

Antimicrobial activity of metal oxide nanoparticles against Gram-positive and Gram-negative bacteria: a comparative study

Ameer Azam^{1,2}
Arham S Ahmed²
Mohammad Oves³
Mohammad S Khan³
Sami S Habib¹
Adnan Memic¹

¹Centre of Nanotechnology, King Abdulaziz University, Jeddah, Saudi Arabia; ²Centre of Excellence in Materials Science (Nanomaterials), ³Department of Agricultural Microbiology, Aligarh Muslim University, Aligarh, India

Background: Nanomaterials have unique properties compared to their bulk counterparts. For this reason, nanotechnology has attracted a great deal of attention from the scientific community. Metal oxide nanomaterials like ZnO and CuO have been used industrially for several purposes, including cosmetics, paints, plastics, and textiles. A common feature that these nanoparticles exhibit is their antimicrobial behavior against pathogenic bacteria. In this report, we demonstrate the antimicrobial activity of ZnO, CuO, and Fe₂O₃ nanoparticles against Gram-positive and Gram-negative bacteria.

Methods and results: Nanosized particles of three metal oxides (ZnO, CuO, and Fe₂O₃) were synthesized by a sol-gel combustion route and characterized by X-ray diffraction, Fourier-transform infrared spectroscopy, and transmission electron microscopy techniques. X-ray diffraction results confirmed the single-phase formation of all three nanomaterials. The particle sizes were observed to be 18, 22, and 28 nm for ZnO, CuO, and Fe₂O₃, respectively. We used these nanomaterials to evaluate their antibacterial activity against both Gram-negative (*Escherichia coli* and *Pseudomonas aeruginosa*) and Gram-positive (*Staphylococcus aureus* and *Bacillus subtilis*) bacteria.

Conclusion: Among the three metal oxide nanomaterials, ZnO showed greatest antimicrobial activity against both Gram-positive and Gram-negative bacteria used in this study. It was observed that ZnO nanoparticles have excellent bactericidal potential, while Fe₂O₃ nanoparticles exhibited the least bactericidal activity. The order of antibacterial activity was demonstrated to be the following: ZnO > CuO > Fe₂O₃.

Keywords: nanomaterial, ZnO, CuO, Fe₂O₃, antibacterial activity, metal oxides

Introduction

The emergence of infectious diseases in general poses a serious threat to public health worldwide, especially with the emergence of antibiotic-resistant bacterial strains. Generally, both Gram-positive and Gram-negative bacterial strains are thought to present a major public health problem. Over the years, antibiotics have been used to control infections resulting from both community and hospital environments.¹⁻³ Current advances in the field of nanobiotechnology, particularly the ability to prepare metal oxide nanomaterials of specific size and shape, are likely to lead to the development of new antibacterial agents. The functional activities of nanoparticles are influenced largely by the particle size. Therefore, nanoparticles have received great attention due to their unique physical, chemical, and effective biological properties in various fields, including medicine. The properties of nanoparticles can easily be altered by reducing or changing their size, especially when the manipulations are done at the

Correspondence: Ameer Azam
Centre of Nanotechnology, King Abdulaziz University, Abdullah Sulayman, Jeddah 22254, Saudi Arabia
Email azam222@rediffmail.com

nanometer scale.⁴⁻⁷ Similarly, tailoring of materials at the atomic level in order to attain unique properties has been widely reported. Considering these unique properties, nano-sized organic and inorganic particles are being generated for ultimate use in medical practices, such as metal oxides of zinc, copper, and iron in biomedical research.^{8,9} In addition, nanoparticles with smaller particle size have been reported to show good antimicrobial activity.¹⁰ Antimicrobial activity of nanoparticles has largely been studied with human pathogenic bacteria such as *Escherichia coli*¹¹ and *Staphylococcus aureus*.¹² Moreover, these microbes seem to be highly sensitive to ZnO and CuO nanoparticles.^{10,13} Bactericidal activity of such nanoparticles in part depends on (1) size, (2) stability, and (3) concentration in the growth medium. While growing in medium amended with nanoparticles, the bacterial population growth can be inhibited by specific nanoparticle interactions.⁷ In general, bacterial cell size is in the micrometer range, while its outer cellular membranes have pores in the nanometer range. Since nanoparticles can be smaller in size than bacterial pores, they will have a unique ability of crossing the cell membrane. There lies a strong challenge in preparing metal oxide nanomaterials stable enough to restrict bacterial growth significantly while in nutrient medium.

In comparison to published reports on physical and chemical properties, very limited information is available on the antimicrobial properties of metal oxide nanoparticles. Realizing the potential antimicrobial applications of metal oxide nanoparticles, we designed experiments to synthesize ZnO, CuO, and Fe₂O₃ nanoparticles using a gel-combustion method and subsequently tested their antibacterial activities against both Gram-positive (*S. aureus* and *Bacillus subtilis*) and Gram-negative (*Pseudomonas aeruginosa* and *E. coli*) bacterial strains. Furthermore, the antibacterial behavior of these metal oxide nanoparticles was compared.

Materials and methods

Synthesis and characterization

In a typical synthesis procedure, metal nitrates of Zn, Cu, and Fe and citric acid were dissolved in distilled water with a molar ratio of 1:1. The solutions were stirred with a magnetic stirrer at 100°C. Stirring continues till the formation of gel for approximately 2 hours. As the gel was formed, it was allowed to burn at 200°C. A light fluffy mass was obtained as a result of combustion, which was further annealed at 400°C for 1 hour to obtain the respective crystalline metal oxide nanoparticles.¹⁴ The metal oxide nanoparticles thus obtained were characterized by X-ray diffraction (XRD), Fourier-transform infrared (FTIR) spectroscopy, and transmission

electron microscopy (TEM). Crystallinity, structure, and crystallite size of nanoparticles were determined by XRD using a Rigaku (Tokyo, Japan) Miniflex X-ray diffractometer with Cu-K α radiations ($\lambda = 0.15406$ nm) in the 2θ range from 20° to 80°. FTIR spectra of the samples were obtained using a PerkinElmer (Waltham, MA) FTIR spectrophotometer in the KBr matrix. TEM analysis was carried out using a 200 kV JEOL (Tokyo, Japan) transmission electron microscope.

Determination of antibacterial activity by well-diffusion method

Antimicrobial activities of the synthesized metal oxide nanoparticles were performed against both Gram-negative (*E. coli* and *P. aeruginosa*) and Gram-positive (*B. subtilis* and *S. aureus*) bacteria. The antibacterial activity was done by modified Kirby-Bauer disk diffusion method.¹⁵ In brief, the pure cultures of organisms were subcultured in Müller-Hinton broth at 35°C \pm 2°C on a rotary shaker at 160 rpm. For bacterial growth, a lawn of culture was prepared by spreading the 100 μ L fresh culture having 10⁶ colony-forming units (CFU)/mL of each test organism on nutrient agar plates with the help of a sterile glass-rod spreader. Plates were left standing for 10 minutes to let the culture get absorbed. Then 8 mm wells were punched into the nutrient agar plates for testing nanomaterial antimicrobial activity. Wells were sealed with one drop of molten agar (0.8% agar) to prevent leakage of nanomaterials from the bottom of the wells. Using a micropipette, 100 μ L (50 μ g) of the sample of nanoparticle suspension was poured onto each of five wells on all plates. After overnight incubation at 35°C \pm 2°C, the different levels of zone of inhibition were measured. Solvent blank was used as negative control. Antibiotic tetracycline was used as a positive control.

Determination of minimal bactericidal concentrations

Bacterial minimum bactericidal concentration (MBC) for metal oxide nanoparticles was determined by the broth-dilution method.¹⁶ In the present experiment, we used both Gram-positive and Gram-negative bacterial strains. Control experiments were also carried out in the presence of known standard antibiotics (tetracycline). A 10 mL nutrient broth medium amended with metal oxide nanomaterials (10–100 μ g/mL) was prepared separately. Each set was inoculated aseptically with 100 μ L of respective bacterial suspension (10⁶ CFU/mL). The inoculated sets were incubated at 35°C \pm 2°C for 24 hours. Viable bacterial colonies were counted and recorded by the naked eye determining the lowest concentration that locked bacteria growth, defining

this as the MBC. The experiments were carried out in triplicate, and averages were reported.

Results

Morphological analysis

The typical XRD patterns of the ZnO, CuO, and Fe₂O₃ nanoparticles annealed at 400°C are shown in Figure 1. The peak positions of samples exhibit the hexagonal, monoclinic, and rhombohedral structures of ZnO, CuO, and Fe₂O₃, which were confirmed from the International Centre for Diffraction Data card numbers 80-0075, 80-1916, and 85-0987, respectively. Furthermore, no impurity peaks were observed in the XRD patterns, as all of the three metal oxides showed single-phase sample formation. The crystallite size was calculated using the Scherrer formula,

$$D = \frac{0.9\lambda}{\beta \cos \theta} \quad (1)$$

where λ is the wavelength of X-ray radiation and β is the full width at half maximum of the peaks at the diffracting angle θ . Crystallite sizes were calculated to be 18 nm, 22 nm, and 26.1 nm for ZnO, CuO, and Fe₂O₃ nanoparticles, respectively.

Figure 2 exhibits TEM images and histograms of particle-size distribution of ZnO, CuO, and Fe₂O₃ nanoparticles

sintered at 400°C. Average particle sizes obtained from TEM images were found to be 19.89 ± 1.43 nm, 29.11 ± 1.61 nm, and 35.16 ± 1.47 nm for ZnO, CuO, and Fe₂O₃ nanoparticles, respectively. The average particle sizes determined by TEM images were very close to the crystallite size calculated from XRD results. Thus, the TEM results correlate well with XRD results.

FTIR spectra were recorded in solid phase using the KBr pellet technique in the regions of 3500–400 cm⁻¹. FTIR spectra of ZnO, CuO, and Fe₂O₃ nanoparticles are shown in Figure 3. FTIR spectra of all three metal oxide (ZnO, CuO, and Fe₂O₃) nanoparticles exhibited vibrations in the region 400–600 cm⁻¹, which can be attributed to the vibrations of M–O (M = Zn, Cu, and Fe) which confirms the formation of ZnO, CuO and Fe₂O₃ nanoparticles. A weak band at around 2300 cm⁻¹ may be attributed to the vibrations of atmospheric CO₂. In the case of Fe₂O₃, the bands appearing at 1632 cm⁻¹ can be attributed to the angular deformation of water δ H–OH, while the band appearing at 3436 cm⁻¹ can be assigned to the O–H stretching of water. The present findings agree well with the values reported in the available literature.^{17–21}

Antimicrobial properties

Antibacterial activity results revealed that ZnO and CuO nanoparticles acted as excellent antibacterial agents against both Gram-positive and Gram-negative bacteria when

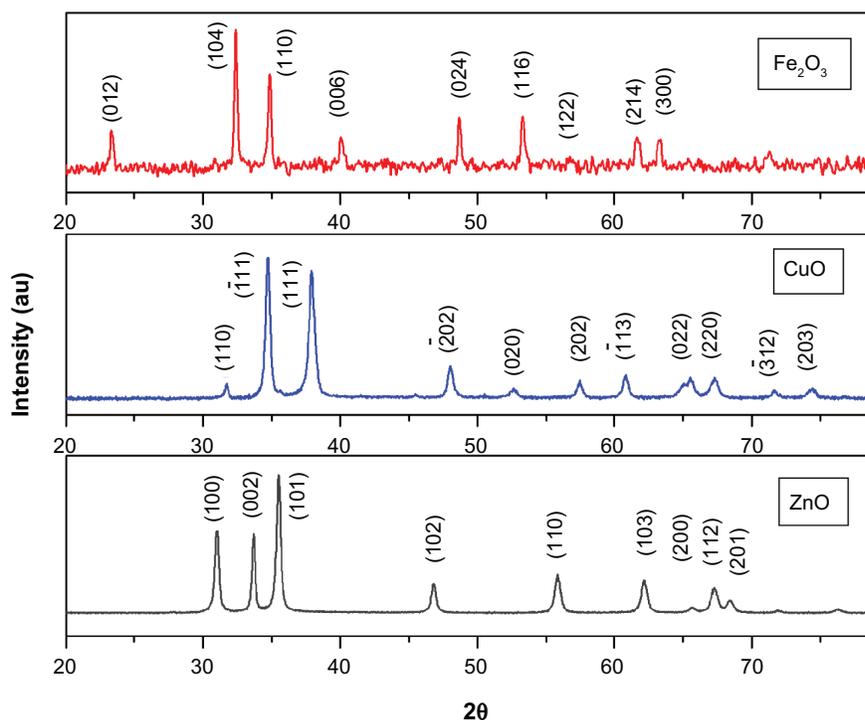


Figure 1 X-ray diffraction spectra of ZnO, CuO, and Fe₂O₃ nanoparticles.

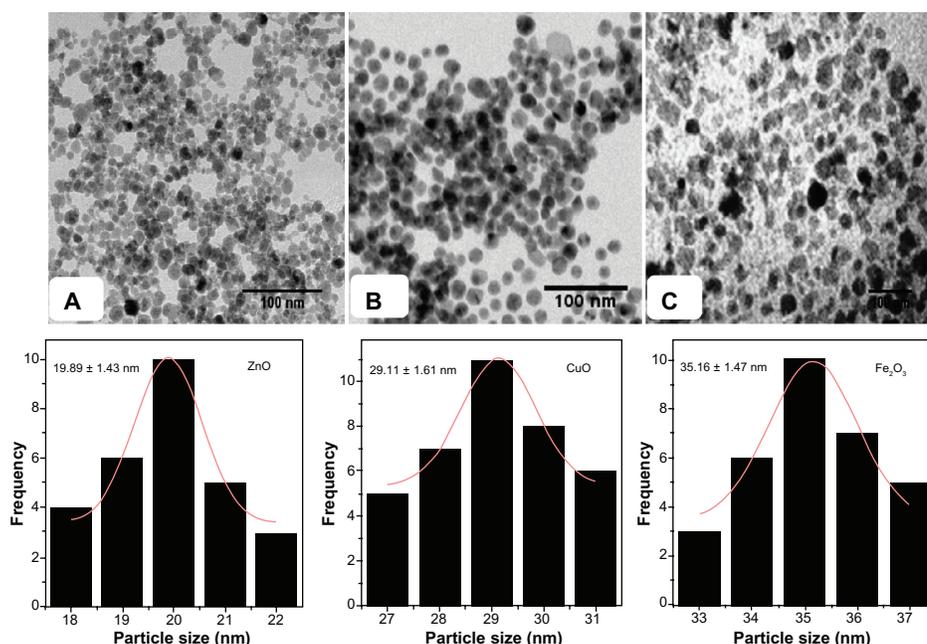


Figure 2 Transmission electron microscopy images of (A) ZnO, (B) CuO, and (C) Fe₂O₃ nanoparticles and histogram of particle-size distribution for different metal oxide nanoparticles.

compared to Fe₂O₃ nanoparticles. It is clear from the XRD and TEM results that ZnO nanoparticles are smaller in size compared to CuO and Fe₂O₃. Figure 4 shows the zone of inhibition produced by different metal oxide nanoparticles against both Gram-positive and Gram-negative bacterial strains. ZnO (19.89 ± 1.43 nm) nanoparticles exhibited maximum

(25 mm) bacterial growth inhibition against *B. subtilis*, in the form of zone-of-inhibition studies, where diffusion of nanoparticles on nutrient agar plates inhibits growth. In contrast, CuO and Fe₂O₃ showed zones of inhibition of 21 and 15 mm, respectively, against *B. subtilis*. In the case of *E. coli* maximum growth, inhibition zones were found to be

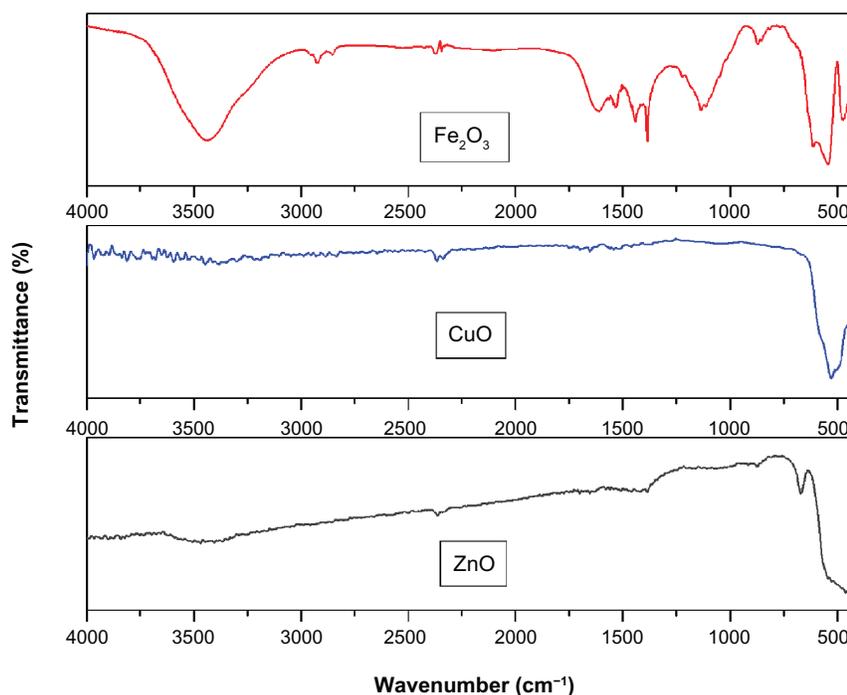


Figure 3 Fourier-transform infrared spectra of ZnO, CuO, and Fe₂O₃ nanoparticles.

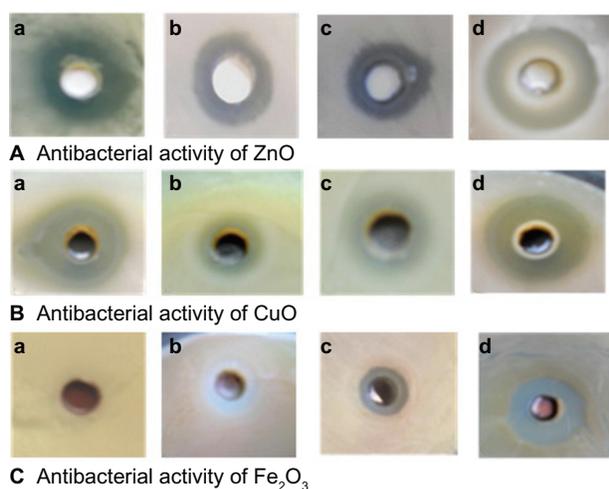


Figure 4 Zone of inhibition produced by different metal oxide nanoparticles against both Gram-positive and Gram-negative bacterial strains. Antibacterial activity of (A) ZnO; (B) CuO; and (C) Fe₂O₃ of bacterial strains (a), *Escherichia coli*, (b) *Staphylococcus aureus*, (c) *Pseudomonas aeruginosa*, and (d) *Bacillus subtilis*.

the following: 19, 15, and 3 mm for ZnO, CuO, and Fe₂O₃, respectively (Figure 5). Similar patterns were observed in the case of *P. aeruginosa* and *S. aureus*, where the maximum zone of inhibition was exhibited by ZnO followed by CuO and Fe₂O₃. It appears that the antibacterial activity of the nanomaterials increased with increase in surface-to-volume ratio due to the decrease in size of nanoparticles.

Discussion

We demonstrated that the order of antibacterial activities of nanomaterials was ZnO (19.89 ± 1.43 nm) > CuO (29.11 ± 1.61 nm) > Fe₂O₃ (35.16 ± 1.47 nm), which indicates the size of the nanoparticles might also play a role in the antibacterial activity of each sample. Similar activity observations have been made for nanoparticles composed of a single metal oxide.^{6,7,10} However, it should also be noticed that Gram-negative bacterial strains of *E. coli* and

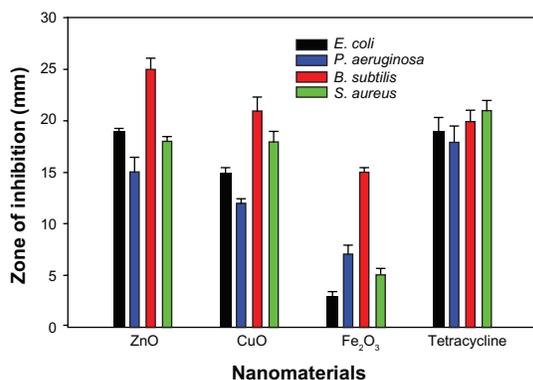


Figure 5 Bar graphs showing zone of inhibition introduced by different metal oxides against various microorganisms.

P. aeruginosa had inhibition-zone sizes that were 24% and 16% lower than Gram-positive bacterial strains of *B. subtilis* and *S. aureus* in the case of ZnO nanoparticles. And in the case of CuO nanoparticles, same Gram-negative bacterial strains of *E. coli* and *P. aeruginosa* had zone sizes that were 28% and 33% lower than Gram-positive *B. subtilis* and *S. aureus* bacterial strains, respectively. This observation could also be indicative of higher Gram-negative strain resistance/tolerance against such nanomaterials over Gram-positive bacterial strains. Our finding is in agreement with Premanathan et al, who reported that the ZnO nanoparticle effect is more pronounced against Gram-positive bacterial strains than Gram-negative bacterial strains.²²

Furthermore, previous studies have shown that the smaller the ZnO particle size, the greater the efficacy in inhibiting the growth of bacteria, involving both the production of reactive oxygen species and the accumulation of nanoparticles.^{7,10,23} However, nanoparticles of ZnO were previously reported to act both as bactericidal agents²⁴ and bacteriostatic agents,²⁵ perhaps thereby limiting their biomedical use.

In another experiment, we analyzed the MBC of different metal oxide nanoparticles against both Gram-negative and Gram-positive bacterial strains (Table 1). ZnO nanoparticles were also found to be most bactericidal compared to Fe₂O₃ and CuO nanoparticles. In our MBC study of ZnO nanoparticles, ZnO was 72%, 80%, 88%, and 84% more effective than Fe₂O₃, while 28%, 31%, 27%, 50%, and 40% more bactericidal than CuO against against *E. coli*, *S. aureus*, *P. aeruginosa*, and *B. subtilis*, respectively. The bactericidal pattern of our synthesized nanomaterials against both Gram-negative and Gram-positive bacterial strains was again ZnO > CuO > Fe₂O₃. Our results are supported by Baek and An²⁶ and Wang et al,²⁷ who reported that ZnO was the most toxic nanomaterial among ten other nanomaterials. As previously observed with zone-of-inhibition studies, ZnO nanoparticles had 11% and 12% more bactericidal activity against Gram-positive *S. aureus* and *B. subtilis* than Gram-negative *E. coli* and *P. aeruginosa*, respectively. CuO nanoparticles were 12% and 21% more active against Gram-positive *S. aureus* and *B. subtilis* than Gram-negative *E. coli* and *P. aeruginosa*, respectively. Overall, our observations are that Gram-positive bacterial strains are more sensitive in comparison to Gram-negative strains against the nanomaterials tested.

Thus, in this report, ZnO nanoparticles have shown the best antibacterial behavior compared to CuO and Fe₂O₃ nanoparticles. However, it is important to identify the key physicochemical properties of nanometal oxides that govern antibacterial activity and cytotoxicity to mammalian cells,

Table 1 Minimum bactericidal concentration (MBC) of different metal oxide nanoparticles against Gram-negative and Gram-positive bacteria

Nanoparticles	MBC values ($\mu\text{g/mL}$)			
	<i>Escherichia coli</i>	<i>Pseudomonas aeruginosa</i>	<i>Staphylococcus aureus</i>	<i>Bacillus subtilis</i>
ZnO	18 \pm 0.5	14 \pm 0.6	16 \pm 0.2	12 \pm 0.5
CuO	25 \pm 0.3	28 \pm 0.5	22 \pm 0.5	20 \pm 0.7
Fe ₂ O ₃	65 \pm 1.5	120 \pm 2.3	80 \pm 1.5	78 \pm 1.4
Tetracycline*	30 \pm 0.4	30 \pm 1.2	28 \pm 1.2	25 \pm 1.5

Notes: Values are mean of three replicates; *standard antibiotic.

as has been previously initiated. It is essential to assess the contribution of the size, shape, morphology, and electronic properties on cytotoxicity if these particles are to have wide biomedical applications.^{28,29} Our expectation is first to see such nanomaterials applied as surface disinfectants, as their stability would allow long-term storage and prolonged activity.

Conclusion

The nanosized particles of pure ZnO, CuO, and Fe₂O₃ were synthesized by the sol-gel combustion method. XRD and TEM results showed that ZnO nanoparticles were smallest (18 nm) in size compared to CuO (22 nm) and Fe₂O₃ (26 nm). Furthermore, the antibacterial activity of all the three synthesized nanomaterials was compared and varied considerably. Antimicrobial activity increased with increase in surface-to-volume ratio due to a decrease in particle size of nanoparticles. Here, ZnO nanoparticles showed excellent bactericidal potential, while iron oxide nanoparticles had the least bactericidal activity. Our results indicate that nanomaterials were most effective against Gram-positive bacterial strains compared to Gram-negative bacterial strains.

Acknowledgments

Mr Arham S Ahmed and Mr M Oves are thankful to CSIR, New Delhi, for providing financial support in the form of SRF. Dr Adnan Memic would like to thank the Strategic Technologies Program of King Abdulaziz City for Science and Technology, grant number 10-NAN1081-3, for their partial support and funding of this project.

Disclosure

The authors report no conflicts of interest in this work.

References

1. Lowy F. *Staphylococcus aureus* infections. *N Engl J Med*. 1998;339:520–532.
2. Komolafe OO. Antibiotic resistance in bacteria – an emerging public health problem. *Malawi Med J*. 2003;15:63–67.
3. Hawkey PM. The growing burden of antimicrobial resistance. *J Antimicrob Chemother*. 2008;62 Suppl 1:i1–i9.
4. Lewis K, Klibanov AM. Surpassing nature: rational design of sterile-surface materials. *Trends Biotechnol*. 2005;23:343–348.
5. Rosi NL, Mirkin CA. Nanostructures in biodiagnostics. *Chem Rev*. 2005;105:1547–1562.
6. Azam A, Ahmed AS, Oves M, Khan MS, Memic A. Size-dependent antimicrobial properties of CuO nanoparticles against Gram-positive and -negative bacterial strains. *Int J Nanomedicine*. 2012;7:3527–3535.
7. Raghupati KR, Koodali RT, Manna AC. Size-dependent bacterial growth inhibition and mechanism of antibacterial activity of zinc oxide nanoparticles. *Langmuir*. 2011;27:4020–4028.
8. Mahapatra O, Bhagat M, Gopalakrishnan C, Arunachalam KD. Ultrafine dispersed CuO nanoparticles and their antibacterial activity. *J Exp Nanosci*. 2008;3:185–193.
9. Tran N, Mir A, Mallik D, Sinha A, Nayar S, Webster TJ. Bactericidal effects of iron oxide nanoparticles on *Staphylococcus aureus*. *Int J Nanomedicine*. 2010;5:277–283.
10. Jones N, Ray B, Ranjit KT, Manna AC. Antibacterial activity of ZnO nanoparticles suspensions on a broad spectrum of microorganisms. *FEMS Microbiol Lett*. 2008;279:71–76.
11. Yoon KY, Byeon JH, Park JH, Hwang J. Susceptibility constants of *Escherichia coli* and *Bacillus subtilis* to silver and copper nanoparticles. *Sci Total Environ*. 2007;373:572–575.
12. Ruparelia JP, Chatterjee AK, Duttagupta SP, Mukherji S. Strain specificity in antimicrobial activity of silver and copper nanoparticles. *Acta Biomater*. 2008;4:707–716.
13. Heinlaan M, Ivask A, Blinova I, Dubourguier HC, Kahru A. Toxicity of nanosized and bulk ZnO, CuO and TiO₂ to bacteria *Vibrio fischeri* and crustaceans *Daphnia magna* and *Thamnocephalus platyurus*. *Chemosphere*. 2008;71:1308–1316.
14. Jundale DM, Pawar SG, Patil SL, Chougule MA, Godse PR, Patil VB. Effect of annealing on structure, morphology and optoelectronic properties of nanocrystalline CuO thin films. *AIP Conf Proc*. 2011;1391: 573–575.
15. Bauer AW, Kirby WMM, Sherris JC, Truck M. Antibiotic susceptibility testing by standardized single disk method. *Am J Clin Pathol*. 1966;45:493–496.
16. Wikler MA. *Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically: Approved Standard*. 5th ed. Wayne, PA: National Committee for Clinical Laboratory Standards (NCCLS); 2000:M7–M5.
17. Arshad M, Azam A, Ahmed AS, Mollah S, Naqvi AH. Effect of Co substitution on the structural and optical properties of ZnO nanoparticles synthesized by sol-gel route. *J Alloys Compd*. 2011;509:8378–8381.
18. Jagminas A, Kuzmarskyte J, Niaura G. Electrochemical formation and characterization of copper oxygenous compounds in alumina template from ethanolamine solutions. *Appl Surf Sci*. 2002;201:129–137.
19. Jagminas A, Niaura G, Kuzmarskyte J, Butkiene R. Surface-enhanced Raman scattering effect for copper oxygenous compounds array within the alumina template pores synthesized by ac deposition from Cu(II) acetate solution. *Appl Surf Sci*. 2004;225:302–308.

20. Zhang YC, Tang JY, Wang GL, Zhang M, Hu XY. Facile synthesis of submicron Cu₂O and CuO crystallites from a solid metallorganic molecular precursor. *J Cryst Growth*. 2006;294:278–282.
21. Ansari SA, Azam A, Naqvi AH. Structural and morphological study of Fe₂O₃ nanoparticles. *Asian J Res Chem*. 2011;4:1638–1642.
22. Premanathan M, Karthikeyan K, Jeyasubramanian K, Manivannan G. Selective toxicity of ZnO nanoparticles toward Gram-positive bacteria and cancer cells by apoptosis through lipid peroxidation. *Nanomedicine*. 2011;7:184–192.
23. Salah N, Habib SS, Khan ZH, et al. High-energy ball milling technique for ZnO nanoparticles as antibacterial material. *Int J Nanomedicine*. 2011;6:863–869.
24. Xie Y, He Y, Irvin PL, Jin T, Shi X. Antibacterial activity and mechanism of action of zinc oxide nanoparticles against *Campylobacter jejuni*. *Appl Environ Microbiol*. 2011;77:2325–2331.
25. Gajjar P, Pettee B, Britt DW, Huang W, Johnson WP, Anderson AJ. Antimicrobial activities of commercial nanoparticles against an environmental soil microbe, *Pseudomonas putida* KT2440. *J Biol Eng*. 2009;3:9.
26. Baek Y, An Y. Microbial toxicity of metal oxide nanoparticles (CuO, NiO, ZnO, and Sb₂O₃) to *Escherichia coli*, *Bacillus subtilis*, and *Streptococcus aureus*. *Sci Total Environ*. 2011;409:1603–1608.
27. Wang Z, Lee Y, Wu B, et al. Anti-microbial activities of aerosolized transition metal oxide nanoparticles. *Chemosphere*. 2010;80:525–529.
28. Xu M, Fujita D, Kajiwara S, et al. Contribution of physicochemical characteristics of nano-oxides to cytotoxicity. *Biomaterials*. 2010;31:8022–8031.
29. Brayner R. The toxicological impact of nanoparticles. *Nano Today*. 2008;3:48–55.

International Journal of Nanomedicine

Dovepress

Publish your work in this journal

The International Journal of Nanomedicine is an international, peer-reviewed journal focusing on the application of nanotechnology in diagnostics, therapeutics, and drug delivery systems throughout the biomedical field. This journal is indexed on PubMed Central, MedLine, CAS, SciSearch®, Current Contents®/Clinical Medicine,

Journal Citation Reports/Science Edition, EMBase, Scopus and the Elsevier Bibliographic databases. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <http://www.dovepress.com/international-journal-of-nanomedicine-journal>