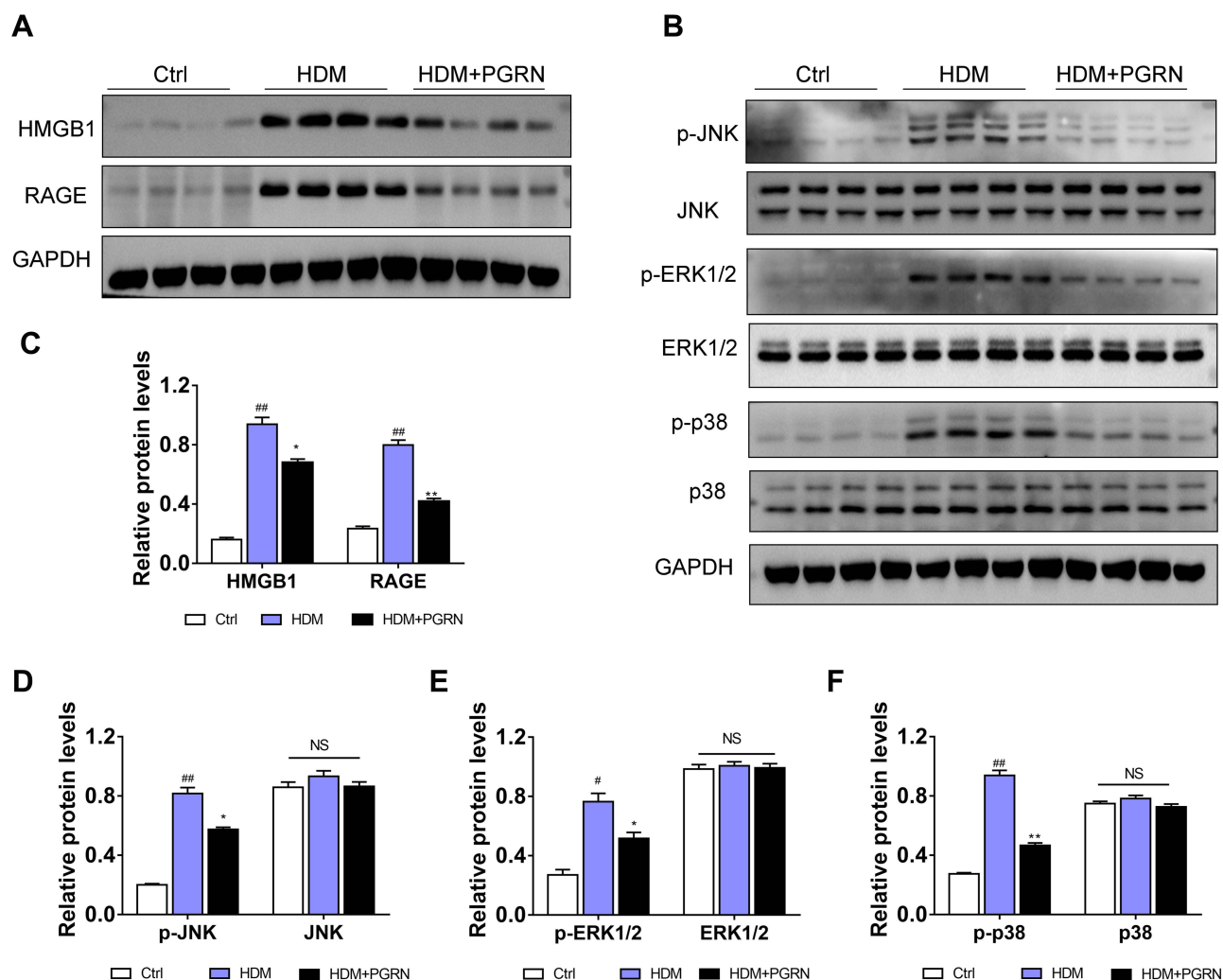


Figure 5 Effect of PGRN on HMGB1, RAGE and MAPK pathways in HDM-challenged mice. **(A)** PGRN downregulated HMGB1 and RAGE protein expression in mice lungs upon HDM exposure. **(B)** The effect of PGRN on phosphorylated and total JNK, ERK1/2, and p38 expression were evaluated by immunoblotting analysis. **(C–F)** Quantitative densitometric ratio of the above proteins normalized to GAPDH in each group. For immunoblotting analysis, data are expressed as means±SEM of three independent experiments with four mice per group. [#] $p<0.05$ vs control group, ^{##} $p<0.01$ vs control group, ^{*} $p<0.05$ vs HDM group, ^{**} $p<0.01$ vs HDM group.

after PGRN treatment. HDM-elicited phosphorylations of JNK, ERK1/2 and p38 were obviously decreased in the PGRN-treated asthmatic mice (Figure 5). These results indicated that PGRN treatment blocked HMGB1 expression and its downstream signaling during HDM-induced allergic asthma.

HMGB1 is Required for the Suppressive Effect of PGRN on Autophagy Under Fibrotic Conditions

To ascertain whether HMGB1 is involved in the anti-fibrosis functions of PGRN on airway epithelia, we first deleted HMGB1 via siRNA. After knockdown of HMGB1 expression, the BEAS-2B cells were exposure to 10ng/mL TGF- β 1 for 48h with or without 100ng/mL PGRN

treatment. We first evaluated the role of HMGB1 in autophagy of airway epithelia following TGF- β 1 exposure. The immunoblotting analysis revealed that HMGB1 deficiency crucially restrained autophagy activation, shown as the downregulation of autophagy proteins LC3, Beclin-1 and p62, and further suppressed overexpression of α -SMA, collagen-I and collagen-III induced by TGF- β 1 (Figure 6A and B). Moreover, the double immunofluorescent staining was used to assess the degree of fibrosis together with autophagic activity. As illustrated in Figure 6C and D, the staining intensity of dual-labeled LC3 (red) and α -SMA (green) was substantially decreased in the TGF- β 1+PGRN co-culture groups compared with TGF- β 1-treated alone groups, which were in line with our in vivo results. Particularly, in the presence of PGRN

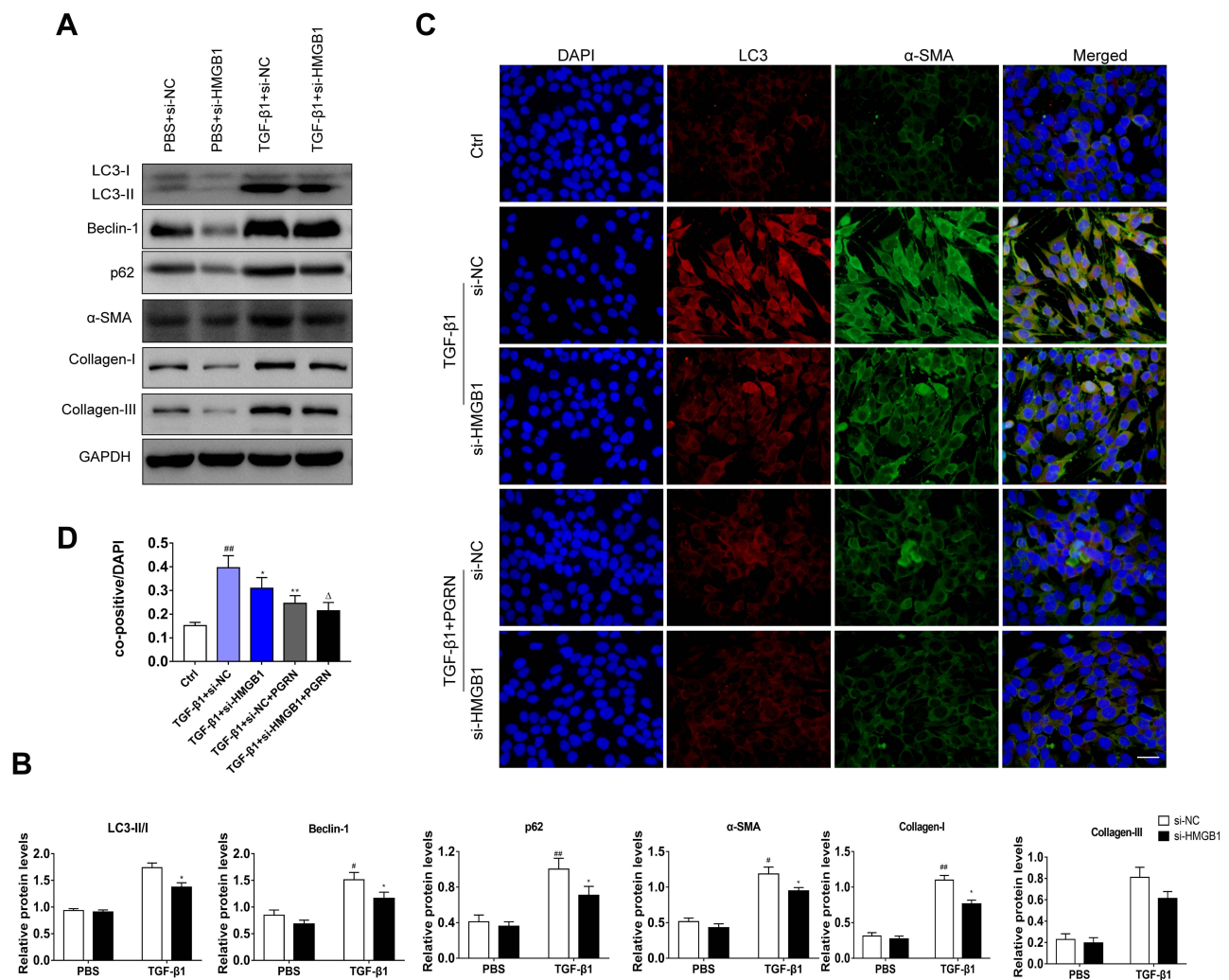


Figure 6 HMGB1 is responsible for the negative modulation of PGRN on autophagy under fibrotic conditions. (A) After silencing HMGB1, the expressions of autophagy (LC3, p62, Beclin-1) and fibrosis markers (α -SMA, collagen-I, collagen-III) in BEAS-2B cells were measured by Western blot. (B) Greyscale analysis of the above proteins. (C) Double immunofluorescence staining with LC3 and α -SMA in TGF- β 1-treated BEAS-2B cells transfected with si-HMGB1 or si-NC in the presence or absence of PGRN. (D) Co-dying data (LC3 and α -SMA) was summarized as bar graph. Scale Bar=100 μ m. Data are expressed as means $\pm$ SEM of three independent experiments. [#] $p<0.05$ vs control group, ^{##} $p<0.01$ vs control group, ^{*} $p<0.05$ vs TGF- β 1+si-NC group, ^{**} $p<0.01$ vs TGF- β 1+si-NC group, ^Δ $p<0.05$ vs TGF- β 1+si-HMGB1 group.

treatment, the cells transfected with si-HMGB1 had weaker co-staining of LC3 and α -SMA elicited by TGF- β 1 stimulation than the si-NC group. In this regard, HMGB1 contributed to TGF- β 1-mediated fibrosis as a possible target of PGRN. To further clarify the relationship between HMGB1, PGRN, fibrosis and autophagy, we pretreated BEAS-2B cells with 500ng/mL recombinant HMGB1 (rHMGB1) (Sigma-Aldrich, Stockholm, Sweden) or PBS vehicle for 12h prior to treatment with TGF- β 1 in the presence or the absence of PGRN. Under the same fibrotic conditions, BEAS-2B cells were additionally subjected to 5mM autophagy inhibitor 3-methyladenine (3-MA) (Sigma-Aldrich, St. Louis, USA). As expected, PGRN treatment reversed the increased

expression of autophagy proteins, thereby reducing collagen accumulation and fibrosis, which was similar with the results under 3-MA treatment. Nevertheless, rHMGB1 pretreatment aggravated autophagic activity in airway epithelia exposed to TGF- β 1, thus facilitating the fibrotic process. In addition, the negative effects of PGRN on autophagy and fibrosis upon TGF- β 1 insult were obviously blunted in the cells pretreated with rHMGB1 (Figure 7A and B). Meanwhile, PGRN-exerted inhibitory effect on autophagy flux was significantly reversed by rHMGB1 treatment (Figure 7C and D). Correspondingly, the decreased counts of autophagosomes as well as apoptotic cell death in PGRN-treated cells were also compromised after rHMGB1 treatment (Supplementary Figure S3A and

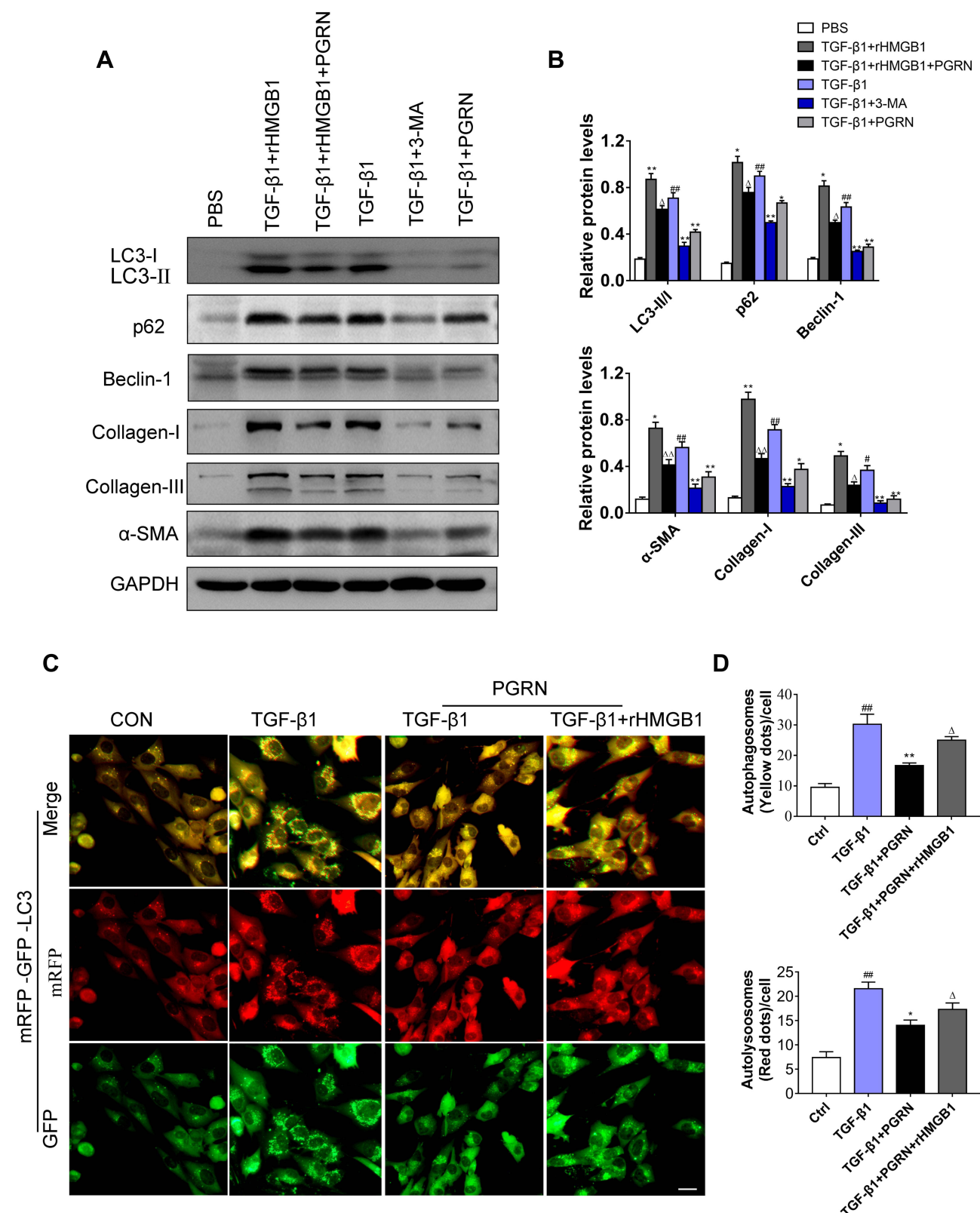


Figure 7 Increased extracellular HMGB1 offsets the inhibition of PGRN on TGF-β1-driven fibrosis through facilitating autophagy. **(A)** Immunoblotting analysis showed that PGRN or autophagy inhibitor 3-MA restrained expression of autophagy and fibrosis markers in BEAS-2B cells upon TGF-β1 exposure, while enhancement of extracellular HMGB1 by rHMGB1 pretreatment overturned this effect. **(B)** Relative densitometry of the indicated proteins to GAPDH was displayed as statistical graphs. **(C)** Representative images of mRFP-GFP-LC3 dual-fluorescence assay showed autophagic flux in BEAS-2B cells with different treatments. Scale bar= 20μm. **(D)** Quantification of the counts of autophagosomes (yellow dots) and autolysosomes (red dots) in merged images. Data are expressed as means±SEM of three independent experiments. #p<0.05 vs control group, ##p<0.01 vs control group, *p<0.05 vs TGF-β1 group, **p<0.01 vs TGF-β1 group, Δp<0.01 vs TGF-β1+PGRN group, ΔΔp<0.01 vs TGF-β1+PGRN group.

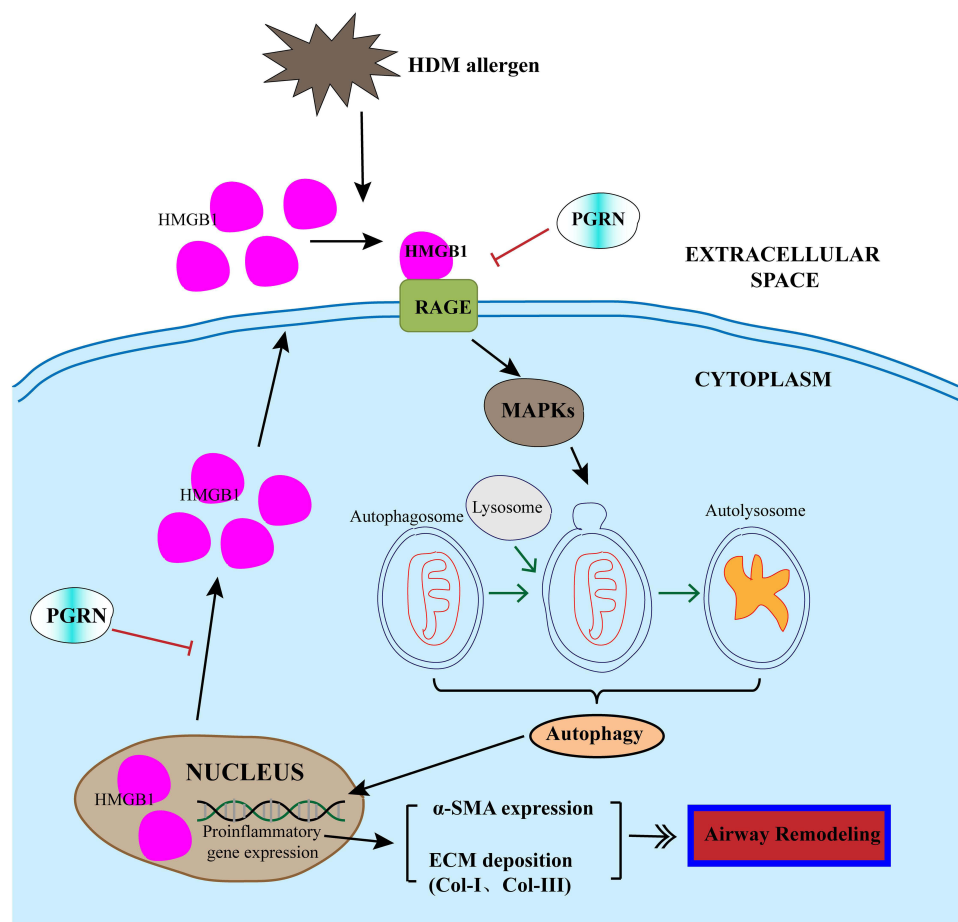


Figure 8 Schematic diagram illustrated the mechanism that HMGB1 is involved in the inhibition of PGRN on autophagic activity in HDM-induced airway remodeling. HDM exposure triggered the activation of HMGB1 and subsequent RAGE/MAPKs signaling pathway in airway epithelial cells, thus potentiating autophagy process and upregulating the expression of α -SMA and collagen proteins, then the formation of airway remodeling. PGRN could overturn HDM-induced HMGB1 overexpression, and further restrained autophagy and alleviated ECM production. Black lines: facilitation; Red lines: inhibition.

B). Altogether, the results show that PGRN prevents fibrotic process in airway epithelia through the reduction of autophagy in a manner dependent on HMGB1 suppression, as illustrated in Figure 8.

Discussion

Airway remodeling, an independent and major pathology that describes the disruption of airway wall tissues, is an important factor for aggravating the progression of asthma. Hitherto, there have not been clinically satisfactory treatments for asthmatic airway remodeling. PGRN is regarded as a therapeutic agent for its anti-inflammatory and anti-fibrotic abilities in various diseases.^{27,28} But little is known about the action of PGRN in airway remodeling under asthmatic conditions. In this study, we provided the evidence that PGRN attenuated collagen synthesis and fibrotic process in HDM-induced chronic asthma model. Moreover, PGRN could downregulate autophagy via

HMGB1 suppression that was considered to mainly contribute to airway remodeling in asthma. Although PGRN has not yet been applied to asthmatic patients, our study may provide a new theoretical and experimental basis for clinical treatment of asthma.

PGRN, also known as granulatin-epithelium precursor (GEP), proelithelin, or prostate cancer cell-derived growth factor (PCDGF), is an autocrine growth factor with multiple biological functions.⁶ Notably, many studies have suggested the defensive properties of PGRN on inflammatory response in asthma.²⁹ Intranasal administration of recombinant PGRN successfully reduced airway hyperresponsiveness by inhibiting the degradation of I κ B- α in ovalbumin-challenged mice.³⁰ Park et al reported that serum PGRN levels were lower in asthmatic subjects compared with those in healthy controls, thus serving as an indicator for the severity of neutrophil inflammation in asthma.³¹ Based on these aforementioned observations, we speculate that decreased endogenous PGRN

may be a potential inducer for asthma pathogenesis. Therefore, in the present study, we applied exogenous PGRN to the airways of HDM-challenged asthmatic mice. Our results revealed that PGRN treatment significantly suppressed airway inflammation, collagen accumulation, smooth muscle hyperplasia/hypertrophy, and submucosal gland hyperproduction in asthmatic mice upon HDM irritations, accompanied by the down-regulation of fibrosis markers α -SMA and collagen-I in mice lungs. These findings suggested the protective ability of PGRN against HDM-induced airway remodeling and fibrosis.

It is known that HDM allergens are a key source of environmental allergens. HDM-derived allergens such as Derp 1 and Derp 2 are considered to be major triggers that activate airway epithelial cells and promote the release of pro-inflammatory cytokines via their protease activities, which initiate the primary phase during asthmatic process.³² For another, damaged epithelial cells exposed to HDM produce a lot of pro-fibrotic mediators such as TGF- β 1 that modulate the formation of ASM and ECM proteins. TGF- β 1 renders positive influence on subepithelial fibrosis, EMT induction, and myofibroblast differentiation, which is strongly involved in the changes of airway structures.³³ Consistently, higher expression of TGF- β 1 and another mesenchymal marker MMP9 were detected in HDM-challenged mice compared with the controls. In the *in vivo* experiments of the current study, we found that TGF- β 1 stimulation drove matrix production and collagen accumulation through upregulating ECM and EMT markers expression in BEAS-2B cells. TGF- β 1-induced EMT in airway epithelial cells is an underlying cause which leads to fibrosis of the airways. Following HDM allergens insult, damaged airway epithelium can be induced into mesenchymal transdifferentiation which promotes disrupted adherens junctions. This inappropriate airway repair further facilitates fibrogenesis involved in airway remodeling.³⁴ Additionally, in this study, more serious tendencies of fibrotic process were displayed in airway epithelia with longer-term TGF- β 1 stimuli, which were yet abrogated by PGRN treatment. Collectively, our results confirm the inhibitory impact of PGRN on TGF- β 1-mediated fibrosis that essentially contributes to abnormal transformations in airway wall composition in HDM-induced chronic asthma.

Autophagy is a “self-eating” regulatory mechanism that disposes metabolic waste and maintains cellular homeostasis in different physiological conditions. Nevertheless, aberrant autophagy can induce type II programmed cell death and may lead to the occurrence of various diseases.³⁵ Emerging evidence verify the role of autophagy in asthma pathogenesis. Poon et al showed that severe asthmatics had more

autophagosomes in epithelial cells of bronchial biopsy tissues than healthy subjects.³⁶ Elevated expression of autophagy-related gene 5 (Agt5) was found to greatly result in airway collagen synthesis in asthma patients.¹⁸ Moreover, autophagy activation is considered as a crucial promoter for extracellular matrix deposition in asthmatic airway remodeling.³⁷ In the present study, increased expression of autophagy markers LC3, Beclin-1 and p62 was discovered in asthmatic mice and TGF- β 1-treated cells, which was also actively correlated with collagen accumulation and fibrotic process in airway epithelia. Intriguingly, PGRN alleviated airway remodeling coupled with the suppression of autophagy both in our *in vivo* and *in vitro* experiments, which was also confirmed by an autophagy inhibitor 3-MA treatment. These findings demonstrate that negative regulation of PGRN on autophagy is a possible mechanism of reducing airway remodeling. Actually, accumulating data provided the evidence supporting the function of PGRN as a novel regulator on the modulation of autophagic activity. PGRN deficiency in aged mice led to aggregation of the autophagy markers p62/ubiquitin in neuronal and glial cells via disturbing autophagy-lysosomal pathway.^{38,39} But a research from Zhou et al reported that PGRN prevented podocyte injuries by promoting autophagic activity in diabetic nephropathy.⁴⁰ Therefore, the discrepancy about the role of PGRN on autophagy might depend on diverse types of tissues and cells under specific pathophysiological conditions.

Mechanically, there are many signaling pathways involved in the regulation of autophagy. As a recognized inflammatory mediator, HMGB1 released by injured cells plays critical roles in immunity, inflammation and fibrosis by eliciting autophagy.^{21,22,25} A study by Hou et al showed that elevated HMGB1 activity led to airway remodeling in OVA-challenged asthma model.⁴¹ Likewise, we herein demonstrated that HMGB1 expression was upregulated following HDM stimuli in mice, whilst silencing HMGB1 inhibited TGF- β 1-mediated collagen synthesis and fibrosis. Also, increased levels of extracellular HMGB1 improved autophagy flux in airway epithelia under fibrotic conditions, which were concomitant with fibrotic process. These results identified the functions of HMGB1 on HDM-induced airway remodeling and pathological fibrosis through upregulating autophagy. Recently, some researchers sought to link the protective role of PGRN to HMGB1 suppression. In LPS-induced acute kidney injury model, PGRN pretreatment effectively reduced the expression and nucleocytoplasmic translocation of HMGB1 in renal tubular epithelial cells of mice.⁴² Similarly, our data in the present study clarified that HDM-mediated HMGB1 overexpression in airway tissues of asthmatic mice was obviously abolished by PGRN

treatment. It was thus likely that HMGB1 was a target in PGRN-exerted antagonistic effect against airway remodeling in HDM-induced asthma. We further demonstrated that the improvement of collagen synthesis and autophagic activity by PGRN was offset when extracellular HMGB1 levels were elevated. These findings indicate that the protection of PGRN against asthmatic airway remodeling via downregulating autophagy is undoubtedly attributed to HMGB1 suppression. In addition, our results revealed the reduction of RAGE expression and the inactivation of MAPKs (JNK, ERK1/2, p38) signaling following HMGB1 suppression in PGRN-treated asthmatic mice. Notably, RAGE is a well-accepted receptor bound by HMGB1 to mediate allergic inflammation in asthma.⁴³ HMGB1-RAGE axis has also been proved to induce autophagy under diverse disorders.^{44,45} So we speculated that PGRN may render protective ability through inhibiting HMGB1-RAGE and downstream MAPKs signaling during HDM-induced asthma, which was partly in accordance with a previous study by Liu et al.⁴⁶ There were some limitations that we only evaluated the changes of RAGE expression in PGRN-treated mice lungs under HDM exposure. The correlation between PGRN and other HMGB1-mediated signaling pathways is warranted to be elucidated in further study.

Conclusions

In conclusion, inhaled HDM allergens trigger aberrant autophagy of airway epithelial cells to accelerate airway remodeling. PGRN effectively ameliorates the HDM-induced airway remodeling in terms of collagen production and fibrosis, which is associated with reduction of autophagy by inhibiting HMGB1. Our data provides evidence for PGRN as a promising therapeutic option of HDM-induced chronic asthma.

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Disclosure

The authors report no conflicts of interest in this work.

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