

Association of microsatellite polymorphisms of the GPDS1 locus with normal tension glaucoma in the Japanese population

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Background: To investigate whether the GPDS1 locus, a potential causative locus of pigment-dispersal syndrome, is associated with normal-tension glaucoma (NTG) in Japanese patients.

Materials and methods: We used polymerase chain reaction amplification with sequence-specific primers to analyze 20 polymorphic microsatellite markers in and around the GPDS1 locus with an automated DNA analyzer and automated fragment detection by fluorescent-based technology. The DNA samples used for these analyses were obtained from ethnicity- and gender-matched patients, including 141 Japanese patients with NTG and 101 healthy controls. Patients exhibiting a comparatively early onset were selected as this suggests that genetic factors may show stronger involvement.

Results: One allele of D7S2462 exhibited a frequency that was significantly decreased in NTG cases compared to controls ($P = 0.0013$, $P_c = 0.019$, $OR = 0.48$, 95% CI = 0.30–0.75). Alleles at another six microsatellite loci were positively or negatively associated with NTG, but these associations did not retain statistical significance after Bonferroni correction ($P < 0.05$, $P_c > 0.05$).

Conclusion: Our study showed a significant association between the GPDS1 locus and NTG, suggesting that there may be some genetic risk factor(s) in the development of NTG.

Keywords: microsatellite, normal tension glaucoma, glaucoma-related pigment dispersion syndrome, GPDS1, DPP6

Background

Glaucoma is the second most common cause of blindness worldwide.¹ This condition has a high socioeconomic impact, affecting over 67 million people in the western world.² The disease is characterized by a loss of visual field corresponding to the excavation of the optic disc. It is considered to be a multifactorial disorder that occurs because of the interaction of genetic and environmental factors. In addition to an association with intraocular pressure (IOP), the onset and progression of glaucoma is involved with several other risk factors, including ischemia, refraction, systemic illness, and genetic predisposition.

Primary open-angle glaucoma (POAG) is usually associated with elevated IOP. The pathogenesis of glaucomatous optic neuropathy is not completely understood, although elevated IOP is a generally accepted major risk factor for this condition. Normal-tension glaucoma (NTG) is an important subtype of POAG and accounts for approximately one-third of all POAG cases in Caucasians.³ Notably, the incidence of NTG is higher in Japan than in the western world, and it occurs in approximately 92% of all Japanese POAG patients, as reported in the Tajimi Study.⁴ The IOP of NTG patients consistently lies within the statistically normal range; thus, although IOP is suggested to play a role in the development of NTG, its involvement may be less in NTG than that in high-tension glaucoma (HTG). Other pathogenic factors such as vasospasm,

ischemia, and vascular insufficiencies are indicated to be associated with the development of NTG.^{5–8}

Pigment dispersion syndrome (PDS) and pigmentary glaucoma (PG) are characterized by a disruption of iris pigment epithelium and deposition of the dispersed pigment granules throughout the anterior segment. The classic diagnostic triad consists of corneal pigmentation (Krukenberg spindle); slit-like, radial, or mid-peripheral iris transillumination defects; and dense trabecular pigmentation.⁹ PDS/PG usually affects individuals under the age of 30 years. In the early stages of the disease, affected individuals may exhibit clinical evidence of dispersed pigment without an associated elevation of IOP or optic nerve degeneration. However, as the disease progresses, some patients develop increased IOP and degeneration of the optic nerve, causing permanent vision loss. Recent linkage studies have identified several genes or chromosomal loci in some types of glaucoma.^{10–12} In PDS-affected families, significant linkage has been observed between the disease phenotype and markers located on several chromosomal loci. GPDS1 (OMIM ID 600510) was the first genetic locus on chromosome 7q35–q36 found to be associated with PDS.¹³

Although the GPDS1 locus may include a gene associated with NTG, an association between the GPDS1 locus and NTG had not been previously investigated. The purpose of this study is to determine whether the GPDS1 locus is one of the candidate loci for developing NTG. We explored this potential in an association study using 20 microsatellite markers around the GPDS1 locus.

Materials and methods

Participants

We recruited 242 Japanese participants from the Yokohama City University, Yamanashi University, Gifu University, Kobe University, Yamaguchi University, Kumamoto University, Hokkaido University, Tokyo University, Niigata University, Kanazawa University, Hiroshima University, Tajimi Municipal Hospital, and Tokai University in Japan. Of these participants, 141 were diagnosed with NTG, and 101 served as controls. The criteria applied for the diagnosis of NTG were those proposed in our previous study.¹⁴ The mean patient age was 49.2 ± 9.6 years, and the male:female ratio was 0.87. The mean refraction value was -3.75 ± 3.02 diopters (D), and the mean deviation observed in the Humphrey® static visual field determination (Carl Zeiss Meditec, Oberkochen, Germany) was -10.13 ± 8.04 dB. The control participants were of the same age and sex as the NTG patients, and they were not affected by glaucoma or any local or systemic illnesses that

could result in optic disc or visual field changes. Furthermore, the control cases either had no myopia or had mild myopia with refractive errors of -3.00 D or less. All patients and control participants were of Japanese ethnicity, with similar social backgrounds and residing in the same urban area. Informed consent was obtained from all participants, and the study was conducted in accordance with the Declaration of Helsinki and its subsequent revisions.

Analysis of repeat polymorphisms for 12 microsatellite loci

Genomic DNA was extracted using the QIAamp DNA Blood Mini Kit (Qiagen, Hilden, Germany) or the guanidine method. In this association study, we selected 20 microsatellite loci that are located around the GPDS1 region on chromosome 7q35–36.3 and that have been established as informative polymorphic genetic markers (Figure 1). The markers were determined based on the National Center for Biotechnology Information guidelines for fine mapping. A polymerase chain reaction (PCR) was performed in a reaction mixture with a total volume of 12.5 μ l, containing PCR buffer, genomic DNA, 0.2 mM dinucleotide triphosphates (dNTPs), 0.5 μ M primers, and 0.35 U Taq polymerase. The PCR conditions were as follows: 94 °C for 5 min, followed by 30 cycles of denaturation at 94 °C for 30 sec, annealing at 56 °C for 30 sec, extension at 72 °C for 1 min, and a final elongation step at 72 °C for 10 min. The reaction was carried out in a PCR thermal cycler (GeneAmp System 9700, Applied Biosystems, Foster City, CA, USA). The forward primer was labeled with 6-FAM (Sigma-Aldrich, St. Louis, USA) at the 5' end (Table 1). To determine the number of microsatellite repeats, the PCR products were denatured at 97 °C for 2 min, mixed with formamide, and electrophoresed in an ABI3130 Genetic Analyzer (Applied Biosystems). The number of microsatellite repeats was estimated automatically using GeneScan 672 software (Applied Biosystems) and the local Southern method with a size marker of GS500 TAMRA (Applied Biosystems).

Statistical analysis

Allele frequencies were estimated by direct counting. The significance of allele frequencies between the patient and control groups was evaluated by the Fisher's exact test. The probability of association was corrected by the Bonferroni inequality method, ie, by multiplying the obtained P values with the number of alleles compared. A P value of <0.05 was considered statistically significant. Statistical analyses were performed on a computer using SPSS software (version 10.1; SPSS Inc., Chicago, IL, USA).

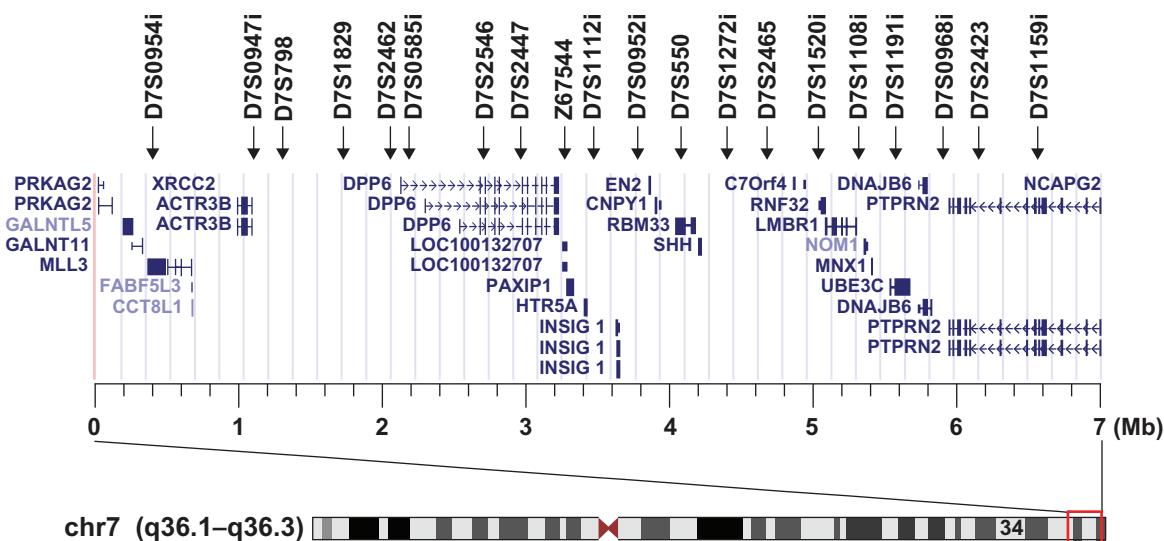


Figure 1 Location of 20 microsatellite markers used in this study and the genes around the GPDS1 locus. Display of genes and chromosome seven results in the UCSC genome browser on Human Mar. 2006 Assembly at chr7: 151,120,000–158,120,000.

Results

We analyzed 20 microsatellite markers localized around GPDS1 for all patients and control participants (Figure 1). All allele frequencies were found to be in Hardy–Weinberg equilibrium proportion. Table 2 lists the allele frequencies of microsatellite markers in patients and control participants, with each allele designated by the size of its amplification. The 132 allele of D7S2462 was significantly associated with NTG, and the allele frequency was decreased in NTG cases as compared to controls (14.5% vs 26.2%, $P = 0.0013$, $P_c = 0.019$, OR = 0.48, 95% CI = 0.30–0.75). Another two markers, D7S2465 and D7S2423, were also negatively associated with the risk of NTG. On the other hand, four markers – D7S2546, Z67544, D7S1272i, and D7S1108i – were positively associated with the risk of NTG. However, these two negative associations and four positive associations did not retain statistical significance after Bonferroni correction ($P < 0.05$, $P_c > 0.05$).

Discussion

Criteria for PDS were two of the following: midperipheral, radial iris transillumination defects; Krukenberg spindle; heavy trabecular meshwork pigmentation; and wide-open angle.⁹ Criteria for PG were PDS and two or more of three findings: IOP greater than 21 mmHg, optic nerve damage, or visual field loss. The probability of converting to pigmentary glaucoma was 10% at 5 years and 15% at 15 years. Young, myopic men were most likely to have pigmentary glaucoma. The most significant risk factor for conversion to pigmentary

glaucoma was an IOP greater than 21 mmHg at initial examination, whereas age, refractive error, and family history of glaucoma were not correlated with conversion.¹⁵

NTG is a critical and major subtype of POAG and shares many of the same pathologic characteristics with HTG except for elevated IOP. Notably, many other diseases cause atrophy of the optic nerve similar to that caused by NTG. Since the coexistence of these different diseases decreases statistical validation in the case-control analysis, we carefully selected the NTG patients using highly stringent criteria to select and distinguish the NTG phenotype from the other diseases that cause optic nerve atrophy.

The GPDS1 locus is responsible for PDS.¹³ An association between the locus and NTG, however, had not been previously investigated. Therefore, in this study, we sequenced the repeat polymorphisms of 20 microsatellite loci within and around the GPDS1 locus to determine whether these repeat polymorphisms are associated with NTG in a Japanese population.

The 20 microsatellite markers selected for this study were located almost 100–700 kb apart from each other on chromosome 7q35–36.3; they were chosen to achieve dense coverage of the regions located around GPDS1 and to detect susceptible loci related to NTG. As a result, a relatively increased allele frequency was observed in the patient group for the four microsatellite markers D7S2546, Z67544, D7S1272i, and D7S1108i. Another three markers – D7S2462, D7S2465, and D7S2423 – were associated with lower risk of NTG. Of these markers, only D7S2462 remained

Table I Primers used in polymerase chain reaction for amplification of microsatellite markers in this study

| Location | Orientation ^a | Sequence |
|----------|--------------------------|-------------------------------|
| D7S0954i | F (NED) | 5' TTACATAAGGTACTTCTGGTCAT3' |
| | R | 5' TGGTCTCCAGCTACATCAG3' |
| D7S0947i | F (NED) | 5' CTACTTCCGGTGTCTAGCTT3' |
| | R | 5' GATAACAGCTGATAGGTAGATCAG3' |
| D7S798 | F (VIC) | 5' AGCTGCAAAATAGTGGAAAGTAG3' |
| | R | 5' CATCAATTACACATAATGACCG3' |
| D7S1829 | F (VIC) | 5' GGGGGTTTCAGAAGAGATGT3' |
| | R | 5' CCTCCAAAATTCTGAGCAA3' |
| D7S2462 | F (FAM) | 5' GCCAAGATTGCACCAACTAC3' |
| | R | 5' TGTTAAAAGTCCCCAAGC3' |
| D7S0585i | F (NED) | 5' AATGAGCATTCTCAGTTGAATAC3' |
| | R | 5' ATCTGGTGAACGAGTAATAACAC3' |
| D7S2546 | F (FAM) | 5' GGAGGTTGAACAACCTCTGAATAC3' |
| | R | 5' CACGCCAGGGTCTATCTT3' |
| D7S2447 | F (NED) | 5' GTCAAGATGTAGGAACCAAGC3' |
| | R | 5' TCTCACTGTACTCAGCCTGT3' |
| Z67544 | F (VIC) | 5' CAACCTAAAATACGCTGTTACC3' |
| | R | 5' GCTTTTCAGACCAAATAACTTAC3' |
| D7S1112i | F (NED) | 5' GAGAATCTAATGAAAGCTGTGG3' |
| | R | 5' TTCAGGGCATCCAAGAAG3' |
| D7S0952i | F (NED) | 5' AGATATTTGGCTAACATGGTC3' |
| | R | 5' TCTTCAGAGATACAGAAGCAATAG3' |
| D7S550 | F (FAM) | 5' TCTCATCTGTGAATGCACTATC3' |
| | R | 5' GCAGTTGGTTATTCAAGTC3' |
| D7S1272i | F (PET) | 5' AATGTCCTGGAGGCTGAG3' |
| | R | 5' AAAGACTTGGTGAGACCTTCTC3' |
| D7S2465 | F (PET) | 5' ACCTGGCAACAGAGTGAG3' |
| | R | 5' CTTCAAAGAGTTATGCTTATGTGG3' |
| D7S1520i | F (PET) | 5' GCTGTTAGCATCTGGTAATTAC3' |
| | R | 5' AAACTGAAGTCTGCCATCTATTAG3' |
| D7S1108i | F (FAM) | 5' ACTTACTTATCCTACTGATCCGTG3' |
| | R | 5' TGAACCATAAATTACCTCATTAC3' |
| D7S1191i | F (VIC) | 5' CACTCTATTGCATGGTGAAC3' |
| | R | 5' TCCAGTAGAGGGAGCTCAG3' |
| D7S0968i | F (NED) | 5' CACATCCTTATACCTACATTCAAG3' |
| | R | 5' TAATTTATCTAGAAGGTAGCAA3' |
| D7S2423 | F (NED) | 5' CTTCAGACCTTCAGTTGATGAT3' |
| | R | 5' GCTCTCAGACACATTTCCA3' |
| D7S1159i | F (VIC) | 5' CTTGGAGTCAAAGAGGCC3' |
| | R | 5' CTTCCCTGACTCCTGATTAGAG3' |

Note: ^aDye label.

significantly associated with NTG after Bonferroni correction ($P_c < 0.05$). Therefore, there is one statistically appreciable difference between the NTG group and the control group in these 20 microsatellite markers located over a 6-Mb genomic region around GPDS1.

The dipeptidyl-peptidase 6 (DPP6) gene is the nearest gene to the D7S2462 marker. DPP6 is a type II membrane protein that binds voltage-gated neuronal potassium channels (A-type) and alters their expression and biophysical properties.¹⁶ Recently, genetic variants in DPP6 have been

Table 2 Frequencies of 20 microsatellite markers in normal-tension glaucoma (NTG) patients and healthy participants

| Marker | Occurrence of allele | Allele ^a | Frequencies (%) | | P | Pc | Odds ratio (95% CI) |
|----------|-------------------------|---------------------|----------------------------|---------------------------------------|--------|-------|------------------------|
| | | | NTG patients (2n = 282) | Healthy participants (2n = 202) | | | |
| D7S0954i | 9 | 310 | 3.5 | 2.0 | 0.31 | | |
| D7S0947i | 7 | 151 | 3.2 | 2.5 | 0.64 | | |
| D7S798 | 8 | 211 | 7.8 | 4.0 | 0.08 | | |
| D7S1829 | 13 | 391 | 6.0 | 4.0 | 0.31 | | |
| D7S2462 | 15 | 132 | 14.5 | 26.2 | 0.0013 | 0.019 | 0.48 |
| D7S0585i | 9 | 324 | 4.3 | 3.0 | 0.46 | | |
| D7S2546 | 8 | 239 | 13.8 | 6.4 | 0.0096 | 0.077 | 2.33 |
| D7S2447 | 4 | 175 | 53.5 | 52.5 | 0.82 | | |
| Z67544 | 6 | 151 | 6.4 | 1.5 | 0.0091 | 0.055 | 3.99 |
| D7S1112i | 6 | 121 | 44.7 | 42.1 | 0.57 | | |
| D7S0952i | 3 | 450 | 1.2 | 0.5 | 0.21 | | |
| D7S550 | 13 | 192 | 1.2 | 0.5 | 0.21 | | |
| D7S1272i | 6 | 156 | 20.2 | 13.4 | 0.0499 | 0.30 | 1.64 |
| D7S2465 | 14 | 175 | 22.7 | 32.2 | 0.020 | 0.28 | 0.62 |
| D7S1520i | 8 | 300 | 3.2 | 3.0 | 0.58 | | |
| D7S1108i | 14 | 188 | 3.2 | 0.5 | 0.040 | 0.56 | 6.63 |
| D7S1191i | 6 | 181 | 0.7 | 0.5 | 0.77 | | |
| D7S0968i | 6 | 357 | 50.0 | 44.6 | 0.24 | | |
| D7S2423 | 5 | 250 | 19.9 | 29.2 | 0.017 | 0.086 | 0.60 |
| D7S1159i | 8 | 115 | 13.8 | 9.4 | 0.14 | | |

Note: ^aAlleles found to exhibit the most difference in frequencies between patients and healthy participants for each marker are shown. Each allele was designated by the size of its amplification.

reported to show a significant association with human neural diseases such as amyotrophic lateral sclerosis and autism.^{17–21} Since potassium channels are considered to play an important role in protecting against ischemia and the development of apoptosis,²² DPP6 variants may act as risk factors in the development of NTG.

In conclusion, we performed an association analysis of the GPDS1 locus using microsatellite markers in NTG patients and found that there was an NTG-associated region in the GPDS1 locus. Therefore, further association studies using single nucleotide polymorphisms in the resistant region might help to identify the pathogenic gene(s) related to NTG.

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