

Inter-alpha-trypsin inhibitor heavy chain 4: a novel biomarker for environmental exposure to particulate air pollution in patients with chronic obstructive pulmonary disease

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Abstract: Chronic obstructive pulmonary disease (COPD) is a chronic inflammatory disease that is correlated with environmental stress. Particulate matter $\leq 10 \mu\text{m}$ (PM_{10}) is considered to be a risk factor for COPD development; however, the effects of PM_{10} on the protein levels in COPD remain unclear. Fifty subjects with COPD and 15 healthy controls were recruited. Gene ontology analysis of differentially expressed proteins identified immune system process and binding as the most important biological process and molecular function, respectively, in the responses of PM_{10} -exposed patients with COPD. Biomarkers for PM_{10} in COPD were identified and compared with the same in healthy controls and included proteoglycan 4 (PRG4), inter-alpha-trypsin inhibitor heavy chain 4 (ITIH4), and apolipoprotein F (APOF). PRG4 and ITIH4 were associated with a past 3-year PM_{10} exposure level. The receiver operating characteristic curve analysis showed that ITIH4 is a sensitive and specific biomarker for PM_{10} exposure (area under the curve [AUC] = 0.690, $P=0.015$) compared with PRG4 (AUC = 0.636, $P=0.083$), APOF (AUC = 0.523, $P=0.766$), 8-isoprostane (AUC = 0.563, $P=0.405$), and C-reactive protein (CRP; AUC = 0.634, $P=0.086$). ITIH4 levels were correlated with CRP ($r=0.353$, $P=0.005$), suggesting that ITIH4 may be involved in an inflammatory mechanism. In summary, serum ITIH4 may be a PM_{10} -specific biomarker in COPD and may be related to inflammation.

Keywords: air pollution, apolipoprotein F, C-reactive protein, inflammation, proteoglycan 4, oxidative stress

Introduction

Epidemiological and toxicological research has identified adverse human health effects due to particulate air pollution exposure. Particulate matter $\leq 10 \mu\text{m}$ in aerodynamic diameter (PM_{10}) is associated with increased hospital admissions and mortality due to pulmonary and cardiovascular diseases.¹ Air pollutants participate in the development of pulmonary disease by interfering with both nonspecific and specific lung defenses.² Chronic obstructive pulmonary disease (COPD), for instance, is recognized as an environmentally related pulmonary disease that is associated with exposure to air pollution.³ Schikowski et al⁴ in 2014, provided evidence that the chronic effects of air pollution affect the prevalence and incidence of COPD among adults. Kumar et al⁵ (2013) noted the association between acute COPD exacerbation and particulate air pollution, finding that the risk of acute COPD exacerbation increased by 2.3% following a unit increase in $\text{PM}_{2.5}$ ($\leq 2.5 \mu\text{m}$). However, the effects of PM_{10} on protein expression in COPD are not well understood.

A protein biomarker is capable of objectively and quantitatively acting as an indicator to identify normal or pathological processes and the corresponding exposure to environmental stresses.⁶ For example, 8-isoprostane and C-reactive protein (CRP), the biomarkers for oxidative stress and inflammation, respectively, have been widely used to evaluate the cardiopulmonary toxicity of PM₁₀.^{7,8} However, they are nonspecific PM₁₀ biomarkers for oxidative inflammatory reactions. Thus, an increasing number of studies have attempted to identify more specific biomarkers for the assessment of specific pollutant exposures. For example, urinary p-nitrophenol has been reported to be a specific biomarker for occupational exposure to methyl parathion,⁹ and DNA modifications, such as *F2RL3* methylation, have been suggested as biomarkers for tobacco exposure.¹⁰ Ideally, a quality biomarker can be used for treatment and intervention decisions. However, biomarker measurement may also vary according to exposure sources and susceptible populations.

A useful biomarker not only differentiates stable disease from exacerbations but also predicts the severity of pollutant exposure. The objective of this study was to identify biomarkers for PM₁₀ exposure in patients with COPD. This study identified and assessed the use of serum biomarkers in the evaluation of PM₁₀ compared with the existing biomarkers of inflammation, namely 8-isoprostane and CRP. We used a proteomics approach to comprehensively screen protein profiles in healthy subjects and patients with COPD exposed to low and high levels of PM₁₀ over the past 3 years. The identified protein candidates were determined in serum samples obtained from 50 patients with COPD and 15 subjects without COPD. The specific and sensitive values of 8-isoprostane, CRP, and identified biomarkers were compared.

Materials and methods

Study population

We recruited 50 patients with COPD and 15 subjects without COPD (nonsmokers and smokers) in a medical center in Taiwan between January 2013 and August 2014. All subjects were between 40 years and 80 years of age at the time of inclusion. Patients identified with current cancer or active inflammatory disease or who had an exacerbation during the 4 weeks before the study were excluded. All subjects with COPD were informed of diagnosis and exhibited postbronchodilation forced expiratory volume in 1 second (FEV₁)/forced vital capacity (FVC) ratios of <70%. Subjects without COPD exhibited an FEV₁/FVC ratio of ≥75% and FEV₁ ≥80% of predicted value. The Ethics Committees of

Taipei Medical University-Joint Institutional Review Board approved the study protocol (number 201310027). The Chinese Clinical Trial Register number is ChiCTR-OCC-13004025. All subjects received written and oral information before inclusion and provided informed consent.

Data collection

Before recruitment into the study, a physical examination was performed, as was a clinical interview regarding smoking habits, comorbidities, and medicinal use. All subjects with COPD continued with a stable regimen of medications throughout the study. Lung function parameters were assessed at the time of recruitment using a Vitalograph Spirotac V™. Postbronchodilation measurements for FEV₁ and FVC were taken, and FEV₁/FVC was calculated. Serum samples were obtained and stored at -80°C for analysis.

PM₁₀ exposure

PM₁₀ data were obtained from 25 monitoring stations (operated by the Taiwan Environmental Protection Administration, Taiwan) throughout northern Taiwan. The daily concentrations of PM₁₀ were collected, corresponding to subject exposure. If a subject resided within 10 km of multiple monitoring stations, the weighted average was used to calculate the PM₁₀ levels. The PM₁₀ data were used to estimate the 1-year, 2-year, and 3-year effects of PM₁₀ on COPD.

Protein digestion

To determine the effects of PM₁₀ on protein expression in COPD, the five subjects with the highest average 3-year PM₁₀ values were classified as the High Ambient Particles (HAP) group (exposure range =63.2–64.5 µg/m³), and the five subjects with the lowest average 3-year PM₁₀ served as the Low Ambient Particles (LAP) group (exposure range =35.4–43.9 µg/m³). Serum samples from healthy controls, HAP, and LAP were collected and pooled together following protein digestion.¹¹ After protein depletion to remove albumin and immunoglobulin, the serum samples were diluted and denatured with 8 M urea/10 mM ammonium bicarbonate for 1 hour; they were then alkylated with 50 mM iodoacetamide for 30 minutes, tryptically digested in a solution of 50 mM ammonium bicarbonate at 37°C for 18 hours, and desalted on C18 columns.

Mass spectrometry and protein identification

Tryptic samples were analyzed using a mass spectrometer (LTQ Orbitrap XL; Thermo Fisher Scientific, Bremen,

Germany) coupled with an Ultimate 3000 RSLC system (Dionex, Sunnyvale, CA, USA) using previously documented protocols.¹² Briefly, peptides were separated using a 150 mm length $\times$ 100 mm inner diameter self-pulled spray tip (5 μ m tip opening) and packed with C18 material (ReproSil; 3 μ m; Dr Maisch, Ammerbuch, Germany). The linear gradient was from 5% to 10% of mobile phase B (mobile phase A: 0.5% acetic acid, mobile phase B: 80% acetonitrile/0.5% acetic acid) for 5 minutes, 10% to 40% of mobile phase B for 30 minutes, and 40% to 99% of mobile phase B for 5 minutes, with a total of 75 minutes' separation time. Five ranges of full mass spectrometry (MS) scans (m/z 350–2,000, m/z 350–600, m/z 600–800, m/z 800–1,200, and m/z 1,200–2,000) were followed by the selection of the ten most intense ions for MS/MS scans. Data were analyzed using the Proteome Discoverer 1.4 program to perform a Mascot database search of the National Center for Biotechnology Information and UniProt databases both for prediction as well as a literature search. The maximum mass tolerance was set to 10 ppm for precursor ions and 0.05 Da for fragment ions. Variable modifications for deamidation (NQ), oxidation (M), and fixed modification for carbamidomethyl (C) were selected for the search parameters.

Protein functional analyses

Unique proteins expressed in the HAP and LAP groups were analyzed using the Protein ANalysis THrough Evolutionary Relationships (PANTHER) classification system.^{13,14} The UniProt accession database was used to relate the expressed proteins identified in this study to related biological processes and molecular functions.

Enzyme-linked immunosorbent assay

Enzyme-linked immunosorbent assays (ELISAs) were used to determine the serum levels of 8-isoprostane (Cayman, Ann Arbor, MI, USA), CRP (CRP; R&D Systems, Minneapolis, MN, USA), proteoglycan 4 (PRG4), inter-alpha-trypsin

inhibitor heavy chain 4 (ITIH4), and apolipoprotein F (APOF; MyBioSource, San Diego, CA, USA) according to the manufacturer's instructions.

Statistical analysis

All ELISA experiments were performed at least three times. Data are expressed as the mean $\pm$ standard deviation. For multiple comparison analysis, one-way analysis of variance with Tukey's post hoc was used. Spearman's rank correlation coefficient was used to examine the correlation of quintiles of PM₁₀ with PRG4, ITIH4, and APOF, as well as the correlation of 8-isoprostane and CRP with PRG4, ITIH4, and APOF. Receiver operating characteristic (ROC) curve analysis was used to determine the specificity and sensitivity of PRG4, ITIH4, and APOF to PM₁₀ ($\geq 50 \mu\text{g}/\text{m}^3$ and $< 50 \mu\text{g}/\text{m}^3$) in COPD. Statistical analyses were performed using GraphPad Version 5 (La Jolla, CA, USA) for Windows. The level of significance was set to $P < 0.05$.

Results

Characteristics of study subjects

Fifty subjects with COPD, ten healthy controls, and five healthy smoking volunteers were enrolled. Detailed baseline characteristics of the 65 subjects in the study population are presented in Table 1. The ages of healthy controls, healthy smokers, and subjects with COPD were 67.8 ± 6.2 years, 59.8 ± 14.6 years, and 70.7 ± 8.4 years, respectively. The majority of the study population consisted of men with body mass indexes that ranged between $22.5 \text{ kg}/\text{m}^2$ and $24.8 \text{ kg}/\text{m}^2$. The current smoking percentage among the subjects with COPD was 9.1%. The FEV₁ percentages of predicted values for the healthy controls, healthy smokers, and subjects with COPD were $88.0\% \pm 11.5\%$, $85.5\% \pm 14.9\%$, and $42.6\% \pm 12.4\%$, respectively. The levels of 8-isoprostane and CRP for subjects with COPD were $13.4 \pm 3.0 \text{ pg}/\text{mL}$ and $5.6 \pm 6.4 \text{ mg}/\text{L}$, respectively. The average PM₁₀ concentrations for the study subjects were estimated, as shown in Table 2, and were $46.0 \pm 8.4 \mu\text{g}/\text{m}^3$.

Table 1 Baseline characteristics of healthy control subjects, smokers, and patients with COPD

Categorical variables	Healthy controls (n=10)	Healthy smokers (n=5)	Patients with COPD (n=50)
Age, years	67.8 $\pm$ 6.2	59.8 $\pm$ 14.6	70.7 $\pm$ 8.4
Men, %	75.0	75.0	95.5
BMI, kg/m ²	22.5 $\pm$ 2.9	24.8 $\pm$ 4.0	23.8 $\pm$ 3.7
Current smoking, %	0	100	9.1
FEV ₁ , % predicted	88.0 $\pm$ 11.5	85.5 $\pm$ 14.9	42.6 $\pm$ 12.4
8-Isoprostane, pg/mL	ND	ND	13.4 $\pm$ 3.0
CRP, mg/L	ND	ND	5.6 $\pm$ 6.4

Note: Data is presented as mean $\pm$ SD.

Abbreviations: BMI, body mass index; FEV₁, forced expiratory volume in 1 second; CRP, C-reactive protein; ND, no data.

Table 2 Environmental exposure to PM₁₀ over the previous 1 year, 2 years, and 3 years in the study subjects with COPD

	PM ₁₀ , µg/m ³ (Mean ± SD)
1-year average	46.0±8.4
2-year average	48.5±7.2
3-year average	50.8±7.0

Abbreviations: PM₁₀, particulate matter ≤10 µm; SD, standard deviation.

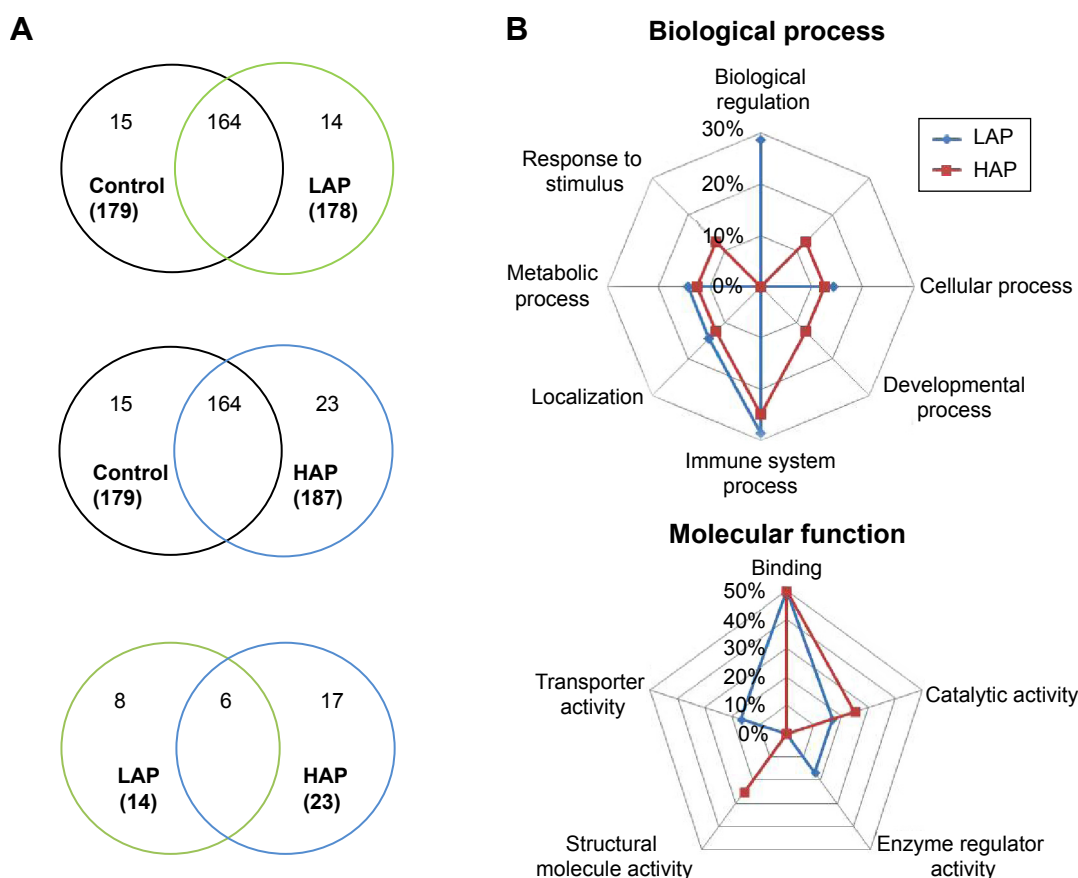
for the 1-year average, 48.5±7.2 µg/m³ for the 2-year average, and 50.8±7.0 µg/m³ for the 3-year average.

Selection of potential biomarkers in patients with COPD

Overlaps in protein profiles between healthy controls and the LAP- and HAP-exposed subjects with COPD were compared to investigate protein expression in COPD (Figure 1A). A total of 179 proteins were identified in the healthy controls, and 178 and 187 proteins were identified

in the LAP- and HAP-exposed subjects with COPD, respectively (Table S1). A total of 14 and 23 proteins were unique to LAP- and HAP-exposed subjects with COPD, respectively. Six serum proteins were common to both LAP- and HAP-exposed subjects with COPD: PRG4, ITIH4, APOF, Ig kappa chain V-I region DEE, type I cytoskeletal 16 keratin, and type II cytoskeletal 6A keratin.

PANTHER analysis was conducted on the proteins unique to LAP- and HAP-exposed subjects with COPD to understand their biological context (Figure 1B). The common biological processes identified in the LAP and HAP groups included biological regulation (29% for LAP), cellular component organization or biogenesis (13% for HAP), cellular process (14% for LAP and 13% for HAP), developmental process (13% for HAP), immune system process (29% for LAP and 25% for HAP), localization (14% for LAP and 13% for HAP), metabolic process (14%

**Figure 1** Protein expression and biological process and molecular functions determined by PANTHER.

Notes: (A) Venn diagram of proteins that are common among or unique to patients with COPD exposed to low ambient particles (LAP; range: 35.4–43.9 µg/m³) and high ambient particles (HAP; range: 63.2–64.5 µg/m³) compared with healthy controls exposed to either LAP or HAP. Six proteins were commonly expressed in the serum samples: proteoglycan 4 (PRG4), inter-alpha-trypsin inhibitor heavy chain 4 (ITIH4), apolipoprotein F (APOF), Ig kappa chain V-I region DEE, type I cytoskeletal 16 keratin, and type II cytoskeletal 6A keratin. (B) The biological processes and molecular functions of the unique proteins identified in the LAP and HAP groups. Immune system process and binding are the important biological process and molecular function, respectively, in response to PM₁₀ in COPD.

Abbreviation: COPD, chronic obstructive pulmonary disease.

for LAP and 13% for HAP), and response to stimulus (13% for HAP), with immune system process terms being the most common. The common molecular functions identified between the LAP and HAP groups included binding (50% for LAP and 50% for HAP), catalytic activity (17% for LAP and 25% for HAP), enzyme regulator activity (17% for LAP), structural molecule activity (25% for HAP), and transporter activity (17% for LAP), with binding terms being the most common.

Validation of PRG4, ITIH4, and APOF in patients with COPD and control subjects

The levels of PRG4, ITIH4, and APOF in healthy controls, smokers, and subjects with COPD are presented in Figure 2. There were no differences in the PRG4, ITIH4,

and APOF levels between healthy controls and healthy smokers. The PRG4 levels in the subjects with COPD were significantly higher than those in healthy controls or smokers ($P<0.05$). The ITIH4 levels in subjects with COPD were lower than those in healthy controls ($P<0.05$) but not those of healthy smokers. There was no significant difference in the APOF levels across subjects with COPD, healthy controls, or smokers.

Correlation of PM_{10} with PRG4, ITIH4, and APOF

The changes in PM_{10} for the 1-year, 2-year, and 3-year averages in quintiles were related to PRG4, ITIH4, and APOF (Figure 3). The PRG4 levels were correlated with 2-year ($r=0.278$, $P=0.033$) and 3-year ($r=0.349$, $P=0.007$)

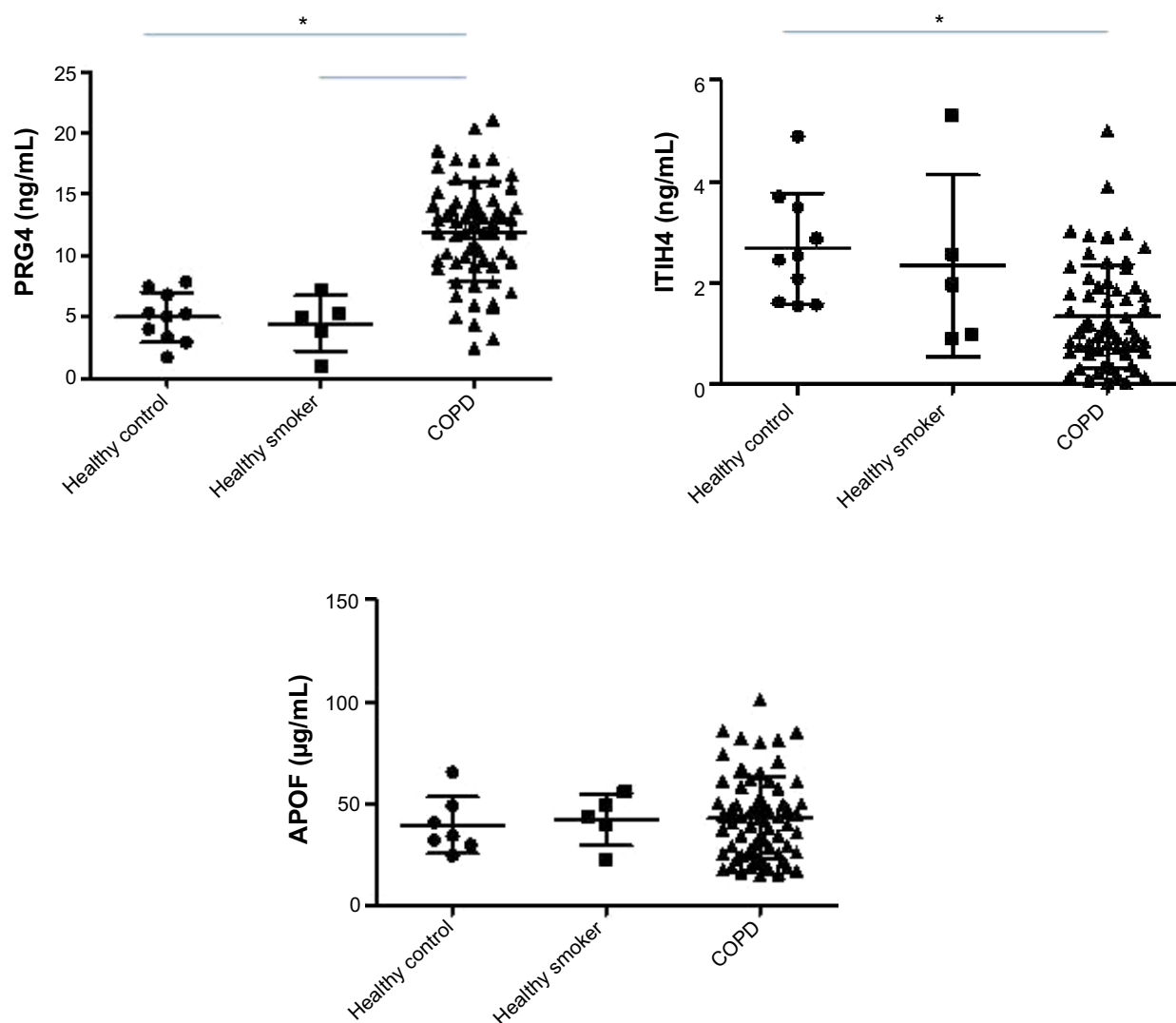


Figure 2 Serum PRG4, ITIH4, and APOF levels in healthy controls, smokers, and subjects with COPD.

Notes: The PRG4 levels in subjects with COPD were significantly higher than in healthy controls and smokers, whereas the ITIH4 level in COPD was lower than in healthy controls (* $P<0.05$).

Abbreviations: APOF, apolipoprotein F; COPD, chronic obstructive pulmonary disease; ITIH4, inter-alpha-trypsin inhibitor heavy chain 4; PRG4, proteoglycan 4.

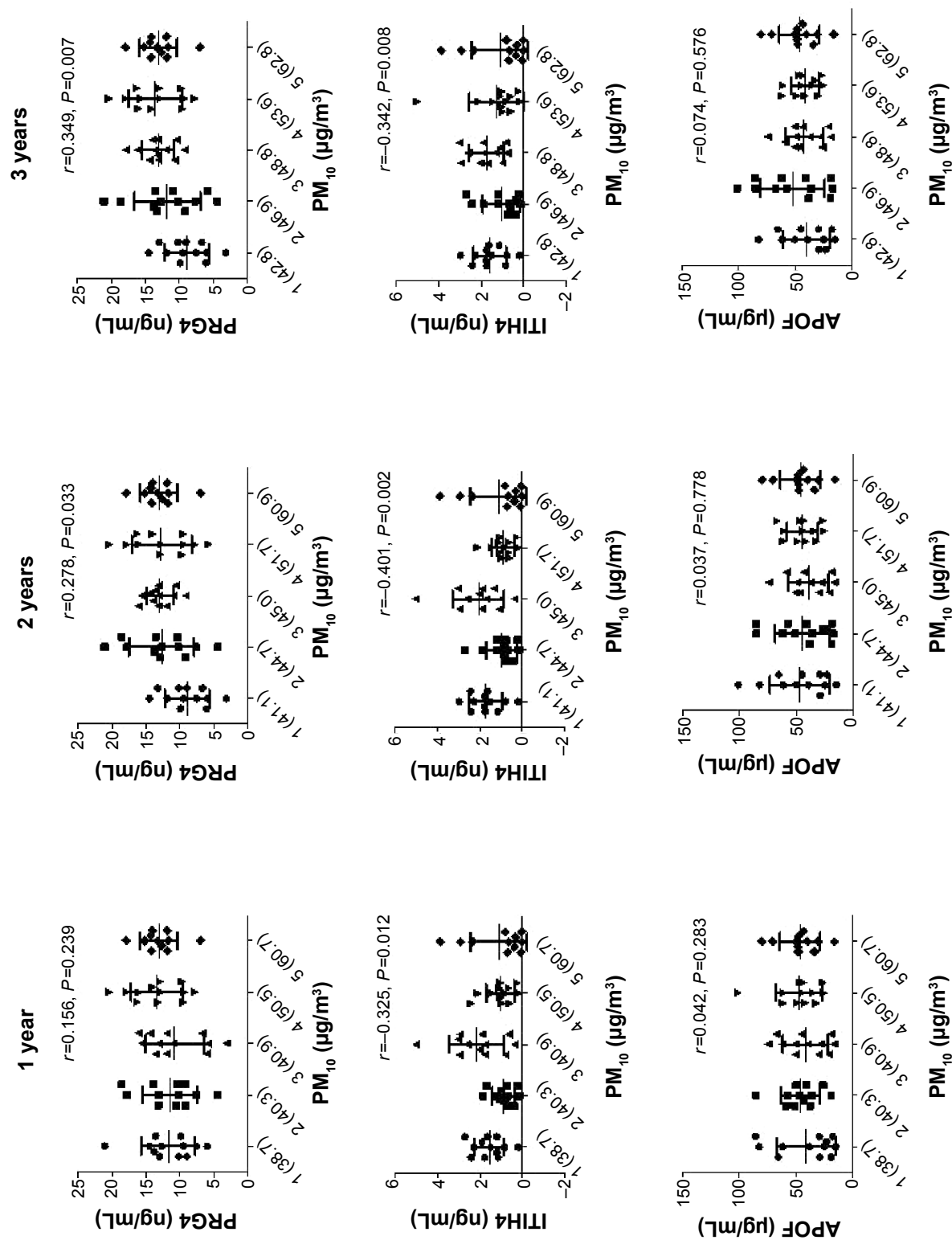


Figure 3 The relationship of serum PRG4, ITIH4, and APOF levels to 1-year, 2-year, and 3-year changes in PM_{10} levels.

Notes: Study subjects were divided equally based on quintiles of PM_{10} . The values in parentheses along the X-axis represent the mean PM_{10} values for each quintile group. Spearman's rank correlation coefficient was used to examine the correlation of quintiles of PM_{10} with PRG4, ITIH4, and APOF. PRG4 levels were correlated with the 2-year ($r=0.278$, $P=0.033$) and 3-year PM_{10} averages ($r=0.349$, $P=0.007$), whereas the ITIH4 levels were correlated with the 1-year ($r=-0.325$, $P=0.012$), 2-year ($r=-0.401$, $P=0.002$), and 3-year ($r=-0.342$, $P=0.008$) PM_{10} averages.

Abbreviations: APOF, apolipoprotein F; ITIH4, inter-alpha-trypsin inhibitor heavy chain 4; PM_{10} , particulate matter $\leq 10 \mu m$; PRG4, proteoglycan 4.

PM₁₀ averages, whereas the ITIH4 levels were correlated with the 1-year ($r=-0.325$, $P=0.012$), 2-year ($r=-0.401$, $P=0.002$), and 3-year PM₁₀ averages ($r=-0.342$, $P=0.008$). There was no correlation between APOF and PM₁₀ levels over the previous 3 years.

Figure 4 shows the ROC curve of the sensitivity and specificity obtained for the PRG4, ITIH4, and APOF levels when discriminating between the $\geq 50 \mu\text{g}/\text{m}^3$ and $< 50 \mu\text{g}/\text{m}^3$ average 3-year PM₁₀ levels in patients with COPD. The area under the curve (AUC) value was 0.563 (95% confidence

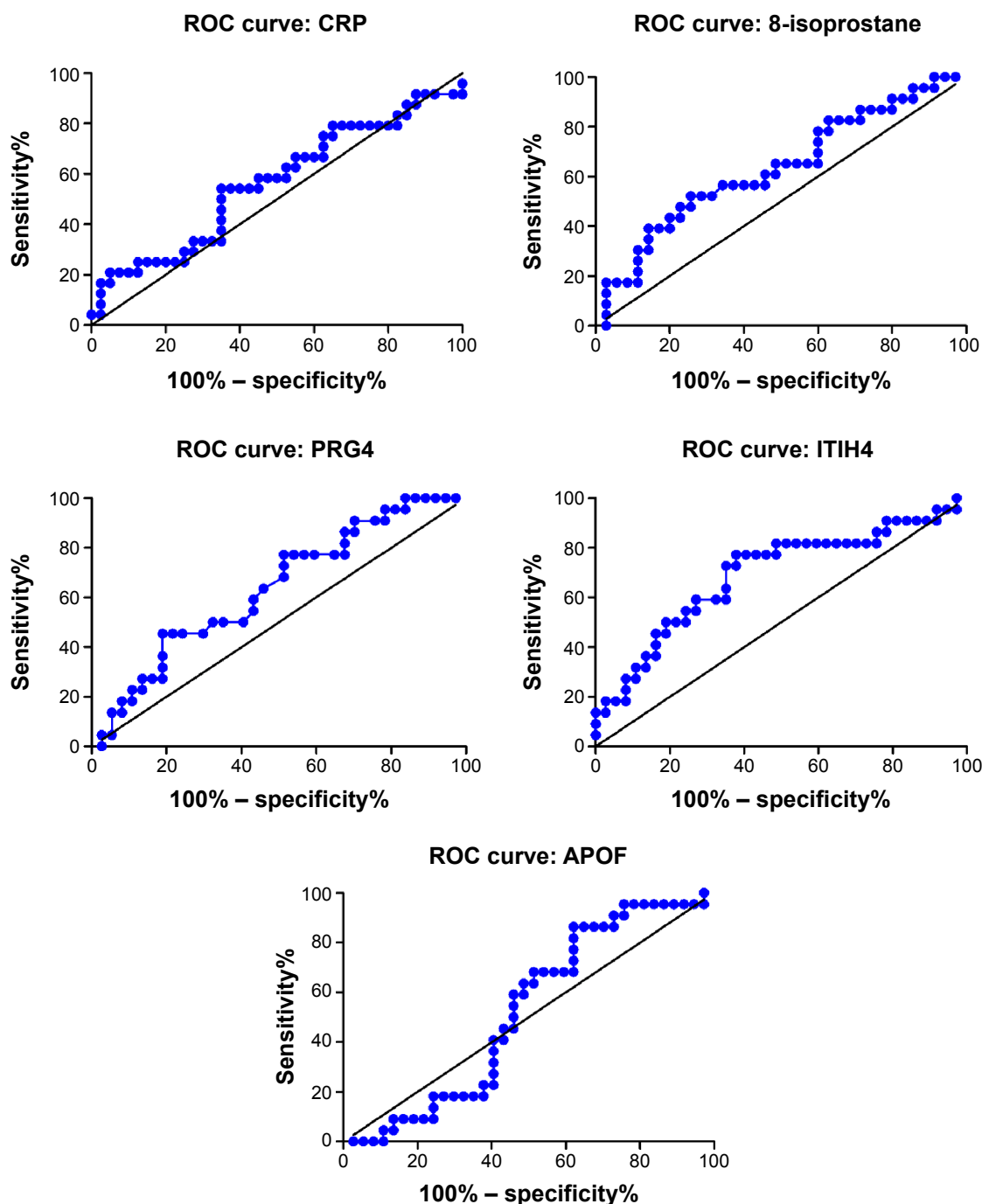


Figure 4 Diagnostic performance of serum 8-isoprostane, CRP, PRG4, ITIH4, and APOF in receiver operating characteristic (ROC) curve analyses.

Notes: ROC curve of the sensitivity and specificity of PRG4, ITIH4, and APOF levels for discriminating between the $\geq 50 \mu\text{g}/\text{m}^3$ and $< 50 \mu\text{g}/\text{m}^3$ average 3-year PM₁₀ value in COPD. The area under the curve value was 0.563 (95% CI: 0.413–0.713, $P=0.405$) for 8-isoprostane, 0.634 (95% CI: 0.486–0.782, $P=0.086$) for CRP, 0.636 (95% CI: 0.492–0.780, $P=0.083$) for PRG4, 0.690 (95% CI: 0.545–0.836, $P=0.015$) for ITIH4, and 0.523 (95% CI: 0.376–0.671, $P=0.766$) for APOF.

Abbreviations: APOF, apolipoprotein F; CI, confidence interval; COPD, chronic obstructive pulmonary disease; CRP, C-reactive protein; ITIH4, inter-alpha-trypsin inhibitor heavy chain 4; PM₁₀, particulate matter $\leq 10 \mu\text{m}$; PRG4, proteoglycan 4.

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Correlation of 8-isoprostane and CRP with PRG4, ITIH4, and APOF

Correlations between 8-isoprostane and CRP with PRG4, ITIH4, and APOF for COPD were determined (Figure 5).

There were no statistical associations of 8-isoprostane and CRP to PRG4 and APOF in patients with COPD. Only ITIH4 was correlated with the CRP levels ($r=0.353$, $P=0.005$).

Discussion

COPD, a chronic inflammatory disease, is recognized by irreversible airflow obstruction and by structural modifications in the proximal and peripheral airways.¹⁵ An abnormal inflammatory response of the lung to PM₁₀ appears to play a central role in the pathogenesis of COPD;^{16,17} however, few

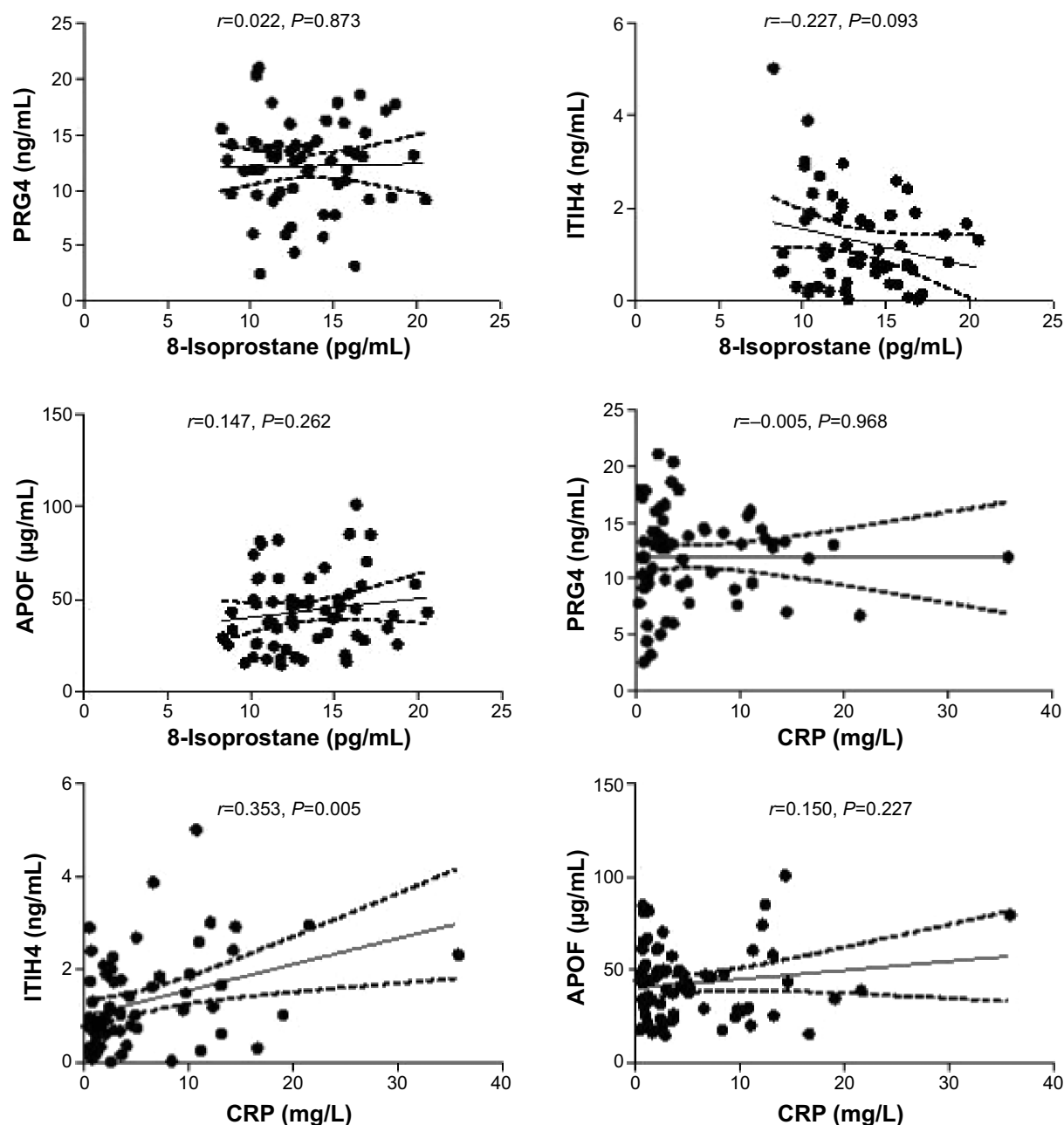


Figure 5 Correlation of 8-isoprostane and CRP to PRG4, ITIH4, and APOF in subjects with COPD.

Notes: The Spearman's rank correlation coefficient for CRP and ITIH4 was $r=0.353$ ($P=0.005$).

Abbreviations: APOF, apolipoprotein F; COPD, chronic obstructive pulmonary disease; CRP, C-reactive protein; ITIH4, inter-alpha-trypsin inhibitor heavy chain 4; PM₁₀, particulate matter $\leq 10 \mu\text{m}$; PRG4, proteoglycan 4.

studies have investigated protein profiles in patients with COPD residing in polluted urban areas. The present study discovered differences in protein expression in patients with COPD who were exposed to low and high levels of PM₁₀ (LAP and HAP). Three major findings are reported in the present study: 1) immune system process and binding are the most important biological process and molecular function, respectively, in the response to PM₁₀ in patients with COPD, 2) ITIH4 is a biomarker for PM₁₀ in COPD, and 3) ITIH4 is associated with inflammatory response.

Oxidative stress and inflammatory reactions are considered common biological responses that result from the inhalation of PM₁₀. Toxicology exposure experiments in animals and humans have identified several biological mechanisms that may be affected by the inhalation of PM₁₀, including oxidative stress and systemic inflammation.¹⁸ The present study identified the immune system process as an important biological process in the response to PM₁₀ in COPD and the binding terms as important molecular functions. Immune response is considered an important determinant for the exacerbation of COPD,^{19,20} and immune disorder commonly associated with PM₁₀ contributes to lung function impairment and mortality. Systemic inflammation is considered a common link between COPD and immune response.²¹ Furthermore, a link between binding and COPD has been reported.²² For example, a heavy subunit with a regulatory light subunit can modulate glutamate cysteine ligase activity.²³ Notably, a key common feature between PM₁₀ exposure and COPD is “inflammatory response.” Oxidative inflammatory reactions may be a result of exposure-related variances in protein regulation; therefore, an investigation of the effect of PM₁₀ on protein profiling in COPD is important.

Discovering biomarkers for susceptible populations assists with human environmental exposure assessment and risk evaluation. Despite increased efforts to identify COPD biomarkers, however, the gap between clinical and environmental medicine is substantial. We identified six biomarker candidates from healthy controls and from LAP- and HAP-exposed patients with COPD using a proteomics approach on serum samples: PRG4, ITIH4, APOF, Ig kappa chain V-I region DEE, type I cytoskeletal 16 keratin, and type II cytoskeletal 6A keratin. Type I cytoskeletal 16 keratin and type II cytoskeletal 6A keratin are associated with epithelial tissues from nail beds, esophagus, tongue, and hair follicles,^{24,25} which are less likely related to the mechanisms of PM₁₀ and COPD. Thus, we next examined PRG4, ITIH4, and APOF as candidate biomarkers. We observed that the PRG4 levels in subjects with COPD were significantly higher than those in healthy controls and smokers,

whereas the ITIH4 levels were significantly lower than those in healthy controls. These results suggest PRG4 and ITIH4 could be the potential biomarkers for COPD and that APOF is not sensitive as a potential biomarker for COPD and PM₁₀ exposure. We then correlated the PM₁₀ levels for the previous 1-year, 2-year, and 3-year averages with these three biomarker candidates. The correlations showed that PRG4 and ITIH4 may be biomarkers for alteration in PM₁₀ levels, particularly ITIH4. Previously, 8-isoprostane and CRP were characterized as biomarkers of oxidative stress and inflammation, respectively, in response to particulate air pollution.^{7,26} Therefore, we used them for the comparison with PRG4, ITIH4, and APOF in COPD. The US Environmental Protection Agency’s health-based national air quality standard for PM₁₀ (annual average of 50 µg/m³) was used as the cutoff level for the ROC analysis. We discovered that ITIH4 is the only significant biomarker for PM₁₀ in COPD; no significance was observed for 8-isoprostane, CRP, PRG4, and APOF. This finding suggests that ITIH4 is a good biomarker for evaluating PM₁₀ exposure in subjects with COPD.

To understand the potential mechanisms underlying PM₁₀-regulated COPD in the present study, 8-isoprostane and CRP were correlated with PRG4, ITIH4, and APOF. Statistical significance was observed between CRP and ITIH4 in COPD, suggesting that ITIH4 may be involved in inflammatory responses. ITIH4 is a type II acute phase protein and has been associated with inflammatory responses.^{27,28} Our findings are consistent with those of a previous study that observed correlation between the ITIH4 and CRP levels.^{27,28} However, the correlations between PM₁₀ and ITIH4 are negatively associated with increasing PM₁₀ concentrations. CRP is commonly used in the assessment of several acute clinical conditions, such as COPD.²⁹ Notably, CRP levels are also elevated in many other conditions, including infective, inflammatory, and neoplastic pathologies.^{30–32} Because of CRP’s involvement in multiple pathologies, ITIH4 may be a more specific biomarker for PM₁₀ in COPD than CRP.

Conclusion

In conclusion, circulating ITIH4 is an important biomarker to assess PM₁₀ exposure in patients with COPD and could be involved in postexposure inflammatory responses. Furthermore, metabolic process and catalytic activity are the important biological process and molecular function, respectively, in response to PM₁₀ exposure in COPD. The limitation of the present study is the sex distribution in the study subjects. The sex difference may associated with the biomarker identification for PM₁₀ exposure. Future studies are

required to investigate the potential mechanisms underlying PM₁₀ exposure and to determine whether ITIH4 can provide a specific index for evaluating PM₁₀ exposure in COPD.

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Author contributions

All authors have contributed substantially to the concept and design of the study, the drafting of the article, and the critical revision of the manuscript for important intellectual content. All authors have read and approved the final version of the manuscript for publication.

Disclosure

The authors report no conflicts of interest in this work.

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