

Synthesis and extracellular accumulation of silver nanoparticles by employing radiation-resistant *Deinococcus radiodurans*, their characterization, and determination of bioactivity

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Abstract: There has been rapid progress in exploring microorganisms for green synthesis of nanoparticles since microbes show extraordinary diversity in terms of species richness and niche localization. Microorganisms are easy to culture using relatively inexpensive and simple nutrients under varied conditions of temperature, pressure, pH, etc. In this work, *Deinococcus radiodurans* that possesses the ability to withstand extremely high radiation and desiccation stress has been employed for the synthesis of silver nanoparticles (AgNPs). *D. radiodurans* was able to accumulate AgNPs in medium under various conditions, and process optimization was carried out with respect to time, temperature, pH, and concentration of silver salt. AgNPs were characterized using UV/vis spectroscopy, scanning electron microscopy, transmission electron microscopy, X-ray diffraction, energy-dispersive X-ray spectroscopy, and Fourier transform infrared spectroscopy. The microbially synthesized AgNPs exhibited good antimicrobial activity against both Gram-negative and Gram-positive organisms and anti-biofouling activity. Their ability to inhibit growth and proliferation of cancer cell line was also examined, and it could be seen that AgNPs synthesized using *D. radiodurans* exhibited excellent anticancer activity.

Keywords: *Deinococcus radiodurans*, silver nanoparticles, anticancer, radiation resistance, antibacterial, anti-biofouling

Introduction

Nanotechnology has gained tremendous public interest due to the possibility of applications of nanomaterials in many areas such as industry, agriculture, medicine, diagnostics, and drug delivery. Nanomaterials act as a connecting link between bulk materials and molecular and atomic structures. They exhibit completely new and improved properties when compared with the bulk material, particularly with respect to specific characteristics such as size, distribution, morphology, and spectral and electrical properties. Silver nanoparticles (AgNPs), in particular, have been demonstrated to have diverse potential applications. The strong antibacterial activity against a wide range of pathogens and their potential in wound healing are among the most exploited applications of AgNPs in the medical field, due to which they are used in advanced bandages and dressing burn wounds.¹ AgNPs have been exploited in designing silica-silver core-shell nanoparticles and utilized for rapid detection of microbes, proteins, and antibodies.^{2,3} AgNPs have also been used in bone cements in artificial joint replacements, since their presence drastically reduced the wear and tear of the polymer.^{4,5} Although AgNPs have promising role in biomedical applications, human toxicity and environmental hazards posed by

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them cannot be neglected. It is reported that indiscriminate use of AgNPs and silver-containing medications has led to increased cases of argyria.⁶ In addition, nano-antimicrobials when released in natural systems may pose a serious threat to the beneficial bacteria in these systems, thereby leading to eutrophication.⁷ Therefore, a better understanding about the biological interactions of these nanoparticles is required to develop safe nano-antimicrobials.

Currently, many methods have been reported for the synthesis of AgNPs using chemical, physical, photochemical, and biological routes.⁸ The chemical and physical methods of nanosilver production are often expensive and also involve the use of toxic and hazardous chemicals, which may pose potential environmental and biological risks. To reduce the dreadful effects of chemical synthesis and to obtain novel nanoparticles at a cheaper rate, green synthesis is increasingly being explored. An ecofriendly solvent system and availability of biocompatible reducing and capping agents are the basic requirements for green synthesis.⁹ In addition, biological methods are inexhaustible, economic, and can be operated at ambient temperature and pressure conditions. Various microbes are known to reduce the silver (Ag^+) ions to form AgNPs, most of which are found to be spherical particles.¹⁰ Studies have indicated that culture supernatants of several bacteria like *Escherichia coli*, *Enterobacter cloacae*, *Klebsiella pneumoniae*, *Bacillus* spp., *Lactobacillus acidophilus*, *Staphylococcus aureus*, and several psychrophilic bacteria like *Pseudomonas antarctica*, *Pseudomonas proteolytica*, *Pseudomonas meridi-ana*, *Arthrobacter kerguelensis*, and *Arthrobacter gangot-riensis* can be used to generate AgNPs.^{8,11,12} Some of these microorganisms can survive and grow even at high metal ion concentrations. Metal-reducing bacteria are exposed to extreme environmental conditions and possess specific defense mechanisms to quell such stresses, including the toxicity of foreign metal ions or metalloids^{13,14} and hence can be utilized in the synthesis of nanoparticles. Dissimilatory metal-reducing bacteria *Geobacter sulfurreducens* have shown the ability of actively reducing soluble Pd(II) to Pd(0) extracellularly, thus reducing the toxicity of metal ion.¹⁵ Fe(III)-reducing bacterium *Shewanella algae* reduces Au^{3+} ions in anaerobic environments to form 10–20 nm gold nanoparticles¹⁶ and is capable of producing extracellular M-substituted magnetite nanoparticles using akaganeite and dopants in soluble form.¹⁷ *Deinococcus radiodurans*, the most radiation-resistant organism having a potential to sustain the presence of various heavy metals such as Fe(III), Cr(VI), U(VI), and Tc(VII),¹⁸ has not yet been explored for its ability to synthesize nanoparticles. Genome manipulation has been extensively used to clone and express

genes encoding bioremediation functions in *Deinococcus*, for example, the use of *merA* to detoxify highly toxic, thiol-reactive Hg(II), to much less toxic and nearly inert elemental and volatile Hg(0),¹⁹ the *tod* and *xyl* operons to oxidize toluene and reduce Cr(VI) in sediment microcosms,²⁰ and *phoN* that is capable of bioprecipitation and biorecovery of uranium.²¹ Previously, the S-layer lattices from *D. radiodurans* have been successfully used as a biotemplate for guided self-assembly of commercially procured hexagonal and honeycomb-ordered arrays of the dendrimer-encapsulated platinum nanoparticles, citrate-capped gold nanoparticles, and various species of CdSe/ZnS core-shell quantum dots.²² The present study involves the synthesis and extracellular accumulation of AgNPs using the bacterium *D. radiodurans*. UV/vis spectroscopy, transmission electron microscopy (TEM), scanning electron microscopy (SEM), and energy-dispersive spectroscopy (EDX) were used to characterize these AgNPs. In addition, Fourier transform infrared spectroscopy (FTIR) was performed for functional group identification, and crystal structure studies were performed using X-ray diffraction (XRD).

This report highlights, for the first time, the use of radiation- and desiccation-resistant *D. radiodurans* for the synthesis of AgNPs. These nanoparticles exhibit excellent antibacterial, anti-biofouling, and anticancer activity.

Materials and methods

Materials

D. radiodurans strain used was R1 ATCC BAA-816. Silver nitrate (AgNO_3) was purchased from Merck (Mumbai, India). Tryptone glucose yeast (TGY) extract broth and Luria-Bertani (LB) broth were purchased from HiMedia (Mumbai, India). *E. coli* NCIM 2739, *Proteus vulgaris* NCIM 2027, and *Pseudomonas aeruginosa* NCIM 2948, which were procured from National Collection of Industrial Microorganisms (Pune, India), and *S. aureus* and *Bacillus subtilis* (soil isolates) were used for antibacterial assay. MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) was purchased from Sigma-Aldrich Co. (St Louis, MA, USA) for cytotoxicity studies.

Strain and culture conditions

D. radiodurans strain R1 ATCC BAA-816 was grown aerobically in TGY (1% bacto tryptone, 0.5% bacto yeast extract, 0.1% glucose) broth with agitation (160 rpm), or on TGY-agar plates (1.5% bacto agar) at 32°C. Bacterial growth was assessed by measuring turbidity (optical density at 600 nm, OD_{600}) of broth cultures or by determining colony-forming units (CFU) on TGY agar plates incubated at 32°C for 48 hours.

Synthesis and extracellular accumulation of AgNPs

Overnight-grown culture of *D. radiodurans* was spun at 5,000 rpm for 10 minutes, and the pellet thus obtained was resuspended in fresh TGY and incubated at 32°C with 160 rpm till the culture attained an OD₆₀₀ of 0.6. The midlog phase culture of *D. radiodurans* was adapted for 2 hours using 1 mM AgNO₃. The adapted culture was spun at 5,000 rpm for 10 minutes, and the pellet was resuspended in the TGY with different concentrations of AgNO₃ (1 mM, 2.5 mM, 5 mM, and 10 mM) for 24 hours. The bioreduction and extracellular accumulation of the silver ions in the solution using *D. radiodurans* were monitored at regular intervals by sampling the supernatant and measuring the absorption spectrum of the solution using UV/vis spectrophotometer at a resolution of 1 nm. Appropriate controls (inoculated medium without AgNO₃ and uninoculated medium with AgNO₃ salt) were run simultaneously. UV/vis spectra of these sample aliquots were recorded as a function of time of reaction from 400 nm to 800 nm on a UV/vis dual-beam spectrophotometer at room temperature (28°C).

Characterization of AgNPs

The preliminary characterization of the AgNPs was performed using UV/vis spectroscopy technique (Jasco dual-beam spectrophotometer V-630). UV/vis spectrum of the *D. radiodurans* supernatant (different time interval cultures were spun at 5,000 rpm for 10 minutes) was taken in order to check the optimum time when maximum production of AgNPs was observed using different concentrations of AgNO₃ solution (1 mM, 2.5 mM, 5 mM, and 10 mM). Different pH ranges (4.8, 6.8, and 8.8) and temperatures (20°C, 32°C, and 37°C) were also optimized. For FTIR analysis, 500 µL of the synthesized AgNPs (2.5 mM) was lyophilized for 30 minutes, and the pellet thus obtained was resuspended in chloroform. The size and morphological characterization of the synthesized AgNPs were carried out using SEM and TEM (Technai F-30). TEM sample was prepared on 400 mesh carbon-coated copper grid, and silicon wafer was used for SEM sample preparation. Elemental analysis to confirm the presence of AgNPs in the test sample was performed using SEM-EDX (Quanta-200, 3D dual beam, source used – tungsten thermionic emission). EDX spectrum was recorded in the spot-profile mode by focusing the electron beam onto a region on the surface coated with silicon wafer. A thick film of synthesized AgNPs was made on a piece of glass slide; film was allowed to dry and used for XRD analysis.

Antibacterial assay

Agar well diffusion method

Three Gram-negative bacteria *E. coli* NCIM 2739, *P. vulgaris* NCIM 2027, and *P. aeruginosa* NCIM 2948, and two Gram-positive bacteria *S. aureus* and *B. subtilis* (soil isolates) were grown overnight in LB medium and spun at 5,000 rpm for 5 minutes. The pellet was washed with phosphate-buffered saline and resuspended in LB medium. The antibacterial activity was tested using agar well diffusion method.²³ One hundred microliters of the suspended culture was spread uniformly on LB plates. The solid medium was then gently punctured with the help of cork borer to make wells, and 50 µL of the synthesized AgNPs (150 µg/mL) was added in the respective well. The plates were incubated at 37°C for 24 hours. The antibacterial activity was assayed by measuring the diameter of the inhibition zone formed around the well. Kanamycin (5 mg/mL), ampicillin (5 mg/mL), and streptomycin (5 mg/mL) were used as positive controls, while sterile water served as a negative control.

Standard dilution micromethod

Antimicrobial activities of the synthesized AgNPs were assessed using the standard dilution micromethod with *E. coli* and *S. aureus* as representative Gram-negative and Gram-positive organisms. A concentration of 10⁶ CFU/mL of each bacterial culture was exposed to 150 µg/mL concentration of AgNP solution; in the corresponding control, deionized water was added, and all flasks were incubated for 3 hours at 37°C with continuous shaking (200 rpm). Aliquot from each culture was withdrawn after 3 hours, and the growth of bacteria was monitored by plating the culture on Luria agar plates and measuring the CFU per milliliter. The counts from three independent experiments corresponding to a particular sample were averaged.

Anti-biofouling assay

The *S. aureus* and *P. aeruginosa* cultures were grown overnight in 96-well microtiter plate at 37°C. One hundred microliters of bacterial suspension along with different concentrations of AgNPs (15–120 µg) was pipetted to each well of a 96-well microtiter plate and incubated at 37°C. After 24 hours of incubation, the medium was discarded, and wells were thoroughly washed with phosphate-buffered saline (pH 7.2). One hundred microliters of 0.1% crystal violet was added to each well and left for 30 minutes. The stain was then discarded, and the plate was thoroughly washed. For quantification of attached cells, the crystal violet was solubilized in absolute ethanol, and the absorbance was measured

at 570 nm.²⁴ The OD value was used as an index to observe the ability of these organisms to form biofilm. Reduction of the biofilm was correlated with the AgNPs treatment to the cells. These experiments were carried out in triplicates, and the average values were calculated.

Anticancer activity

Cytotoxicity assay

The mammalian breast cancer cell line (MCF-7) was utilized to examine the cytotoxic effect of synthesized AgNPs by using MTT assay and colony-forming assay.²⁵ MCF-7 cells (7,000 cells/well) were seeded into a 96-well plate and exposed to a concentration range of 0–30 µg of AgNPs. After 24 hours of incubation, 20 µL of 5 mg/mL MTT was added to each well and further incubated for 3 hours. Medium was removed, and 150 µL of dimethyl sulfoxide was added. Plate was covered with foil and incubated on orbital shaker for 15 minutes. Absorbance was read at 550 nm with a reference filter of 660 nm. The percent viability was calculated using the following formula.

$$\% \text{ Viability} = \frac{\Delta A_{550} (\text{treated})}{\Delta A_{550} (\text{control})} \times 100$$

Colony-forming assay

The breast cancer cells (MCF-7) (50 cells/well) were seeded into a 24-well plate and incubated overnight. These cells were treated with different concentration (0–7.5 µg) values below lethal dose (LD₅₀) of nanoparticles for 10 days, medium was removed, and cells were fixed using chilled ethanol. Cells were stained with crystal violet (0.1% in ethanol) for 30 minutes. The plate was washed with water and allowed to dry. Numbers of colonies were counted. Percent viability was calculated using the following formula.

$$\% \text{ Viability} = \frac{\text{Colony count in treated}}{\text{Colony count in untreated}} \times 100$$

Statistical analysis

Statistically significant differences between groups were determined using the Student's *t*-test and analysis of variance.

Results and discussion

Microbial synthesis of AgNPs

Microbially synthesized AgNPs were reddish brown in color. Figure 1A shows a color change from yellow to reddish brown after incubation of *D. radiodurans* in the presence of AgNO₃, which was indicative of the synthesis of AgNPs.

Spectrophotometric absorption measurements (UV/vis) in the wavelength range of 400–450 nm are used in characterizing the AgNPs.^{26,27} Figure 1B represents the UV/vis spectrum of culture supernatant containing AgNPs indicating the surface plasmon resonance centered at approximately 426 nm, confirming the extracellular accumulation of AgNPs in the solution. Microbial synthesis of nanoparticles with different sizes and shapes depends on the organisms involved, concentration of AgNO₃, temperature, pH, and incubation period.²⁸ To determine the influence of these factors on AgNPs synthesis, the experiment was carried out with variable concentrations of AgNO₃ salt, temperatures, times of incubation, and pH. A prominent absorption peak was seen at 426 nm, which is characteristic of AgNPs synthesized at 32°C (Figure 1C), pH 6.8 (Figure 1D), and an AgNO₃ concentration of 2.5 mM (Figure 1E) when incubated for 24 hours (Figure 1F). A progressive increase in the characteristic peak with increase in reaction time is a clear indication of nanoparticles formation. It is well studied that though AgNPs have highly spherical and face-centered cubic crystal structure, various anisotropic shapes can be grown by controlling the assembly of metal atoms in the solution.^{26,29,30} Different sizes and shapes of AgNPs exhibit different spectra and can be used in different applications.

SEM and TEM analysis

Microscopic techniques such as SEM and TEM were utilized to characterize the morphology of nanoparticles. The SEM analysis showed that the synthesized AgNPs were spherical in shape (Figure 2A). To obtain better resolution, the AgNPs were scanned using TEM (Figure 2B). From the TEM image, it is evident that the entire surface of the grid was evenly spread, and the AgNPs were well dispersed. The particle size histogram revealed that the average particle size of the AgNPs was 16.82 nm as represented in Figure 2C. The size of nanoparticles ranges from 4 nm to 50 nm with maximum percentage of particles of 15–20 nm and some particles of 5–8 nm sizes, while a very small percentage with diameters ranging from 45 nm to 50 nm could be visualized (data not shown). These variations in shape and size of nanoparticles synthesized by biological systems are common.³¹ TEM images for other preparation conditions using different concentrations of AgNO₃, 5 mM and 10 mM, were also obtained. The average size observed for 2.5 mM was 17 nm (Figure 3A), which was consistent with previous data. The concentration of 5 mM exhibited an average size of 14.41 nm (Figure 3B), while 10 mM showed an average size of 13.45 nm (Figure 3C). There was no significant difference observed in size of AgNPs synthesized using 2.5 mM AgNO₃ salt and other concentrations used

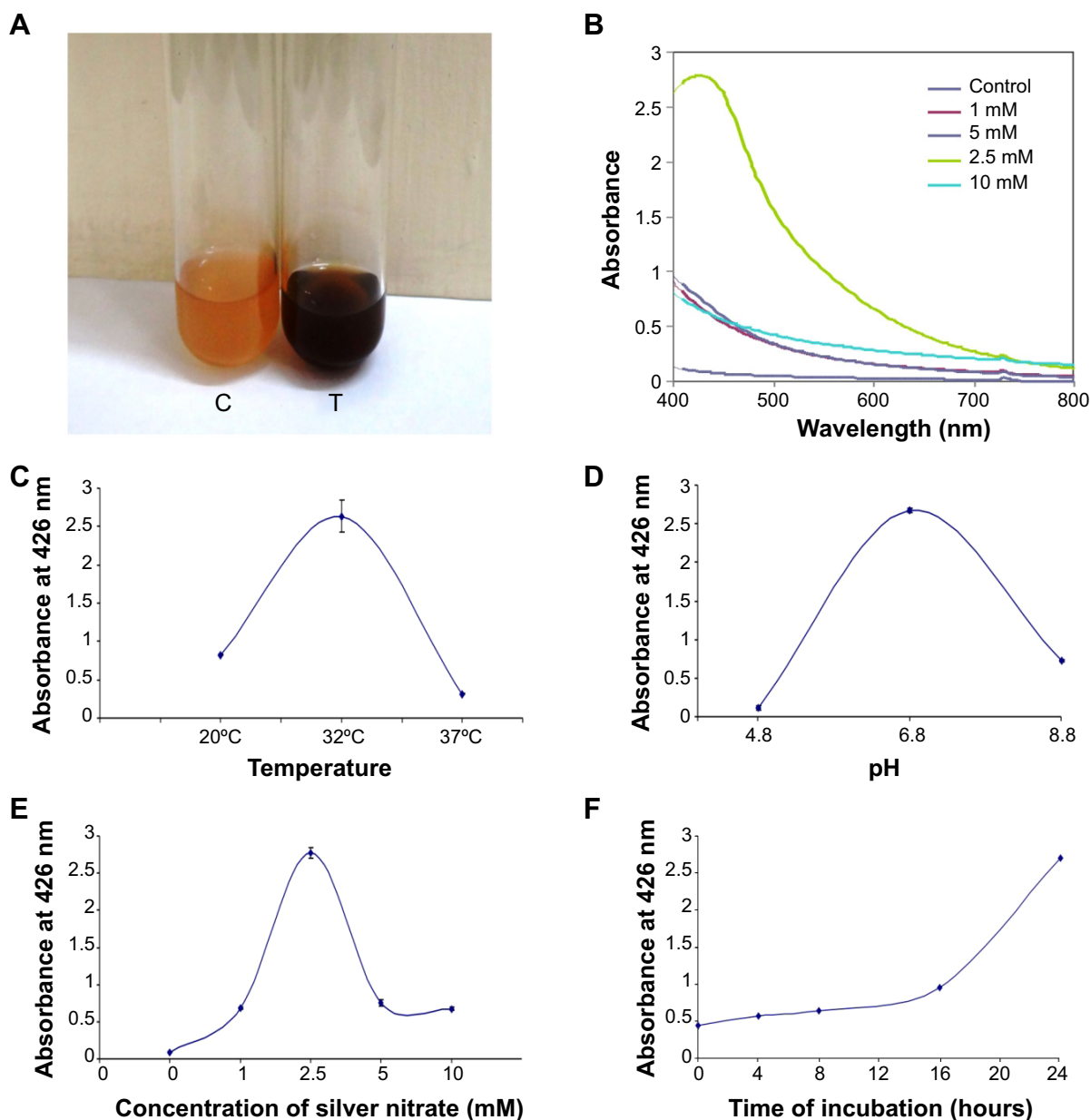


Figure 1 Synthesis and optimization of AgNPs.

Notes: (A) Tubes containing the culture supernatant before (C – control) and after (T – test) immersion of *Deinococcus radiodurans* in 2.5 mM silver nitrate solution for 24 hours. (B) UV/vis spectral analysis of culture supernatant containing the AgNPs over a wavelength range of 400–800 nm after 24 hours of reaction. Prominent peak for UV/vis spectra is visualized at 426 nm. Optimization of physicochemical parameters for silver nanoparticle synthesis: (C) temperature, (D) pH, (E) concentration of silver nitrate, and (F) incubation time. The error bars represent mean \pm standard deviation.

Abbreviation: AgNPs, silver nanoparticles.

(5 mM and 10 mM AgNO_3 salt). Earlier reports have shown that AgNPs synthesized using various microbes like *Pseudomonas stutzeri* AG259, *Bacillus megaterium*, *K. pneumoniae*, and *Bacillus licheniformis* exhibit a larger particle size of 200 nm, 80–98.56 nm, 28.2–122 nm, and 50 nm, respectively;^{32–35} however, biologically synthesized AgNPs using *D. radiodurans* are comparatively of smaller size (16.82 nm). Nanoparticles with smaller size can have promising applications in biomedicine due to the advantage of increased surface area.

FTIR

FTIR measurements were performed to identify the possible biomolecules responsible for bioreduction, capping, and/or stabilizing the AgNPs synthesized using *D. radiodurans*. In order to obtain good signal-to-noise ratio of AgNPs, the spectrum was taken in the range 500–3,100 cm^{-1} . Figure 4A shows the prominent intensity peaks at 1,591.30 cm^{-1} , 1,395.39 cm^{-1} , and 3,197.97 cm^{-1} . The functional groups such as C–H stretch, CH_3 –R, N–H, C–O–C, aromatic C–C skeletal

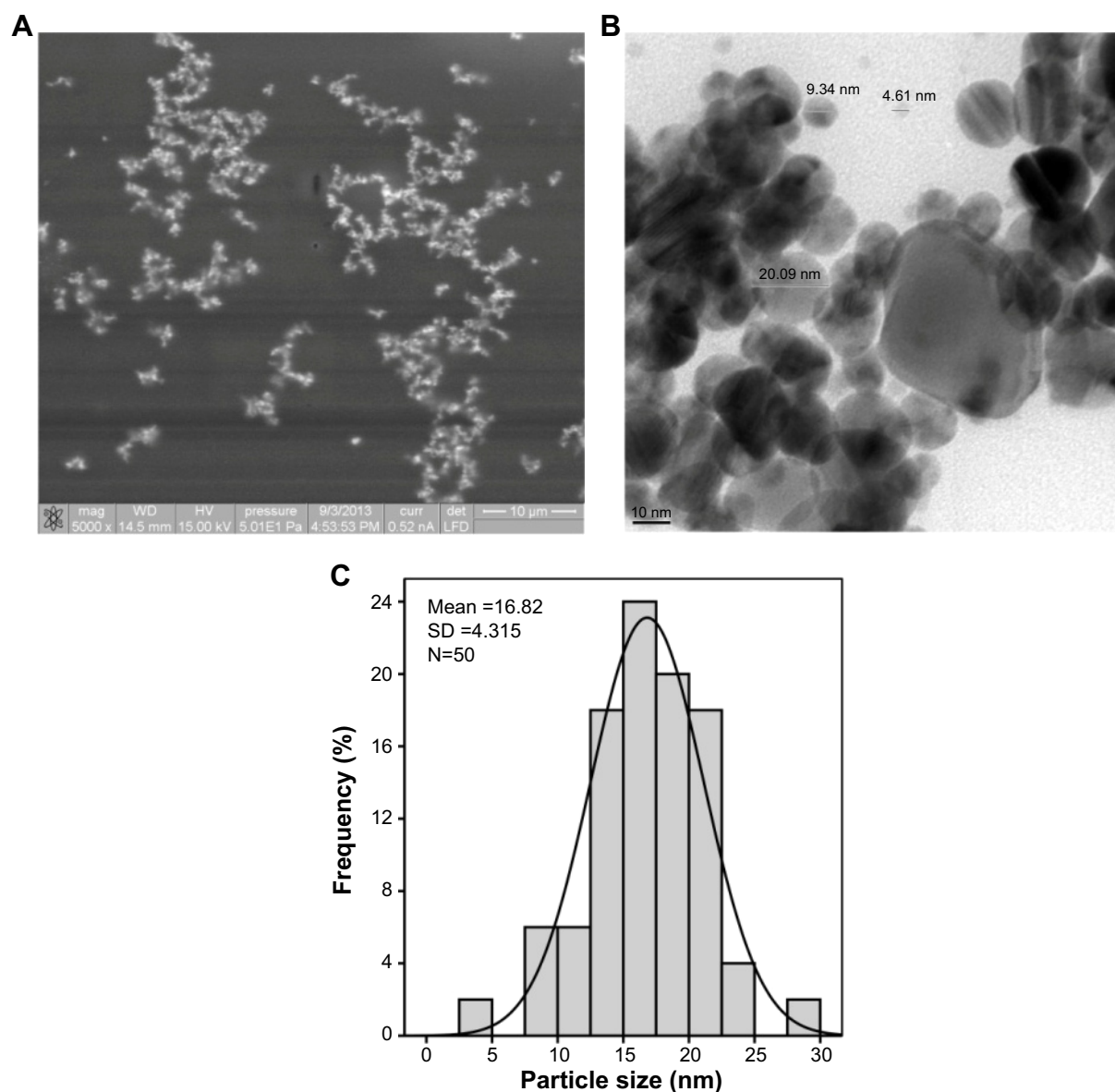


Figure 2 Morphology of synthesized AgNPs.

Notes: (A) Scanning electron microscopic observation of AgNPs at $\times 5,000$; insert bar corresponds to 10 μ m. (B) TEM image of AgNPs; insert bar corresponds to 10 nm. (C) Particle size histogram evaluated from corresponding TEM micrograph is shown in (B).

Abbreviations: AgNPs, silver nanoparticles; TEM, transmission electron microscopy.

vibrations, $-\text{COO}$, $-\text{NO}_3$, thioester, and S-S stretch were observed at different wave numbers. Sastry et al reported that functional groups like $-\text{C}-\text{O}-\text{C}-$, $-\text{C}=\text{C}-$, and $-\text{COO}$ are derived from heterocyclic compounds like proteins present in the fungal extract and are capping ligands of nanoparticles.³⁶ In our study, the amide linkage and other functional groups revealed through the FTIR analysis may probably play a role in the interaction of synthesized nanoparticles with the proteins/peptides of *D. radiodurans*, thereby stabilizing the AgNPs.

Energy-dispersive spectroscopy

Elemental analysis performed using EDX (Figure 4B) confirmed the presence of metallic AgNPs. In addition, the Si

signal observed was most likely caused by X-ray emission from silicon wafer support used in EDX analysis.²⁷

XRD measurements

The XRD technique is used to establish the metallic nature of particles. Figure 4C depicts the XRD pattern of AgNP films deposited on a glass substrate. XRD analysis of AgNPs synthesized using *D. radiodurans* showed clear diffraction peaks at 38.36, 45.67, 44.64, and 77.420, indexing the planes 211, 111, 200, and 103 of the crystalline silver. Planes 211, 111, and 200 correspond to cubic structure of nanoparticles, while the plane 103 corresponds to hexagonal phase.³⁷ It therefore suggests that the AgNPs synthesized

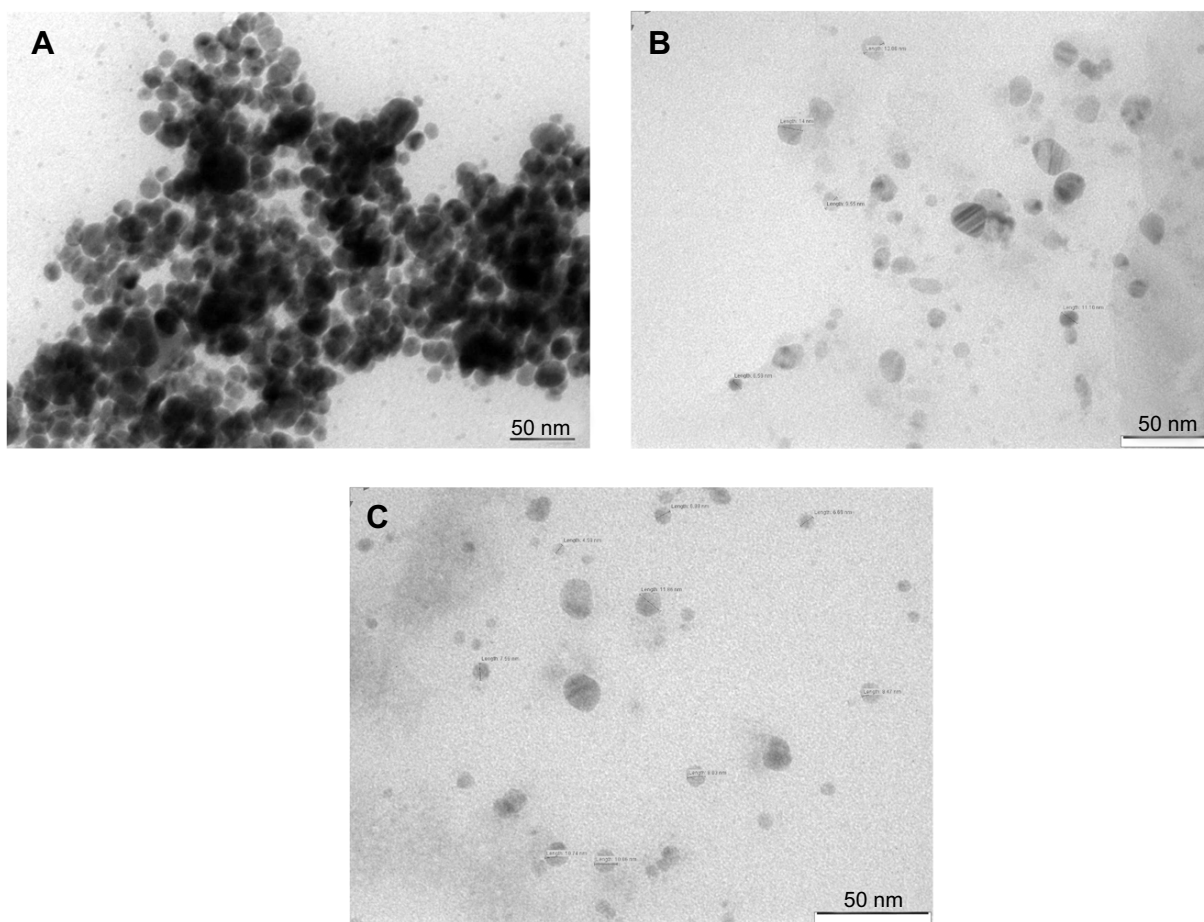


Figure 3 Transmission electron microscopic images of AgNPs.

Notes: AgNPs synthesized using different concentrations of AgNO_3 : (A) 2.5 mM, (B) 5 mM, and (C) 10 mM. Insert bar corresponds to 50 nm.

Abbreviation: AgNPs, silver nanoparticles.

using *D. radiodurans* are biphasic in nature. This slight shift in the peak positions indicated the presence of strain in the crystal structure, which is a characteristic of nanocrystallites. The data obtained were matched with the database of Joint Committee on Powder Diffraction Standards (file numbers 870718 and 721426). According to this match, the peaks 2.344, 2.027, and 1.984 correspond to silver, while the peak 2.799 corresponds to silica as the sample was prepared and scanned on a glass slide. The XRD data clearly indicated the crystalline structure of the AgNPs. Thus, the XRD pattern along with UV/vis spectra and TEM images provides an evidence for the presence of AgNPs.

Antibacterial activity

Several studies have demonstrated that biosynthesized AgNPs alone or in combination with antibiotics tend to have strong bactericidal activity against Gram-negative and Gram-positive bacteria³⁸ including multidrug-resistant strains such as *E. coli*, *P. aeruginosa*, and *S. aureus*.³⁹ In this study, the antibacterial activity of the synthesized AgNPs was

investigated using the agar well diffusion method and standard dilution micromethod. The results for the inhibition zones and their average values obtained from agar well diffusion assay are shown in Figure 5A. In liquid growth experiments, 91% growth inhibition was observed when *E. coli* (10^6 CFU/mL) was treated with 150 $\mu\text{g/mL}$ concentration of AgNPs for 3 hours, and a decline in CFU per milliliter was seen from 1.54×10^7 (untreated control) to 1.42×10^6 (AgNPs treated), while in the case of Gram-positive *S. aureus* (10^6 CFU/mL), treatment with 150 $\mu\text{g/mL}$ of AgNPs for 3 hours exhibited a 46% growth inhibition with CFU per milliliter reducing from 8.6×10^7 (untreated control) to 4.64×10^7 (AgNPs treated). Therefore, it was apparent that all bacterial cultures used in this study were inhibited by AgNPs. Gram-negative bacteria were highly suppressed by the AgNPs compared to Gram-positive bacteria. Among the Gram-negative bacteria, *P. vulgaris* was inhibited the most followed by *E. coli* and *P. aeruginosa*, while among the Gram-positive bacteria, *S. aureus* was more susceptible followed by *B. subtilis*. Antibacterial activity of AgNPs (160–200 $\mu\text{g/mL}$) against

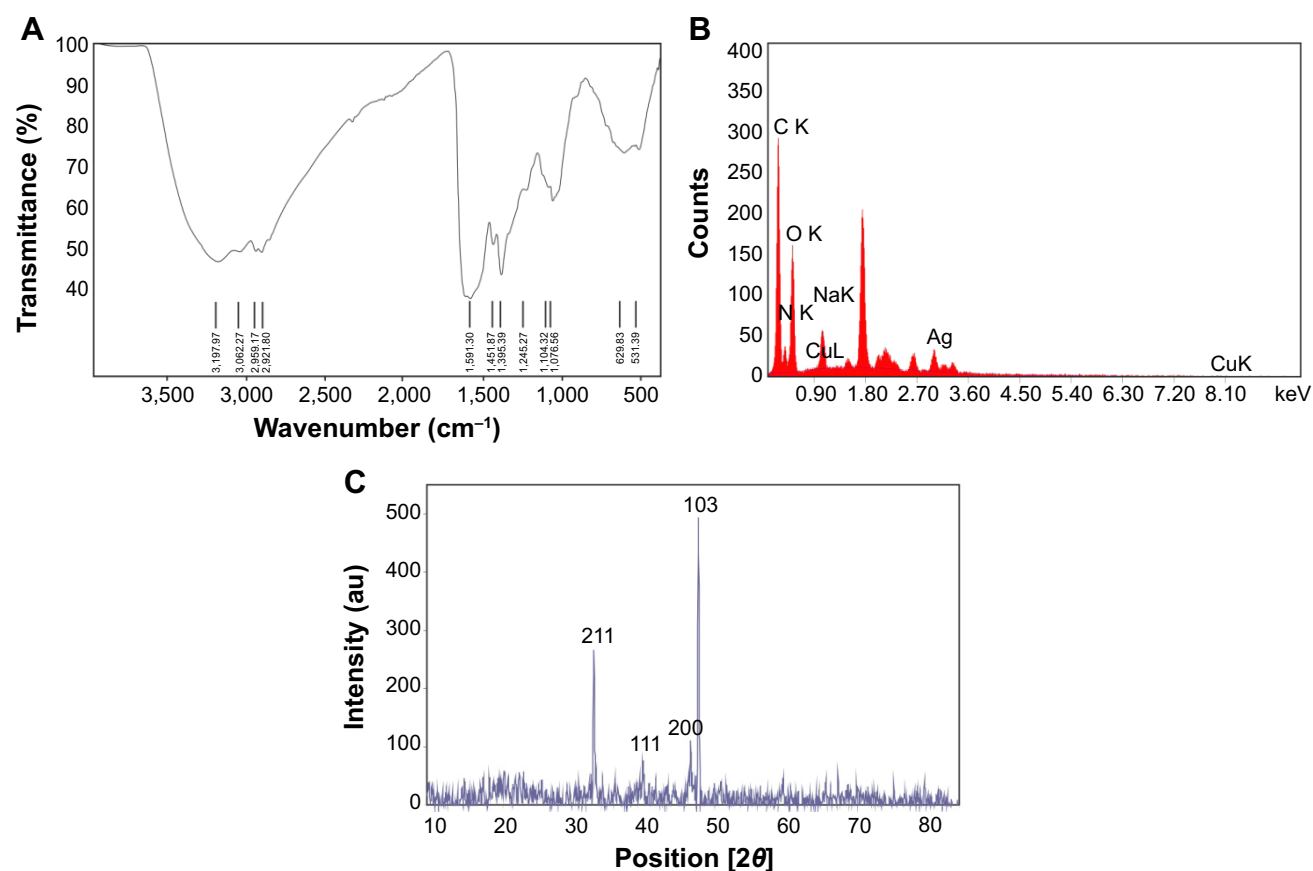


Figure 4 Characterization of silver nanoparticles.

Notes: (A) FTIR spectrum recorded from microbially synthesized silver nanoparticle powder using *Deinococcus radiodurans*. (B) EDX observation of biosynthesized AgNPs with their corresponding strong Ag signals at around 3 KeV. (C) XRD pattern of the AgNPs synthesized by the bacterium *D. radiodurans* in the form of a thin film deposited on a glass substrate. The Bragg reflections are identified in the XRD pattern.

Abbreviations: FTIR, Fourier transform infrared spectroscopy; EDX, energy-dispersive X-ray spectroscopy; AgNPs, silver nanoparticles; XRD, X-ray diffraction.

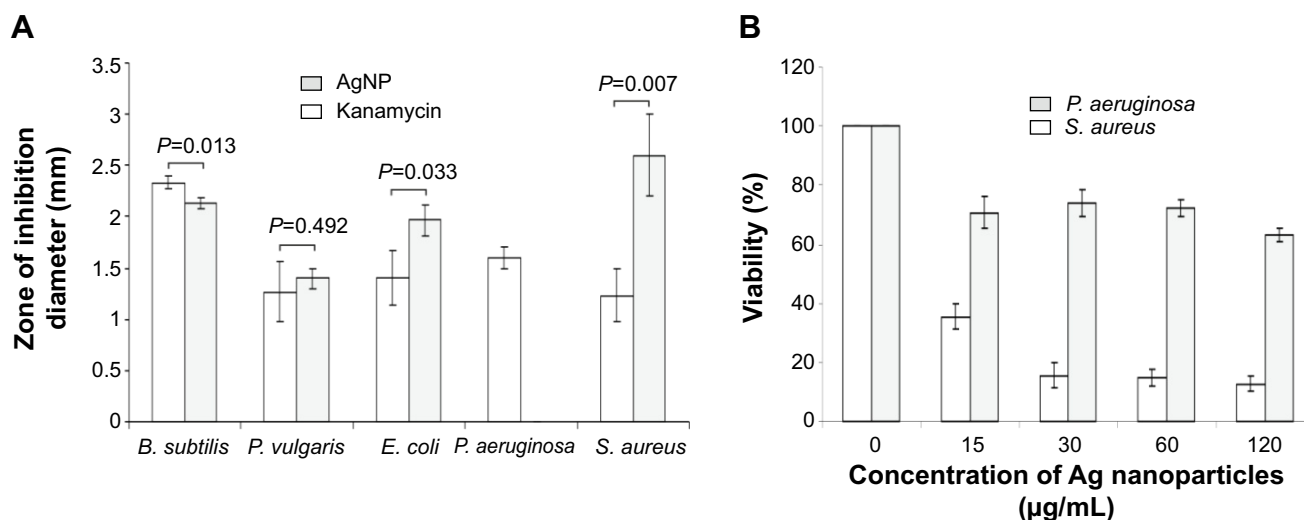


Figure 5 Antibacterial and Anti-biofouling activity of silver nanoparticles.

Notes: (A) Representative results of antimicrobial activity of nanoparticles against Gram-positive *Bacillus subtilis* and *Staphylococcus aureus* and Gram-negative *Escherichia coli*, *Proteus vulgaris*, and *Pseudomonas aeruginosa*. (B) Anti-biofouling activity of silver nanoparticles against Gram-negative *P. aeruginosa* and Gram-positive *S. aureus*. The figure depicts a graph showing percent viability of the *P. aeruginosa* (statistically significant $P=1.72 \times 10^{-6}$ [ANOVA] and $P<0.01$ [t-test]) and *S. aureus* ($P=4.64 \times 10^{-11}$ [ANOVA] and $P<0.0001$ [t-test]) against the varying concentrations of silver nanoparticles.

Abbreviation: ANOVA, analysis of variance.

B. subtilis, *Bacillus cereus*, *E. coli*, and *P. aeruginosa* has been reported.^{40–42} AgNPs synthesized using *D. radiodurans* exhibited an effective antibacterial activity at a concentration of 150 µg/mL. Antibiotic resistance is the biggest challenge to the medical field for the treatment of infectious diseases, particularly due to the emergence of multidrug-resistant pathogenic strains. Thus, AgNPs synthesized in the present study could offer a potential as an effective antibacterial agent alone or in combination for the management of antibiotic-resistant bacterial diseases after completing the successful clinical trials. In earlier reports on antibacterial activity of AgNPs, it has been observed that maximum inhibition of AgNPs was against Gram-negative microorganisms as compared to Gram-positive microorganisms,⁴³ which may be probably associated with the differences in their cell wall composition.^{39,44} However, the exact mechanism by which the AgNPs exert their antibacterial effect remains to be elucidated.

Anti-biofouling assay

Along with antibacterial activity, we evaluated the potential of AgNPs as an anti-biofouling agent against *S. aureus* and multidrug-resistant *P. aeruginosa*. AgNPs were capable of inhibiting the biofilm formation in both the microorganisms, *S. aureus* being more susceptible as compared to *P. aeruginosa*. The biofilm formation by the bacterial strains was inhibited in a dose-dependent manner (Figure 5B). Decrease in biofilm formation was observed with increasing concentration of AgNPs. This indicates that AgNPs

synthesized by *D. radiodurans* may provide a good alternative source of anti-biofouling compounds.

Anticancer activity

Microbially synthesized AgNPs have been well characterized as an antimicrobial agent. Although there are several reports on antitumor/anticancer activity of AgNPs synthesized using various plants,⁴⁵ there is limited information about antitumor/anticancer potential of AgNPs generated using microbial synthesis methods. This study is focused on determining cytotoxic effects of AgNPs on MCF-7 human breast cancer cells.

Optical microscopic study

MCF-7 cells exposed to different concentrations of AgNPs had alterations in cell shape and morphology indicating unhealthy cells, whereas untreated control cells exhibited no change in morphology (Figure 6). Nanoparticle-treated cells appeared to be clustered, and cell-spreading pattern was restricted as compared to untreated cells.

MTT assay

Cell viability and metabolic activity studies were conducted using MTT assay by exposing MCF-7 cells to AgNPs at 0–30 µg concentrations for 24 hours. The results of MTT assay showed a dose-dependent decrease in percent viability of the cells (Figure 7A). In the present study, cytotoxicity data were fitted to a sigmoidal curve, and a four-parameter nonlinear logistic model was used to calculate the LD₅₀ of

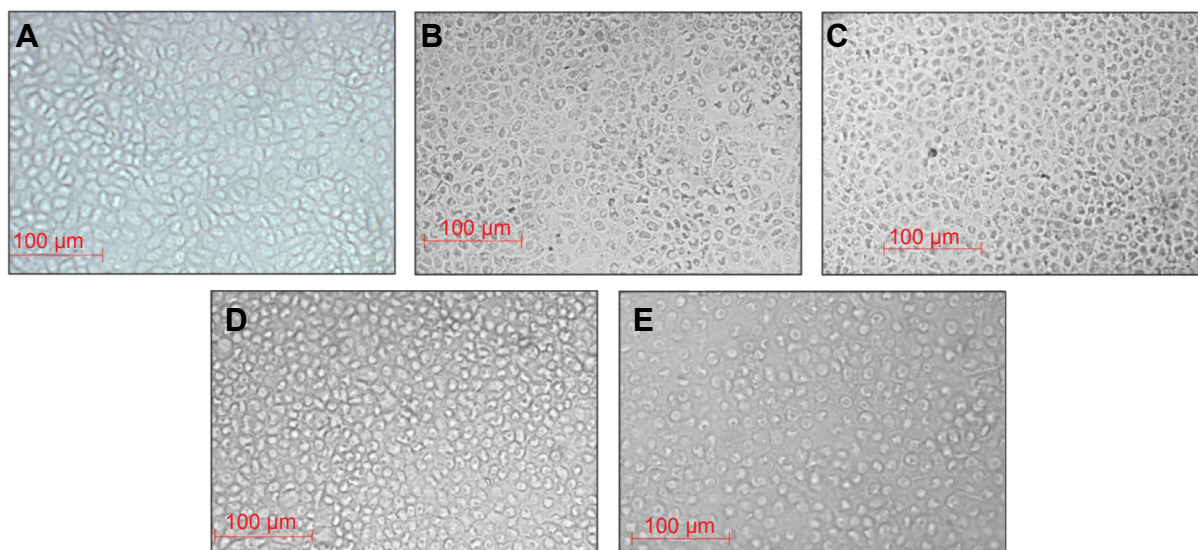


Figure 6 Cytotoxic activity of AgNPs.

Notes: Effect of AgNPs on human breast cancer cell line (MCF-7) at different concentrations: (A) control, (B) 1.65 µg/mL, (C) 3.75 µg/mL, (D) 7.5 µg/mL, and (E) 15 µg/mL.

Abbreviation: AgNPs, silver nanoparticles.

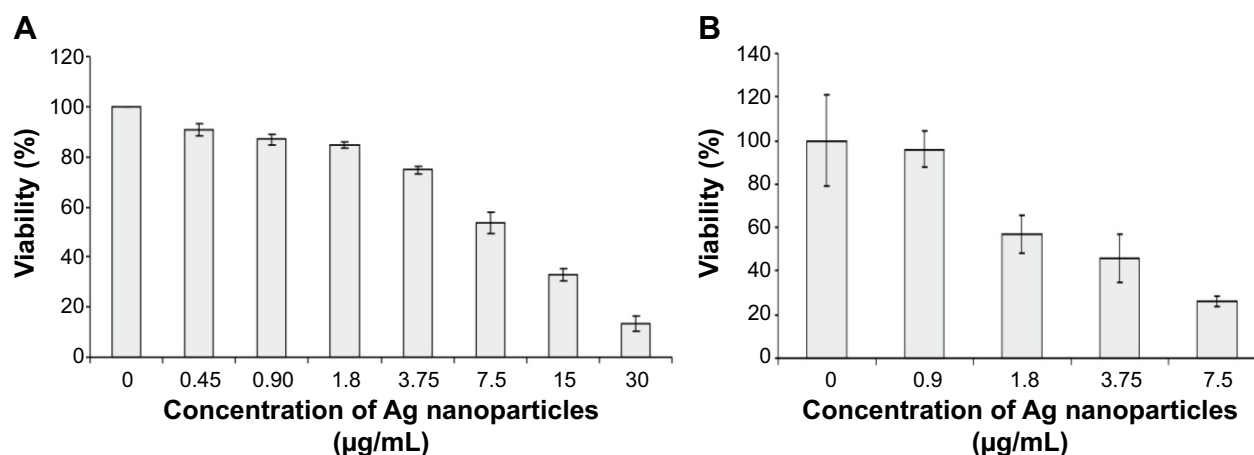


Figure 7 Percent viability of MCF-7 human breast cancer cells upon treatment with AgNPs.

Notes: (A) Graph showing percent viability of human breast cancer cell line (MCF-7) against various concentrations of the synthesized silver nanoparticles ($P=2.15 \times 10^{-17}$ [ANOVA]). (B) Colony-forming assay representing the percent viability of MCF-7 cell line against different concentrations of AgNPs below LD₅₀ after 10 days.

Abbreviations: ANOVA, analysis of variance; AgNPs, silver nanoparticles; LD₅₀, lethal dose.

nanomaterials that caused a 50% inhibition in comparison to untreated controls, which was 7.9 µg/mL. All LD₅₀ values were calculated using the average cytotoxicity data of the three independent experimental results and their associated errors. Exposure to increasing concentrations of AgNPs showed a dose-dependent cytotoxicity on the cancer cell line. It is reported that the LD₅₀ of AgNPs synthesized using various methods against MCF-7 breast cancer cell lines is in the range of 10–30 µg/mL.⁴⁶ In the present study, the LD₅₀ of AgNPs synthesized using *D. radiodurans* was found to be 7–8 µg/mL, which suggests that these nanoparticles are effective at a lower concentration against MCF-7 human breast cancer cell line compared to AgNPs synthesized using other sources. It is well studied that cytotoxicity of nanoparticles is size dependent with smaller nanoparticles entering the cells more easily and thus are more efficient than larger ones.⁴⁷ It is likely that the smaller size of AgNPs (average size of 16.82 nm) generated in this study using *D. radiodurans* may be one of the factors, and these particles could be further exploited for potential applications in cytotoxicity and anticancer activity.

Clonogenic survival

Clonogenic assay indicates the proliferative capacity of a single cell upon nanoparticle exposure for 10 days. MCF-7 cells were incubated with AgNPs and then stained with crystal violet stain. The recorded colony count represents the number of viable cells. The results of the colony-forming assay for cells treated with the AgNPs below LD₅₀ for 24 hours are presented in Figure 7B. It represents the dose response indicating the decrease in the colony number with the increasing concentration of AgNPs, thus demonstrating

the effective inhibition of growth and proliferation of the MCF-7 cancer cell line.

Conclusion

D. radiodurans is able to withstand 5,000× more radiation than any other known living organism and demonstrates ability to withstand exposure to desiccation. It is known to possess the ability to repair double-strand breaks, and metals such as manganese have been shown to have a role in radiation resistance. We report here a simple, rapid, and efficient method for the synthesis and extracellular accumulation of AgNPs using *D. radiodurans*. The synthesized AgNPs are spherical in shape and have an average particle size of 16.82 nm. We have demonstrated the potential of these AgNPs for several applications. They exhibited excellent broad-spectrum antimicrobial activity against both Gram-positive and Gram-negative bacteria and anti-biofouling activity against *P. aeruginosa* and *S. aureus*. Inhibitory effect of AgNPs on multidrug-resistant *P. aeruginosa* was also evident. AgNPs also showed effective anticancer activity against human breast cancer cell line (MCF-7). The cytotoxicity and mortality data highlight these AgNPs as potential anticancer agents. It will be possible to scale up AgNPs synthesis using *D. radiodurans* and explore possible applications based on its anticancer and antiproliferative activity. The biosynthesized AgNPs have proved to be potential candidates for medical applications where antifouling, antimicrobial, and cytotoxic activities are highly essential. Since *D. radiodurans* can sustain radiations, it may offer advantage over other organisms for on-field applications where the nanoparticle synthesis ability can be explored for bioremediation. In recent years, radioactive nanoparticles

have been explored as cancer therapeutic and imaging agents. Till date, fabrication of radioactive nanoparticles has not been performed using microorganism; therefore, *D. radiodurans* may serve as a promising candidate for the synthesis of radioactive nanoparticles.

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Authors contribution

RK and NS contributed equally in carrying out all experiments and jointly analyzed the data sets. DND and DDD conceived the experiments, supervised, and provided major intellectual inputs. All authors assisted with drafting the article and critically revised it, and all have approved the final article.

Disclosure

The authors report no conflicts of interest in this work.

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