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REVIEW

Endocannabinoid system as a regulator of tumor cell malignancy – biological pathways and clinical significance

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Abstract: The endocannabinoid system (ECS) comprises cannabinoid receptors (CBs), endogenous cannabinoids, and enzymes responsible for their synthesis, transport, and degradation of (endo)cannabinoids. To date, two CBs, CB1 and CB2, have been characterized; however, orphan G-protein-coupled receptor GPR55 has been suggested to be the third putative CB. Several different types of cancer present abnormal expression of CBs, as well as other components of ECS, and this has been shown to correlate with the clinical outcome. Although most effects of (endo)cannabinoids are mediated through stimulation of classical CBs, they also interact with several molecules, either prosurvival or proapoptotic molecules. It should be noted that the mode of action of exogenous cannabinoids differs significantly from that of endocannabinoid and results from the studies on their activity both in vivo and in vitro could not be easily compared. This review highlights the main signaling pathways involved in the antitumor activity of cannabinoids and the influence of their activation on cancer cell biology. We also discuss changes in the expression pattern of the ECS in various cancer types that have an impact on disease progression and patient survival. A growing amount of experimental data imply possible exploitation of cannabinoids in cancer therapy.

Keywords: cannabinoids, cancer, cell signaling, cannabinoid receptor, delta-9-tetrahydrocannabinol

Introduction

Cannabinoids are active components of Cannabis sativa, which has been used for medical and recreational purposes for many years. Their activity depends mainly on activation of members of GPR family, the so-called cannabinoid receptors (CBs).¹ Two subtypes of this molecule have been isolated so far - CB1 and CB2 - although a third putative CB, CB3, has been suggested.²⁻⁴ The main difference between them seems to be their tissue expression pattern. CB1 receptors are found mainly in the central nervous system, particularly in those regions of the brain that are involved in the control of memory (hippocampus), emotions (amygdala), motor activity (basal ganglia and cerebellum), cognition (cortex), and appetite (hypothalamus).⁵ CB1 receptors are also present in peripheral tissues, such as uterus, testes, ovaries, and prostate.^{1,6} In contrast, CB2 receptors predominate in the immune system, both in cells such as lymphocytes and macrophages and in tissues such as spleen, tonsils, and lymph nodes.^{1,6} Recently, it has been reported that some effects of cannabinoids are non-CB1/CB2 mediated.^{7,8} Orphan GPR GPR55 has been suggested to be the third CB according to some reports.^{9,10} The expression of genes encoding a CB3 receptor has been shown in several peripheral tissues, such as spleen, tonsils, and mammae, and in endothelial

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cells.^{8,11} It has been discovered only recently that cannabinoids exert an array of anticancer effects via interfering with signaling pathways involved in neoplastic transformation and tumor progression. Along with cannabinoids produced endogenously, termed endocannabinoids, and enzymes responsible for synthesis, transport, and degradation, CBs comprise the endocannabinoid system (ECS).

Among the range of endocannabinoids, two major ones are 2-arachidonoylglycerol (2-AG) and anandamide (N-arachidonoylethanolamide [AEA]), both lipid-signaling molecules, containing long-chain polyunsaturated fatty acids, amides, and esters.^{1,12,13} Other minor compound that belongs to the "endocannabinoid family" is arachidonyl glyceryl ether named noladin.^{12,13} AEA originates from the phospholipid precursor, phosphatidylethanolamine, which is transformed into N-acylphosphatidylethanolamine (NAPE) by a calcium-dependent NAT. NAPE is subsequently converted to anandamide via four alternative enzymatic pathways; the best studied and the most direct is through catalysis by an NAPE-PLD.^{14,15} Similarly, 2-AG is synthesized in a twostep process. First, a phospholipase C enzyme catalyses the hydrolysis of inositol phospholipids to diacylglycerols, which are next transformed into 2-AG by a DAGL.14,15

Endocannabinoids are not stored in cellular vesicles but are rapidly synthesized following the increase in the intracellular calcium level.^{1,13,16} Released from depolarized postsynaptic neurons, endocannabinoids act as retrograde messengers, binding and activating CB1 receptors at presynaptic terminals.^{15–17} Their action results in the inhibition of neurotransmitter release, mostly glutamate and γ-aminobutyric acid.¹⁶ After activation of CBs, AEA and 2-AG undergo cellular reuptake and are degraded to arachidonic acid and ethanolamine or glycerol, respectively.¹⁶ AEA is metabolized by the enzyme FAAH, whereas 2-AG is hydrolyzed predominantly by MAGL and to a lesser extent by ABHD-6 and ABHD-12.^{18–20} Endocannabinoids might also be degraded through oxidation by COX, LOX, or cytochrome P450.¹⁵

Numerous studies have been performed to explain the biological role of the ECS in health and disease and its potential exploitation in therapy.¹ Owing to diverse pharmacological properties, cannabinoids have attracted significant attention in recent years. In particular, they offer potential applications as antitumor drugs, based on the ability to inhibit cell proliferation and to induce tumor cell death by some members of the cannabinoid family.⁶ It is important to point out that the action of endocannabinoids does not match that of exogenous cannabinoids and results from research Synthetic cannabinoids as well as plant-derived and endogenous analogs are not only oriented at activation of classical CBs but also interact with several molecules, either prosurvival or proapoptotic molecules. It has been suggested that stimulation of calcium-permeable ion channel TRPV1 or PPAR γ and inhibition of COX-2 may mediate the proapoptotic and antiproliferative effects of anandamide and synthetic cannabinoids in cancer.^{21–23}

A variety of mechanisms are likely to contribute to the anticancer effect of CB ligands. CB1 and/or CB2 receptors are coupled to several signaling pathways directly involved in cell survival, proliferation, and apoptosis, including p38 MAPK, cyclic AMP, PI3K-Akt, RhoA, JNK, EGFR, ERK,²⁴ and ceramide pathways.^{1,19,25-30}

Ceramide seems to be the key mediator of cannabinoidmediated anticancer effects. Cannabinoids induce sustained production of sphingolipid ceramide, which is commonly found in the cell membrane and is generated de novo by ceramide synthase or through sphingomyelin hydrolysis.^{29,31} Ceramide generation has been implicated in the action of cannabinoids in MG63 osteosarcoma cells, C6.9 glioma cells, and primary astrocytes through stimulation of ERK, MAPK, and/or JNK pathways.24,32,33 This effect can be mediated via CB1 and/or CB2 receptors.^{24,31,34} In addition, there are several reports describing the influence of cannabinoids on ERK cascade in cancer. Although ERK is considered as a rather proproliferative signaling molecule,²⁹ it has been recently shown that sustained activation of ERK induces apoptosis in astrocytes.35 Regarding cannabinoid influence on the ERK activation, conflicting data have been reported, demonstrating either upregulation or reduction of this signaling pathway in response to CB1/CB2 agonist.36,37

As shown earlier, mechanisms implicated in the anticancer effect of cannabinoids are quite complex; thus, in order to broaden our knowledge, further investigation of this subject is warranted.

Role of cannabinoids in cancer – cultures and in vivo studies

Currently, cannabinoids are used in the palliative therapy of cancer patients. In particular, Δ^9 -tetrahydrocannabinol (THC; Marinol), its analog nabilone (Cesamet), and Sativex (Δ^9 -THC and cannabidiol [CBD]) were brought onto the market in several countries owing to their ability to inhibit chemo- and radiotherapy-induced side effects.³⁸ Palliative properties of the aforementioned drugs include inhibition of vomiting and nausea, pain relief, stimulation of appetite, and reduction in weight loss, commonly seen in patients undergoing conventional cancer treatment.^{6,38,39}

However, cannabinoids have been proved to affect proliferation, viability, and invasiveness of cancer cells in cultures and in vivo.^{1,6,38-40} A considerable body of evidence has accumulated since 1975 when Munson et al⁴¹ demonstrated for the first time the antitumorigenic action of C. sativa-derived cannabinoids in in vitro and in vivo experiments. For example, after 20 days of consecutive oral administration of Δ^8 -THC and cannabinol, reduction in tumor growth was seen in the Lewis lung adenocarcinoma mouse model. Antitumoral activities of CB ligands include several distinct mechanisms, such as inhibition of proliferation, induction of autophagy and apoptosis, cell-cycle arrest, reduction in metastasis outgrowth through diminishing secretion of proangiogenic molecules, and stimulation of inflammatory response against cancer.42-53 Figure 1 summarizes the influence of synthetic, plant-derived, and endogenous cannabinoids on signaling molecules, leading to tumor growth inhibition and/or cancer cell death.

Influence on apoptosis and proliferation of cancer cells

It has recently been shown that WIN 55,212-2, a synthetic nonselective CB1/CB2 agonist, induces dose- and time-dependent apoptotic effects on HT29, HCT116, and Caco-2 colon cancer cells as demonstrated by loss of mitochondrial potential and activation of caspase-3 and -9.50 These effects were accompanied by elevated endoplasmic reticulum (ER) stress and a subsequent increase in the level of specific markers, such as GRP78, CHOP, and TRB3. It is noteworthy that induction of GRP78 and CHOP was required for apoptosis of cancer cells and overexpression of CHOP, which were correlated with better prognosis in cancer patients.54-56 Also, upregulation of TRB3 expression was shown to be critical for inhibition of cancer cell proliferation.57 CHOP and TRB3 are genes that have been demonstrated to be critically involved in cellular response to stress stimuli after THC treatment as shown by Carracedo et al.58 They have reported THC-induced proapoptotic activity in several cancer cell lines due to generation of ceramide and upregulation of downstream target - stressregulated protein p8 – which subsequently mediated the increase in the expression of CHOP and TRB3.58

Contrarily, in osteosarcoma, despite the fact that WIN treatment induced ER stress markers, such as GRP78, CHOP, and TRB3, and cellular vacuolization, it did not contribute to apoptosis initiation when used alone.⁵⁹ However, incubation

of WIN-treated cells with TRAIL led to apoptotic death (~80% of cell viability decreased after 36 hours) as shown in morphological and biochemical hallmarks of apoptosis and concomitant translocation of GRP78 to cell surface and upregulation of tumor suppressor factor PAR-4.⁵⁹ In human gastric cancer cells, WIN was able to arrest the cell cycle in the G0/G1 phase, thus reducing proliferation and inducing apoptosis, and both these effects were mediated via down-regulation of pAkt.^{49,53}

Interestingly, WIN-induced killing of colon cancer cells depends upon the downregulation of PPARy, at both mRNA and protein levels. Indeed, pharmacological or genetic inhibition of PPARy exerted similar effect to WIN, inducing cell death and moreover in combination with WIN treatment, even potentiated this effect.⁵⁰ This observation is somewhat unexpected, since the majority of reports show that cannabinoid-induced apoptosis relies on upregulation of PPARy expression and this effect could be attenuated by either genetic ablation or pharmacological inhibition of the PPARy pathway.^{23,60} On the other hand, AEA-induced apoptosis in tumorigenic JWF2 keratinocytes depends upon activation of COX-2, which then induces ER stress.⁵² Furthermore, it was determined that COX-2 metabolizes AEA to J-series prostaglandins and this mechanism is required for ER stress-associated apoptotic process activated by AEA.⁵² Cannabinoid-induced apoptosis may be, actually, paralleled by both PPARy and COX-2 upregulation.61

In LNCaP prostate carcinoma cells, CBD significantly reduced viability and induced apoptosis, which was partly due to antagonism of TRPM8 and was accompanied by diminished expression of androgen receptor, p53 activation, and elevated levels of reactive oxygen species (ROS).⁶² The proapoptotic effect of CBD was confirmed by upregulation of PUMA and CHOP, markers of apoptotic intrinsic pathways, and elevated intracellular Ca²⁺ levels.⁶² In colon cancer cell lines, inhibition of proliferation induced by CBD has been demonstrated, and this effect was CB1, TRPV1, and/or PPARγdependent.⁶² Likewise, standardized *C. sativa* extracts (CBD botanical drug substance [BDS]) with high content of CBD were shown to be effective in decreasing proliferation of cancer cells as shown by Romano et al.⁶³

Anticancer activity of some synthetic compounds has also been indicated in research. In a study by Ortega et al, CP55-940, a CB agonist, was the most effective agent to induce antitumor action in C6 (rat) and U373 (human) glioma tumor lines over synthetic cannabinoid WIN 55,212-2 and the endogenous agonist AEA.⁶⁴ Moreover, this study proposes apoptosis as a major mechanism contributing

Apoptosis			
Mediator	Mediator Effect		
WIN-55,212-2	↓ ∆Ψm, ↑ caspase-3, ↑ GRP78, ↑ CHOP, ↑ TRB3	50	
AEA	↑ ER stress	52	
WIN-55,212-2	↓ pAkt	53	
THC	↑ ceramide, ↑ p8, ↑ ER stress, ↑ CHOP, ↑ TRB3	58	
WIN-55,212-2 + TRAIL	↑ PAR-4	59	
CBD	 ↓ PUMA, ↑ CHOP, ↓ TRPM8, ↑ intracellular Ca²⁺, activation of p53, ↑ ROS, ↓ AR 	62	
O-1602	↑ BAX, ↑ p53	65	
Met-F-AEA + URB597	↓ pro-caspase 9, ↓ pro-PARP	75	

Proliferation			
Mediator	Effect	Reference	
WIN-55,212-2	↓ pAkt	53	
WIN-55,212-2	$\begin{array}{l} \downarrow pAkt, \uparrow p\text{-}ErRK1/2, \downarrow pAkt, \\ \uparrow Kip1/p27, \uparrow Cip1/WAF1/p21, \\ \downarrow cyclin D1, \downarrow cyclin E, \downarrow Cdk 2, \\ \downarrow Cdk 4, \downarrow Cdk 6, \downarrow phospho-Rb, \\ \downarrow E2F-1 \end{array}$	49	
CBD	↓ NF-κB translocation, ↓ pEGFR, ↓ p-ERK, ↓ pAkt	47	
WIN-55,212-2	↑ GRP78, ↑ CHOP, ↑ TRB3	59	
O-1602	↓ PCNA, $↓$ STAT3, ↓ p-NFκB p65, $↓$ TNFα	65	
Met-F-AEA + URB597	\downarrow p-EGFR, \downarrow p-ERK, \downarrow p-Akt \downarrow cyclin D1, \downarrow CDK1	75	
Met-F-AEA	↓ p-ERK	93	

Neovascularization and angiogenesis		
Mediator	Effect	Reference
WIN-55,212-2, JWH-133	↓ VEGF, ↓ PIGF, ↓ Ang2, ↓ EGF-R	44
WIN-55,212-2, JWH-133	↓ VEGF, ↓ VEGFR2	43
CBD	$\begin{array}{c} \downarrow MMP-2, \downarrow MMP-9, \\ \downarrow TIMP1, \\ \downarrow SERPINE1/PAI1, \\ \downarrow uPA, \downarrow CXCL16, \\ \downarrow IL-8, \downarrow ET-1, \\ \downarrow PDGF-AA \end{array}$	94
Met-F-AEA	↓ MMP-2	93

Metastasis formation			
Mediator	Effect	Reference	
WIN-55,212-2	↓ MMP-2, ↓ VEGF-A	53	
CBD	↓ MMP-2, ↓ MMP-9	47	
Met-F-AEA + URB597	↓ MMP-2	75	
CBD, O-1663	↓ ld1	84	
THC	↓ MMP-2	34	
AM404	$\begin{array}{l} \downarrow \text{ NFAT, } \downarrow \text{ NF-}\kappa\text{B, } \downarrow \text{ MMP-1,} \\ \downarrow \text{ MMP-3, } \downarrow \text{ MMP-7} \end{array}$	92	
CBD, ∆9-THC, MA, JWH-133	↑ ICAM-1, ↑ TIMP-1	96	
∆9-THC, MA	↑ TIMP-1	98	
CBD	↑ ICAM-1, ↑ TIMP-1	103	

Autophagy			
Mediator	Effect	Reference	
ACPA + GEM, GW405833+ GEM, SR141716+ GEM	↑ ROS, ↑ XBP-1, ↑ Grp78, ↑ CHOP	46	
ACPA, GW405833	$ \begin{tabular}{l} $$ \uparrow ROS, $$ \uparrow AMP/ATP ratio, $$ \uparrow AMPK, $$ $$ $$ $$ $$ $$ $$ $$ $$ $$ $$ $$ $$$	45	
∆9-THC	Ceramide accumulation, ↑ eIF2α, ↓ ER stress, ↑ p8/TRB3, ↓ Akt/mTORC1	51	
WIN-55,212-2	↑ LC3-II, ↓ p-Akt	50	
∆9-THC, JWH-015	↑ p8/TRB3, ↑ LC3-II, ↓ Akt/mTORC1, ↓ p-S6, ↑ AMPK	80	
Q-1663	↑ I C3-II	84	

Figure 1 Influence of synthetic, plant-derived, and endogenous cannabinoids on signaling molecules leading to tumor growth inhibition and/or cancer cell death. Notes: \uparrow , increased; \downarrow , decreased.

Abbreviations: ER, endoplasmic reticulum; ROS, reactive oxygen species; AMPK, 5'AMP-activated protein kinase; Ang, Angiopoietin; AEA, N-arachidonoylethanolamide; CBD, Cannabidiol; CXCL, Chemokine (c-x-c motif) ligand; EGFR, Epidermal growth factor receptor; ERK, Extracellular signal-regulated kinase; GAPDH, Glyceraldehyde 3-phosphate dehydrogenase GEM, Gemcitabine; GRP, Glucose-regulated protein; Met-F-AEA, 2-methyl-20-F-anandamide; MMP, Matrix metalloproteinases; NF-κB, nuclear factor kappa-light-chain-enhancer of activated B cells; PCNA, Proliferating cell nuclear antigen; STAT, Signal transducer and activator of transcription; TIMP, Tissue inhibitor matrix metalloproteinases; THC, Tetrahydrocannabinol; TRB, Tribblespseudokinase; TRPM, Transient receptor potential cation channel subfamily M member; uPA, Urokinase-type plasminogen activator; VEGF, Vascular endothelial growth factor; VEGFR, Vascular endothelial growth factor receptor.

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to cannabinoid-induced death of cancer cells, at least for CP55-940.⁶⁴ The atypical cannabinoid O-1602 inhibited cancer cell proliferation and induced apoptosis in two colon cancer cell lines, HT-29 and SW480,⁶⁵ and reduced viability of melanoma cells via GPR55 activation.⁶⁶ In addition, antineoplastic effect was accompanied by decreased level of proliferation marker PCNA and depression of oncogenic transcription factors NF- κ B and STAT3 and protumorigenic cytokine TNF, while proapoptotic molecules BAX and p53 increased.⁶⁵ This is noteworthy, since TNF has

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been previously shown to promote cancer cell motility and invasiveness, which is attributed to activation of NF- κ B signaling.⁶⁷ In addition, TNF may contribute to mutations in tumor supressor genes, such as p53.⁶⁵

In human bladder carcinoma (ECV304), dramatic, time-related decline in proliferation of cells was obtained in response to an inverse agonist of CB1 receptor, AM281. As the authors suggested, delay in progression of each cell-cycle phase, rather than growth inhibition, underlies the antiproliferative effect of AM281.⁴⁸ It has been shown that radiation combined with WIN treatment induced a growth-inhibitory effect, without causing death of breast cancer cells.⁶⁸ Furthermore, antiproliferative effects of combination of WIN and radiation therapy were more effective than use of either agent separately.⁶⁸

The EGFR is a cell surface receptor tyrosine kinase, in which activation increases tumor growth, cellular differentiation, and migration and inhibits apoptosis. 69-71 Overexpression of EGFR is found in diverse cancer types and is often associated with poor prognosis.69,72 Some authors suggest that EGFR expression may serve as a negative prognostic factor.^{72,73} Several lines of evidence have shown antiproliferative activity of (endo)cannabinoids due to downregulation of EGFR signaling route, although, in contrast, Fiori et al74 reported an enhanced expression of EGFR in AM251-treated cells (CB1 inverse agonist). Synthetic and natural cannabinoids (2-methyl-20-F-anandamide [Met-F-AEA] and CBD) inhibit EGF-induced proliferation and chemotaxis of different types of cancer cells, which is related to decreased expression of EGFR and its downstream targets Akt, ERK, and NF-KB signaling.47,75 In non-small cell lung cancer and human cutaneous melanoma, inhibition of AEA-degrading enzyme (FAAH) with URB597 potentiated antitumorigenic effect of AEA and its analog Met-F-AEA.^{66,75} Also, inhibition of 2-AG hydrolysis with the use of MAGL inhibitor, JZL184, produced an antiproliferative effect in EGF-treated prostate cancer cells.⁷⁶

Antiproliferative and proapoptotic activities of several cannabinoids have also been proved in preclinical studies. Aviello et al⁴² demonstrated chemopreventive effect of nonpsychotropic CBD in colon carcinoma-bearing mice treated with azoxymethane (AOM). CBD reduced preneoplastic lesion and tumor formation induced by AOM and counteracted the upregulation of pAkt.⁴² Indeed, in colorectal cancer, CBD BDS significantly reduced the formation of AOM-induced aberrant crypt foci and polyps, as well as growth of xenograft tumor obtained by injection into nude mice.⁶³ A similar result was observed in prostate cancer, where CBD BDS reduced the size of xenografts generated from LNCaP cells.⁶²

Anticancer activity of some synthetic compounds has also been indicated in research. Treatment with the atypical cannabinoid – O-1602 – led to >30% reduction in tumor incidence and >50% decrease in tumor volume.⁶⁵ Further investigation revealed that these effects were associated with antiproliferative and proapoptotic activities of cannabinoids as shown by alterations in PCNA (decreased) and proapoptotic markers, such as BAX and p53, (increased) levels.⁶⁵

A combination treatment with the use of the synthetic compound Met-F-AEA and FAAH inhibitor URB597 resulted in inhibition of EGF signaling cascade in xenograftbearing mice, leading to reduced tumor growth.⁷⁵ Additionally, the use of Met-F-AEA combined with URB597 was shown to induce superior therapeutic response when compared to each substance alone.⁷⁵

Induction of autophagy

Recently, autophagy has drawn an increasing interest due to its dual role, depending on a cellular context and strength and duration of stimulating signal.^{77,78} In regard to cancer, it has been demonstrated that autophagy may either promote survival or induce cell death in the process termed programmed cell death type II.⁷⁸ In HT29 colon cancer cells, WIN treatment triggered an autophagic process, as shown by accumulation of p62 and LC3; however, prolonged incubation with WIN blocked an autophagic flux after autophagosome formation. Apparently, although autophagy has been implicated in the proapoptotic effect of several cannabinoids, in this case, WIN-induced death of colon cancer cells and autophagy seemed to play a prosurvival role.^{45,50,77,79}

In contrast, Donadelli et al⁴⁶ reported that gemcitabine (GEM) in combination with synthetic cannabinoids (arachidonoylcyclopropamide, GW405833, or SR141716) induced synergistic reduction in pancreatic adenocarcinoma cell growth by ROS-dependent autophagic cell death in an in vitro experiment. Importantly, combinations of GEM and cannabinoids exerted greater efficacy in cell growth inhibition in GEMresistant cell lines compared to GEM-sensitive ones, depending on the increase in intracellular ROS.⁴⁶ Since pancreatic cell lines have been shown to be more resistant to GEM due to lower basal levels of ROS, the combination therapy utilizing cannabinoids may help to overcome GEM resistance at least in pancreatic cancer cells.⁷⁹ It has been further evaluated that cannabinoid-induced autophagy is closely related to an increased AMP/ATP ratio and subsequent activation of AMPK in Panc1 cells.⁴⁵

There is evidence for cross talk between autophagy and apoptosis, and factors characteristic of the latter (GRP78, CHOP, TRB3) might also play a crucial role in the autophagic process.^{46,59,80,81} Moreover, autophagy is often suggested to

precede apoptosis and some autophagy-related molecules may facilitate apoptosis induction.^{49,82} In an in vitro study on human glioma cells, Δ^9 -THC evoked cancer cell death through the induction of autophagy, which was associated with accumulation of ceramide de novo and phosphorylation of eIF2 α , a protein related to ER stress response.^{51,83} THC and/or JWH-015 were shown to upregulate p8/TRB3 cascade, which further led to decreased phosphorylation of prosurvival Akt kinase and inhibition of Akt/mTORC1 axis in glioma and hepatocellular carcinoma (HCC) cells.^{51,80} The authors proposed that the latter is crucial for cannabinoidinduced autophagy of cancer cells.⁵¹ In breast cancer cells, O-1663, a THC and CBD analog, was reported to stimulate autophagy, as shown in LC3-II accumulation, and was even more active in inducing apoptosis than THC.⁸⁴

Unfortunately, our knowledge about the cannabinoidinduced autophagy of cancer cells in vivo is still scarce. In fact, it has been demonstrated only in a few animal cancer models. Treatment of mice bearing BRAF wild-type melanoma xenografts with the use of Sativex-like preparation (a mixture of equal doses of THC and CBD) induced substantial decrease in melanoma viability and tumor growth, paralleled by an activation of the autophagic process.⁸⁵

Similarly, THC administered intraperitoneally to U87MG xenograft-bearing mice produced decrease in tumor growth via the induction of autophagy.⁵¹ Regarding clinical research, analysis of tumors of two patients with recurrent glioblastoma multiforme demonstrated features of autophagic phenotype after the intracranial inoculation of THC.⁵¹ No side effects were observed in clinical trial patients or in tumor-bearing mice.⁵¹

Impact on metastasis formation

Cannabinoids are thought to modulate secretion of several proteins, leading to decrease in metastasis formation. Matrix metalloproteinases (MMPs) have long been associated with extracellular matrix degradation, the process intimately involved in the premetastatic niche development.⁸⁶ In particular, activation of TGF-β by MMP-2, MMP-9, and MMP-14 favors invasion of tumor cells and promotes metastasis outgrowth.⁸⁷ Overexpression of MMPs has been shown in various cancer types, and it had an unfavorable impact on patients' survival.^{34,88–91} The downregulation of MMP by cannabinoids has been demonstrated in several types of neoplasm.^{53,92–94} Solinas et al⁹⁴ showed a concentration-dependent decrease in proliferation and migration of human umbilical vein endothelial cells (HUVECs) after incubation with CBD, an effect associated with downregulation of

several angiogenesis-related proteins, particularly MMP-2 and MMP-9, and also TIMP-1, SERPINE1/PAI1, uPA, CXCL16, IL-8, ET-1, and PDGF-AA. In gastric cancer, WIN 55,212-2 treatment produced dose-dependent decrease in the expression of VEGF-A and MMP-2, but not in the levels of MMP-7 and MMP-9 proteins.⁵³

The metabolically stable anandamide analog Met-F-AEA was effective in inhibiting endothelial cells sprouting, and it was partly due to diminished MMP-2 secretion after treatment with Met-F-AEA.⁹³ Of note, exposition of HUVECs to SR141716, a CB1 antagonist, reversed the inhibitory effect of Met-F-AEA, confirming its dependency on activation of CB1 receptor.⁹³ In neuroblastoma cells, *N*-arachidonoylphenolamine (AM404), a paracetamol lipid metabolite, inhibited transcription factors NFAT and NF-κB, which resulted in decreased levels of MMP-1, MMP-3, and MMP-7 and subsequent inhibition in cell migration and invasion.⁹² Despite the fact that AM404 is known as a dual agonist of CB1 and TRPV1, its antimigratory activity was evoked independently of CB1 and TRPV1 binding.⁹²

Recently, ICAM-1 and its downstream target TIMP-1 were proposed to play a pivotal role in inhibiting carcinoma invasion and metastasis.95-98 Increased levels of ICAM-1 reduce the rate of tumor growth, and an inverse correlation between ICAM-1 expression and metastasis incidence was noted in several oncologic disorders.99-101 In line with this notion, there are data showing antimigratory and antiangiogenic activities of cannabinoid, accompanied by the upregulation of both ICAM-1 and TIMP-1.96-98 Ramer et al96 observed that CBD, Δ^9 -THC, and R(+)-methanandamide or JWH-133, a CB2 agonist, decreased migration and sprout formation of HUVECs suspended in conditioned media of A549 lung cancer cells. The same group of researchers also found time- and dose-dependent suppression of human cervical cancer (HeLa) cell invasion induced by methanandamide and THC.98 In both cases, observed phenomena were accompanied by increased expression of TIMP-1 and ICAM-1 molecules.96,98

In A549 and H460 lung cancer cell lines, CBD-induced upregulation of ICAM-1 enhanced susceptibility of tumor cells to adhesion of lymphokine-activated killer (LAK) cells and LAK cell-mediated cytotoxicity. Both effects were reversed by pharmacological or genetic blocking of ICAM-1 or by using antagonists of CB1, CB2, or TRPV1.¹⁰² Also adhesion molecules, such as P- and E-selectins, exhibit increased surface exposure on cancer cells in response to cannabinoids, which is related to enhanced recruitment of immune cells to the tumor microenvironment.^{48,103} Coupling to CB1 and CB2 receptors, 2-AG triggered an immune response against

bladder carcinoma cells through increased secretion of P- and E-selectins and enhanced TNF release by these cells and subsequent adhesion of Jurkat T lymphocytes.⁴⁸

Cannabinoids have shown antimetastatic effects in in vivo cancer models. A strong correlation between metastatic capacity of cancer cells and the expression of transcriptional regulator Id1 was suggested in research.¹⁰⁴ Id1 is considered as a tumor promoter and has been previously reported to induce formation of breast cancer metastasis.105,106 In one study, CBD, a natural component of C. sativa, reduced metastasis in advanced stages of breast cancer in vivo, mainly through the downregulation of transcriptional regulator Id1 and upregulation of ROS.84 Further investigation revealed that O-1663, a derivative of CBD and agonist of the CB2 receptor, is even more potent than its parent compound in inhibiting total metastasis in human MDA-MB231 and mouse 4T1 models of breast cancer metastasis. Furthermore, O-1663 appeared to be efficacious in prolonging survival in both tested preclinical models.84

It is already well known that neovascularization and angiogenesis play a prominent role in metastasis formation in tumors and contribute to tumor progression.¹⁰⁷ Strong migratory potential is attributed to invasive carcinoma cells, which additionally produce angiogenic factors, like VEGF, a key regulator of new blood vessels growth.^{107,108} VEGF has a major impact on tumor development, mainly through coupling and activating the tyrosine kinase receptor VEGFR2.¹⁰⁹ Accumulating evidence demonstrated antimigratory and antiangiogenic properties of plant-derived cannabinoids and cannabinoid-based drugs, although proangiogenic activity of some endocannabinoids has also been observed.¹¹⁰

Local administration of WIN 55,212-2 (CB1/CB2 agonist) or JWH-133 (CB2 agonist) resulted in considerable inhibition of tumor growth in a mouse model of skin tumor. This was accompanied by decrease in the size of blood vessels and decreased secretion of proangiogenic factor VEGF, placental growth factor, and angiopoietin 2, indicating impairment of vascularization.44 As shown in mouse gliomas, cannabinoid treatment (WIN 55,212-2 or JWH-133) lowered VEGF pathway, and similar results were observed in two patients with glioblastoma multiforme.43 Similarly, CBD inhibited VEGF-induced formation of capillary-like structures from HUVECs in the in vivo Matrigel sponge angiogenesis assay.94 In this experiment, antiangiogenic properties of CBD were demonstrated in decreased hemoglobin content of pellets obtained from C57/BL6 male mice.94 Similarly, Met-F-AEA exhibited an antiangiogenic effect, inducing a twofold or

eightfold reduction in the number of new capillaries, shown in the in vivo CAM assay.⁹³

The ability to modulate the tumor microenvironment by cannabinoids was also confirmed in animal cancer models. In the case of breast cancer, Elbaz et al⁴⁷ reported significantly decreased recruitment of tumor-associated macrophages, especially protumorigenic M2 type, to primary tumor and secondary tumor metastatic sites after the peritumoral administration of CBD. The authors suggested that CBD affects the cytokine secretion profile of cancer cells, which was further confirmed by lowered levels of CCL3 and GM-CSF, molecules implicated in the stimulation of macrophage chemotactic activity and formation of metastasis in various tumor types.^{47,111–113}

Expression of CBs and its prognostic value

To date, CBs have been detected in many cancer types and are proposed to be potential therapeutic agents due to their involvement in controlling survival and death of cells.^{40,114,115} Several studies have shown that the components of ECS present diverse expression in cancer, although the level and direction of this diversity are not always the same.^{116–119} Interestingly, CBs have been reported to be overexpressed in malignant cells of many cancer types compared to their nontransformed counterparts. Apparently, a lowered expression of CB in normal cells appears to be a protection mechanism from the proapoptotic and antiproliferative effects of some CB receptor ligands. It is hypothesized that increased CB expression may be associated with some features of cancer aggressiveness (Table 1).

The ECS has been studied most extensively in case of prostate cancer and central nervous system tumors. Sánchez et al¹²⁰ examined biopsies from human astrocytoma and reported that 70% of them expressed elevated levels of CB1 and CB2 and upregulated CB2 was directly related to tumor malignancy. Data presented by Ellert-Miklaszewska et al showing increased CB2 receptor in higher histologic grade of glioblastoma were in line with these results.¹²¹ Some data present an opposite expression of both CBs in glial cancers where significantly lower CB1 and greater CB2 immunoreactivities were seen in the membranes of glioblastoma multiforme in comparison to normal postmortem brain tissue.¹²² In this case, the authors speculate whether an increased expression of CB2 might be related to an increase in macrophages invading brain tumor.¹²²

Regarding prostate malignancy, elevated levels of CB1 and CB2 expression were reported in CA-human

Number	Receptor expression	Tumor	Influence	References
I	↑СВІ	Prostate cancer (stage IV)	↑ Gleason score, ↑ incidence of metastases at diagnosis, ↑ tumor size, ↑ rate of proliferation, ↓ disease-specific survival	40
2	↑ СВІ, ↑ СВ2	НСС	↑ disease-free survival	115
3	↑ СВ2	Colorectal cancer	\downarrow disease-free survival, \downarrow overall survival, \uparrow tumor growth	118
4	↑ СВ2	Astrocytoma	↑ tumor malignancy	120
5	↑ CB2	Glioblastoma	\uparrow histologic grade	121
6	↑ CBI	Prostate cancer	\uparrow Gleason score, \uparrow incidence of metastases at diagnosis,	123
			\uparrow tumor stage, \uparrow Ki67 index	
7	↑ CBI	Colorectal cancer	\downarrow disease outcome	124
8	↑ CBI	Colorectal cancer (stage II MSS)	\downarrow disease-specific survival	125
9	↑ СВ2	Head and neck squamous cell carcinoma	\downarrow disease-specific survival	128
10	↑ CB2	Glioma	↑ tumor malignancy	130
11	↓ CBI	PDAC	\downarrow survival	146

Table I CB expression in human tumors and its clinical relevance

Notes: \uparrow , increased; \downarrow , decreased.

Abbreviations: CB, cannabinoid receptor; HCC, hepatocellular carcinoma; MSS, microsatellite stable; PDAC, pancreatic ductal carcinoma.

papillomavirus-10 (virally transformed cells derived from adenocarcinoma of human prostate tissue) and other human prostate cancer cell lines (LNCaP, DU145, PC-3, CWR22Rv1) than in normal prostate epithelial cells and PZ-HPV-7 (virally transformed cells derived from normal human prostate tissue) cells.114 Similar results were obtained in other studies on prostate carcinoma where upregulated expression of CB1 and/or CB2 receptors was detected in cancer cells compared to their nonmalignant counterparts.^{40,123} With regard to prognosis, a high expression of CB1 in prostate tumor tissue, but not in normal tissue, is correlated with a higher Gleason score and presence of metastases at diagnosis.123 After a long follow-up period, it has been shown that patients with prostate carcinoma had significantly shorter disease-specific survival when malignant epithelial cells expressed a high CB1-immunoreactivity (CB1-IR) score.⁴⁰ This finding indicates association between CB1 and disease severity and outcome.³⁹ Likewise, high CB1-IR is an indicator of a poorer prognosis for patients (following surgery) in stage IV colorectal cancer as well as in stage II microsatellite stable colorectal cancer, which additionally is associated with shorter survival.^{124,125}

In human HCC, in situ hybridization analysis revealed overexpression of mRNAs for CB1 and CB2 in HCC samples (62% and 53%, respectively).¹¹⁵ Similarly, immunohistochemical analysis showed high expression of CB1 in 29 (45%) and CB2 in 33 (52%) of 64 HCC samples. In contrast to aforementioned studies, further evaluation indicated significantly longer disease-free survival time in HCC patients with higher CB1 and CB2 expression, implying usefulness of CB expression as a positive prognosis indicator in cancer.¹¹⁵ Such divergent results indicate that CB receptor expression and its prognostic value in human neoplasms depend strongly on the cancer type.

In addition to CB1 expression, high levels of other cannabinoid-like molecules and signaling pathways interacting with ECS have been shown in several types of malignancy.^{66,126,127} The data are shown in Table 2. Members of the GPRs family, in particular, GPR55, GPR119, and GPR18, were found in hCMEC/D3 human brain endothelial cells and A2058 melanoma cells.127 These findings were further supported by the results of two other reports. There was found a predominant expression of CB1 in Hodgkin-Reed-Sternberg cells of classical Hodgkin's lymphoma, whereas the surrounding, reactive nontumor lymphatic infiltrate was negative.¹²⁶ In the same study, GPR55 was strongly expressed in Hodgkin's lymphoma and non-Hodgkin's lymphoma cell lines. In melanoma cell line A375, not only CB1 and GPR55 but also TRPV1 and COX-2 receptors were highly expressed.⁶⁶ TRPV1 correlates with the degree of malignancy, as shown in PCC epithelial tissues, where elevated levels of CB1 and TRPV1 were observed.123

A strong expression of the CB2 receptor may contribute to poor prognosis in cancer patients as shown in primary squamous cell carcinoma of the head and neck.¹²⁸ It is most probably due to immunosuppressive properties of (endo) cannabinoids mediated via the CB2 receptor. In fact, it has been reported that THC administration to lung cancerbearing mice induced accelerated growth of tumor implant, mainly due to the augmentation of IL-10 and TGF- β and downregulation of IFN- γ release, indicating reduction in host immune response.¹²⁹ Furthermore, both T lymphocytes and antigen-presenting cells had impaired capability to generate alloreactivity against tumor.¹²⁹

Recently, augmented levels of CB1 and CB2 were observed in glioma tissue, and further analysis revealed a

Group	Molecule Primary cancer type/cancer cell line		References
GPRs	GPR55	HL: L428, L540, L1236, HDLM2, and KM-H2	126
		NHL: Karpas 422, BJAB, SUDHL8, and Farage	
	↑ GPR55	Melanoma: A375	66
	GPR18	Melanoma: A2058	127
	GPR55		
	GPR119		
Enzymes	↑ FAAH	Non-small cell lung cancer: A459, A549, CALUI, H460, and H1299	75
	↑ FAAH	Melanoma: A375	66
	\downarrow NAPE-PLD	Glioma	130
	↓ MAGL		
	\downarrow FAAH		
	↑ FAAH	Prostate: LNCaP and DU-145	138

Notes: \uparrow , increased; \downarrow , decreased.

Abbreviations: HL, Hodgkin's lymphoma; NHL, non-Hodgkin's lymphoma.

positive correlation between CB2 expression and tumor malignancy.¹³⁰ An increased expression of CB1 and CB2 was observed in high-grade glioma (World Health Organization [WHO] grades III–IV) compared with lowgrade glioma (WHO grades I–II).¹³⁰ With regard to ovarian cancer, immunohistochemical examination indicated that the expression of CB1 receptor was moderate in the benign and borderline epithelial tumors, while it was strong in the invasive tumors.¹³¹ It is quite an interesting observation since CB1 receptor (as well as FAAH) was found expressed in mouse ovarian surface epithelium, which is a major source of ovarian cancer.¹³² In human, CB1-IR was observed in ovarian cortex and medulla; however, the possibility of cannabinoid signaling contribution to ovarian carcinogenesis still exists and further investigation needs to be undertaken.¹³³

In colon cancer, it has been shown that CB2 overexpression might be considered as a poor prognostic factor in patients with more advanced stage, involvement of lymph nodes, and vascular invasion.¹¹⁸ The authors pointed out that since patients in advanced stage of cancer are often submitted to adjuvant therapy, CB2 may be considered as a marker of treatment resistance.¹¹⁸ This notion may be supported by the fact that CB2 expression is correlated with the expression of SNAIL1, a transcription factor known for its role in mediating epithelial-mesenchymal transition (EMT).^{118,134} EMT is a process by which cancer cells acquire migratory properties and ability to metastasize to distant sites.¹³⁵ The primary mechanism of SNAIL1-induced EMT is repression of epithelial markers, such as E-cadherin, and this effect promotes migratory, mesenchymal phenotype.^{134,135} It has been shown in several types of cancer that upregulation of SNAIL1 and further induction of EMT are associated with acquisition of radio- and chemoresistance.¹³⁴⁻¹³⁶ In colon

cancer cells, a CB2 receptor agonist induced delocalization of E-cadherin from membrane to cytoplasm and increase in SNAIL1 expression.¹¹⁸ The changes that are related to the EMT process might explain the reason why treatment of patients in advanced stages who overexpress CB2 could be less effective.

Interesting findings were provided by Schley et al¹³⁷ who reported abundant expression of CB2 in the endothelial cells of glioblastoma vessels. Given that active neovascularization is one of the most characteristic features of tumor invasion, the presence of CB2 in these areas may imply the possible exploitation of CB2 activation in cancer treatment.^{44,137} In fact, this assumption was further supported by the study in which two CB2 receptor agonists impaired growth of new skin tumor vessels.⁴⁴

Recently, other components of the ECS in human neoplasms have drawn great attention. Androgen-dependent LNCaP cells exhibit a high level of FAAH mRNA and protein expression, and FAAH immunoreactivity has been subsequently shown to be inversely correlated with disease severity and outcome.138,139 A case was made that FAAH immunoreactivity is regulated by the local tumor microenvironment. It is supported by the facts that the IL-4 receptor was found on epithelial cells expressing FAAH and the incubation of prostate cells with this cytokine increased their FAAH activity.¹³⁹ In fact, it was previously shown that lymphocyte treatment with IL-4 and IL-10 had a stimulating effect on FAAH, while incubation with IL-12 and IFN-y depressed the activity and lowered the expression of FAAH.¹⁴⁰ It is already well known that progression of malignant disease is associated with Th1/Th2 cytokine imbalance and immune response polarization toward production of immunosuppressive IL-4, IL-10, and TGF-B.141,142 An increased expression of Th2 cytokines has been shown in tumor tissue and infiltrating lymphocytes in several cancer types such as renal cancer, pulmonary carcinoma, and glioma.^{141,142} In some of them, increased levels of IL-10 were considered as an unfavorable prognostic factor and were correlated with stage and histological grade.^{143,144} Accordingly, a gradual loss of Th1 population and an increase in Th2 may contribute to high FAAH expression level in cancer. These findings also raise the question whether upregulation of this molecule due to Th2-dominant cytokine profile is responsible for poor prognosis in cancer patients.

In glial malignancy, decreased expression and activity of NAPE-PLD and MAGL were found.¹³⁰ Since NAPE-PLD and MAGL are the enzymes responsible for biosynthesis of anandamide and hydrolysis of 2-AG, respectively, their expression reflects the level of endogenous substrates in glioma tissues, namely, reduced level of anandamide and elevated level of 2-AG. However, with respect to FAAH, an anandamide-hydrolyzing enzyme, a significant down-regulation was reported. It is plausible that in this case, the depression of FAAH might be a compensatory response to a low level of its substrate, therefore suggesting that MAGL has a greater influence on AEA level than FAAH.¹³⁰

Although in most cases CB1 and CB2 are upregulated in cancer cells, there are reports indicating the opposite. Quantitative reverse transcription polymerase chain reaction and Western blot analyses revealed substantially reduced expression of CB1 receptor in colorectal carcinoma due to aberrant methylation within CpG islands of the CB1 promoter.¹¹⁹ This result may explain the general mechanism of CB1 expression loss, not only in colorectal cancer but also in all cancer types. Some interesting findings were demonstrated by Larrinaga et al¹¹⁶ who reported 12-fold downregulation of CB1 mRNA, whereas immunohistochemical analysis revealed positive staining of this protein in chromophobe renal cell carcinoma. A previous study of this group provided contradictory results and showed presence of mRNA encoding CB1 in normal and tumor tissues of clear cell renal carcinoma, while Western blot and immunohistochemistry failed to demonstrate any content of CB1 protein.117 It indicates the importance of not relying only on mRNA levels when evaluating the expression of certain molecules, as mRNA expression may not reflect the protein status in the tissue due to modifications at the posttranscriptional level.¹¹⁷ A similar lack of correlation between mRNA and protein levels has been demonstrated in studies on other kidney diseases.145

Regarding the prognostic value, in pancreatic ductal adenocarcinoma, a low level of CB1 is related to better

outcome, and this relationship is underlined by a significant correlation of high pain scores (18 out of 27 tissue specimens showing clinical pain demonstrated low receptor levels in nerves) with increased survival (median survival 11 months in low pain scores vs 24.5 months in high pain scores).¹⁴⁶ This result may be explained by the fact that pain is a warning symptom of ongoing disease process and it contributes to earlier diagnosis and treatment initiation.¹⁴⁶

An association of high expression of CB1 and CB2 receptors with poor prognosis in cancer patients is quite unexpected, considering the well-known antitumoral activity of CB ligands. A possible explanation for this ambiguity was given in a study on mouse astrocytoma cells.¹⁴⁷ Cudaback et al¹⁴⁷ showed that the level of CB1 and CB2 expression determines the efficacy of cannabinoids in cancer cells. It is suggested that the ECS mediates proapoptotic effects in cancer cells exhibiting low CB expression, while at high expression levels, it promotes cell survival due to activation of the Akt signaling pathway, known for its antiapoptotic effect.^{123,147} As summarized earlier, the ECS presents a heterogeneous pattern of expression in human cancers and this fact precludes its potential use as a prognostic marker in oncologic pathologies.

Conclusion

This review focused on current literature concerning ECS expression and the influence of its activation on cancer cell biology. As demonstrated earlier, CBs and other molecules closely related to ECS display a heterogeneous pattern of expression in human oncological pathologies, which probably impede its exploitation as prognostic markers in these diseases. Nevertheless, the fact that CB receptors are overexpressed in malignant cells compared to their nontransformed counterparts encouraged scientists to evaluate the influence of their ligands on several hallmarks of cancer. Indeed, synthetic and plant-derived cannabinoids, as well as endogenous compounds, exert a wide range of anticancer effects, including induction of autophagy and apoptosis, inhibition of proliferation, and reduction of metastasis outgrowth via antiangiogenic and antimigratory activities. They have also been found to stimulate immune response against cancer through enhanced recruitment of immune cells and, on the other hand, decreased migration of tumor-associated macrophages (particularly, protumorigenic M2 type) to the tumor microenvironment. Clearly, cannabinoids possess strong antiproliferative, proapoptotic, and antimetastatic properties, which have been confirmed in in vitro studies and in animal models. Despite a large amount of promising

evidence for cannabinoid-induced antitumoral action, there is a lack of clinical research addressing this issue. Currently, cannabinoids are used clinically only in palliative therapy in cancer patients owing to antiemetic and analgesic properties of these compounds. The only clinical trial undertaken so far concerned patients with recurrent glioblastoma multiforme.¹⁴⁸ In this study, intracranial delivery of Δ^9 -THC displayed a fair drug safety profile and did not induce overt psychoactive side effects.¹⁴⁸ Therefore, future research should be directed toward possible application of these compounds in therapy, at least as adjunctive treatment. In particular, combination therapy utilizing cannabinoids and conventional treatments may be a beneficial option for patients who do not respond to common therapies.

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Disclosure

The authors report no conflicts of interest in this work.

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